

Enabling Next Generation Pharmaceutical Safety Assessment: An Evolution

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Opinions expressed are those of the author rather than the employer

Key components

- Motive
- Opportunity
- Strategy
- Partnership

KEYFACTS 2015

RESEARCH AND DEVELOPMENT (R&D)¹

Average time to develop a drug = more than 10 years
 Percentage of drugs entering clinical trials resulting in an approved medicine = less than 12%

DEVELOPMENT COSTS

Average cost to develop a drug (including the cost of failures):²

- 2000s–early 2010s = \$2.6 billion
- 1990s–early 2000s = \$1.0 billion*
- 1980s = \$413 million
- 1970s = \$179 million



R&D SPENDING

Year	PhRMA members ³
2014	\$51.2 billion (est.)
2013	\$51.6 billion
2012	\$49.6 billion
2011	\$48.6 billion
2010	\$50.7 billion
2009	\$46.4 billion
2008	\$47.4 billion
2007	\$47.9 billion
2006	\$43.0 billion
2005	\$39.9 billion
2000	\$26.0 billion
1990	\$8.4 billion
1980	\$2.0 billion

SALES

Generic share of prescriptions filled:⁴
 2000 = 49%
 2013 = 88%



Motive

There are good reasons for us to be interested in novel ways of working!

An analysis of the attrition of drug candidates from four major pharmaceutical companies

Michael J. Waring¹, John Arrowsmith², Andrew R. Leach³, Paul D. Leeson^{3,4}, Sam Mandrell², Robert M. Owen⁵, Garry Pairaudeau¹, William D. Pennie^{6,7}, Stephen D. Pickett³, Jibo Wang⁸, Owen Wallace^{8,9} and Alex Weir²

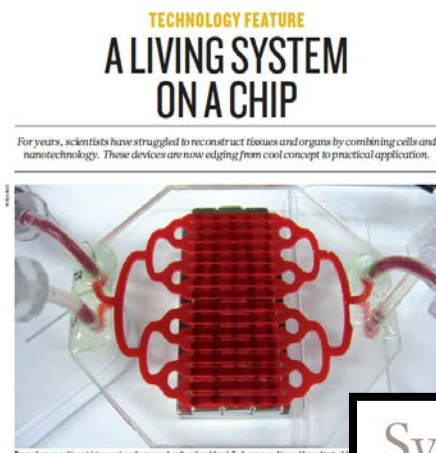
Nat Rev Drug Disc 14: 475, 2015

Table 1 | Populations of the primary cause of failure categories for terminated compounds*

Termination reason	Overall	Period		Phase		
		2000–2005	2006–2010	Candidate nomination	Phase I	Phase II
Clinical safety	68 (11%)	48 (13%)	20 (8%)	5 (1%)	40 (25%)	22 (25%)
Commercial	40 (7%)	23 (6%)	17 (7%)	26 (7%)	10 (6%)	4 (4%)
Efficacy	55 (9%)	45 (11%)	10 (4%)	10 (3%)	14 (9%)	31 (35%)
Formulation	9 (1%)	4 (1%)	5 (2%)	8 (2%)	1 (0.6%)	0
Non-clinical toxicology	240 (40%)	144 (40%)	96 (40%)	211 (59%)	21 (13%)	7 (8%)
Patent issue	1 (0.2%)	0	1 (0.4%)	1 (0.3%)	0	0
Pharmacokinetics or bioavailability	29 (5%)	19 (5%)	10 (4%)	3 (0.8%)	25 (16%)	1 (1%)
Rationalization of company portfolio	124 (21%)	46 (13%)	78 (32%)	75 (21%)	29 (18%)	19 (21%)
Regulatory	2 (0.3%)	2 (0.6%)	0	1 (0.3%)	1 (0.6%)	0
Scientific	33 (5%)	28 (8%)	5 (2%)	13 (4%)	15 (10%)	5 (6%)
Technical	3 (1%)	3 (1%)	0	2 (0.6%)	1 (0.6%)	0
Other	1 (0.2%)	0	1 (0.4%)	1 (0.3%)	0	0
Total	605	362	243	356	157	89

