

Development and Implementation of Human Intestinal Organoid Models for GI Toxicity

Julia Co, PhD

Sr. Principal Scientist, Complex *in vitro* Systems (CiS)

Safety Assessment, Genentech



COMPLEX IN VITRO
— SYSTEMS —

Genentech
A Member of the Roche Group

Goals for Intestinal *in vitro* Models

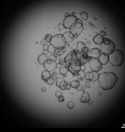
Goals

1. Enable modeling human GI biology with user-defined levels of complexity
2. Replace existing models (Caco-2, MDCK)
3. Reduce and sometimes replace animal models
4. Inform pre-clinical species selection

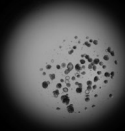
Safety Assessment

- De-risking and predicting GI tox
- Mechanisms of Tox
- Understanding GI tox in the clinic
- Stratifying patient populations/understanding patient diversity

Control

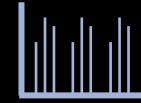
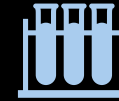


Treated



Biomarkers

- Identify novel biomarkers to determine target engagement and disease progression in the clinic



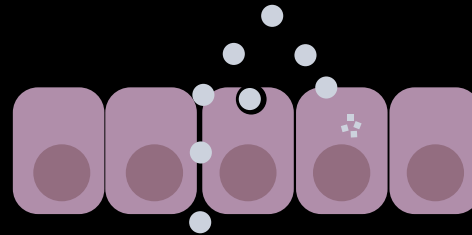
Disease Modeling and Discovery

- Determine mechanisms of action/pathogenesis
- Target discovery
- Screen libraries in a relevant GI model



