

The Updated ICCVAM Recommended Murine LLNA Test Method Protocol

J Matheson¹, M Wind¹, A Jacobs², D Allen³, T Burns³, E Salicru³, J Strickland³, R Tice⁴, and W Stokes⁵

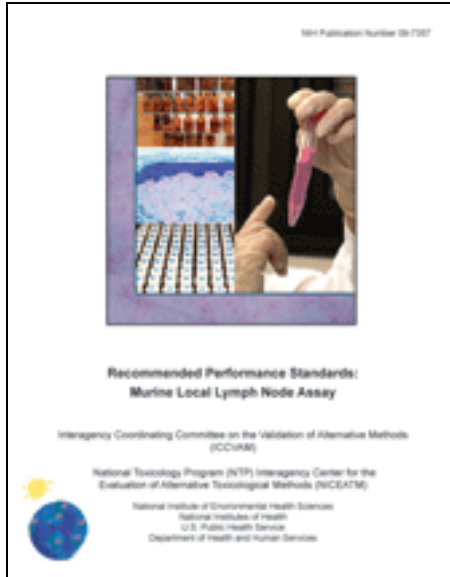
¹U.S. Consumer Product Safety Commission (CPSC), Bethesda, MD, USA; ²U.S. Food and Drug Administration (FDA), Silver Spring, MD, USA; ³ILS, Inc., Contractor Supporting the National Toxicology Program Interagency Center for the Evaluation of Alternative Toxicological Methods (NICEATM), RTP, NC, USA; ⁴National Institute of Environmental Health Science (NIEHS)/NIH/DHHS, RTP, NC, USA; ⁵NICEATM/NIEHS/NIH/DHHS, RTP, NC, USA

Introduction

United States and international regulatory authorities currently accept the murine local lymph node assay (LLNA) as an alternative to guinea pig test methods for skin sensitization testing. In March 2008, Interagency Coordinating Committee on the Validation of Alternative Toxicological Methods (ICCVAM) and the National Toxicology Program Interagency Center for the Evaluation of Alternative Toxicological Methods (NICEATM) convened an independent international peer review panel (the “Panel”) to evaluate the usefulness and limitations of new versions and applications of the LLNA. The Panel provided several recommendations that were applicable to the LLNA protocol. ICCVAM subsequently finalized an updated LLNA protocol to include:

1. Reducing the required number of animals from five to four per group
2. Rationale for collection of individual animal data
3. Guidance for use of a concurrent positive control group
4. Guidance on evaluating local irritation and systemic toxicity to establish the appropriate highest dose to test

