

Cobalt-Related Exposures

The Report on Carcinogens includes two separate listings (i.e., profiles) for cobalt-related exposures: Cobalt and Cobalt Compounds That Release Cobalt Ions *In Vivo* and Cobalt-Tungsten Carbide: Powders and Hard Metals. Cobalt and cobalt compounds as a class are listed for the first time in the *Fourteenth Report on Carcinogens*, and this listing includes and supersedes the listing for cobalt sulfate, which first appeared in the *Eleventh Report on Carcinogens*. Cobalt-tungsten carbide was first listed in the *Twelfth Report on Carcinogens*. The profiles for these listings follow this introduction.

Cobalt and Cobalt Compounds That Release Cobalt Ions *In Vivo*

CAS No. 7440-48-4 (Cobalt metal)

No separate CAS No. assigned for cobalt compounds as a class
 Reasonably anticipated to be human carcinogens
 First listed in the *Fourteenth Report on Carcinogens* (2016)

Introduction

This listing of the class of cobalt and cobalt compounds that release cobalt ions *in vivo* (as defined below) supersedes the previous listing of cobalt sulfate in the Report on Carcinogens. The compound cobalt sulfate was first listed in the *Eleventh Report on Carcinogens* in 2004 as *reasonably anticipated to be a human carcinogen* based on sufficient evidence of carcinogenicity in experimental animals.

Carcinogenicity

Cobalt and cobalt compounds that release cobalt ions *in vivo* are *reasonably anticipated to be human carcinogens* based on sufficient evidence of carcinogenicity from studies in experimental animals and supporting data from studies on mechanisms of carcinogenesis. Mechanistic data indicate that the release of cobalt ions *in vivo* is a key event for cobalt-induced carcinogenicity. The available data show that cobalt metal and cobalt compounds that release cobalt ions *in vivo* (regardless of their solubility in water) act via similar modes of action to cause similar types of effects, including cell death, DNA

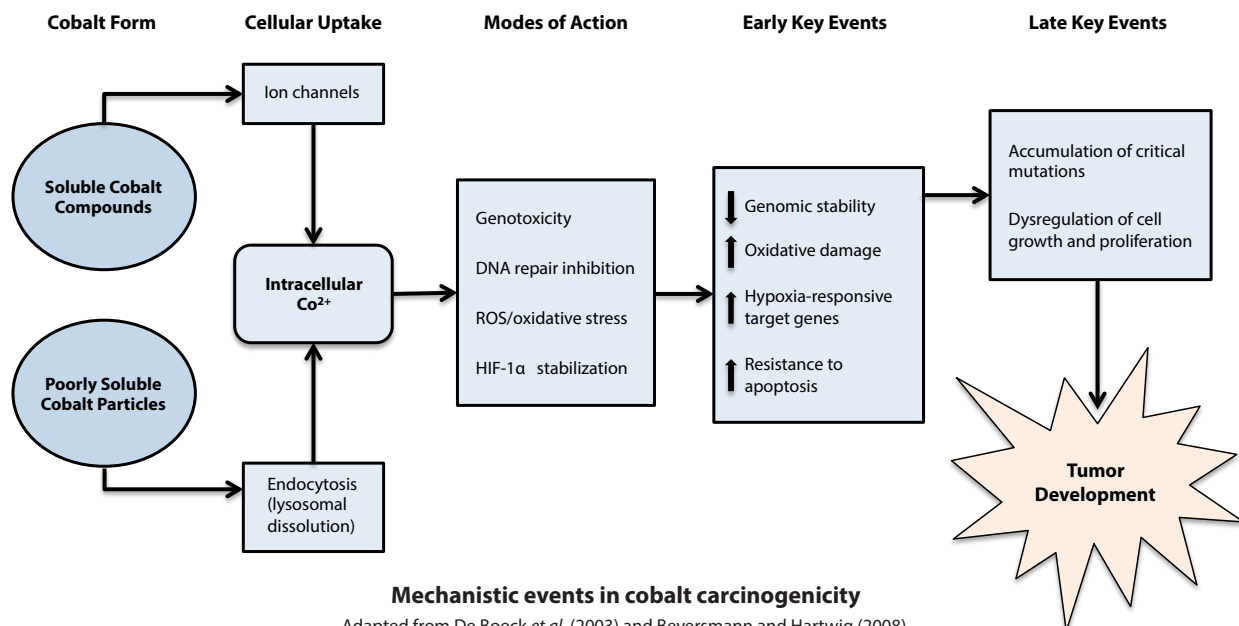
damage, and cancer, and that the cobalt ion is largely responsible for the toxicity and carcinogenicity (NTP 1998, 2014, IARC 2006).

Both water-soluble cobalt compounds and poorly water-soluble cobalt particles are included in this class, as both types of cobalt species can release cobalt ions *in vivo*, although they differ in the mechanisms by which the cobalt ions enter cells. Vitamin B₁₂, which is an essential cobalt-containing nutrient, does not meet the criteria for this listing, because the vitamin does not release cobalt ions, but passes through the body intact while bound to specific carrier proteins (Neale 1990). It is not possible to determine the quantitative carcinogenic risk from cobalt ions released from surgical implants because of limitations in the available cancer studies of cobalt alloy implants in experimental animals and of patients with cobalt-containing surgical implants.

Mechanisms of Carcinogenesis and Other Relevant Data

The key events related to toxicity and carcinogenicity are thought to include cellular uptake of cobalt, intracellular release of cobalt ions from particles, and immediate and downstream biological responses related to the proposed modes of action. The first step in the carcinogenicity or toxicity process is the release of cobalt ions *in vivo*. Water-soluble cobalt compounds release cobalt ions into fluids outside the cell, and the ions enter the cell through ion channels within the cell membrane. In contrast, poorly soluble particulate cobalt compounds are taken up by specific organelles (lysosomes) in the cell via a process called endocytosis; cobalt is then solubilized in the acidic environment in the lysosomes, and the ions are released inside the cell. Evidence for cellular uptake of the different forms of cobalt is provided by studies evaluating their solubility in biological fluids *in vitro* (e.g., in gastric and lysosomal fluids) (see Properties) and *in vitro* studies measuring levels of cobalt ions within cells (Peters *et al.* 2007, Ortega *et al.* 2014, Sabbioni *et al.* 1994, Smith *et al.* 2014).

Although the mechanism(s) of action for cobalt-induced carcinogenic effects are not completely understood, several key events have been identified that are related to biologically plausible modes of action and are applicable to all cobalt forms that release cobalt ions *in vivo*. These events include inhibition of DNA repair, genotoxicity, generation of reactive oxygen species (ROS) resulting in oxidative damage, and stabilization of hypoxia-inducible factor 1α (HIF-1α), a protein that increases the expression of genes that promote sur-



vival of cells when they receive less oxygen. The proposed modes of action are summarized in the diagram below.

Cobalt is considered to be a clastogen, because in *in vitro* assays in mammalian cells, it primarily causes chromosome damage and DNA strand breaks. Only a few genotoxicity studies in experimental animals were available, but the results were generally consistent with those of *in vitro* studies. Two potential mechanisms for genotoxicity include (1) direct induction of oxidative damage to DNA by cobalt(II) ions and (2) an indirect effect through inhibition of DNA repair (Smith *et al.* 2014, Lison 2015).

Cobalt is one of a group of metals (transition metals, like iron and nickel) that promote oxidation and reduction (redox) reactions through transfer of electrons. *In vitro* studies have shown that cobalt particles and ions can induce ROS in mammalian cells, with cobalt metal and cobalt oxide particles having a greater effect than ions. It has been proposed that ROS can play a role in the tumor development process at several stages, including initiating the process by inducing mutations and promoting proliferation of these mutated cells by deregulating controls on cell growth, leading to tumors. Studies in rats have shown that cobalt causes oxidative stress and oxidative DNA damage in several tissues, including kidney, liver, and lung (Kasprzak *et al.* 1994), which supports this proposed pathway for cobalt-induced carcinogenicity. Also, a higher frequency of a specific mutation in the *K-ras* oncogene, a gene with the potential to cause cancer, was found in cobalt-induced lung tumors in mice and rats than in spontaneous lung tumors (NTP 1998, 2014, IARC 2006). This mutation involves substitution of one nucleotide for another in a G to T transversion, which is a mutation commonly associated with oxidative DNA damage. In addition, cobalt-induced oxidative stress (via the production of ROS) can activate genes and proteins (specifically, the transcription factors NF- κ B, AP1, p53, and Nrf2) that in turn regulate the expression of many genes that play a role in carcinogenicity, such as those involved in inflammation and control of the cell cycle (Valko *et al.* 2005, 2006, Beyersmann and Hartwig 2008, Shukla *et al.* 2012, Davidson *et al.* 2015, PubChem 2015).

Finally, a well-established biological effect of cobalt is to mimic oxygen deficiency in cells by stabilizing HIF-1 α (Maxwell and Salnikow 2004, Greim *et al.* 2009, Saini *et al.* 2010a,b, Galán-Cobo *et al.* 2013, Gao *et al.* 2013, Nyga *et al.* 2015). HIF-1 α plays a central role in regulating more than 100 hypoxia-responsive genes and is a major regulator of the adaptation of cancer cells to oxygen deficiency. HIF-1 α overexpression has been linked to cancer initiation and progression and is a common characteristic of many human cancers (Paul *et al.* 2004, Galanis *et al.* 2008, 2009, Cheng *et al.* 2013).

Although most of the toxicological effects of cobalt are attributed to the cobalt ion, direct toxic effects of cobalt particles also contribute, as evidenced by the greater toxicity of cobalt metal than of cobalt sulfate in National Toxicology Program (NTP) rodent bioassays (NTP 1998, 2014, Behl *et al.* 2015). Differences in the relative toxicity reported for cobalt particles and ions may be partially explained by differences in the mechanisms by which cobalt enters the cell and in the subsequent accumulation and distribution of cobalt within the cell, as well as a synergistic effect between the particles and metal on ROS production (Peters *et al.* 2007, Sabbioni *et al.* 2014, Smith *et al.* 2014).

Cancer Studies in Experimental Animals

Exposure of experimental animals to cobalt metal or cobalt compounds caused tumors in two rodent species, at several different tissue sites, and by several different routes of exposure. This conclusion is based on studies in rats and mice exposed to cobalt metal (five studies), water-soluble cobalt compounds (two studies with cobalt sulfate and one study with cobalt chloride), and poorly water-

soluble cobalt compounds (four studies with cobalt oxide). Studies of cobalt alloys and radioactive cobalt in experimental animals were not considered to be informative, because of potential confounding by other carcinogens.

Inhalation exposure of rats and mice to cobalt metal (NTP 2014) or cobalt sulfate (NTP 1998) or intratracheal instillation of cobalt oxide in rats (Steinhoff and Mohr 1991) caused lung tumors (alveolar/bronchiolar adenoma and carcinoma). In addition, inhalation exposure of rats to cobalt metal caused squamous-cell tumors of the lung (primarily cystic keratinizing epithelioma) in females and possibly in males.

In inhalation studies of cobalt metal in rats (NTP 2014), tumors were also induced at sites distant from the lung, including tumors of the pancreas (islet-cell adenoma or carcinoma combined) in males and of the hematopoietic system (mononuclear-cell leukemia) in females, indicating a systemic effect. Increased incidences of kidney tumors (adenoma or carcinoma combined) in male rats and pancreas (carcinoma) in female rats may have been related to cobalt metal inhalation; however, the findings were not conclusive. Inhalation exposure to cobalt metal (NTP 2014) or cobalt sulfate (NTP 1998) induced adrenal-gland tumors (benign and malignant pheochromocytoma), which could have been caused by direct or indirect mechanisms.

In rats, local injection of cobalt at various anatomic locations caused tumors at the injection sites. Although these studies were less robust than the inhalation studies, and sarcomas are common in rats following injection of a variety of compounds, the consistency of the tumor types and findings across different cobalt forms provides supporting evidence for the carcinogenicity of cobalt. Intraperitoneal or intramuscular injection of the poorly water-soluble compound cobalt oxide caused histiocytoma or sarcoma at the injection site (Gilman and Ruckerbauer 1962, Steinhoff and Mohr 1991), and subcutaneous injection of the water-soluble compound cobalt chloride caused fibrosarcoma (Shabaan *et al.* 1977). Intramuscular or intrathoracic injection of cobalt metal (Heath 1956, Heath and Daniel 1962) or nanoparticles (Hansen *et al.* 2006) caused various types of sarcoma (primarily rhabdomyofibrosarcoma, rhabdomyosarcoma, or fibrosarcoma). In the study of nanoparticles, no tumors were observed after implantation of substances (e.g., titanium dioxide and silicon dioxide) with the same physical characteristics (i.e., surface-to-volume ratio) as cobalt, suggesting that the tumors were due to carcinogenic properties of cobalt and not just to a reaction to any physical implant.

A few studies in rodents (Gilman and Ruckerbauer 1962, Jasmin and Riopelle 1976, Wehner *et al.* 1977) found no tumors at certain tissue sites following exposure to the same forms of cobalt that caused tumors in other studies; however, these studies generally lacked sensitivity to detect an effect, because of the use of a less sensitive animal model, shorter study duration, or lower exposure levels.

Cancer Studies in Humans

The data available from studies in humans are inadequate to evaluate the relationship between human cancer and exposure specifically to cobalt and cobalt compounds that release cobalt ions *in vivo*. The data relevant to the evaluation were from studies primarily evaluating lung cancer in five independent cohorts of workers in different types of industries and two population-based case-control studies of esophageal cancer and other cancers of the respiratory and upper digestive (aerodigestive) tract, one in Ireland (O'Rorke *et al.* 2012) and the other in the state of Washington (Rogers *et al.* 1993). Studies of cobalt alloys in humans (primarily joint implants) were not considered to be informative, because they were not specific to cobalt exposure, and the extent of any cobalt exposure was unknown.