ADME

- Drug uptake/transport/bioavailability
- Drug metabolism

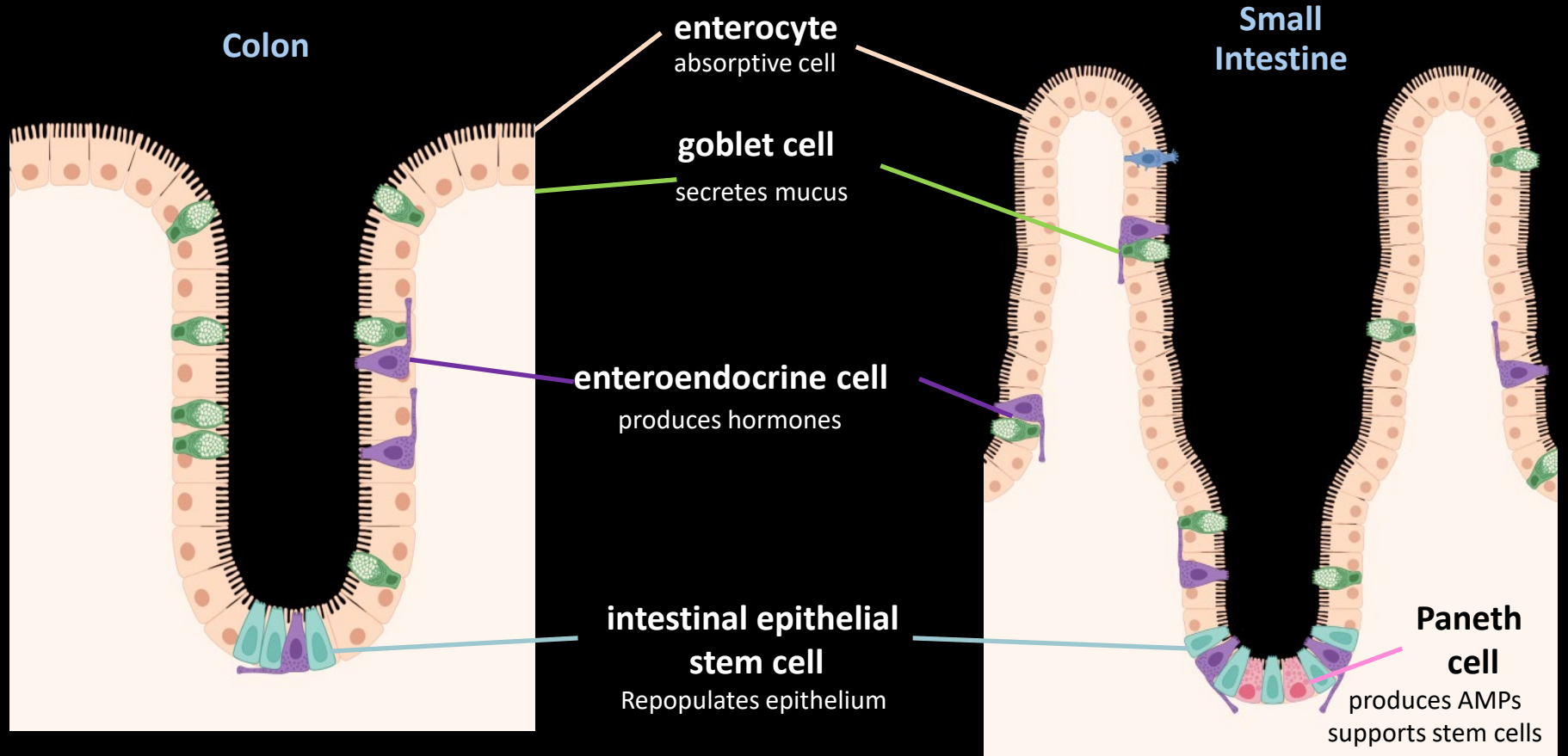


D&I

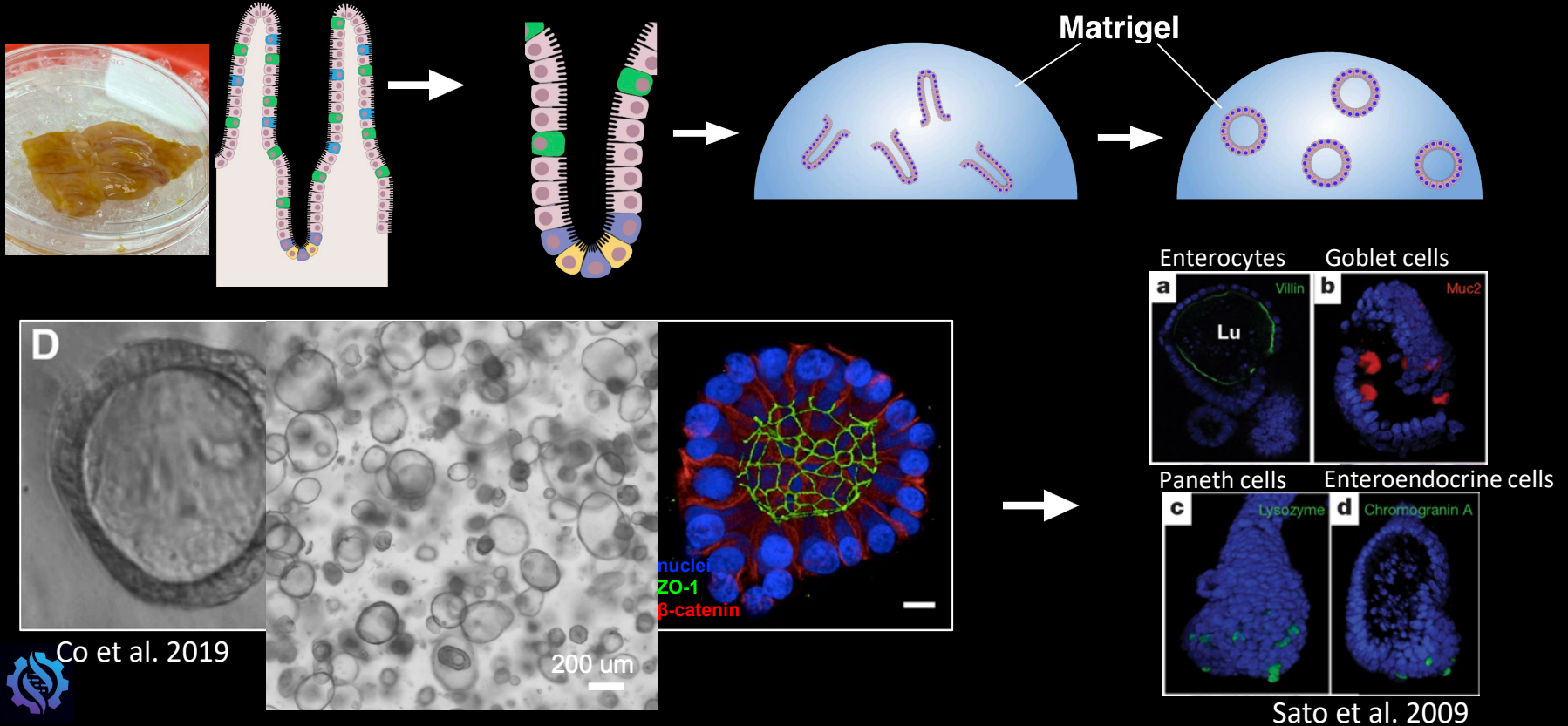
- Diversity of biological responses to molecules/compounds



Intestinal Epithelial Cells



Human Primary Tissue-Derived Intestinal Epithelial Organoids



Intestinal *in vitro* Models

ORGANOID CULTURE

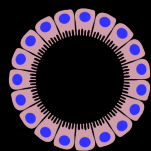
ORGANOID-DERIVED MODELS



Tissue



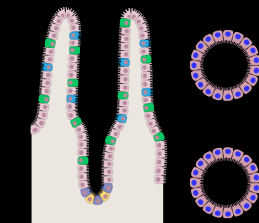
Freeze tissue fragments



Derive/grow organoids



Freeze organoids



Organoid spheroids

Stem cell organoids (crypt-like)

Differentiated organoids (villus-like)



Nuclei
ZO-1
Actin

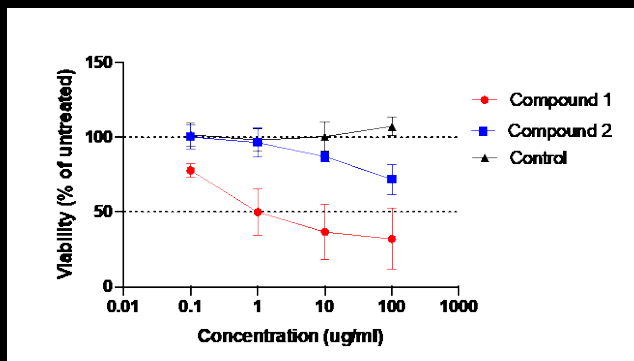
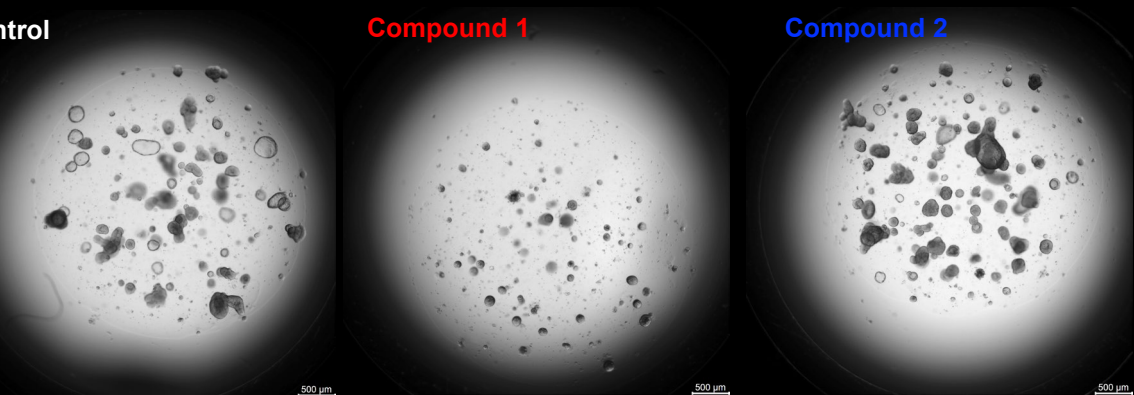