Updated ICCVAM Recommended Test Method Protocol for the LLNA

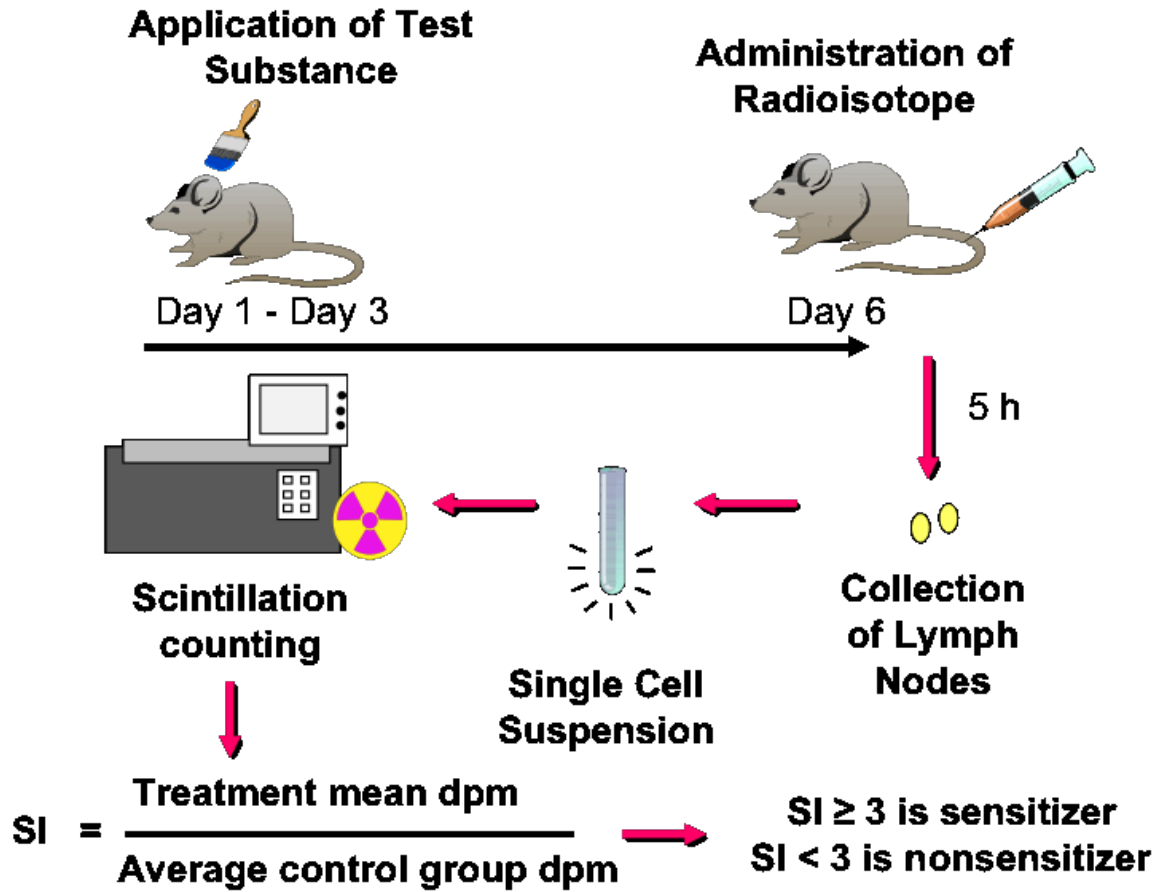


- U.S. Federal agencies are interested in a standardized protocol for skin sensitization hazard classification.
- NICEATM submitted a proposed update to Organisation for Economic Co-operation and Development (OECD) Test Guideline (TG) 429 (OECD 2002).
- The test method protocol was a key component of performance standards for the LLNA (ICCVAM 2009a)

Basic Principles of the LLNA

- Sensitizers induce proliferation of lymphocytes in the lymph nodes draining the site of substance application.
- Proliferation is a function of *in vivo* radioisotope incorporation into the DNA of dividing lymphocytes.
- Proliferation is proportional to the dose applied, and provides a means of obtaining an objective, quantitative measurement of sensitization.

Methodology of the LLNA



ICCVAM Immunotoxicity Working Group

U.S. Consumer Product Safety Commission

Joanna Matheson, Ph.D. (Co-chair)
Marilyn Wind, Ph.D.

U.S. Environmental Protection Agency

Office of Pesticide Programs

Jonathan Chen, Ph.D.
Masih Hashim, D.V.M., Ph.D.
Marianne Lewis
Deborah McCall
Timothy McMahan, Ph.D.
John Redden, M.S.
Jenny Tao, Ph.D.

Office of Prevention, Pesticides, and Toxic Substances

Elizabeth Margosches, Ph.D.
Ronald Ward, Ph.D.

Office of Research and Development

Marsha Ward, Ph.D.

Office of Science Coordination and Policy

Karen Hamernik, Ph.D.

Food and Drug Administration

Center for Devices and Radiological Health

Vasant G. Malshet, Ph.D., D.A.B.T.
Jeffrey Toy, Ph.D.

Center for Drug Evaluation and Research

Paul Brown, Ph.D.
Abigail Jacobs, Ph.D. (Co-chair)
Jiaqin Yao, Ph.D.

Center for Veterinary Medicine

Ruth Barratt, Ph.D., D.V.M.

Office of Science and Health Coordination

Suzanne Fitzpatrick, Ph.D., D.A.B.T.

National Institute of Environmental Health Sciences

Dori Germolec, Ph.D.
William Stokes, D.V.M., D.A.C.L.A.M.

National Institute for Occupational Safety and Health

B. Jean Meade, D.V.M., Ph.D.

National Library of Medicine

Pertti (Bert) Hakkinen, Ph.D.

European Centre for the Validation of Alternative Methods — Liaison

Silvia Casati, Ph.D.
Alexandre Angers, Ph.D.

Japanese Center for the Validation of Alternative Methods — Liaison

Hajime Kojima, Ph.D.

Reducing from Five to Four Animals per Group

- 83 LLNA studies (275 dose groups) from six laboratories were evaluated.
 - The average agreement between N = 4 and N = 5 was 97.5%.
 - A reduction in the sample size N = 5 to N = 4 is unlikely to have a significant impact on the results of an LLNA study (**Table 1**).