Although increased risks of lung cancer were found in most of the cohort studies, it is unclear that the excess risks were due to exposure specifically to cobalt, because of potential confounding from exposures to known lung carcinogens or other study limitations. In the cohort studies, hard-metal (Moulin *et al.* 1998, Wild *et al.* 2000) and nickel-refinery workers (Grimsrud *et al.* 2005) were also exposed to known lung carcinogens. The findings of an increased risk of lung cancer among porcelain painters exposed to cobalt was complicated by a somewhat similar increase in risk among female pottery workers who were not thought to be exposed to cobalt (Tüchsen *et al.* 1996). In studies of a cohort of cobalt production workers, the excess risk found in the first report of this cohort (Mur *et al.* 1987) was no longer present in an update of the cohort (Moulin *et al.* 1993). No association between cobalt exposure and lung cancer was found in a study of stainless- and alloyed-steel workers in France (Moulin *et al.* 2000). Most of the studies had limited sensitivity to detect a true risk, because of small numbers of lung-cancer cases among exposed workers, crude methods of exposure assessment, or potential healthy-worker-related effects (due to the fact that workers are healthier on average than the general population).

Increased risks of esophageal cancer were suggested in two case-control studies; however, it is unclear whether cobalt exposure contributed to the cancer excess. In both studies, cobalt exposure was assessed from a single sample of toenail clippings taken at or several months after diagnosis of esophageal cancer. Measurements of cobalt in toenails reflect an integrated exposure that occurred 12 to 18 months before clipping, raising the question of whether levels found in toenails close to or, in many cases, after cancer diagnosis reflected the relevant period of exposure for long-latency cancer.

Properties

As a class, cobalt and cobalt compounds that release cobalt ions *in vivo* are related largely by their chemical properties, specifically bioavailability. (The different valence states of cobalt are described below, under Chemical Characteristics.)

Bioavailability

The carcinogenic and toxic effects of cobalt and cobalt compounds begin with the release of cobalt ions *in vivo*. The bioavailability of a metal species can be predicted by its solubility in biological fluids, such as synthetic equivalents of gastric and intestinal fluids (for ingestion exposure) or lung (alveolar, interstitial, and lysosomal) fluids (for inhalation exposure), and by studies in cultured cells. Results from studies testing solubility in synthetic biological fluids are shown in the table below, along with other chemical and physical properties of cobalt metal and these cobalt compounds. These studies demonstrated that cobalt metal and both water-soluble and poorly water-soluble cobalt compounds can dissolve and release cobalt ions in some biological fluids (Brock and Stopford 2003, Stopford *et al.* 2003, Cobalt Development Institute personal communication 21 Jul and 19 Oct 2015), suggesting that they will release ions *in vivo*.

Very low values ($\leq 2\%$) for bioaccessibility have been reported for the sulfide and mixed (II,III) oxide (Co_3O_4), and intermediate values (14% to 55%) for stearate and oxalate under the same test conditions. However, other, more informative tests with more physiologically relevant test conditions (e.g., two-week studies with 0.3- μm particles in culture medium in the presence of alveolar macrophages) have reported 50% solubility for Co_3O_4 . In addition, Ortega *et al.* (2014) found that intracellular concentrations of solubilized cobalt ions were

Physical and chemical properties of cobalt metal and some cobalt compounds

Form ^a	CAS No.	Formula	Molec. weight	Physical form	Density or specific gravity	Water solubility (g/100 cc) ^b	Bioaccessibility (% solubility in gastric/lysosomal fluids)
<i>Cobalt metal</i>	7440-48-4	Co ^c	58.9 ^c	grey hexagonal or cubic metal ^c	8.92 ^c	0.00029 ^d	100/100 ^e
Water-soluble compounds							
<i>Acetate (org.)</i>	71-48-7	Co(C ₂ H ₃ O ₂) ₂ ^f	249.1 ^f	red-violet, monoclinic ^f	1.70 ^f	34.8 ^d	98/80 ^d
<i>Chloride</i>	7646-79-9	CoCl ₂ ^g	129.8 ^g	blue hexagonal leaflets ^g	3.36 ^g	45 ^g	100/100 ^e
<i>Nitrate</i>	10141-05-6	CoN ₂ O ₆ ^c	182.9 ^c	red powder or crystals ^c	2.49 ^c	67.0 ^d	96/100 ^d
<i>Sulfate heptahydrate</i>	10026-24-1	CoSO ₄ ·7H ₂ O ^f	281.1 ^f	red pink, monoclinic ^f	1.95 ^f	60.4 ^f	100/100 ^e
Poorly water-soluble compounds							
<i>Carbonate (org.)</i>	513-79-1	CoCO ₃ ^f	118.9 ^f	red, trigonal ^f	4.13 ^f	0.00114 ^d	100/100 ^e
<i>2-Ethylhexanoate (org.)</i>	136-52-7	Co(C ₈ H ₁₅ O ₂) ₂ ^f	173.7 ^h	blue liquid (12% Co) ^f	1.01 ^f	0.630 ^d	100/100 ^e
<i>Hydroxide</i>	21041-93-0	Co(OH) ₂ ^f	93.0 ^f	rose-red, rhombic ^f	3.60 ^f	0.00032 ^f	95/98 ^d
<i>Naphthenate (org.)</i>	61789-51-3	Co(C ₁₁ H ₇ O ₂) ₂ ^c	401.3 ^c	purple liquid (6% Co) ^f	0.97 ^f	0.0293 ^d	100/100 ^e
<i>Oxalate (org.)</i>	814-89-1	CoC ₂ O ₄ ^f	147.0 ^f	white or reddish ^f	3.02 ^f	0.00322 ^d	37/55 ^d
<i>Oxide</i>	1307-96-6	CoO ^f	74.9 ^f	green-brown cubic ^f	6.45 ^f	0.00049 ^d	100/92.4 ^e
<i>(II,III) Oxide</i>	1308-06-1	Co ₃ O ₄ ^f	240.8 ^f	black, cubic ^f	6.07 ^f	0.00016 ^d	2/2 ^d (50%) ⁱ
<i>Propionate (org.)</i>	1560-69-6	Co(C ₃ H ₅ O ₂) ₂ ^c	205.1 ^c	reddish solid ^d	–	7.49 ^d	91/94 ^d
<i>Stearate (org.)</i>	1002-88-6	Co(C ₁₈ H ₃₅ O ₂) ₂ ^c	625.9 ^c	grey solid ^d	–	0.00705 ^d	14/16 ^d
<i>Sulfide</i>	1317-42-6	CoS ^f	91.0 ^f	reddish octahedral ^f	5.45 ^f	0.00038 ^f	1/1 ^d

^aCobalt compounds selected for inclusion in the table are those with toxicological data or of commercial importance. All compounds contain Co(II) except where noted. Forms in italics have been tested for carcinogenicity or genetic toxicity or have mechanistic data; org. = organic compound; all others are inorganic.

^bSolubility data were converted to grams per 100 cubic centimeters as necessary.

^cPubChem 2015, ^dCobalt Development Institute personal communication 21 Jul and 19 Oct 2015, ^eStopford *et al.* 2003, ^fCDI 2006, ^gHSDB 2012, ^hHSDB 2004.

ⁱKreyling *et al.* 1990. Bioaccessibility was assessed by release of cobalt ions into culture medium in the presence of canine alveolar macrophages after two weeks of culture.

similar for Co_3O_4 and cobalt chloride in human lung cells *in vitro*, suggesting that Co_3O_4 would release cobalt ions *in vivo*. Results with other biological fluids, such as serum and intestinal, alveolar, and interstitial fluids, indicate that the species of cobalt compound, particle size and surface area, and pH of the surrogate fluid all can affect the solubility of cobalt in biological fluids.

The solubility of cobalt compounds in water depends largely on pH, and cobalt is generally more mobile in acidic solutions than in alkaline solutions (IARC 1991, Paustenbach *et al.* 2013). Sulfates, nitrates, and chlorides of cobalt tend to be soluble in water, whereas oxides (including the mixed oxide, Co_3O_4), hydroxides, and sulfides tend to be poorly soluble or insoluble in water (Lison 2015). Organic cobalt compounds can be either soluble, as is cobalt(II) acetate, or insoluble, as are cobalt(II) carbonate and cobalt(II) oxalate (CDI 2006). In addition to low pH, solubilization of some poorly water-soluble compounds in biological fluids may be enhanced in the presence of binding proteins (IARC 2006).

Chemical Characteristics

Cobalt (Co) is a naturally occurring transition element with magnetic properties. It is the 33rd most abundant element, making up approximately 0.0025% of the weight of Earth’s crust. Cobalt is a component of more than 70 naturally occurring minerals, including arsenides, sulfides, and oxides. The only stable and naturally occurring cobalt isotope is ^{59}Co (ATSDR 2004, WHO 2006). Metallic cobalt, Co(0), exists in two crystalline forms, hexagonal and cubic, which are stable at room temperature (IARC 1991, ATSDR 2004, WHO 2006). Cobalt predominantly occurs in two oxidation states, Co(II) and Co(III). Co(II) is much more stable than Co(III) in aqueous solution (Nilsson *et al.* 1985, Paustenbach *et al.* 2013) and is present in the environment and in most commercially available cobalt compounds (e.g., cobalt chloride, sulfide, and sulfate). Co(III) also is present in some commercially available cobalt compounds, including the mixed oxide (Co_3O_4) (IARC 1991, Paustenbach *et al.* 2013, Lison 2015) and some simple salts of Co(III) (e.g., Co_2O_3). Important salts of carboxylic acids include formate, acetate, citrate, naphthenate, linoleate, oleate, oxalate, resinate, stearate, succinate, sulfamate, and 2-ethylhexanoate.

Use

Cobalt and cobalt compounds are used in numerous commercial, industrial, and military applications. On a global basis, the largest use of cobalt is in rechargeable battery electrodes (Shedd 2014b); however, U.S. production of rechargeable batteries has been very limited (Brodd 2005). In 2012, the reported U.S. consumption of cobalt and cobalt compounds was approximately 8,420 metric tons, the majority used for superalloys (Shedd 2014b). Major uses for metallic cobalt include production of superalloys, cemented carbides, and bonded diamonds. Cobalt nanoparticles are used in medical applications (e.g., sensors, magnetic resonance imaging contrast enhancement, and drug delivery), and cobalt nanofibers and nanowires are used in industrial applications. Cobalt compounds are used as pigments for glass, ceramics, and enamels (oxides, sulfate, and nitrate), as driers for paints, varnishes, or lacquers (hydroxide, oxides, propionate, acetate, tallate, naphthenate, and 2-ethylhexanoate), as catalysts (hydroxide, oxides, carbonate, nitrate, acetate, oxalate, and sulfide), as adhesives and enamel frits (naphthenate, stearate, and oxides), and as trace mineral additives in animal diets (carbonate, sulfate, nitrate, oxides, and acetate). U.S. consumption of cobalt and cobalt compounds in 2012 is summarized in the following table.

The fastest-growing use for cobalt in recent years has been in high-capacity, rechargeable batteries, including nickel-cadmium, nickel-metal hydride, and lithium-ion batteries for electric vehicles

End use	Metric tons of cobalt content	Percent of total consumption
Superalloys	4,040	48.0
Chemicals and ceramics	2,300	27.3
Cemented carbides	774	9.2
Other alloys ^a	699	8.3
Steels	548	6.5
Miscellaneous and unspecified	63	0.7

Source: Shedd 2014b.

^aIncludes magnetic, nonferrous, and wear-resistant alloys and welding materials.

and portable electronic devices such as smartphones and laptops (Maverick 2015). Many other uses for cobalt exist, including in integrated circuit contacts and semiconductor production. An emerging use is as a key element in several forms of “green” energy technology applications, including gas-to-liquids and coal-to-liquids processes, oil desulfurization, clean coal, solar panels, wind and gas turbines, and fuel cells, and in cobalt-based catalysts for sunlight-driven water-splitting to convert solar energy into electrical and chemical energy.

Production

Cobalt metal is produced as a by-product from ores associated with copper, nickel, zinc, lead, and platinum-group metals and is most often chemically combined in its ores with sulfur and arsenic (Davis 2000, CDI 2006). The largest cobalt reserves are in the Congo (Kinshasa), Australia, Cuba, Zambia, Canada, Russia, and New Caledonia, with very limited production in the United States in recent years (Shedd 2014a). Except for a negligible amount of by-product cobalt produced from mining and refining of platinum-group metal ores, the United States did not refine cobalt in 2012 (Shedd 2014b). Cobalt has not been mined in the United States in over 30 years (ATSDR 2004); however, a primary cobalt mine, mill, and refinery were being established in Idaho in 2015 (Farquharson 2015). In 2012, 2,160 metric tons of cobalt was recycled from scrap. No cobalt has been sold from the National Defense Stockpile since 2009.

Metallic cobalt and several cobalt compounds are high-production-volume chemicals, based on their annual production or importation into the United States in quantities of at least 1 million pounds. Volumes of U.S. production, imports, and exports of cobalt metal and high-production-volume cobalt compounds are listed in the following table.

Cobalt category	Quantity (lb)		
	Production (2015)	Imports (2017)	Exports (2017)
Metal (excluding alloys)	10 million to 50 million	–	–
Compounds			
Acetates	1 million to 10 million	434,399	438,720
Carbonates	1 million to 10 million	–	–
Chlorides	1 million to 10 million	79,657	12,890
2-Ethylhexanoate	1 million to 10 million	–	–
Hydroxide	1 million to 10 million	–	–
Oxides	1 million to 10 million ^a	4,663,291 ^b	441,396 ^b
Propionate	1 million to 10 million	–	–

Sources: EPA 2016 (production volume = production + imports), USITC 2018 (imports and exports).

– = no data found.

^aThe reported value is for cobalt oxide (CoO).

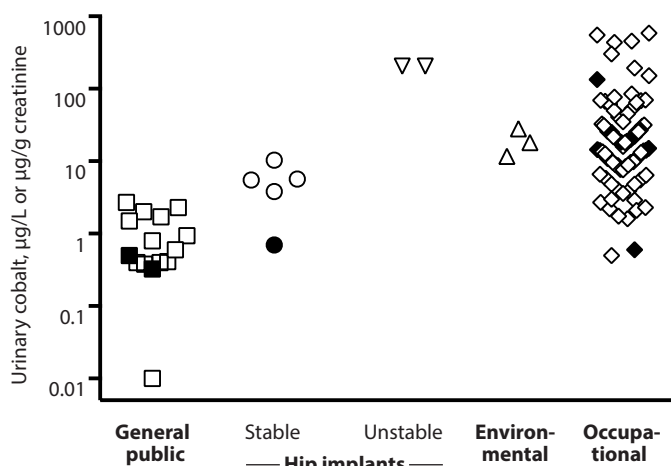
^bThe reported value is for cobalt hydroxide and oxides combined.