Organoids for Predictive GI Toxicity Studies

Control

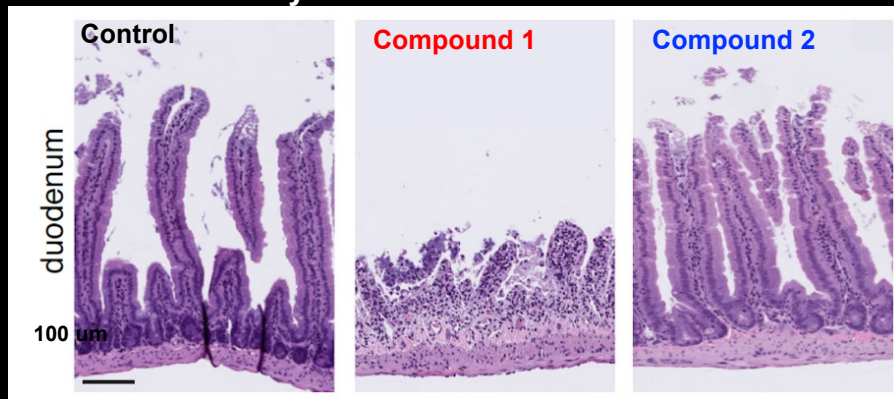
Compound 1

Compound 2



n=4 plates in 3 independent experiments, Error bars are S.D.
Viability determined by CellTiter Glo 3D ATP assay

in vivo mouse study



**Organoids facilitate
lead optimization and
reduction in scale of animal studies**



Intestinal Organoid Models for Safety Assessment



Jessica Klein



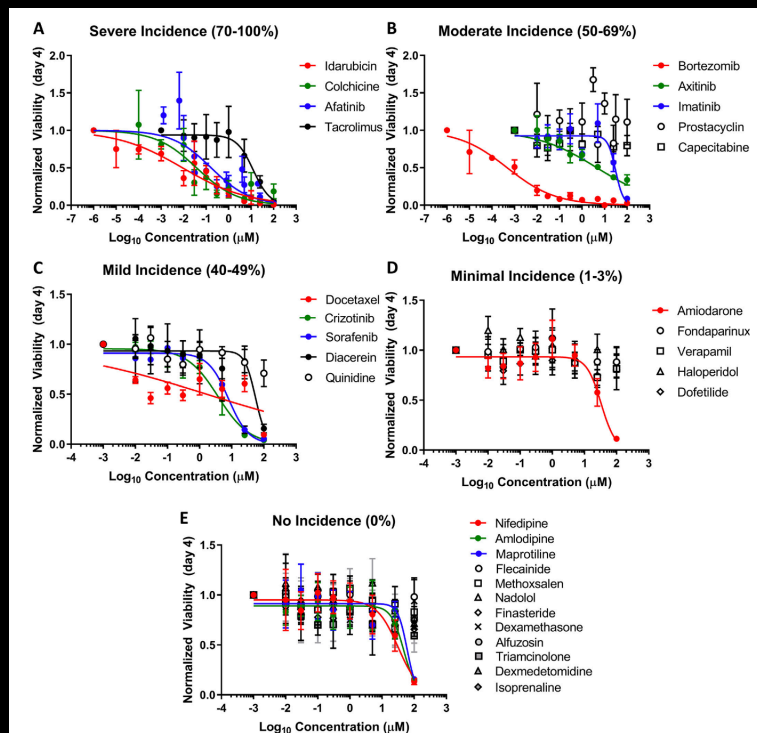
Julia Heidmann



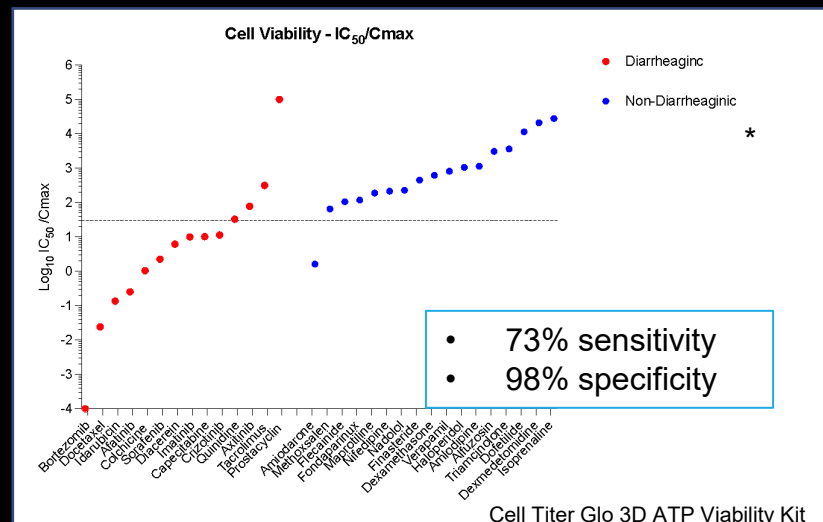
Lanlan Yu



Tomo Kiyota



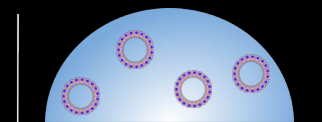
Validating Context-of-Use Assays



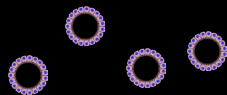
*where no IC₅₀ was achievable, maximum concentration tested was used to calculate ratio
 Compound list derived from Belair et al. Toxicol In Vitro 2020



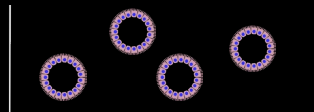
Organoid Polarity can be Reversed for Apical Access



