# DRAFT

# **Report on Carcinogens Background Document for**

# Formaldehyde

September 3, 2009



U.S. Department of Health and Human Services Public Health Service National Toxicology Program Research Triangle Park, NC 27709

This DRAFT background document is distributed solely for the purpose of public comment and predissemination peer review. It should not be construed to represent final NTP determination or policy. This Page Intentionally Left Blank

#### FOREWORD

The Report on Carcinogens (RoC) is prepared in response to Section 301 of the Public Health Service Act as amended. The RoC contains a list of identified substances (i) that either are known to be human carcinogens or are reasonably be anticipated to be human carcinogens and (ii) to which a significant number of persons residing in the United States are exposed. The Secretary, Department of Health and Human Services (HHS), has delegated responsibility for preparation of the RoC to the National Toxicology Program (NTP), which prepares the report with assistance from other Federal health and regulatory agencies and nongovernmental institutions.

Nominations for (1) listing a new substance, (2) reclassifying the listing status for a substance already listed, or (3) removing a substance already listed in the RoC are reviewed in a multi-step, scientific review process with multiple opportunities for public comment. The scientific peer-review groups evaluate and make independent recommendations for each nomination according to specific RoC listing criteria. This background document was prepared to assist in the review of formaldehyde. The scientific information used to prepare Sections 3 through 5 of this document must come from publicly available, peer-reviewed sources. Information in Sections 1 and 2, including chemical and physical properties, analytical methods, production, use, and occurrence may come from published and/or unpublished sources. The NTP will provide a reference for all published and unpublished sources used in this document. For each study cited in the background document from the peer-reviewed literature, information on funding sources (if available) and the authors' affiliations will be provided in the reference section. Any interpretive conclusions, comments, or statistical calculations made by the authors or peer reviewers of this document that are not contained in the original citation are identified in brackets []. This draft document will be peer reviewed in a public forum by an *ad hoc* expert panel of scientists from public and private sectors with relevant expertise and knowledge selected by the NTP in accordance with the Federal Advisory Committee Act and HHS guidelines and regulations. This document will be finalized based on the peer-review recommendations of the expert panel and public comments received for this draft document.

A detailed description of the RoC nomination review process and a list of all substances under consideration for listing in or delisting from the RoC can be obtained by accessing the 12th RoC at <u>http://ntp.niehs.nih.gov/go/9732</u>. The most recent RoC, the 11th Edition (2004), is available at <u>http://ntp.niehs.nih.gov/go/19914</u>.

#### CONTRIBUTORS

#### **Project Managers, Authors, and Principal Reviewers**

# National Toxicology Program (NTP) and National Institute of Environmental Health Sciences (NIEHS)

Ruth Lunn, Dr.P.H.	Director, Report on Carcinogens Center
Gloria Jahnke, D.V.M.	Health Scientist, Report on Carcinogens Center
Diane Spencer, M.S.	Health Scientist, Report on Carcinogens Center
C.W. Jameson, Ph.D.	Report on Carcinogens Center (former Director; currently at CWJ Consulting, LLC)

#### SRA International, Inc. (Support provided through NIEHS Contract Number NO1-ES-35505)

Sanford Garner, Ph.D.Principal InvestigatorStanley Atwood, M.S., DABTGreg Carter, M.E.M.Andrew Ewens, Ph.D.Dana Greenwood, B.S.Jennifer Ratcliffe, Ph.D.

#### **Consultants**

Tania Desrosiers, M.P.H. Joe Haseman, Ph.D. University of North Carolina Independent Consultant

#### Administrative Support

Ella Darden, B.S. Tracy Saunders, B.S. Jenaya Brown SRA International, Inc. SRA International, Inc. Report on Carcinogens Center, NIEHS

#### Criteria for Listing Agents, Substances or Mixtures in the Report on Carcinogens

#### U.S. Department of Health and Human Services

#### **National Toxicology Program**

The criteria for listing an agent, substance, mixture, or exposure circumstance in the RoC are as follows:

#### Known To Be Human Carcinogen:

There is sufficient evidence of carcinogenicity from studies in humans<sup>\*</sup>, which indicates a causal relationship between exposure to the agent, substance, or mixture, and human cancer.

#### Reasonably Anticipated To Be Human Carcinogen:

There is limited evidence of carcinogenicity from studies in humans<sup>\*</sup>, which indicates that causal interpretation is credible, but that alternative explanations, such as chance, bias, or confounding factors, could not adequately be excluded,

or

there is sufficient evidence of carcinogenicity from studies in experimental animals, which indicates there is an increased incidence of malignant and/or a combination of malignant and benign tumors (1) in multiple species or at multiple tissue sites, or (2) by multiple routes of exposure, or (3) to an unusual degree with regard to incidence, site, or type of tumor, or age at onset,

or

there is less than sufficient evidence of carcinogenicity in humans or laboratory animals; however, the agent, substance, or mixture belongs to a well-defined, structurally related class of substances whose members are listed in a previous Report on Carcinogens as either known to be a human carcinogen or reasonably anticipated to be a human carcinogen, or there is convincing relevant information that the agent acts through mechanisms indicating it would likely cause cancer in humans.

Conclusions regarding carcinogenicity in humans or experimental animals are based on scientific judgment, with consideration given to all relevant information. Relevant information includes, but is not limited to, dose response, route of exposure, chemical structure, metabolism, pharmacokinetics, sensitive sub-populations, genetic effects, or other data relating to mechanism of action or factors that may be unique to a given substance. For example, there may be substances for which there is evidence of carcinogenicity in laboratory animals, but there are compelling data indicating that the agent acts through mechanisms which do not operate in humans and would therefore not reasonably be anticipated to cause cancer in humans.

This evidence can include traditional cancer epidemiology studies, data from clinical studies, and/or data derived from the study of tissues or cells from humans exposed to the substance in question that can be useful for evaluating whether a relevant cancer mechanism is operating in people.

## **Executive Summary**

#### Introduction

1 Formaldehyde is a high-production-volume chemical with a wide array of uses. The 2 predominant use of formaldehyde in the United States is in the production of industrial 3 resins (mainly urea-formaldehyde, phenol-formaldehyde, polyacetal, and melamine-4 formaldehyde resins) that are used to manufacture products such as adhesives and binders 5 for wood products, pulp and paper products, plastics, and synthetic fibers, and in textile 6 finishing. Formaldehyde is also used as a chemical intermediate. Resin production and 7 use as a chemical intermediate together account for over 80% of its use. Other, smaller 8 uses of formaldehyde that may be important for potential human exposure include use in 9 agriculture, medical use as a disinfectant and preservative (for pathology, histology, and 10 embalming), and use in numerous consumer products as a biocide and preservative. 11 Formaldehyde (gas) is listed in the *Eleventh Report on Carcinogens* (RoC) as *reasonably* 12 anticipated to be a human carcinogen based on limited evidence of carcinogenicity in 13 humans and sufficient evidence of carcinogenicity in laboratory animals (NTP 2005a); it 14 was first listed in the 2nd RoC (NTP 1981). Formaldehyde (all physical forms) was

15 nominated by NIEHS for possible reclassification in the 12th RoC based on the 2004

16 review by the International Agency for Research on Cancer (IARC 2006), which

17 concluded that there was sufficient evidence for the carcinogenicity of formaldehyde in

18 humans.

#### Human Exposure

Formaldehyde has numerous industrial and commercial uses and is produced in very large amounts (billions of pounds per year in the United States) by catalytic oxidation of methanol. Its predominant use, accounting for roughly 55% of consumption, is in the production of industrial resins, which are used in the production of numerous commercial products. Formaldehyde is used in industrial processes primarily as a solution (formalin) or solid (paraformaldehyde or trioxane), but exposure is frequently to formaldehyde gas, which is released during many of the processes. Formaldehyde gas is also created from 1 the combustion of organic material and can be produced secondarily in air from

2 photochemical reactions involving virtually all classes of hydrocarbon pollutants. In

3 some instances, secondary production may exceed direct air emissions. Formaldehyde is

4 also produced endogenously in humans and animals.

5 Formaldehyde is a simple, one-carbon molecule that is rapidly metabolized, is 6 endogenously produced, and is also formed through the metabolism of many xenobiotic agents. Because of these issues, typical biological indices of exposure, such as levels of 7 8 formaldehyde or its metabolites in blood or urine, have proven to be ineffective measures 9 of exposure. Formaldehyde can bind covalently to single-stranded DNA and protein to 10 form crosslinks, or with human serum albumin or the N-terminal value of hemoglobin to 11 form molecular adducts, and these reaction products of formaldehyde might serve as 12 biomarkers for exposure to formaldehyde.

13 Occupational exposure to formaldehyde is highly variable and can occur in numerous

14 industries, including the manufacture of formaldehyde and formaldehyde-based resins,

15 wood-composite and furniture production, plastics production, histology and pathology,

16 embalming and biology laboratories, foundries, fiberglass production, construction,

17 agriculture, and firefighting, among others. In fact, because formaldehyde is ubiquitous, it

18 has been suggested that occupational exposure to formaldehyde occurs in all work places.

19 Formaldehyde is also ubiquitous in the environment and has been detected in indoor and

20 outdoor air; in treated drinking water, bottled drinking water, surface water, and

21 groundwater; on land and in the soil; and in numerous types of food.

The primary source of exposure is from inhalation of formaldehyde gas in indoor settings(both residential and occupational); however, formaldehyde also may adsorb to respirable

24 particles, providing a source of additional exposure. Major sources of formaldehyde

25 exposure for the general public have included combustion sources (both indoor and

26 outdoor), automobile emissions, off-gassing from numerous construction and home

27 furnishing products, off-gassing from numerous consumer goods, and cigarette smoke.

28 Ingestion of food and water can also be a significant source of exposure to formaldehyde.

- 1 Numerous agencies, including the Department of Homeland Security, CPSC, EPA, FDA,
- 2 HUD, the Mine Safety and Health Administration, OSHA, the Pipeline and Hazardous
- 3 Materials Safety Administration, ACGIH, and NIOSH, have developed regulations and
- 4 guidelines to reduce exposure to formaldehyde.

#### **Human Cancer Studies**

5 A large number of epidemiological studies have evaluated the relationship between 6 formaldehyde exposure and carcinogenicity in humans. The studies fall into the following 7 main groups: (1) historical cohort studies and nested case-control studies of workers in a 8 variety of industries that manufacture or use formaldehyde, including the chemical, 9 plastics, fiberglass, resins, and woodworking industries, as well as construction, garment, 10 iron foundry, and tannery workers; (2) historical cohort studies of health professionals, 11 including physicians, pathologists, anatomists, embalmers, and funeral directors; and (3) 12 population-based or occupationally-based case-control incidence or mortality studies of 13 specific cancer endpoints. In addition, several studies have re-analyzed data from specific 14 cohort or case-control studies or have conducted pooled analyses or meta-analyses for 15 specific cancer endpoints

16 The largest study available to date is the combined cohort mortality study of mixed 17 industries conducted by the National Cancer Institute (NCI). This cohort includes 26,561 18 male and female workers, enrolled from ten different formaldehyde-producing or using 19 industries, employed before 1966 and followed most recently to 1994 and 2004, most of 20 the workers were exposed to formaldehyde (Hauptmann et al. 2003, 2004 and Beane 21 Freeman et al. 2009). Quantitative exposure data were used to construct job exposure 22 matrices for individual workers, some of whom experienced peak exposures to 23 formaldehyde > 4 ppm. This cohort is the only study in which exposure-response 24 relationships for peak, average, cumulative, and duration of exposures and mortality for 25 multiple cancer sites were investigated. Two other large cohort studies are available: (1) a 26 large multi-plant cohort study (N = 14,014) of workers in six chemical manufacturing 27 plants in the United Kingdom (Coggon et al. 2003), which calculated SMRs among ever-28 exposed and highly exposed workers for formaldehyde, and (2) a NIOSH cohort of 29 garment workers (N = 11,039), which evaluated mortality for duration of exposure, time

1 since first exposure, and year of first exposure to formaldehyde for selected cancer sites. 2 The other cohorts (both for industrial and professional health workers) were smaller, and 3 in general only reported mortality or incidence for ever-exposed workers in external 4 (SMR or PMR) analyses, although some of the studies of professional health workers 5 attempted indirect measures of exposure (such as length in a professional membership) as 6 a proxy for exposure duration. In general, the majority of the nested case-control and 7 other studies attempted to look at exposure-response relationships, but most were semi-8 quantitative. Since most of the cohorts have relatively low statistical power to evaluate 9 rare cancers such as sinonasal and nasopharyngeal cancer, case-control studies are 10 generally more informative for these outcomes. Findings across studies for cancer sites 11 that have been the principal focus of investigation are summarized below.

#### 12 Sinonasal cancers

13 There are two major histological types of sinonasal cancer (adenocarcinomas and 14 squamous-cell carcinomas). Sinonasal cancers are rare, and the majority of cohort studies 15 have insufficient numbers of exposed workers to be informative; many of the cohort 16 studies did not report findings or did not observe any deaths for this specific endpoint. 17 Increased risks of sinonasal cancers were observed among male (SPICR = 2.3, 95% CI = 18 1.3 to 4.0, 13 exposed cases) and female (SPICR = 2.4, 95% CI = 0.6 to 6.0, 4 exposed cases) Danish workers exposed to formaldehyde (Hansen and Olsen 1995, 1996) and 19 20 among formaldehyde-exposed workers in the NCI cohort (SMR = 1.19, 95% CI = 0.38 to 21 3.68, 3 deaths) (Hauptmann et al. 2004). No increase in risk was found among 22 formaldehyde-exposed workers in the large cohort of British chemical workers, based on 23 two observed deaths (Coggon et al. 2003). Of the six case-control studies reviewed, four 24 (Olsen et al. 1994, Olsen and Asnaes 1986, Hayes et al. 1986, Roush et al. 1987, and 25 Luce et al. 1993) reported an association between sinonasal cancers and formaldehyde 26 exposure; statistically significant risks were found in three studies (for ever exposed or 27 individuals with higher measures of exposure) (Olsen et al. 1994, Hayes et al. 1986, Luce 28 et al. 1993). Stronger associations were found for adenocarcinomas, and higher risks of 29 adenocarcinomas were found among individuals with higher average and cumulative 30 exposure, duration of exposure, and earlier dates of first exposure (Luce *et al.* 1993).

1 Wood dust is an established cause of sinonasal cancer, particularly adenocarcinomas 2 (NTP 2005) and is a possible confounder in studies of woodworking industry workers; 3 however, elevated risks for formaldehyde exposure were found among workers with low 4 or no exposure to wood dust (Hayes et al. 1986, Olsen et al. 1994, Olsen and Asnaes 5 1986, Luce et al. 1993) and a possible synergistic effect was suggested in the latter two 6 studies. A pooled analysis of 12 case-control studies of sinonasal cancer from seven 7 countries (Luce et al. 2002) found an increase in adenocarcinomas among formaldehyde-8 exposed cases, adjusted for wood dust exposure, with increasing level of estimated 9 exposure (OR = 3.0, 95% CI = 1.5 to 5.7, 91 exposed cases for men and OR = 1.5, 95%10 CI = 0.6 to 3.8, 6 exposed cases for women; both in the highest exposure groups). For 11 squamous-cell carcinomas, the association with formaldehyde exposure was weaker, 12 except among men with 30 or more years of exposure (OR = 1.4, 95% CI = 0.9 to 2.3, 13 number of cases not specified; not adjusted for wood dust exposure).

#### 14 Nasopharyngeal cancers

15 As in the case of sinonasal cancer, nasopharyngeal cancers are rare, and the majority of 16 cohort studies have insufficient numbers of exposed workers to be informative. Several 17 cohort studies did not report findings for nasopharyngeal cancer, or observed one or no 18 cases or deaths, for this tumor site. A statistically significant increase in mortality from 19 nasopharyngeal cancer was observed in the large NCI cohort (SMR = 2.10, 95% CI = 20 1.05 to 4.21, 8 exposed cases, one subsequently reclassified as oropharygneal cancer) 21 (Hauptmann et al. 2004). Statistically non-significantly elevated risks were observed 22 among white embalmers from the United States (SMR = 1.89, 95% CI = 0.39 to 5.48, 323 deaths) (Hayes et al. 1990), and among male Danish workers exposed to formaldehyde 24 (SPICR = 1.3, 95% CI = 0.3 to 3.2, 4 exposed cases) (Hansen and Olsen 1995, 1996). In 25 the British chemical workers cohort, one death was observed (SMR not reported) 26 (Coggon *et al.* 2003).

27 Exposure-response relationships between formaldehyde exposure and nasopharyngeal

28 cancers risk were evaluated in the large NCI cohort study. Among seven exposed deaths,

- 29 relative risks of nasopharyngeal cancers increased with peak exposure ( $P_{\text{trend}} < 0.001$ ),
- 30 average exposure ( $P_{\text{trend}} = 0.066$ ) and cumulative exposure ( $P_{\text{trend}} = 0.025$ ); tests for trend

1 among combined, exposed, and unexposed workers were  $P_{\text{trend}} = 0.044, 0.126, \text{ and } 0.029,$ 2 respectively. Adjustment for duration of exposure to a number of potentially confounding 3 substances did not substantively alter the findings. An analysis adjusted for plant type 4 found statistically significant trends among exposed workers for peak and cumulative 5 exposure and duration of exposure. Marsh and colleagues studied one of the plants, in 6 which five of the nasopharyngeal cancers deaths had occurred, separately (Marsh et al. 7 2002, 2007a). These authors also reanalyzed the nasopharyngeal cancers cancer findings 8 in the NCI cohort (Marsh et al. 2007b) and concluded that external employment in metal 9 working may have partly explained the findings for nasopharyngeal cancers in this

10 cohort.

Х

11 Six of the seven available case-control studies reported increases in nasopharyngeal 12 cancers in association with probable exposure to formaldehyde or at higher levels or 13 duration of estimated exposure (Olsen et al. 1984 [women only], Vaughan et al. 1986, 14 Roush et al. 1987, West et al. 1993, Vaughan et al. 2000, and Hildesheim et al. 2001). 15 Risks of nasopharyngeal cancers increased with exposure duration and cumulative 16 exposure in two population based case-control studies (Vaughan et al. 2000, Hildesheim 17 et al. 2001). In a meta-analysis of case-control and cohort studies (Collins et al. 1997), a 18 statistically significant increased risk for nasopharyngeal cancers and formaldehyde 19 exposure was estimated (mRR = 1.3, 95% CI = 1.2 to 1.5), and a pooled analysis of 20 SMRs from three cohort mortality studies (Bosetti et al. 2008) reported an overall 21 increase in the SMR of 1.33 (95% CI = 0.61 to 2.53, 9 deaths).

#### 22 Other head and neck cancers, and respiratory cancer

23 Most cohort studies reported risk estimates for cancers of the buccal cavity, pharynx, 24 larynx, and lung or combinations of these cancers. Most of these studies, including two of 25 the three larger cohorts (Pinkerton et al. 2004 and Coggon et al. 2003), three of the professional health worker studies (Hayes et al. 1990, Walrath and Freumeni 1983 and 26 27 1984), and two of the smaller industrial cohorts (Anjelkovich et al. 1995 and Hansen and 28 Olsen 1995, 1996) found elevated (between approximately 10% and 30%) but 29 statistically non-significant risks for cancers of the buccal cavity or buccal cavity and 30 pharynx combined; risk estimates were usually based on small numbers of deaths or

1 cases. In the NCI cohort, no association between buccal cavity and formaldehyde 2 exposure was observed; however, a statistically significant increased risk for all upper 3 respiratory cancers combined was found among workers with the highest average 4 exposure (> 1 ppm) compared with the lowest exposure group (RR = 2.21, 15 deaths) 5 (Hauptmann et al. 2004). Relative risks increased somewhat with increasing average and 6 peak (but not cumulative) exposure, but the trends were not statistically significant. Most 7 of the case-control studies that reported on head and neck cancers found elevated (usually 8 statistically non-significant) risks for formaldehyde exposure and cancers of the buccal 9 cavity and pharynx (or parts of the pharynx) (Vaughan et al. 1986, Merletti et al. 1991, 10 Gustavsson et al. 1998, Laforest et al. 2000, Marsh et al. 2002, Wilson et al. 2004). 11 Positive exposure-response relationships with probability and duration of exposure for 12 cancers of the hypopharynx and larynx combined were reported by Laforest et al. (2000) 13 and for combined probability and intensity of exposure and salivary cancer by Wilson et 14 al. (2004). No clear association between formaldehyde exposure and hypopharyngeal or 15 laryngeal cancer was observed by Berrino *et al.* (2003) or for combined head and neck 16 cancers by Tarvainen et al. (2008). Most of the cohort studies and two of the three 17 available case-control studies found no association between formaldehyde exposure and 18 laryngeal cancer. Bosetti et al. (2008) calculated a combined estimated RR (using a 19 weighted average of SMRs and/or PMRs) for combined buccal cavity and pharynx of 20 1.09 (95% CI = 0.88 to 1.34, 88 deaths) among industrial workers and 0.96 (95% CI = 21 0.75 to 1.24, 61 deaths) among health professional workers exposed to formaldehyde in a 22 pooled analysis of 10 occupational cohort mortality studies.

23 Five of the industrial cohort studies reported increases in the risk of lung or respiratory

system cancers (Andjelkovich et al. 1995, Bertazzi et al. 1986, Dell and Teta 1995,

25 Hansen and Olsen 1996 [women only]) including the large cohort of British chemical

26 workers, which reported a statistically significant increased risk (SMR = 1.22, 95% CI =

1.12 to 1.32, 594 deaths, all workers) (Coggon et al. 2003). In this study, risks increased

28 with increasing exposure level ( $P_{\text{trend}} < 0.001$ ) but not with duration of exposure. No

association was observed in the other two large cohorts (Pinkerton et al. 2004,

30 Hauptmann et al. 2004), in several of the smaller occupational cohorts (Hansen and

31 Olsen 1995, 1996 [in men, although a small increase was seen in women], Edling *et al.* 

1 1987b, Stellman et al. 1998, Stern et al. 1987), or in the six studies of health professional 2 workers. Findings from case-control studies were also mixed: statistically significant 3 increased risks were found among fiberglass manufacturing workers who were ever 4 exposed to formaldehyde (OR = 1.61, 95% CI = 1.02 to 2.57, 591 cases) (Marsh *et al.* 5 2001) and among formaldehyde-exposed individuals in a population-based case-control 6 study (Coggon et al. 1984), although risks were not increased among workers with higher 7 exposure. Three studies reported statistically non-significant elevated risks for lung 8 cancer, but no clear exposure response patterns were observed (Gerin *et al.* 1989, 9 Andjelkovich et al. 1994, Chiazze et al. 1997). No association of lung cancer with 10 formaldehyde exposure was reported in three other occupational case-control studies and 11 one population-based study (Bond et al. 1986, Jensen and Andersen 1982, Partanen et al. 12 1990, Brownson et al. 1993). In a pooled analysis of 14 occupational mortality studies of 13 formaldehyde exposure, which included an analysis of lung cancers, Bosetti et al. (2008) 14 calculated a combined RR of 1.06 (95% CI = 0.92 to 1.23, 1.459 deaths) among 15 industrial workers and 0.63 (95% CI = 0.47 to 0.84, 562 deaths) among health professional workers. 16

17 Lymphohematopoietic cancers

18 Among workers in the NCI cohort study, peak exposure to formaldehyde was associated with increased mortality for several types of lymphohematopoietic cancer (Beane 19 20 Freeman et al. 2009). With respect to all lymphohematopoietic cancers combined and 21 leukemias, relative risks increased with increasing peak exposure and statistically 22 significant increased risks were found among workers with the highest peak exposure ( $\geq$ 23 4ppm) vs. the lowest exposed category for all lymphohematopoietic cancers (OR = 1.37, 24 95% CI = 1.03 to 1.81, 108 deaths,  $P_{\text{trend}} = 0.02$ ) and statistically non-significant increases 25 in risk were observed for all leukemia and peak exposure  $\geq$  4ppm (RR = 1.42, 95% CI = 26 0.92 to 2.18, 48 deaths,  $P_{\text{trend}} = 0.02$ ) and for myeloid leukemia (RR = 1.78, 95% CI = 27 0.87 to 3.64, 19 deaths,  $P_{\text{trend}} = 0.13$ ). No association was found with cumulative or 28 average exposure. Leukemias observed in the earlier (1984) NCI follow-up (Hauptmann 29 et al. 2003) were re-analyzed by Marsh and Youk (2004) using different exposure

1 assessments; these authors reported no statistically significant trends with exposure, 2 although risks remained elevated for all leukemias (combined) and myeloid leukemia. 3 Increases in all lymphohematopoietic cancers were also observed in other studies. Each 4 of the studies of health professionals found elevated mortality for all 5 lymphohematopoietic cancers combined and for leukemia (Hall et al. 1991, Hayes et al. 6 1990, Stroup et al. 1986, Levine et al. 1984 and Walrath and Fraumeni 1983, 1984). 7 Most estimates were statistically non-significant, except for those of Hayes *et al.* (1990), 8 and Stroup et al. (1986), where statistically significant excess mortality was found for all 9 leukemia or myeloid leukemia. An excess of leukemia, especially myeloid leukemia, was 10 also found among garment workers in the large NIOSH cohort (Pinkerton et al. 2004), 11 but not in the British chemical workers cohort (Coggon et al. 2003). In the NIOSH 12 cohort, risks for leukemia, myeloid leukemia, and acute myeloid leukemia were higher 13 among workers with longer duration of exposure (> 10 yrs), longer time since first 14 exposure (> 20 years), and among those exposed prior to 1963 (when formaldehyde 15 exposure was thought to be higher). In the smaller industrial cohort studies, some studies 16 reported excesses for lymphohematopoietic cancers combined (Bertazzi et al. 1986, 17 Stellman et al. 1998) or leukemia (Hansen and Olsen 1995, 1996, Stern et al. 1987), but 18 others observed no associations among formaldehyde-exposed workers for all 19 lymphohematopoietic cancers (Pinkerton et al. 2004, Andjelkovich et al. 1995) or 20 leukemia (Stellman et al. 1998). Of the three available case-control studies, a population-21 based study found no association between leukemia and exposure to formaldehyde (Blair 22 et al. 2001), and two nested case control studies reported statistically non-significant 23 increases in risk based on small numbers of exposed cases (Partanen et al. 1993, and Ott 24 et al. 1989).

Few cohort studies reported findings for other types of lymphohematopoietic cancers. Most of the cohort studies had relatively low power to detect effects, and either did not report findings or did not evaluate exposure-response relationships. The NCI study was the only cohort that observed an association between formaldehyde exposure and Hodgkin's lymphoma (Beane Freeman *et al.* 2009). Among exposed workers, relative risks increased with increasing peak ( $P_{trend} = 0.01$ ) and average exposure ( $P_{trend} = 0.05$ ),

1	but not with cumulative exposure; statistically significant risks were found for the highest
2	peak ( $\geq$ 4.0 ppm) vs. lowest formaldehyde exposure category (RR = 3.96, 95% CI = 1.31
3	to 12.02, 11 deaths). In external analyses, a statistically non-significant elevation in
4	mortality was observed (SMR = 1.4, 95% CI = 0.96 to 2.10, 25 deaths). For non-
5	Hodgkin's lymphoma (NHL), almost all the cohort studies that reported results observed
6	no increases in mortality or incidence. Two nested case-control studies (Partanen et al.
7	1993, Ott et al. 1989) reported increases in NHL risk, but these studies had very small
8	numbers of exposed cases. In the population case-control studies, the risk of NHL
9	increased with increasing probability and intensity combined ( $P < 0.001$ ) in a large U.S.
10	study (Wang et al. 2008), but most of the other studies found no clear association (Gerin
11	et al. 1989, McDuffie et al. 2001, Tatham et al. 1997). For multiple myeloma, peak
12	exposure was associated with a statistically significant increase in risk in the NCI cohort
13	(RR= 2.04, 95% CI = 1.01 to 4.12, 21 deaths, $P_{\text{trend}} = 0.08$ ) (Beane Freeman <i>et al.</i> 2009),
14	and increased risks were seen among British chemical workers (Coggon et al. 2003),
15	abrasive materials workers (Edling et al. 1987b), and U.S. embalmers (Hayes et al.
16	1990). Other studies did not find associations. Small but non-significant increases in risks
17	were also observed in three case-control studies (Boffetta et al. 1989, Heineman et al.
18	1992, Pottern et al. 1992).
19	Bosetti et al. 2008 conducted a pooled analysis of 12 cohort mortality studies and
20	reported a pooled estimated RR for all lymphohematopoietic cancers of 0.85 (95% $CI =$
21	0.74 to 0.96, 234 deaths) for industrial workers and 1.31 (95% CI = 1.16 to 1.48, 263
22	deaths) for health professional workers. The corresponding pooled RRs for leukemia
23	were 0.90 (95% CI = 0.75 to 1.07, 122 deaths) and 1.39 (95% CI = 1.15 to 1.68, 106
24	deaths), respectively. A meta-analysis by Collins and Lineker (2004) of leukemia and
25	formaldehyde exposure among 12 cohort and case-control studies reported an mRR of

- 1.1 (95% CI = 1.0 to 1.2). Zhang *et al.* (2009a) conducted a meta-analysis of data from 26
- 27 studies of occupations with known high formaldehyde exposures, and found an mRR of
- 28 1.25 (95% CI = 1.09 to 1.43) for all lymphohematopoietic cancers (19 studies), an mRR
- of 1.31 (95% CI = 1.02 to 1.67, P = 0.02, 9 studies) for multiple myeloma, and an mRR
- 30 of 1.54 (95% CI =1.18 to 2.00, P < 0.001, 15 studies) for leukemia in association with

1 formaldehyde exposure. The highest risk in the latter group was among myeloid

2 leukemias (mRR = 1.90, 95% CI = 1.31 to 2.76, P = 0.001, 6 studies).

#### 3 Other cancer sites

4 In general, few of the cohort studies reported consistently elevated risks for cancers at 5 other sites. [Not all studies reported findings for all cancer sites and few studies included 6 women.] Few case-control studies of other cancer endpoints have been conducted. An 7 excess of mortality from brain and central nervous system cancers have been reported in 8 all six of the cohort studies of health professionals; statistically significant SMR/PMRs 9 (1.68 to 2.7) were reported in three studies (Stroup et al. 1986, Walrath and Fraumeni 10 1983, 1984). Higher risks were found among workers with longer employment as 11 estimated by length of professional membership (Stroup et al. 1986). No increases in 12 these cancers have been observed in the industrial cohort studies that have reported 13 findings, although a small increased risk was reported among garment workers exposed 14 20 years since first exposure (SMR = 1.20, CI not reported, 13 deaths), and among those 15 whose first exposure was prior to 1963 (Pinkerton et al. 2004). A pooled analysis of 16 cohorts by Bosetti et al. (2008) found an increase of 1.56 (95% CI = 1.24 to 1.96, 74 17 deaths) among professional health workers but not among industrial cohorts.

Several industrial studies have reported increases in stomach, colon, rectal, and kidney cancers, and a case-control study of pancreatic cancer (Kernan *et al.* 1999) suggested an increase in this endpoint at higher levels of formaldehyde exposure. Two meta-analyses of pancreatic cancer (Ojajarvi *et al.* 2000, Collins *et al.* 2001) showed no consistent increase in risk across studies, with the possible exception of a statistically significant increase among pathologists, anatomists and embalmers.

#### **Studies in Experimental Animals**

24 Formaldehyde has been tested for carcinogenicity in mice, rats, and hamsters. Studies

- 25 reviewed include chronic and subchronic inhalation studies in mice, rats, and hamsters;
- 26 chronic and subchronic drinking-water studies in rats; and one chronic skin-application
- 27 study in mice. No chronic studies in primates were found, but one subchronic inhalation
- study and one acute/subacute inhalation study in monkeys were reviewed.

1 Formaldehyde exposure resulted in nasal tumors (primarily squamous-cell carcinoma) in 2 rats when administered chronically by inhalation (Kerns *et al.* 1983, Appelman *et al.* 3 1988, Woutersen et al. 1989, Sellakumar et al. 1985, Monticello et al. 1996, Kamala et 4 al. 1997). Only two inhalation studies in mice or hamsters were found. No tumors were reported in C3H mice exposed to formaldehyde at 200 mg/m<sup>3</sup> for 1 hour/day, 3 5 6 days/week, for 35 weeks (Horton et al. 1963), but squamous-cell carcinoma of the nasal 7 cavity occurred in 2 of 120 B6C3F<sub>1</sub> male mice exposed at 14 ppm for 6 hours/day, 5 8 days/week, for 104 weeks (Kerns et al. 1983). The authors concluded that the tumors 9 were exposure-related, although the increase was not statistically significant. No tumors 10 were reported in Syrian golden hamsters exposed at 10 ppm for life (Dalbey 1982) or 11 2.95 ppm for 26 weeks (Rusch et al. 1983). No tumors occurred in monkeys exposed at 12 2.95 ppm for 26 weeks (Rusch et al. 1983) or 6 ppm for 6 weeks (Monticello et al. 1989); 13 however, squamous metaplasia and hyperplasia in the nasal passages and respiratory 14 epithelia of the trachea and major bronchi occurred.

15 Male rats administered formaldehyde in drinking water at 5,000 ppm for 32 weeks 16 developed forestomach tumors (squamous-cell papillomas) in one study (Takahashi et al. 17 1986); however, in two other drinking-water studies, no tumors were reported in either 18 male or female rats administered formaldehyde at concentrations ranging from 20 to 19 5,000 ppm for two years (Til et al. 1989, Tobe et al. 1989). In another study, male and 20 female breeder rats administered formaldehyde at 2,500 ppm in drinking water had 21 slightly increased incidences of hemolymphoreticular neoplasms (Soffritti et al. 1989). 22 Offspring of these breeder rats exposed transplacentally beginning on gestation day 13 23 and postnatally via drinking water for life showed increased incidences of benign and 24 malignant tumors of the gastrotinestinal tract, particularly intestinal leiomyosarcoma. 25 Male rats administered formaldehyde at concentrations up to 1,500 ppm showed 26 increased incidences (compared with control groups given tap water or tap water 27 containing 15 mg/L methanol) of the number of animals bearing malignant tumors, 28 hemolymphoreticular neoplasms (leukemia and lymphoma combined), and testicular 29 tumors (interstitial-cell adenoma) (Soffritti et al. 2002a). Female rats showed higher 30 incidences of mammary-gland adenocarcinoma and hemolymphoreticular neoplasms than 31 the tap-water control group; however, the incidences were not significantly higher than in 1 the tap-water-plus-methanol control group. In addition, some rare stomach and intestinal

2 tumors occurred in a few male and female rats in the exposed groups but not in the

3 control groups.

4 Other studies examined the promoting effects of formaldehyde when administered after

5 initiation with DBMA, DEN, MNU, or MNNG or cocarcinogenic effects when

6 administered with coal tar, benzo[a]pyrene, wood dust, and hydrogen chloride. Some of

7 these studies did not show an enhanced tumor response. However, a few studies,

8 including a skin-painting study in mice (Iverson et al. 1986), a drinking-water study in

9 rats (Takahashi et al. 1986), and inhalation studies in rats (Albert et al. 1982, Holmstorm

10 et al. 1989a) and hamsters (Dalbey et al. 1986), indicated that formaldehyde could act as

11 a tumor promoter or act as a cocarcinogen when administered with other substances.

#### 12 Adsorption, distribution, metabolism, and excretion

13 Formaldehyde is a metabolic intermediate that is essential for the biosynthesis of purines, 14 thymidine, and some amino acids. The metabolism of formaldehyde is similar in all 15 mammalian species studied. Differences in distribution following inhalation exposure can 16 be related to anatomical differences. For example, rats are obligate nose breathers while 17 monkeys and humans are oronasal breathers. Thus, in humans, some inhaled 18 formaldehyde will bypass the nasal passages and deposit directly into the lower 19 respiratory tract. The endogenous concentrations in the blood of humans, rats and 20 monkeys are about 2 to 3  $\mu$ g/g and do not increase after ingestion or inhalation of 21 formaldehyde from exogenous sources. Although formaldehyde is rapidly and almost completely absorbed from the respiratory or gastrointestinal tracts, it is poorly absorbed 22 23 from intact skin. When absorbed after inhalation or ingestion, very little formaldehyde 24 reaches the systemic circulation because it is rapidly metabolized at the site of absorption 25 to formate, which is excreted in the urine or oxidized to carbon dioxide and exhaled. 26 Although the metabolic pathways are the same in all tissues, the data indicate that route 27 of absorption does affect the route of elimination. When inhaled, exhalation is the 28 primary route of elimination; however, when ingested, urinary excretion as formate is 29 more important. Unmetabolized formaldehyde reacts non-enzymatically with sulfhydryl

1 groups or urea, binds to tetrahydrofolate and enters the single-carbon intermediary

2 metabolic pool, or reacts with macromolecules to form crosslinks (primarily between

3 protein and single-stranded DNA).