Exposure

A significant number of people living in the United States are exposed to cobalt, based on several lines of evidence, including biological monitoring data demonstrating exposure in occupationally and non-occupationally exposed populations. Data from the U.S. Envi-

ronmental Protection Agency’s Toxics Release Inventory (TRI) indicate that production- and use-related releases of cobalt compounds have occurred at numerous industrial facilities in the United States.

In biomonitoring studies that measured cobalt in the urine of people exposed to cobalt from various sources, the highest levels generally were due to occupational exposures and failed hip implants; lower levels were due to exposure from normal implants or the environment. The lowest levels were observed in the general population (with unknown sources of exposure). The graph below shows the mean or median levels of urinary cobalt for the general population and for groups with known exposures. Data are reported for both U.S. and non-U.S. exposures; occupational and medical implant exposures outside the United States can be informative because of the similarity of production methods and implant compositions worldwide.



Urine levels of cobalt for various exposed groups

Source: NTP 2015.
 Filled symbols = U.S. data; open symbols = non-U.S. data.
 Each data point represents a different study.

Urinary cobalt measurements in the U.S. general population were similar in 1999 and 2014, with geometric mean values between 0.316 and 0.391 µg/L, according to the National Health and Nutrition Examination Survey (CDC 2018). Urinary cobalt is considered a good indicator of absorbed cobalt (IARC 2006, WHO 2006), especially from recent exposures (ATSDR 2004). Levels of cobalt in blood (including whole blood, plasma, and serum) show a pattern similar to that for urinary cobalt levels (CDC 2018).

Occupational Exposure

The primary route of occupational exposure to cobalt is via inhalation of dust, fumes, mists, or gaseous cobalt carbonyl. Dermal contact with cemented carbide (i.e., hard-metal) powders and cobalt salts can result in systemic uptake. Occupational exposure to cobalt occurs in the following industries: (1) production of cobalt metal or salts, (2) metallurgical-related industries, (3) cemented carbides and bonded diamonds, (4) chemicals and pigments, and (5) electronics, “green” energy, and recycling. Occupational exposure has been documented by measurements of cobalt in ambient workplace air (as shown in the following table) and in blood, urine (as shown in the figure above), nails, and hair, and lung tissue from workers or deceased workers (IARC 1991, ATSDR 2004, IARC 2006, CDC 2013). The highest levels of cobalt in workplace air were generally for hard-metal manufacture involving cobalt metal powders (> 1,000 µg/m³ in some instances) (NTP 2009), production of cobalt salts, and metallurgical-related industries (> 10,000 µg/m³ in some instances) (IARC 2006). The high-

est cobalt levels in urine, blood, hair, and nails also were associated with exposure to cobalt powders.

Industry	Cobalt in workplace air (range, µg/m ³)
Production of cobalt metal or salts	2–50,000
Metallurgical-related industries ^a	ND–21,000 ^b
Cemented carbides and bonded diamonds ^a	ND–1,622
Chemicals and pigments ^a	ND–80
Electronics, “green” energy, and recycling ^a	ND–10

Sources: IARC 2006, NIOSH 2015. ND = not detected.
^aThe range for cobalt in workplace air includes U.S. data from NIOSH Hazard Evaluation and Technical Assistance surveys.
^bOne higher value was reported; however, the Occupational Safety and Health Administration noted that the sample appeared to have been tampered with.

Surgical Implants

Total hip implants consist of (1) a femoral head attached to a stem that is inserted in the thigh bone (usually made of ceramic or metal) and (2) a socket or cup that is anchored in the pelvis (made of metal, ceramic, or polyethylene). Cobalt-chromium-molybdenum (CoCrMo) alloy is the predominant alloy used in metal-containing implants, such as metal-on-metal implants (in which both articulating surfaces are metal), polyethylene-on-metal implants, and metal-on-ceramic implants. Other metals, such as nickel, tungsten, iron, aluminum, and titanium, may also be used in implants. Knee implants may also contain cobalt metal; however, unlike some hip implants with metal-to-metal contact, knee implants are designed so that metal surfaces do not contact each other. Cobalt ions may be released into the body throughout the lifetime of a cobalt-containing device (Sampson and Hart 2012, Devlin *et al.* 2013). Urinary levels of cobalt identified from studies of hip implants reported as stable or that did not specifically address stability ranged from approximately 0.7 to 12 µg/L, compared with a range of 0.01 to 4.2 µg/L for the general population (as shown in the previous graph). Implants may fail because of excessive wear or corrosion by body fluids, increasing the levels of cobalt released from the implants (Sampson and Hart 2012). Dunstan *et al.* (2005) reported blood cobalt levels of 19 and 52 µg/L in two individuals with unstable (radiologically loose) metal-on-metal implants. In rare cases, high levels of cobalt from failed implants may be associated with toxicity. Recommended levels of blood cobalt for further clinical investigation and action were set at 7 µg/L in the United Kingdom (MHRA 2012) and 10 µg/L in the United States by the Mayo Clinic (2015).

Environmental Exposure

The TRI reported that in 2013, on- and off-site industrial releases of cobalt and cobalt compounds totaled approximately 5.5 million pounds from 723 facilities in the United States (TRI 2014a). Calculations based on media-specific release data from the TRI indicate that releases to land accounted for 82% of total releases in 2013 (TRI 2014b,c). Worldwide, approximately 75,000 metric tons of cobalt enters the environment annually, with similar amounts coming from natural sources (40,000 metric tons) and sources related to human activities (35,000 metric tons) (Shedd 1993, CDI 2006). Recycling of electronic and electrical waste can result in release of cobalt to the environment; however, releases from this source are less of a concern in the United States than in other global regions where recycling is more common and less controlled (Julander *et al.* 2014).

The average concentration of cobalt in ambient air in the United States has been reported to be approximately 0.4 ng/m³ (ATSDR 2004). Levels can be orders of magnitude higher near source areas (e.g., near facilities processing cobalt-containing alloys and compounds) reported from outside the United States. The median co-

balt concentration in U.S. drinking water has been reported to be less than 2.0 µg/L; however, levels as high as 107 µg/L have been reported (ATSDR 2004). Cobalt concentrations have been reported to range from 0.01 to 4 µg/L in seawater and from 0.1 to 10 µg/L in fresh water and groundwater (IARC 2006). Studies have reported cobalt soil concentrations ranging from 0.1 to 50 ppm. However, soils near ore deposits, phosphate rock, or ore-smelting facilities or soils contaminated by airport or highway traffic or near other source areas may contain higher concentrations (IARC 2006).

Data for individuals exposed to cobalt from the environment are limited, but a study of metal exposure from mining and processing of nonferrous metals in Katanga, Democratic Republic of Congo, found that geometric mean urinary cobalt concentrations were 4.5-fold higher for adults and 6.6-fold higher for children in urban and rural communities near mines and metal smelters than in rural communities without mining or industrial activities (Cheyns *et al.* 2014).

Other Sources of Exposure of the General Population

The general population can be exposed to low levels of cobalt primarily through consumption of food and to a lesser degree through inhalation of ambient air and ingestion of drinking water (ATSDR 2004). The daily cobalt intake from food in the United States was estimated to range from 3.4 to 11.6 µg based on analyses of 234 foods in the 1984 U.S. Food and Drug Administration Total Diet Study (Pennington and Jones 1987). Although this amount includes cobalt as part of both vitamin B₁₂ and other cobalt compounds (ATSDR 2004), green, leafy vegetables and fresh cereals generally contain the most cobalt (IARC 1991), and these plant sources of cobalt do not contain vitamin B₁₂. In the 1960s, some breweries added cobalt salts to beer to stabilize the foam (resulting in cobalt exposures of 0.04 to 0.14 mg/kg of body weight), but cobalt is no longer added to beer (ATSDR 2004). Higher cobalt intake may result from consumption of over-the-counter or prescription mineral preparations containing cobalt compounds.

Other potential sources of exposure include consumer products and tobacco smoking. Cobalt is present in only a few consumer products, including cleaners, detergents, soaps, car waxes, and a nickel metal hydride battery (5% to 10% cobalt) (ATSDR 2004, HPD 2014). Various brands of tobacco have been reported to contain cobalt at concentrations ranging from less than 0.3 to 2.3 µg/g of dry weight, and 0.5% of the cobalt content is transferred to mainstream smoke (WHO 2006). However, urinary cobalt levels (unadjusted for creatinine) for cigarette-smoke-exposed and unexposed NHANES participants for survey years 1999 to 2004 did not differ significantly (Richter *et al.* 2009).

Regulations

Coast Guard (Dept. of Homeland Security)

Minimum requirements have been established for safe transport of cobalt naphthenate in solvent naphtha on ships and barges.

Department of Transportation (DOT)

Numerous cobalt compounds are considered hazardous materials, and special requirements have been set for marking, labeling, and transporting these materials.

Environmental Protection Agency (EPA)

Clean Air Act

National Emission Standards for Hazardous Air Pollutants: Cobalt compounds are listed as hazardous air pollutants.

Clean Water Act

Cobalt discharge limits are imposed for numerous processes during the production of cobalt at secondary cobalt facilities processing tungsten carbide scrap raw materials. Discharge limits for cobalt are imposed for numerous processes during the production of cobalt at primary cobalt facilities; for numerous processes during the production of batteries; and for numerous processes during the production of cobalt salts.

Discharge limits for cobalt are imposed for wastewater discharges from centralized waste treatment facilities except discharges and activities exempted in 40 CFR 437.1(b), (c), and 40 CFR 421, Subpart AC.

Cobaltous bromide, formate, and sulfamate are designated as hazardous substances.

Comprehensive Environmental Response, Compensation, and Liability Act

Reportable quantity (RQ) = 1,000 lb for cobaltous bromide, formate, and sulfamate.

Emergency Planning and Community Right-To-Know Act

EPCRA Section 302: Threshold planning quantity (TPQ) = 100 lb for cobalt, ((2,2'-(1,2-ethanediyldibis (nitrilomethylidene))bis(6-fluorophenolato))(2-)-N,N',O,O')- (also called fluomine) (solids in powder form with particle size < 100 µm or solution or molten form); = 10,000 lb for all other forms of fluomine; = 10 lb for cobalt carbonyl (solids in powder form with particle size < 100 µm or solution or molten form); = 10,000 lb for all other forms of cobalt carbonyl.

EPCRA Section 304: Reportable quantity (RQ) = 100 lb for fluomine; = 10 lb for cobalt carbonyl.

Toxics Release Inventory: Cobalt and cobalt compounds are listed substances subject to reporting requirements.

Federal Insecticide, Fungicide, and Rodenticide Act

Boiled linseed oil (containing no more than 0.33% manganese naphthenate and no more than 0.33% cobalt naphthenate) is exempt from the requirement of a tolerance when used as a coating agent for 5-ethyl hexahydro-1H-azepine-1-carbothioate. No more than 15% of the pesticide formulation may consist of boiled linseed oil, and this exemption is limited to use on rice before edible parts form.

Food and Drug Administration (FDA, an HHS agency)

Cobaltous salts are prohibited from use in human food.

All drugs containing cobalt salts (except radioactive forms of cobalt and its salts and cobalamin and its derivatives) have been withdrawn from the market because they were found to be unsafe or not effective, and they may not be compounded.

Chromium–cobalt–aluminum oxide used as a color additive for linear polyethylene surgical sutures used in general surgery must comprise no more than 2% by weight of the suture material, not migrate to surrounding tissue, and conform to labeling requirements in 21 CFR 70.25.

Chromium cobalt–aluminum oxide may be used as a color additive in contact lenses in amounts not to exceed the minimum reasonably required to accomplish the intended coloring effect.

Ferric ammonium ferrocyanide and ferric ferrocyanide used to color externally applied drugs (including those for use in the area of the eye) must not contain more than 200 ppm cobalt (as Co) and conform to labeling requirements in 21 CFR 70.25.

21 CFR 369 contains recommended drug labeling statements for over-the-counter cobalt preparations containing ≥ 0.5 mg cobalt as a cobalt salt per dosage unit and which recommend administration rates of ≥ 0.5 mg per dose and ≥ 2 mg per 24-hour period.

An approved new drug application is required for marketing cobalt preparations intended for use by man.

21 CFR 872, 874, and 888 identify class designations (Class I, II, or III) of various cobalt-containing dental prosthetic device alloys, cobalt–chromium–alloy–based facial prosthetics, and cobalt–chromium–molybdenum orthopedic devices that determine the type of premarketing submission or application required for FDA clearance to market.

Cobalt naphthenate may be used in quantities that do not exceed those reasonably required as an accelerator in the production of cross-linked polyester resins used as articles or components of articles intended for repeated use in contact with food.

Cobalt aluminate may be safely used as a colorant in the manufacture of articles or components of articles intended for use in producing, manufacturing, packing, processing, preparing, treating, packaging, transporting, or holding of food at levels not to exceed 5% by weight of all polymers except in resinous and polymeric coatings complying with 21 CFR 175.300, melamine-formaldehyde resins in molded articles complying with 21 CFR 177.1460, xylene-formaldehyde resins complying with 21 CFR 175.380, ethylene-vinyl acetate copolymers complying with 21 CFR 177.1350, and urea-formaldehyde resins in molded articles complying with 21 CFR 177.1900.

Occupational Safety and Health Administration (OSHA, Dept. of Labor)

This legally enforceable PEL was adopted from the 1968 ACGIH TLV-TWA shortly after OSHA was established; it may not reflect the most recent scientific evidence and may not adequately protect worker health.

Permissible exposure limit (PEL) (8-h TWA) = 0.1 mg/m³ for cobalt metal, dust, and fume (as Co).

