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Subject: [NTP Web] NTP Interagency Center for the Evaluation of Alternative Toxicological Methods Public Comments Submission [81FR42718-42719]
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Attachments: [2016 Hunt C. elegans in Tox Testing.pdf](#)
[2015 Boyd ToxCast C. elegans vs zebrafish rats rabbits.pdf](#)
[2016 Harlow C. elegans to predict mammalian mechanism of tox.pdf](#)
[2009 Sprando rank toxicity compounds in worms.pdf](#)
[2000 Olson Tox concordance humans and animals.pdf](#)

Attention NTP Interagency Center for the Evaluation of Alternative Toxicological Methods
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Name: Piper Hunt

Telephone: [REDACTED]

Email: Piper.Hunt@fda.hhs.gov

Affiliation Type: Government

Affiliation:

Additional Contact Information:

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Comments: *Caenorhabditis elegans* is a small nematode that can be maintained at low cost and handled using standard in vitro techniques. Unlike toxicity testing using cell cultures, *C. elegans* toxicity assays provide data from a whole animal with intact and metabolically active digestive, reproductive, endocrine, sensory, and neuromuscular systems. Toxicity ranking screens in *C. elegans* have repeatedly been shown to be as predictive of rat LD50 ranking as mouse LD50 ranking [1]. Additionally, many instances of conservation of mode of toxic action have been noted between *C. elegans* and mammals. These consistent correlations make the case for inclusion of *C. elegans* assays in early safety testing and as one component in tiered or integrated toxicity testing strategies.

In a recent study of hundreds of compounds from the ToxCast™ libraries, *C. elegans* larval growth identified rat or rabbit developmental toxins with a balanced accuracy of 52-53%, while the concordance for developmental toxicity between rat and rabbit was 58% [2]. These levels of predictivity are consistent with an earlier meta-study which found that a single-species rodent study alone predicted human toxicity less than 50% of the time [3]. Unlike a study in mammals however, a *C. elegans* larval growth assay that evaluates multiple compounds or mixtures at multiple concentrations can be conducted by a single technician in less than a week.

Interestingly, the sensitivity of the aforementioned larval growth assay for the detection mammalian developmental toxins was high, but the balanced accuracy was brought down by low specificity [2]. In contrast, in a recent study evaluating *C. elegans* egg viability using 72 compounds of known developmental activity in mammals, the specificity of the egg viability test was high while the sensitivity was low [4]. Thus, it is possible that the *C. elegans* larval growth and egg viability assays could be used together to improve the detection of mammalian developmental toxins. Additionally, both of these assays used *E. coli* as a feeder organism, and neither publication discusses efforts at Good *C. elegans* Culture Practice (GCeCP) which can substantially alter test results [1]. While early *C. elegans* axenic media formulations slowed growth, newer media produce identical growth rates to cultures fed the feeder organism, without the complication of xenobiotic metabolism [5]. It may be that the use of a combination of axenic media and GCeCP could improve the predictivity of both the *C. elegans* larval growth and egg viability assays, making them a useful addition to developmental toxicant testing strategies.

1. Hunt, P.R., The *C. elegans* model in toxicity testing. *Journal of Applied Toxicology*, 2016.
2. Boyd, W.A., et al., Developmental Effects of the ToxCast Phase I and II Chemicals in and Corresponding Responses in Zebrafish, Rats, and Rabbits. *Environ Health Perspect*, 2015. 124: p. 586-593.
3. Olson, H., et al., Concordance of the toxicity of pharmaceuticals in humans and in animals. *Regul Toxicol Pharmacol*, 2000. 32(1): p. 56-67.
4. Harlow, P.H., et al., The nematode *Caenorhabditis elegans* as a tool to predict chemical activity on mammalian development and identify mechanisms influencing toxicological outcome. *Sci Rep*, 2016. 6: p. 22965.
5. Sprando, R.L., et al., A method to rank order water soluble compounds according to their toxicity using *Caenorhabditis elegans*, a Complex Object Parametric Analyzer and Sorter, and axenic liquid media. *Food Chem*

Toxicol, 2009. 47(4): p. 722-8.
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