- This change is important since most animal-use regulations require that the minimum number of animals be used in studies.
- The current OECD TG 429 specifies four animals/group for pooled data and five animals/group for individual animal data
 - Only pooled data are collected in many countries because it requires fewer animals.
 - An update to TG 429 is being proposed so that only four animals are required for collection of individual animals.

Table 1. Agreement Between Sample Sizes for LLNA Outcomes of 275 Dose Groups

SI	Frequency of SI	Agreement of Outcomes ¹ (%)
< 2.1	154	100.0
2.1 – 2.5	16	90.1
2.6	2	85.0
2.7	3	73.3
2.8	2	59.5
3.1	1	56.0
3.2	2	55.5
3.3	4	73.5
3.4	1	88.0
3.5	1	68.0
3.6	1	84.0
3.7	1	90.0
3.8	1	100.0
4.0 – 4.7	16	97.9
> 4.7	70	100.0
Total	275	97.5

Abbreviation: SI = stimulation index.

¹ Proportion of samples with SI \geq 3 and SI < 3.

Rationale for Collection of Individual Animal Data

- Allows for the identification of outlier responses
 - Avoids false negative results for weaker sensitizers
- Allows for the assessment of interanimal variability
- Allows for a statistical comparison of the test substance and vehicle control group measurements
- Allows for evaluation of statistical power for different group sizes
- Allows for an evaluation of the impact of reducing the number of animals in the positive control group

Guidance for Use of a Concurrent Positive Control Group

- A concurrent positive control to ensure the appropriate performance of the assay
- Positive control dose should be reproducible but not excessive
- There may be certain regulatory situations where it is necessary to test the positive control substance in both a standard and a non-standard vehicle (e.g., a clinically/chemically relevant formulation) to test for possible interactions

Highlights from the LLNA Peer Review Panel Meeting



Panel Meeting

- A public meeting of an independent scientific peer review Panel organized by ICCVAM and NICEATM was held on March 4-6, 2008.
- The Panel report is available at http://iccvam.niehs.nih.gov/docs/immunotox_docs/LLNAPRPREpt2008.pdf

Conclusions Relevant to the LLNA Test Method Protocol

- Data should be collected at the level of individual animals to allow an estimate of the variance within control and treatment groups.
- A concurrent positive control should be run with each test substance to ensure that the system is operating as expected and technical errors are not occurring.
- If a laboratory has extensive historical data indicating that the positive control consistently yields statistically bioequivalent results in the LLNA, then, on a regular periodic basis, evaluation of a positive control could be recommended.

Independent Scientific Peer Review Panel

Michael Luster, Ph.D., (Panel Chair), Senior Consultant to the National Institute for Occupational Safety and Health, Morgantown, WV

Nathalie Alépée, Ph.D., L'Oréal Research and Development, Aulnay sous Bois, France

Anne Marie Api, Ph.D., Research Institute for Fragrance Materials, Woodcliff Lake, NJ

Nancy Flournoy, M.S., Ph.D., University of Missouri – Columbia, Columbia, MO

Thomas Gebel, Ph.D., Federal Institute for Occupational Safety & Health, Dortmund, Germany

Sidney Green, Ph.D., Howard University, Washington, DC

Kim Headrick, B.Admin., B.Sc., Health Canada, Ottawa, Ontario, Canada

Dagmar Jírová, M.D., Ph.D., National Institute of Public Health, Prague, Czech Republic

David Lovell, Ph.D., University of Surrey, Guilford, United Kingdom

Howard Maibach, M.D., University of California – San Francisco, San Francisco, CA

James McDougal, Ph.D., Wright State University, Dayton, OH

Michael Olson, Ph.D., GlaxoSmithKline, Research Triangle Park, NC

Raymond Pieters, Ph.D., Utrecht University, Utrecht, The Netherlands

Jean Regal, Ph.D., University of Minnesota Medical School, Duluth, MN

Jon Richmond, M.D., Home Office, London, United Kingdom

Peter Theran, V.M.D., Consultant, Novato, CA

Stephen Ullrich, Ph.D., M.D. Anderson Cancer Center, Houston, TX

Michael Woolhiser, Ph.D., Dow Chemical, Midland, MI

Takahiko Yoshida, M.D., Ph.D., Asahikawa Medical College, Hokkaido, Japan

Guidance on Relating Local Irritation and Systemic Toxicity with the Highest Dose to Test