#### 4 Toxic effects

Formaldehyde is a highly reactive chemical that causes tissue irritation and damage on
contact. Because of its reactivity and rapid metabolism, toxicity is generally limited to
local effects. *In vitro* studies have demonstrated that formaldehyde is cytotoxic and
affects cell viability, cell differentiation and growth, cell proliferation, gene expression,
membrane integrity, mucociliary action, apoptosis, and thiol and ion homeostasis.
Furthermore, cells depleted of glutathione are more susceptible to formaldehyde toxicity.

11 Formaldehyde concentrations that have been associated with various toxic effects in 12 humans show wide interindividual variation and are route dependent. Symptoms are rare 13 at concentrations below 0.5 ppm; however, upper airway and eye irritation, changes in 14 odor threshold, and neurophysiological effects (e.g., insomnia, memory loss, mood 15 alterations, nausea, fatigue) have been reported at concentrations  $\leq 0.1$  ppm. The most 16 commonly reported effects include eye, nose, throat and skin irritation. Other effects 17 include allergic contact dermatitis, histopathological abnormalities (e.g., hyperplasia, 18 squamous metaplasia, and mild dysplasia) of the nasal mucosa, occupational asthma, 19 reduced lung function, and altered immune response. Some studies suggest that long-term 20 exposure to formaldehyde can decrease the number of white blood cells, and possibly 21 lower platelet and hemoglobin, and other studies have shown that formaldehyde exposure 22 affects changes in the percentage of lymphocyte subsets. Higher rates of spontaneous 23 abortion and low birth weights have been reported among women occupationally exposed 24 to formaldehyde. Oral exposure is rare, but there have been several suicides and 25 attempted suicides where individuals drank formaldehyde. These data indicate that the 26 lethal dose is 60 to 90 mL. Formaldehyde ingestion results in severe corrosive damage to 27 the gastrointestinal tract followed by CNS depression, myocardial depression, circulatory 28 collapse, metabolic acidosis, and multiple organ failure.

1 The toxic effects of formaldehyde in experimental animals include irritation, cytotoxicity,

- 2 and cell proliferation in the upper respiratory tract, ocular irritation, pulmonary
- 3 hyperactivity, bronchoconstriction, gastrointestinal irritation, and skin sensitization.
- 4 Histopathological lesions of the upper respiratory tract and cell proliferation have not
- 5 been reported at concentrations less that 2 ppm. Other reported effects include oxidative
- 6 stress, neurotoxicity, immunotoxicity, testicular toxicity, and decreased liver, thyroid
- 7 gland, and testis weights.

#### 8 Carcinogenicity of metabolites and analogues

9 Formic acid (formate  $+ H^+$ ), the major metabolite of formaldehyde, has not been tested 10 for carcinogenic effects. Acetaldehyde, an analogue of formaldehyde, is listed as 11 reasonably anticipated to be a human carcinogen by the NTP. Acetaldehyde induced 12 respiratory tract tumors in rats (adenocarcinoma and squamous-cell carcinoma of the 13 nasal mucosa) and laryngeal carcinoma in hamsters. In addition, epidemiological data 14 provide some evidence that acetaldehyde may be associated with oral, esophageal, 15 pharyngeal, laryngeal, and bronchial tumors in humans. Glutaraldehyde and 16 benzaldehyde have also been tested for carcinogenicity in 2-year bioassays by the NTP. 17 Glutaraldehyde was not considered to be carcinogenic in rats and mice, and benzaldehyde 18 was not considered to be carcinogenic in rats. The NTP concluded that there was some 19 evidence of carcinogenicity for benzaldehyde in mice based on an increased incidence of 20 squamous-cell papillomas and hyperplasias in the forestomach of male and female mice.

21 Genetic and related effects

22 Formaldehyde is a direct-acting genotoxic compound that affects multiple gene 23 expression pathways, including those involved in DNA synthesis and repair and 24 regulation of cell proliferation. Most studies in bacteria were positive for forward or 25 reverse mutations without metabolic activation and for microsatellite induction. Studies 26 in non-mammalian eukaryotes and plants also were positive for forward and reverse 27 mutations, dominant lethal and sex-linked recessive lethal mutations, and DNA single-28 strand breaks. In vitro studies with mammalian and human cells were positive for DNA 29 adducts, DNA-protein crosslinks, unscheduled DNA synthesis, single-strand breaks, 30 mutations, and cytogeneic effects (chromosomal aberrations, sister chromatid exchange,

1 and micronuclei induction). In *in vivo* studies, formaldehyde caused DNA-protein cross 2 links (in the nasal mucosa and fetal liver but not bone marrow), DNA strand breaks 3 (lymphocytes and liver), dominant lethal mutations, chromosomal aberrations 4 (pulmonary lavage cells and bone marrow in one of two studies), and micronuclei 5 induction in the gastrointestinal tract; however it did not induce sister chromatid 6 exchange or chromosomal aberrations in lymphocytes. P53 mutations were detected in 7 nasal squamous-cell carcinomas from rats. Inhalation exposure of formaldehyde also 8 induced DNA-protein cross links in the nasal turbinates, nasopharynx, trachea, and 9 bronchi of rhesus monkeys. In mice, formaldehyde exposure did not cause dominant 10 lethal mutations, micronuclei induction, or chromosomal aberrations when exposed by 11 intraperitoneal injection, but did induced heritable mutations when exposed by inhalation.

In studies of lymphocytes humans exposed to formaldehyde, increased frequencies of chromosomal aberrations were observed in seven of twelve reviewed studies, sister chromatid aberrations in six of thirteen studies, and micronuclei induction in fifteen of sixteen studies reviewed. Increased frequencies of micronuclei were also observed in the buccal or oral epithelium, nasal epithelium in all but one of the available studies. DNAprotein cross links and DNA strand breaks have also been observed in lymphocytes from medical personnel exposed to formaldehyde.

#### 19 Mechanistic considerations

Although the biological mechanisms associated with formaldehyde-induced cancer are
not completely understood, it is important to recognize that chemicals can act through
multiple toxicity pathways and mechanisms to induce cancer or other health effects.
Potential carcinogenic modes of actions for formaldehyde include DNA reactivity
(covalent binding), gene mutation, chromosomal breakage, aneuploidy, and epigenetic
effects.

- 26 Studies evaluating nasal tumors in rats have shown that, regional dosimetry, genotoxicity,
- 27 and cytotoxicity are believed to be important factors. Computational fluid dynamics
- 28 models have been developed to predict and compare local flux values in the nasal
- 29 passages of rats, monkeys, and humans. Regions of the nasal passages with the highest

1 flux values are the regions most likely affected by formaldehyde exposure. Similar flux 2 values were predicted for rats and monkeys for regions of the nasal passages with 3 elevated cell proliferation rates, thus providing support for the hypothesis that 4 formaldehyde flux is a key factor for determining toxic response. Furthermore, DNA-5 protein crosslinks and cell-proliferation rates are correlated with the site specificity of 6 tumors. Cell proliferation is stimulated by the cytotoxic effects of formaldehyde. 7 Increased cell proliferation may contribute to carcinogenesis by increasing the probability 8 of spontaneous or chemically induced mutations. The dose-response curves for DNA-9 protein crosslinks, cell proliferation, and tumor formation show similar patterns with sharp increases in slope at concentrations greater than 6 ppm. The observed sequence of 10 11 nasal lesions is as follows: rhinitis, epithelial dysplasia, squamous metaplasia and 12 hyperplasia, and squamous-cell carcinoma.

13 Biological mechanisms have been proposed for the possible association between 14 lymphohematopoietic cancers and formaldehyde exposure. Proposed mechanisms for 15 formaldehyde-induced leukemia are: (1) direct damage to stem cells in the bone marrow, 16 (2) damage to circulating stem cells, (3) damage to pluripotent stem cells present in the 17 nasal turbinate or olfactory mucosa. Evidence in support of the potential for DNA 18 damage to circulating hematopoietic stem cells is that DNA-protein crosslinks have been 19 identified in the nasal passages of laboratory animals exposed to formaldehyde and 20 increased micronuclei have been identified in the nasal and oral mucosa of formaldehyde-21 exposed humans. In addition, olfactory epithelial cells obtained from rat nasal passages 22 contain hematopoietic stem cells, which have been shown to re-populate the 23 heamtopoietic tissue of irradiated rats. However, some authors have questioned the 24 biologically plausibility of an association between formaldehyde exposure and leukemia, 25 because formaldehyde is rapidly metabolized and would not enter the systemic 26 circulation. They state that formaldehyde does not cause bone marrow toxicity or 27 pancytopenia, which are common features of known leukemogen, and that the genotoxic 28 and carcinogenic effects in animals and humans are limited to local effects.

## Abbreviations

ACGIH:	American Conference of Governmental Industrial Hygienists
ADC:	adenocarcinoma
ADCN:	adenocarcinoma
ADH:	alcohol dehydrogenase
AGT:	$O^{6}$ -alkylguanine DNA alkyltransferase (also known as MGMT)
AIPH:	2,2'-azobis-[2-(2-imidazolin-2-yl)propane] dihydrochloride
ALDH:	aldehyde dehydrogenase
AML:	Acute myelogenous leukemia
ANOVA:	analysis of variance
AOPC:	all other pharyngeal cancers
ATSDR:	Agency for Toxic Substances and Disease Registry
b.w.:	body weight
BCF:	bioconcentration factor
BEAM:	Boston Exposure Assessment in Microenvironments
BEI:	biological exposure indices
BLS:	Bureau of Labor Statistics
BMCR:	binucleated micronucleated cell rate
BRCA1:	breast cancer 1, early onset gene
BrdUrd:	5-bromodeoxyuridine
C:	control
CA:	chromosomal aberrations
Cal/OSHA:	California Division of Occupational Safety and Health
CAS:	Chemical Abstracts Service
CBI:	covalent binding index

CC1b:	Clara-cell specific protein
CDC:	Centers for Disease Control and Prevention
CEH:	Chemical Economics Handbook
CFD:	computational fluid dynamics
CHO:	Chinese hamster ovary
CLL:	chronic lymphocytic leukemia
cm:	centimeter
CMBN:	cytokinesis-blocked micronucleus assay
CML:	chronic myeloid leukemia
CNS:	central nervous system
CPBI:	cytokinesis proliferation block index
CR:	creatinine
CYP:	cytochrome P450
Cyt-B:	cytochalasin B
Da:	Dalton
DC:	decarboxylase
dm:	decimeter
DNA:	deoxyribonucleic acid
DOT:	Department of Transportation
dpm:	disintegrations per minute
E.U.:	European Union
E:	exposed
EBV:	Epstein-Barr virus
EPA:	Environmental Protection Agency
EPHX:	epoxide hydrolase

ESTR:	expanded simple tandem repeats
ETS:	environmental tobacco smoke
F:	female
FDA:	Food and Drug Administration
FDH:	formaldehyde dehydrogenase
FEMA:	Federal Emergency Management Agency
FISH:	fluorescence in-situ hybridization
FR:	frequency ratios
g:	gram
GGT:	gamma-glutamyl transpeptidase
GI:	gastrointestinal
GPA:	glycophorin A
GSH:	glutathione
GSTM1:	glutathione S transferase M1
GSTT1:	glutathione S transferase T1
h:	hour
HA:	hydroxylapatite
HazDat:	Hazardous Substances Release and Health Effects Database
HCHO:	formaldehyde
HE:	human erythrocytes
HEL:	human embryonic lung
HFC:	high-frequency cells
Hg:	mercury
HIC:	highest ineffective concentration
HID:	highest ineffective dose

HMMECs:	human mucosal microvascular endothelial cells
HPLC:	high performance liquid chromatography
HR:	hazard ratio
HSA:	human serum albumin
HSDB:	Hazardous Substances Data Bank
Hz:	Hertz
i.p.:	intraperitoneal
IARC:	International Agency for Research on Cancer
ICAM:	intercellular adhesion molecule
ICD:	International Classification of Diseases
IFN:	interferon
IgG:	immunoglobin G
IgM:	immunoglobin M
IMIS:	Integrated Management Information System
IRR:	incidence rate ratio
IUPAC:	The International Union of Pure and Applied Chemistry
JEM:	job-exposure matrix
kBq:	1,000 becquerel (units of radioactivity)
kg:	kilogram
K <sub>oc</sub> :	soil organic carbon-water partitioning coefficient
K <sub>ow</sub> :	octanol-water partition coefficient
L:	liter
LC:	liquid chromatography
LD <sub>50</sub> :	lethal dose for 50% of the population
LEC:	lowest effective concentration

xxvi	Draft Background Document for Formaldehyde
LED:	lowest effective dose
LH:	lymphohematopoietic
LHC:	lymphohematopoietic cancer
LWAE:	lifetime weighted average exposure
M:	male or molar
m <sup>3</sup> :	cubic meter
MA:	mandelic acid
MAK:	maximum workplace concentration
MAPKs:	mitogen-activated protein kinases
mCi:	millicurries
MDF:	medium density fiberboard
MDS:	myelodysplastic syndrome
mEH:	microsomal epoxide hydrolase
MF:	melamine-formaldehyde
mg:	milligram, 10 <sup>-3</sup> gram
MGMT:	O <sup>6</sup> -methylguanine DNA methyltransferase (also known as AGT)
mL:	milliliter
mm:	millimeter
mM:	millimolar
MM:	multiple myeloma
MN:	micronuclei
mol wt:	molecular weight
mRNA:	messenger RNA
mRR:	meta relative risk
MS:	mass spectrometry

••
X X V11
AAVII

MTT:	methylthiazole tetrazolium
MUF:	melamine-urea-formaldehyde
N:	sample size
NA:	not available
NA-AAF:	N-acetoxy-2-acetylaminofluorene
NAcT:	N-acetyltransferase
NADPH:	nicotinamide adenine dinucleotide phosphate, reduced form
NALT:	nasal associated lymph tissue
NAP:	not applicable
NCEs:	micronucleated normochromatic erythrocytes
NCHS:	National Center for Health Statistics
NCI:	National Cancer Institute
ND:	not detected
NDMA:	N-nitrosodimethylamine
NDT:	not determined
NF-ĸB:	nuclear factor kappa B
ng:	nanogram
NGF:	nerve growth factor
NHANES:	National Health and Nutrition Examination Survey
NHL:	non-Hodgkin's lymphoma
NI:	not identified
NIEHS:	National Institute of Environmental Health Sciences
NIOSH:	National Institute for Occupational Safety and Health
NLM:	National Library of Medicine
NMR:	nuclear magnetic resonance

xxviii	Draft Background Document for Formaldehyde
NNK:	4-(N-nitrosomethylamino)-1-(3-pyridyl)-1-butanone
NOS:	not otherwise specified
NPC:	nasopharyngeal cancer
NQ:	not quantified
NR:	not reported
NRC:	National Response Center
NS:	not significant
NT:	not tested
NTP:	National Toxicology Program
OH:	hydroxyl
OHPC:	oro- or hypopharyngeal
OPC:	oropharyngeal
OR:	odds ratio
OSB:	oriented strandboard
OSHA:	Occupational Safety and Health Administration
OVA:	ovalbumin
PAH:	polycyclic aromatic hydrocarbon
PAMA:	phenacylmercapturic acid
PBL:	peripheral blood lymphocytes
PBPK:	physiologically based pharmacokinetic model
PCEs:	micronucleated polychromatic erythrocytes
PCMR:	proportionate cancer mortality ratio
PCR:	polymerase chain reaction
PEL:	permissible exposure limit
PF:	phenol-formaldehyde

PGA:	phenylglyoxylic acid
PHA:	phytohemagglutinin
PHEMA:	phenylhydroxyethyl mercapturic acids
PMR:	proportionate mortality ratio
ppb:	parts per billion
ppbv:	parts per billion by volume
ppm:	parts per million
<i>r</i> :	correlation coefficient
REL:	recommended exposure limit
RLU:	relative light units
RNA:	ribonucleic acid
RoC:	Report on Carcinogens
RR:	relative risk
RTECS:	Registry of Toxic Effects of Chemical Substances
s.c.:	subcutaneous
SCC:	squamous-cell carcinoma
SCE:	sister chromatid exchange
SD:	standard deviation
SDH:	sorbitol dehydrogenase
SE:	standard error of the mean
SEER:	Surveillance, Epidemiology and End Results program
SIR:	standardized incidence ratio
SMR:	standardized mortality ratio
SNC:	sinonasal
SOC:	Standard Occupational Classification

SOCMI:	Synthetic Organic Chemical Manufacturing Industry
SPICR:	standardized proportionate incidence cancer ratio
SSB:	single-strand breaks
STEL:	short-term exposure limit
TLV:	threshold-limit value
TRI:	Toxics Release Inventory
TSH:	thyroid stimulating hormone
TWA:	time-weighted average
UDS:	unscheduled DNA synthesis
UF:	urea-formaldehyde
UFFI:	urea-formaldehyde foam insulation
USITC:	United States International Trade Commission
VCAM:	vascular cell adhesion molecule
VOC:	volatile organic chemical
VPT:	vinylphenol
WHO:	World Health Organization
XO:	xanthine oxidase
XPC:	xeroderma pigmentosum, complementation group C
XPD:	xeroderma pigmentosum, complementation group D
XPG:	xeroderma pigmentosum, complementation group G
XRCC:	X-ray repair cross-complementing group
yr:	year
γ-GT:	gammaglutamyl transpeptidase
μg:	microgram; 10 <sup>-6</sup> gram

## **Table of Contents**

1	Introduction1			1	
	1.1 Chemical identification			2	
	1.2	Physical-chemical properties			
	1.3	Formale	lehyde Polymers	4	
	1.4	Metabo	5		
2	Human Exposure				
	2.1	Use			
	2.2	Product	ion		
		2.2.1	Industrial production		
		2.2.2	Other production sources	15	
		2.2.3	Endogenous production	15	
	2.3 Biological indices of exposure			16	
	2.4	Occupational exposure		19	
		2.4.1	Formaldehyde and formaldehyde-based resin production		
		2.4.2	Wood-based products and paper production		
		2.4.3	Manufacture of textiles and garments		
		2.4.4	Foundries		
		2.4.5	Production of formaldehyde-based plastic products	40	
		2.4.6	Embalming		
		2.4.7	Histology	47	
		2.4.8	Construction-related exposures	49	
		2.4.9	Fiberglass and mineral-wool insulation manufacturing	51	
		2.4.10	Firefighting and other combustion-related exposures	52	
		2.4.11	Agriculture and aquaculture	55	
		2.4.12	Office buildings and nonindustrial work places	56	
		2.4.13	Other occupational exposures	58	
	2.5	Environ	mental occurrence and fate	59	
		2.5.1	Air	60	
		2.5.2	Water	77	
		2.5.3	Land and soil	83	
		2.5.4	Food	83	
	2.6	5 Exposure estimates			
	2.7	2.7 Regulations and Guidelines			
		2.7.1	Regulations		
		2.7.2	Guidelines		
	2.8 Summary				
3	Human	Cancer S	tudies		
	3.1	Description of head and neck cancers			
	3.2	Cohort standardized and proportionate mortality and incidence studies			

		3.2.1	National Cancer Institute (NCI) Cohort: mixed industries	. 103
		3.2.2	National Institute for Occupational Safety and Health (NIOSH)	
			cohort: garment industry	. 116
		3.2.3	British Chemical Workers Study	. 118
		3.2.4	Studies of fiberglass workers	. 120
		3.2.5	Studies of woodworking and related industries	. 124
		3.2.6	Miscellaneous studies: abrasive material manufacturing, Iron	
			foundry, mixed industry and chrome leather tannery workers	. 129
		3.2.7	Studies of resin, chemical, and plastics manufacturing workers	. 133
		3.2.8	Studies of health professionals, embalmers, and funeral directors	. 137
	3.3	Case-coi	ntrol studies	. 145
		3.3.1	Cancers of the paranasal sinuses and nasal cavity	. 146
		3.3.2	Cancer of the nasopharynx	. 155
		3.3.3	Other head and neck cancers	. 164
		3.3.4	Lung cancer	. 173
		3.3.5	Lymphohematopoietic malignancies	. 178
		3.3.6	Cancers at other sites	. 185
	3.4	Summar	y by tumor site	. 191
		3.4.1	Cancers of the paranasal sinuses and nasal cavity	. 192
		3.4.2	Cancer of the nasopharynx	. 203
		3.4.3	Other head and neck cancers	. 215
		3.4.4	Respiratory cancers or lung cancer	. 229
		3.4.5	Lymphohematopoietic cancers	. 239
		3.4.6	Cancers of the brain and central nervous system	. 256
		3.4.7	Cancer at other sites	. 260
	3.5	Summar	у	. 264
		3.5.1	Sinonasal cancers	. 265
		3.5.2	Nasopharyngeal cancers	. 266
		3.5.3	Other head and neck cancers, and respiratory cancer	. 267
		3.5.4	Lymphohematopoietic cancers	. 269
		3.5.5	Other cancer sites	. 272
4	Studies	of Cancer	in Experimental Animals	. 274
	4.1	Inhalatic	on	. 274
		4.1.1	Mice	. 275
		4.1.2	Rats	. 277
		4.1.3	Hamsters	. 288
		4.1.4	Monkeys	. 289
		4.1.5	Summary of inhalation studies	. 290
	4.2	Oral and	dermal administration	. 294
		4.2.1	Drinking-water studies	. 294
		4.2.2	Skin application	. 302

		4.2.3	Summary of oral and dermal exposure studies		
	4.3	Co-exp	bosure with other substances		
		4.3.1	Mice		
		4.3.2	Rats		
		4.3.3	Hamsters		
		4.3.4	Summary of promotion and cocarcinogenicity studies		
	4.4	Summa	ary		
5	Other Relevant Data				
	5.1	Absorption, distribution, and excretion			
		5.1.1	In vitro studies		
		5.1.2	In vivo studies		
	5.2	Airway	v deposition models		
	5.3	Metabo	olism		
	5.4	Toxic e	effects		
		5.4.1	In vitro toxicity studies		
		5.4.2	Toxic effects in humans		
		5.4.3	Toxic effects in experimental animals		
	5.5	Carcine	ogenicity studies of metabolites and analogues		
	5.6	Genetie	c and related effects		
		5.6.1	Prokaryotes		
		5.6.2	Non-mammalian eukaryotes		
		5.6.3	Mammalian systems		
		5.6.4	Human in vivo studies		
		5.6.5	Gene expression		
	5.7	Mechanistic considerations			
		5.7.1	Genotoxicity		
		5.7.2	Glutathione depletion and oxidative stress		
		5.7.3	Mutational spectra		
		5.7.4	Epigenetic effects		
		5.7.5	Nasal tumors		
		5.7.6	Other tumors		
	5.8	Summary			
		5.8.1	Adsorption, distribution, metabolism, and excretion		
		5.8.2	Toxic effects		
		5.8.3	Carcinogenicity of metabolites and analogues		
		5.8.4	Genetic and related effects		
		5.8.5	Mechanistic considerations		
6	Reference	ces			
Gl	ossary of	Terms			

### List of Tables

Table 1-1. Chemical identification of formaldehyde	2
Table 1-2. Physical and chemical properties of formaldehyde	
Table 1-3. Chemical identification and physical and chemical properties of paraformaldehyde and trioxane	5
Table 1-4. Some low-molecular weight formaldehyde analogues	6
Table 2-1. Formaldehyde exposure levels associated with formaldehyde production and formaldehyde-based resin production	
Table 2-2. Formaldehyde exposure levels associated with the production of wood-based composites.	27
Table 2-3. Formaldehyde exposure levels associated with the manufacture of plywood and laminates	
Table 2-4. Formaldehyde exposure levels associated with wood furniture manufacturing	32
Table 2-5. Formaldehyde exposure levels associated with the manufacture of paper and paper products	34
Table 2-6. Formaldehyde exposure levels associated with the textile and garment industries	
Table 2-7. Formaldehyde exposure levels associated with foundries	40
Table 2-8. Formaldehyde exposure levels associated with production of plastics and plastic products	42
Table 2-9. Formaldehyde exposure levels associated with embalming or autopsies or in anatomy laboratories	45
Table 2-10. Formaldehyde exposure levels associated with histology and pathology           laboratories	49
Table 2-11. Formaldehyde levels associated with construction-related activities	51
Table 2-12. Formaldehyde exposure levels associated with fiberglass manufacturing	52
Table 2-13. Formaldehyde exposure levels associated with firefighting and other combustion sources	54
Table 2-14. Formaldehyde exposure levels associated with agriculture and aquaculture	56
Table 2-15. Formaldehyde exposure levels in offices and other nonindustrial work places	5 57
Table 2-16. Occurrence of formaldehyde in outdoor air in the United States	65
Table 2-17. Formaldehyde off-gassing emission rates from building materials, home furnishings, and consumer products	68
Table 2-18. Occurrence of formaldehyde in U.S. residential indoor air	
Table 2-19. Formaldehyde levels associated with cigarette smoke	76
Table 2-20. Formaldehyde concentrations in drinking water	79
Table 2-21. Formaldehyde levels in U.S. environmental water	82
Table 2-22. Formaldehyde levels in food	
Table 2-23. Estimated formaldehyde exposure levels    87	7
--	---
Table 3-1. Summary of cohort studies and nested case-control studies	3
Table 3-2. Lymphohematopoietic (LH) cancers in formaldehyde-exposed workers (NCI cohort and peak exposure: 1994 and 2004 updates)	3
Table 3-3a. Summary of cohort studies of formaldehyde exposure and cancer of the sinus and nasal cavities (SNC)	1
Table 3-3b. Summary of case-control studies investigating formaldehyde exposure and sinonasal cancer       199	)
Table 3-4a. Summary of cohort studies of formaldehyde exposure and nasopharyngeal cancers	3
Table 3-4b. Summary of case-control studies (including nested case-control studies) and cancer registry studies of formaldehyde exposure and nasopharyngeal cancer.         211	L
Table 3-5a. Summary of cohort studies of formaldehyde exposure and cancers of the oral cavity, pharynx, and larynx	)
Table 3-5b. Summary of case-control studies (including nested case-control studies) and cancer registry studies of formaldehyde exposure and cancers of the oral cavity, pharynx, and larynx	3
Table 3-6a. Summary of cohort studies of formaldehyde exposure and cancers of the lung 232	)
Table 3-6b. Summary of case-control studies (including nested case-control)         investigating formaldehyde exposure and lung or respiratory cancer	5
Table 3-7a. Summary of cohort studies of formaldehyde exposure and         lymphohematopoietic cancers	5
Table 3-7b. Summary of case-control studies (including nested case-control)         investigating formaldehyde exposure and lymphohematopoietic cancers	L
Table 3-8. Summary of industrial SMR and PMR studies of formaldehyde exposure and         Brain and CNS cancers         258	3
Table 4-1. Histologic changes in the lungs of C3H mice exposed to formaldehyde by         inhalation for up to 35 weeks	5
Table 4-2. Neoplastic responses in the nasal cavity of male Wistar rats exposed toformaldehyde by inhalation for 4 to 13 weeks	)
Table 4-3. Nasal tumors in F344 rats exposed to formaldehyde by inhalation for up to 24 months	L
Table 4-4. Apparent sites of origin of squamous-cell carcinomas in the nasal passages ofF344 rats exposed to formaldehyde by inhalation for up to 24 months	2
Table 4-5. Neoplastic responses in the nasal cavity of male albino Wistar rats, with and without damaged nasal mucosa, exposed to formaldehyde by inhalation for 3 or 28 months	Ļ
Table 4-6. Neoplastic responses in the nasal cavity of male F344 rats exposed toformaldehyde by inhalation for up to 24 months	,

Table 4-7. Proliferative lesions and neoplastic responses in the nasal cavity of male F344         rats exposed to formaldehyde by inhalation for up to 28 months	288
Table 4-8 Summary of inhalation studies of formaldehyde in experimental animals	291
Table 4-9. Non-neoplastic responses in Wistar rats given formaldehyde in drinking water         for 24 months	295
Table 4-10. Tumor incidences in Sprague-Dawley rats exposed to formaldehyde indrinking water at two different ages for up to 104 weeks	297
Table 4-11. Total malignant tumors in Sprague-Dawley rats exposed to formaldehyde in drinking water for up to 104 weeks	298
Table 4-12a. Incidences of mammary, testicular, and hemolymphoreticular tumors in Sprague-Dawley rats exposed to formaldehyde in drinking water for up to 104 weeks	300
Table 4-12b. Incidences of stomach and intestinal tumors in Sprague-Dawley rats         exposed to formaldehyde in drinking water for up to 104 weeks	301
Table 4-13. Summary of oral and dermal carcinogenicity studies of formaldehyde in experimental animals	303
Table 4-14. Incidences of squamous-cell lung tumors in C3H mice exposed to formaldehyde and coal tar by inhalation	304
Table 4-15. Skin tumor promotion study of formaldehyde in Oslo hairless mice	306
Table 4-16. Proliferative and neoplastic lesions in the nasal cavity of male Sprague-         Dawley rats exposed to formaldehyde and hydrogen chloride	309
Table 4-17. Effects of formaldehyde on gastric carcinogenesis in male Wistar rats         initiated with MNNG	311
Table 4-18. Effects of formaldehyde on induction of respiratory-tract tumors by DEN in male Syrian hamsters	313
Table 4-19. Co-exposure carcinogenicity studies of formaldehyde and other substances in experimental animals	314
Table 4-20. Summary of neoplasms associated with formaldehyde exposure in experimental animals	317
Table 5-1. Disposition of inhaled $^{14}$ C-formaldehyde in male F344 rats (% radioactivity ± SD)	321
Table 5-2. Concentrations of formaldehyde in human blood before and after exposure to         1.9 ppm for 40 minutes	323
Table 5-3. Formaldehyde and formic acid concentrations detected in body fluids and tissues following formaldehyde ingestion	326
Table 5-4. Distribution of <sup>14</sup> C-labelled formaldehyde in rodents and monkeys during the first 72 h after topical administration	328
Table 5-5. Formaldehyde concentrations associated with various health effects	338
Table 5-6. Irritant effects of formaldehyde following acute inhalation exposures	339
Table 5-7. Effects on the nasal mucosa from chronic exposure to formaldehyde	343

Table 5.9. Studies of accurational asthma and formal debude avacuum	245
Table 5-8. Studies of occupational astima and formaldenyde exposure	545
Table 5-9. Effects of formaldehyde exposure on peripheral lymphocyte subsets in anatomy students	352
Table 5-10. Summary of blood cell counts in Chinese workers with formaldehyde exposure reported by Tang <i>et al.</i> (2000)	356
Exposure reported by range $er ar. (2009)$	350
Table 5-11. Reproductive effects of formaldehyde in humans	358
Table 5-12. Seminiferous tubular diameter and height in Wistar rats	369
Table 5-13. Mean seminiferous tubular diameters and testosterone serum levels after 13-         week exposure to formaldehyde by inhalation in rats	370
Table 5-14. In vivo effect of formaldehyde on spermatozoa	371
Table 5-15. Genetic effects of formaldehyde in bacteria	375
Table 5-16. Genetic effects of formaldehyde in non-mammalian eukaryotes	376
Table 5-17. In vitro studies of DNA adducts, DNA-protein crosslinks and strand breaks         in mammalian systems	380
Table 5-18. In vivo studies of DNA-protein crosslinks and strand breaks in mammalian           systems	385
Table 5-19. In vitro studies of cytogenetic effects of formaldehyde in mammalian cells	389
Table 5-20. Cytogenetic effects of formaldehyde in mammals	391
Table 5-21. Mutagenic effects of formaldehyde in mammalian systems	392
Table 5-22. Other genetic effects of formaldehyde in mammalian systems	393
Table 5-23. DNA-protein crosslinks and pantropic p53 protein levels in medical workers         exposed to formaldehyde	395
Table 5-24. Distribution of autopsy service and pathology department workers with mutagenic or toxic urine samples	399
Table 5-25. Chromosomal aberrations in peripheral blood lymphocytes from humans         exposed to formaldehyde	405
Table 5-26. Sister chromatid exchange in peripheral blood lymphocytes from humans         exposed to formaldehyde	412
Table 5-27. Micronuclei in various cell types from humans exposed to formaldehyde	420
Table 5-28. Formaldehyde exposure, DNA-protein crosslinks, and nasal tumor incidence	428
Table 5-29. Formaldehyde exposure, cell proliferation, and nasal tumor incidence	445

## List of Figures

Figure 1-1. Chemical structure of formaldehyde	2
Figure 1-2. Chemical structures of hydrated and polymeric formaldehyde	4
Figure 2-1. Major uses of formaldehyde in the United States	. 12
Figure 2-2. Formaldehyde production in the United States	. 14

Figure 3-1. Upper respiratory system	96
Figure 4-1. Midsagittal section of the rat nose showing the anatomical levels typically	
examined in inhalation studies	275
Figure 5-1. Metabolism and fate of formaldehyde	334
Figure 5-2. Biological reactions of formaldehyde	335
Figure 5-3. Sagital (A) and cross-section (B) through the rat nose.	441

1

## 1 1 Introduction

2 Formaldehyde is a high-production-volume chemical with a wide array of uses. The 3 predominant use of formaldehyde in the United States is in the production of industrial 4 resins (mainly urea-formaldehyde [UF], phenol-formaldehyde [PF], polyacetal, and 5 melamine-formaldehyde [MF] resins) that are used to manufacture products such as 6 adhesives and binders for wood products, pulp and paper products, plastics, and synthetic 7 fibers, and in textile finishing. Formaldehyde is also used as a chemical intermediate. 8 Resin production and use as a chemical intermediate together account for over 80% of its 9 use. Other, smaller uses of formaldehyde that may be important for potential human 10 exposure include use in agriculture, medical use as a disinfectant and preservative (for 11 pathology, histology, and embalming), and use in numerous consumer products as a 12 biocide and preservative.

13 Formaldehyde is present in outdoor air as a result of its formation from the combustion of 14 organic materials (e.g., in automobiles, forest fires, and power plants), its formation from 15 the breakdown of hydrocarbons in the air, and releases from industrial facilities. In indoor 16 air, it is present as a result of off-gassing from formaldehyde-containing materials such as 17 wood products, carpets, fabrics, paint, and insulation, and it is formed from combustion 18 sources such as wood stoves, gas stoves, kerosene heaters, open fireplaces, and furnaces, 19 through cooking, and in cigarette smoke. It has been found in numerous foods and 20 beverages, including drinking water.

21 Formaldehyde (gas) is listed in the Eleventh Report on Carcinogens (RoC) as reasonably 22 anticipated to be a human carcinogen based on limited evidence of carcinogenicity in 23 humans and sufficient evidence of carcinogenicity in laboratory animals (NTP 2005a); it 24 was first listed in the 2nd RoC (NTP 1981). Formaldehyde (all physical forms) was 25 nominated by NIEHS for possible reclassification in the 12th RoC based on the 2004 26 review by the International Agency for Research on Cancer (IARC 2006), which 27 concluded that there was sufficient evidence for the carcinogenicity of formaldehyde in 28 humans.

## 1 **1.1 Chemical identification**

- 2 Formaldehyde is the simplest aldehyde. It is a highly reactive gas and is formed by
- 3 oxidation or incomplete combustion of hydrocarbons (ChemIDPlus 2009a). Figure 1-1
- 4 shows the chemical structure of formaldehyde, and Table 1-1 provides some chemical
- 5 identifying information.



## Figure 1-1. Chemical structure of formaldehyde

- 6 Commercially, formaldehyde is most often available as 30% to 50% (by weight) aqueous
- 7 solutions commonly referred to as formalin (IARC 2006), to which have been added
- 8 stabilizers, generally up to 15% methanol or lower concentrations (usually several
- 9 hundred milligrams per liter) of various amine derivatives. In the absence of stabilizers,
- 10 formaldehyde in solution oxidizes slowly to form formic acid and polymerizes to form
- 11 oligomers, including paraformaldehyde (HSDB 2009a).

Table 1-1. Chemical identification of formaldehyde

Characteristic	Information	References
CAS Registry number	50-00-0	HSDB 2009a
IUPAC systematic name	methanal	IARC 2006
Molecular formula	CH <sub>2</sub> O	HSDB 2009a
Synonyms	Fannoform, Formalith, formalin, formic aldehyde, Lysoform, methanal, methyl aldehyde, methylene oxide, Morbicid, oxomethane, oxymethylene, Superlysoform	HSDB 2009a

## 12 **1.2** Physical-chemical properties

Formaldehyde exists at room temperature as a flammable, nearly colorless gas with a pungent, suffocating odor (ATSDR 1999, HSDB 2009a). Formaldehyde gas is generally stable in the absence of water, but it is flammable and can be ignited by heat, sparks, or flame. Vapors form explosive mixtures with air. Formaldehyde gas reacts violently with strong oxidizing agents and with bases and reacts explosively with nitrogen dioxide at

2

- 1 around 180°C (Akron 2009). It reacts with hydrochloric acid to form bis(chloromethyl)
- 2 ether (which is listed in the RoC as *known to be a human carcinogen*). In its pure state,
- 3 formaldehyde is not easily handled, because it is extremely reactive and polymerizes
- 4 readily.
- 5 The physical and chemical properties of formaldehyde are summarized in Table 1-2.