Guidelines

American Conference of Governmental Industrial Hygienists (ACGIH)

Threshold limit value – time-weighted average (TLV-TWA) = 0.02 mg/m³ for cobalt and inorganic compounds; = 0.1 mg/m³ for cobalt carbonyl and cobalt hydrocarbonyl.

Biological exposure index (BEI) = 15 µg/L for cobalt in urine for cobalt and inorganic compounds, including cobalt oxides but not combined with tungsten carbide, for end of shift at end of workweek.

Consumer Product Safety Commission (CPSC)

The CPSC has issued guidance regarding the potential hazards of specific cobalt- or cobalt-compound-containing art and craft materials (e.g., glazes, glass colorants, paints, toners, pigments, and dyes) and specific precautions to take when using them.

Environmental Protection Agency (EPA)

Regional Screening Levels (formerly Preliminary Remediation Goals): residential soil = 23 mg/kg; industrial soil = 350 mg/kg; residential air = 0.00031 µg/m³; industrial air = 0.0014 µg/m³; tap water = 6 µg/L.

National Institute for Occupational Safety and Health (NIOSH, CDC, HHS)

Recommended exposure limit (REL) (10-h TWA) = 0.05 mg/m³ for cemented tungsten carbide containing > 2% Co (as Co); = 0.05 mg/m³ for cobalt metal dust and fume (as Co); = 0.1 mg/m³ for cobalt carbonyl (as Co) and cobalt hydrocarbonyl (as Co).

Immediately dangerous to life and health (IDLH) limit = 20 mg/m³ for cobalt metal dust and fume (as Co).

References

- ATSDR. 2004. *Toxicological Profile for Cobalt*. Atlanta, GA: Agency for Toxic Substances and Disease Registry. pp. 207-E203. <http://www.atsdr.cdc.gov/ToxProfiles/TP.asp?id=373&tid=64>.
- Behl M, Stout MD, Herbert RA, Dill JA, Baker GL, Hayden BK, Roycroft JR, Bucher JR, Hooth MJ. 2015. Comparative toxicity and carcinogenicity of soluble and insoluble cobalt compounds. *Toxicology* 333: 195-205.
- Beyersmann D, Hartwig A. 2008. Carcinogenic metal compounds: Recent insight into molecular and cellular mechanisms. *Arch Toxicol* 82(8): 493-512.
- Brock T, Stopford W. 2003. Bioaccessibility of metals in human health risk assessment: evaluating risk from exposure to cobalt compounds. *J Environ Monit* 5(4): 71N-76N.
- Brodd RJ. 2005. *Factors Affecting U.S. Production Decisions: Why Are There No Volume Lithium-Ion Battery Manufacturers in the United States?* ATP Working Paper 05-01, prepared for the National Institute of Standards and Technology Advanced Technology Program. <http://www.atp.nist.gov/eao/wp05-01/wp05-01.pdf>. 92 pp.
- CDC. 2013. *Biomonitoring Summary: Cobalt*. Centers for Disease Control and Prevention. Last updated: 12/4/13. http://www.cdc.gov/biomonitoring/Cobalt_BiomonitoringSummary.html.
- CDC. 2018. Cobalt. In *Fourth National Report on Human Exposure to Environmental Chemicals*, Updated Tables, March 2018, vol. 1. Atlanta, GA: U.S. Department of Health and Human Services, Centers for Disease Control and Prevention. pp. 299-305.
- CDI. 2006. Cobalt in chemicals. In *Cobalt Facts*. Cobalt Development Institute. http://theccdi.com/cdi/images/documents/facts/COBAL_FACTS-Chemicals%202015.pdf.
- Cheng Y, Chen G, Hong L, Zhou L, Hu M, Li B, Huang J, Xia L, Li C. 2013. How does hypoxia inducible factor-1α participate in enhancing the glycolysis activity in cervical cancer? *Ann Diagn Pathol* 17(3): 305-311.
- Cheyns K, Banza Lubaba Nkulu C, Ngombe LK, Asosa JN, Haufroid V, De Putter T, et al. 2014. Pathways of human exposure to cobalt in Katanga, a mining area of the D.R. Congo. *Sci Total Environ* 490: 313-321.
- Davidson T, Ke Q, Costa M. 2015. Selected molecular mechanisms of metal toxicity and carcinogenicity. In *Handbook on the Toxicology of Metals*, 4th ed., vol. 1. Nordberg GF, Fowler BA, Nordberg M, eds. Waltham, MA: Elsevier. pp. 173-196.
- Davis JR, ed. 2000. The cobalt industry: Occurrence, recovery, and consumption. In *Nickel, Cobalt, and Their Alloys*. Materials Park, OH: ASM International. pp. 345-348.
- De Boeck M, Kirsch-Volders M, Lison D. 2003b. Cobalt and antimony: Genotoxicity and carcinogenicity. *Mutat Res* 533(1-2): 135-152.
- Devlin JJ, Pomerleau AC, Brent J, Morgan BW, Deitchman S, Schwartz M. 2013. Clinical features, testing, and management of patients with suspected prosthetic hip-associated cobalt toxicity: A systematic review of cases. *J Med Toxicol* 9(4): 405-415.
- Dunstan E, Sanghrajka AP, Tilley S, Unwin P, Blunn G, Cannon SR, Briggs TW. 2005. Metal ion levels after metal-on-metal proximal femoral replacements: A 30-year follow-up. *J Bone Joint Surg Br* 87(5): 628-631.
- EPA. 2014. *Non-confidential 2012 Chemical Data Reporting (CDR) Database*. U.S. Environmental Protection Agency. Last updated: 6/14. http://java.epa.gov/oppt_chemical_search and select CDR tab and search on CAS number.
- EPA. 2016. *Chemical Data Reporting Summary: Cobalt*. U.S. Environmental Protection Agency. <https://chemview.epa.gov/chemview> and search on CAS number or substance name and select Manufacturing, Processing, Use, and Release Data Maintained by EPA and select Chemical Data Reporting Details.
- Farquharson JP. 2015. *Formation Metals Inc. - President's Letter to Shareholders*. Formation Metals, Inc. <http://www.formationmetals.com/s/CobaltNews.asp?ReportID=697830>.
- Galán-Cobo A, Sánchez-Silva R, Serna A, Abreu-Rodríguez I, Muñoz-Cabello AM, Echevarría M. 2013. Cellular overexpression of Aquaporins slows down the natural HIF-2α degradation during prolonged hypoxia. *Gene* 522(1): 18-26.
- Galanis A, Pappa A, Giannakakis A, Lanitis E, Dangaj D, Sandaltzopoulos R. 2008. Reactive oxygen species and HIF-1 signalling in cancer. *Cancer Lett* 266(1): 12-20.
- Galanis A, Karapetsas A, Sandaltzopoulos R. 2009. Metal-induced carcinogenesis, oxidative stress and hypoxia signalling. *Mutat Res* 674(1-2): 31-35.
- Gao S, Zhou J, Zhao Y, Toselli P, Li W. 2013. Hypoxia-response element (HRE)-directed transcriptional regulation of the rat lysyl oxidase gene in response to cobalt and cadmium. *Toxicol Sci* 132(2): 379-389.
- Gilman JP, Ruckerbauer GM. 1962. Metal carcinogenesis. I. Observations on the carcinogenicity of a refinery dust, cobalt oxide, and colloidal thorium dioxide. *Cancer Res* 22: 152-157.
- Greim H, Hartwig A, Reuter U, Richter-Reichhelm HB, Thielmann HW. 2009. Chemically induced pheochromocytomas in rats: Mechanisms and relevance for human risk assessment. *Crit Rev Toxicol* 39(8): 695-718.
- Grimsrud TK, Berge SR, Haldorsen T, Andersen A. 2005. Can lung cancer risk among nickel refinery workers be explained by occupational exposures other than nickel? *Epidemiology* 16(2): 146-154.
- Hansen T, Clermont G, Alves A, Eloy R, Brochhausen C, Boutrand JP, Gatti AM, Kirkpatrick CJ. 2006. Biological tolerance of different materials in bulk and nanoparticulate form in a rat model: Sarcoma development by nanoparticles. *J R Soc Interface* 3(11): 767-775.
- Heath JC. 1956. The production of malignant tumours by cobalt in the rat. *Br J Cancer* 10(4): 668-673.
- Heath JC, Daniel MR. 1962. The production of malignant tumours by cobalt in the rat: Intrathoracic tumours. *Br J Cancer* 16(3): 473-478.
- HPD. 2014. *Household Products Database*. National Library of Medicine. Last updated: 8/14. <http://householdproducts.nlm.nih.gov/advancedsearch.htm> and select Ingredient and search on CAS number.
- HSDB. 2004. *Hazardous Substances Database. Cobalt Bis(2-Ethylhexanoate)*. National Library of Medicine. Last updated: 3/5/04. <http://toxnet.nlm.nih.gov/cgi-bin/sis/htmlgen?HSDB> and search on CAS number.
- HSDB. 2012. *Hazardous Substances Database. Cobaltous Chloride*. National Library of Medicine. Last updated: 3/23/12. <http://toxnet.nlm.nih.gov/cgi-bin/sis/htmlgen?HSDB> and search on CAS number.
- IARC. 1991. Cobalt and cobalt compounds. In *Chlorinated Drinking-water; Chlorination By-products; Some Other Halogenated Compounds; Cobalt and Cobalt Compounds*. IARC Monographs on the Evaluation of Carcinogenic Risks to Humans, vol. 52. Lyon, France: International Agency for Research on Cancer. pp. 363-434.
- IARC. 2006. Metallic cobalt particles (with or without tungsten carbide). In *Cobalt in Hard-metals and Cobalt Sulfate, Gallium Arsenide, Indium Phosphide and Vanadium Pentoxide*, IARC Monographs on the Evaluation of Carcinogenic Risk of Chemicals to Humans, vol. 86, Lyon, France: International Agency for Research on Cancer. pp. 39-155.
- Jasmin G, Riopelle JL. 1976. Renal carcinomas and erythrocytosis in rats following intrarenal injection of nickel subsulfide. *Lab Invest* 35(1): 71-78.
- Julander A, Lundgren L, Skare L, Grandér M, Palm B, Vahter M, Lidén L. 2014. Formal recycling of e-waste leads to increased exposure to toxic metals: An occupational exposure study from Sweden. *Environ Int* 73: 243-251.
- Kasprzak KS, Zastawny TH, North SL, Riggs CW, Diwan BA, Rice JM, Dizdaroğlu M. 1994. Oxidative DNA base damage in renal, hepatic, and pulmonary chromatin of rats after intraperitoneal injection of cobalt(II) acetate. *Chem Res Toxicol* 7(3): 329-335.
- Kreyling WG, Godleski JJ, Kariya ST, Rose RM, Brain JD. 1990. *In vitro* dissolution of uniform cobalt oxide particles by human and canine alveolar macrophages. *Am J Respir Cell Mol Biol* 2(5): 413-422.
- Lison D. 2015. Cobalt. In *Handbook on the Toxicology of Metals*, 4th ed., vol. II. Nordberg GF, Fowler BA, Nordberg M, eds. Waltham, MA: Elsevier. pp. 743-763.
- Maverick T. 2015. Cobalt shortage put brakes on electric car. *Wall Street Daily*. <http://www.wallstreetdaily.com/2015/01/13/cobalt-electric-car-battery>.
- Maxwell P, Salnikow K. 2004. HIF-1: An oxygen and metal responsive transcription factor. *Cancer Biol Ther* 3(1): 29-35.
- Mayo Clinic. 2015. *Test ID: COS. Cobalt, serum*. Mayo Medical Laboratories. <http://www.mayomedicallaboratories.com/test-catalog/Clinical+and+Interpretive/80084>. Last accessed: 4/10/15.
- MHRA. 2012. *Medical Device Alert: All Metal-on-Metal (MoM) Hip Replacements*. MDA/2012/036. Medicines and Healthcare Products Regulatory Agency, National Joint Registry of England, Wales, and Northern Ireland. <https://assets.digital.cabinet-office.gov.uk/media/5485abf6ed915d4c10000273/con155767.pdf>.
- Moulin JJ, Wild P, Mur JM, Fournier-Betz M, Mercier-Gallay M. 1993. A mortality study of cobalt production workers: an extension of the follow-up. *Am J Ind Med* 23(2): 281-288.
- Moulin JJ, Wild P, Romazini S, Lasfargues G, Peltier A, Bozec C, Deguerry P, Pellet F, Perdrix A. 1998. Lung cancer risk in hard-metal workers. *Am J Epidemiol* 148(3): 241-248.
- Moulin JJ, Clavel T, Roy D, Dananché B, Marquis N, Févotte J, Fontana JM. 2000. Risk of lung cancer in workers producing stainless steel and metallic alloys. *Int Arch Occup Environ Health* 73(3): 171-180.
- Mur JM, Moulin JJ, Charruyer-Seinerra MP, Lafitte J. 1987. A cohort mortality study among cobalt and sodium workers in an electrochemical plant. *Am J Ind Med* 11(1): 75-81.
- Neale G. 1990. B12 binding proteins. *Gut* 31(1): 59-63.
- Nilsson K, Jensen BS, Carlsen L. 1985. The migration chemistry of cobalt. *Eur Appl Res Rept - Nucl Sci Technol* 7(1): 23-86.
- NIOSH. 2015. *NIOSH Health Hazard Evaluations*. https://java.epa.gov/oppt_chemical_search and search on cobalt. Last accessed: 7/17/15.
- NTP. 1998. *Toxicology and Carcinogenesis Studies of Cobalt Sulfate Heptahydrate in F344/N Rats and B6C3F1 Mice (Inhalation Studies)*. NTP Technical Report Series No. 471. Research Triangle Park, NC: National Toxicology Program. 268 pp.
- NTP. 2009. Section 2: Human exposure. In *Report on Carcinogens Background Document for Cobalt-Tungsten Carbide Powders and Hard Metals*. National Toxicology Program. http://ntp.niehs.nih.gov/ntp/roc/twelfth/2010/finalbds/hardmetalsbd20100408_508.pdf. pp. 7-37.
- NTP. 2014. *Toxicology Studies of Cobalt Metal (CAS No. 7440-48-4) in F344/N Rats and B6C3F1/N Mice and Toxicology and Carcinogenesis Studies of Cobalt Metal in F344/N Tac Rats and B6C3F1/N Mice (Inhalation Studies)*. Technical Report Series No. 581. NIH Publication No. 14-5932. Research Triangle Park, NC: National Toxicology Program. 308 pp.
- NTP. 2015. Appendix B.2: Exposure. In *Report on Carcinogens Monograph on Cobalt and Cobalt Compounds That Release Cobalt Ions In Vivo*. Research Triangle Park, NC: National Toxicology Program. http://ntp.niehs.nih.gov/ntp/about_ntp/monoperrvw/2015/july/cobalt_finalmonograph_508.pdf. pp. B-2-B-15.