Grow organoids submerged in Matrigel

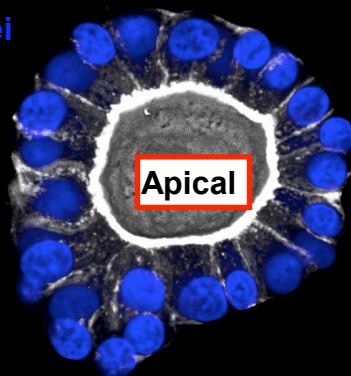


Dissociate ECM to isolate organoids



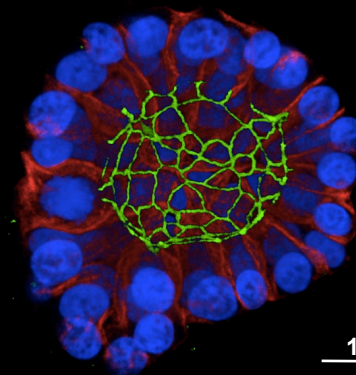
Suspend organoids in growth media

nuclei
actin



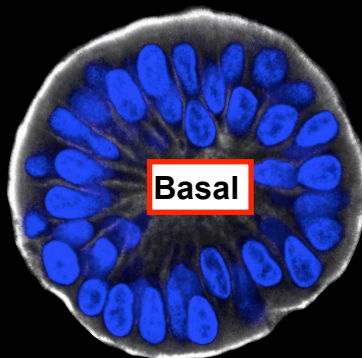
Basal

nuclei
ZO-1
(apical)
 β -catenin
(basolateral)

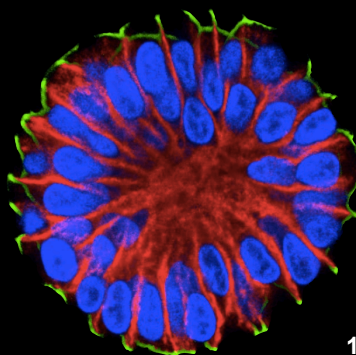


10 μ m

Basal



Apical

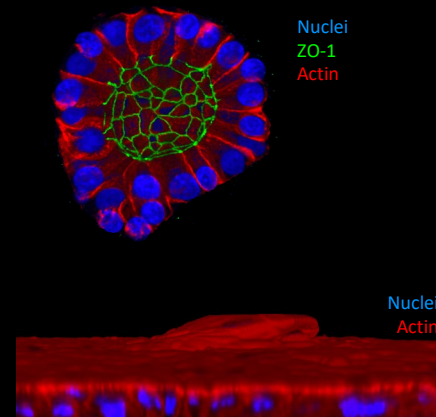
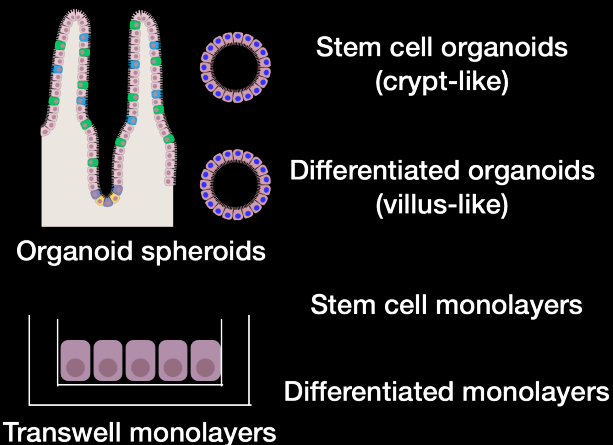
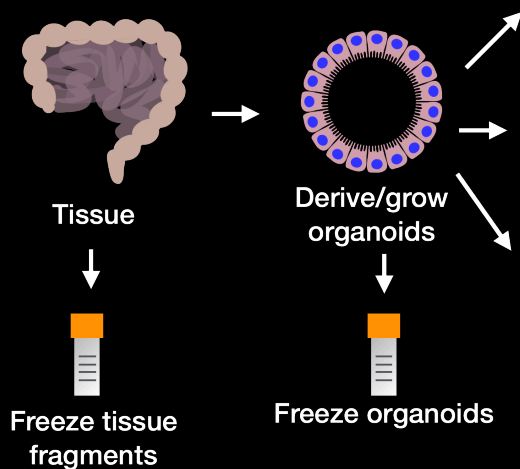


10 μ m

Intestinal *in vitro* Models

ORGANOID CULTURE

ORGANOID-DERIVED MODELS

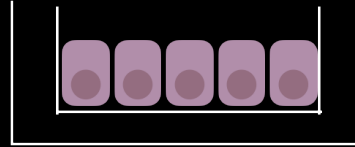


GI Organoid-Derived Transwell Monolayers

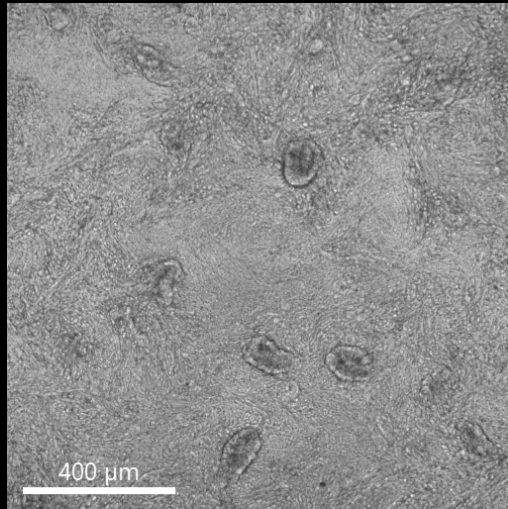
Cultivate organoids
in matrix



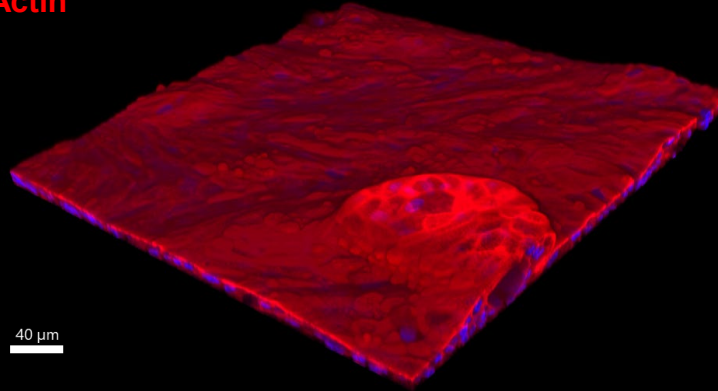
Dissociate to
single cells with
TrypLE Express



