



Centers for Disease Control  
and Prevention (CDC)  
Atlanta, GA 30341-3724

July 11, 2018

Dr. Warren Casey  
Director  
NICEATM  
[Warren.casey@nih.gov](mailto:Warren.casey@nih.gov)  
(984) 287-3118

Dear Dr. Casey,

This letter is in response to the recent request for data and information on technologies used to detect and measure botulinum neurotoxin (Fed Reg. 83(114): pages 27622-27623, June 13, 2018). The Centers for Disease Control and Prevention (CDC) is the premier public health institute in the United States and considers the accurate diagnosis of human botulism to be an important public health goal. To that end, we have developed a method to detect botulinum neurotoxin (BoNT) in clinical and selected food matrices.

The method is called the Endopep-MS method and detects the neurotoxin based on its enzymatic activity. The Endopep-MS method involves concentration and isolation of BoNT from a complex matrix using carefully selected monoclonal antibodies, followed by incubation with a substrate that is enzymatically cleaved by BoNT. The resulting substrate product is analyzed by mass spectrometry to determine the presence or absence of the BoNT serotype-specific peptide cleavage products indicating the presence of BoNT in the original test sample. Because each serotype produces peptide cleavage products of different mass, the method easily differentiates between all seven confirmed serotypes.

The method was first published in 2005, describing the ability to detect the neurotoxin and differentiate all seven serotypes in buffer [1,2]. Next, the method was adapted to detect BoNT in milk [3], serum [4], stool [4], culture supernatants [5], drinking water [6], and foods [7]. The method has been modified to increase sensitivity and specificity through substrate modification [8-13]. The method can be quantitative [14,15], but is typically run in the qualitative mode to confirm a diagnosis of botulism.

Endopep-MS has undergone extensive validation both within CDC as well as in outside laboratories [16,17]. Within CDC, the validation has consisted of a variety of tests, beginning with a spiked limit of detection study in buffer, serum, stool, and culture supernatants. The limit of detection varies according to matrix and serotype. Diagnostic sensitivity (a positive result from a true positive sample) is in the range of 0.1 mLD50 to 1 mLD50 at the 95% or above confidence interval, or below the limit of detection of the mouse bioassay. In addition to being more sensitive than the mouse bioassay, the assay has smaller sample volume requirements than the mouse bioassay. Endopep-MS has been shown to detect more than just the commercially-available subtypes [18-21], a critical factor for public health testing and part of diagnostic sensitivity that many in vitro assays ignore. To date, the Endopep-MS method has been used to successfully test all subtypes which we have been able to collect, including all of the eight BoNT/A subtypes, seven of the eight BoNT/B subtypes, six of the twelve BoNT/E subtypes, all seven of the BoNT/F subtypes, two BoNT/C subtypes, two BoNT/D subtypes, mosaics of these toxins, and BoNT/G. Endopep-MS has been used in the discovery of novel subtypes [22-28]; the method was used to discover the first new BoNT cleavage site in over 20 years [22].

Diagnostic specificity (a negative result from a true negative sample) was also part of our validation work; diagnostic specificity was found to be 100% as determined using the following samples: negative buffer (minimum of 20 samples), negative serum (minimum of 50 separate specimens), stool extract (minimum of

50 separate specimens), culture supernatants (minimum of 20 samples); and near neighbors of BoNT including tetanus toxin and culture supernatants from related non-BoNT producing *C. botulinum*.

Following validation of the Endopep-MS method, the method was certified as a CLIA diagnostic for human botulism in our laboratory and has been in use for almost a decade, testing clinical specimens (serum, stool, foods, and culture supernatants) in parallel with the mouse bioassay, the standard for BoNT detection, when sample volumes permit the use of Endopep-MS. To date, over 400 samples have been analyzed with both methods with a concordance rate of approximately 99%; a remarkable feat given the increased sensitivity of the Endopep-MS method compared to the mouse bioassay. The method has been applied successfully to a number of public health investigations of botulism [29,30] and was reported as the suggested strategy for detection of BoNT by mass spectrometry [31].