- Nyga A, Hart A, Tetley TD. 2015. Importance of the HIF pathway in cobalt nanoparticle-induced cytotoxicity and inflammation in human macrophages. *Nanotoxicology* 9(7): 905-917.
- O'Rorke MA, Cantwell MM, Abnet CC, Brockman AJ, Murray LJ, Group FS. 2012. Toenail trace element status and risk of Barrett's oesophagus and oesophageal adenocarcinoma: Results from the FINBAR study. *Int J Cancer* 131(8): 1882-1891.
- Ortega R, Bresson C, Darolles C, Gautier C, Roudeau S, Perrin L, et al. 2014. Low-solubility particles and a Trojan-horse type mechanism of toxicity: The case of cobalt oxide on human lung cells. *Part Fibre Toxicol* 11:14. 18 pp.
- Paul SAM, Simons JW, Mabeesh NJ. 2004. HIF at the crossroads between ischemia and carcinogenesis. *J Cell Physiol* 200(1): 20-30.
- Paustenbach DJ, Tvermoes BE, Unice KM, Finley BL, Kerger BD. 2013. A review of the health hazards posed by cobalt. *Crit Rev Toxicol* 43(4): 316-362.
- Pennington JAT, Jones JW. 1987. Molybdenum, nickel, cobalt, vanadium, and strontium in total diets. *J Am Diet Assoc* 87(12): 1644-1650.
- Peters K, Unger RE, Gatti AM, Sabbioni E, Tsaryk R, Kirkpatrick CJ. 2007. Metallic nanoparticles exhibit paradoxical effects on oxidative stress and pro-inflammatory response in endothelial cells *in vitro*. *Int J Immunopathol Pharmacol* 20(4): 679-689.
- PubChem. 2015. *PubChem Compound*. National Center for Biotechnology Information. <http://www.ncbi.nlm.nih.gov/pccompound> and search on CAS number. Last accessed: 3/24/15.
- Richter PA, Bishop EE, Wang J, Swahn MH. 2009. Tobacco smoke exposure and levels of urinary metals in the U.S. youth and adult population: The National Health and Nutrition Examination Survey (NHANES) 1999-2004. *Int J Environ Res Public Health* 6(7): 1930-1946.
- Rogers MA, Thomas DB, Davis S, Vaughan TL, Nevissi AE. 1993. A case-control study of element levels and cancer of the upper aerodigestive tract. *Cancer Epidemiol Biomarkers Prev* 2(4): 305-312.
- Sabbioni E, Minoia C, Pietra R, Mosconi G, Forni A, Scansetti G. 1994a. Metal determinations in biological specimens of diseased and non-diseased hard metal workers. *Sci Total Environ* 150(1-3): 41-54.
- Sabbioni E, Fortaner S, Farina M, Del Torchio R, Petrarca C, Bernardini G, et al. 2014. Interaction with culture medium components, cellular uptake and intracellular distribution of cobalt nanoparticles, microparticles and ions in Balb/3T3 mouse fibroblasts. *Nanotoxicology* 8(1): 88-99.
- Saini Y, Greenwood KK, Merrill C, Kim KY, Patial S, Parameswaran N, Harkema JR, LaPres JJ. 2010a. Acute cobalt-induced lung injury and the role of hypoxia-inducible factor 1 α in modulating inflammation. *Toxicol Sci* 116(2): 673-681.
- Saini Y, Kim KY, Lewandowski R, Bramble LA, Harkema JR, LaPres JJ. 2010b. Role of hypoxia-inducible factor 1 α in modulating cobalt-induced lung inflammation. *Am J Physiol Lung Cell Mol Physiol* 298(2): L139-147.
- Sampson B, Hart A. 2012. Clinical usefulness of blood metal measurements to assess the failure of metal-on-metal hip implants. *Ann Clin Biochem* 49(Pt 2): 118-131.
- Shabaan AA, Marks V, Lancaster MC, Dufeu GN. 1977. Fibrosarcomas induced by cobalt chloride (CoCl₂) in rats. *Lab Anim* 11(1): 43-46.
- Shedd KB. 1993. *The Materials Flow of Cobalt in the United States*. Bureau of Mines Information Circular 9350. United States Department of the Interior. <http://pubs.usgs.gov/usbmic/ic-9350/ic-9350.pdf>.
- Shedd KB. 2014a. Cobalt. In *Mineral Commodity Summaries 2014*. U.S. Geological Survey. <http://minerals.usgs.gov/minerals/pubs/commodity/cobalt/mcs-2014-cobal.pdf>.
- Shedd KB. 2014b. *USGS 2012 Minerals Yearbook: Cobalt [Advance Release]*. <http://minerals.usgs.gov/minerals/pubs/commodity/cobalt/myb1-2012-cobal.pdf>.
- Shukla SJ, Huang R, Simmons SO, Tice RR, Witt KL, Vanleer D, Ramabhadran R, Austin CP, Xia M. 2012. Profiling environmental chemicals for activity in the antioxidant response element signaling pathway using a high throughput screening approach. *Environ Health Perspect* 120(8): 1150-1156.
- Smith LJ, Holmes AL, Kandpal SK, Mason MD, Zheng T, Wise JP, Sr. 2014. The cytotoxicity and genotoxicity of soluble and particulate cobalt in human lung fibroblast cells. *Toxicol Appl Pharmacol* 278(3): 259-265.
- Steinhoff D, Mohr U. 1991. On the question of a carcinogenic action of cobalt-containing compounds. *Exp Pathol* 41(4): 169-174.
- Stopford W, Turner J, Cappellini D, Brock T. 2003. Bioaccessibility testing of cobalt compounds. *J Environ Monit* 5(4): 675-680.
- TRI. 2014a. *TRI Explorer Chemical Report. TRI On-Site and Off-site Reported Disposed of or Otherwise Released (in pounds) for Cobalt and Cobalt Compounds*, U.S. U.S. Environmental Protection Agency. <http://www.epa.gov/triexplorer>. Last accessed: 10/14/14.
- TRI. 2014b. *TRI Explorer Chemical Report. TRI On-Site and Off-Site Reported Disposed of or Otherwise Released (in pounds), Trend Report for Facilities in All Industries for Cobalt Chemical, U.S.* U.S. Environmental Protection Agency. <http://www.epa.gov/triexplorer>. Last accessed: 10/14/14.
- TRI. 2014c. *TRI Explorer Chemical Report. TRI On-site and Off-site Reported Disposed of or Otherwise Released (in pounds), Trend Report for Facilities in All Industries, for Cobalt Compounds Chemical, U.S.* U.S. Environmental Protection Agency. <http://www.epa.gov/triexplorer>. Last accessed: 10/13/14.
- Tüchsen F, Jensen MV, Villadsen E, Lyng E. 1996. Incidence of lung cancer among cobalt-exposed women. *Scand J Work Environ Health* 22(6): 444-450.
- USITC. 2018. *USITC Interactive Tariff and Trade DataWeb*. United States International Trade Commission. http://dataweb.usitc.gov/scripts/user_set.asp and search on HTS no. 2915293000, 2827396000, and 2822000000.
- Valko M, Morris H, Cronin MT. 2005. Metals, toxicity and oxidative stress. *Curr Med Chem* 12(10): 1161-1208.
- Valko M, Rhodes CJ, Moncol J, Izakovic M, Mazur M. 2006. Free radicals, metals and antioxidants in oxidative stress-induced cancer. *Chem Biol Interact* 160(1): 1-40.
- Wehner AP, Busch RH, Olson RJ, Craig DK. 1977. Chronic inhalation of cobalt oxide and cigarette smoke by hamsters. *Am Ind Hyg Assoc J* 38(7): 338-346.
- WHO. 2006. *Cobalt and Inorganic Cobalt Compounds*. Concise International Chemical Assessment Document 69. International Programme on Chemical Safety. <http://www.inchem.org/documents/cicads/cicads/cicad69.htm>.
- Wild P, Perdrix A, Romazini S, Moulin JJ, Pellet F. 2000. Lung cancer mortality in a site producing hard metals. *Occup Environ Med* 57(8): 568-573.

Cobalt–Tungsten Carbide: Powders and Hard Metals

CAS No.: none assigned

Reasonably anticipated to be a human carcinogen

First listed in the *Twelfth Report on Carcinogens* (2011)

Also known as Co/WC, WC/Co

Carcinogenicity

Cobalt–tungsten carbide powders and hard metals are *reasonably anticipated to be human carcinogens* based on limited evidence of carcinogenicity from studies in humans and supporting evidence from studies on mechanisms of carcinogenesis.

Cancer Studies in Humans

Epidemiological studies provide evidence for the carcinogenicity of cobalt–tungsten carbide powders and hard metals based on (1) consistent findings of excess lung-cancer mortality among cobalt–tungsten carbide hard-metal manufacturing workers across studies, (2) higher risks among individuals with higher exposure levels, and (3) positive exposure-response relationships that cannot be explained by confounding with tobacco smoking. However, the epidemiological data are limited, because there are few studies of independent populations.

The published epidemiological literature consists of mortality studies of two independent multi-plant cohorts of cobalt–tungsten carbide hard-metal manufacturing workers, one in France (Moulin *et al.* 1998) and one in Sweden (Hogstedt and Alexandersson 1990), and cohort studies of two individual factories included in the French multi-plant cohort (Lasfargues *et al.* 1994, Wild *et al.* 2000). The French multi-plant cohort included all 10 cobalt–tungsten carbide manufacturing plants in France; in addition, a nested case-control study of lung cancer was conducted within this cohort. The nested case-control study is most informative for evaluating cancer risk, because it used a semi-quantitative exposure scale to evaluate exposure-response relationships and considered potential confounding by exposure to tobacco smoking and other known or suspected occupational carcinogens. The cohort study of the largest French factory shares these advantages; however, because the workers were included in the multi-plant study, it does not provide independent evidence for carcinogenicity. In these two studies, four metrics of exposure were evaluated: (1) exposure level, which was the highest exposure score experienced during an individual's work history (on a scale of 0 to 9), (2) duration of exposure at a level of 2 or higher, (3) unweighted cumulative dose, which assigned the same level to occasional and full-time exposure, thus favoring peak exposure, and (4) frequency-weighted cumulative dose, which weighted exposure level by the frequency of exposure, thus reducing the effect of occasional exposure. The Swedish study, although limited in size, provides supporting information for an independent population.

Excess lung-cancer mortality (of approximately 30%) was found in both multi-plant cohort studies (Hogstedt and Alexandersson 1990, Moulin *et al.* 1998); risk estimates were significantly higher among individuals with higher measures of exposure or longer time since first exposure (latency). In the nested case-control study (Moulin *et al.* 1998), lung cancer risk was significantly higher (odds ratio [OR] = 1.93, 95% CI = 1.03 to 3.62, 35 exposed cases) among workers exposed to cobalt–tungsten carbide (exposure level ≥ 2) than among workers with little or no exposure (exposure level < 2). In exposure-response analyses using workers in the lowest exposure category as the comparison group, lung-cancer risk was significantly higher (by up to fourfold) for workers in the highest categories of both measures of cumulative dose, and an elevated risk of borderline statistical significance was found for workers in the highest exposure-level category. Positive exposure-response relationships were observed for all four measures of exposure: duration ($P_{\text{trend}} = 0.03$), unweighted cumulative dose ($P_{\text{trend}} = 0.01$), frequency-weighted cumulative dose ($P_{\text{trend}} = 0.08$), and exposure level ($P_{\text{trend}} = 0.08$). Adjustment for tobacco smoking or exposure to known or suspected carcinogens did not change the results. The Swedish study had limited ability to evaluate exposure-response relationships because of small numbers of exposed workers with lung cancer. Nevertheless, the risk of lung cancer mortality was significantly increased for workers with exposure duration of over 10 years and latency of over 20 years (standardized mortality ratio [SMR] = 2.78, 95% CI = 1.11 to 5.72, 7 exposed cases). Analyses restricted to workers with at least 10 years' exposure or at least 20 years' latency found somewhat higher SMRs for "high-exposed" than "low-exposed" workers (Hogstedt and Alexandersson 1990).

Excess risks of lung-cancer mortality were also found in studies of the two individual French factories. Wild *et al.* (2000) reported significantly elevated SMRs (by approximately twofold) for lung cancer among all male workers and among male workers ever employed in presintering workshops or with exposure levels of at least 2. The highest SMRs were observed for male workers in the highest exposure categories of all four exposure metrics (level, duration, and both measures of cumulative dose), although the trends were not statistically significant, and the risk estimates were imprecise. In the study by Lasfargues *et al.* (1994), the entire cohort had a significantly increased risk of lung cancer, and the risk was highest among workers in the highest exposure-level category. Although small, this study provides supporting evidence that the findings for the French industry-wide cohort were not due solely to the results for the large factory studied by Wild *et al.*

Both the French multi-plant cohort study (Moulin *et al.* 1988) and the larger study of an individual French factory (Wild *et al.* 2000) found higher risks of lung cancer for exposure to cobalt–tungsten carbide before sintering than after sintering (see Production). The authors stated that exposure was highest during presintering processes; however, there is no evidence of toxicological differences between presintered and sintered materials, and both materials release similar amounts of cobalt ions (see Studies on Mechanisms of Carcinogenesis).