Seed on matrix-coated
transwell membrane



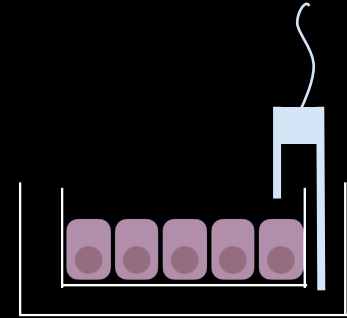
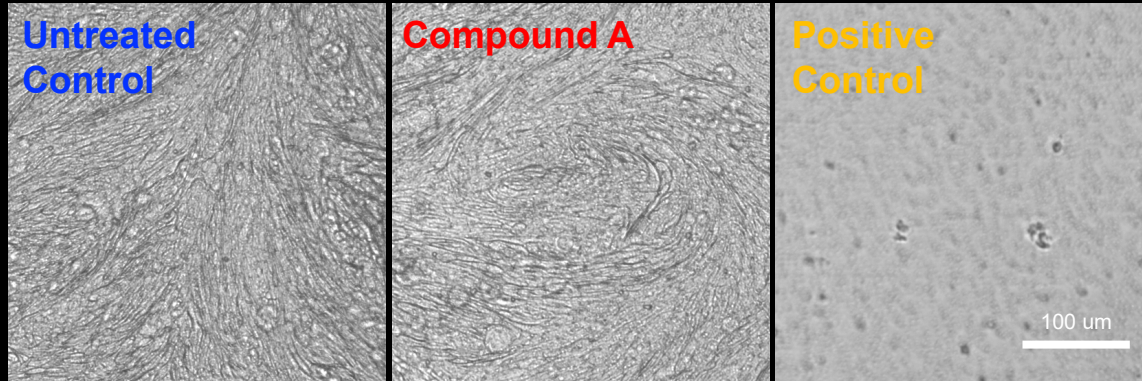
Nuclei
Actin



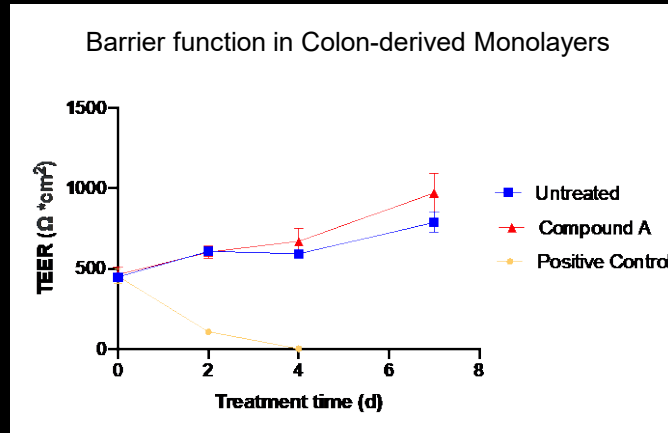
- Requires a lot of cells
- Enables apical/basolateral access
- Compatible with kinetic barrier function measurements



Organoid-Derived Transwell Monolayers for GI Toxicity



Trans-Epithelial Electrical Resistance (TEER)
as a measurement of barrier function



n = 4 replicates, Error bars represent S.D.

Organoid models enable
compound evaluation in
conditions not achievable *in vivo*



Intestinal *in vitro* Models

ORGANOID CULTURE

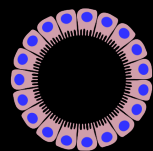
ORGANOID-DERIVED MODELS



Tissue



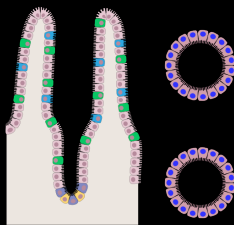
Freeze tissue fragments



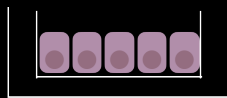
Derive/grow organoids



Freeze organoids



Organoid spheroids



Transwell monolayers



MPS chip

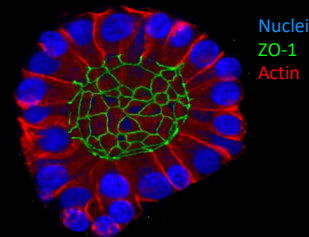
Stem cell organoids (crypt-like)

Differentiated organoids (villus-like)

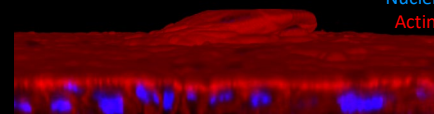
Stem cell monolayers

Differentiated monolayers

Differentiated MPS (+/- stretch)