We are currently seeking FDA clearance for the Endopep-MS assay as a 510(k) De Novo In Vitro Diagnostic for human botulism. We have already submitted 2 pre-submission packets of information from our analytical study, our multi-center evaluation, and our clinical study. We are currently engaged in a multi-center validation of the assay involving state public health laboratories within our laboratory response network (LRN). To date, we have trained a number of laboratories on the Endopep-MS method, including 9 laboratories within the LRN, two FDA laboratories, and laboratories in France, Belgium, Germany, Sweden, Czech Republic, and Canada (both PHAC and Health Canada).

In short, the Endopep-MS method is a viable alternative to the mouse bioassay to diagnose human botulism. We are happy to provide additional information upon request.

#### References:

1. A.E. Boyer, H. Moura, A.R. Woolfitt, S.R. Kalb, L.G. McWilliams, A. Pavlopoulos, J.G. Schmidt, and J.R. Barr, "From the Mouse to the Mass Spectrometer: Novel Detection and Differentiation of the Endoproteinase Activities of Botulinum Neurotoxins A-G by Mass Spectrometry", *Anal. Chem.*, 2005, 77, 3916-3924.
2. J.R. Barr, H. Moura, A.E. Boyer, A.R. Woolfitt, S.R. Kalb, A. Pavlopoulos, L.G. McWilliams, J.G. Schmidt, R.A. Martinez, and D.L. Ashley, "Fast Detection and Differentiation of Botulinum Neurotoxins A,B,E, and F, Using Endopep-MS, a Mass Spectrometry-Based Method", *Emerg. Infect. Dis.*, 2005, 11(10), 1578-1583.
3. S.R. Kalb, M.C. Goodnough, C.J. Malizio, J.L. Pirkle, and J.R. Barr, "Detection of Botulinum Neurotoxin A in a Spiked Milk Sample with Subtype Identification Through Toxin Proteomics", *Anal. Chem.*, 2005, 77, 6140-6146.
4. S.R. Kalb, H. Moura, A.E. Boyer, L.G. McWilliams, J.L. Pirkle, and J.R. Barr, "The Use of Endopep-MS for the Detection of Botulinum Toxins A, B, E, and F in Serum and Stool Samples", *Anal. Biochem.*, 2006, 351, 84-92.
5. S.R. Kalb, T.J. Smith, H. Moura, K. Hill, J. Lou, C. Garcia-Rodriguez, J.D. Marks, L.A. Smith, J.L. Pirkle, and J.R. Barr, "The Use of Endopep-MS to Detect Multiple Subtypes of Botulinum Neurotoxins A, B, E, and F", *International Journal of Mass Spectrometry*, 2008, 278, 101-108.
6. B.H. Raphael, M. Lautenschlager, A. Kahler, S. Pai, B.A. Parks, S.R. Kalb, S.E. Maslanka, S. Shah, M. Magnuson, and V. Hill, "Ultrafiltration improves ELISA and Endopep MS analysis of botulinum neurotoxin type A in drinking water", *Journal of Microbiological Methods*, 2012, 90(3), 267-272.
7. S.R. Kalb, J.C. Krilich, J.K. Dykes, C. Luquez, S.E. Maslanka, and J.R. Barr, "Endopep-MS for the Detection of Botulinum Toxins A, B, E, and F in Foods", *Journal of Agricultural and Food Chemistry*, 2015, Feb 4; 63(4):1133-1141.
8. D. Wang, J. Baudys, S.R. Kalb, and J.R. Barr, "Improved Detection of Botulinum Neurotoxin Type A in Stool by Mass Spectrometry", *Analytical Biochemistry*, 2011, 412, 67-73.
9. S.R. Kalb, J. Baudys, C. Egan, T.J. Smith, L.A. Smith, J.L. Pirkle, and J.R. Barr, "Clostridium baratii Type F Neurotoxin Cleaves Synaptobrevin-2 with Different Substrate