It is unlikely that the excess risks of lung cancer found in the French studies were due to confounding by tobacco smoking or co-exposure to other known carcinogens. In the multi-plant study, the smoking-adjusted odds ratio for cobalt–tungsten carbide exposure (OR = 2.6, 95% CI = 1.16 to 5.82) was similar to the unadjusted risk (OR = 2.29, 95% CI = 1.08 to 4.88). Neither study found increased risks of smoking-related diseases, such as chronic bronchitis and emphysema, and adjustment for smoking or exposure to other occupational carcinogens did not change the findings in the exposure-response analyses (Moulin *et al.* 1988, Wild *et al.* 2000). Neither the

Swedish multi-plant study (Hogstedt and Alexandersson 1990) nor the small French cohort study (Lasfargues *et al.* 1994) adjusted for smoking; however, surveys of smoking habits among a subset of workers found smoking rates similar to those in the general population. Overall, the studies are limited by the lack of quantitative exposure assessment and potential confounding; however, exposure misclassification would most likely reduce the likelihood of detecting a true effect.

Studies on Mechanisms of Carcinogenesis

The findings from epidemiological studies are supported by studies on mechanisms of carcinogenesis. Although the mechanism(s) by which cobalt–tungsten carbide causes cancer have not been fully elucidated, it has been shown that (1) cobalt–tungsten carbide releases cobalt ions, (2) cobalt ions affect biochemical pathways related to carcinogenicity, (3) cobalt compounds are carcinogenic in experimental animals, (4) cobalt–tungsten carbide increases the production of reactive oxygen species (ROS) and causes greater cytotoxic, toxic, and genotoxic effects than does cobalt alone, (5) cobalt–tungsten carbide causes key events related to carcinogenesis, including genotoxicity, cytotoxicity, inflammation, and apoptosis (programmed cell death), and (6) the oxidative stress response resulting from increased ROS production may play a role in these key events and may also interfere with cells' ability to repair damage caused by cobalt–tungsten carbide. The combination of the effects from cobalt ions and the oxidative stress response from ROS production provide plausible modes of action for the carcinogenicity of cobalt–tungsten carbide.

Studies in biological fluids, *in vitro* systems, experimental animals, and humans have demonstrated that cobalt is rapidly solubilized from cobalt–tungsten carbide. Cobalt dissolution rates were similar for presintered and sintered cobalt–tungsten carbide incubated in various artificial biological fluids (Stopford *et al.* 2003). Tungsten is not rapidly solubilized from cobalt–tungsten carbide, but can be phagocytized by macrophages (Lombaert *et al.* 2004). Cobalt was also released from hard-metal dust incubated with plasma and lung tissue (Edel *et al.* 1990). In experimental animals administered cobalt–tungsten carbide by intratracheal administration, cobalt was solubilized rapidly, cleared from the lung, distributed in the body, and excreted in the urine (Lison 1996). Rats exposed intratracheally to cobalt–tungsten carbide had more cobalt in the urine than did rats administered cobalt alone, suggesting that tungsten carbide increases the bioavailability of cobalt (Lasfargues *et al.* 1992). Several biomonitoring studies detected elevated levels of cobalt in the urine, lungs, and other tissues of workers exposed to cobalt–tungsten carbide hard metals (Rizzato *et al.* 1986, Nicolaou *et al.* 1987, Gallorini *et al.* 1994, Sabbioni *et al.* 1994b, Scansetti *et al.* 1994, 1998, Linnainmaa and Kiilunen 1997, Goldoni *et al.* 2004).

Soluble cobalt compounds are genotoxic and carcinogenic in experimental animals. Cobalt and cobalt compounds that release cobalt ions *in vivo* are listed as *reasonably anticipated to be human carcinogens* in the Report on Carcinogens based on sufficient evidence of carcinogenicity from studies of cobalt metal, cobalt sulfate, cobalt chloride, and cobalt oxide in experimental animals and supporting evidence from studies on mechanisms of carcinogenesis. Cobalt ions produce ROS, which cause oxidative DNA damage and act on a number of cancer-related molecular targets. Cobalt ions disrupt cell-signaling pathways (Murata *et al.* 1999), inhibit DNA repair (Hartwig 2000, Hartwig *et al.* 2002), regulate genes involved in the response to hypoxia (Beyersmann 2002), replace or mimic essential divalent metal ions, thus altering cellular reactions (Nackerdien *et al.* 1991, Beyersmann and Hartwig 1992, Kawanishi *et al.* 1994, Lloyd *et al.* 1998), and interfere with mechanisms involved in cell-cycle control and modulation of apoptosis (DeBoeck *et al.* 2003b,c).

Numerous *in vitro* studies (reviewed in NTP 2009) and *in vivo* studies (Huaux *et al.* 1995, Lasfargues *et al.* 1995) have shown greater cytotoxic effects (measured primarily by lactate dehydrogenase release) for cobalt–tungsten carbide than for either cobalt powder or tungsten carbide alone. The mixture's greater *in vitro* toxicity to macrophages is not fully explained by greater bioavailability of cobalt (Lison and Lauwerys 1992, 1994). Respirable samples collected at various stages of the hard-metal manufacturing process (including powders for pressing, presintered materials, and powders from grinding of sintered materials) caused cytotoxicity and pathological changes in the lungs of rats after intratracheal injection (Adamis *et al.* 1997). In addition, cobalt–tungsten carbide causes a type of respiratory toxicity (“hard-metal disease”) that is not observed with exposure to cobalt alone. Hard-metal disease is characterized by a giant-cell interstitial pneumonia that can develop into lung fibrosis (Lison 1996, Lison *et al.* 1996).

There is some evidence that the greater toxicity of cobalt–tungsten carbide may result from a physicochemical reaction that takes place at the interface between certain carbides and cobalt particles (Lison and Lauwerys 1992). The structural features of the two particles may help to explain the effects. Cobalt metal can reduce ambient oxygen, but only at a low rate of reaction, because of the particles' surface characteristics. Tungsten carbide is inert and does not react with oxygen but is a good electron conductor. When cobalt and tungsten carbide particles are associated, the cobalt electrons are transferred to the carbide surface, allowing increased oxygen reduction and thus increased production of ROS. Biochemical studies on the production of ROS have shown that cobalt's capacity to generate hydroxyl radicals is greatly increased by association with tungsten carbide. Formation of the ROS results directly from the interaction of cobalt with tungsten carbide or indirectly from the cobalt ions generated from the Fenton-like reaction of the cobalt metal with the carbide (Lison and Lauwerys 1993, Lison *et al.* 1995). In oxygen-radical-generating systems, post-sintered powders sampled from final machining (grinding) of cobalt–tungsten carbide products produced higher levels of ROS than did pre-sintered samples of cobalt and tungsten carbide separately or as mixtures (Stefaniak *et al.* 2010).

Metal-induced generation of ROS in cellular test systems leads to oxidative stress as a result of increased free radicals and insufficient antioxidative defense. Protective mechanisms include cellular antioxidant systems, the stress-protein response, and the involvement of DNA excision and repair enzymes (Kasten *et al.* 1997, Shi *et al.* 2004, Lombaert *et al.* 2008). Fenoglio *et al.* (2008) studied oxidation of the antioxidant glutathione and cysteine sulfhydryl groups by cobalt–tungsten carbide dust-induced ROS and reported dust-concentration-dependent generation of thyl radicals at particle surface sites. Depletion of cellular antioxidant defenses could further exacerbate cellular oxidative damage caused by ROS generated by cobalt–tungsten carbide particles.

Regulation of gene expression, including apoptotic, stress-protein, and immune-response pathways, also can be affected by ROS. Lombaert *et al.* (2008) evaluated the effects of cobalt–tungsten carbide exposure *in vitro* on patterns of gene expression in human peripheral-blood mononucleated cells and reported statistically significant up-regulation of apoptosis and stress or defense response pathways and down-regulation of immune-response pathways.

Apoptosis has been associated with exposure to a number of known carcinogens (arsenic, cadmium, chromium, nickel, and beryllium) and possible carcinogens (cobalt and lead). Cobalt chloride has been shown to induce apoptosis through formation of ROS in both human alveolar macrophages and a rat pheochromocytoma cell line (PC12); co-administration of antioxidants suppressed ROS

production and restored cell viability (Zou *et al.* 2001, Araya *et al.* 2002). Cobalt–tungsten carbide, tungsten carbide, and cobalt ions induced apoptosis in human lymphocytes; the effect of the mixture was significantly greater than that of tungsten carbide or cobalt alone (Lombaert *et al.* 2004).

Cobalt–tungsten carbide is genotoxic *in vitro* and causes mutations in the lungs of rats exposed *in vivo*. Its genotoxicity (clastogenic effects) may be caused by increased ROS production from the interaction between cobalt and tungsten carbide, from ionic cobalt, or from both. In addition, cobalt ions inhibit DNA repair, which may also contribute to cobalt–tungsten carbide's genotoxic effects. Specifically, cobalt–tungsten carbide caused DNA strand breaks in mouse 3T3 fibroblasts and human peripheral-blood lymphocytes (Anard *et al.* 1997) and micronucleus formation in human peripheral-blood lymphocytes (Van Goethem *et al.* 1997, De Boeck *et al.* 2003c). In these studies, cobalt–tungsten carbide was more genotoxic than cobalt alone. In rats exposed by intratracheal instillation, cobalt–tungsten carbide caused DNA damage and micronucleus formation in the lung (type II pneumocytes) (De Boeck *et al.* 2003a). No increase in DNA damage or micronucleus formation was observed in rat peripheral-blood lymphocytes; however, it is unclear whether circulating lymphocytes are a good reporter for monitoring genotoxic effects from inhaled particles. In humans, neither DNA damage nor micronucleus formation was increased in lymphocytes of cobalt–tungsten carbide hard-metal workers, compared with unexposed workers; however, this study was limited by small sample size (De Boeck *et al.* 2000). Multiple regression analyses (Mateuca *et al.* 2005) indicated that both end points were associated with an interaction between tobacco smoking and exposure, and that micronucleus formation was associated with smoking, working in a cobalt–tungsten carbide plant, and having variant forms of genes coding for DNA repair enzymes (X-ray repair cross-complementing group 3 and 8-oxoguanine DNA glycosylase).

In addition, although the pathogenesis of hard-metal disease is not fully understood, it may involve differences in the susceptibility (genetic and/or health-related) of affected individuals to the toxic effects of increased ROS production due to cobalt–tungsten carbide exposure. Further, the mechanisms for fibrosing alveolitis and lung cancer in hard-metal workers may be related, conceivably involving oxidative damage and/or inflammatory events (IARC 2006).

Cancer Studies in Experimental Animals

No studies in experimental animals were identified that evaluated the relationship between cancer and exposure specifically to cobalt–tungsten carbide powders or hard metals.

Properties

This listing includes powders and dusts (either unsintered or sintered) containing both cobalt and tungsten carbide and hard metals containing both cobalt and tungsten carbide. Powders containing both cobalt and tungsten carbide may result from combination of these materials during manufacture of hard metals, and dusts containing both materials may result from production, finishing, or maintenance (e.g., sharpening or grinding) of cobalt–tungsten carbide hard-metal products. Cobalt–tungsten carbide hard metals are composites of tungsten carbide particles (either alone or in combination with smaller amounts of other carbides) with a metallic cobalt powder as a binder, pressed into a compact, solid form at high temperatures by a process known as “sintering.” Cobalt–tungsten carbide hard metals are commonly referred to as “cemented carbides” in the United States, but the term “sintered carbide” also may be used, and some sources

refer to cobalt–tungsten carbide products simply as “tungsten carbides” (Brookes 2002).

The physical properties of cobalt–tungsten carbide hard metals vary with the relative proportions of cobalt, tungsten carbide, and other carbides, but they have common properties of extreme hardness, abrasion resistance, and toughness. Tungsten carbide is hard (able to resist cutting, abrasion, penetration, bending, and stretching) but brittle; cobalt is soft but tough (able to withstand great strain without tearing or breaking). The composition of commercial-grade cobalt–tungsten carbide hard metals can vary greatly; it generally ranges from 50% to 97% tungsten carbide (along with other metallic carbides such as titanium carbide or tantalum carbide) and from 3% to 16% cobalt, with variations in grain size and additives. The proportion of cobalt as the binding metal in the composite hard metal depends on the intended use (Azom 2002). Cobalt–tungsten carbide hard metals for various uses have Vickers hardness values (a measure of the resistance of a substance to indentation by a diamond penetrator of special profile) typically ranging from 1250 to 1900 (Brookes 1998).

The crystalline structure of cobalt–tungsten carbide includes the structures individually of cobalt, which can exist as either hexagonal or cubic crystals, and tungsten carbide, which consists primarily of W_2C , WC , and possibly other carbides (Upadhyaya 1998b). The phase diagram for the combination of cobalt and tungsten carbide is extremely complex, as tungsten can form a solid solution in cobalt, and cobalt can form carbides with carbon; the overall relationship varies with the concentrations of the major components and the temperature.

Mixtures of cobalt and tungsten carbide are more active than the individual components in adsorption of water vapor (with respect to both the amount adsorbed and the interaction energy) and in the catalytic decomposition of hydrogen peroxide (Zanetti and Fubini 1997). Physical and chemical properties of tungsten carbide and cobalt are listed in the following table.

Property	Cobalt	Tungsten carbide
Molecular or atomic weight	58.9	195.9
Density	8.92	15.6
Melting point	1,495°C	2,785°C
Boiling point	2,927°C	6,000°C
Vapor pressure	1 Pa at 1,517°C (0.0075 mmHg)	NR

Source: HSDB 2010. NR = not reported.

Use

About 70% of cobalt–tungsten carbide hard-metal production is used for cutting tools and 30% for wear-resistant materials, primarily for tools for mining and grinding operations (Santhanam 2003). Hard-metal grades for machining are assigned International Organization for Standardization (ISO) codes beginning with “P” for machining of steel, “M” for multiple purposes, including machining of steel, nickel-based superalloys, and higher-tensile-strength (ductile) cast iron, and “K” for cutting of lower-tensile strength (gray) cast iron, nonferrous metals, and nonmetallic materials.