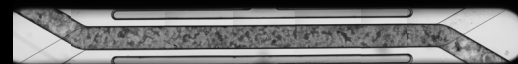


Nuclei
ZO-1
Actin



Nuclei
Actin

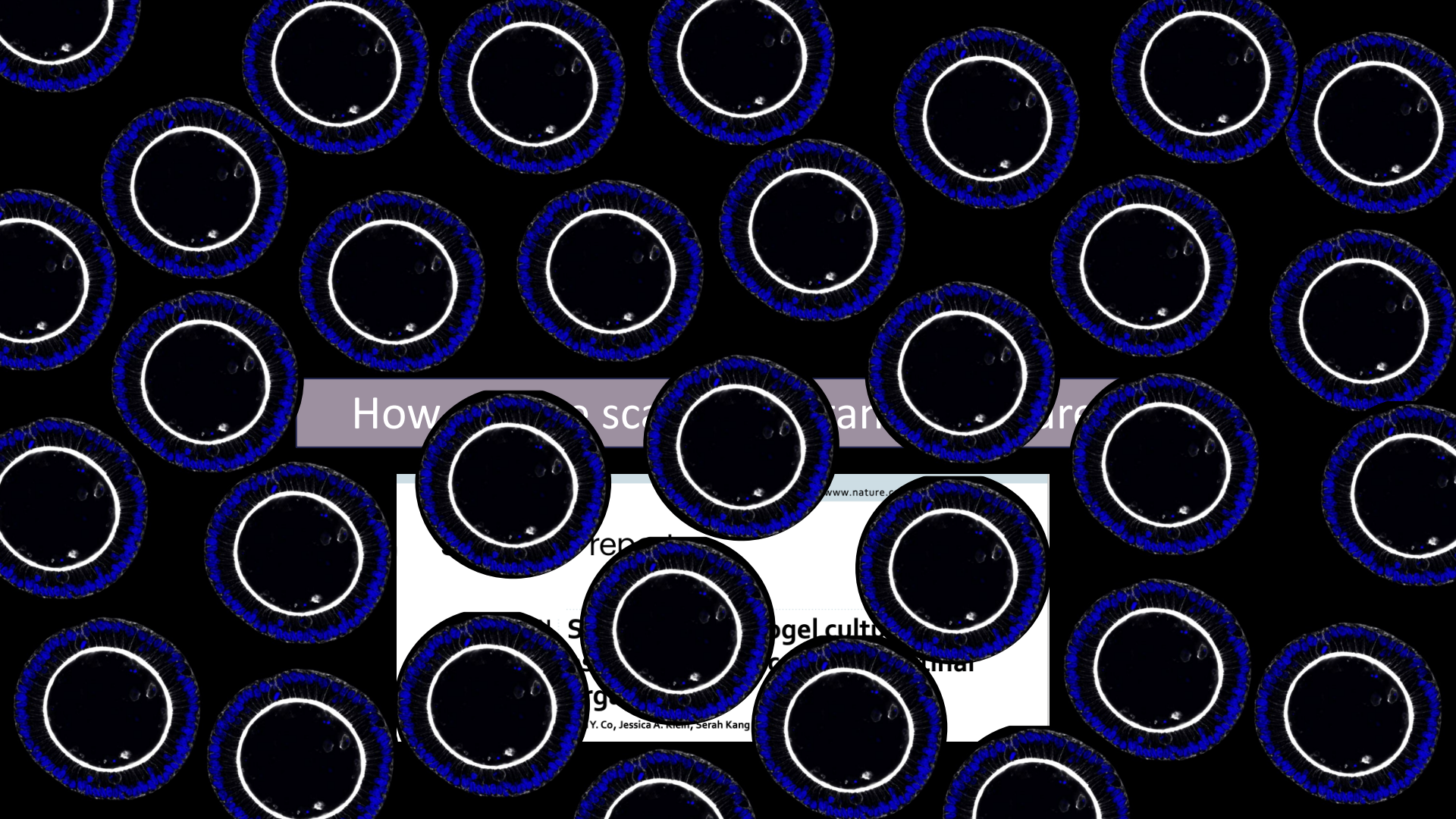
Emulate Colon Chip



Model Selection Considerations

- Scale per sample
- Scale – number of samples
- Apical/basal epithelium access
- Mechanical properties (fluid flow, stretch, etc.)
- Co-culture compatibility
- Multi-organ linkage
- Readout compatibility
 - Assay type
 - Live vs endpoint
 - Sample collection volume





How to scale up your culture

www.nature.com

rep

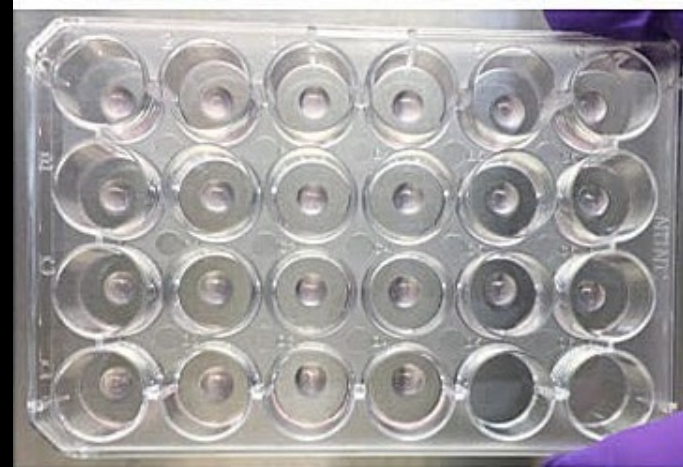
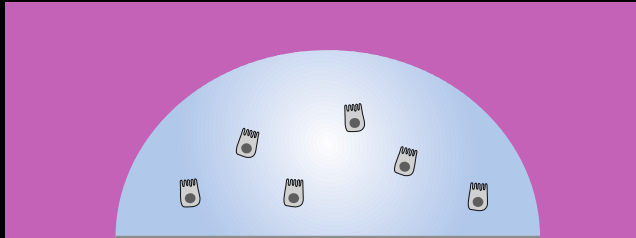
St. S. gel cultu