- Recognition Requirement than Clostridium botulinum Type F Neurotoxin, Applied and Environmental Microbiology, 2011, 77, 1301-1308.
10. D. Wang, J. Baudys, Y. Ye, J.C. Rees, J.R. Barr, J.L. Pirkle, and S.R. Kalb, Improved Detection of Botulinum Neurotoxin Serotype A by Endopep-MS through Peptide Substrate Modification, Analytical Biochemistry, 2013, Jan 15; 432(2):115-23.
  11. D. Wang, J.C. Krilich, J. Baudys, J.R. Barr, and S.R. Kalb, "Optimization of peptide substrates for botulinum neurotoxin E improves detection sensitivity in the Endopep-MS assay", Analytical Biochemistry, 2014, 468C, 15-21.
  12. D. Wang, J.C. Krilich, J. Baudys, J.R. Barr, and S.R. Kalb, "Enhanced detection of type C botulinum neurotoxin by the Endopep-MS assay through optimization of peptide substrates", Bioorganic and Medicinal Chemistry, 2015, Jul 1; 23(13), 3667-73.
  13. D. Wang, J. Baudys, K.M. Hoyt, J.R. Barr, and S.R. Kalb, "Further Optimization of Peptide Substrate Enhanced Assay Performance for BoNT/A Detection by MALDI-TOF Mass Spectrometry", Analytical and Bioanalytical Chemistry, 2017, Aug; 409(20), 4779-4786.
  14. B.A. Parks, J.D. Shearer, J. Baudys, S.R. Kalb, D.C. Sanford, J.L. Pirkle, and J.R. Barr, Quantification of Botulinum Neurotoxin Serotypes A and B from Serum using Mass Spectrometry, Analytical Chemistry, 2011, 83(23), 9047-9053.
  15. D. Wang, J. Baudys, J.C. Krilich, T.J. Smith, J.R. Barr, and S.R. Kalb, "A Two-Stage Multiplex Method for Quantitative Analysis of Botulinum Neurotoxin type A, B, E, and F by MALDI-TOF Mass Spectrometry", Analytical Chemistry, 2014, 86(21), 10847-54.
  16. K. Bjornstad, A.T. Aberg, S.R. Kalb, D. Wang, J.R. Barr, U. Bondesson, and M. Hedeland, "Validation of the Endopep-MS method for qualitative detection of active botulinum neurotoxins in human and chicken serum", Analytical and Bioanalytical Chemistry, 2014, 406(28), 7149-61.
  17. M.J. Perry, D.A. Centurioni, S.W. Davis, G.E. Hannett, K.A. Musser, and C.T. Egan, "Implementing the Bruker MALDI Biotyper in the Public Health Laboratory for C. botulinum Neurotoxin Detection", Toxins, 2017, Mar 9; 9(3), E94.
  18. S.R. Kalb, T.J. Smith, H. Moura, K. Hill, J. Lou, C. Garcia-Rodriguez, J.D. Marks, L.A. Smith, J.L. Pirkle, and J.R. Barr, The Use of Endopep-MS to Detect Multiple Subtypes of Botulinum Neurotoxins A, B, E, and F, International Journal of Mass Spectrometry, 2008, 278, 101-108.
  19. S.R. Kalb, J. Lou, C. Garcia-Rodriguez, I.N. Geren, T.J. Smith, H. Moura, J.D. Marks, L.A. Smith, J.L. Pirkle, and J.R. Barr, Extraction and Inhibition of Enzymatic Activity of BoNT/A1, /A2, and /A3 by a Panel of Monoclonal Anti-BoNT/A Antibodies, PLoS One, 2009, 4(4), e5355.
  20. S.R. Kalb, C. Garcia-Rodriguez, J. Lou, J. Baudys, T.J. Smith, J.D. Marks, L.A. Smith, J.L. Pirkle, and J.R. Barr, Extraction of BoNT/A, /B, /E, and /F with a single, high affinity monoclonal antibody for detection of botulinum neurotoxin by Endopep-MS, PLoS One, 2010, 5(8), e12237.
  21. S.R. Kalb, W.I. Santana, I.N. Geren, C. Garcia-Rodriguez, J. Lou, T.J. Smith, J.D. Marks, L.A. Smith, J.L. Pirkle, and J.R. Barr, Extraction and inhibition of enzymatic activity of botulinum neurotoxins/B1, /B2, /B3, /B4, and /B5 by a panel of monoclonal anti-BoNT/B antibodies, BMC Biochemistry, 2011, 12(58), 1-12.
  22. S.R. Kalb, J. Baudys, R.P. Webb, P. Wright, T.J. Smith, L.A. Smith, R. Fernandez, B.H. Raphael, S.E. Maslanka, J.L. Pirkle, and J.R. Barr, Discovery of a Novel Enzymatic Cleavage Site for Botulinum Neurotoxin F5, FEBS Letters, 2012, 586(2), 109-115.
  23. S.R. Kalb, J. Baudys, J.C. Rees, T.J. Smith, L.A. Smith, C.H. Helma, K.K. Hill, S. Kull, S. Kirchner, M.B. Dorner, B.G. Dorner, J.L. Pirkle, and J.R. Barr, De novo Subtype and Strain Identification of Botulinum Neurotoxin Type B through Toxin Proteomics, Analytical and Bioanalytical Chemistry, 2012, 403(1), 215-226.
  24. B.H. Raphael, M. Lautenschlager, S.R. Kalb, L.I.T. de Jong, M. Frace, C. Luquez, J.R. Barr, R.A. Fernandez, and S.E. Maslanka, Analysis of a unique Clostridium botulinum strain from the Southern hemisphere producing a novel BoNT/E subtype, BMC Microbiology, 2012, Oct 31; 12(1), 245.