Production

Cobalt–tungsten carbide hard metals were developed in Germany during and after World War I and marketed commercially by a German company in 1927 as Widia, which consisted of tungsten carbide with 6% cobalt as binder (Brookes 1998, Upadhyaya 1998a). Cobalt–tungsten carbide hard-metal manufacturing processes vary somewhat, but all involve production of cobalt and tungsten carbide powders, which are mixed, pressed into a compact, solid form, and sintered by heating to about 1,500°C. The manufacturing pro-

cess consists of three steps: Step 1, producing the cobalt and tungsten carbide powders; Step 2, mixing, drying, pressing, presintering, shaping the presintered hard metal, and sintering; and Step 3, finishing the sintered products, which includes grinding and sharpening.

Worldwide use of cemented carbides has increased steadily over the years, from about 10 tons in 1930 to 30,000 tons per year in the early 2000s (Azom 2002). In 2004, estimated U.S. production of hard-metal products totaled 5,527 metric tons (6,080 tons) (Hsu 2004). The U.S. Geological Survey (USGS 2008a,b) estimated that 792 metric tons (873 tons) of cobalt (9.3% of total U.S. cobalt consumption) and 6,610 metric tons (7,286 tons) of tungsten (56% of total U.S. tungsten consumption) was used in the production of cemented carbides in the United States in 2007. In 2008, 127 U.S. and Canadian companies were identified that produced or supplied cobalt–tungsten carbide and materials made from cobalt–tungsten carbide (ThomasNet 2008), and the Cemented Carbide Producers Association had 22 U.S. members and partner members (CCPA 2008). In 2017, the United States imported about 7.5 million pounds and exported about 4.4 million pounds of tungsten carbide (USITC 2018); no data specific for U.S. imports or exports of cobalt–tungsten carbide were found.

Exposure

The major source of exposure to cobalt–tungsten carbide powders and hard metals is occupational. However, people who live in the vicinity of hard-metal production or maintenance facilities could be exposed to cobalt–tungsten carbide hard-metal dusts. Although no exposure levels for the general population were found, some studies provided data on possible environmental contamination from the manufacture or maintenance of hard-metal products. Soil sampled from the rear of a cemented carbide tool-grinding plant contained cobalt at concentrations of up to 12,780 mg/kg (Abraham and Hunt 1995). The concentrations of tungsten and cobalt in airborne particulates in Fallon, Nevada, and four nearby towns were characterized by Sheppard *et al.* (2006), who found higher levels of tungsten (0.1 to 40.9 ng/m³) and cobalt (0.02 to 0.16 ng/m³) in Fallon than in the other towns. The authors suggested that a hard-metal facility located in Fallon could be a candidate source for airborne exposure to the metals, a suggestion that has been disputed (see NTP 2009).

Sources of occupational exposure to cobalt–tungsten carbide during the manufacture of hard metals include the processes of mixing, drying, pressing, presintering, shaping, and sintering (parts of Step 2, as described under Production) and the processes of grinding and sharpening sintered products (parts of Step 3, as described under Production). Exposure to cobalt–tungsten carbide hard metals can also occur from other miscellaneous manufacturing operations, during processing of hard-metal scrap for recycling, and during end use and maintenance of hard-metal tools. Particle size (and hence respirable fraction), morphology, and concentrations of airborne dusts and bulk dusts were found to differ among production areas (Stefaniak *et al.* 2007). For cobalt-containing particles, the minimum mass median aerodynamic diameter (MMAD) was 6 µm (for dry grinding), and the maximum MMAD was over 18 µm (for scrap reclamation and pressing operations); the MMAD for powder mixing was around 10 µm, which is generally considered the maximum diameter for respirable particles in humans. Inhalable, thoracic, and respirable particles were found in all work areas of three facilities that together carried out the cobalt–tungsten carbide manufacturing process, with the highest levels reported for the powder-mixing area (Stefaniak *et al.* 2009). Cobalt and tungsten have been detected in workers’ urine, blood, hair, toenails, and bronchoalveolar lavage fluid, and through open lung and transbronchial biopsy (NTP 2009).

Step 2 processes, particularly powder-processing operations, generally are associated with the highest airborne exposures; several studies reported cobalt concentrations approaching or exceeding 5,000 $\mu\text{g}/\text{m}^3$ (NTP 2009). A maximum mean cobalt air concentration of 32,740 $\mu\text{g}/\text{m}^3$ (range = 44 to 438,000 $\mu\text{g}/\text{m}^3$) was reported during weighing and mixing operations in a U.S. manufacturing facility (Sprince *et al.* 1984). An Italian study reported a mean tungsten air concentration of 26 $\mu\text{g}/\text{m}^3$ (Sabbioni *et al.* 1994a), and a German study reported a maximum single measurement of 254 $\mu\text{g}/\text{m}^3$ (Kraus *et al.* 2001). Among workers involved in Step 2 manufacturing processes, cobalt was detected in the urine (at up to 2,100 $\mu\text{g}/\text{L}$), blood or serum (at up to 32 $\mu\text{g}/\text{L}$), and hair (at up to 25.8 ppm), and tungsten was detected in urine (at up to 169 $\mu\text{g}/\text{L}$).

Cobalt air concentrations reported for Step 3 processes (including tool finishing, grinding, and reconditioning operations) have generally been lower than those for Step 2, but have exceeded 1,000 $\mu\text{g}/\text{m}^3$ in some studies (NTP 2009). For Step 3 processes, a maximum mean cobalt air concentration of 1,292 $\mu\text{g}/\text{m}^3$ and a maximum single measurement of 2,440 $\mu\text{g}/\text{m}^3$ were reported, both for dry-grinding operations. For tungsten in air, a maximum mean concentration of 5,160 $\mu\text{g}/\text{m}^3$ and a maximum single measurement of 12,800 $\mu\text{g}/\text{m}^3$ were reported. Among workers involved specifically in Step 3 processes, cobalt was detected in urine (at up to 730 $\mu\text{g}/\text{L}$), blood (at up to 39 $\mu\text{g}/\text{L}$), and hair (at up to 9.11 ppm). Tungsten also was detected in urine (at up to 1,000 $\mu\text{g}/\text{L}$) and blood (at up to 60 $\mu\text{g}/\text{L}$).

A few studies reported on exposure for jobs outside of the cobalt-tungsten carbide production process. McDermott (1971) reported airborne cobalt concentrations during packing operations (10 to 250 $\mu\text{g}/\text{m}^3$), equipment cleaning (40 to 820 $\mu\text{g}/\text{m}^3$), and miscellaneous operations (10 to 6,700 $\mu\text{g}/\text{m}^3$), but the nature of these operations was not defined further. Maintenance activities (including housekeeping) were reported by Scansetti *et al.* (1985) to result in airborne cobalt concentrations exceeding 50 $\mu\text{g}/\text{m}^3$, and Kraus *et al.* (2001) reported urinary levels associated with maintenance activities ranging from 1.3 to 4.7 $\mu\text{g}/\text{L}$ for cobalt and 1.5 to 5.3 $\mu\text{g}/\text{L}$ for tungsten.

Information on exposure from the end use of hard-metal tools is limited; however, exposure appears to be minimal. Pellet *et al.* (1984) reported cobalt air concentrations of 180 to 193 $\mu\text{g}/\text{m}^3$ and a mean urinary cobalt concentration of 11.7 $\mu\text{g}/\text{L}$ associated with use of hard metal; however, no additional information was provided for these data. No other information was found that directly demonstrated exposure to cobalt-tungsten carbide powders and hard metals by end users of products containing the material. The Washington State Department of Labor, in a Hazard Alert issued in March 1995, stated that there was no evidence of substantial exposure to cobalt during the use of tools containing tungsten carbide or other hard metals (WSDLI 1995).

Several studies found significant correlations between cobalt concentrations in air and in workers' blood or urine (Ichikawa *et al.* 1985, Scansetti *et al.* 1985, Lison *et al.* 1994, Sabbioni *et al.* 1994b). Urinary cobalt levels for hard-metal workers have been reported to increase through the workday (Torra *et al.* 2005) and workweek (Lison *et al.* 1994, Scansetti *et al.* 1998, Torra *et al.* 2005). In one study, urinary cobalt concentrations were significantly higher ($P < 0.005$) at the end of a shift than at the beginning of the shift, with significant increases "day in and day out" during the workweek (Torra *et al.* 2005).

Regulations

U.S. Environmental Protection Agency (EPA)

Clean Water Act

Tungsten and cobalt discharge limits are imposed for numerous processes during the production of tungsten or cobalt at secondary tungsten and cobalt facilities processing tungsten or tungsten carbide scrap raw materials.

Discharge limits for tungsten are imposed for numerous processes during the production of tungsten at primary tungsten facilities.

Discharge limits for cobalt are imposed for numerous processes during the production of cobalt at primary cobalt facilities.

Emergency Planning and Community Right-To-Know Act

Toxics Release Inventory: Cobalt and cobalt compounds are listed substances subject to reporting requirements.

Occupational Safety and Health Administration (OSHA, Dept. of Labor)

While this section accurately identifies OSHA's legally enforceable PELs for this substance in 2018, specific PELs may not reflect the more current studies and may not adequately protect workers.

Permissible exposure limits (PEL) (8-h TWA) = 0.1 mg/m^3 for cobalt metal, dust, and fume (as Co); = 5 mg/m^3 for insoluble tungsten compounds (as W).

Short-term exposure limits (STEL) = 10 mg/m^3 for insoluble tungsten compounds (as W).

Guidelines

American Conference of Governmental Industrial Hygienists (ACGIH)

Threshold limit value – time-weighted average (TLV-TWA) = 0.02 mg/m^3 for cobalt and inorganic cobalt compounds; = 5 mg/m^3 for tungsten metal and insoluble compounds.

Threshold limit value – short-term exposure limit (TLV-STEL) = 10 mg/m^3 for tungsten metal and insoluble compounds.

Biological exposure index (BEI) (end of shift at end of workweek) = 15 $\mu\text{g}/\text{L}$ for cobalt in urine.

National Institute for Occupational Safety and Health (NIOSH, CDC, HHS)

Recommended exposure limit (REL) (10-h TWA) = 0.05 mg/m^3 for cemented tungsten carbide containing > 2% Co (as Co); = 0.05 mg/m^3 for cobalt metal dust and fume (as Co); = 5 mg/m^3 for tungsten and insoluble tungsten compounds (as W).

Immediately dangerous to life and health (IDLH) limit = 20 mg/m^3 for cobalt metal dust and fume (as Co).

Short-term exposure limit (STEL) = 10 mg/m^3 for tungsten and insoluble tungsten compounds (as W).

References

- Abraham JL, Hunt A. 1995. Environmental contamination by cobalt in the vicinity of a cemented tungsten carbide tool grinding plant. *Environ Res* 69(1): 67-74.
- Adamis Z, Tatrai E, Honma K, Karpati J, Ungvary G. 1997. A study on lung toxicity of respirable hard metal dusts in rats. *Ann Occup Hyg* 41(5): 515-526.
- Anard D, Kirsch-Volders M, Elhajouji A, Belpaeme K, Lison D. 1997. *In vitro* genotoxic effects of hard metal particles assessed by alkaline single cell gel and elution assays. *Carcinogenesis* 18(1): 177-184.
- Angerer J, Heinrich R. 1988. Chapter 20: Cobalt. In *Handbook on Toxicity of Inorganic Compounds*. Seiler HG, Sigel H, eds. New York: Marcel Dekker. pp. 251-264.
- Araya J, Maruyama M, Inoue A, Fujita T, Kawahara J, Sassa K, *et al.* 2002. Inhibition of proteasome activity is involved in cobalt-induced apoptosis of human alveolar macrophages. *Am J Physiol Lung Cell Mol Physiol* 283(4): L849-L858.
- Azom. 2002. *Tungsten Carbide — An Overview*. The A to Z of Materials. <http://www.azom.com/Details.asp?ArticleID=1203>.
- Beyersmann D, Hartwig A. 1992. The genetic toxicology of cobalt. *Toxicol Appl Pharmacol* 115(1): 137-145.
- Beyersmann D. 2002. Effects of carcinogenic metals on gene expression. *Toxicol Lett* 127(1-3): 63-68.
- Brookes K. 2002. Through the looking glass—the rather odd world of hardmetals. *Metal Powder Report* 57(5): 28-29.
- Brookes KJA. 1998. *Hardmetals and Other Hard Materials*, 3rd ed. East Barnet, Hertfordshire, England: International Carbide Data. 220 pp.
- CCPA. 2008. *Cemented Carbide Producers Association — Members*. Cemented Carbide Producers Association. <http://www.ccpa.org/pages/members.html>. Last accessed: 10/6/08.
- De Boeck M, Lardau S, Buchet JP, Kirsch-Volders M, Lison D. 2000. Absence of significant genotoxicity in lymphocytes and urine from workers exposed to moderate levels of cobalt-containing dust: a cross-sectional study. *Environ Mol Mutagen* 36(2): 151-160.
- De Boeck M, Hoet P, Lombaert N, Nemery B, Kirsch-Volders M, Lison D. 2003a. *In vivo* genotoxicity of hard metal dust: induction of micronuclei in rat type II epithelial lung cells. *Carcinogenesis* 24(11): 1793-1800.
- De Boeck M, Kirsch-Volders M, Lison D. 2003b. Cobalt and antimony: Genotoxicity and carcinogenicity. *Mutat Res* 533(1-2): 135-152.
- De Boeck M, Lombaert N, De Backer S, Finsy R, Lison D, Kirsch-Volders M. 2003c. *In vitro* genotoxic effects of different combinations of cobalt and metallic carbide particles. *Mutagenesis* 18(2): 177-186.
- De Boeck M, Kirsch-Volders M, Lison D. 2004. Corrigendum to "Cobalt and antimony: genotoxicity and carcinogenicity" [Mutat Res 533 (2003) 135-152]. *Mutat Res* 548(1-2): 127-128.