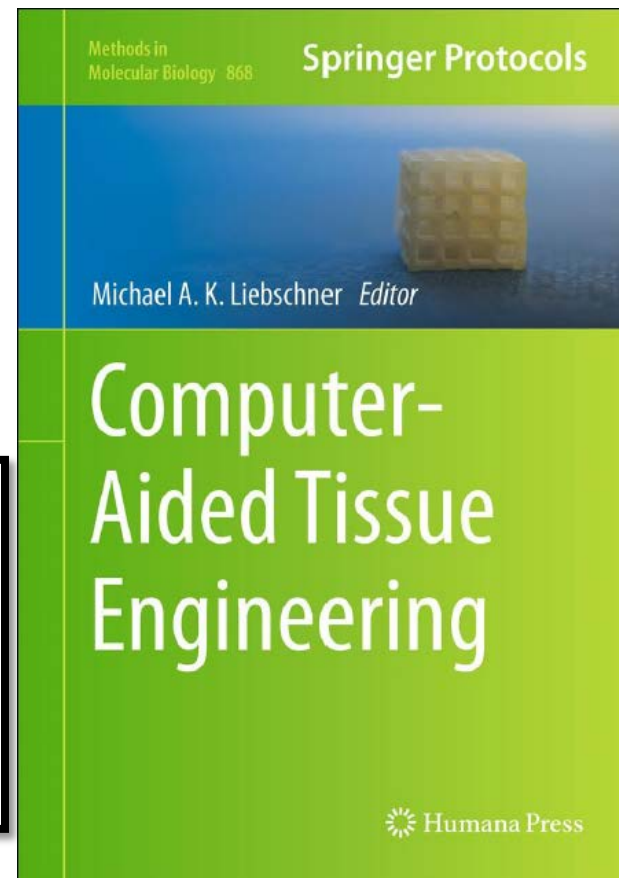
*Table entries for each column indicate the total number and the percentage in parentheses.

Lots of new opportunity!



Systems Pharmacology to Predict Drug Toxicity: Integration Across Levels of Biological Organization*

Jane P.F. Bai and Darrell R. Abernethy



ASSOCIATE EDITOR: ERIC L. BARKER

Computational Methods in Drug Discovery

Gregory Sliwoski, Sandeepkumar Kothiwale, Jens Meiler, and Edward W. Lowe, Jr.

Meiler Laboratory, Center for Structure Biology, Vanderbilt University, Nashville, Tennessee

Leveraging the opportunity

- Alignment on the gaps and the opportunities best suited to fill those gaps
- Clearly defined context of use for alternative test systems
- Sense of what it takes to build confidence in a different approach
 - biological relevance, data, experience
- Willingness to accept managed risk
- Freedom to operate
- Collaboration with shared goals

Challenges of applying a novel modeling **strategy** early in discovery

- Sufficient biological complexity with adequate throughput
- Accounting for the biology not in the platform (novel platforms, though less reductionist, will be reductionist)
- Defining what you're measuring against
- Accounting for dose/exposure extrapolation
- Interpreting more mechanistic endpoints
- Considering rapid changes in scope of drug modalities (small molecules, Ab therapies, oligonucleotides, cell/gene therapy)
- Designing a different intellectual framework
- Impacts on cost and time
- **Building a reason to believe!**

We know where to start

- IQ Dru Safe Attrition of Pharmaceuticals during Preclinical Development
 - 282 compounds
 - 16 pharma contributors
 - contemporary data
- Primary target organs
 - CV, liver, kidney, GI, CNS, testes
 - ~70% of preclinical safety attrition
 - We don't have to replicate the 15K data points in a 28d rat study

Design-Test-Implement- IQ/NCATS/FDA partnership as a model



IQ Microphysiological Systems Working Group Organotypic Standards- Cardiovascular

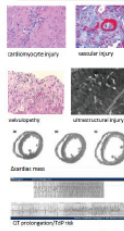
Berridge, B. R., IQ MPS Working Group, HESI Cardiac Safety Technical Committee

Introduction

- The cardiovascular system is dynamic and integrated with significant adaptive function, intra-system interdependencies and non-CV influences
- It is an important source of drug safety liabilities for which the in vivo mammalian system has been important for identifying and characterizing
- The myocardium, heart valves, and intra- and extra-cardiac arteries are primary targets of both structural and functional toxicity
- Contemporary assessments of CV safety include assessment of cardiac rhythm and contractility, systemic blood pressure, heart mass and morphology of the heart and blood vessels
- An appropriately robust, human-relevant MPS system could provide near term opportunities for early screening as well as a means to differentiate species-specific effects
- MPS systems are likely to be more mechanistically informative

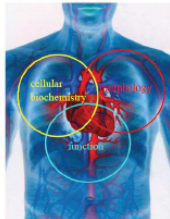
What does cardiovascular toxicity look like?