- The highest LLNA dose is the maximum soluble concentration that does not induce systemic toxicity or excessive local irritation.
- In the absence of such information, a prescreen test should be performed using three dose levels of the test substance.
 - The prescreen test is conducted under identical conditions as the main LLNA study, except there is no assessment of lymph node cell proliferation.
 - Six mice (two per concentration) are used.
 - All mice are observed daily for any clinical signs of systemic toxicity or local irritation at the application site.
 - Body weights are recorded pre-test and on Day 6.
 - Both ears of each mouse are observed for erythema and scored using Table 2.
 - Ear thickness measurements are recorded on Day 1 (pre-dose), Day 3 (~48 hours after first dose), and Day 6.
 - Excessive irritation is indicated by an erythema score ≥ 3 and/or ear swelling of $\geq 25\%$ (Reeder et al. 2007; ICCVAM 2009b).

Table 2. Erythema Scores

Observation	Value
No visual effect	0
Slight erythema (barely perceptible)	1
Well-defined erythema	2
Moderate to severe erythema (beet redness)	3
Eschar (i.e., piece of dead tissue that is cast off from the surface of the skin)	4

- The following clinical observations, which are based on test guidelines and current practices (ICCVAM 2009c), may indicate systemic toxicity when used as part of an integrated assessment and therefore may indicate that the maximum dose recommended for the LLNA has been exceeded:
 - Changes in nervous system function (e.g., piloerection, ataxia, tremors, and convulsions)
 - Changes in behavior (e.g., aggressiveness, change in grooming activity, marked change in activity level)

- Changes in respiratory patterns (i.e., changes in frequency and intensity of breathing such as dyspnea, gasping, and rales)
- Changes in food and water consumption
- Lethargy and/or unresponsiveness
- Any clinical signs of more than slight or momentary pain and distress
- Reduction in body weight >10% from Day 1 to Day 6
- Mortality

International Acceptance

- A proposal to update OECD TG 429 (the LLNA) has been submitted to include recommendations described herein and is currently under review by the OECD Test Guidelines Program.
 - A minimum of four animals per test group with a collection of individual animal data rather than pooled data
 - Guidance for a prescreen test to select the highest dose for testing based on the maximum soluble concentration and the absence of systemic toxicity and/or excessive local irritation

Conclusions

- ICCVAM recommended LLNA test method protocol
 - Reduces the minimum number of animals required from five to four per group
 - Includes rationale for collecting individual animal data and using a concurrent positive control in each LLNA study
 - Provides guidance for determining the appropriate highest dose to test

References

ICCVAM. 2009a. Recommended Performance Standards: Murine Local Lymph Node Assay. NIH Publication No. 09-7357.

<http://iccvam.niehs.nih.gov/methods/immunotox/PerfStds/llna-ps.htm>

ICCVAM. 2009b. Nonradioactive Murine Local Lymph Node Assay: Flow Cytometry Test Method Protocol (LLNA: BrdU-FC) Revised Draft Background Review Document.

<http://iccvam.niehs.nih.gov/methods/immunotox/fcLLNA/BRDcomplete.pdf>

ICCVAM. 2009c. Report on the ICCVAM-NICEATM/ECVAM/JaCVAM Scientific Workshop on Acute Chemical Safety Testing: Advancing In Vitro Approaches and Humane Endpoints for Systemic Toxicity Evaluations.

http://iccvam.niehs.nih.gov/methods/acutetox/Tox_workshop.htm

OECD. 2002. Guideline for the Testing of Chemicals 429 – Skin Sensitisation: Local Lymph Node Assay. Paris: OECD. <http://www.oecd.org>

Reeder MK, Broomhead YL, DiDonato L, DeGeorge GL. 2007. *Toxicologist* 96 (S-1):235.

Acknowledgments

The Intramural Research Program of the NIH, National Institute of Environmental Health Sciences supported this poster; ILS staff supported by NIEHS contract N01-ES 35504.

ICCVAM and NICEATM gratefully acknowledge the following institutions that submitted data used to support the updates to the LLNA test method protocol:

BASF – The Chemical Company, U.S.

BAuA (The Federal Institute for Occupational Safety and Health), Germany

Dow AgroSciences, U.S.

DuPont, U.S.

ECPA (European Crop Protection Association), Belgium

EFfCI (European Federation for Cosmetic Ingredients), Belgium

This poster reflects the views of the authors and has not been reviewed or approved by the U.S. Consumer Product Safety Commission or other agencies.