Property	Information	References
Molecular weight	30.0	HSDB 2009a
Melting point (°C)	-92	HSDB 2009a
Boiling point (°C)	-19.5	HSDB 2009a
Specific gravity	0.815 at -20°C/4°C	O'Neil et al. 2006
Vapor pressure (mm Hg)	3,890 at 25°C	HSDB 2009a
Vapor density	1.067 (air = 1)	HSDB 2009a
Critical temperature (°C)	137.2 to 141.2	HSDB 2009a
Solubility		HSDB 2009a
water at 20°C	400 g/L	
acetone, alcohol, benzene, ether	soluble	
Octanol-water partition coefficient	0.35	HSDB 2009a
(log K <sub>ow</sub> )		
Dissociation constant (pK <sub>a</sub> )	13.27 at 25°C	HSDB 2009a
Henry's law constant	$3.4 \times 10^{-7} \text{ atm-m}^3/\text{mol}$	HSDB 2009a
Unit conversion (air concentrations)	$mg/m^3 = 1.23 \times ppm$	IARC 2006

Table 1-2. Physical and chemical properties of formaldehyde

- 6 The primary form of formaldehyde in dilute aqueous solutions is its monomeric hydrate,
- 7 methylene glycol (Figure 1-2), and the primary forms in concentrated solutions are
- 8 oligomers and polymers of polyoxymethylene glycols (IARC 2006). Formaldehyde can
- 9 also exist as paraformaldehyde, a polymer with 8 to 100 units of formaldehyde, and as
- 10 1,3,5-trioxane, a cyclic trimer (Figure 1-2).



#### Figure 1-2. Chemical structures of hydrated and polymeric formaldehyde

#### 1 **1.3 Formaldehyde Polymers**

- 2 Paraformaldehyde is a white crystalline powder with the odor of formaldehyde. It has the
- 3 molecular formula  $(CH_2O)_n$  and is a mixture of linear polyoxymethylene glycols
- 4 containing 90% to 99% formaldehyde (HSDB 2009b, O'Neil et al. 2006).
- 5 Paraformaldehyde dissolves slowly in cold water and more readily in hot water, with
- 6 evolution to formaldehyde. It is soluble in fixed alkali hydroxide solution, but insoluble
- 7 in alcohol and ether. Paraformaldehyde is used as an engineering plastic because it has
- 8 good resistance to wear, chemicals, and temperature, a low coefficient of friction, and
- 9 good mechanical properties of strength and stiffness (Inventro 2009). Trioxane is a white
- 10 crystalline solid with a cholorform-like odor and the molecular formula (CH<sub>2</sub>O)<sub>3</sub> (HSDB
- 11 2009c). It is stable and easily handled. In acidic solutions, it will decompose to
- 12 formaldehyde. Both paraformaldeyde and trioxane are used as low-water-content sources
- 13 of formaldehyde. Table 1-3 shows chemical identifying information and some physical
- 14 and chemical properties of paraformaldehyde and trioxane.

Characteristic/Property	Paraformaldehyde	1,3,5-Trioxane	
CAS Registry number	30525-89-4	110-88-3	
Molecular formula	(CH <sub>2</sub> O) <sub>n</sub> <sup>a</sup>	C <sub>3</sub> H <sub>6</sub> O <sub>3</sub>	
Synonyms	Aldicide, Paraform, polyacetal, polyformaldehyde, polymethylene oxide, polyoxymethylene <sup>b</sup>	metaformaldehyde, <i>s</i> - trioxane, trioxymethylene	
Molecular weight	30.03 (monomer) <sup>a</sup>	90.08	
Melting point (°C)	164 (decomposes)	64	
Boiling point (°C)	slowly sublimes, forming formaldehyde gas <sup>c</sup>	114.5 @ 759 mm Hg	
Density	1.46 at 15°C	1.17 @ 65°C	
Vapor pressure (mm Hg)	10.5 at 25°C	NR	
Vapor density	1.03 <sup>c</sup>	3.1 <sup>d</sup>	
Water solubility at 18°C	$\begin{array}{c} 2\times10^5 \text{ mg/L} \\ 500 \text{ mg/L}^{\text{e,f}} \end{array}$	$1.7 \times 10^5 \text{ mg/L}$	
$\begin{array}{c} Octanol-water \ partition \ coefficient \\ (log \ K_{ow}) \end{array}$	NR	-0.43 <sup>g</sup>	
Dissociation constant (pK <sub>a</sub> )	15.50 at 25°C	NR	
Henry's law constant	NR	$1.97 \times 10^{-7g}$	

 Table 1-3. Chemical identification and physical and chemical properties of paraformaldehyde and trioxane

Source: HSDB 2009b,c unless otherwise noted. NR = not reported. <sup>a</sup>O'Neil *et al.* 2006. <sup>b</sup>PolymerProcessing 2009 and HSDB 2009b <sup>c</sup>Mallinckrodt 2009. <sup>d</sup>ScienceLab 2009a. <sup>e</sup>ScienceLab 2009b. <sup>f</sup>The higher-molecular-weight polymers are insoluble in water (ScienceLab 2009b). <sup>g</sup>ChemIDPlus 2009b.

### 1 **1.4 Metabolites and analogues**

- 2 Formaldehyde is an endogenous metabolic product of *N*-, *O*-, and *S*-demethylation
- 3 reactions and an essential metabolic intermediate in all cells (ATSDR 1999, Feick et al.
- 4 2006, IARC 2006). It is oxidized to formate, primarily by glutathione-dependent
- 5 formaldehyde dehydrogenase. Formate may be excreted in the urine, further metabolized
- 6 to carbon dioxide and water, or incorporated into the folic acid metabolic pathway for
- 7 synthesis of nucleic and amino acids. Further discussion of formaldehyde metabolism and
- 8 other biological reactions is provided in Section 5.2.

NOT FOR DISTRIBUTION OR ATTRIBUTION	

Draft Background Document for Formaldehyde	

- 1 Analogues of formaldehyde include other low-molecular-weight aldehydes, such as
- 2 acetaldehyde, propionaldehyde, butyraldehyde, *n*-pentanal, glutaraldehyde, and
- 3 benzaldehyde. The chemical structures and molecular weights of these compounds are
- 4 shown in Table 1-4, and carcinogenicity data for these analogues are discussed in
- 5 Section 5.4.

6

Compound	Molecular weight	Chemical structure
Acetaldehyde	44.1	О    H <sub>3</sub> С
Propionaldehyde	58.1	H <sub>3</sub> C CH
Butyraldehyde	72.1	H <sub>2</sub> H <sub>3</sub> C H <sub>2</sub> C H <sub>2</sub> C H <sub>2</sub> C H
<i>n</i> -Pentanal	86.1	$H_{3}C$ $H_{2}$ $C$ $H_{2}$ $C$ $H_{2}$ $H_{2}$ $C$ $H_{2}$ $C$ $H_{2}$ $C$ $H_{2}$ $C$ $H_{2}$ $C$ $C$ $H_{2}$ $C$
Glutaraldehyde	100.1	$HC \xrightarrow{C} H_2 \xrightarrow{C} H_2$
Benzaldehyde	106.1	СН

Table 1_/	Some low-mol	ocular waight	formoldohydo	analogues
1 avic 1-4.	Some tow-mor	cculal weight	101 maiuchyuc	analogues

## 1 2 Human Exposure

2 Formaldehyde is an important chemical with numerous industrial and commercial uses. 3 Annual U.S. industrial production in the early to mid 2000s averaged nearly 5 million 4 tons. In addition to intentional industrial production, formaldehyde is produced 5 unintentionally from human activities and from natural sources through the breakdown of 6 hydrocarbons and other precursors. Formaldehyde is also produced endogenously in 7 humans and other animals. Workers can be exposed to formaldehyde during its 8 production or during the production or use of derivative products. The general population 9 can be exposed to formaldehyde primarily from breathing indoor or outdoor air, from ingestion of food and water, from tobacco smoke, and from use of cosmetic products 10 11 containing formaldehyde. In the natural environment, formaldehyde has been detected in 12 indoor and outdoor air, surface water, rainwater, fog water, groundwater, soil, and food. 13 Numerous U.S. federal agencies, including the Environmental Protection Agency (EPA), 14 Food and Drug Administration (FDA), Department of Housing and Urban Development 15 (HUD), and Occupational Safety and Health Administration (OSHA), have enacted 16 regulations aimed at reducing formaldehyde exposures.

17 This section begins with a discussion of formaldehyde's various uses (Section 2.1).

18 Section 2.2 discusses industrial production of formaldehyde and formalin, natural sources

19 of formaldehyde, and endogenous production of formaldehyde in living organisms.

20 Section 2.3 discusses the issues surrounding biological indices of exposure to

21 formaldehyde. Occupational exposure levels are presented in Section 2.4 and

22 environmental levels in Section 2.5. Section 2.6 provides data from studies that have

estimated intake of formaldehyde by the general public from various sources. Section 2.7

24 provides regulations and guidelines that have been established with the intent of reducing

25 exposure. Section 2 concludes with a summary (Section 2.8).

26 Several organizations have prepared review articles on formaldehyde (e.g., IARC, WHO,

27 ATSDR); the most recent being a 2006 IARC monograph. These review articles have

- 28 been used extensively in this section for information for the period before 2006. In
- addition to the review articles, an extensive literature search was conducted as recently as

1 March 2009, and that literature was reviewed for inclusion. Throughout this section,

2 when data are cited from a review article, the primary citation is provided when

3 available.

4 The occupational epidemiology studies presented in Section 3 of this document include a

5 number of international studies; therefore, international occupational exposure data are

6 included in Section 2.4 (Occupational Exposure) in addition to U.S. data. For

7 environmental media, only U.S. levels are provided with the exception of levels that have

8 been measured in food and bottled water because a possibility of exposure to these

9 substances exists for the U.S. general public.

10 **2.1** Use

Formaldehyde has many and varied uses; however, its predominant use in the United States is in the production of industrial resins, accounting for over 50% of formaldehyde use in the early to mid 2000s (Bizzari 2007, ICIS 2007). Other major uses include as a chemical intermediate (~29%), various agricultural uses (~5%), paraformaldehyde production (~3%), production of chelating agents (~3%), and various minor uses (~5%) such as in the medical field, in funeral homes, in histology, and in numerous consumer products (see Figure 2-1).

18 The predominant formaldehyde-based industrial resins consumed in the United States are 19 urea-formaldehyde (UF) resins, accounting for 22% of the total formaldehyde consumed 20 in 2006 (Bizzari 2007). The largest use of UF resins is as a wood adhesive in the 21 manufacture of composite wood products, mainly particleboard and medium-density 22 fiberboard (MDF). Bizzarri (2007) reported that UF resins account for over 95% of the 23 adhesives used in manufactured particleboard and that 45% of U.S. UF consumption in 24 2006 was for particleboard manufacture. Wood adhesives made of UF resins are also 25 used to produce MDF, hardwood plywood, and other composite-wood products. UF 26 resins have also been used in the production of glass fiber roofing mats, as urea-27 formaldehyde foam for insulation (UFFI) in buildings, and in mining, where hollow areas

are filled with foam (ATSDR 1999).

1 Three other major resins are produced from formaldehyde: phenol-formaldehyde (PF) 2 resins, polyacetal resins, and melamine-formaldehyde (MF) resins. In the United States, 3 PF resins accounted for roughly 18%, polyacetal resins for nearly 12%, and MF resins for 4 roughly 3% of total formaldehyde consumption in 2006 (Bizzari 2007). Forecasts of U.S. 5 demand through 2011 show little change in these patterns. Demand for PF, MF, and 6 polyacetal resins is expected to grow between 0.1% and 3% annually through 2011, while 7 consumption of UF resins is expected to decline by approximately 0.3% annually, 8 primarily as a result of decreased particleboard production in the United States (Bizzari 9 2007).

10 Formaldehyde is also used as a chemical intermediate in the production of other

11 chemicals and products. In 2006, the predominant chemicals produced from

12 formaldehyde (based on the amount of formaldehyde consumed in production) were 1,4-

13 butanediol (10% of total U.S. consumption) and methylenebis(4-phenyl isocyanate) (11%

14 of total U.S. consumption) (Bizzari 2007). Formaldehyde is also used in the manufacture

15 of chelating agents (2.7% of total U.S. consumption in 2006), primarily in the

16 manufacture of ethylenediaminetetraacetic acid (EDTA) (57%), diethylenetriamine

17 pentaacetic acid (DTPA) (20%), hydroxyethylethylenediaminetriacetic acid (HEDTA)

18 (7%), and nitrilotriacetic acid (NTA) (16%) (Bizzari 2007).

19 Formaldehyde has many other varied uses that account for a small percentage of its total

20 consumption. It has been used as a disinfectant in hospital wards and operating rooms

and is used as a tissue preservative and disinfectant in embalming fluids (ATSDR 1999,

22 Dascalaki *et al.* 2008, IARC 2006). It is used as an antimicrobial in many cosmetic

23 products, at reported levels of up to 0.5% in lotions, creme rinses, and bubble-bath oils,

and up to 4.5% in nail hardeners. Other cosmetic products that may contain formaldehyde

25 include suntan lotions, hand creams, bath products, mascara and eye make-up, cuticle

26 softeners, nail creams, vaginal deodorants, shaving creams, soaps, shampoos, hair

27 preparations, deodorants, and mouthwashes. The Agency for Toxic Substances and

28 Disease Registry (ATSDR 1999) also noted that trace levels of formaldehyde may exist

29 in cosmetic products as a result of its use as a disinfectant for the equipment used to

30 manufacture the product. Formaldehyde has been used as a preservative in many

1 consumer goods, including household cleaning agents, dishwashing liquids, fabric 2 softeners, shoe-care agents, car shampoos and waxes, and carpet-cleaning agents; these 3 products generally contain less than 1% formaldehyde. It has been found in moist toilet 4 tissues for babies at levels exceeding 100  $\mu$ g/g (100 ppm) (WHO 2002). It also has been 5 added to fingerpaint as a preservative and has been measured at levels of 441 to 6 793 mg/kg in two types of fingerpaints; formaldehyde was undetectable (limit of 7 detection = 189 ng) in two other types (Garrigós et al. 2001). It has been used in pet-care 8 products at levels less than 0.5% and in various glues, epoxies, and adhesives intended 9 for household use at levels up to 9% (HPD 2009).

10 In the food industry, formaldehyde has been used for preserving dried foods, disinfecting 11 containers, preserving fish and certain oils and fats, and modifying starch for cold 12 swelling (ATSDR 1999). Formaldehyde has been used as a bacteriostatic agent in cheese 13 and other foods and in juice production, and paraformaldehyde has been implanted into 14 maple syrup tap holes to deter bacterial growth. Formaldehyde has been used as a 15 chemical germicide to control bacterial contamination in water distribution systems 16 (IARC 2006). It has also been used in the animal feed industry as a preservative and to 17 improve handling characteristics of feed (WHO 2002).

18 Although formaldehyde has many medical uses, consumption of formaldehyde in this 19 industry is relatively small, reflecting only about 1.5% of total U.S. volume in the late 20 1980s (ATSDR 1999). Formaldehyde is used as an antibacterial agent delivered via 21 hydrolysis of formaldehyde-releasing prodrugs, such as methenamine, used to treat 22 urinary-tract infections (FDA 2006, MedScape 2006). Rectal instillation, topical 23 application, and other techniques for administration of formalin solutions (typically 4% 24 formalin) have been used to treat radiation proctitis (Haas et al. 2007, Leiper and Morris 25 2007). The synergy between doxorubicin and formaldehyde-releasing prodrugs in killing 26 cancer cells has been shown to be due predominantly to formaldehyde (Rephaeli et al. 27 2007). Rephaeli *et al.* reported that these prodrugs also protected neonatal rat 28 cardiomyocytes and adult mice against the toxicity of doxorubicin.

11

1 Other reported minor medicinal applications for formaldehyde have included its use

2 during vasectomies, as a treatment for athlete's foot, as a sterilant for *Echinococcus* 

3 (tapeworm) cysts prior to their surgical removal, and in dentistry (IARC 1982, 2006).

4 Formaldehyde has had many uses in agriculture, including use as a fumigant, for 5 prevention of mildew in spelt wheat and rot in oats, as a preservative in fodder, as a 6 preplanting soil sterilant in mushroom houses, as a germicide and fungicide for plants 7 and vegetables, as an insecticide for flies and other insects, as a disinfectant in brooding 8 houses, in the production of herbicides, for seed treatment, and in the manufacture of 9 controlled-release fertilizers (used in agriculture and on residential lawns) (ATSDR 1999, 10 WHO 2002). Formaldehyde is also used to produce glyphosate, which is the active 11 ingredient in the herbicide Roundup (Bizzari 2007).

12 Additional uses of formaldehyde have been reported for the manufacture of glass mirrors, 13 explosives, artificial silk, and dyes; as a bactericide in coating agents and other chemicals 14 used in paper mills; for tanning and preserving animal hides; for hardening gelatin plates 15 and papers, toning gelatin-chloride papers, and chrome printing and developing in the 16 photography industry; as a biocide for latex, an adhesive additive, and an anti-oxidizer 17 additive for synthetic rubber in the rubber industry; as a biocide in oil-well drilling fluids 18 and as an auxiliary agent in petroleum refining; in chemical toilets; in the manufacture of 19 crease-resistant and flame-retardant fabrics; as an anticorrosive agent for metals; and in 20 formaldehyde-based resins often used as core binders in foundries (ATSDR 1999, WHO 21 2002).



Figure 2-1. Major uses of formaldehyde in the United States

Resins = UF, PF, MF and polyacetal resins; Chemical Intermediates = 1,4-butanediol, methylenebis(4phenyl isocyanate), pentaerythritol, hexamethylenetetramine, trimethylolpropane; Agriculture = controlledrelease fertilizers and herbicides; Chelating Agents = EDTA, DTPA, HEDTA, and NTA Source: Bizzari 2007

- 1 Because formaldehyde is fairly easy to make, is costly to transport, and can become
- 2 unstable during transport, it usually is produced to satisfy captive requirements for the
- 3 production of derivatives or to supply local merchant sales (Bizzari 2007). The uses for
- 4 formaldehyde vary regionally within the United States. Almost all formaldehyde
- 5 produced in the West is consumed for wood adhesives; formaldehyde produced in the
- 6 Gulf region is used primarily in chemical derivatives and to a lesser extent for wood
- 7 adhesives; and production in the South and Southeast is used primarily for wood
- 8 adhesives and to a lesser extent in chemical derivatives.
- 9 Paraformaldehyde is a high-formaldehyde-content product that is commercially available
- 10 as 91% or 95% prills; roughly 2.6 metric tons of 37% formaldehyde are required to
- 11 produce 1 metric ton of paraformaldehyde (Bizzari 2007). The main applications for
- 12 paraformaldehyde are foundry resins and applications where the presence of water could

interfere with a production process. Being a solid, paraformaldehyde is preferred over
 aqueous formaldehyde for shipping over long distances (Bizzari 2007).

3 Paraformaldehyde has been used as a fumigant to decontaminate laboratories and to

4 disinfect sickrooms, clothing, and linen; in pesticide applications; for making varnish

5 resins, thermosets, and foundry resins; in the synthesis of chemical and pharmaceutical

6 products; in the preparation of disinfectants and deodorants; and in the production of

7 textile products. In 2006, the production of paraformaldehyde accounted for almost 3%

8 of U.S. formaldehyde consumption (Bizzari 2007, EPA 2007).

9 Formaldehyde is also marketed in solid form as its cyclic trimer, trioxane (Bizzari 2007).

10 In acidic solutions, trioxane decomposes to generate three formaldehyde molecules

11 (HSDB 2009c). Trioxane and hexamine ( $C_6H_{12}N_4$ ) are the main components of solid fuel

12 tablets, commonly known as Esbit, which are used by campers, hobbyists, the military,

13 and relief organizations primarily for boiling water and cooking (ZenStoves 2009).

14 Trioxane is also used in the production of polyacetal resins (Bizzari 2007) and has many

15 other potential industrial applications (BASF 2006).

16 Some preservatives break down and release formaldehyde as the active agent (WHO

17 2002). The levels of decomposition and formaldehyde release depend mainly on

18 temperature and pH. Products most often containing formaldehyde releasers are industrial

19 and household cleaning agents, soaps, shampoos, paints, lacquers, and cutting fluids,

20 based on a review of the Danish Product Register Data Base (WHO 2002). Examples of

21 formaldehyde-releasing antimicrobial agents used in metalworking fluids are

22 tris(hydroxymethyl)nitromethane and hexahydro-1,3,4, tris(2-hydroxyethyl)-S-triazine

23 (NIOSH 2001). No data were found on formaldehyde levels resulting from formaldehyde

24 releasers.

## 25 2.2 Production

## 26 2.2.1 Industrial production

27 Formaldehyde has been produced commercially since 1889 by catalytic oxidation of

28 methanol. Currently, the two predominant production processes are a silver catalyst

29 process and a metal oxide catalyst process (Bizzari 2007).

Formaldehyde is produced and consumed at various concentrations; the data on industrial 1 2 levels presented here are based on a concentration of 37% unless otherwise noted. In 3 2006, worldwide formaldehyde production was around 28 million metric tons [31 million 4 tons], with Western Europe being the highest producer, at 7.8 million metric tons 5 [8.6 million tons], and China the second-highest producer, at 7 million metric tons 6 [7.7 million tons] (Bizzari 2007). In the United States, production has gradually but 7 steadily increased from 0.9 million metric tons [1 million tons] in 1960 to 4.5 million 8 metric tons [5 million tons] in 2006. Figure 2-2 shows U.S. formaldehyde production 9 from 1960 through 2006. Bizzari reported in 2007 that U.S. formaldehyde production





Figure 2-2. Formaldehyde production in the United States Source: Bizzari 2007

- 11 In the United States in 2009, formaldehyde was reported to be produced at 39
- 12 manufacturing plants (SRI 2009a) by an estimated 12 companies [estimate based on
- 13 Bizzari 2007], and paraformaldehyde and trioxane were each produced at one U.S.
- 14 manufacturing facility (SRI 2009b, 2009c). In 2009, 36 suppliers of formaldehyde, 25

1 suppliers of paraformaldehyde, and 11 suppliers of trioxane were identified in the United

2 States; identified internationally were 152 formaldehyde suppliers in 25 countries, 59

3 paraformaldehyde suppliers in 15 countries, and 21 trioxane suppliers in 9 countries

4 (ChemSources 2009a, 2009b, 2009c).

Because of transportation and storage issues associated with formaldehyde, it usually is
produced close to the point of consumption; international trade in formaldehyde is
therefore minimal, accounting for approximately 2% of worldwide production in 2006
(Bizzari 2007). In the United States, formaldehyde imports in 2006 were about 10,000
metric tons [11,000 tons], or roughly 0.2% of consumption, while exports were about
14,000 metric tons [15,400 tons], or about 0.3% of production.

## 11 2.2.2 Other production sources

12 In addition to intentional industrial production, formaldehyde is produced unintentionally 13 from natural sources and from human activities. Combustion processes account either 14 directly (i.e., release of formaldehyde) or indirectly (i.e., release of chemicals that are 15 reduced to formaldehyde in the environment) for most of the formaldehyde entering the 16 environment (ATSDR 1999, Howard 1989). Combustion sources include automobiles 17 and other internal combustion engines, power plants, incinerators, refineries, forest fires, 18 wood stoves, and cigarettes. Photochemical oxidation of hydrocarbons and other 19 precursors released from combustion processes can be a significant indirect source of 20 formaldehyde. Formaldehyde may also be produced in the atmosphere by the oxidation 21 of methane; this is probably the predominant source of formaldehyde in regions remote 22 from hydrocarbon emissions. Formaldehyde is also formed in the early stages of 23 decomposition of plant residues in soil (IARC 2006).

### 24 2.2.3 Endogenous production

In humans and other animals, formaldehyde is an essential metabolic intermediate in all cells and is produced endogenously from serine, glycine, methionine, and choline, and from the demethylation of *N*-, *O*-, and *S*-methyl compounds (IARC 2006) (see Section 5.1). Zhang *et al.* (2009a) reported that the endogenous concentration of formaldehyde in the blood of humans, monkeys, and rats is approximately 2 to 3 mg/L.

#### 1 **2.3** Biological indices of exposure

2 Direct measures of exposure to formaldehyde normally would involve determination of 3 formaldehyde or its major metabolite formic acid (or formate) in blood or urine of 4 exposed individuals. Neither formaldehyde nor formate has been very useful for direct 5 biological monitoring, for several reasons. Levels of both of these molecules show large 6 intrapersonal and interpersonal variation even in the absence of formaldehyde exposure 7 (ATSDR 1999). Because both formaldehyde and formate are simple one-carbon 8 molecules that are rapidly metabolized and incorporated into the one-carbon pathway or 9 oxidized to carbon dioxide (Shaham et al. 2003), most of the formaldehyde taken into the 10 body becomes unidentifiable as the parent molecule or major metabolite. A further 11 complication is the formation of formaldehyde in vivo from the metabolism of many 12 xenobiotics, including carbon tetrachloride, endrin, paraquat, dioxins, and 13 dichloromethane (ATSDR 1999). Formate can also be part of the metabolic pathways of 14 chemicals such as methanol, halomethanes, and acetone (ATSDR 1999, Shaham et al. 15 2003).

16 Formaldehyde can bind covalently to single-stranded DNA and protein to form crosslinks 17 or with human serum albumin (HSA) or the N-terminal valine of hemoglobin to form 18 molecular adducts, and these reaction products of formaldehyde might serve as 19 biomarkers for exposure to formaldehyde. Pala et al. (2008) reported a significant 20 relationship between levels of exposure to airborne formaldehyde and formaldehyde-21 HSA conjugate (FA-HSA); however, no relationship was observed between exposure 22 levels and chromosomal aberrations, micronuclei, or sister chromatid exchanges. 23 Metabolism of formaldehyde and adduct formation are discussed in Section 5, and the 24 potential for these molecules as biomarkers for formaldehyde exposure is described in the 25 remainder of this section.

Shaham *et al.* (1996a, 1997) conducted a pilot study to investigate the use of DNAprotein crosslinks as a biomarker for formaldehyde exposure in humans. DNA-protein crosslinks were measured in white blood cells from 12 exposed workers (physicians and technicians) and 8 unexposed controls. The workers had been exposed to formaldehyde

30 from 2 to 31 years, with a mean of 13 years. Formaldehyde concentrations were

1 measured in the room air and in personal samples. Concentrations ranged from about 1.4 2 to 3.1 ppm [1.7 to 3.8 mg/m<sup>3</sup>]. The levels of crosslinks were significantly higher (P =3 0.03) in exposed workers than in controls and significantly higher (P < 0.05) in the most-4 exposed workers (technicians) than in less-exposed workers (physicians). Furthermore, 5 the years of exposure and levels of crosslinks were linearly related. Smoking did not 6 influence the results. The authors concluded that DNA-protein crosslinks can be used as a 7 method for biological monitoring of formaldehyde exposure.

8 Shaham et al. (2003) conducted a follow-up study of the relationship of DNA-protein 9 crosslinks to occupational exposure to formaldehyde. This study also investigated effects 10 on p53 protein expression (see Section 5.5.4.1). The workers included physicians, 11 laboratory assistants and technicians, and hospital orderlies at 14 hospital pathology 12 departments, and the workers had a mean exposure period of 15.9 years (range = 1 to 51 13 years). The exposed group included 59 men and 127 women, who were further divided 14 into low- and high-exposure subgroups. The low-exposure group, which consisted of 15 laboratory assistants and technicians, had exposure levels ranging from 0.04 to 0.7 ppm 16  $[0.05 \text{ to } 0.86 \text{ mg/m}^3]$ , while the high-exposure group, which consisted of physicians and 17 orderlies, had exposure levels ranging from 0.72 to 5.6 ppm  $[0.88 \text{ to } 6.9 \text{ mg/m}^3]$ . [Note 18 that characterization of the exposure levels of physicians and technicians as being high or 19 low differed between the two studies by Shaham et al.] The control group included 213 20 administrative workers (127 men and 86 women) at the same hospitals. Age distribution, 21 sex, origin, and education differed significantly between the exposed and control groups; 22 therefore, the data were adjusted for these variables. DNA-protein crosslinks were 23 measured in the mononuclear-cell fraction of peripheral blood. The adjusted mean 24 number of crosslinks was significantly higher (P < 0.01) in the total exposed group than 25 in the control group. The mean number of crosslinks did not differ significantly by level of exposure or median years of exposure ( $\leq 16$  vs. > 16 years). 26

27 Pharmacokinetic modeling suggests that the rate of formation of DNA-protein crosslinks

is dose-dependent (IARC 2006), and it has been suggested that this rate can serve as a

surrogate for the delivered dose of formaldehyde (Casanova et al. 1991, Shaham et al.

2003). DNA-protein crosslinks are also a marker for effect of exposure and are discussed
further in Section 5.

3 Madison et al. (1991) reported that levels of immunoglobin M (IgM) and 4 immunoglobin G (IgG) isotypes to FA-HSA were significantly higher in a group of 5 subjects exposed to formaldehyde from an urea-formaldehyde spill than in a non-exposed 6 group (see Section 5.3.2.4 for additional details). Carraro et al. (1999) later developed an 7 indirect competitive enzyme immunoassay to titrate serum anti-FA-HSA antibodies 8 using FA-HSA adducts conjugated in vitro. The assay was used to examine two groups of 9 roughly 90 healthy adults each, using adducts with a different ratio of formaldehyde to 10 HSA for each group (5:1 and 10:1). The assay was more sensitive and specific with the 11 10:1 adduct than with the 5:1 adduct. The authors noted that the results of this study 12 supported the assertion that the FA-HSA adduct is a good marker for formaldehyde 13 exposure and concluded that this assay appeared to be able to evaluate immunological 14 response against this adduct, in particular when the adduct with the 10:1 ratio was used. 15 They suggested that the assay could be a useful tool for investigating formaldehyde 16 exposure; however, no follow-up to this study was found in the literature.

17 Bono et al. (2006) found that the prevalence of N-methylenvaline (a molecular adduct 18 formed by addition of formaldehyde to the N-terminal value of hemoglobin) in blood 19 was significantly higher in exposed workers than in non-exposed controls, and that levels 20 of N-methylenvaline in blood were positively related to formaldehyde exposures. The 21 authors concluded that its measurement in blood could be useful as a biomarker for 22 occupational exposure to formaldehyde. For this study, 21 volunteers occupationally 23 exposed to formaldehyde were recruited from a plywood factory and a laminate factory; 24 30 non-exposed workers served as a control group. The procedure for each subject 25 consisted of the administration of a questionnaire, application of a passive sampler for 26 one eight-hour working day, collection of a venous blood sample for N-methylenvaline 27 determination, and collection of a urine sample to investigate the presence of cotinine (a 28 biomarker for tobacco smoke exposure). Formaldehyde levels in personal air samples 29 were significantly higher (P = 0.0001) for workers at both factories than for the controls, 30 whereas the difference between the two factories was not statistically significant. Mean

19

1 exposure levels were 0.092 mg/m<sup>3</sup> for the plywood factory and 0.076 mg/m<sup>3</sup> for the 2 factory producing laminates. *N*-Methylenvaline distribution in blood showed a direct 3 positive relationship to formaldehyde exposure (r = 0.465), and prevalence of the 4 molecular adduct (as nanomoles per gram of globin) was significantly higher (P < 0.04) 5 in the exposed group than in the control group.

6 Li et al. (2007a) investigated the formation of antibodies against formaldehyde-protein 7 conjugates in rats as a potential biological marker for formaldehyde exposure. Male 8 Sprague-Dawley rats were exposed to formaldehyde in their drinking water (1.6 mg/mL) 9 for up to 6 months. Blood samples were collected at 3 and 6 months, and antibodies were 10 measured in the serum. Antibodies were detected in half the animals at both 3 and 6 11 months, but the antibody titer was higher at 6 months. The antibodies were highly 12 specific and did not cross-react with malondialdehyde or other albumin adducts. The 13 antibody against formaldehyde-albumin adducts also recognized formaldehyde-human 14 albumin conjugates, but only with about one-third the binding affinity. The authors 15 concluded that anti-formaldehyde-protein conjugate antibodies are a potential biomarker 16 for formaldehyde exposure.

### 17 **2.4 Occupational exposure**

18 No current data were found on the number of U.S. employees who are exposed to 19 formaldehyde; however, in the late 1980s, the Occupational Safety and Health 20 Administration (OSHA) estimated that over 2 million U.S. workers were exposed to 21 formaldehyde, with about 45% of these working in the garment industry (ATSDR 1999). 22 OSHA estimated that about 1.9 million workers were exposed to formaldehyde at 23 concentrations between 0.1 and 0.5 ppm  $[0.12 \text{ and } 0.61 \text{ mg/m}^3]$ , about 123,000 at concentrations between 0.5 and 0.75 ppm [0.61 and 0.92  $mg/m^3$ ], about 84,000 at 24 25 concentrations between 0.75 and 1 ppm [0.92 and 1.23 mg/m<sup>3</sup>], and about 107,000 at concentrations greater than 1 ppm  $[1.23 \text{ mg/m}^3]$ . It has been suggested that because 26 27 formaldehyde is ubiquitous, occupational exposure occurs in all workplaces (WHO 28 2002).

1 OSHA (1990) stated that formaldehyde exposure can occur in three ways: (1) exposure to 2 liquid or solid formaldehyde (paraformaldehyde) and the accompanying vapors, 3 (2) exposure to formaldehyde during primary processing of formaldehyde resins and 4 other chemicals manufactured from formaldehyde, and (3) exposure to formaldehyde 5 released from products that contain formaldehyde-based resins. In occupational 6 environments, formaldehyde occurs mainly as a gas; however, formaldehyde particulates 7 can be inhaled when paraformaldehyde or powdered resins are used, or when 8 formaldehyde adsorbs to other particulates such as wood dust (IARC 1995). Dermal 9 exposure also is possible when formalin solutions or liquid resins come in contact with 10 the skin; however, no data were found on dermal exposures.

11 IARC (2006) noted that in the past, the highest continuous exposures have been 12 measured during the varnishing of furniture and wooden floors, in the finishing of 13 textiles, in the garment industry, during the treatment of furs, and in certain jobs within 14 manufactured board mills and foundries. Short-term exposures to high levels have been 15 reported for embalmers, pathologists, and paper workers. Lower levels have usually been 16 encountered during the manufacture of synthetic vitreous fibers, abrasives, and rubber, 17 and in formaldehyde production industries. A very wide range of exposure levels has 18 been observed in the production of resins and plastic products.

19 Lavoué et al. (2008) extracted OSHA personal exposure monitoring data for

20 formaldehyde (N = 5,228) from the U.S. Integrated Management Information System

21 (IMIS) in order to develop a retrospective assessment of formaldehyde exposure and to

22 determine what factors affect exposure levels. The authors noted that overall, short-term

23 measurements were higher than time-weighted average (TWA) measurements. Short-

term measurements decreased 18% per year until 1987, the year in which the OSHA

25 permissible exposure limit (PEL) was implemented (see Section 2.7.1), and then 5% per

year after that. TWA measurements decreased at a rate of 5% per year until 1987 and 4%
per year thereafter.

28 Formaldehyde concentrations from IMIS were analyzed with a linear mixed-effects

29 model, and TWA and short-term levels were estimated for numerous industries. The

21

1 highest estimated TWA concentrations were for the reconstituted wood products,

2 structural wood members, and wood dimension and flooring industries (geometric mean

 $3 = 0.2 \text{ mg/m}^3$ ), and the highest estimated short-term levels were for the funeral service and

4 crematory and reconstituted wood products industries (geometric mean =  $0.35 \text{ mg/m}^3$ ).

5 The authors noted that very low and very high temperatures were associated with higher

6 exposure levels.

7 In a review of formaldehyde exposure in China, Tang et al. (2009) noted that the wood

8 processing industry had the highest average industrial formaldehyde air concentration,

9 caused in part by unventilated workshops and a lack of employee safety precautions.

10 This section provides information on various industries where occupational exposure to

11 formaldehyde occurs: these include formaldehyde and formaldehyde-based resin

12 production, wood-based products and paper production, manufacture of textiles and

13 garments, foundries, production of formaldehyde-based plastics, embalming, histology,

14 construction activities, fiberglass and mineral wool insulation production, firefighting and

15 combustion-related exposures, agriculture, office-building exposures, and other

16 exposures. Tables are provided with exposure levels; where available, information on

17 sources of exposure and exposure reduction methods is included in the text. In addition to

18 the review articles discussed above (i.e., WHO 1989, ATSDR 1999, and IARC 2006),

19 Tang *et al.* (2009) performed an extensive review of occupational exposure to

20 formaldehyde in China, and this article is used throughout the occupational exposure

21 section. As with the other review articles, the primary reference is provided for the data

from Tang *et al*.

Often, information on the specific resin used in a process was not provided in the source document; where available, this information is provided with the exposure levels. Within the exposure-level tables, the data generally are sorted by industry and then by year of publication of the study. Throughout the tables in this section, concentrations are presented in units of milligrams per cubic meter. If the concentrations were presented in parts per million in the source document, values were multiplied by a conversion factor of 1.23.