- Edel J, Sabbioni E, Pietra R, Rossi A, Torre M, Rizzato G, Fraioli P. 1990. Trace metal lung disease: *In vitro* interaction of hard metals with human lung and plasma components. *Sci Total Environ* 95: 107-117.
- Fenoglio I, Corazzari I, Francia C, Bodoardo S, Fubini B. 2008. The oxidation of glutathione by cobalt/tungsten carbide contributes to hard metal-induced oxidative stress. *Free Radic Res* 42(8): 737-745.
- Gallorini M, Edel J, Pietra R, Sabbioni E, Mosconi G. 1994. Cobalt speciation in urine of hard metal workers—a study carried out by nuclear and radioanalytical techniques. *Sci Total Environ* 150(1-3): 153-160.
- Goldoni M, Catalani S, De Palma G, Manini P, Acampa O, Corradi M, Bergonzi R, Apostoli P, Mutti A. 2004. Exhaled breath condensate as a suitable matrix to assess lung dose and effects in workers. *Environ Health Perspect* 112(13): 1293-1298.
- Hartwig A. 2000. Recent advances in metal carcinogenicity. *Pure Appl Chem* 72(6): 1007-1014.
- Hartwig A, Asmuss M, Ehleben I, Herzer U, Kostelac D, Pelzer A, Schwerdtle T, Bürkle A. 2002. Interference by toxic metal ions with DNA repair processes and cell cycle control: molecular mechanisms. *Environ Health Perspect* 110(Suppl 5): 797-799.
- Hogstedt C, Alexandersson R. 1990. Dödsorsaker hos Hardmetallarbetare. *Arbete och Hälsa* 21: 1-26.
- HSDB. 2010. *Hazardous Substances Data Bank*. National Library of Medicine. <http://toxnet.nlm.nih.gov/cgi-bin/sis/htmlgen?HSDB> and search on cobalt, elemental; and search on tungsten carbide. Last accessed: 4/15/10.
- Hsu WY. 2004. Hsu WY, Kennametal, Inc., Latrobe, PA, letter to Jameson CW, National Toxicology Program, Research Triangle Park, NC, July 16, 2004.
- Huax F, Lasfargues G, Lauwerys R, Lison D. 1995. Lung toxicity of hard metal particles and production of interleukin-1, tumor necrosis factor- α , fibronectin, and cystatin-c by lung phagocytes. *Toxicol Appl Pharmacol* 132(1): 53-62.
- IARC. 2006. *Cobalt in Hard-metals and Cobalt Sulfate, Gallium Arsenide, Indium Phosphide and Vanadium Pentoxide*, IARC Monographs on the Evaluation of Carcinogenic Risk of Chemicals to Humans, vol. 86, Lyon, France: International Agency for Research on Cancer. 330 pp.
- Ichikawa Y, Kusaka Y, Goto S. 1985. Biological monitoring of cobalt exposure, based on cobalt concentrations in blood and urine. *Int Arch Occup Environ Health* 55(4): 269-276.
- Kasten U, Mullenders LH, Hartwig A. 1997. Cobalt(II) inhibits the incision and the polymerization step of nucleotide excision repair in human fibroblasts. *Mutat Res* 383(1): 81-89.
- Kawanishi S, Inoue S, Yamamoto K. 1994. Active oxygen species in DNA damage induced by carcinogenic metal compounds. *Environ Health Perspect* 102(Suppl 3): 17-20.
- Kraus T, Schramel P, Schaller KH, Zöbelein P, Weber A, Angerer J. 2001. Exposure assessment in the hard metal manufacturing industry with special regard to tungsten and its compounds. *Occup Environ Med* 58(10): 631-634.
- Lasfargues G, Wild P, Moulin JJ, Hammon B, Rosmorduc B, Rondeau du Noyer C, Lavandier M, Moline J. 1994. Lung cancer mortality in a French cohort of hard-metal workers. *Am J Ind Med* 26(5): 585-595.
- Lasfargues G, Lardot C, Delos M, Lauwerys R, Lison D. 1995. The delayed lung responses to single and repeated intratracheal administration of pure cobalt and hard metal powder in the rat. *Environ Res* 69(2): 108-121.
- Linnainmaa M, Kiilunen M. 1997. Urinary cobalt as a measure of exposure in the wet sharpening of hard metal and stellite blades. *Int Arch Occup Environ Health* 69(3): 193-200.
- Lison D. 1996. Human toxicity of cobalt-containing dust and experimental studies on the mechanism of interstitial lung disease (hard metal disease). *Crit Rev Toxicol* 26(6): 585-616.
- Lison D, Lauwerys R. 1992. Study of the mechanism responsible for the elective toxicity of tungsten carbide-cobalt powder toward macrophages. *Toxicol Lett* 60(2): 203-210.
- Lison D, Lauwerys R. 1993. Evaluation of the role of reactive oxygen species in the interactive toxicity of carbide-cobalt mixtures on macrophages in culture. *Arch Toxicol* 67(5): 347-351.
- Lison D, Lauwerys R. 1994. Cobalt bioavailability from hard metal particles: Further evidence that cobalt alone is not responsible for the toxicity of hard metal particles. *Arch Toxicol* 68(8): 528-531.
- Lison D, Buchet JP, Swennen B, Molders J, Lauwerys R. 1994. Biological monitoring of workers exposed to cobalt metal, salt, oxides, and hard metal dust. *Occup Environ Med* 51(7): 447-450.
- Lison D, Carbonnelle P, Mollo L, Lauwerys R, Fubini B. 1995. Physicochemical mechanism of the interaction between cobalt metal and carbide particles to generate toxic activated oxygen species. *Chem Res Toxicol* 8(4): 600-606.
- Lison D, Lauwerys R, Demedts M, Nemery B. 1996. Experimental research into the pathogenesis of cobalt/hard metal lung disease. *Eur Respir J* 9(5): 1024-1028.
- Lloyd DR, Carmichael PL, Phillips DH. 1998. Comparison of the formation of 8-hydroxy-2'-deoxyguanosine and single- and double-strand breaks in DNA mediated by Fenton reactions. *Chem Res Toxicol* 11(5): 420-427.
- Lombaert N, De Boeck M, Ecordier I, Undari E, Lison D, Irsch-Volders M. 2004. Evaluation of the apoptogenic potential of hard metal dust (WC-Co), tungsten carbide, and metallic cobalt. *Toxicol Lett* 154: 23-34.
- Lombaert N, Lison D, Van Hummelen P, Kirsch-Volders M. 2008. *In vitro* expression of hard metal dust (WC-Co)-responsive genes in human peripheral blood mononucleated cells. *Toxicol Appl Pharmacol* 227: 299-312.
- Mateuca R, Aka PV, De Boeck M, Hauspie R, Kirsch-Volders M, Lison D. 2005. Influence of hOGG1, XRCC1 and XRCC3 genotypes on biomarkers of genotoxicity in workers exposed to cobalt or hard metal dusts. *Toxicol Lett* 156(2): 277-288.
- McDermott FT. 1971. Dust in the cemented carbide industry. *Am Ind Hyg Assoc J* 32(3): 188-193.
- Moulin JJ, Wild P, Romazini S, Lasfargues G, Peltier A, Bozec C, Deguerry P, Pellet F, Perdrix A. 1998. Lung cancer risk in hard-metal workers. *Am J Epidemiol* 148(3): 241-248.
- Murata M, Gong P, Suzuki K, Koizumi S. 1999. Differential metal response and regulation of human heavy metal-inducible genes. *J Cell Physiol* 180(1): 105-113.
- Nackerdien Z, Kasprzak KS, Rao G, Halliwell B, Dizdaroglu M. 1991. Nickel(II)- and cobalt(II)-dependent damage by hydrogen peroxide to the DNA bases in isolated human chromatin. *Cancer Res* 51(21): 5837-5842.
- Nicolaou G, Pietra R, Sabbioni E, Mosconi G, Cassina G, Seghizzi P. 1987. Multielement determination of metals in biological specimens of hard metal workers: a study carried out by neutron activation analysis. *J Trace Elem Electrolytes Health Dis* 1(2): 73-77.
- NTP. 2009. *Report on Carcinogens Background Document for Cobalt-Tungsten Carbide Powders and Hard Metals*. National Toxicology Program. http://ntp.niehs.nih.gov/ntp/roc/twelfth/2010/finalbds/hardmetalsbd20100408_508.pdf.
- Pellet F, Perdrix A, Vincent M, Mallion JM. 1984. Biological levels of urinary cobalt. *Arch Mal Prof* 45: 81-85 (as cited in Angerer and Heinrich 1988).
- Pulido MD, Parrish AR. 2003. Metal-induced apoptosis: mechanisms. *Mutat Res* 533(1-2): 227-241.
- Rizzato G, Lo Cicero S, Barberis M, Torre M, Pietra R, Sabbioni E. 1986. Trace of metal exposure in hard metal lung disease. *Chest* 90(1): 101-106.
- Sabbioni E, Minoia C, Pietra R, Mosconi G, Forni A, Scansetti G. 1994a. Metal determinations in biological specimens of diseased and non-diseased hard metal workers. *Sci Total Environ* 150(1-3): 41-54.
- Sabbioni E, Mosconi G, Minoia C, Seghizzi P. 1994b. The European Congress on cobalt and hard metal disease. Conclusions, highlights and need of future studies. *Sci Total Environ* 150(1-3): 263-270.
- Santhanam AT. 2003. Carbides, cemented. In *Kirk-Othmer Encyclopedia of Chemical Technology*, vol. 4. Online edition. New York: John Wiley & Sons. pp. 655-674.
- Scansetti G, Lamon S, Talarico S, Botta GC, Spinelli P, Sulotto F, Fantoni F. 1985. Urinary cobalt as a measure of exposure in the hard metal industry. *Int Arch Occup Environ Health* 57(1): 19-26.
- Scansetti G, Botta GC, Spinelli P, Reviglione L, Ponzetti C. 1994. Absorption and excretion of cobalt in the hard metal industry. *Sci Total Environ* 150(1-3): 141-144.
- Scansetti G, Maina G, Botta GC, Bambace P, Spinelli P. 1998. Exposure to cobalt and nickel in the hard-metal production industry. *Int Arch Occup Environ Health* 71(1): 60-63.
- Sheppard PR, Ridenour G, Speakman RJ, Witten ML. 2006. Elevated tungsten and cobalt in airborne particulates in Fallon, Nevada: possible implications for the childhood leukemia cluster. *Appl Geochem* 21: 152-165.
- Shi H, Hudson LG, Liu KJ. 2004. Oxidative stress and apoptosis in metal ion-induced carcinogenesis. *Free Radic Biol Med* 37(5): 582-593.
- Sprince NL, Chamberlin RI, Hales CA, Weber AL, Kazemi H. 1984. Respiratory disease in tungsten carbide production workers. *Chest* 86(4): 549-557.
- Stefaniak AB, Day GA, Harvey CJ, Leonard SS, Schwegler-Berry DE, Chipera SJ, Sahakian NM, Chisholm WP. 2007. Characteristics of dusts encountered during the production of cemented tungsten carbides. *Ind Health* 45:793-803.
- Stefaniak AB, Virji MA, Day GA. 2009. Characterization of exposures among cemented tungsten carbide workers. Part I: Size-fractionated exposures to airborne cobalt and tungsten particles. *J Expo Sci Environ Epidemiol* 19(5): 475-491.
- Stefaniak AB, Harvey CJ, Bukowski VC, Leonard SS. 2010. Comparison of free radical generation by pre- and post-sintered cemented carbide particles. *J Occup Environ Hyg* 7: 23-34.
- Stopford W, Turner J, Cappellini D, Brock T. 2003. Bioaccessibility testing of cobalt compounds. *J Environ Monit* 5(4): 675-680.
- Sueker JK. 2006. Comment on "Elevated tungsten and cobalt in airborne particulates in Fallon, Nevada: Possible implications for the childhood leukemia cluster," by Sheppard PR, Ridenour G, Speakman RJ, and Witten ML. *Appl Geochem* 21: 1083-1085.
- ThomasNet. 2008. *Metals: Carbide*. Thomas Publishing. <http://www.thomasnet.com/products/tungsten-carbide-89540207-1.html>. Last accessed: 9/24/08.
- Torra M, Fernández J, Rodamilans M, Navarro AM, Corbella J. 2005. Biological monitoring of cobalt exposure: results in a non-exposed population and on workers of a hard metal manufacture. *Trace Elem Electrolytes* 22(3): 174-177.
- Upadhyaya GS. 1998a. Classification and applications of cemented carbides. In *Cemented Tungsten Carbides. Production, Properties, and Testing*. Westwood, NJ: Noyes Publications. pp. 288-293.
- Upadhyaya GS. 1998b. Crystal structure and phase equilibria. In *Cemented Tungsten Carbides. Production, Properties, and Testing*. Westwood, NJ: Noyes Publications. pp. 7-54.
- USGS. 2008a. *Mineral Industry Surveys: Cobalt in October, November and December 2007*. Reston, VA: U.S. Geological Survey.
- USGS. 2008b. *Mineral Industry Surveys: Tungsten in January 2008*. Reston, VA: U.S. Geological Survey.
- USITC. 2018. *USITC Interactive Tariff and Trade DataWeb*. United States International Trade Commission. http://dataweb.usitc.gov/scripts/user_set.asp and search on HTS no. 2849903000.
- Van Goethem F, Lison D, Kirsch-Volders M. 1997. Comparative evaluation of the *in vitro* micronucleus test and the alkaline single cell gel electrophoresis assay for the detection of DNA damaging agents: genotoxic effects of cobalt powder, tungsten carbide and cobalt-tungsten carbide. *Mutat Res* 392(1-2): 31-43.
- Wild P, Perdrix A, Romazini S, Moulin JJ, Pellet F. 2000. Lung cancer mortality in a site producing hard metals. *Occup Environ Med* 57(8): 568-573.

WSDLI. 1995. *Hard-Metal Workers Face Risks from Cobalt, Cadmium*. WISHA Hazard Alert. State of Washington Department of Labor and Industries. <http://www.lni.wa.gov/Safety/Basics/HazAlerts/951a.asp>.

Zanetti G, Fubini B. 1997. Surface interaction between metallic cobalt and tungsten carbide particles as a primary cause of hard metal lung disease. *J Mater Chem* 7(8): 1647-1654.

Zou W, Yan M, Xu W, Huo H, Sun L, Zheng Z, Liu X. 2001. Cobalt chloride induces PC12 cells apoptosis through reactive oxygen species and accompanied by AP-1 activation. *J Neurosci Res* 64(6): 646-653.