Nonclinical CV Toxicity



Δ BP
Δ HR
Δ contractility

Clinical CV Toxicity



- "Adverse CV events" (Veno, Durocent)
- Compensative Heart Failure (T2Ds, T1Ds)
- Decreased EF (T1Ds)
- Widulopathy (Fen-Phen)
- QT prolongation
- Non-QT arrhythmias
- Increased blood pressure (toxicologic)

Important physiological features of the CV system

- Integrated system with significant interdependence, reserve capacity and dynamic function
- Significant biological integration at the system (heart and blood vessels), tissue (cardiomyocytes and capillaries) and cellular (cardiomyocyte ion channels) levels
- High energy requirements- substrates = fatty acids > glucose, oxygen, high number of mitochondria
- Relative lack of regenerative capacity in the myocardium
- Functional changes manifest in changes in fluid dynamics. Changes in fluid dynamics can induce structural changes.
- Cardiomyocytes express drug transporters and they can be mediators of toxicity but may be poorly well characterized.

Cardiovascular response to toxic injury, endpoints and test compounds

Cellular components	Structural features	Functional features	Response/mechanisms of injury	Relevant endpoints	Test compounds
Cardiomyocyte	Tightly-coupled with various phenotypes (e.g. contractile apparatus, Ca ²⁺ regulatory proteins, mitochondrial mass, DNA, transporter and channel expression)	Highly contractile with reserve capacity (paced beating with positive force- frequency relation potential; troponin, Sarin/Troponin)	Neuroinjury/apoptosis	LDH, cTnT, TUNEL	LFs, CEM, Doxorubicin, Ixabepilone
			Vasodilation (pulmo, into swelling, ER dilation, lumenal necrosis, glycogen accrual)	microbubbles, lipid stain, into dyes, lactonase stain	retroviral (SDF1), doxorubicin (SDF1, chloroquine (SDF1), Ixabepilone (SDF1), phenacetol (SDF1), phenacetol (SDF1), I)
			Myocarditis	Calcium staining	
			Hypertrophy	Cell size quantitation	iodobutanol, phenylephrine, dobutamine
			Dysrhythmia (e.g. arrhythmogenic dysrhythmia)	Cellular stress, ATP production, Ca ²⁺ concentration	
			Isotropic dysfunction	contractile force/shortening	propranolol, amrinone, thapsigargin, thapsigargin, digoxin
			Luotropic dysfunction	rate of relaxation (S)	nitroglycerine
			Chronotropic dysrhythmia	heart rate	isoproterenol, Muscarinic
			Myofibrillar dysfunction	action potential measures	terfenadine, digoxin
Fibrocyte	Vesicle expression, no DNA	Contractile through generally not-actin/collagen production	Activation to myofibrillar, proliferation, increased collagen production	DNA expression, proliferation	TGFβ
Microvascular cells	Matrix phenotype expressing lamin, fibronectin, ESM production, SMC2B expression	ECM production, contractile through not-shrinking	Activation to myofibrillar, proliferation, increased ECM production	DNA expression, proliferation	AUX-5 inhibitors, losartan/phenacetol
Capillary endothelial cells	Matrix phenotype (ECM expression, vWF, fenestrated endothelial bridges, Weibel-Pellets bodies, pinocytotic vesicles)	selective permeability, anti-coagulation	Neuroinjury/apoptosis	LDH, morphology	LFs, tubule inhibitors, cytophosphamide
			Pro-inflammatory activation	PECAM expression	LFs
			Retraction	Loss of fenestrular bridging, actin	Lu-27
			Hypertrophy	Cell size quantitation	
			Pro-thrombotic activation	PKC, βactin/immunoprecipitation	
			Altered NO/ET expression	platelet adhesion, surface markers	LFs
				NO, ET	endothelial inhibitors, L-NMMA
Skeletal muscle cells	Matrix phenotype (DNA expression, vWF, fenestrated endothelial bridges, Weibel-Pellets bodies, pinocytotic vesicles)	contractile and responsive to NO, ET adrenergic signals	Neuroinjury/apoptosis	LDH, TUNEL	
			Lipid accumulation	lipid stain	T2Ds
			lumenal necrosis	lactonase marker	
			contraction/relaxation	Functional measure	NO, endothelin, hydrochloric, nitroglycerin, losartan
Arterial endothelial cells	Matrix phenotype (ECM expression, vWF, fenestrated endothelial bridges, Weibel-Pellets bodies, pinocytotic vesicles)	NO/ET production, autocrine/paracrine	Neuroinjury/apoptosis, activation, pro-thrombotic activation, hypertrophy	See capillary endothelial cells	