#### 1 2.4.1 Formaldehyde and formaldehyde-based resin production

As noted in Section 2.2.1, most industrial production of formaldehyde is in the form of
formalin; an aqueous solution of formaldehyde with small amounts of stabilizers such as

4 methanol added to prevent polymerization. The predominant industrial use of

5 formaldehyde is in the production of urea-, phenol-, and melamine-formaldehyde resins,

6 which are used primarily as binders for wood products such as particleboard, MDF,

7 plywood, and wood-molding compounds and as laminates for flooring, cabinets,

8 countertops, furniture, and similar items (Bizzari 2007). Another major use of

9 formaldehyde is for the production of polyacetal resins, which are used widely in the

10 production of plastics, industrial machinery, automotive components, and various

11 consumer and industrial goods (Bizzari 2007, IARC 2006) (see Section 2.5.5).

12 Jobs with potential exposure during the production of formaldehyde or formaldehyde-

13 based resins include machine operator, reception and shipping clerk, maintenance

14 worker, laboratory technician, foreman, and office worker (IRSST 2006). Tasks that may

15 result in formaldehyde exposure include collecting product samples for analysis,

16 maintenance and repair operations, filter replacement, bagging, and filling trucks and

17 barrels. The main factors that affect occupational exposures to formaldehyde include the

18 condition of the piping and equipment, the presence and efficiency of fume hoods or

19 local collection systems at the source of the emissions, and the efficiency of the general

20 ventilation system.

21 IARC (2006) reported that mean air levels of formaldehyde were less than 1 ppm

22 [1.23 mg/m<sup>3</sup>] during the manufacture of formaldehyde and ranged from less than 1 ppm

23  $[1.23 \text{ mg/m}^3]$  to more than 10 ppm  $[12.3 \text{ mg/m}^3]$  during the manufacture of

24 formaldehyde-based resins. Table 2-1 presents exposure data for formaldehyde and

25 formaldehyde-based resin production. IARC (2006) noted that while obvious differences

26 have been seen in formaldehyde air levels among factories producing formaldehyde-

27 based resins, no consistent seasonal variation has been demonstrated. Workers in

28 formaldehyde production may also be exposed to methanol, carbon monoxide, carbon

29 dioxide, and hydrogen as process gases.

1 In Canada, formaldehyde production is done in a continuous closed circuit and is

- 2 completely automated (IRSST 2006); however, no information was found on processes
- 3 used in the United States for formaldehyde or formaldehyde-resin production or the

4 potential for releases to air.

5 The major steps that can be taken to reduce exposure in this industrial sector include

6 confining operations that may result in formaldehyde exposure, such as sample

7 collection, barrel filling, filter cleaning, and tanker-truck filling operations, and installing

8 hoods above the emission sources. Ensuring proper general ventilation with outside air

9 will also help reduce exposure levels, and personal protective equipment should be used

10 where exposure levels are high (IRSST 2006).

		Exposure level	
		mean (range)	Reference
Industry (year measured)	Ν	(mg/m³)	Location
Formaldehyde production			
Formaldehyde production (2001)	48	1.07 (0.5–3.5)	Li and Chen 2002 <sup>a</sup>
			China
Formaldehyde production (1988–			Zhang et al. 1999 <sup>a</sup>
1997)			China
Oxidation	196	1.2 (0.01–2.1)	
Storage	206	1.3 (0.02–1.8)	
Formaldehyde workshops			
(1994)	22	0.985 (NR)	Cheng et al. 1995 <sup>a</sup>
(1995)	NR	NR (0–2.88)	Huan et al. 2001 <sup>a</sup>
(1995)	NR	NR (0–3.66)	Huan et al. 2001 <sup>a</sup>
(1996)	12	2.53 (0.24-8.03)	Wang et al. 1997 <sup>a</sup>
(2006)	21	0.029 (0.022-0.044)	Yang 2007a <sup>a</sup>
			China
Factory producing formaldehyde	62	0.3 (0.05–0.5)	Holmström et al. 1989b <sup>b</sup>
and resins (1979–1985)			Sweden
Formaldehyde manufacture			Stewart et al. 1987a <sup>b</sup>
(1983)	15	$0.7 (0.04 - 2.3)^{c}$	United States
Plant 2 summer	9	$0.9 (0.7 - 1.0)^{c}$	Childe States
Plant 10 summer			
Formaldehyde production (1980s)	9	0.3 (NR)	Rosen et al. 1984 <sup>b</sup>
			Sweden
Paraformaldehyde packaging			Blade 1983 <sup>d</sup>
(NR)			United States
Personal sampling	10	0.66 (< 0.30–1.02)	
Area sampling	8	1.4 (0.34–4.08)	
Formaldehyde production (NR)			NIOSH 1980a <sup>d</sup>
Production operator	NR	1.68	United States
Laboratory technician	NR	1.57	

# Table 2-1. Formaldehyde exposure levels associated with formaldehyde production and formaldehyde-based resin production

		Exposure level mean (range)	Reference
Industry (year measured)	N	(mg/m <sup>3</sup> )	Location
Formaldehyde-based resin produ	ction		
Resin production (1981–1982)			Heikkila et al. 1991 <sup>b</sup>
Furan resin production	3	2.9 (1.3-4.2)	Finland
Maintenance	4	3.6 (1.8-6.9)	
UF resin production	7	0.9 (0.7–1.1)	
Resin production (NR)	NR	0.3 (0.05–0.5)	Holmström et al. 1989a <sup>e</sup>
			United States
Resin production (1983–1984)			Stewart et al. 1987a <sup>b</sup>
Plant 1 summer	24	$4.2 (0.3-16.2)^{c}$	United States
Plant 6 summer	6	0.3 (0.1–0.3) <sup>c,f</sup>	
Plant 7 summer	9	$0.3 (0.1-0.4)^{c}$	
Plant 7 winter	9	$0.7 (0.5 - 1.1)^{c}$	
Plant 8 summer	13	0.7 (0.3–1.0) <sup>c,f,g</sup>	
Plant 8 winter	9	0.1 (0.1–0.3) <sup>c,f,g</sup>	
Plant 9 summer	8	17.5 (5.0–37.5) <sup>c,f,g</sup>	
Plant 9 winter	9	2.1 (1.4–3.1) <sup>c</sup>	
Plant 10 summer	23	$0.9 (0.4 - 1.5)^{c,g}$	
Resin production (1980s)	22	0.6 (NR)	Rosen et al. 1984 <sup>b</sup>
			Sweden
Resin and plastic materials	NR	1.67 (NR) <sup>h</sup>	NIOSH 1980a <sup>d</sup>
production (NR)			United States

NR = not reported.

<sup>a</sup>Cited in Tang *et al.* 2009.

<sup>b</sup>Cited in IARC 2006.

<sup>c</sup>Mean and range of geometric means.

<sup>d</sup>Cited in WHO 1989.

<sup>e</sup>Cited in ATSDR 1999.

<sup>f</sup>Some of the sampling results were affected by simultaneous occurrence of phenol, which interferes with the measurement method, leading to artificially low values.

<sup>g</sup>Some of the sampling results were affected by a simultaneous occurrence of particulates "that contained nascent formadehyde (leading to high values)."

<sup>h</sup>Data also presented in Table 2-8.

### 1 2.4.2 Wood-based products and paper production

2 The predominant use for formaldehyde-based resins is in the production of wood-based

3 composites; UF, MF, melamine-urea-formaldehyde (MUF), and PF resins all can be

- 4 used, depending on the product being manufactured. Plywood and other laminated wood
- 5 products often are referred to as composite-wood products; however, in this section, they
- 6 are discussed separately from other wood-based composites, because of important

1 differences in manufacturing processes and exposure potential. Wood furniture and

2 paper-product manufacturing also are discussed in this section.

3 2.4.2.1 Wood-based composites

4 The product class of wood-based composites includes particleboard, fiberboard, and 5 oriented strandboard (OSB), which are differentiated primarily by the type of wood fiber 6 used (i.e., from large particles to small fibers). Regardless of the type of fiber used, the 7 manufacturing process is basically the same: (1) the wood fiber is bonded together with a 8 thermosetting resin to form a mat, (2) the mat is hot-pressed, and (3) the pressed mat is 9 then cooled and allowed to mature (IRSST 2006). The wood fibers typically are bonded 10 with UF, MF, MUF, or PF resins. During hot-pressing, the mat is heated and compacted 11 to the desired density and thickness, and the resin polymerizes to bind the particles and 12 stabilize the panel.

13 UF resins are primarily used in the manufacture of products where dimensional 14 uniformity and surface smoothness are of primary concern. Conner (2001) reported that 15 over 70% of the UF resin produced is used by the forest industry in the production of 16 particleboard (61%), MDF (27%), hardwood plywood (5%), and as a laminating adhesive 17 (7%). The popularity of UF resins results from a number of factors, including low cost, 18 ease of use, water solubility, hardness, and lack of color. However, moist conditions, 19 especially when combined with heat, lead to a reversal of the bond-forming reactions and 20 result in the release of formaldehyde. For this reason, UF resins are unsuitable for most 21 outdoor uses and are used almost exclusively for products intended for indoor use. MF 22 and MUF resins are more resistant to breakdown in moist environments; however, 23 melamine is much more expensive than urea. MF resins are used primarily for decorative 24 laminates. PF resins are the most resistant to breakdown from moisture and thus typically 25 are used in products requiring some degree of outdoor exposure durability, such as OSB. 26 PF resins also have a darker color, making them generally less suitable for decorative 27 products such as paneling and furniture (USDA 1999).

- 28 The major determinants of worker exposure levels are the type of resin used and the
- 29 molar ratio of formaldehyde to the other components (IRSST 2006). IRSST noted that
- 30 the emission rate is highest for UF resin and lowest for PF resin. Other parameters that

1 affect exposure levels include process operating conditions, such as temperature, pressing 2 time, panel thickness, and maturation time; the presence and efficiency of fume hoods or 3 other collection systems; and the level of general ventilation. Production areas and 4 processes associated with formaldehyde exposure include gluing (both glue preparation 5 and application), board press operations, board cooling operations, maturing and drying, 6 and storage. Jobs that may result in formaldehyde exposure include resin preparer, press 7 operator, finisher, laminator, laboratory technician, and maintenance and office 8 personnel. The main means of controlling exposure to formaldehyde are substitution 9 (e.g., isocyanate-based products can be used for some applications but have high 10 toxicity), the use of resins with lower emission rates, confinement of production steps 11 that produce formaldehyde emissions, the use of hoods and capture devices, good general 12 ventilation, and the use of personal protection where formaldehyde levels are high. 13 Process- and product-related changes over the past few decades have led to general 14 reductions in levels of occupational exposure to formaldehyde, which is reflected in the 15 data presented by Kauppinen and Niemelä (1985) (as cited in IARC 2006) (see 16 Table 2-2). Lower mean exposure levels were seen for all operations that were assessed 17 during the 1975 to 1984 time period when compared with the 1965 to 1974 time period. 18 These data indicate that tasks with the highest exposure levels include glue preparation,

19 hot pressing, and sawing.

Industry (year measured)	N	Exposure level mean (range) (mg/m³)	Reference Location
Particleboard production MDF production OSB production	332 42 2	0.56 <sup>a</sup> (NR) 0.41 <sup>a</sup> (NR) 0.05 <sup>a</sup> (NR)	Lavoue <i>et al.</i> 2007 Compiled data from various locations
Fiberboard production (2003) (2005)	60 NR	0.42 (0.11–0.86) 0.41 (0.14–3.2)	Geng <i>et al.</i> 2004 <sup>b</sup> Jiang <i>et al.</i> 2006 <sup>b</sup> China

 
 Table 2-2. Formaldehyde exposure levels associated with the production of woodbased composites

		Exposure level	Peference
		mean (range)	Kelelence
Industry (year measured)	N	(mg/m°)	Location
Blocking			
(2002)	40	1.13 (0.35–2.6)	Fan <i>et al.</i> $2004^{\text{b}}$
(2005)	NR	0.18 (NR)	Shi <i>et al.</i> 2006 <sup>°</sup>
			China
Fiberboard sawing and sanding	46	$0.04-0.13 (0.01-0.17)^{c}$	Chung et al. 2000 <sup>d</sup>
(1990s)			United Kingdom
OSB plant (1990s) <sup>e</sup>	20	$\leq$ 0.06 (NR)	Herbert et al. 1995 <sup>d</sup>
			Canada
Particleboard mill (NR)	9	3.0 (1.5–4.3)	Malaka and Kodama 1990 <sup>d</sup>
			Indonesia
Blockboard mill (NR)	6	0.6 (0.5–0.7)	Malaka and Kodama 1990 <sup>d</sup>
			Indonesia
Chipboard production (1980–1988)	24	1.9 (< 0.01–10)	Triebig et al. 1989 <sup>d</sup>
			Germany
Particleboard and molded plastics	NR	$0.85 (0.21 - 3.6)^{\rm f}$	Horvath <i>et al.</i> 1988 <sup>g</sup>
plant (NR)			United States
Two particleboard plants and a	NR	NR (0.1–1.11) <sup>h</sup>	Edling et al. 1988 <sup>g</sup>
laminate plant (1980s)			Sweden
Particleboard sanding (NR)	NR	NR (0.23–0.96)	Stumpf et al. 1986 <sup>g</sup>
			United States
Particleboard mills (1965–1984)			Kauppinen and Niemela 1985 <sup>d</sup>
Glue preparation 1975–1984	10	2.7 (0.4–6.0)	Finland
Blending 1965–1974	10	1.2 (0.1–2.5)	
Blending 1975–1984	8	0.9 (< 0.1–1.7)	
Forming 1965–1974	26	2.1 (< 0.6–5.7)	
Forming 1975–1984	32	1.7 (0.1–5.9)	
Hot press 1965–1974	35	4.2 (1.4–11.7)	
Hot press 1975–1984	61	2.1 (0.25–5.7)	
Sawing 1965–1974	17	5.9 (0.9–11.3)	
Sawing 1975–1984	36	1.2 (< 0.1–4.1)	
Coating 1965–1974	7	1.2 (0.6–2.2)	
Coating 1975–1984	12	0.5 (0.1–1.5)	
Particleboard and MDF production	40	0.3–0.4 (NR)	Rosen <i>et al.</i> 1984 <sup>d</sup>
(19808)			Sweden
Cork compression (1985)	28	3.01 (0.33-46.14)	Gao et al. 1988 <sup>b</sup>
			China

NR = not reported. <sup>a</sup>Median geometric mean from data compiled from 13 studies.

<sup>b</sup>Cited in Tang et al. 2009.

<sup>c</sup>Includes both gaseous formaldehyde and formaldehyde extracted from dust for various products; maximum levels are for formaldehyde extracted from dust. <sup>d</sup>Cited in IARC 2006. <sup>e</sup>Includes debarking, pre-heat conveyor, post-heat conveyor, and packaging and storage. <sup>f</sup>Mean and range of TWAs. Data also presented in Table 2-8. <sup>g</sup>Cited in ATSDR 1999.

<sup>h</sup>Data from the particleboard and laminate plants are not segregated. Presented is a range of estimated TWAs; peaks of up to  $5 \text{ mg/m}^3$  were reported.

1 2.4.2.2 Plywood and other laminated veneer

- 2 This industrial sector involves the manufacture of plywood, veneer, laminated wood, and
- 3 panel coating and generally involves gluing together panels of wood veneer or other
- 4 materials. Regardless of the end product, the process generally consists of five steps:
- 5 gluing, pressing, drying, finishing, and storage. Adhesives used in this industry can be
- 6 made of UF, MF, MUF, or PF resins. UF, MF, or MUF resins are used primarily for
- 7 decorative products intended for indoor use, while PF resins are used for structural
- 8 plywood (softwood plywood) and weather-resistant materials (USDA 1999, WSDE
- 9 1998). Methods of applying the adhesives include spraying, curtain coating, roller
- 10 coating, extrusion, and foaming (USDA 1999). The veneer panels are laid up by hand,
- 11 machine, or a combination of both. The glue is then allowed to partially cure under
- 12 pressure. Pressing operations can include cold pressing (pressing at ambient
- 13 temperatures), hot pressing (pressing at high temperatures), or a combination of the two.
- Hot pressing is used for some UF glues and for all PF glues (WSDE 1998). Pressing 14
- 15 times range from a few minutes to several hours depending on the temperature of the
- 16 press, the size of the product, and the type of glue used.

17 Sources of exposure within this sector include glue preparation and application, press 18 operations, drying and storage, maintenance operations, finishing operations, and 19 packaging and transportation operations. The main factors that affect worker exposure 20 include the type of resin and the molar ratio used; process operating conditions, such as 21 temperature, amount of pressure applied and duration of pressing, panel thickness, and 22 type of wood coating; the presence and efficiency of fume hoods and local collection 23 systems; and the efficiency of the general ventilation system (IRSST 2006). Measures to 24

- control exposure include product substitution (e.g., isocyanate resins are available, but
- 25 their toxicity is high), the use of resins with lower emission rates (PF resins release less

1 formaldehyde during curing than UF resins), confinement of production steps that

- 2 produce formaldehyde emissions, installation of fume hoods above the sources of
- 3 emissions, sufficient levels of ventilation in the finishing and storage areas to dissipate
- 4 residual formaldehyde emissions, and the use of personal protection where exposure
- 5 levels are high.
- 6 Numerous process- and product-related changes over the past few decades have led to
- 7 general reductions in occupational exposure levels, as can be seen in Table 2-3. Of
- 8 particular interest are data reported for several different processes for the periods 1965–
- 9 74 and 1975–84 by Kauppinen (1986) (as cited in IARC 2006); mean exposure levels for
- 10 all operations assessed during 1975–84 had decreased from 1965–74. Based on these
- 11 data, tasks with the highest exposure levels include glue preparation and hot pressing, and
- 12 major exposure-level reductions were seen for these tasks.

		Exposure level mean (range)	Reference
Industry (year measured)	N	(mg/m <sup>3</sup> )	Location
Plywood panels production	8	0.092 (NR)	Bono et al. 2006
Laminates production	13	0.076 (NR)	NR
Plywood mill (2000)			Fransman et al. 2003 <sup>b</sup>
Dryers	14	$0.07^{a}$ (NR)	New Zealand
Composers	2	$0.03^{a}$ (NR)	
Pressing	5	$0.16^{a}$ (NR)	
Finishing end	1	$0.04^{a}$ (NR)	
Plywood mill (1996–1997)			Makinen et al. 1999 <sup>b</sup>
Patching	6	0.07 (0.03-0.10)	Finland
Feeding of drying machine	6	0.06 (0.01-0.15)	- Innuncu
Forklift driving	6	0.07 (0.02-0.20)	
Scaring [scarfing]	6	0.14 (0.07-0.24)	
Assembly (machine 1)	4	0.30 (0.10-0.81)	
Assembly (machine 2)	6	0.15 (0.10-0.27)	
Hot pressing	5	0.13 (0.08-0.23)	
Glue preparation	2	0.15 (0.07-0.23)	
Finishing	4	0.09 (0.07-0.14)	
Carrying plywood piles	2	0.06 (0.05-0.07)	
Finishing	2	0.05 (0.01-0.07)	

## Table 2-3. Formaldehyde exposure levels associated with the manufacture of plywood and laminates

		Exposure level	Reference
Industry (year measured)	N	(mg/m <sup>3</sup> )	Location
Plywood factory (NR)			Ballarin et al. 1992 <sup>c</sup>
Warehouse	3	0.39 (0.21-0.60)	Italy
Shearing press	8	0.10 (0.08-0.14)	
Sawmill	1	0.09 (1 sample)	
Plywood mill (NR)	40	0.8 (0.3–2.8)	Malaka and Kodama 1990 <sup>b</sup>
			Indonesia
Plywood paneling manufacture			Stewart et al. 1987a <sup>b</sup>
(1983–1984)			United States
Winter	27	0.3 <sup>a</sup> (0.1–0.5)	
Summer	26	0.1 <sup>a</sup> (0.01–0.6)	
Plywood mills (1964–1984)			Kauppinen 1986 <sup>b</sup>
Glue prep 1965–1974	15	2.7 (0.7-6.2)	Finland
Glue prep 1975–1984	19	0.9 (0.1–2.8)	1 mund
Assembly 1965–1974	32	1.9 (< 0.1–5.4)	
Assembly 1975–1984	55	0.7 (0.03-8.3)	
Hot press 1965–1974	41	2.5 (< 0.1–9.5)	
Hot press 1975–1984	43	0.6 (0.07–2.6)	
Sawing 1965–1974	5	0.6 (0.4–1.0)	
Sawing 1975–1984	12	0.1 (0.03–0.3)	
Coating 1965–1974	7	1.2 (0.6–2.2)	
Coating 1975–1984	28	0.4 (0.03–0.7)	
Plywood production (1980s)	47	0.4 (NR)	Rosen et al. 1984 <sup>b</sup>
			Sweden

NR = not reported.

<sup>a</sup>Geometric mean.

<sup>b</sup>Cited in IARC 2006.

<sup>c</sup>Cited in IARC 2006 and ATSDR 1999; data presented are from the original article, because of discrepancies between data presented in the IARC and ATSDR papers.

### 1 2.4.2.3 Wood furniture

2 Most furniture is manufactured from either wood-based composite or hardwood, and the

3 manufacturing process can be generalized into four steps: (1) processing (sawing,

4 sanding, assembly, inspection), (2) painting, staining, or varnishing (mixing, applying,

5 drying, sanding, repair), (3) upholstery and installation of hardware, and (4) packaging

6 and shipping (IRSST 2006). IRSST (2006) noted that most of the adhesives used in the

7 industry do not emit formaldehyde; although wood-based composites and veneers may

8 emit some formaldehyde, the main source of formaldehyde in this industry originates

9 from finishes used on the furniture. Formaldehyde-based resins often are used to

crosslink more flexible resins, providing finishes that have good scratch and chemical
 resistance for use in furniture surface coatings (TIG 2005).

3 Exposure determinants include the type of varnish used; process operating conditions, 4 such as the nature of the spraying systems, drying time, and the location of operations; 5 work methods employed; the presence and efficiency of varnishing booths and other 6 local collection systems at the source; and the efficiency of the general ventilation system 7 (IRSST 2006). Tasks that can result in formaldehyde exposure include paint preparation, 8 application of primers and varnishes, sanding between coats, unloading of furniture from 9 ovens, repair tasks, installation of hardware, cleaning of application guns, and maintenance. Sources of formaldehyde release include releases from varnish use and 10 11 storage, paint booths, furniture drying operations, and furniture storage. Jobs that may 12 result in exposure include laborer, painter, finish operator, repair and maintenance personnel, finisher/shipper, supervisor, and office personnel. 13

14 Exposure control measures can include product substitution (i.e., use of formaldehyde-

15 free coatings), confinement of operations with high emissions (e.g., preparation and

16 application of varnish and paint in booths), good local and general ventilation, good work

17 methods (such as proper use of capture devices), and the use of personal protection where

18 formaldehyde levels are high (IRSST 2006). Table 2-4 provides formaldehyde levels that

19 have been measured in the wood furniture manufacturing industry.

 Table 2-4. Formaldehyde exposure levels associated with wood furniture manufacturing

Operation (year measured)	N	Exposure level mean (range) (mg/m <sup>3</sup> )	Reference Location
Wood processing			
(1995)	104	3.07 (0.7–19.2)	Feng et al. 1996 <sup>a</sup>
(1990–1998)	72	0.92 (NR)	Pan <i>et al</i> . 2000 <sup>a</sup>
(1990–1998)	90	0.87 (NR)	Pan <i>et al</i> . 2000 <sup>a</sup>
			China
		Exposure level	Reference
---	------	-----------------------------------	---
Operation (year measured)	N	(mg/m <sup>3</sup> )	Location
Woodworking shops (1990s)		(9,)	Abdel Hameed <i>et al.</i> 2000 <sup>b</sup>
Ventilated workshop	14	0.52 (0.34–0.66)	Fount
Unventilated workshop	14	0.79 (0.59–1.03)	Едурі
Manufacture of furniture (NR)			Vinzents and Laursen 1993 <sup>b</sup>
Painting	43	$0.2 (2.25)^{c}$	Denmark
Gluing	68	0.15 (2.87) <sup>c</sup>	Dominark
Furniture factories (1981–1986)			Heikkila <i>et al.</i> 1991 <sup>b</sup>
Gluing	73	0.4 (0.09–1.2)	Finland
Machining in finishing department	9	0.4 (0.1–1.1)	1 mana
Varnishing	150	1.4 (0.1–7.9)	
Furniture factory (NR)	NR	0.25 <sup>d</sup> (0.2–0.5)	Holmström et al. 1989b <sup>e</sup>
			NR
Furniture factories, finishing with paints (NR)			Alexandersson and Hedenstierna 1988 <sup>b</sup>
Paint mixer/supervisor	6	0.3 (0.2–0.5)	Sweden
Mixed duties on the line	5	0.5 (0.3–0.6)	
Assistant painter	3	0.6 (0.2–0.9)	
Spray painter	10	0.5 (0.2–1.3)	
Feeder/receiver	13	0.3 (0.1–0.9)	
Furniture factory (1975–1984)			Priha et al. 1986 <sup>b</sup>
Feeding painting machine	14	1.4 (0.4–3.3)	Finland
Spray painting	60	1.2 (0.3–5.0)	- Innund
Spray painting assistant	10	1.2 (0.3–2.0)	
Curtain painting	18	1.4 (0.3–7.5)	
Before drying of varnished furniture	34	1.8 (0.1–5.2)	
After drying of varnished furniture	14	1.7 (0.3–6.6)	
Furniture factory, varnishing (1980s)	32	0.9 (NR)	Rosen et al. 1984 <sup>b</sup>
			Sweden
Wood furniture manufacture (NR)	> 33	0.14–3.3 (0.01–7.68) <sup>f</sup>	Herrick <i>et al.</i> 1983 <sup>g</sup>
			NR
Cabinetmaking (NR)	48	max. = < 0.1	Sass-Kortsak et al. 1986 <sup>b</sup>
			Canada

<sup>a</sup>Cited in Tang *et al.* 2009. <sup>b</sup>Cited in IARC 2006. <sup>c</sup>Geometric mean and standard deviation. <sup>d</sup>Median. <sup>e</sup>Cited in ATSDR 1999. <sup>f</sup>Range of means and full range across four datasets. <sup>g</sup>Cited in WHO 1989.

### 1 2.4.2.4 Paper products

- 2 Formaldehyde-based products can be used for various purposes in paper production. UF
- 3 and MF resins can be added to fiber slurries before pressing to increase paper strength,
- 4 and UF, MF, and PF resins often are used as coatings for various types of paper products
- 5 (IARC 2006, TIG 2005). UF resins are used as adhesives in paper bags, cardboard, and
- 6 sandpaper, and formaldehyde is used as a bactericide in some paper-coating agents.
- 7 In paper-coating operations, the primary sources of emissions are from the dipping or
- 8 coating operations and from drying ovens (WSDE 1998), which is reflected in the data
- 9 presented in Table 2-5. Emissions from storage tanks and from areas where resin blends
- 10 are prepared can also be a source of exposure. In a large epidemiological study of
- 11 workers in 12 countries employed in the production departments of paper and paperboard
- 12 mills and recycling plants, the highest exposure levels were observed during the
- 13 calendering or on-machine coating operations (IARC 2006).

		Exposure level	Reference
Industry (year measured)	N	(mg/m <sup>3</sup> )	Location
Pulp and paper industry (1950–1994)			Korhonen et al. 2004 <sup>a</sup>
Pulping, refining of stock	25	0.6 (0.0–3.8)	12 countries [specific
Newsprint and uncoated paper machine	7	0.18 (0.05-0.57)	countries not reported
Fine and coated paper machine	51	1.4 (0.01–12.2)	by IARC]
Paperboard machine	8	0.6 (0.2–2.7)	
Paper/paperboard machine	228	0.5 (0.0-8.1)	
Calendering or on-machine coating	166	5.2 (0.0-61.5)	
Winding, cutting, and grading	111	0.3 (0.0–1.4)	
Repulping of waste paper	8	0.3 (0.06–0.5)	
Paper mill (1968–1973)			FIOH 1994 <sup>a</sup>
Gluing, hardening, lamination, and rolling of paper	12	1.1 (0.4–3.1)	Finland
Impregnation of paper with phenol resin	38	9.1 (< 1.1-40.6)	
Paper storage, diesel truck traffic	5	0.4 (0.25–0.5)	
Paper mill (1975–1984)			Heikkila et al. 1991 <sup>a</sup>
Coating of paper	30	0.9 (0.5–39)	Finland
Gum paper production	4	0.5 (0.3–0.8)	
Impregnation of paper with amino resin	6	3.9 (0.6–16)	
Impregnation of paper with phenol resin	20	0.1 (0.06–0.4)	

## Table 2-5. Formaldehyde exposure levels associated with the manufacture of paper and paper products

Industry (year measured)	N	Exposure level mean (range) (mg/m <sup>3</sup> )	Reference Location
Lamination and impregnation of paper with MF and PF resins (1983)			Stewart <i>et al.</i> 1987a <sup>a</sup> United States
Winter	53 39	$0.9^{\circ} (< 0.01 - 9.1)$ $0.4^{\circ} (0.06 - 0.9)$	
Paper production (1980s) Laminated paper Offset paper	23 8	0.4 (NR) 0.2 (NR)	Rosen <i>et al.</i> 1984 <sup>a</sup> Sweden
Paper and paperboard manufacture, coating preparation (NR)	11	0.61, 1.2 (< 0.01–3.6) <sup>c</sup>	NIOSH 1980a <sup>d</sup> United States
Manufacture of treated paper products (NR)	101	0.41, 0.7 <sup>e</sup> (0.17–1.19) <sup>c</sup>	NIOSH 1979b <sup>d</sup> United States
Paper and paperboard manufacture, resin impregnation (NR)	62	0.06–0.1 (0.01–0.34) <sup>c</sup>	NIOSH 1976b <sup>d</sup> United States
Map printing (1985)	28	0.64 (0.04–1.79)	Gao <i>et al.</i> 1988 <sup>f</sup> China

<sup>a</sup>Cited in IARC 2006.

<sup>b</sup>Geometric mean. The authors noted that the simultaneous occurrence of phenol in summer interfered with the measurement method, resulting in artificially low values, and that occurrence of particulates (regardless of season) resulted in some high values due to off-gassing of formaldehyde from dust.

<sup>c</sup>Range of means (or medians if denoted) and full range across two or three sets of data.

<sup>d</sup>Cited in WHO 1989.

<sup>e</sup>Median.

<sup>f</sup>Cited in Tang *et al.* 2009.

### 1 2.4.3 Manufacture of textiles and garments

- 2 Formaldehyde-based resins are used in the textile industry during the chemical finishing
- 3 stage to impart crease-resistant and flame-retardant properties and to prevent shrinkage
- 4 (IRSST 2006). Formaldehyde-based resins have been used for crease resistance since the
- 5 1950s. Early resins contained substantial amounts of extractable formaldehyde; however,
- 6 modifications in the resins have decreased free formaldehyde levels from about 0.4% to
- 7 0.01% or less, which has also resulted in lower occupational exposure levels (IARC
- 8 2006). IARC (2006) reported the results of a study in which formaldehyde air levels
- 9 increased from 0.1 to 1.0 ppm  $[0.12 \text{ to } 1.2 \text{ mg/m}^3]$  when formaldehyde content in the
- 10 fabric increased from 0.015% to 0.04%. In another study, formaldehyde air levels in
- 11 cutting rooms decreased from over 10 ppm  $[12.3 \text{ mg/m}^3]$  in 1968 to less than 2 ppm [2.5]
- $12 mg/m^3$  in 1973 as a result of improvements in resin treatment processes (IARC 2006).

1 The finishing process involves impregnating the fabric in an aqueous solution and then 2 pressing it to remove the excess solution (IRSST 2006). The main factors that affect 3 worker exposure to formaldehyde include the types of processes and products used, the 4 presence and efficiency of fume hoods and emission collection systems, and the level of 5 general ventilation. Jobs that may result in formaldehyde exposure include resin preparer, 6 process operators (various types), colorist, and maintenance worker. The main means of 7 controlling exposure include use of formaldehyde-free finishes, the use of fume hoods at 8 the source of emissions, sufficient general ventilation, and the use of personal protective 9 equipment where formaldehyde levels are high.

10 In addition to gaseous formaldehyde exposure, workers can be exposed to formaldehyde 11 bound to dust. IARC (2006) presented results of a study in a garment production facility in the United States where formaldehyde gas levels ranged from 26 to 36  $\mu$ g/m<sup>3</sup> [0.026 to 12  $0.036 \text{ mg/m}^3$  and levels of formaldehyde bound to dust ranged from 0.2 to 0.7  $\mu$ g/m<sup>3</sup> 13  $[0.0002 \text{ to } 0.0007 \text{ mg/m}^3]$ . Workers in this industry may also be exposed to ammonia, 14 15 dimethylthiourea, textile dyes, flame retardants, carrier agents, textile-finishing agents, 16 and solvents (IARC 2006). The use of formaldehyde in garments can also result in 17 formaldehyde exposure in retail shops and potentially of end users (IARC 2006, ATSDR 18 1999). Formaldehyde exposure levels associated with textile and garment manufacture 19 are presented in Table 2-6.

		Exposure level	
		mean (range)	Reference
Operation (year measured)	Ν	(mg/m³)	Location
Textile Industry			
Textile and shoe industry			
Resin collar (1989, summer)	18	NR (0.22–0.62)	Tao <i>et al.</i> 1990 <sup>a</sup>
Resin collar (1989, winter)	9	NR (1.39–5.59)	Tao <i>et al.</i> 1990 <sup>a</sup>
Paint/production (2000)	56	1.92 NR (0.4–4.3)	Pan <i>et al.</i> 2001 <sup>a</sup>
			China
Textile mills (1980s)			Rosen et al. 1984 <sup>b</sup>
Crease-resistance treatment	29	0.2 (NR)	Sweden
Flame-retardant treatment	2	1.5 (NR)	Sweden
Textile manufacture (NR)	19	0.64, 0.83 (0.13–1.6) <sup>c</sup>	NIOSH 1981 <sup>d</sup>
			United States
Textile plant (1975–1978)			Nousiainen and Lindqvist
Finishing department mixing	8	1.1 (< 0.2-> 6.0)	1979 <sup>b</sup>
Crease-resistance treatment	52	0.5 (< 0.2->4.0)	Finland
Flame-retardant treatment	67	2.5 (< 0.2->11.0)	
Other finish treatment	17	0.4 (max. = 1.5)	
Fabric store	6	1.1 (0.1–1.6)	
Textile warehouse (NR)	22	$0.30, 0.37 (0.05 - 0.88)^{c}$	NIOSH 1979a <sup>d</sup>
			United States
Textile facilities (NR)	43	$0.84, 0.96 (< 0.12 - 1.68)^{e}$	NIOSH 1979b <sup>d</sup>
			United States
Garment Industry			1
Garment manufacturing (NR)	32	0.19–0.3 (0.17–0.37) <sup>c</sup>	Echt and Burr 1997 <sup>b</sup>
			United States
Cut & spread and turn & ticket	48	$< 0.01 - 0.05 (NR)^{f}$	Kennedy et al. 1992 <sup>b</sup>
operations (NR)	10		ND
Commont in ductory (1081, 1080)	50	$0.1, 0.2, (0.02, 0.0)^{\circ}$	
Garment industry (1981–1986)	50	0.1-0.3 (0.03-0.9)	Heikkila et al. 1991
	_		Finland
Sewing plant (NR)			Luker and Van Houten
0.04% formaldehyde fabric	9	1.2 (0.6–1.4)	1990°
0.015% formaldehyde fabric	9	0.1 (< 0.1–0.3)	United States
Shirt manufacturing (NR)	NR	NR (0.12–1.2)	Stayner <i>et al.</i> 1985, Stayner <i>et al.</i> 1988 <sup>g</sup>
			NR
Use of fabric treated with	326	~0.25 (< 0.1–0.5)	Elliott et al. 1987 <sup>b</sup>
formaldehyde-based resins (1980s)			United States

# Table 2-6. Formaldehyde exposure levels associated with the textile and garment industries

		Exposure level mean (range)	Reference
Operation (year measured)	Ν	(mg/m <sup>3</sup> )	Location
Use of crease-resistant cloth (NR)	181	NR (< 0.1–1.1)	Blade 1983 <sup>b</sup>
			United States
Garment manufacturing (NR)	168	$0.23 - 0.55 (< 0.04 - 1.34)^{c}$	Blade 1983 <sup>d</sup>
			NR
Clothing production warehouse (NR)	22	$0.14, 0.47 (0.05 - 0.68)^{c}$	NIOSH 1979a <sup>d</sup>
			United States
Sewing machine operators (NR)	57	0.86, 1.44 (0.36–2.16) <sup>c</sup>	NIOSH 1979a <sup>d</sup>
			United States
Clothing pressers (NR)	40	0.08 (0.006–1.14)	NIOSH 1976a <sup>d</sup>
			United States
Permanent-press clothing production	41	0.37, 0.89 (0.0–3.24) <sup>c</sup>	USDHEW 1966, 1968 <sup>d</sup>
(NR)			United States
Shops			
Fabric shops (NR)	77	0.17 (0.04–0.34)	McGuire et al. 1992 <sup>b</sup>
			United States
Fabric shops (1985–1987)	3	0.21 (0.15-0.3)	Priha <i>et al</i> . 1988 <sup>b</sup>
			Finland
Retail dress shops (1959)	NR	NR (0.1–0.6)	Elliott <i>et al.</i> 1987 <sup>b</sup>
			United States

<sup>a</sup>Cited in Tang et al. 2009.