g

Y. Co, Jessica A. Klein, Serah Kang

Existing Organoid Culture Method

Attached BME Dome Culture

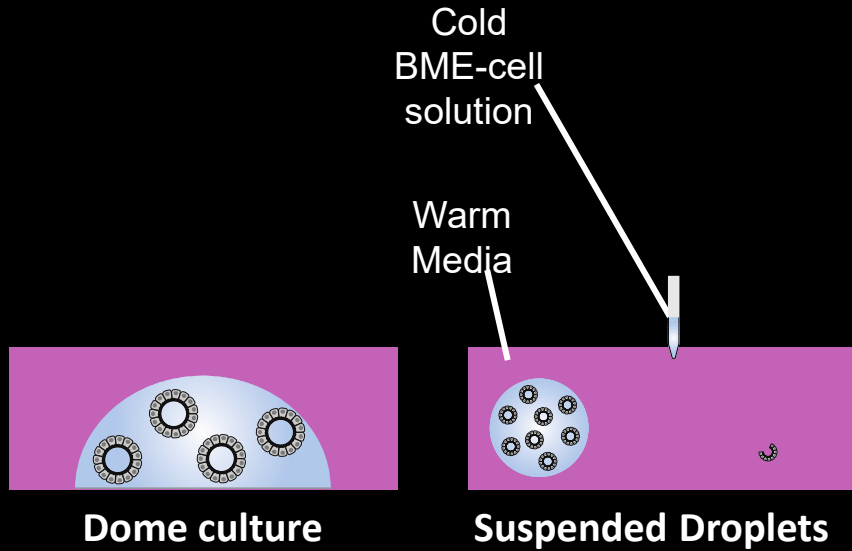


SigmaAldrich.com

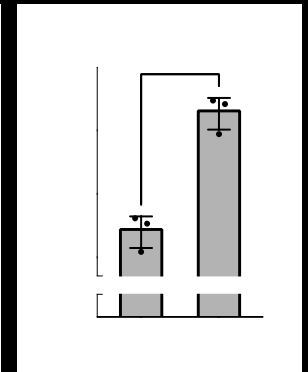
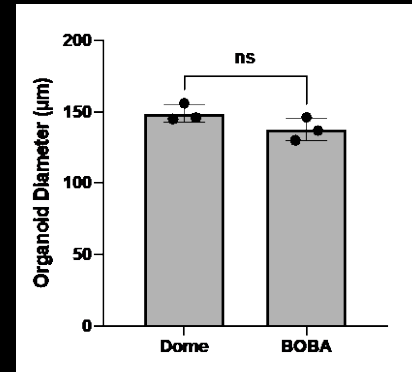
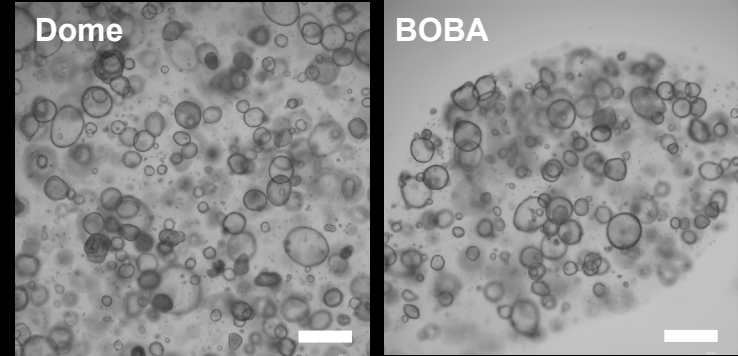
1. Deposit cells + cold BME/Matrigel (liquid) in center of well.
2. Incubate at 37C to cure BME/Matrigel.
3. Add media to each well.



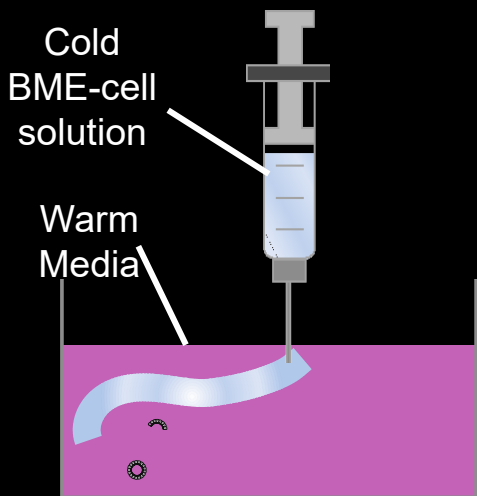
Suspended BME Hydrogel Organoid Culture



BOBA
(BME-embedded
Organoid Bead
Assemblies)



Suspended BME Hydrogel Organoid Culture

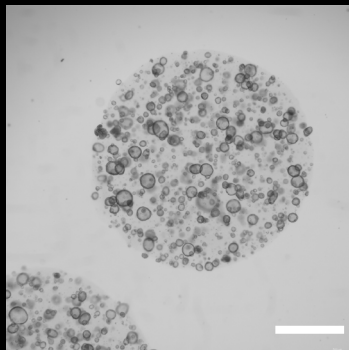


Suspended Filaments

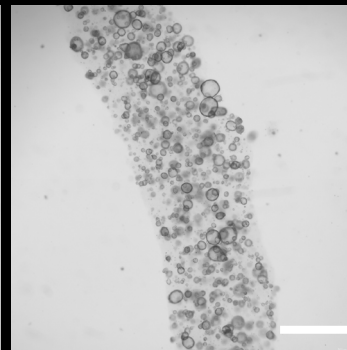
SOBA
(Syringe-extruded
Organoid BME
Assemblies)



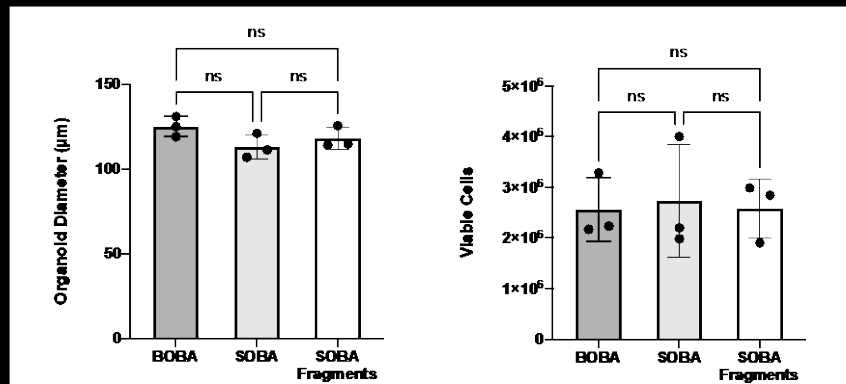
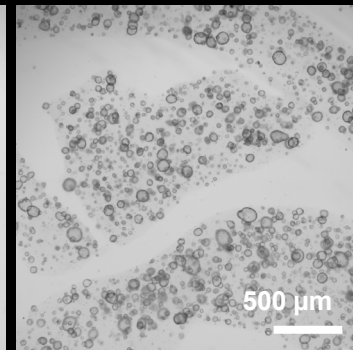
BOBA (droplets)