25. S.R. Kalb, J. Baudys, T.J. Smith, L.A. Smith, and J.R. Barr, "Three Enzymatically Active Neurotoxins of Clostridium botulinum Strain Af84: BoNT/A2, /F4, and /F5" Analytical Chemistry, 2014, 86(7), 3254-62.
26. B.H. Raphael, M. Bradshaw, S.R. Kalb, L. Joseph, C. Luquez, J.R. Barr, E.A. Johnson, and S.E. Maslanka, "Characterization of Clostridium strains producing BoNT/F4 and BoNT/F5", Applied and Environmental Microbiology, 2014, 80(10), 3250-7.
27. S. Kull, K.M. Schultz, J. Weisemann, S. Kirchner, T. Schreiber, A. Bollenbach, P.W. Dabrowski, A. Nitsche, S.R. Kalb, M.B. Dorner, J.R. Barr, A. Rummel, and B.G. Dorner, "Isolation and Functional Characterization of the Novel Clostridium botulinum Neurotoxin A8 Subtype", PLoS One, 2015, Feb 6; 10(2), e0116381.
28. S.R. Kalb, J. Baudys, B.H. Raphael, J.K. Dykes, C. Luquez, S.E. Maslanka, and J.R. Barr, "Functional Characterization of Botulinum Neurotoxin Serotype H as a Hybrid of Known Serotypes F and A (BoNT F/A)", Analytical Chemistry, 2015, Apr 7; 87(7), 3911-3917.
29. L. McCrickard, M. Marlow, J. Self, L.F. Watkins, K. Chatham-Stephens, J. Anderson, S. Hand, K. Taylor, J. Hanson, K. Patrick, C. Luquez, J. Dykes, S.R. Kalb, K. Hoyt, J.R. Barr, T. Crawford, A. Chambers, B. Douthit, R. Cox, M. Craig, J. Spurzem, J. Doherty, M. Allswede, P. Byers, and T. Dobbs, "Botulism Outbreak from Drinking Prison-Made Illicit Alcohol in a Federal Correctional Facility—Mississippi, June 2016", MMWR, 2017; 65: 1491-2.
30. B.Freund, L. Hayes, L. Rivera-Lara, C. Sumner, V. Chaudhry, K. Chatham-Stephens, K. Benedict, S. Kalb, D. Blythe, R. Brooks, and J.C. Probasco, "Adult intestinal colonization botulism mimicking brain death", Muscle and Nerve, 2017, Oct; 56(4), E27-28.
31. S.R. Kalb, J. Baudys, D. Wang, and J.R. Barr, "Recommended mass spectrometry-based strategies to identify botulinum neurotoxin-containing samples", Toxins, 2015, May 19; 7(5), 1765-1778.

Sincerely,



Dr. John R. Barr  
Chief, Clinical Chemistry Branch  
(770) 488-7848  
[jbarr@cdc.gov](mailto:jbarr@cdc.gov)



Dr. Suzanne R. Kalb  
Research Chemist  
(770) 488-7931  
[skalb@cdc.gov](mailto:skalb@cdc.gov)

National Center for Environmental Health  
Centers for Disease Control and Prevention  
4770 Buford Hwy NE  
MS-F50  
Atlanta, GA 30341