<sup>b</sup>Cited in IARC 2006.

<sup>c</sup>Means or range of means and full range across two to four datasets.

<sup>d</sup>Cited in WHO 1989.

<sup>e</sup>Medians and full range across two datasets.

<sup>f</sup>Range of means for different measurements of formaldehyde as gas and bound to particulates. <sup>g</sup>Cited in ATSDR 1999.

1 2.4.4 Foundries

2 The foundry process consists of pouring molten metal into a mold to obtain a cast product

3 of specific shape. The mold can also contain a core that determines the dimensions of any

- 4 internal cavity of the final product. Formaldehyde-based resins (both UF and PF) are
- 5 commonly blended with sand to produce the molds and cores used in foundries (IARC
- 6 2006). Important manufacturing steps in the foundry process include manufacturing and
- 7 assembling the molds and cores, melting the metal, pouring the metal into the mold,
- 8 cooling the molded part, removing the mold and core (shake-out), and dressing and
- 9 deflashing (IRSST 2006).

39

1 Tasks with potential formaldehyde exposure include molding-sand preparation, mold and 2 core preparation, pouring of the molten metal into the mold, and shakeout operations 3 (IRSST 2006). The main factors affecting worker exposure to formaldehyde include 4 production variables (i.e., the molding and core-making processes employed and the 5 types of metals processed), the percentage of free formaldehyde in the binder, the sizes of 6 the molds and cores, the presence and efficiency of fume hoods and other emission 7 collection systems, and the level of general ventilation (IRSST 2006). The main means of 8 controlling formaldehyde exposure include use of mold and core-making materials that 9 do not contain formaldehyde, replacement of hot-mold production processes with cold-10 hardening processes, using resins with lower emission rates, confinement of production 11 steps that produce formaldehyde emissions, installation of fume hoods at emission 12 sources, sufficient general ventilation, and use of personal protective equipment for tasks where the formaldehyde concentration is high. In a study assessing formaldehyde levels 13 14 in foundry sand, Oliva-Teles et al. (2009) reported that formaldehyde content in used 15 foundry sands decreased with time, as formaldehyde was released to the occupational 16 environment. Data presented by Heikkilä et al. (1991) (as cited in IARC 2006) show 17 major reductions in formaldehyde exposure levels for core-making operations from the 18 1970s to the 1980s (see Table 2-7).

Other chemicals to which workers potentially are exposed in the foundry industry include
silica and other mineral dusts, polycyclic aromatic hydrocarbons, asbestos, metal fumes
and dusts, carbon monoxide, isocyanates, phenols, organic solvents, and amines (IARC
2006).

		Exposure level mean (range)	Reference
Operation (year measured)	Ν	(mg/m <sup>3</sup> )	Location
Foundries (before 1975 through 1986)			Heikkila et al. 1991 <sup>a</sup>
Core-making before 1975	43	3.4 (< 0.1-> 11)	Finland
Core-making 1981–1986	17	0.4 (0.03–1.8)	
Casting 1981–1986	10	0.2 (0.03–0.8)	
Molding 1981–1986	25	0.4 (0.05–2.5)	
Foundry molder (NR)	36	0.1 (0.02–0.27)	Ahman <i>et al.</i> 1991 <sup>a</sup>
			Sweden
Foundry (1980s)			Rosen et al. 1984 <sup>a</sup>
Hot-box method	5	1.9 (NR)	Sweden
Molding	17	0.1 (NR)	
Iron foundry core machine operator (NR)	14	0.52 <sup>b</sup> (< 0.02–22.0)	NIOSH 1979b <sup>c</sup>
			United States
Bronze foundry, core machine operator (NR)	15	$0.47, 0.64 (0.14-0.96)^d$	NIOSH 1976c <sup>c</sup>
			United States

Table 2-7. Formaldehyde exposure levels associated with foundries

<sup>a</sup>Cited in IARC 2006. <sup>b</sup>Median. <sup>c</sup>Cited in WHO 1989.

<sup>d</sup>Means and full range across two datasets.

### 1 2.4.5 Production of formaldehyde-based plastic products

2 Formaldehyde-based resins (UF, MF, and PF) are used as hardenable molding materials

- 3 in plastics that are used to produce a number of end products, including electrical
- 4 insulation, melamine tableware, lawn and garden equipment, plumbing fixtures, and
- 5 various other products (ATSDR 1999, IARC 2006, OSHA 1990, WHO 1989). A growing
- 6 application for UF and MF molded compounds is to cut the cured resin into particle-sized
- 7 pieces for use as an alternative to sand in sandblasting operations (TIG 2005).
- 8 Polyoxymethylene (also called acetal resin, polytrioxane, or paraformaldehyde) is a very
- 9 strong and hard plastic that is formed through the polymerization of formaldehyde and is
- 10 an important engineering polymer commonly used to make gears, bushings, and other
- 11 mechanical parts (ATSDR 1999, DuPont 2009, WHO 1989). Because polyoxymethylene
- 12 is lightweight and harder, tougher, and longer lasting than other plastics, it is used in
- 13 many applications where metals previously were used, such as in motor vehicles,
- 14 machine parts, household appliances, and plumbing fixtures. Formaldehyde also has been
- 15 used for synthesizing polyols, such as pentaerythritol and trimethylolpropane, which are

41

1 used to manufacture polyurethane plastic and alkydes (KEMI 1993); however, no 2 information on formaldehyde release or occupational exposure was found for this use. 3 In 1990, OSHA noted that the plastics industry was the second-largest user of 4 formaldehyde, behind the compressed-wood industry, and that formaldehyde-based 5 resins used in the production process were capable of releasing formaldehyde when 6 subjected to heat or compression during the molding process (OSHA 1990). IRSST 7 (2006) noted that the plastics production industry is continually evolving and that various starting materials and manufacturing processes are used; however, regardless of the 8 9 process or the type of plastic being manufactured, the heating stage will result in the most 10 significant formaldehyde emissions.

Exposure levels depend primarily on the materials used, the processes employed, the presence and efficiency of emissions collection systems, and the level of general ventilation at the production facility (IRSST 2006). Exposure-reduction methods include confinement of production steps that produce formaldehyde emissions, installation of fume hoods above the emission sources, adequate general ventilation, and the use of personal protective equipment for tasks where formaldehyde concentrations are high.

17 IARC (2006) noted that plastic dust and fumes may be present in the atmosphere of 18 molded-plastic plants, and exposures in these facilities are usually considerably higher 19 than those in facilities where the products are used. It also was noted that workers in 20 these plants might have been exposed to pigments, lubricants, and fillers (e.g., asbestos 21 and wood flour) during some production processes. Table 2-8 presents formaldehyde 22 exposure levels for this industry.

		Exposure level mean (range)	Reference
Industry (year measured)	N	(mg/m <sup>3</sup> )	Location
Vinylon production	NR	2.51 (0.95-5.72)	Jin and Zhu 1992 <sup>a</sup>
			China
Hexamine workshop	NR	0.787 (NR)	Dai and Bao 1999 <sup>a</sup>
			China
Polyacetal workshop	NR	1.023 (NR)	Dai and Bao 1999 <sup>a</sup>
			China
Plastics manufacturing (NR)	9	max. < 0.12	Tikuisis et al. 1995 <sup>b</sup>
			Canada
Plastics production (1981–1986)			Heikkila et al. 1991 <sup>b</sup>
Casting of polyacetal resin	10	0.4 (0.08–0.8)	Finland
Casting of UF resin	4	0.5 (0.3–0.6)	
Casting of other plastics	29	< 0.1 (< 0.1–0.3)	
Particleboard and molded plastics plant (NR)	NR	$0.85 (0.21 - 3.6)^{c}$	Horvath <i>et al.</i> 1988 <sup>d</sup>
			United States
Production of molded plastic products (1983–1984)			Stewart et al. 1987a <sup>b</sup>
Phenol resin	10	$0.6^{e} (0.1 - 1.1)$	United States
Melamine resin	13	$11.3^{\rm e}$ ( < 0.01–32.6)	
Molding compound manufacture (1983–1984)			Stewart et al. 1987a <sup>b</sup>
Plant 9, winter	9	3.4 <sup>e</sup> (0.05–8.2)	United States
Plant 9, summer	18	$47.0^{\rm e} (11.7-74.8)^{\rm f}$	
Plant 1, winter	12	$1.8^{e}$ (1.1–2.1)	
Plant 1, summer	24	11.9 <sup>e</sup> (4.7–17.7)	
Plant 8, winter	13	0.4 <sup>e</sup> (0.09–0.9)	
Plant 7, summer	43	$0.4^{e} (0.06-0.8)$	
Plant 2, summer	15	$8.0^{e} (0.4-25.3)$	
Resin and plastic materials production (NR)	NR	1.67 <sup>g</sup> (NR)	NIOSH 1980a <sup>h</sup>
			United States

 Table 2-8. Formaldehyde exposure levels associated with production of plastics and plastic products

<sup>a</sup>Cited in Tang et al. 2009.

<sup>b</sup>Cited in IARC 2006.

<sup>c</sup>Mean and range of TWAs. Data also presented in Table 2-2.

<sup>d</sup>Cited in ATSDR 1999.

<sup>e</sup>Geometric mean.

<sup>f</sup>Some results were affected by the simultaneous occurrence in samples of particulates containing

formaldehyde, leading to high values.

<sup>g</sup>Data also presented in Table 2-1.

<sup>h</sup>Cited in WHO 1989.

#### 1 2.4.6 Embalming

2 Embalming is a procedure that delays the decomposition of a cadaver. To accomplish 3 this, the embalmer injects into either the common carotid or femoral artery usually 12 to 4 18 L of aqueous solutions of formaldehyde at concentrations ranging from about 1.25% 5 to 32%, depending on how much the body has changed since death (IRSST 2006). 6 Formaldehyde is used as a tissue preservative and disinfectant in the embalming fluids, 7 which contain smaller amounts of other chemicals such as methanol, diethylene glycol, 8 propylene glycol, phenol, benzoic acid, and fragrances (IARC 2006, ATSDR 1999). 9 Although embalming was one of formaldehyde's first and best-known uses, it now 10 accounts for less than 1% of total consumption (GI 2006).

11 Exposure to formaldehyde can occur during the solution preparation and during the 12 embalming operation. The main factors affecting exposure include the concentration of 13 formaldehyde in the embalming fluid, the quantity of solution used, the number of 14 workstations and the number of bodies handled daily, physical characteristics of the 15 cadaver (e.g., condition, size, time since death), presence and efficiency of fume hoods or 16 local collection systems at the emission source, and the level of general ventilation. 17 Embalming of a normal intact body generally is completed within 1 to 1.5 hours, with 10 18 to 35 minutes spent using formaldehyde (IRSST 2006). In the case where the cadaver is 19 in an advanced state of putrefaction or has undergone an autopsy, embalming can take up 20 to 3 hours, with up to 2 hours spent using formaldehyde. Formaldehyde-based or 21 paraformaldehyde-based jellies or powders can be prepared and applied to wounds of the 22 cadaver.

IARC (2006) noted that mean formaldehyde exposure levels from embalming operations
are generally around 1 ppm [1.2 mg/m<sup>3</sup>]. Embalming of autopsied bodies generally
results in higher exposure levels than embalming of intact bodies. Airborne formaldehyde
concentrations in seven funeral homes in the United States in 1980 ranged from 0.12 to
0.42 mg/m<sup>3</sup> during the embalming of non-autopsied bodies and from 0.6 to 1.4 mg/m<sup>3</sup>
during the embalming of autopsied bodies (Williams *et al.* 1984, as cited in WHO 1989).
Table 2-9 summarizes exposure levels associated with embalming operations.

1 Methods to reduce formaldehyde exposure include product substitution and modifications 2 of work areas and work practices. Although embalming solutions are available that do 3 not contain formaldehyde (e.g., phenoxyethanol), none is the subject of consensus in the 4 embalming industry (IRSST 2006). Work-station modifications that can reduce exposure 5 include confining difficult embalming cases; physically separating embalming tasks from 6 restoration tasks (i.e., aesthetic care and dressing in funeral homes); installation and 7 proper use of capture equipment at the source, such as hoods over the injection 8 equipment; and design of work stations to ensure adequate ventilation. In one study of 22 9 funeral-service embalming operations, formaldehyde levels were significantly lower (P =10 0.0001) when general ventilation was turned on during the procedure (0.21 ppm [0.26 11  $mg/m^{3}$ ) than when general ventilation was turned off (0.55 ppm [0.68 mg/m<sup>3</sup>]) (Holness 12 and Nethercott 1989). 13 General work practices that will reduce exposure include closing jars promptly when not 14 in use, prompt disposal of formaldehyde soaked rags, proper storage and disposal of 15 products, and periodic equipment inspections (IRSST 2006), and use of personal 16 protective equipment during procedures where formaldehyde concentrations are high. 17 Embalmed cadavers and animals used in gross human and veterinary anatomy 18 laboratories usually are prepared with a formaldehyde-based embalming fluid. During the 19 process of dissection, formaldehyde vapors are emitted from the cadavers, resulting in the 20 exposure of medical students and their instructors to potentially elevated formaldehyde levels (Ohmichi et al. 2006b). Levels have been shown to increase when body-cavity or 21 22 deep structures were being dissected. Levels have also been shown to be higher in the 23 center of the room than in the corners. Various types of exposure reduction technologies 24 have been reported in the literature (Nacher et al. 2007, Ohmichi et al. 2007, Whitehead 25 and Savoia 2008). Tang et al. reported that even when anatomy laboratories were not in use, minimum formaldehyde concentrations were still above  $0.25 \text{ mg/m}^3$  with one 26 measurement as high as  $20.94 \text{ mg/m}^3$ . Table 2-9 provides exposure levels seen in 27 28 anatomy laboratories.

		Exposure level	Poforonco
		mean (range)	
Operation (year measured)	N	(mg/m )	Location
Embalming	1	Γ	
Embalming in funeral homes (NR)			Korczynski 1996 <sup>a</sup>
Personal samples	4	0.19 (NR)	United States
Area samples	4	NR (< 0.1–0.19)	
Embalming (NR)			Korczynski 1994 <sup>a</sup>
Personal samples	48	0.8 (0.1–5.6)	Canada
Area samples	72	0.6 (0.05-8.4)	
Embalming (NR)	75	2.7-3.2 (0.3-10.7) <sup>b</sup>	Stewart et al. 1992 <sup>a</sup>
			United States
Embalming in mortuaries (NR)	NR	1.4 (0.04–3.9)	Lamont Moore and Ogrodnik
		0.2 (0.01–0.6) (TWA)	1986 <sup>a</sup>
			United States
Embalming in funeral homes (1980)			Williams <i>et al.</i> 1984 <sup>a</sup>
Intact bodies	8	$0.4 (0.2-0.4)^{c}$	NP
Autopsied bodies	15	1.1 (0–2.6)	INK
Embalming in funeral homes (NR)	13	1.32, 3.24 (0.24–4.79) <sup>b</sup>	NIOSH 1980c <sup>d</sup>
			United States
Embalming in funeral homes:	187	0.9 (0.1–6.5)	Kerfoot and Mooney 1975 <sup>a,d</sup>
6 facilities (NR)			United States
Anatomy and biology laboratories a	nd autor	psies	
Medical college anatomy labs			
(1998)	2	4.13 (NR)	Li <i>et al.</i> 1999 <sup>e</sup>
(1999)	12	1.07 (NR)	Ye <i>et al.</i> 2000 <sup>e</sup>
(2002)	3	8.35 (5.87–11.13)	Peng et al. 2003 <sup>e</sup>
(2002)	2	NR (12.95–20.94)	Zhang et al. 2007d <sup>e</sup>
(2006)	9	0.33 (0.037–3.98)	Lu <i>et al</i> . 2007 <sup>e</sup>
			China
Medical college teacher offices			
(1998)	2	0.386 (NR)	Li <i>et al.</i> 1999 <sup>e</sup>
(1999)	12	0.2 (NR)	Ye <i>et al.</i> 2000 <sup>e</sup>
(2006)	9	0.04 (NR)	Lu et al. 2007 <sup>e</sup>
			China
Medical college corridors			
(1999)	14	0.315 (NR)	Ye <i>et al.</i> 2000 <sup>e</sup>
(2006)	9	0.056 (NR)	Lu et al. 2007 <sup>e</sup>
			China

Table 2-9.	. Formaldehyde exposure	levels associated	with embalming or	: autopsies or
in anatom	ıy laboratories			

		Exposure level	Deference
		mean (range)	Reference
Operation (year measured)	Ν	(mg/m³)	Location
Anatomy laboratory, dissecting (NR)	NR	NR (0.14–0.76)	Tanaka et al. 2003 <sup>a</sup>
			Japan
Biology laboratory, dissecting (NR)	36	$0.25, 0.63 (0.11 - 1.5)^{b}$	Dufresne <i>et al.</i> 2002 <sup>a</sup>
			Canada
Anatomy laboratory, dissecting (NR)	15	1.1 (0.3–3.1)	Keil et al. 2001 <sup>a</sup>
			United States
Anatomy laboratory, dissecting (NR)	NR	NR (< 5.0)	Burgaz et al. 2001 <sup>a</sup>
			Turkey
Anatomy laboratory, dissecting (NR)	NR	0.27 (0.13-0.41)	Wantke et al. 2000 <sup>a</sup>
			Austria
Anatomy/histology laboratory,	48	3.7 (0.2–11.2)	Kim <i>et al.</i> 1999 <sup>a</sup>
dissecting (NR)			NR
Anatomy laboratory, dissecting (NR)	25	0.5 (0.07–1.28)	Ying et al. 1997, Ying et al. 1999 <sup>a</sup>
	NR	2.9 (NR)	He <i>et al.</i> 1998 <sup>a</sup>
			China
Anatomy laboratory, dissecting (NR)			Akbar-Khanzadeh and Mlynek
Personal samples	44	2.3 (0.4–5.5)	1997 <sup>ª</sup>
Area samples	76	1.2 (0.7–2.1)	United States
Anatomy laboratory, dissecting (NR)			Akbar-Khanzadeh et al. 1994 <sup>a</sup>
Personal samples	32	1.5 (0.09–3.6)	United States
TWA personal samples	NR	0.5 (0.11–1.17)	
Area samples	13	1.7 (1.1–2.2)	
TWA area samples	2	2.0 (1.2-2.8)	
Anatomy laboratory, dissecting (NR)	NR	0.15 (0.07–0.27)	Wantke et al. 1996b <sup>a</sup>
			Austria
Autopsy (1981–1986)	5	0.8 (< 0.1–1.7)	Heikkila et al. 1991 <sup>a</sup>
			Finland
Anatomical theater (1980–1988)	29	1.4 <sup>f</sup> (0.9–2.2)	Triebig et al. 1989
			Germany
Anatomy laboratory, dissecting			Korky <i>et al.</i> 1987 <sup>a</sup>
(1982–1983) Laboratori			United States
Laboratory	NR	NR (8.6–20.3)	
Stock room	NR	NR (2.4–3.2)	
Public hallway	NR	NR (< 1.2)	
Animal dissection laboratory (NR)	24	0.18, 0.22 (0.13–1.25) <sup>b</sup>	Blade 1983 <sup>d</sup>
			NR

		Exposure level mean (range)	Reference
Operation (year measured)	Ν	(mg/m <sup>3</sup> )	Location
Autopsy (NR)			Coldiron <i>et al.</i> 1983 <sup>a</sup>
Personal samples	27	1.7 (0.5–4.0)	United States
Area samples	23	5.0 (0.1–16.7)	
Anatomy classrooms, 1998	4	2.514 (NR)	Li <i>et al</i> . 1999 <sup>e</sup>
Biology teaching (NR)	8	9.96 (3.3–17.76)	EPA 1981 <sup>d</sup>
			United States
Pathology autopsy room (NR)	10	5.76 (0.07–9.5)	Covino 1979 <sup>d</sup>
			NR
Pathology autopsy room (NR)	6	5.22 (2.64–9.5)	NIOSH 1979b <sup>d</sup>
			United States
Autopsy room (NR)			Makar et al. 1975 <sup>d</sup>
Personal sampling for a resident	10	1.9 (NR)	NR
Personal sampling for a pathologist	9	1.5 (NR)	
Personal sampling for a technician	2	0.68 (NR)	
Area sampling for assistants	23	0.86 (0.16–16.28)	

<sup>a</sup>Cited in IARC 2006.

<sup>b</sup>Range of means and full range across two to three datasets.

<sup>c</sup>No explanation provided for the mean being equal to the high end of the range.

<sup>d</sup>Cited in WHO 1989.

<sup>e</sup>Cited in Tang *et al.* 2009. <sup>f</sup>Median.

- 1 2.4.7 Histology
- 2 Histopathology laboratories receive organ, tissue, or cell specimens in which to study
- 3 structural modifications in support of diagnosis and prognosis of disease, and formalin is
- 4 commonly used to preserve these samples (IARC 2006, IRSST 2006). The main steps in
- 5 the process include preparing formaldehyde solutions (diluting the formalin solution to
- 6 roughly 4% formaldehyde), macroscopic examination of the specimen with the naked
- 7 eye, placing the samples in cassettes, and microscopic observation (IRSST 2006).
- 8 Specific tasks that may result in exposure to formaldehyde include prepararing the
- 9 formalin solution, handling and disposing of specimens, handling waste (such as draining
- 10 specimens), handling and cleaning used jars, handling bags of medical waste,
- 11 maintaining equipment, and recycling and discarding formalin solution. Equipment leaks
- 12 are another potential source of exposure (e.g., leaks from the tissue preparer,
- 13 formaldehyde recycler, specimen storage, and storage of new and waste formaldehyde

1 solutions). Workers who might be occupationally exposed include pathologists,

2 technicians, technical assistants, and administrative personnel (IRSST 2006).

3 IARC (2006) noted that the typical mean formaldehyde exposure level in pathology

4 operations is approximately 0.5 ppm [0.1 mg/m<sup>3</sup>]. Table 2-10 summarizes exposure

5 levels associated with histology operations.

6 One way in which formaldehyde exposure can be reduced in histology operations is

7 through substitution of other chemicals. Because of increasing concern about health

8 effects associated with formaldehyde exposure, a number of proprietary fixatives have

9 been developed that do not contain formaldehyde. Although a number of these fixatives

10 have been successfully used in the United States, none are the subject of consensus, and

11 formaldehyde-based fixatives generally are considered superior (IRSST 2006, Titford

12 and Horenstein 2005). Other exposure-reduction methods include the use of hoods and

13 other ventilation methods and wearing of personal protective equipment for tasks where

14 the formaldehyde concentration is high (IRSST 2006).

		Exposure level	Deference
		mean (range)	Reference
Operation (year measured)	N	(mg/m³)	Location
Hospital pathology rooms			
(2005)	8	NR (0.086–2.0)	Li <i>et al</i> . 1999 <sup>a</sup>
(2003)	40	NR (0.184–0.931)	Cheng et al. 2004 <sup>a</sup>
(2003)	85	1.6 (0.18–5.84)	Fan <i>et al.</i> $2006^{a}$
			China
Histology laboratory (NR)			Shaham <i>et al.</i> 2002 <sup>b</sup>
Laboratory assistants/technicians	NR	0.5 (0.05–0.9)	Israel
Physicians and orderlies	NR	2.8 (0.9–7.0)	
Pathology laboratory (NR)	10	NR (max. < 2.5)	Burgaz et al. 2001 <sup>b</sup>
			Turkey
Medical college specimen workshops	2	1.1 (NR)	Li <i>et al</i> . 1999 <sup>a</sup>
(1998)			China
Medical college specimen rooms (1998)	2	12.783 (NR)	Li <i>et al</i> . 1999 <sup>a</sup>
			China
Histopathology teaching laboratory (NR)	16	0.4 (NR)	Tan <i>et al.</i> 1999 <sup>b</sup>
			United States
Histology laboratory (NR)			Shaham <i>et al</i> . 1996a, 1996b <sup>b</sup>
Area samples	NR	NR (1.7–2.0)	Israel
Personal samples	NR	NR (3.4–3.8)	
Hospital histopathology laboratories	80	0.6 (0.01–9.1)	Heikkila et al. 1991 <sup>b</sup>
(1981–1986)			Finland
Pathology laboratories (1980–1988)	21	0.6 <sup>c</sup> (< 0.01–1.6)	Triebig et al. 1989 <sup>b</sup>
			Germany
Histology laboratory, tissue specimen	NR	NR (0.25–2.3)	Kilburn <i>et al.</i> 1985a <sup>b</sup>
preparation and sampling (NR)			United States
Pathology laboratory (1980s)	13	0.7 (NR)	Rosen et al. 1984 <sup>b</sup>
			Sweden

Table 2-10. Formaldehyde exposure levels associated with histology and pathology laboratories

NR = not reported.<sup>a</sup>Cited in Tang *et al.* 2009. <sup>b</sup>Cited in IARC 2006. <sup>c</sup>Median.

1 2.4.8 Construction-related exposures

- 2 There are many potential sources of exposure to formaldehyde in the construction
- 3 industry; however, data are limited on exposure levels for most of these sources.
- 4 Construction workers who varnish floors can have high exposures. IARC (2006) noted

1 that formaldehyde levels during varnishing with UF-based varnishes have been measured

2 at levels ranging from 2.5 to  $6.2 \text{ mg/m}^3$  during a 30-minute application period, and that

3 workers may apply 5 to 10 coats per day. These workers are also potentially exposed to

4 wood dust and various solvent vapors from varnishes, putties, and adhesives.

5 Working with UFFI or fiberglass insulation manufactured using formaldehyde-based

6 resins also can result in formaldehyde exposure (IARC 2006); however, no data on

- 7 exposure levels associated with this activity.
- 8 Since the 1980s, glass-fiber mats have become an important material for roof shingles,

9 asphalt roofing tiles, and roll roofing (TIG 2005). UF and occasionally PF resins are used

10 as binders to hold the glass fibers together until an asphalt coating is applied. No

11 information was found on exposure levels from their use.

12 Machining of wood-based composites and other formaldehyde-containing wood products

13 are other sources of exposure in the construction industry; however, IARC (2006) noted

14 that formaldehyde exposure levels from this activity are consistently low. Formaldehyde

15 exposure levels associated with construction-related activities are presented in

16 Table 2-11.

		Exposure level mean (range)	Reference
Operation (year measured)	Ν	(mg/m <sup>3</sup> )	Location
Varnishing parquet with UF varnish (1976 & 1987)	16	3.6, 5.3 (0.4–8.1) <sup>a</sup>	Heikkila <i>et al.</i> 1991 <sup>b</sup> and Riala and Riihimaki 1991 <sup>b</sup>
			Finland
Insulating buildings with UFFI	6	0.2 (NR)	Rosen et al. 1984 <sup>b</sup>
(1980s)			Sweden
UFFI dealing and installation (NR)	NR	NR (0.08–2.4)	Herrick et al. 1983 <sup>c</sup>
			NR
UFFI dealing and installation (NR)	82	1.26–1.87 (0.36–6.36) <sup>d</sup>	NIOSH 1979b <sup>c</sup>
			United States
Fiberglass insulation installation	13	0.028 (0.008-0.04)	NIOSH 1980a <sup>c</sup>
(NR)			United States
Sawing particleboard at construction	5	< 0.6 (NR)	FIOH 1994 <sup>b</sup>
site (1967)			Finland

Table 2-11. Formaldehyde levels associated with construction-related activities

<sup>a</sup>Means and full range across two studies.

<sup>b</sup>Cited in IARC 2006.

<sup>c</sup>Cited in WHO 1989.

<sup>d</sup>Range of means and full range across three datasets.

### 1 2.4.9 Fiberglass and mineral-wool insulation manufacturing

- 2 PF resins commonly are used to bind fiberglass, mineral wool, or shredded waste
- 3 products such as cotton, wool, or polyester for use as structural and acoustical insulation
- 4 for residential and commercial buildings, pipes, and industrial equipment. Fiberglass
- 5 insulation accounts for 90% of formaldehyde consumption in this industry (Bizzari
- 6 2007). In fiberglass and mineral-wool insulation, UF resins often are used in conjunction
- 7 with PF resins to inhibit the burning potential of the PF resins (TIG 2005).
- 8 Fiberglass insulation manufacturing involves six general steps: melting glass, spinning
- 9 the molten glass into fibers, cooling and coating the fibers with a binder, forming the
- 10 fibers into a pad, curing the binder (i.e., heating at 400°F to 600°F to set the binder), and
- 11 packaging the insulation (Milton *et al.* 1996). The primary sources of formaldehyde
- 12 release are from the fiber-coating process and the curing process. IARC (2006) described
- 13 measurements taken in the 1980s and noted that very high levels occasionally were
- 14 measured in close proximity to these two operations. Measured formaldehyde levels

- 1 associated with fiberglass insulation are presented in Table 2-12. No data were found on
- 2 exposure levels associated with manufacture of insulation from materials other than
- 3 fiberglass or synthetic vitreous fibers.

Industry		Exposure level mean (range)		Reference
(year measured)	Ν	(mg/m <sup>3</sup> )	Comment	Location
Fiberglass manufacturing plant (NR)				Milton <i>et al.</i> 1996
Area sampling	50	0.05–0.52 (max. = 1.25)	Range of means for area sampling at four different locations; maximum concentration found at forehearth.	United States <sup>a</sup>
Personal sampling	197	0.022–0.086 (NR)	Range of mean TWA concentrations from personal sampling of 37 workers.	
Synthetic vitreous fiber plant (1981–1986)	60	0.11, 0.25 (0.01–1.7)	Means and full range across production and form-pressing operations.	Heikkila <i>et al.</i> 1991 <sup>b</sup> Finland
Insulation manufacture				Tao <i>et al</i> .
(1989, summer)	8	NR (0.15–0.39)		1990°
(1989, winter)	8	NR (0.64–0.93)		China
Synthetic vitreous fiber plant (1980s)	20	0.19, 0.20 (NR)	Mean values for production and form-	Rosen <i>et al.</i> 1984 <sup>b</sup>
			pressing operations.	Sweden

Table 2-12. Formaldehyde exposure levels associated with fiberglass manufacturing

<sup>a</sup>Cited in ATSDR 1999 and IARC 2006; data presented here are from the original article, which was reviewed because of questions raised during review of IARC and ATSDR documents. <sup>b</sup>Cited in IARC 2006. <sup>c</sup>Cited in Tang *et al.* 2000

<sup>c</sup>Cited in Tang *et al.* 2009.

- 4 2.4.10 Firefighting and other combustion-related exposures
- 5 As noted in Section 2.2.2, combustion processes are one of the major sources of
- 6 formaldehyde in the environment. IARC (2006) reviewed three studies that assessed
- 7 firefighters' levels of personal exposure to formaldehyde during various stages of
- 8 firefighting, with concentrations measured up to  $10.2 \text{ mg/m}^3$  (see Table 2-13).
- 9 Formaldehyde was detected in 6 of 24 samples (25%) in one study and 73% of samples
- 10 in a second study; the percentage was not reported for the third study. In a comprehensive
- 11 air-monitoring study to characterize exposure of firefighters during 25 structure fires,

formaldehyde levels exceeded 0.1 ppm  $[0.12 \text{ mg/m}^3]$  [which was cited as the National 1 2 Institute for Occupational Safety and Health (NIOSH) ceiling recommended exposure 3 limit (see Section 2-7)] at 22 of the 25 fires. Firefighters might also be exposed while 4 fighting wildfires. Results of two studies, in which formaldehyde was detected in all 5 samples, showed concentrations that ranged from 0.02 to 0.42 mg/m<sup>3</sup>. 6 Because formaldehyde is emitted from internal combustion engines, workers in any 7 occupation that involves exposure to exhaust from automobile or other internal 8 combustion engines potentially are exposed to formaldehyde. In a study of occupational 9 exposure to volatile organic compounds (VOCs) and aldehydes in the U.S. trucking 10 industry, Davis et al. (2007) measured formaldehyde at the perimeter of trucking terminal 11 yards (i.e., considered background levels), at indoor work areas (i.e., at loading docks and 12 mechanic shops), and in on-road truck cabs (i.e., driver exposures). The mean background level was reported to be 3.33  $\mu$ g/m<sup>3</sup> [0.003 mg/m<sup>3</sup>], and higher exposure 13 levels were reported for the indoor work areas than in on-road truck cabs (Table 2-13). 14 15 Zhang et al. (2003) (as cited in IARC 2006) reported a slightly higher mean level for automobile garages  $(0.04 \text{ mg/m}^3)$  than the mean level for the mechanic shop (13.72)16  $\mu g/m^3$  [0.0137 mg/m<sup>3</sup>]) reported by Davis *et al.* Pang and Mu (2007) assessed carbonyl 17 18 exposures from public vehicles in Beijing, China, noting that taxi and bus drivers can 19 have high levels of formaldehyde exposure as a result of high concentrations and long 20 work hours. They also noted that in-vehicle carbonyl concentrations were loosely 21 associated with vehicular service years and type of fuel used. All drivers were asked to 22 refrain from smoking during this study. Formaldehyde exposure levels for these studies 23 are presented in Table 2-13.

IARC (2006) reported exposure levels ranging up to 0.6 mg/m<sup>3</sup> for lumberjacks using
chainsaws and up to 0.021 mg/m<sup>3</sup> in personal air samples from French policemen
working close to traffic. Pilidis *et al.* (2009) reported exposure levels for policemen in
outdoor environments (car, motorcycle, and foot patrol, guards, and traffic regulation)
that ranged from about 0.003 to 0.02 mg/m<sup>3</sup>.

		Exposure level	
		Mean (range)	Reference
Operation (year measured)	Ν	(mg/m³)	Location
Firefighting, city fire (1998)	96	0.31 (0.02–1.5)	Bolstad-Johnson <i>et al.</i> 2000 <sup>a</sup>
			United States
Firefighting, city fire (NR) Knockdown <sup>b</sup> Overhaul <sup>b</sup>	(22 fires)	NR (ND–9.8) NR (ND–0.5)	Jankovic <i>et al.</i> 1991 <sup>a</sup> United States
Inside mask		NR (ND-0.4)	
Firefighting, city fire (1986)	24	0.68 (0.1–10.2) <sup>c</sup>	Brandt-Rauf <i>et al.</i> 1988 <sup>a</sup>
			United States
Wildland fire fighting (1990 and 1989)	35	0.06, 0.16 (0.02–0.42) <sup>d</sup>	Reh <i>et al.</i> 1994 <sup>a</sup> and Materna <i>et al.</i> 1992 <sup>a</sup>
			United States
Trucking industry (2004–2006)			Davis et al. 2007
In cab (nonsmokers)	234	0.0083 (NR)	United States
In cab (smokers)	62	0.0096 (NR)	
Loading dock	65	0.0254 (NR)	
Mechanic shop	17	0.0137 (NR)	
Public transportation vehicles			Pang and Mu 2007
Taxis	35	0.024, 0.028 (0.013–0.034)	China
Buses	15	0.016-0.04 (0.013-0.094)	
Chain-sawing (NR)	NR	< 0.1 (< 0.1–0.6)	Heikkila <i>et al.</i> 1991 <sup>a</sup>
			Finland
Chain-sawing (NR)	NR	0.06 (0.03–0.13)	Hagberg <i>et al.</i> 1985 <sup>a</sup>
			Sweden
Automobile garage (NR)	53	0.04 (NR)	Zhang et al. 2003 <sup>a</sup>
			NR
Policemen working close to traffic			Maitre et al. 2002 <sup>a</sup>
center (NR)			France
Summer	[32] <sup>e</sup>	$0.014^{\rm f}$ (NR)	
Winter	[32] <sup>e</sup>	$0.021^{\rm f}$ (NR)	

# Table 2-13. Formaldehyde exposure levels associated with firefighting and other combustion sources

Operation (year measured)	N	Exposure level Mean (range) (mg/m³)	Reference Location
Policemen (2006)			Pilidis et al. 2009
Vehicle patrol	5	~0.024–0.034 (0.020–0.038) <sup>g</sup>	Greece
Motorcycle patrol	4	~0.027-0.034 (0.020-0.039) <sup>g</sup>	Gibber
Foot patrol	2	~0.018, 0.019 (0.017–0.030) <sup>g</sup>	
Guards	2	~0.014, 0.023 (0.012–0.026) <sup>g</sup>	
Traffic regulation	3	~0.021–0.037 (0.018–0.042) <sup>g</sup>	

NR = not reported, ND = not detected.

<sup>a</sup>Cited in IARC 2006.

<sup>b</sup>"Knockdown" is when the main body of the fire is brought under control; "overhaul" refers to searching for and extinguishing hidden fires.

<sup>c</sup>The mean and range do not include 18 values that were noted as 0 in the original paper.

<sup>d</sup>Means and full range across two studies.

<sup>e</sup>Personal sampling performed for 8 policemen, four days each in summer and winter.

<sup>f</sup>Median.

<sup>g</sup>Estimated from graph.