Relating in vitro to in vivo endpoints

In vivo	In vitro	Comments
Heart rate	Beat rate	= autonomic influence
ECC	Fluid potential, conduction velocity	Ability to model re-entrant action potentials
QA interval/dP/dT max	Force of contraction, contraction velocity, calcium flux	Direct contractility measures
Blood pressure	NO production, blood vessel diameter	Measure of direct vasoactivity
Cardiomyocyte hypertrophy-beat weight	Cell size, gene expression	Measure of direct anabolism or catabolism
Cardiomyocyte injury-microscopic morphology, CTN, special staining procedures	Cell size, morphology and viability, gene expression, protein content	Restricted to direct effects but potentially mechanistically insightful
Endothelial injury-microscopic morphology, coagulation factors, inflammation	Gene expression, barrier function	

Contextualizing other capabilities

Bridging Functional and Structural Cardiotoxicity Assays Using Human Embryonic Stem Cell-Derived Cardiomyocytes for a More Comprehensive Risk Assessment
John Clements, Paul Miller, Angela S. Williams, and Stan Kubota
TOXICOLOGICAL SERVICES, 1480, 2015, 30-06

Structural and functional screening in human induced-pluripotent stem cell-derived cardiomyocytes accurately identifies cardiotoxicity of multiple drug types
Kendrick K. Stokely*, Devonson E. Calvert, Patricia E. Trank, Burnard M. Moore, Scott A. Goff, Sarah Beava
Toxicology and Applied Pharmacology 301 (2015) 11-20

Building Confidence

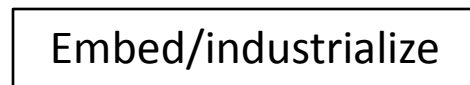
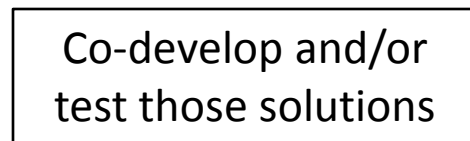
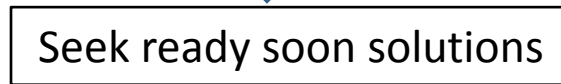
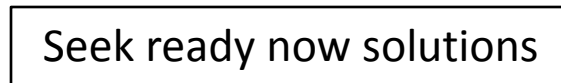
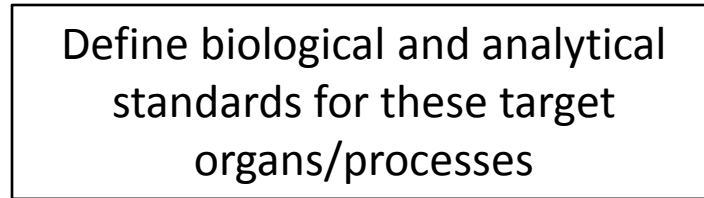
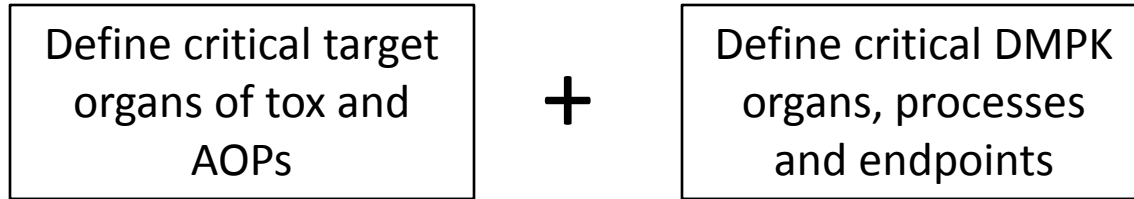
- Relate the platform to a specific component of the integrated system
- Demonstrate relevant in vivo morphologic and functional attributes
- Test known positive and negative compounds for specific outcomes
 - representing a spectrum of mechanisms
 - at relevant in vivo doses
 - with pre-defined success criteria
- Know how to respond to the data- e.g.
 - stop development
 - needs further characterization
 - mitigates animal data
- Be able to represent appropriate context of use, throughput, value proposition, weaknesses