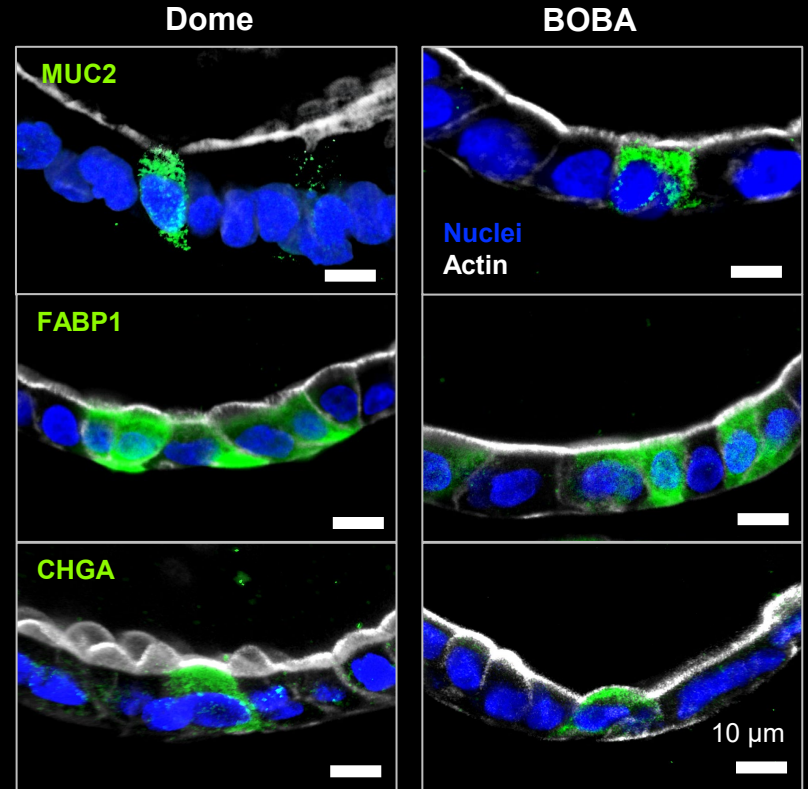
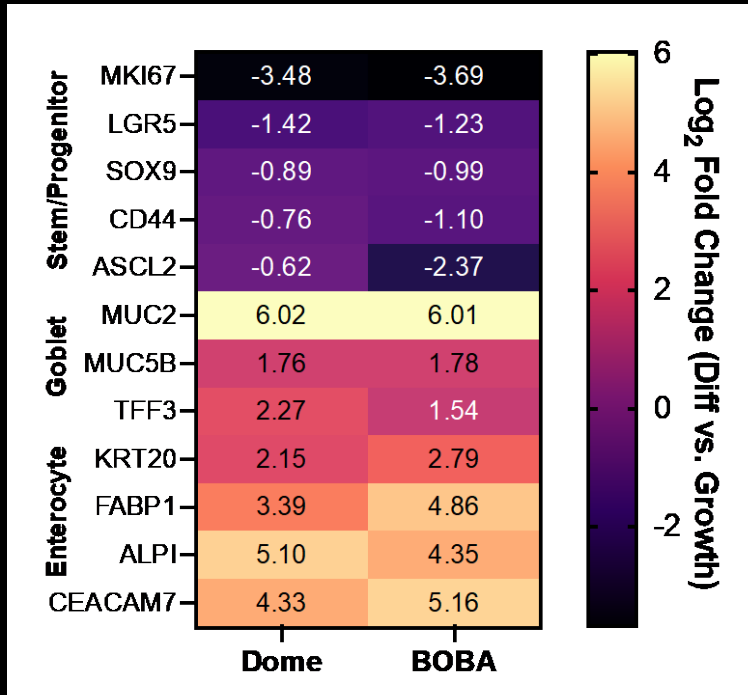
SOBA (filaments)



SOBA Fragments



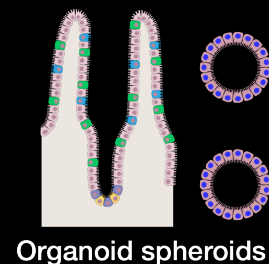
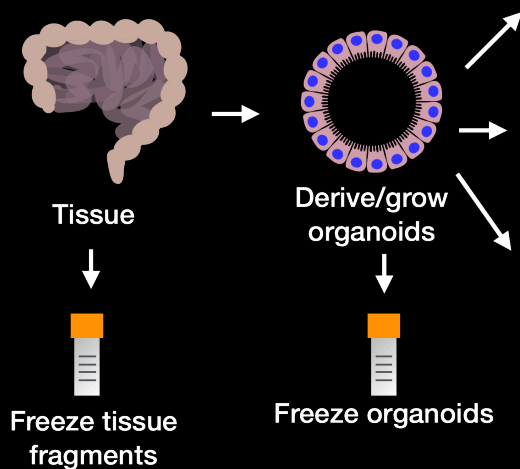
Organoid differentiation in Dome and BOBA culture is comparable



Intestinal *in vitro* Models

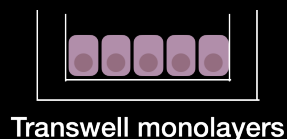
ORGANOID CULTURE

ORGANOID-DERIVED MODELS



Stem cell organoids
(crypt-like)

Differentiated organoids
(villus-like)

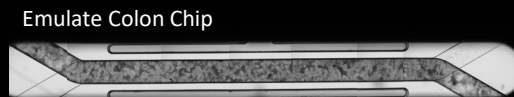
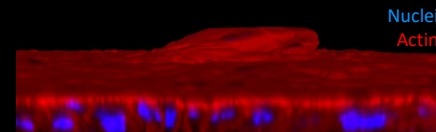


Stem cell monolayers

Differentiated monolayers



Differentiated MPS
(+/- stretch)



Next Steps for the field: Implementation of GI NAMs for Drug Development

- Confidence in the models
 - Comparative characterization of NAMs and physiologic tissue – omics and functional assays
 - Inform model selection for each application
- Confidence in the assays
 - Qualification of context-of-use assays comparing in vitro to clinical outcomes
 - Protocol standardization
- Building large datasets
 - Shared and accessible data
- Using NAMs to model human diversity
 - Accessibility to organoid cells from diverse donors



Acknowledgements

Complex in vitro Systems

Kim Homan
Jessica Klein
Serah Kang
Sarah Madira
Elias Kahn

Investigative Toxicology

Aaron Fullerton
Tomo Kiyota
Julia Heidmann
Lanlan Yu

Toxicology

Donna Lee
Catherine Ruff
Michelle Lepherd
Nina Ljumanovic

SA Pathology

Catherine Ruff
Michelle Lepherd

PD PBS

Vidhyalakshmi Arumugam
Sugantha Balasubbu
Melanie Dela Cruz
Aayushi Trivedi

Felipe de Sousa e Melo
JT Koerber
Mary Keir
Loryn Holokai