1 2.4.11 Agriculture and aquaculture

2 In agricultural settings, formaldehyde has been used as a preservative for fodder,

3 disinfectant in brooding houses, sterilant in mushroom houses, and preservative for

4 produce (IARC 2006, ATSDR 1999). Levels as high as 9.6 mg/m<sup>3</sup> have been reported

5 when formaldehyde is used for disinfection of eggs in brooding houses; however, IARC

6 (2006) noted that annual exposures are likely to be low, because the operation is

7 performed only intermittently (roughly 5 to 10 times per year). Formalin solutions have

8 been used in aquaculture to treat fish eggs to control infection (IARC 2006), with

9 treatment times ranging from 15 to 90 minutes. Urea-formaldehyde concentrates are used

10 in the manufacture of controlled-release fertilizers (Bizzari 2007); however, no

11 information was found on exposure to formaldehyde from application of these products.

12 [Although there is the potential for occupational exposure from agricultural applications

13 of controlled-release fertilizers, their primary uses are nonagricultural, such as on lawns

14 and turfs and in nurseries (Bizzari 2007)]. Formaldehyde exposure levels associated with

15 agriculture and aquaculture are presented in Table 2-14.

Operation (year measured)	N	Exposure level mean (range) (mg/m³)	Reference Location
Handling of fodder (1982)	NR	NR (0.03–0.5)	Heikkila <i>et al</i> . 1991 <sup>a</sup>
			Finland
Disinfection of eggs (1981–1986)	11	3.2 (0.3–9.6)	Heikkila et al. 1991 <sup>a</sup>
			Finland
Mushroom farming (NR)	18	3.22 (ND-> 12.0) <sup>b</sup>	NIOSH 1980b <sup>c</sup>
			United States
Fish hatchery, treating fish eggs (NR)			Lee and Radtke 1998 <sup>a</sup>
Personal monitoring of 6 employees	6	NR (NQ-1.0)	United States
Area monitoring during treatment operations	6	NR (< 0.062–0.84)	
TWA concentrations	6	0.02 (0.007-0.05)	

 Table 2-14. Formaldehyde exposure levels associated with agriculture and aquaculture

NR = not reported, NQ = not quantifiable, ND = not detected.

<sup>a</sup>Cited in IARC 2006.

<sup>b</sup>Upper end of range reported as "12+" in WHO 1989. Range is across three datasets; the mean was reported for only one of these datasets.

<sup>c</sup>Cited in WHO 1989.

### 1 2.4.12 Office buildings and nonindustrial work places

- 2 There are numerous sources of formaldehyde in office buildings, restaurants, commercial
- 3 buildings, and other nonindustrial work places. These sources include paint and varnish,
- 4 carpeting, wallpaper, insulation, furniture, and laser printers (IARC 2006, ATSDR 1999).
- 5 In a study that assessed exposure of policemen performing several types of activities (i.e.,
- 6 vehicle or foot patrol, traffic regulation, guarding outside the police station building, and
- 7 office work), Pilidis *et al.* (2009) found that officers working indoors had significantly
- 8 higher exposure than those working outdoors. Table 2-15 presents exposure-level data for
- 9 offices and other nonindustrial work places. IARC (2006) noted that laser printers have
- 10 been found to be a source of formaldehyde exposure as a result of ozonolysis reactions
- 11 with VOCs emitted from the toner. IARC (2006) also noted that newer-technology laser
- 12 printers did not produce detectable levels of formaldehyde.

		Exposure level	Reference
Operation (year measured)	N	(mg/m <sup>3</sup> )	Location
Office buildings: 23 buildings for which air- quality complaints had been filed but for which there were no clear, unusual sources for chemical pollutants (2001–2006)	76	0.011 (0.044 max)	Salonen <i>et al.</i> 2009 Finland
Offices: summary of results from 9 studies (1996–2005)	351	0.256 (0.058–2.25) (Overall mean and range of individual means)	Tang <i>et al.</i> 2009 China
Office buildings: 5 buildings, 8-hour average concentrations (NR)	54	0.14–1.19 (NR)	Wu <i>et al.</i> 2003 <sup>a</sup> Taiwan, China
Office buildings: 6 buildings (1996–1997)	72	0.002–0.013 <sup>b</sup> (NR)	Reynolds <i>et al.</i> 2001 <sup>a</sup> United States
Offices (NR) Conventional offices (18 sites) Portable office buildings (20 sites)	NR 40	0.027 (0.012–0.096) 1.4 (0.52–2.6)	Dingle <i>et al.</i> 2000 <sup>a</sup> Australia
Offices (1995–1996) Recently painted with low-emitting paint Three months after painting Control	NR	0.018 (0.016–0.02) 0.008 (0.007–0.01) 0.008 (0.008–0.009)	Wieslander <i>et al.</i> 1999a <sup>a</sup> Sweden
Offices (1995)	11	0.04 (0.012–0.1)	Brickus <i>et al.</i> 1998 <sup>a</sup> Brazil
Nonindustrial workplaces and restaurants (1995)	12	0.02 (0.005–0.06)	Miguel <i>et al.</i> 1995 <sup>a</sup> Brazil
Office work (NR)	NR	0.086 <sup>c</sup> (0.086–0.16) <sup>d</sup>	Holmström <i>et al.</i> 1989b <sup>e</sup> NR
Offices (1981–1984)	25	0.08 <sup>c</sup> (NR)	Shah and Singh 1988 <sup>a</sup> United States
Office building (NR) Nonsmoking office Office that allowed smoking	NR	NR (ND-0.27) NR (ND-0.74)	Sterling <i>et al.</i> 1987 <sup>e</sup> NR
Offices Aged 1 to 3 years Aged 11 to 43 years	NR	0.143 (NR) 0.087 (NR)	Kalinic <i>et al.</i> 1985 <sup>f</sup> Yugoslavia
Offices (NR) Smokers Nonsmokers	NR	NR (0.01–0.13) NR (0.02–0.1)	Prescher 1984 <sup>f</sup> Germany
Office work (NR)	48	< 0.05, 0.07 (0.02–0.14) <sup>g</sup>	Blade 1983 <sup>f</sup> NR

Table 2-15. Formaldehyde exposure levels in offices and other nonindustrial work places

Operation (year measured)	N	Exposure level mean (range) (mg/m³)	Reference Location
Offices and commercial buildings: 4 establishments (NR)	NR	NR (0.012–1.24)	Konopinski 1983 <sup>e</sup> United States
Commercial buildings (NR)	NR		Kuljac 1983 <sup>f</sup>
Offices		1.083 (NR)	Yugoslavia
Stores		2.60 (NR)	1 480014114
Furniture stores		0.15 (NR)	

NR = not reported, ND = not detected.

<sup>a</sup>Cited in IARC 2006.
<sup>b</sup>Geometric means.
<sup>c</sup>Median.
<sup>d</sup>The median is a year-round median concentration, but the range is only for late summer.
<sup>e</sup>Cited in ATSDR 1999.
<sup>f</sup>Cited in WHO 1989.
<sup>g</sup>Means for two studies. The range is from one study; the other study reported the range as < 0.05 mg/m<sup>3</sup>.

1 2.4.13 Other occupational exposures

2 Formaldehyde has been used in the treatment of furs and leather (IARC 2006). Its use in

3 the treatment of furs resulted in the highest formaldehyde exposure levels for all jobs and

4 industries studied in a large Swedish survey in the early 1980s. The eight-hour TWA

5 concentration of formaldehyde was reported to be 1.0 to  $2.0 \text{ mg/m}^3$ , and high peak

6 exposures occurred several times per day. Formaldehyde concentrations of 0.5 to 7 ppm

7 [0.61 to 8.6 mg/m<sup>3</sup>] have been measured in leather-tanning facilities (ATSDR 1999), and

8 a mean level of 0.3 mg/m<sup>3</sup> has been reported for taxidermy operations in Sweden (Rosén

9 *et al.* 1984).

- 10 Formaldehyde has been used extensively in hospitals and healthcare facilities (IARC
- 11 2006). ATSDR (1999) noted that numerous types of healthcare professionals (e.g.,

12 pharmacists, physicians, veterinarians, dentists, nurses) can be exposed to formaldehyde

- 13 vapors during the preparation, administration, or cleanup of various medicines. IARC
- 14 (2006) reported exposure levels associated with the use of formaldehyde as a disinfectant
- 15 in hospitals, showing mean levels ranging from 0.06 to  $1.1 \text{ mg/m}^3$ , with levels as high as
- 16  $6.3 \text{ mg/m}^3$ . Formaldehyde levels as high as  $0.288 \text{ mg/m}^3$  were measured in a hospital
- 17 operating room where it was used as a disinfectant (Dascalaki et al. 2008). Formaldehyde
- 18 has also been detected in the plume of surgical smoke produced by electrocautery,
- 19 harmonic scalpel, and argon beaming (Krones *et al.* 2007).

1 Formaldehyde has been used as a biocide in the oil processing industry (Steinsvag *et al.* 

2 2007); however, the authors noted that formaldehyde appears to have been replaced by

3 other biocides and phased out before 2002. Mean measured airborne exposure levels

4 were 0.13 mg/m<sup>3</sup> (range = 0.06 to 0.29 mg/m<sup>3</sup>) for personal sampling and 0.21 mg/m<sup>3</sup>

5  $(range = 0.05 \text{ to } 0.53 \text{ mg/m}^3)$  for stationary monitoring of Norwegian offshore oil drilling

6 installations during 1999 and 2000.

7 In a study assessing exposure of nail technicians to formaldehyde and toluene, a mean

8 airborne formaldehyde exposure level of 0.022 ppm [0.027 mg/m<sup>3</sup>] was calculated based

9 on personal air sampling at 30 nail salons in California (McNary and Jackson 2007).

10 Formaldehyde has been measured in studies assessing exposure of workers to

11 metalworking fluids in a secondary aluminum plant (Godderis *et al.* 2008) and in

12 machine shops (Lillienberg *et al.* 2008). Godderis *et al.* reported airborne formaldehyde

13 at a concentration of 0.03 mg/m<sup>3</sup>, and Lillienberg *et al.* reported mean levels of 0.003,

14 0.012, and 0.128  $mg/m^3$  for three facilities (the full range across the three facilities was

15 0.001 to 0.154 mg/m<sup>3</sup>). Lillienberg *et al.* suggested that use of recirculating air probably

16 was responsible for the higher levels observed in one machine shop. Godderis *et al.* 

17 postulated that the airborne formaldehyde in the aluminum plant originated either from

18 the combustion of metalworking fluids or from formaldehyde-releasing triazines used as

19 biocides.

20 Formaldehyde levels in spacecraft have been found to consistently exceed 0.05 mg/m<sup>3</sup>

21 (IARC 2006). ATSDR (1999) noted that the laser cutting of felt, woven fabrics, formica,

22 plexiglass, and acrylic materials has been found to release formaldehyde; however, no air

23 levels were identified for these activities. Concentrations ranging from less than 0.01 to

 $24 \quad 2.0 \text{ mg/m}^3$  have been measured at coal and pitch-coking plants in the former

25 Czechoslovakia. Levels up to  $1.1 \text{ mg/m}^3$  have been measured at plants producing

26 photographic film.

### 27 **2.5 Environmental occurrence and fate**

28 Formaldehyde is ubiquitous in the environment and can occur in outdoor and indoor air,

29 drinking water, groundwater, surface water, sediment, soil, and food. This section

1 discusses the sources of formaldehyde, its fate and transport, and occurrence of

2 formaldehyde in air (Section 2.5.1), water (Section 2.5.2), land and soil (Section 2.5.3),

3 and food (Section 2.5.4).

4 A potential source of contamination for all environmental media and for general 5 population exposure is from inadvertent spills of formaldehyde-containing materials. A 6 2009 search of the National Response Center (NRC 2009) on-line database using the 7 keyword "formaldehyde" yielded 802 results. The NRC serves as the sole national point 8 of contact for the reporting of all oil, chemical, radiological, biological, and etiological 9 (i.e., biologically hazardous) spills into the environment anywhere in the United States 10 and its territories. The level of information provided in the query results was not 11 sufficient to estimate the extent of environmental contamination or the number of people 12 exposed; however, it does suggest the potential for environmental contamination and 13 general public exposure from inadvertent spills of formaldehyde or chemical mixtures 14 containing formaldehyde.

### 15 2.5.1 Air

16 In air, formaldehyde is a gaseous pollutant that is produced both naturally and from 17 human activities and occurs as a primary or secondary pollutant. In outdoor air, primary 18 sources include direct emissions of formaldehyde from industrial processes and products 19 and its release during the combustion of organic materials. Occurrence of formaldehyde 20 as a secondary pollutant results from the photochemical breakdown of hydrocarbons, 21 which occur both naturally and as a result of human activities. In indoor air, the main 22 sources of formaldehyde are indoor combustion sources, including tobacco smoke, and 23 off-gassing from various materials.

Because formaldehyde air levels generally are higher in occupational settings than in
nonoccupational settings, this section reports air concentrations in units of micrograms
per cubic meter rather than the milligrams per cubic meter used to describe occupational
exposure (Section 2.4). If the source document reported concentrations in parts per
billion, values were multiplied by a conversion factor of 1.23.

1 Four studies were found in the literature that estimated time-weighted daily exposure 2 levels for indoor and outdoor exposures. Probabilistic methods were used to estimate a 3 24-hour TWA exposure concentration for the general Canadian public, taking into 4 account the amount of time spent indoors and outdoors and the associated formaldehyde 5 concentrations (WHO 2002). Although this study applies specifically to the Canadian 6 population, it was noted that the sources of formaldehyde are ubiquitous and are likely 7 similar in most countries, and the overall magnitude of relative contributions from indoor 8 air and outdoor air are expected to be similar in other parts of the world. Based on two 9 different assumptions regarding the statistical distribution of formaldehyde concentrations, mean values were 24 and 29  $\mu$ g/m<sup>3</sup>, median values were 33 and 36  $\mu$ g/m<sup>3</sup>, 10 11 and 95th-percentile values were 94 and 80  $\mu$ g/m<sup>3</sup>.

12 More recently, in a review of production, consumption, exposure levels, and health

13 effects of formaldehyde in China, Tang *et al.* (2009) provided data from numerous

14 studies that had measured formaldehyde air levels. From these data, Tang *et al.* 

15 calculated average concentrations of formaldehyde in various locations including outdoor

16 air, in new remodeled homes, new office buildings, and public places. Based on these

17 levels and time-activity pattern assumptions, the authors estimated an effective

18 concentration for a hypothetical person of 0.21 mg/m<sup>3</sup> during workdays and 0.17 mg/m<sup>3</sup>

19 over the course of the weekend. The authors noted that this level of exposure was higher

20 than the WHO recommended indoor level of  $0.1 \text{ mg/m}^3$ . They further noted that higher

21 levels would be associated with occupational exposures: 0.58 mg/m<sup>3</sup> per day for

industrial exposures and  $0.61 \text{ mg/m}^3$  per day for professional exposures (e.g., exposures

associated with anatomy labs or pathology).

24 Dodson *et al.* (2007) developed a personal exposure model using VOC data (including

25 data on formaldehyde) collected for teachers and office workers as part of the Boston

26 Exposure Assessment in Microenvironments study. Included in the final model were data

27 on participants' time-activity and concentration measurements for residential outdoor,

28 residential indoor, and workplace microenvironments, along with average concentrations

29 in various dining, retail, and transportation microenvironments. The authors noted that

30 even with the full model, exposures to formaldehyde were not fully characterized, based

1 on comparison with personal monitoring data; they emphasized the need for additional 2 time-activity and concentration data. Measured time-weighted personal exposure levels ranged from roughly 8 to 88  $\mu$ g/m<sup>3</sup> [0.008 to 0.088 mg/m<sup>3</sup>] across 62 observations. 3 4 Boström *et al.* (1994) derived ratios of nitrogen oxide  $(NO_x)$  levels to levels of other 5 pollutants in urban air, including formaldehyde, and used time-activity data together with 6 NO<sub>x</sub> levels to estimate exposure of the Swedish population to various pollutants. The overall mean exposure level for formaldehyde was estimated at 1.2  $\mu$ g/m<sup>3</sup> [0.001 mg/m<sup>3</sup>]. 7 8 The remainder of this section discusses outdoor air and indoor air separately. 9 2.5.1.1 Outdoor air 10 Formaldehyde in outdoor air has many natural and anthropogenic sources. Natural 11 sources of formaldehyde include forest fires, animal wastes, microbial products of 12 biological systems, and plant volatiles. In Riverside, CA, airborne formaldehyde levels 13 were twice as high during a wildfire as after the wildfire had ended (Na and Cocker 14 2008). However, the majority of formaldehyde in outdoor air is from anthropogenic 15 activities, primarily combustion processes; therefore, higher levels are seen in urban 16 environments than in rural environments (ATSDR 1999, WHO 2002). Major 17 anthropogenic sources of formaldehyde in outdoor air include power plants, refineries, 18 manufacturing facilities, incinerators, automobile exhaust, and other combustion sources. 19 In 2007, U.S. industrial air emissions of more than 9.2 million pounds of formaldehyde 20 were reported to the U.S. EPA's Toxics Release Inventory (TRI) as either fugitive 21 (1 million pounds) or point-source (8.2 million pounds) emissions (TRI 2009). Total air 22 emissions reported to TRI trended downward slightly between 1988 and 2007, with a 23 maximum of 13.2 million pounds in 1989 and a minimum of 9 million pounds in 2006. 24 Reported emissions were lowest in 2005, 2006, and 2007. 25 It has been suggested that formaldehyde levels due to secondary formation might be 26 much larger than levels from direct emissions. One study reviewed by the World Health 27 Organization (WHO 2002), estimated that 70% to 90% of atmospheric formaldehyde was 28 the result of secondary formation.

63

1 Formaldehyde is not present in gasoline; however, it is a product of incomplete 2 combustion and is therefore released from internal combustion engines (WHO 2002). 3 Automobiles are a major source of formaldehyde in outdoor air through direct 4 formaldehyde emissions and through emission of precursors that form formaldehyde via 5 atmospheric oxidation. Formaldehyde levels have been found to be correlated with traffic 6 activity (ATSDR 1999). In the mid 1970s, the U.S. EPA estimated that automobiles 7 emitted about 610 million pounds of formaldehyde annually. Emission levels depend on 8 the fuel composition, the type of engine, the type of emission controls, the operating 9 temperature, and the age and state of repair of the vehicle; therefore, emission rates are 10 quite variable. The introduction of catalytic converters reduced automobile emissions of 11 formaldehyde; however, the use of oxygenated fuels increases emissions. With the 12 increased use of both catalytic converters and oxygenated fuels, the net effect on 13 formaldehyde emissions is uncertain. Tractors and back-up generators are additional 14 sources of substantial amounts of formaldehyde in outdoor air (Sawant et al. 2007). 15 In a study of emissions from diesel engines operating on standard diesel fuel or on 16 various blends of biodiesel, Liu et al. (2009a) reported that emissions of carbonyl 17 compounds (including formaldehyde) increased when the engines were run on biodiesel 18 fuels; however, the total concentration of the emitted carbonyls did not increase with 19 biodiesel content. Sawant et al. (2007) noted that for tractors and back-up generators, 20 engine operating mode and application appear to strongly influence the absolute mass

21 emission rate of carbonyls (including formaldehyde); however, they do not appear to

exert as strong an influence on the relative mass emission rates of individual carbonylcompounds.

No consistent seasonal variation has been demonstrated for formaldehyde levels, which could be explained in part by the fact that photo-oxidation is both an important source of formaldehyde (i.e., photo-oxidative breakdown of hydrocarbons to form formaldehyde) and an important pathway for degradation of formaldehyde.

28 Chen *et al.* (2004) measured formaldehyde levels continuously over several days and 29 reported that peak formaldehyde levels occurred during daylight hours due to photochemical oxidation of VOCs caused by intense sunlight, and that minimum levels
 occurred during nighttime (Chen *et al.* 2004).

3 Formaldehyde half-lives in air can vary considerably under different conditions (WHO 4 2002). Atmospheric residence times in several U.S. cities ranged from 0.3 hours under 5 conditions typical of a rainy winter night to 250 hours under conditions typical of a clear 6 summer night. ATSDR (1999) reported half-lives in the atmosphere ranging from 1.6 to 7 19 hours. Reaction with the hydroxyl radical is the most important photo-oxidation 8 process in the degradation of formaldehyde (WHO 2002). Factors that influence 9 formaldehyde's atmospheric half-life, such as time of day, intensity of sunlight, and 10 temperature, are mainly those factors that affect the availability of the hydroxyl radical. 11 Based on hydroxyl radical reaction rate constants, the atmospheric half-life of 12 formaldehyde has been calculated to be between 7.1 and 71.3 hours. Photolysis is another 13 degradation pathway; however, it accounts for only about 2% to 5% of formaldehyde 14 removal. At night, the degradation of formaldehyde is expected to occur through 15 reactions with nitrate radicals. This process tends to be more significant in urban areas, 16 where concentrations of the nitrate radical are higher.

Formaldehyde is highly soluble in water and will transfer into clouds, precipitation, and
surface water. WHO (2002) noted that formaldehyde has a washout ratio (concentration
in rain/concentration in air) of 73,000, and thus is expected to be efficiently scavenged
from the atmosphere by atmospheric water.

Table 2-16 summarizes data on outdoor formaldehyde air levels in the United States that have been reported in review articles by Zhang *et al.*  $(2009a)^1$ , IARC (2006), ATSDR (1999), and WHO (1989). Both IARC and Zhang *et al.* reported levels for other countries that were more than an order of magnitude higher than those seen in the United States. The highest mean ambient level reported in the IARC review was 40 µg/m<sup>3</sup> in Rio de Janeiro, Brazil, and the highest single measurement (based on the upper end of the reported range) was 176 ppbv [216 µg/m<sup>3</sup>] in Budapest, Hungary. Ambient levels

<sup>&</sup>lt;sup>1</sup> Due to questions that arose during review of the Zhang *et al.* (2009a) review article, the primary references (Sax *et al.* 2004, Chen *et al.* 2004, and Mohammed *et al.* 2002) were reviewed and are cited in Table 2-16.

- 1 exceeding those reported for the United States were also seen in Italy, China, Mexico,
- 2 France, England, Egypt, and other parts of Brazil, all in urban areas. The highest levels
- 3 reported by Zhang *et al.* were from Rio de Janeiro, Brazil (151 ppb  $[186 \,\mu\text{g/m}^3]$ ) and
- 4 Mexico City, Mexico (110 ppb  $[135 \ \mu g/m^3]$ ). In addition to Brazil and Mexico, Zhang *et*
- 5 *al.* reported concentrations for seven countries that exceeded the maximum U.S.
- 6 concentration. The ATSDR (1999) and WHO (1989) reviews reported similar levels for
- 7 the United States and other countries.

Location (sampling period)	N	Concentration mean (range)	Poforonco
Location (sampling period)		(μ9/11)	Kelelence
$P_{\text{ostop}} MA (1002)$			Poiss at al. 1005 <sup>b</sup>
Winter massurements outside 4 residences	Q	3.81(0, 3.81)	Keiss et al. 1995
Summer measurements outside 9 residences	0 18	3.81(0-3.81) 3.2(1.5, 7.3)	
Number incastrements outside 9 residences	ND	3.2(1.3-7.3)	
New Jersey, 4 cities (1974)	NK	4.7-8.1 (means)	Cleveland <i>et al.</i> 1977 <sup>c</sup>
New York City, NY (1000)		17.2–20.0 (maxima)	Sou of al 2004
Winter	36	21(0541)	Sax el al. 2004
Summer	36	5.3(1.9-13)	
Schenestedy, NV (June, August 1082)	ND	ND (1.22, 28)	Sahulam at al. 1085 <sup>d</sup>
Atlantic CA 4 al anti-August 1985)		NK(1.25-56)	
Atlanta, GA, 4 urban areas (July and August 1992)	217	3.3-3.7  (max. = 10.2)	Grosjean <i>et al.</i> 1993 <sup>*</sup>
Baton Rouge, LA, FEMA trailer-staging area (2006)	NR	6.0 (1.0-87)	ATSDR 2007a
OH urban centers (June–July 1989)	48	3.7 (max. = 19.0)	Spicer et al. 1996 <sup>d</sup>
Houston, TX: Range of peak levels across the 3 sampling periods (2002)	NR	NR (< 8.6–37)	Chen et al. 2004
Denver, CO (1987–1991)	NR		Anderson et al. 1996 <sup>b</sup>
Winter		4.8 (NR)	
Spring		2.8 (NR)	
Summer		3.3 (NR)	
Los Angeles, CA (2000)			Sax et al. 2004
Winter	40	3.9 (2.3-8.4)	
Fall	35	4.4 (2.5–7.8)	
Los Angeles, CA (1999–2000)	69	8.8 (5.3–17.22)	Delfino et al. 2003 <sup>b</sup>
Los Angeles, CA (1993)			Grosjean et al. 1996 <sup>b</sup>
Measured at urban locations during smog season (September)	32	6.5 (1.7–13.0)	
Measured at 1 background location	NR	1.0 (0.9–1.2)	

Table 2-16. Occurrence of formaldehyde in outdoor air in the United States

Location (sampling period)	N	Concentration mean (range) (۲۹۹/m³)	Reference
Los Angeles, CA (Cal State University) (May– June 1980)	NR	NR 2.5–49	Grosjean 1982 <sup>d</sup>
Los Angeles, CA downtown (1960–1961) July–November (1960) September–November (1961)	31	49.1 (NR) 55.3 (NR)	Altschuller and McPherson 1963 <sup>c</sup>
California, during air pollution episode (NR) Lennox Azusa Los Angeles	36 36 20	NR 0.6–48.6 NR 0.9–43 NR 4.5–70.1	Grosjean and Swanson 1983 <sup>c</sup>
Claremont, CA (September–October 1980)	NR	NR 3.7–59	Grosjean 1982 <sup>d</sup>
Riverside, CA (NR)	32	NR (< 5–12)	Tuazon <i>et al.</i> 1978 <sup>c</sup>
Rural		r	
Albany, NY, rural and semi rural (October 1991)	NR	NR (0.74–4.5)	Khwaja 1995 <sup>b</sup>
Whiteface Mountain, Wilmington, NY (1983)	NR	NR (0.98–3.2)	Schulam et al. 1985 <sup>d</sup>
Mixed locations			
USA, mixed locations in TX, LA, VT, and NJ (1996–1997)	NR	NR (1.8–9.1)	Mohammed <i>et al.</i> 2002
USA, mixed locations (1975–1985) Nationwide Urban – mixed locations Suburban – mixed locations Rural and semirural – mixed locations	629 332 281 12	5.0 <sup>e,f</sup> (NR) 8.0 <sup>e</sup> (NR) 3.3 <sup>e</sup> (NR) 3.3 <sup>e</sup> (NR)	Shah and Singh 1988 <sup>b</sup>
United States, ambient air measurements at 58 locations (NR)	1,358	3.07 <sup>e</sup> (NR)	Kelly <i>et al.</i> 1994 <sup>d</sup>
United States, 9 datasets from 8 cities (1980– 1984)	NR	2.8–23.3 (means) 6.8–83 (maxima)	Salas and Singh 1986 and Singh <i>et al.</i> 1982 <sup>d</sup>
Minnesota, 25 sites throughout the state (1991–1998)	2,494	1.7 (< 0.05–21)	Pratt et al. 2000 <sup>b</sup>
California, multiple locations (NR)	NR	3.9-6.0 (NR)	Seiber 1996 <sup>d</sup>

<sup>a</sup>Data within this section are sorted geographically, generally from east to west across the United States. <sup>b</sup>Cited in IARC 2006. <sup>c</sup>Cited in WHO 1989. <sup>d</sup>Cited in ATSDR 1999.

<sup>e</sup>Median.

 $^{\rm f} The$  nationwide mean value was 10.2  $\mu g/m^3.$ 

#### 1 2.5.1.2 Indoor air

- 2 Formaldehyde levels generally are higher in indoor air than in outdoor air, often by an
- 3 order of magnitude or more (IARC 2006, ATSDR 1999). Sources of formaldehyde in

1 indoor air include off-gassing from various products (e.g., building materials, composite-2 wood-based furnishings, carpets, various consumer products, clothing, fabrics, UFFI, and 3 paints and varnishes) and indoor combustion sources (e.g., gas burners and ovens, 4 kerosene heaters, cook stoves, and cigarettes) (ATSDR 1999, IARC 2006, WHO 1989). 5 In indoor air, formaldehyde can form due to reactions of ozone with indoor materials 6 such as latex paints and carpets (Sax et al. 2004) and due to degradation of other organic 7 compounds in indoor air (ATSDR). Important determinants of indoor air levels include 8 the sources of the formaldehyde, the age of the source materials, temperature, humidity, 9 and ventilation rates (IARC 2006).

10 Formaldehyde levels in indoor air have been shown to be associated with the age and 11 structural type of the building; however, these factors are not independent and reflect 12 more fundamental variables such as the overall emission potential of the source materials 13 and the air-exchange rate of the dwelling (WHO 1989). In one study reviewed by WHO 14 (1989), the amount and dynamics of formaldehyde migration into indoor air was assessed 15 in relation to the age of the material, air temperature, and air-exchange rate. Age of the 16 material was found to be the most important factor influencing formaldehyde levels, 17 followed by temperature elevation, and then air-exchange rate.

In a study assessing secondary VOC emissions from flooring material, Kagi *et al.* (2009) exposed a low-formaldehyde type of flooring material to UV radiation and found that chemical transformations occurred resulting in the emission of a number of secondary products, including formaldehyde. Similar results were found when the flooring material was exposed to ozone.

Emission rates due to off-gassing have been assessed for various consumer products and
are presented in Table 2-17. (Measured indoor formaldehyde levels are discussed below.)
The highest emission rates were seen for UF floor finishes; this finding is supported by
data showing high exposure levels for workers who varnish floors (see Section 2.4.8).
Other products with high emission rates include fingernail hardener and polish, various
types of composite wood products (i.e., particleboard, plywood, UF wood products),
latex paints, permanent-press fabrics, and insulation. In general, UF resins have the

- 1 highest emission rates and PF resins the lowest emission rates (IRSST 2006). Generally,
- 2 emission rates from these products decrease over time (WHO 1989). It has been shown
- 3 that formaldehyde emission rates increase with higher ozone concentrations, temperature,
- 4 and relative humidity (Sax *et al.* 2004).

Table 2-17. Formaldehyde off-gassing	emission rates	from building	materials, hon	ne
furnishings, and consumer products				

Product	Emission rate (μg/m² per day)	Comment	Reference		
Building supplies and home furnishings					
Commercially applied UF floor finish Base coat Top coat	[10,104] [25,200,000]	Reported by ATSDR as 421 and 1,050,000 $\mu$ g/m <sup>2</sup> per hour	ATSDR 1999		
Particleboard	36,000–168,000	Range of releases based on varying a number of parameters in a test chamber	Pickrell et al. 1984		
Plywood	31,000–68,000	Range of releases based on varying a number of parameters in a test chamber	Pickrell et al. 1984		
Pressed wood products (including particleboard, plywood, and paneling)	BD-36,000	Minimum is for exterior plywood, and maximum is for paneling	Pickrell et al. 1983		
Bare UF wood products	210-37,900	Results from a variety of products	ATSDR 1999		
Bare PF wood products	100–220		ATSDR 1999		
Coated UF wood products	24–11,100	Results from a variety of products	ATSDR 1999		
Low-formaldehyde- emitting flooring Natural wood flooring without adhesives	96–2,000 2,000–6,900	Rates span flooring material exposed to ozone, infrared lamp, sun lamp, UVA lamp, and UVB lamp. Reference rates were "not detected" for the low-emitting flooring and 48 $\mu$ g/m <sup>2</sup> per day for the natural wood flooring	Kagi <i>et al</i> . 2009		
Insulation products	52–620	Includes various fiberglass products, air ducts, blackface insulation sheathing	Pickrell et al. 1983		
Insulation	3,000	Measured release rate from a test chamber; details on type of insulation not provided	Pickrell et al. 1984		

68
Product	Emission rate (μg/m <sup>2</sup> per day)	Comment	Reference
Carpet	BD-65	Both foam-backed and non-foam- backed carpets (highest level from foam-backed and lowest level from non-foam backed)	Pickrell et al. 1983
Carpet	1,500	Measured release rate from a test chamber (carpet type not specified)	Pickrell et al. 1984
Carpet	440–1,375	Measured rates from a test chamber; the maximum rate was at 24 h, and the minimum rate was at 168 h (carpet type not specified)	ATSDR 1999
Latex paints	7,800–14,200	From two brands of paints; the lower value was for a more expensive paint	ATSDR 1999
Decorative laminates	100–1,200		ATSDR 1999
Consumer products			
Fingernail hardener	5,172,000		ATSDR 1999
Nail polish	496,800		ATSDR 1999
Paper products	75–1,000	Paper plates and cups	Pickrell et al. 1983
Paper grocery bags	10		ATSDR 1999
Clothes	15-550	Unwashed new clothing	Pickrell et al. 1983
Fabric	BD-350	Includes drapery fabric and upholstery fabric of cotton, nylon, olefin, and rayon/cotton blends	Pickrell et al. 1983
Permanent press fabrics	1,000–5,100		ATSDR 1999
Towels	< 7		ATSDR 1999
Fiberglass products	380–770		ATSDR 1999

BD = below detection, UVA = ultraviolet A, UVB = ultraviolet B.

1 Off-gassing from UFFI is another potential source of formaldehyde in indoor air. No

2 emission rates were found in the literature; however, studies have indicated that

3 formaldehyde levels in homes increase immediately after foaming, but return to pre-

4 foaming levels after a few weeks (WHO 1989). As noted above, changes in home-

5 construction methods have significantly reduced the use of UFFI since the mid 1980s.

6 Paint can be a source of formaldehyde in indoor air. In one study, the average

7 formaldehyde level was 18  $\mu$ g/m<sup>3</sup> in office buildings that had recently been painted with

8 a low-formaldehyde-emitting paint. Three months later, the concentration had fallen to 8

9  $\mu g/m^3$ , which was the average level in a control area in the same building that had not

10 been painted (IARC 2006) (data are presented in occupational exposure section,

Table 2-15). A study in Swedish homes showed significantly increased formaldehyde
 levels in houses where wood paint had been used. This study also noted that wall-to-wall
 carpeting had contributed almost the same amounts of formaldehyde to indoor air as paint

4 had  $(13 \,\mu\text{g/m}^3 \,\text{vs.} \,16 \,\mu\text{g/m}^3)$ .

Indoor combustion sources of formaldehyde include wood stoves, gas stoves, kerosene
heaters, open fireplaces, furnaces, and burning tobacco products. Combustion sources
generally are considered to be weak emitters to indoor air, but tobacco smoke can be an
important source of formaldehyde in indoor air, potentially accounting for 10% to 25% of
indoor air exposure (ATSDR 1999) (see below and Table 2-19).

10 Other potential sources of formaldehyde in indoor air include cooking and formation 11 from other chemicals in the air. In one study, an emission rate of 1.38  $\mu$ g/g was estimated 12 for charbroiling meat over a natural-gas-fired grill (WHO 2002). Another study showed 13 emission rates for fish that ranged from 0.48  $\mu$ g/g for mackerel to 5.31  $\mu$ g/g for sardines 14 (IARC 2006). Formaldehyde has also been shown to be released from cooking oils that 15 were heated to 240°C to 280°C [464°F to 536°F].

Formaldehyde may form through degradation of organic compounds commonly found in indoor air. Formaldehyde has been found to form through this process at a rate of 0.87  $\mu$ g/s in winter and 2.43  $\mu$ g/s in summer (ATSDR 1999) [which is reflected in the higher indoor formaldehyde levels in summer than in winter shown in Table 2-18 for studies with measurements in both seasons].

21 Park and Ikeda (2006) found that air levels of VOCs in new homes decreased markedly 22 after one year; however, formaldehyde required a longer flushing period in new homes. 23 The authors concluded that decreases in indoor formaldehyde levels depend more on time 24 than on ventilation rates. Gold et al. (1993) noted that older conventional homes had the 25 lowest indoor concentrations of formaldehyde (compared with new conventional homes and mobile homes), with values typically less than 0.05 ppm [60  $\mu$ g/m<sup>3</sup>]. This is 26 27 consistent with the expected decrease in release of latent formaldehyde from wood-based 28 building materials as they age. Interior remodeling can also result in increased

29 formaldehyde levels. Tang et al. reported that in China, indoor formaldehyde

concentrations typically decrease with time, usually falling below 0.1 mg/m<sup>3</sup> about 6
 months after remodeling; however, the authors noted that levels can remain high even up
 to 1 year after remodeling.