Contact Information

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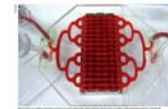
Roadmap



Evolution of Application
Pre-animal Safety Screens

Comparative application with in vivo safety studies

Virtual second safety species

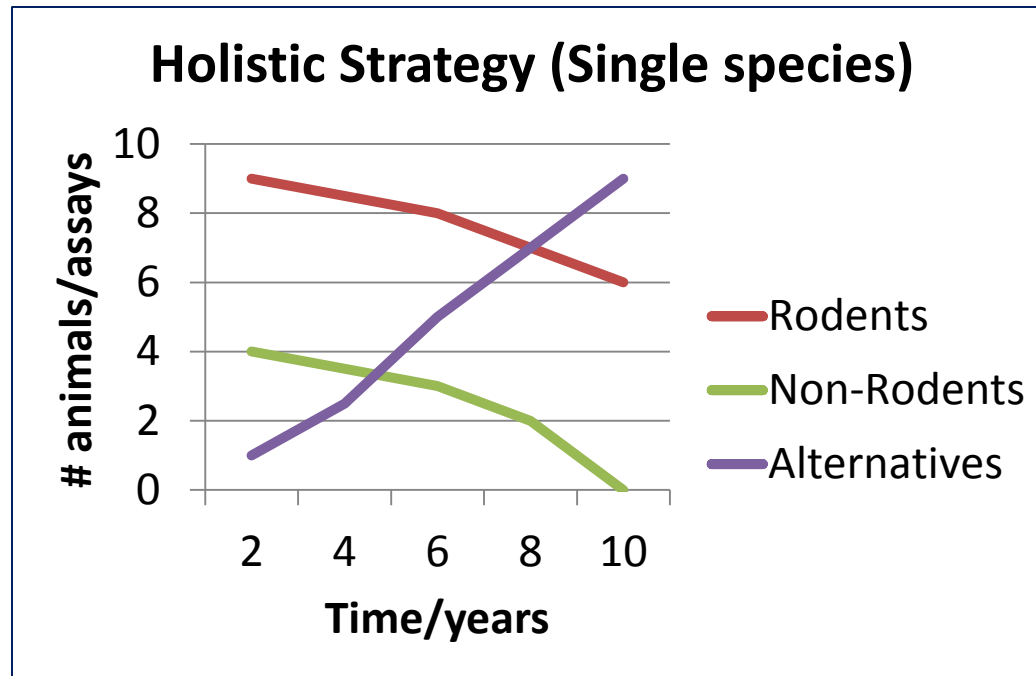


PPP Incubator

The ultimate value proposition

- Improved predictive validity of early preclinical models = lower attrition
 - DMPK + safety in the first instance
 - Parallel application of the principles of this approach to specific disease areas improves efficacy modeling in the second
- Decreased cycle time by bringing the best lead forward the first time (avoids iterative assessment of multiple leads pre-CS)
- Enables early risk:benefit integration
- Decreases animal use
- Lowers development costs
- Efforts in innovation more efficient and impactful in the near term

Alternatives Development Strategy Impact



Incentive-driven

Salient features

- Defined by a bold aspirational goal- i.e. single species safety package
- Alternatives development defined by the prioritized scope of in vivo assessments
- Rate of animal use impact increases with time

Pros

- Deliberate innovation defined by current standards
- Significant alignment and complementarity of investment
- Significant decrease in animal studies- particularly for non-rodents
- Clinical predictivity could/should increase

Cons

- Significant global coordination
- Regulatory acceptance required for full impact
- Structured development and qualification process
- Innovation directed