4 In 2008, CDC released Final Report on Formaldehyde Levels in FEMA-Supplied Travel 5 Trailers, Park Models, and Mobile Homes (CDC 2008). The report summarized a study 6 of a stratified random sample of 519 occupied travel trailers, park models, and mobile 7 homes provided by the Federal Emergency Management Agency (FEMA) for use as 8 temporary shelter for Louisiana and Mississippi residents displaced by hurricanes Katrina 9 and Rita. The overall geometric mean indoor formaldehyde level was 77 ppb [95  $\mu$ g/m<sup>3</sup>] (range = 3 to 590 ppb [3.7 to 726  $\mu$ g/m<sup>3</sup>]). The Centers for Disease Control and 10 11 Prevention (CDC) reported that formaldehyde levels varied by trailer type (travel trailers 12 had significantly higher levels than park models or mobile homes), but all types tested had some levels greater than 100 ppb  $[123 \ \mu g/m^3]$ . Levels also varied by manufacturer. 13 14 Temperature was the most important determinant of indoor levels. Other statistically 15 significant determinants of formaldehyde levels included relative humidity; opened 16 windows, doors, and scuttles; and presence of mold. Indoor cooking and tobacco 17 smoking contributed to formaldehyde levels, although not significantly. The CDC noted 18 that since indoor formaldehyde levels tend to be higher in warmer weather and in newly 19 constructed trailers, the results of this study could have underestimated long-term 20 exposure levels (many of the trailers were around 2 years old, and the study was 21 undertaken in winter).

22 In 2006, ATSDR evaluated data on formaldehyde levels in FEMA temporary housing 23 units in Baton Rouge, LA. Two different ventilation methods were tested in the study: 24 Method A relied on running the air conditioning and opening the bathroom vents only, 25 and Method B relied on opening all windows and vents. The authors found that Method 26 B was more effective at lowering formaldehyde levels (see Table 2-18) (ATSDR 2007a). 27 ATSDR (1999) also noted that the generally increased levels of formaldehyde in mobile 28 homes would be expected because of their generally lower air-exchange rates. IARC 29 noted that formaldehyde in the air of mobile homes has a half-life of about four or five 30 years.

September 3, 2009

1	Residential indoor air levels of formaldehyde have been extensively documented by
2	IARC (2006), ATSDR (1999), and WHO (1989). U.S. levels from these assessments are
3	presented in Table 2-18. Residential indoor air levels reported for other countries were
4	very similar to U.S. levels, and except for one instance (in which $> 500$ ppb was reported
5	in Austrian apartments), all data points fell within the range of concentrations reported
6	for the United States. Zhang et al. (2009a) presented graphs showing indoor
7	formaldehyde air levels for several countries, noting that in general, indoor levels
8	(including U.S levels) were below the WHO recommended indoor limit of $0.1 \text{ mg/m}^3$ .
9	However, mean levels for Cairo, Egypt, and Tianjin, China, were slightly higher than the
10	WHO recommended level (roughly $0.12 \ \mu g/m^3$ for both cities), and levels in Beijing,
11	China, were roughly 0.2 $\mu$ g/m <sup>3</sup> in winter and 0.28 $\mu$ g/m <sup>3</sup> in summer. The ATSDR review
12	included many measurements made in the mid 1980s or earlier; the authors noted that
13	production methods have since been changed to reduce formaldehyde levels in plywood
14	and particleboard, and the use of UFFI has decreased. The authors also noted that
15	formaldehyde levels in mobile homes appear to have been decreasing since about 1980,

16 probably as a result of the use of these reduced-emission products.

Table 2-18. Occurrence of formaldehyde in U.S. residential indoor air

		Concentration mean (range)	
Location (year measured)	N <sup>a</sup>	(µg/m³)	Reference
Manufactured housing			
LA & MS, 519 FEMA-supplied temporary housing units (Dec. 2007–Jan. 2008)	519*	95 (3.7–26)	CDC 2008
Baton Rouge, LA, 96 FEMA-supplied temporary housing units (2006)			ATSDR 2007a
Ventilation with air conditioning and bathroom vents only	1,090	490 (3.4–3,000)	
Ventilation with open windows and vents	1,117	172 (3.0–4,500)	
Florida, new manufactured house (2000)	NR	94.9 (NR)	Hodgson <i>et al.</i> 2002 <sup>b</sup>
United States, East and Southeast (1997–1998)	4		Hodgson et al.
Indoor level		41.8 <sup>c</sup> (25.8–57.7)	2000
Outdoor level		$2.5^{\rm c}$ (NR)	
California, mobile homes (1984–1985)	470	86–110 (NR)	Sexton <i>et al.</i> 1989 <sup>d</sup>

		Concentration	
Location (vear measured)	N <sup>a</sup>	(μα/m <sup>3</sup> )	Reference
Texas, mobile homes whose residents requested		(****** /	Norsted <i>et al.</i>
testing (1979–1982)	443*	(NR) ND-9,830	1985 <sup>d</sup>
Homes < 1 yr old		[> 2,460] for 27% of homes	
Homes $> 1$ yr old		[>2,460] for 11.5% of homes	
United States (NR)	430*	> 1.23 for 4% of samples	Breysse 1984 <sup>e</sup>
		0.61–1.22 for 18% of samples	
		0.12–0.60 for 64% of samples	
		< 0.12 for 14% of samples	
United States (NR)	431*	0.47 (0.012–3.6)	Ulsamer <i>et al.</i> 1982 <sup>e</sup>
United States (NR)			Stone <i>et al</i> .
Complaint homes, WA, < 2 yr old	110*	0.95 (NR)	1981 <sup>e</sup>
Complaint homes, WA, 2–10 yr old	77*	0.58 (NR)	
Complaint homes, MN, < 2 yr old	66*	1.04 (NR)	
Complaint homes, MN, 2-10 yr old	43*	0.34 (NR)	
Complaint homes, WI, < 2 yr old	38*	0.89 (NR)	
Complaint homes, WI, 2–7 yr old	9*	0.56 (NR)	
Random sample, WI, $< 2$ yr old	NR	0.66 (NR)	
Wisconsin, complaint homes, 0.2 to 12 yr old (NR)	65*	$0.59^{ m f}$	Dally <i>et al.</i> 1981 <sup>e</sup>
Traditional housing or unspecified	L		
New York City, NY (1999)			Kinney et al.
Winter	38	12.1 (NR)	2002 <sup>b</sup>
Summer	41	20.9 (NR)	
United States, East and Southeast, site-built houses (1997–1998)	7	44.2° (17.2–71.2)	Hodgson <i>et al.</i> 2000 <sup>b</sup>
Louisiana, 53 houses: 75% urban and 25% rural (NR)	419	460 (ND-6,600)	Lemus <i>et al.</i> 1998 <sup>b</sup>
Boston, MA (1993)			Reiss et al.
Winter, 4 residences	14	13.6 (7.4–19.8)	1995 <sup>b</sup>
Summer, 9 residences	26	19.8 (7.3–66.1)	
Colorado (1992–1993)	9		Lindstrom et
Prior to occupancy		26 <sup>c</sup> (8–66)	<i>al.</i> 1995 <sup>b</sup>
After occupancy for 5 months		49 <sup>c</sup> (33–81)	
New Jersey, residential houses (1992)	6*		Zhang et al.
Indoor		67.01 (NR)	1994b <sup>d</sup>
Outdoor		15.4 (NR)	
Arizona, houses (NR)	202*	31.9 (max. 172)	Krzyzanowski <i>et al.</i> 1990 <sup>d</sup>
United States, residential, various locations (1981–1984)	273	44.0 <sup>f</sup> (NR)	Shah and Singh 1988 <sup>b</sup>

		Concentration mean (range)	
Location (year measured)	N <sup>a</sup>	(μg/m <sup>3</sup> )	Reference
San Francisco, CA, Bay Area (1984)			Sexton et al.
Kitchen	48	50.4 (NR)	1986 <sup>b</sup>
Main bedroom	45	44.2 (NR)	
Pullman, WA, houses (NR)	NR	6.14–88.43 (NR)	Lamb <i>et al.</i> 1985 <sup>d</sup>
United States (NR)			Breysse 1984 <sup>e</sup>
UFFI houses	244*	> 1.23 for 2.8% of samples	
		0.61–1.22 for 1.9% of samples	
		0.12–0.60 for 24.1% of	
		samples	
Non-UFFI houses and apartments	59*	< 0.12 for 71.2% of samples	
		> 1.23 for 1.8% of samples	
		0.61–1.22 for 1.8% of samples	
		0.12–0.60 for 36.3% of	
		< 0.12 for 60.1% of samples	
United States (1092)			Llowthowno of
Houses 0, 30 yr old	40*	$0.076 \pm 0.005^{g}$	$al 1983^{e}$
Houses 0 - 50 yr old	18*	$0.070 \pm 0.093$	<i>un</i> . 1905
Houses 5 15 yr old	10*	$0.103 \pm 0.112$	
Houses $> 15$ yr old	11	$0.032 \pm 0.052$	
Houses $0-5$ yr old	18*	$0.057 \pm 0.052$	
spring	10	$0.107 \pm 0.114^{g}$	
summer		$0.136 \pm 0.125^{g}$	
autumn		$0.058 \pm 0.068^{\text{g}}$	
Houses 5–15 vr old	11*		
spring		$0.053 \pm 0.049^{ m g}$	
summer		$0.060 \pm 0.059^{ m g}$	
autumn		$0.042 \pm 0.043^{g}$	
Houses > 15 yr old	11*		
spring		$0.044 \pm 0.063^{g}$	
summer		$0.036\pm0.046^{\rm g}$	
autumn		$0.032\pm0.028^{\text{g}}$	
United States (1983)			Grimsrud et al.
Energy-efficient new houses	20*	0.076 (NR)	1983 <sup>e</sup>
Low-ventilation modernized houses	16*	0.037 (NR)	
United States (1981)			Ulsamer et al.
Houses without UFFI	41*	0.04 (0.012-0.098)	1982 <sup>e</sup>
Houses with UFFI	636*	0.15 (0.012-4.2)	

Logation (year manufact)	Na	Concentration mean (range)	Poforonoo
	N	(μg/m)	Reference
United States (1980–1981)			Offerman <i>et</i>
Houses averaging 2 yr old	9*		<i>al.</i> 1982 <sup>e</sup>
air-tight construction		$0.044\pm0.02^{\rm g}$	
mechanical ventilation		$0.033\pm0.02^{\rm g}$	
Houses averaging 6 yr old (loose construction)	1*	0.017 (NR)	
United States (1978–1979)	13*	$1.12^{\rm f}$ (NR)	Dally <i>et al.</i> 1981 <sup>e</sup>
United States (1979)	2*		Berk et al.
Energy-efficient house		0.098 (0.04–0.15)	1980 <sup>e</sup>
Unoccupied house without furniture		$0.081 \pm 0.007^{ m g}$	
Unoccupied house with furniture		$0.225\pm0.016^{\text{g}}$	
Occupied house			
day		$0.263 \pm 0.026^{\text{g}}$	
night		$0.141 \pm 0.044^{g}$	

NR = not reported; ND = not detected.

<sup>a</sup>Number of samples unless denoted with an asterisk (\*), which indicates number of houses. <sup>b</sup>Cited in IARC 2006. <sup>c</sup>Geometric mean. <sup>d</sup>Cited in ATSDR 1999. <sup>e</sup>Cited in WHO 1989. <sup>f</sup>Median.

<sup>g</sup>Standard deviation.

#### 1 A number of studies have estimated formaldehyde levels in cigarette mainstream smoke,

2 sidestream smoke, and indoor air due to smoking. Levels in sidestream smoke have been

- 3 estimated to be from 5 to 50 times the levels in mainstream smoke (ATSDR 1999).
- 4 Table 2-19 summarizes formaldehyde levels in tobacco smoke and resultant exposure
- 5 levels.

Source or setting	Average or range	Comment	Reference
Formaldehyde levels in ciga	arettes and cigarette smol	ĸe	·
Total per cigarette	~1,500–2,000 µg	Low end of range reported in WHO 1989 and upper end reported in ATSDR 1999	ATSDR 1999, WHO 1989
Sidestream smoke, total per cigarette	958–2,360 μg (range)	The range represents the minimum and maximum values reported across numerous studies. The low end is the low end of a range from one study. The high end is the mean value from another study (the range for that study was not provided).	WHO 1989, 2002
Mainstream smoke Total per cigarette Total per puff Concentration	8–284 μg 5.1–8.9 μg 60,000–130,000 μg/m <sup>3</sup>	Total per cigarette includes data from numerous studies involving numerous brands and types of cigarettes. Total per puff data from 6 American filter-tip brands.	WHO 2002, 1989, ATSDR 1999
Formaldehyde air concentr	ations due to smoking		·
50-m <sup>3</sup> chamber	120 μg/m <sup>3</sup>	Six cigarettes smoked over 15 minutes; chamber averaged 1 air exchange per hour	WHO 1989
30-m <sup>3</sup> chamber 0.2–0.3 air exchanges/hr 1 air exchange/hr	210–350 μg/m <sup>3</sup> 50–70 μg/m <sup>3</sup>	Formaldehyde yield from 5–10 cigarettes smoked in the chamber at the two different exchange rates	WHO 1989
Nonsmoking office building Smoking section of building	BD-270 μg/m <sup>3</sup> BD-740 μg/m <sup>3</sup>		ATSDR 1999
building			

Table 2-19. Formaldehyde levels associated with cigarette smoke

BD = below detection.

1 The interior of automobiles can be a significant source of formaldehyde exposure as a

2 result of off-gassing from interior materials. Using data from chamber tests that showed

3 an average formaldehyde concentration of 48  $\mu$ g/m<sup>3</sup> at 23°C [73°F], Schupp *et al.* (2005)

4 extrapolated a car concentration of 1,680  $\mu$ g/m<sup>3</sup> at a temperature of 65°C [150°F], which

- 5 is easily reached in the interior of a car sitting in the sun with the windows rolled up.
- 6 Based on air samples taken inside 802 new cars (manufactured in and after 2003) parked
- 7 in an underground parking garage, Zhang et al. (2008b) reported a mean airborne
- 8 formaldehyde level of 80  $\mu$ g/m<sup>3</sup> (range = 20 to 1,110  $\mu$ g/m<sup>3</sup>). Samples were also taken

1 inside 20 older cars (manufactured before 2003) for comparison; levels were slightly

2 lower in the older cars.

#### 3 2.5.2 Water

Formaldehyde has been detected in bottled drinking water, treated drinking water, and
various types of environmental water, including groundwater, surface water, fog, and
mist. This section discusses formaldehyde levels in these various types of water. Because
drinking water is the most likely potential source of exposure, it is discussed first,
followed by a discussion of formaldehyde levels in other types of environmental waters.

#### 9 2.5.2.1 Drinking water

10 Formaldehyde in treated drinking water occurs primarily through the oxidation of organic 11 matter during ozonation or chlorination (WHO 2005); however, formaldehyde can also 12 be present in the water before treatment. Krasner et al. (1989) reported the results of a 13 study on the occurrence of disinfection by-products in U.S. drinking-water supplies. 14 Formaldehyde and several other disinfection by-products were measured both pre- and 15 post-treatment at 35 drinking-water treatment facilities in 1988 and 1989. To ensure that 16 the facilities chosen for analysis were representative, selection was based on the type of 17 source water, type of treatment process, population served, geographic location, and the 18 disinfectants used (i.e., free chlorine, chloramines, chlorine dioxide, or ozone). Levels of 19 disinfection by-products were assessed quarterly (spring, summer, fall, and winter, 1988– 20 1989), and the data for formaldehyde are presented in Table 2-20 [note that formaldehyde 21 was not assessed in spring]. To determine whether the formaldehyde was produced 22 during the disinfection process or originated from the source water, formaldehyde was measured in the influents of all 35 facilities. It was detected in 16 influent samples at 23 24 levels ranging from 1.2 to 13  $\mu$ g/L, with a median of 2.8  $\mu$ g/L. The median for all 25 samples (including samples in which no formaldehyde was detected) was less than 1 26  $\mu$ g/L. The authors suggested that the presence of formaldehyde in treated drinking water 27 depends on a combination of the disinfection process and the influent water quality. It 28 was noted, however, that formaldehyde clearly was a product of the oxidation-29 disinfection process, and that formaldehyde levels were higher at facilities that used 30 ozone treatment.

1 Formaldehyde can also contaminate drinking water through leaching from polyacetal

2 plastic fittings whose protective coatings have been compromised (Owen et al. 1990,

3 Tomkins et al. 1989, WHO 2002). Concentrations ranging from roughly 20 to 100 µg/L

4 have been reported to result from this process; levels were positively associated with the

5 residence time of the water in the pipe (Owen *et al.* 1990).

6 WHO (2002) noted that based on limited U.S. data, formaldehyde concentrations in

7 drinking water may range up to approximately  $10 \mu g/L$  in the absence of contributions

8 from ozone treatment during water treatment or from leaching of formaldehyde from

9 polyacetal plumbing fixtures.

10 Formaldehyde has also been detected in bottled drinking waters. Mutsuga et al. (2006) 11 purchased 20 polyethylene terephthalate (PET) bottles of mineral water and analyzed the 12 water for formaldehyde and acetaldehyde. Of the 20 bottles of water, 6 were bottled in 13 Japan, 11 in Europe, and 3 in North America. All of the Japanese bottled-water samples 14 contained detectable levels of formaldehyde, whereas 3 of the 11 European samples and 15 2 of the 3 North American samples had detectable formaldehyde levels (see Table 2-20). 16 The authors concluded that formaldehyde in the water was due to leaching from the PET 17 bottles. In further investigations to explain the absence of formaldehyde from some of the 18 water samples, the authors discovered that the water samples without formaldehyde were 19 unsterilized and contained heterotrophic bacteria. Based on these findings, the authors 20 suggested that formaldehyde probably had leached from the PET bottles but had been 21 decomposed by the bacteria.

Tsai *et al.* (2003) measured formaldehyde levels in 63 brands of packed drinking water and 13 brands of barreled drinking water in Taiwan. The authors reported that all concentrations were below 129 ppb [129  $\mu$ g/L] [specific levels not reported] and noted that these levels were well below the WHO water-quality guidelines of 900  $\mu$ g/L. No additional information was found specifically for bottled water in the United States.

Water type	Concentrati on (μg/L)	Comments	Reference
U.S. drinking water at treatment facility Summer 1988 Fall 1988 Winter 1988–1989	5.1 <sup>a</sup> 3.5 <sup>a</sup> 2.0 <sup>a</sup>	Formaldehyde was detected at concentrations ranging from 1.2 to 13 $\mu$ g/L in influents of 16 of 35 treatment facilities; however, authors noted that it was also created through treatment by ozonation or chlorination	Krasner <i>et al.</i> 1989
U.S. domestic drinking water	~ 20–100	Concentrations observed in a study assessing the leaching of formaldehyde from domestic polyacetal plumbing fixtures. [The low end is assumed to represent normal conditions and the high end to represent a reasonable worst-case scenario.]	WHO 2002
U.S. domestic drinking water	~ 10	Levels expected without contributions from ozone treatment during water treatment or by leaching from polyacetal plumbing fixtures	WHO 2002
U.S. drinking water	BD	U.S. EPA's 1975 report on National Organics Reconnaissance Survey of Suspected Carcinogens in Drinking Water	ATSDR 1999
Drinking water (location not reported)	< 100	Noted as generally less than this level	WHO 1989
Drinking water (treated with ozone; location not reported)	< 50	Noted as unlikely to exceed this level	WHO 2005
Bottled water Bottled in Japan Bottled in Europe Bottled in North America	10.1–27.9 7.8–13.7 13.6, 19.5	Range of levels detected in water from 20 PET bottles. Detectable levels were found in 6 of 6 Japanese, 3 of 11 European, and 2 of 3 North American bottled waters.	Mutsuga <i>et al.</i> 2006
63 brands of packed drinking water and 13 brands of barreled drinking water in Taiwan	< 129	Specific levels not reported	Tsai <i>et al.</i> 2003

Table 2-20. Formaldehyde concentrations in drinking water

BD = below detection. PET = polyethylene terephthalate. <sup>a</sup>Median; range not reported.

## 1 2.5.2.2 Environmental Water

- 2 Groundwater can be contaminated by formaldehyde leaching from surface soils into the
- 3 water table and through underground injection of wastes. In 2007, underground injection
- 4 of formaldehyde was the predominant source of industrial release to the environment,
- 5 based on TRI reporting data; 11.9 million pounds was released to on-site and off-site
- 6 underground injection wells, accounting for 54% of total U.S. releases reported to TRI

1 (TRI 2009). As a percentage of total releases, underground injection has trended upward

2 since 1988, with a minimum of 29% in 1992 and a maximum of 55% in 2006. ATSDR

3 (1999) reported that formaldehyde had been detected in groundwater at 4 of 26 hazardous

4 waste sites at which at least one environmental medium was contaminated with

5 formaldehyde. No information was found on the fate of formaldehyde in groundwater.

6 Surface water can be contaminated via the direct discharge of formaldehyde-containing

7 wastes, the use of formaldehyde in aquaculture, formaldehyde runoff from hazardous

8 waste sites, and land disposal of formaldehyde-containing wastes. Formaldehyde releases

9 to U.S. surface waters totaling 278,335 pounds were reported to the TRI for 2007 (TRI

10 2009), accounting for roughly 1% of all formaldehyde releases reported to the TRI.

11 Discharges to surface water have declined steadily since 1988 when 904,547 pounds

12 were reported. The minimum amount reported from 1988 through 2007 was 277,083

13 pounds in 2003. Formaldehyde-containing wastes may also be sent to publicly owned

14 treatment works (POTWs) and subsequently released to surface waters. For example,

15 formaldehyde has been found in hospital effluent at a 24-hour average concentration of

16 0.07 mg/L (Boillot et al. 2008). As a result of treatment at POTWs, only a fraction of

17 formaldehyde received is expected to be released to surface waters (ATSDR 1999);

18 however, no data on treatment efficiency or resultant discharge levels were found.

19 Formalin is commonly used in fish-culture activities to treat fish with fungal or

20 ectoparasitic infections; after use, formaldehyde solutions often are discharged into the

21 hatchery effluent (WHO 1989). No data were found on formaldehyde levels in water due

22 to such discharges.

23 In 1999, ATSDR (1999) noted that formaldehyde had been detected in surface water at 5

of 26 hazardous waste sites at which at least one environmental medium was

contaminated with formaldehyde. In 2007, roughly 373,000 pounds of formaldehyde was

26 disposed of in U.S. landfills, surface impoundments, land treatment sites, and other land

disposal sites, accounting for less than 2% of total U.S. releases reported to the TRI for

that year (TRI 2009). No information was available to estimate the impacts to surface

29 water from these land disposals.

1 Although volatilization of formaldehyde from surface waters is expected to be low,

2 biodegradation in surface water is a significant degradation process; formaldehyde is

3 biodegraded to low levels within a few days. In one study, formaldehyde was completely

4 biodegraded in water from a stagnant lake within 30 hours under aerobic conditions and

5 within 48 hours under anaerobic conditions (ATSDR 1999). Based on its low K<sub>ow</sub>,

6 adsorption of formaldehyde to sediment is expected to be low (Howard 1989). Biotic and

7 abiotic degradation are expected to be significant fate processes in sediment.

8 Table 2-21 provides data on formaldehyde levels in U.S. environmental waters.

9 ATSDR's HazDat database provided the only data found for U.S. groundwater levels. 10 [Note that the on-line HazDat database provides only maximum values measured at 11 Superfund sites or other facilities where ATSDR has performed a site assessment.] Three 12 data points were provided for formaldehyde: 0.1 ppm [~0.0001 µg/L] measured in 1979 13 at a facility in New Jersey, 0.0005 µg/L measured in 1980 at a facility in North Carolina, 14 and 140 µg/L at a facility in California [year not reported]. WHO (2002) presented results 15 of groundwater monitoring at two industrial facilities in Canada where groundwater had 16 been contaminated with formaldehyde. For one facility, which produced and used 17 formaldehyde, formaldehyde was detected in 43 samples at concentrations ranging from 18 65 to 690,000  $\mu$ g/L and was not detected in 10 samples (detection limit = 50  $\mu$ g/L). This 19 site was monitored from November 1991 to February 1992 as part of a program to 20 delineate the boundaries of groundwater contamination at the facility. At the other 21 facility, which produced UF resins, quarterly analyses of five on-site monitoring wells in 22 1996 and 1997 showed formaldehyde concentrations ranging from below the limit of 23 detection to 8.200  $\mu$ g/L, with an overall median of 100  $\mu$ g/L. It was noted that 24 concentrations measured in various wells indicated little dispersion from the source of 25 contamination. Groundwater samples collected down gradient from six cemeteries in 26 Ontario, Canada, contained formaldehyde at levels ranging from 1 to 30  $\mu$ g/L (WHO 27 2002).

Water type	Concentration (µg/L)	Comments	Reference
Groundwater	100–500	Range of maximum values from 3 locations in ATSDR's HazDat database	ATSDR 2007b
Surface water	2,100, 7,400	Maximum values from two locations in ATSDR's HazDat database	ATSDR 2007b
Surface water	BD-12	Of 204 sites in 14 heavily industrialized U.S. river basins, 1 site had detectable formaldehyde	Howard 1989
Rainwater	BD-0.06	California	ATSDR 1999
Fog water	1,800 <sup>a</sup> (400–3,000)	Corvallis, OR	ATSDR 1999
Fog water	3,000 <sup>b</sup> (120–6,800)	Riverside, CA	ATSDR 1999
Mist water	250 560	Long Beach, CA Marina del Ray, CA	ATSDR 1999
Snow	18–901	California	WHO 2002

Table 2-21. Formaldehyde levels in U.S. environmental water

BD = below detection

<sup>a</sup>Volume-weighted mean. <sup>b</sup>Median.

1 As with groundwater, ATSDR's HazDat database provided the only data on U.S. surface-

2 water levels of formaldehyde providing maximum levels at two locations in California of

3 7,400 μg/L and 2,100 ppb [~2,100 μg/L].

4 Because of its high solubility in water, formaldehyde is efficiently transferred into

5 clouds, fog, and precipitation, leading to potentially high levels in these media (Table 2-

6 21). WHO (2002) noted that formaldehyde has a washout ratio [concentration in rain to

7 concentration in air] of 73,000, and thus is estimated to be efficiently removed from the

8 atmosphere by atmospheric water. Levels of formaldehyde in rainwater in California

9 have been reported to range from below detection to 0.06  $\mu$ g/L (ATSDR 1999). WHO

10 (1989) reported levels in rainwater ranging from 8  $\mu$ g/L (a mean level reported for the

- 11 central equatorial Pacific Ocean) to  $1,380 \mu g/L$  (location not reported). No information
- 12 was provided that would explain why these levels were so much higher than the levels
- 13 reported by ATSDR (1999).
- 14 No data were found on formaldehyde levels in water sediment.

#### 1 2.5.3 Land and soil

2 Formaldehyde occurs in soil through its use in controlled-release fertilizers, its use as a

- 3 fumigant, and land disposal of industrial, construction, demolition, and other wastes.
- 4 Formaldehyde could be released to soil from hazardous waste sites (ATSDR 1999). It is
- 5 also formed naturally in soil during decomposition of plants (WHO 1989).
- 6 Based on TRI data, 373,000 pounds of formaldehyde were released to land in 2007: 82%
- 7 to landfills, 14% to surface impoundments, 3% to land treatment sites, and 1% to other
- 8 land disposal sites (TRI 2009). Land disposal has declined considerably but has
- 9 fluctuated widely since TRI data were first reported, from a maximum disposal of
- 10 1.25 million pounds in 1988 to a minimum of about 205,000 pounds in 1997. As noted
- 11 above, over 11.9 million pounds of formaldehyde was released to underground injection
- 12 wells in 2007: 98% to on-site wells and 2% to off-site wells. Since 1988 (the first year in
- 13 which data were reported), underground injection releases have ranged from around
- 14 5 million pounds in 1992 to over 13.6 million pounds in 2004.
- 15 Formaldehyde is degradable under both aerobic and anaerobic conditions (Howard
- 16 1989); however, no soil degradation rates were found in the literature. It has a low soil-
- 17 adsorption coefficient, meaning that it is very mobile in soils (WHO 1989). Based on its
- 18 Henry's law constant, it is not expected to volatilize appreciably (Howard 1989).
- 19 Although large amounts of formaldehyde are disposed of on land and in the ground, no
- 20 U.S. soil concentration data were found. In Canada, soil levels were measured in 1991 at
- 21 a plywood manufacturing facility that used PF resins. Six soil samples contained
- formaldehyde concentrations ranging from 73 to 80 mg/kg, with a mean of 76 mg/kg
- 23 (WHO 2002).
- 24 2.5.4 Food
- 25 Formaldehyde can occur in food naturally, through direct addition as a preservative, as a
- 26 result of cooking or smoking of foods, or through inadvertent contamination (e.g., from
- its use as a fumigant or from the use of utensils made from formaldehyde resins)
- 28 (Howard 1989, WHO 1989, ATSDR 1999). Formaldehyde has also been shown to be
- 29 eluted from formaldehyde-resin plastic dishes by water, acetic acid, and ethanol at

temperature-proportionate levels (ATSDR 1999). Formaldehyde levels in fresh fruit have
 been found to increase after refrigeration (Tang *et al.* 2009).

3 As shown in Table 2-22, generally higher formaldehyde levels have been seen in fish and 4 seafood than in other foods, aside from smoked ham. Formaldehyde develops 5 postmortem in marine fish and crustaceans via enzymatic reduction of trimethylamine 6 oxide (WHO 2002). Formaldehyde will accumulate in some fish species, including cod, 7 pollack, and haddock, during frozen storage. The formaldehyde formed in fish reacts with 8 protein, causing muscle toughness, and it has been suggested that fish containing the 9 highest levels of formaldehyde may not be palatable for human consumption. Li et al. 10 (2007b) observed variable formaldehyde levels among four species of squid; levels 11 generally were far higher in viscera than in muscle of frozen squid. The authors also 12 noted that formaldehyde levels increased with increasing cooking temperature.

13 Tang *et al.* (2009) reported that an illegal use of synthetic formaldehyde (Rongalite®

- 14 [i.e., sodium formaldehyde sulfoxylate]) as a food preservative is common in Chinese
- 15 markets, and that formaldehyde-induced food poisoning remains a huge problem in
- 16 China because of this practice. Based on data from seven independent studies, Tang *et al.*
- 17 reported high formaldehyde levels in seafood due to this practice (Table 2.22).

Food	Concentration (mg/kg)	Comment	Reference
Fruits and vegetables			
60 different fresh fruits: Without refrigeration With refrigeration	< 2.74 [< 6.3–10.4]	Reported that fruits had levels below 2.74 but the levels increased 2.3 to 3.8 times with refrigeration	Tang <i>et al.</i> 2009
Pear	38.7, 60	Values based on two different analytical methods	WHO 1989
Apple	17.3, 22.3		WHO 1989
Cabbage	4.7, 5.3		WHO 1989
Carrot	6.7, 10		WHO 1989
Green onion	13.3, 26.3		WHO 1989
Spinach	3.3, 7.3		WHO 1989
Tomato	5.7, 7.3		WHO 1989
White radish	3.7, 4.4		WHO 1989

#### Table 2-22. Formaldehyde levels in food

Food	Concentration (mg/kg)	Comment	Reference
Meat			
Pig	20		WHO 1989
Sheep	8		WHO 1989
Poultry	5.7		WHO 1989
Smoked ham	267	Value for the outer layer of ham	WHO 2002
Milk and milk products			
Goat's milk	1		WHO 1989
Cow's milk	≤ 3.3		WHO 1989
Cow's milk	0.22	Maximum value from cows fed formalin; it was noted that this was roughly 10 times the level in milk from cows without added formalin in the diet.	WHO 2002
Cow's milk (fresh)	0.013-0.057	Higher levels in processed milk were	WHO 2002
Processed 2% milk	0.027 (mean) 0.075–0.255 0.164 (mean)	attributed to processing technique, packaging, and storage.	
Cheese	≤ 3.3		WHO 1989
Fish and seafood			
Squid	10.7–165	Levels across the muscle and viscera and for dried squid thread for 4 species	Li <i>et al.</i> 2007b
Freshwater fish (fumigated)	8.8	Fumigation process not described in the source	WHO 1989
Ocean fish (fumigated)	20		
Cod (frozen)	20		WHO 1989
Shrimp (live)	1		WHO 1989
Crustaceans (Mediterranean)	1–60		WHO 1989
Crustaceans (ocean)	3–98		WHO 1989
Fresh marine products	2.177 ± 1.41 (mean std. dev.)	Includes products such as mackerel, squid, pomfret, hairtail, sea cucumber, red shrimp, yellow croaker, scallop and octopus	Tang <i>et al.</i> 2009
Marine products illegally treated with formaldehyde preservative	~300-4,250	Results of 7 independent studies in 6 Chinese cities	Tang <i>et al.</i> 2009

Food	Concentration (mg/kg)	Comment	Reference
Beverages			
Fruit and vegetable juices	$\leq 800$	It was reported that concentrations up to 800 mg/kg have been reported in fruit and vegetable juices in Bulgaria	WHO 2002
Alcoholic beverages	0.02–3.8 mg/L	Concentrations from a variety of alcoholic beverages from a study in Japan and a study in Brazil	WHO 2002
Canned or bottled beer	0.1–1.5		WHO 2002
Beer	0.1–0.9	Levels in China across domestic and imported beers	Tang <i>et al.</i> 2009
Canned or bottled cola	7.4–8.7		WHO 2002
Brewed coffee	3.4–4.5		WHO 2002
Instant coffee	10–16		WHO 2002
Other			
Shiitake mushroom	40–380	Range of base concentration measurements	Tang <i>et al.</i> 2009
Vermicelli noodles	0.011-3.38	Full range across two studies	Tang <i>et al.</i> 2009
Maple syrup Untreated trees Treated trees	< 1 up to 14	Trees treated with paraformaldehyde to deter bacterial growth	WHO 2002

1 The artificial sweetener aspartame consists of 10% methanol, which Humphries *et al.* 

2 (2008) reported can be converted to formaldehyde and other derivatives. The authors also

3 noted that research has shown that formaldehyde adducts accumulate in the tissues after

4 aspartame ingestion.

5 Formaldehyde can be added to ruminant feeds to improve handling characteristics. It has

6 been estimated that animals may ingest as much as 0.25% formaldehyde in their diets

7 (WHO 2002). Formalin has been added as a preservative to skim milk fed to pigs in the

- 8 United Kingdom and to liquid whey fed to cows and calves in Canada. Formaldehyde
- 9 levels in milk from cows fed formalin at the highest concentration were up to 10 times
- 10 the level in milk from control cows. No data were found on levels in meat due to
- 11 formaldehyde in animals' diets.

#### 1 **2.6 Exposure estimates**

2 Exposure to formaldehyde can occur from breathing of air and tobacco smoke; ingestion 3 of food, drinking water, and other beverages; dermal contact; and, rarely, direct entry of 4 aqueous solution into the bloodstream (e.g., during medical procedures in which 5 machines or tubing have been disinfected with formaldehyde) (IARC 2006, ATSDR 6 1999, WHO 1989). As noted above, there are no widely accepted biomarkers for 7 formaldehyde exposure and, therefore, very few data on human intake levels. Exposure 8 can be estimated by combining media concentration information with assumed ingestion 9 and inhalation rates and making various assumptions about the duration of exposure 10 periods. Exposure estimates found in the literature are provided in Table 2-23.

Source	Intake (mg/day)	Comment	Reference
Food	1.5–14	Range based on meal composition	WHO 1989
Workplace air Without occupational exposure With occupational exposure	0.2–0.8 5.0–8.0	Assumes 25% of day at work. Without occupational exposure assumes normal concentrations in conventional buildings; with occupational exposure assumes 1-mg/m <sup>3</sup> air concentrations. Ranges are across two datasets.	Fishbein 1992, WHO 2002
Tobacco smoke Smoking 20 cigarettes/day Environmental tobacco smoke Home Work	0.9–2.0 0.5–3.5 0.4–2.8	Environmental tobacco smoke exposure assumes 25% of the day at work and 65% of the day at home, with concentrations of 50–350 $\mu$ g/m <sup>3</sup>	WHO 2000
Smoking 20 cigarettes/day Environmental tobacco smoke	1.0 0.1–1.0	Authors noted that environmental tobacco smoke can contribute 10%– 25% of indoor exposure	Fishbein 1992
Residential indoor air Conventional home Mobile home	0.3–0.6 1.0	Assumes 65% of time at home, 30–60 $\mu$ g/m <sup>3</sup> for conventional home, and 100 $\mu$ g/m <sup>3</sup> for mobile home	WHO 2000
Residential indoor air Conventional home Prefabricated home Outdoor air	0.5–2.0 1.0–10.0 0.02	Assumes 65% of day spent in residence and 10% of day spent outdoors	Fishbein 1992
Indoor air Outdoor air	1.0 0.1	Estimates for the Finnish population	HSDB 2007
Outdoor air	0.002–0.04	Assumes 10% of time spent outdoors and 2 m <sup>3</sup> /d intake at 1–20 $\mu$ g/m <sup>3</sup> concentration	WHO 2000

 Table 2-23. Estimated formaldehyde exposure levels

Source	Intake (mg/day)	Comment	Reference
Drinking water	< 0.2	Assumes that concentrations in drinking water are normally less than 0.1 mg/L	WHO 1989
Cosmetics Hand cream Suntan lotion	$0.1^{a}$ $0.85^{a}$	Hand-cream exposure assumes 2-g/application containing 2 mg of formaldehyde and 5% absorption; same assumptions for suntan lotion except 17 g applied	ATSDR 1999

<sup>a</sup>Milligrams absorbed per application.

## 1 **2.7 Regulations and Guidelines**

2 2.7.1 Regulations

## 3 Coast Guard, Department of Homeland Security

- 4 46 CFR 150 and 151 detail procedures for shipping formaldehyde, formaldehyde
- 5 solution, and 1,3,5-trioxane with incompatible chemicals.

## 6 Consumer Product Safety Commission (CPSC)

- 7 Formaldehyde and products containing  $\geq 1\%$  of formaldehyde are considered "strong
- 8 sensitizers" and must contain a warning label.

## 9 U.S. Environmental Protection Agency (EPA)

- 10 Clean Air Act
- 11 Clean-Fuel Vehicles: Formaldehyde emissions limits have been established for various
- 12 classes of clean-fuel vehicles.
- 13 Control of Emissions from New and In-Use Highway Vehicles and Engines:
- 14 Formaldehyde emissions limits have been established for various classes of vehicles.
- 15 National Emissions Standards for Hazardous Air Pollutants: Listed as a hazardous air16 pollutant.
- 17 New Source Performance Standards: Manufacture of formaldehyde is subject to certain18 provisions for the control of VOC emissions.
- 19 Prevention of Accidental Release: Threshold quantity (TQ) = 15,000 lb.
- 20 Regulation of Fuels and Fuel Additives: Under reformulated gasoline certification
- 21 requirements, formaldehyde emissions levels must not be exceeded.
- 22 Clean Water Act
- 23 Designation of Hazardous Substances: Formaldehyde and paraformaldehyde both are
- 24 listed as hazardous substances.
- 25 Comprehensive Environmental Response, Compensation, and Liability Act
- Formaldehyde reportable quantity (RQ) = 100 lb.

- 1 Paraformaldehyde RQ = 1,000 lb.
- 2 *Emergency Planning and Community Right-To-Know Act*
- 3 TRI: Listed substance subject to reporting requirements.
- 4 RQ = 100 lb.
- 5 Threshold planning quantity (TPQ) = 500 lb.
- 6 Radiation Protection Programs
- 7 Health and Environmental Protection Standards for Uranium and Thorium Mill Tailings:
- 8 Formaldehyde will be monitored for in groundwater and shall not exceed either the
- 9 background level or another concentration level determined for that site.
- 10 Resource Conservation and Recovery Act
- 11 Listed as hazardous waste: Waste codes in which listing is based wholly or partly on
- 12 formaldehyde U122, K009, K010, K038, K040, K156, and K157.
- 13 Listed as a hazardous constituent of waste.
- 14 Land disposal restrictions have been promulgated under 40 CFR 268.

## 15 Food and Drug Administration (FDA)

- 16 Numerous formaldehyde-based chemicals may be used as components of adhesives and
- 17 coatings in packaging, transporting, or holding food provided that conditions prescribed
- 18 in 21 CFR 175 are met.
- 19 Numerous formaldehyde-based chemicals may be safely used as articles intended for use
- 20 in contact with food provided that conditions prescribed in 21 CFR 177 are met.
- 21 Numerous formaldehyde-based chemicals may be used in the production of paper
- 22 products intended for use in producing, processing, preparing, treating, packaging,
- transporting, or holding food provided that conditions prescribed in 21 CFR 176 are met.
- 24 Formaldehyde and formaldehyde-based chemicals may be used as adjuvants, production
- aids, and sanitizers that come in contact with foods provided that conditions prescribed in
   21 CFR 178 are met.
- Formaldehyde-based ion-exchange resins may be used in the treatment of food provided
  that conditions prescribed in 21 CFR 173 are met.
- Formaldehyde may be safely used in the manufacture of animal feeds in accordance withconditions prescribed in 21 CFR 573.460.
- Formalin, containing approximately 37% formaldehyde gas by weight, can be used in
- 32 environmental waters for the control of fungi and parasites for certain finfish and
- 33 shellfish given restrictions prescribed in 21 CFR 529.

# 34 U.S. Department of Housing and Urban Development (HUD)

- 35 All plywood and particleboard materials bonded with a resin system or coated with a
- 36 surface finish containing formaldehyde shall not exceed the following emission levels

- 1 when installed in manufactured homes: 0.2 ppm for plywood and 0.3 ppm for
- 2 particleboard.
- 3 Manufactured homes must prominently display a notice which provides information on
- 4 formaldehyde sources, levels, health effects, and remedial actions to reduce indoor levels.

# 5 Mine Safety and Health Administration

- 6 Approval Requirements for Permissible Mobile Diesel-Powered Transportation
- 7 Equipment: Engine exhaust from mobile diesel-powered transportation equipment must
- 8 be diluted with air so that the mixture contains no more than 0.001% by volume of
- 9 aldehydes, calculated as equivalent formaldehyde.

# 10 Occupational Safety and Health Administration (OSHA)

- 11 Permissible exposure limit (PEL) = 0.75 ppm.
- 12 Short-term exposure limit = 2 ppm (15-minute exposure).
- 13 Action level = 0.5 ppm (8-hour TWA).
- 14 Comprehensive standards have been developed for occupational exposure to
- 15 formaldehyde gas, its solutions, and materials that release formaldehyde.
- 16 Requirements for preventing or minimizing the consequences of catastrophic releases of
- toxic, reactive, flammable, or explosive chemicals are prescribed in 29 CFR 1910.119;
- 18 the TQ for formaldehyde is 1,000 lb.

# 19 Pipeline and Hazardous Materials Safety Administration

- 20 Formaldehyde, formalin, and paraformaldehyde are considered hazardous materials, and
- 21 special requirements have been set for marking, labeling, and transporting these
- 22 materials, as prescribed under 49 CFR 172.
- 23 2.7.2 Guidelines

# 24 American Council of Governmental Industrial Hygienists (ACGIH)

- 25 Threshold limit value ceiling (TLV-C) = 0.3 ppm.
- 26 Listed as a suspected human carcinogen.

# 27 National Institute for Occupational Safety and Health (NIOSH)

- 28 Recommended exposure limit (REL) = 0.016 ppm.
- 29 Immediately dangerous to life and health (IDLH) level = 20 ppm.
- 30 Ceiling recommended exposure limit = 0.1 ppm (15-minute exposure).
- 31 Listed as a potential occupational carcinogen.

32

#### 1 2.8 Summary

2 Formaldehyde has numerous industrial and commercial uses and is produced in very 3 large amounts (billions of pounds per year in the United States) by catalytic oxidation of 4 methanol. Its predominant use, accounting for roughly 55% of consumption, is in the 5 production of industrial resins, which are used in the production of numerous commercial 6 products. Formaldehyde is used in industrial processes primarily as a solution (formalin) 7 or solid (paraformaldehyde or trioxane), but exposure is frequently to formaldehyde gas, 8 which is released during many of the processes. Formaldehyde gas is also created from 9 the combustion of organic material and can be produced secondarily in air from 10 photochemical reactions involving virtually all classes of hydrocarbon pollutants. In 11 some instances, secondary production may exceed direct air emissions. Formaldehyde is 12 also produced endogenously in humans and animals.

13 Formaldehyde is a simple, one-carbon molecule that is rapidly metabolized, is 14 endogenously produced, and is also formed through the metabolism of many xenobiotic 15 agents. Because of these issues, typical biological indices of exposure, such as levels of 16 formaldehyde or its metabolites in blood or urine, have proven to be ineffective measures 17 of exposure. Formaldehyde can bind covalently to single-stranded DNA and protein to 18 form crosslinks, or with human serum albumin or the N-terminal value of hemoglobin to 19 form molecular adducts, and these reaction products of formaldehyde might serve as 20 biomarkers for exposure to formaldehyde.

Occupational exposure to formaldehyde is highly variable and can occur in numerous industries, including the manufacture of formaldehyde and formaldehyde-based resins, wood-composite and furniture production, plastics production, histology and pathology, embalming and biology laboratories, foundries, fiberglass production, construction, agriculture, and firefighting, among others. In fact, because formaldehyde is ubiquitous, it has been suggested that occupational exposure to formaldehyde occurs in all work places. 1 Formaldehyde is also ubiquitous in the environment and has been detected in indoor and

2 outdoor air; in treated drinking water, bottled drinking water, surface water, and

3 groundwater; on land and in the soil; and in numerous types of food.

4 The primary source of exposure is from inhalation of formaldehyde gas in indoor settings

5 (both residential and occupational); however, formaldehyde also may adsorb to respirable

6 particles, providing a source of additional exposure. Major sources of formaldehyde

7 exposure for the general public have included combustion sources (both indoor and

8 outdoor), automobile emissions, off-gassing from numerous construction and home

9 furnishing products, off-gassing from numerous consumer goods, and cigarette smoke.

10 Ingestion of food and water can also be a significant source of exposure to formaldehyde.

11 Numerous agencies, including the Department of Homeland Security, CPSC, EPA, FDA,

12 HUD, the Mine Safety and Health Administration, OSHA, the Pipeline and Hazardous

13 Materials Safety Administration, ACGIH, and NIOSH, have developed regulations and

14 guidelines to reduce exposure to formaldehyde.

This Page Intentionally Left Blank

93

# 1 3 Human Cancer Studies

2 This section reviews the body of epidemiologic literature on formaldehyde exposure and 3 human cancer risk. Case reports and other descriptive studies are less informative for 4 evaluating causality and are therefore excluded from this review. Also, some analytic 5 studies are excluded from this review (Andersen et al. 1982, Brinton et al. 1984, Fondelli 6 et al. 2007, Goldoft et al. 1993, Hernberg et al. 1983b, Hernberg et al. 1983a, Linos et al. 7 1990, Nisse *et al.* 2001) due to excessively small sample size, because the evaluation of 8 formaldehyde exposure was not designed to be an *a priori* study hypothesis, or because a 9 more recent study completely subsumes a previous analysis conducted with the same 10 study population. Further exclusions are cited in the corresponding sections relevant to 11 these studies.

12 The vast majority of the epidemiologic literature on formaldehyde and cancer is focused 13 on occupational, rather than recreational or environmental, exposures. Industries known 14 to involve formaldehyde exposure include formaldehyde production or other chemical 15 manufacture using formaldehyde resins; wood, plywood, particleboard, and paper 16 manufacture; garment and other textile manufacture; work in foundries; production of 17 glass fibers, plastics, and rubber products; health professions, including pathology and 18 embalming; and other miscellaneous occupations (see Section 2.4 for more information 19 about exposed occupations). To date, only one study has evaluated residential 20 formaldehyde exposure and cancer risk among individuals living in mobile homes 21 constructed with formaldehyde-treated material (Vaughan et al. 1986b); however, this 22 study is excluded from this review because the exposed number of cases was too small 23 for meaningful analysis.

In 2004, an International Agency for Research on Cancer (IARC) working group

25 concluded that there was significant evidence from studies in humans for the

26 carcinogenicity of formaldehyde and classified formaldehyde as a *known human* 

27 carcinogen (Group 1) (IARC 2006). There have been numerous reviews with conflicting

28 reviews on interpretation of the literature, but these are not discussed in this section.

1 Particular attention is placed in the individual study summaries on results for sites in the

- 2 head and neck that come into direct contact with formaldehyde, including cancers of the
- 3 paranasal sinuses, nasal cavity, and nasopharynx. Section 3.1 briefly describes cancers of
- 4 the upper respiratory system for the purposes of this review.

5 Section 3 is organized primarily by study design. Historical cohort and proportionate

6 mortality studies are first reviewed by major industry in Section 3.2, followed by a

7 review of case-control studies organized by cancer site in Section 3.3. Section 3.4

8 summarizes studies by cancer site.

# 9 **3.1 Description of head and neck cancers**

10 Head and neck cancers associated with the upper respiratory tract include cancers of the 11 paranasal sinuses and nasal cavity, nasopharynx, oral (or buccal) cavity and salivary 12 glands, pharynx, larynx, and trachea. Cancers of the brain, eye, and thyroid are not 13 usually defined as cancers of the head and neck. The National Cancer Institute estimates 14 that head and neck cancers account for up to 5% of all cancers in the United States. Head 15 and neck malignancies, especially sinonasal and nasopharyngeal cancers, are common 16 endpoints for epidemiological investigations of formaldehyde because these sites come 17 into direct contact with both airborne and dust-borne exposure. See Figure 5-1 for an 18 illustration of the upper respiratory system.



# **Figure 3-1. Upper respiratory system** (Illustration prepared by Donna Jeanne Corocran, ImageAssociates.)

2 Sinonasal carcinoma comprises all cancers of the paranasal sinuses and nasal cavity,

- 3 which are small hollow spaces lined with mucosal tissue in and around the nose. The
- 4 histology of these tumors is primarily squamous-cell (60% to 70%). Pharyngeal

5 carcinomas (also known as throat cancer) are also primarily squamous-cell type and

- 6 include nasopharyngeal, oropharyngeal, and hypopharyngeal carcinomas. Oro- and
- 7 hypopharyngeal carcinomas are often grouped together in epidemiologic studies. Most
- 8 studies of formaldehyde exposure and pharyngeal cancer have focused only on
- 9 nasopharyngeal cancers since the nasopharynx is thought to be the primary site of contact
- 10 in the pharynx following inhalation exposure to formaldehyde.

1

#### 1 3.2 Cohort standardized and proportionate mortality and incidence studies 2 This section reviews historical cohort (standardized and proportionate mortality and 3 incidence) studies that examined the association between occupational exposure to 4 formaldehyde and cancer. Case-control analyses nested within cohort studies are also 5 reviewed in this section. Studies are divided by industrial sector and professional groups 6 to respect differences between these study populations with regard to the potential for 7 exposure to formaldehyde, as well as differences between potentially confounding 8 concomitant occupational exposures present in each industry. Information on known 9 confounding factors (e.g., smoking) is noted in each study summary whenever such 10 information was collected by study investigators.

11 Several of the following cohort studies have been updated recently, and the results 12 presented in this review will generally be limited to the most recent findings from each 13 cohort and unique re-analyses within the cohort. Studies conducted in the industrial 14 sector will be reviewed first, including those conducted with workers in the fiberglass, 15 garment, chemical, plastics, iron, and plywood and woodworking industries. A review of 16 proportionate mortality studies of professional groups that use formaldehyde as a tissue 17 preservative follows, including studies of pathologists, anatomists, embalmers, and 18 funeral directors. Notably, none of the studies of professional groups examined cancer 19 risk by estimated level of exposure to formaldehyde; rather, this collection of studies 20 examined cancer outcomes by occupation only. Table 3-1 summarizes the characteristics 21 of the major studies. Findings for the tumor sites of interest from these studies are 22 reported in Table 3-3 to 3-8 (see Section 3.4).

Reference	Study population and follow up	Exposure assessment and exposure levels	Analyses and related studies
Andjelkovich et al. 1994, 1995	Workers at an iron foundry in Michigan, USA N = 8,147 Subcohort of formaldehyde_exposed	Occupational histories obtained from employment records and classified using a JEM <i>Exposure level (ppm)</i> low 0.05 medium 0.5	Standardized mortality analysis on formaldehyde exposed workers Nested case-control study of lung cancer (N
	workers: N = 3,929 1959–87 or 89	high 1.5	= 200) from entire cohort
Beane Freeman <i>et al.</i> 2009 Hauptmann <i>et al.</i> 2003, 2004 (update of Blair 1986)	NCI cohort, USA N = 25,619 Hauptmann et al. 2003 Follow-up 1966–94 median yr 35 Person-yrs 865,708 Beane Freeman et al. Follow-up 1966–2004 median yr 42 Person-yrs 998,106	Occupational histories obtained from company records, interviews, and industrial hygiene monitoring from 1980; exposure was classified by level and frequency of peak exposure, average exposure, cumulative exposure, and duration <i>Exposure levels and duration for</i> <i>exposed workers (median and</i> <i>range)</i> Average intensity (ppm) 0.3 (0.01–4.25) Cumulative (ppm-yrs) 0.6 (0–107.4) Duration 2 yrs (0–46) All workers 82.5% exposed to formaldehyde 4.7% employed in jobs with $\geq 2$ ppm average intensity 22.6 % employed in jobs involving $\geq 4$ ppm peak exposure	Standardized mortality and internal analysis Beane Freeman <i>et al.</i> Lymphohematopoietic malignancies Hauptmann <i>et al.</i> Lymphohematopoietic malignancies and solid tumors Potential confounding from exposure to 11 occupational substances and working as a chemist or lab technician was evaluated Reanalysis of lung, leukemia and NPC by Marsh and Youk 2004, 2005, and Marsh <i>et al.</i> 2007b Follow-up of Wallingford cohort by Marsh <i>et al.</i> 1994a,b, 1996, 2002 and 2007a, cohort findings and nested case-control study on pharyngeal cancer (N = 17)

Table 3-1. Summary of cohort studies and nested case-control studies

Reference	Study population and follow up	Exposure assessment and exposure levels	Analyses and related studies
Bertazzi <i>et al.</i> 1986	Workers at a resin manufacturing plant in Italy N = 1,332	Occupational histories obtained from plant employment records and classified by job title and task	Standardized mortality study for few cancer sites Subcohort exposed to
	1959–86	Air sampling 1974, 1978, 1979 Average 0.13–2.53 ppm	formaldehyde (N not reported but represent 5,731 person years)
		Maximum 0.33–6.5 ppm	Employment length and time since first exposure available for lung and alimentary tract
Bond <i>et al</i> . 1986	Male workers employed at Dow Chemical production facility in Texas	Occupational histories and potential for exposure obtained from records, and information on smoking from interviews	Nested case-control study on lung cancer (N = 308)
	N = 19,608 1940–80	Exposure levels not reported	
Chiazze <i>et al.</i> 1997	Male workers employed at an Owens Corning fiberglass manufacturing plant in South Carolina, USA (N = 4,631) 1951–91	Occupational histories obtained by interview and a historical exposure reconstruction; exposure was classified by a committee of experts <i>Exposure levels</i> Each process was assigned to 1 of 4 exposure levels with mid points ranging from 0.05 to 1.5 ppm	Nested case-control study of lung cancer (N = 47)
		times duration) was estimated for each worker	
Coggon <i>et al.</i> 2003 (update of Acheson <i>et al.</i> 1984)	British Chemical Workers Study, UK N = 14,014 males 1941–2000	Occupational histories obtained from company employment records and classified using plant-specific JEMs <i>Exposure levels</i> Estimated from measurements taken after 1970 and recall of workers' irritant symptomsLevel (ppm) $& 0$ of workers $< 0.1$ 27.6% $0.1-0.5$ 27.2 $0.6-2.0$ 9.7% $> 2.0$ 28.5%Most of which were from the British Industrial Plastics plant	Standardized mortality study SMRs provided for ever exposed and highly exposed; SMR for three levels of exposure, and employment duration provided for lung and stomach

Reference	Study population and follow up	Exposure assessment and exposure levels	Analyses and related studies
Dell and Teta 1995	Male workers employed at a Union Carbide plastics manufacturing plant in New Jersey, USA N = 5,932 1946–88	Occupational histories obtained using employment records Exposure levels not reported	Standardized mortality study Workers exposed to formaldehyde (N = 111)
Edling <i>et al.</i> 1987b	Male and female workers at an abrasive materials manufacturing plant, Sweden N = 506 blue collar workers Mortality 1958–83 Incidence 1958–81	Exposure monitoring in plant from 1970 No individual exposure assessment reported <i>Exposure levels</i> Grinding wheel manufacturing [0.08–0.8 ppm] Abrasive belts (N = 59 workers) Peaks [16–25 ppm)]	Standardized mortality and incidence study Unknown number of workers exposed to formaldehyde in grinding wheel process; 59 making abrasive belts Results reported for males only, and for few cancer sites
Hansen and Olsen 1995, 1996	Danish workers at 265 companies producing or using 1 kg/individual year N = 2,041 men, and 1,263 women 1970–84	Occupational information obtained form Danish product Registry Individuals assigned to low or high exposure based on "white or blue collar" status based on pension records Exposure levels not reported	Standardized proportionate cancer incidence for various cancers Workers were included in study if their longest employment was 10 years prior to cancer diagnosis (Original study population = 126,347 men and women) Findings for some cancer sites provided for low formaldehyde exposure, and formaldehyde and woodworkers (combined)

Reference	Study population and follow up	Exposure assessment and exposure levels	Analyses and related studies
Marsh <i>et al.</i> 2001, Stone <i>et al.</i> 2001, Youk <i>et al.</i> 2001	Workers employed at 10 fiberglass manufacturing facilities in the US N = 32,110 1946–92	Occupational histories obtained from company employment records and relevant industrial hygienic literature; Exposure estimated using job location- weighted measures <i>Exposure level</i> Median average intensity 0.066 ppm Median cumulative exposure 0.173 ppm-yr	Nested case control of cancers of the respiratory system
Ott <i>et al.</i> 1989	Workers employed in 2 Union Carbide Corporation chemical manufacturing facilities and a research and development center, USA N = 29,139 1940–78	Occupational histories obtained from company employment records and classified using a JEM Exposure levels not reported	Nested case-control study of lymphohematopoietic malignancies (N = 129)
Partanen <i>et al.</i> 1985, 1990, 1993	Workers employed in 135 particleboard, plywood and formaldehyde glue factories and sawmills in Finland N = 7,703 1944–65	Occupational histories and air quality monitoring data obtained from company employment records and classified using a JEM <u>Exposure levels determined from</u> <u>hygienic data (ppm)</u> Low 0.1–1 Medium 1–2 Heavy > 2 Workers considered exposed to formaldehyde if minimum exposure was 0.1 ppm and cumulative exposure was > 3 ppm-month 83% of subjects in respiratory case-control study exposed to cumulative exposure of less than < 0.25 ppm-yr	Nested case-control studies of lymphohematopoietic malignancies (N = 24 in 1993 study) and respiratory cancer (N = 136)

Reference	Study population and follow up	Exposure assessment and exposure levels	Analyses and related studies
Pinkerton <i>et al.</i> 2004 (update of Stayner et al. 1985, 1988 PMR and SMR study respectively	NIOSH cohort of garment workers, USA N = 11,039 SMR 1955–1998 PMR 1959–1982	All workers considered exposed; personal exposure levels available from plant monitoring programs <i>Exposure levels</i> 3 plants in 1981 to 1984 Median 8 hr TWA (ppm) 0.15 (0.09–0.20) Median duration = 3.3 years Exposures prior to the 1970s were estimated to be as high as 10 ppm	Standardized mortality study Analysis by duration of exposure, time since first exposure, and time of first exposure performed for a few slected cancer sites PMR study included 285 deaths, PCMR ratios were also calculated to correct for healthy worker effect
Stellman <i>et al.</i> 1998, Boffetta <i>et</i> <i>al.</i> 1989	Workers employed in the wood industry American Cancer Society Cancer Prevention Study, USA N = 362,823 Formaldehyde-exposed workers NR (365 cancer deaths) Formaldehyde-exposed woodworkers (N = 387) 1955–98	Occupational histories obtained by interview and classified by job title and task Exposure levels not reported Findings reported for ever exposed	Mortality study Internal analyses using non-woodworkers or workers not exposed to wood dust as the reference group Nested case-control study of multiple myeloma (N = 282) (Boffetta <i>et al.</i> 1989)
Stern <i>et al.</i> 1987	Workers employed in two chrome leather tannery plants, USA N = 9,365 1940–79 or 1980	Occupational history obtained from industrial hygiene surveys <i>Exposure levels in finishing</i> <i>department (ppm)</i> Mean (range) 2.45 (0.5–7)	Standardized mortality study, including formaldehyde-exposed workers in the finishing department (no. exposed workers not stated; 118 cancer deaths observed)
SMR and PMR cohort studies of professional workers (Pathologists, Anatomists, and Embalmers)			
Hall <i>et al.</i> 1991 (update of Harrington and Shannon 1975, and Harrington and Oakes 1984)	Pathologists, members of professional organizations in the UK 3,872 1974–87	Employment status No information on exposure levels	Standardized mortality study

Reference	Study population and follow up	Exposure assessment and exposure levels	Analyses and related studies
Hayes <i>et al.</i> 1990	Deceased embalmers and funeral directors identified using licensing board records, death certificates, and other sources, USA N = 4,046 1975–85	Employment status No information on exposure levels	Proportionate mortality study
Levine et al. 1984	Licensed embalmers in Ontario, Canada N = 1,413 1950–1977	Licensing records No information on exposure levels	Standardized mortality study
Stroup <i>et al.</i> 1986	Anatomists who were members of the American Association of Anatomists, USA N = 2,317 1888–1979	Employment status No information on exposure levels	Standardized mortality study Findings for brain and lung analyzed by length of membership
Walrath and Fraumeni 1983	All licensed embalmers in New York, USA N = 1,263 1902–80	Licensing records No information on exposure levels Some cancer sites analyzed by age since first license	Proportionate mortality study Findings for a few cancer sites analyzed by latency and type of license (embalmers only and funeral directors and embalmers)
Walrath and Fraumeni 1984	All licensed embalmers in California, USA N = 1,109 1916–80	Licensing records No information on exposure levels Employment duration estimated by length of licensure	Proportionate mortality study

1 3.2.1 National Cancer Institute (NCI) Cohort: mixed industries

2 Blair and colleagues at the National Cancer Institute (NCI) assembled the largest cohort

3 of industrial workers to date to assess the risk of several cancers suspected of being

- 4 associated with exposure to formaldehyde, including leukemia and cancer of the brain,
- 5 lung, oral cavity, and pharynx (Blair *et al.* 1986). This cohort captured workers from
- 6 various industries that used formaldehyde, including plants that manufactured resin,
- 7 plastic, photographic film, and plywood. The authors also measured several concurrent

1 occupational exposures (and potential confounding agents), such as asbestos, wood dust,

2 and solvents.

3 Previous studies (Fayerweather et al. 1983, Liebling et al. 1984, Marsh 1982, Marsh et 4 al. 1994a, Marsh et al. 1994b, Wong 1983) included workers who were later included in 5 the NCI study; the findings of these studies are considered subsumed by NCI analyses for 6 the purposes of this review. Likewise, earlier analyses of the NCI cohort (Blair and 7 Stewart 1989, Blair et al. 1990b, Callas et al. 1996, Marsh et al. 1994a, Marsh et al. 8 1992a, 1992b, Marsh et al. 1994b, Robins et al. 1988, Sterling and Weinkam 1988, 9 1989a, 1989b, 1994, Stewart et al. 1989) will not be discussed in detail since more recent 10 and updated analyses are available on the same study population.

11 Study population and follow-up. Using records from the Formaldehyde Institute, trade 12 organizations, and other sources, including chemical producers, approximately 200 13 companies reported to use or produce formaldehyde were identified. The 10 industrial 14 plants with the largest number of employees and longest history of formaldehyde use 15 were selected for inclusion into the cohort. Three of the plants produced formaldehyde, 16 six produced formaldehyde resins, six produced molding compounds, two produced 17 molded plastic products, two produced photographic film, and one produced plywood 18 (some plants produced more than one product). The study cohort consisted of all workers 19 first employed at the selected plants before January 1, 1966 (N = 26,561; 93% white, 20 12% female). Workers were originally followed through January 1, 1980 to determine 21 vital status and cause of death. Hauptmann et al. (2003, 2004) extended the mortality 22 follow-up through December 31, 1994 for analyses of lymphohematopoietic 23 malignancies (N = 178 deaths) and solid cancers (N = 1,921 deaths), respectively. The 24 NCI cohort was most recently extended through December 31, 2004, resulting in a 25 median follow-up time for workers of 42 years, representing 998,106 person-years of 26 exposure among 25,619 workers, 4,359 of whom were classified as never exposed to 27 formaldehyde. A total of 13,951 deaths were identified from 1943 to December 31, 2004. 28 Beane Freeman et al. (2009) have published the results for lymphohematopoietic cancers 29 from the extended follow-up, which are `described after the results of the Hauptmann et 30 al. analysis, below.
1 *Exposure assessment.* Exposure to formaldehyde was comprehensively reconstructed 2 using work histories collected through 1980 on the basis of job titles, tasks, plant visits by 3 industrial hygienists, information from workers and plant managers, as well as 4 monitoring data. Peak exposures (less than 15 minutes) exceeding the 8-hour time-5 weighted average formaldehyde exposure intensity were estimated by an industrial 6 hygienist using the work histories. In addition to highest peak exposure (unexposed, 0.1 7 to 1.9 ppm, 2.0 to 3.9 ppm,  $\geq$  4 ppm) and frequency of peak exposure (none, hourly, 8 daily, weekly, monthly), time-dependent estimates also were calculated for duration of 9 exposure (years), average exposure (ppm), and cumulative exposure (ppm-years). 10 Exposure-response trends were considered using cut-points at the 60th and 80th 11 percentiles of the distribution of exposure in exposed subjects who died from cancer. 12 Several important cofactors were assessed, including exposure to particulates and 11 13 other widely used chemicals in the plants (i.e., antioxidants, asbestos, carbon black, dyes 14 and pigments, hexamethylenetetramine, melamine, phenol, plasticizers, urea, wood dust, 15 and benzene), routine use of respirators by workers, and duration of employment as a 16 chemist or laboratory technician.

Among jobs considered exposed to formaldehyde (83.4%), the median 8-hour timeweighted average exposure was 0.45 ppm (range = 0.01 to 4.25 ppm); median values were 2 years (range = 0 to 46 years) for duration, 0.3 ppm (range = 0.01 to 4.25 ppm) for average intensity, and 0.6 ppm-years (range = 0.0 to 107.4 ppm-years) for cumulative exposure. Average intensity was 2 ppm or higher for nearly 3% of jobs, and peak exposures reached 4 ppm or higher for over 14% of jobs. Approximately 0.5% (N = 133) of workers ever used a respirator routinely.

The authors noted that smoking information was not available for most of the cohort.
Smoking was not considered to be a source of confounding, however, since analysis of a
sample of workers revealed no major differences in smoking prevalence by cumulative
formaldehyde exposure.

Statistical methods. Standarized mortality ratios (SMRs) were calculated using sex-, race,
 age-, and calendar-year-specific U.S. mortality rates. To investigate the association

1 between exposure to formaldehyde and cancer mortality, log-linear Poisson regression 2 was applied stratified by calendar year, age, sex, race, and pay category. Potential 3 confounding was evaluated for exposure to 11 concomitant occupational substances 4 (ever/never), as well as working as a chemist or lab technician (years). Exposure lags 5 ranging from 2 to 20 years were considered to account for latency; all exposures were 6 subsequently calculated using a 2-year lag interval for the analyses of 7 lymphohematopoietic malignancies (Beane Freeman et al. 2009, Hauptmann et al. 2003) 8 and a 15-year lag interval for the analyses of solid cancers (Hauptmann *et al.* 2004). 9 *Results*. Person-years at risk (456,635) among exposed workers and person-years 10 (409,074) among unexposed workers were compared in external analyses in the 1994 11 cohort update, lagged by 15 years. Compared with the U.S. population, Hauptmann et al. 12 (2004) found that mortality from all cancers was lower than expected both in unexposed 13 (SMR = 0.65, 95%) confidence interval [CI] = 0.56 to 0.75, 166 deaths for 2-year lag) and 14 exposed workers (SMR = 0.90, 95% CI = 0.86 to 0.94, 1,755 deaths for 2-year lag),

15 regardless of length of the exposure lag.

16 *Lymphohematopoetic cancers*. Hauptmann *et al.* (2003) presented data on

17 lymphohematopoietic cancers from the 1994 NCI cohort update, and Beane Freeman et 18 al. (2009) conducted external and internal analyses of lymphohematopoietic cancers 19 through the latest follow-up of the cohort through December 31, 2004, which represents a 20 total of 998,106 person-years of employment among 25,619 workers employed prior to 21 1966 (4,359 of whom were classified as never exposed to formaldehyde). Beane Freeman 22 et al. (2009) noted that a total of 1,004 deaths were identified that were not included in 23 the previous 1980 to 1994 follow-up and 4 subjects were misclassified as deaths but 24 found to be living. In addition, several deaths for lymphohematopoietic cancers that were 25 included in the Hauptmann et al. (2003) analysis were recoded: 6 deaths (one multiple 26 myeloma, one myeloid leukemia, one Hodgkin's lymphoma and three myelofibrosis 27 deaths) were re-classified as non-lymphohematopoetic cancers, and two multiple 28 myelomas were added. The data reported below are confined to the 2004 update reported 29 by Beane Freeman et al. (2009) unless clear differences between findings in this update 30 and the earlier (1994) update were observed. P values for trends in the text refer to the

exposed group only, using the lowest exposure group as the referent, unless otherwise
 stated; *P* values for trends using the unexposed and exposed groups, and exposed groups

3 only are reported in Table 3-2.

4 A total of 319 deaths from all lymphohematopoietic cancers were identified to the end of

5 follow-up in 2004; 286 among ever-exposed and 33 among never-exposed workers. In

6 external analyses, the SMRs for all lymphohematopoietic cancers was similar to national

7 rates in both the exposed and nonexposed groups, using a 2-year lag time for exposure

8 (SMR = 0.94, 95% CI = 0.84 to 1.06, 286 deaths, and SMR = 0.86, 95% CI = 0.61 to

9 1.21, 33 deaths, respectively; compared with U.S. population rates). An increased risk for

10 Hodgkin's lymphoma was observed, but SMRs for other subtypes of

11 lymphohematopoietic cancers among the exposed workers were similar to unexposed

12 rates or the U.S. population. Findings were generally similar to the 1994 findings.

Cancer type	2004 Update RR (95%CI); N <sup>a</sup>	₽ <sub>trend</sub> b	P <sub>trend</sub> c	1994 Update RR (95%CI); N <sup>a</sup>	₽ <sub>trend</sub> b	₽ <sub>trend</sub> c
All LH	1.37 (1.03–1.81); 108	0.02	0.04	1.87 (1.27–2.75); 64	0.002	0.002
All leukemia	1.42 (0.92–2.18); 48	0.12	0.02	2.46 (1.31–4.62); 29	0.004	0.001
Myeloid leukemia	1.78 (0.87–3.64); 19	0.13	0.07	3.46 (1.27–9.43); 14	0.009	0.003
Lymphatic leukemia	1.15 (0.54–2.47); 14	> 0.50	0.30	1.39 (0.46–4.17); 7	> 0.50	0.279
Other leukemia	1.15 (0.53–2.53); 13	> 0.50	0.50	2.47 (0.69–8.87); 7	0.154	0.277
Hodgkin's lymphoma	3.96 (1.31–12.02); 11	0.01	0.004	3.35 (0.97–11.59); 8	0.042	0.014
Multiple myeloma	2.04 (1.01-4.12); 21	0.08	> 0.50	1.67 (0.68–4.12); 11	0.355	> 0.50
NHL	0.91 (0.55–1.49); 28	> 0.50	> 0.50	1.23 (0.59–2.55); 15	> 0.50	> 0.50
LH (lymphoid origin)	1.35 (0.97–1.89); 74	0.06	0.10	NR	NR	NR
LH (nonlymphoid origin)	1.80 (0.91–3.57); 21	0.09	0.09	NR	NR	NR

Table 3-2. Lymphohematopoietic (LH) cancers in formaldehyde-exposed workers(NCI cohort and peak exposure: 1994 and 2004 updates)

Source: Beane Freeman *et al.* 2009, Hauptmann *et al.* 2003; see Table 3-7a for detailed data on all exposure categories and for data on average exposure.

LH = lymphohematopoietic; N = number of deaths; NHL = non-Hodgkin's lymphoma; RR = relative risk. <sup>a</sup> Data for peak ( $\geq$  4 ppm vs. > 0–1.9 ppm) exposures, 2-year exposure lag used.

<sup>b</sup>  $P_{\text{trend}}$  for 2-sided likelihood ratio for exposed person-years only.

 $^{c}P_{trend}$  for 2-sided likelihood ratio for exposed and unexposed person-years.

1 In internal analyses of exposed workers, using Poisson logistic regression stratified by

2 age, sex, race, calendar year, and pay category, peak exposures were associated with a

3 significant increase in all lymphohematopoietic deaths combined (RR = 1.37, 95% CI =

4 1.03 to 1.81, 108 deaths, comparing peaks of  $\geq$  4 ppm with > 0 to 2.0 ppm;  $P_{\text{trend}} = 0.02$ ;

5 Table 3-2). No association was observed for all lymphohematopoietic cancers in the 2004

6 update for average intensity of exposure or cumulative exposure (See Table 3-7a in

7 Section 3.4).

8 With respect to specific subtypes of lymphohematopoietic cancers and peak exposures in

- 9 the latest update, deaths from leukemia were elevated (RR = 1.42, 95% CI = 0.92 to 2.18,
- 10 48 deaths, comparing peaks  $\geq$  4 ppm with > 0 to 2.0 ppm;  $P_{\text{trend}} = 0.12$ ); for the subgroup
- 11 of deaths from myeloid leukemia, the highest peak exposure was associated with a
- 12 slightly higher RR of 1.78 (0.87 to 3.64, 19 deaths,  $P_{\text{trend}} = 0.13$ ). There were no clear

1 trends toward increasing risk with increasing average or cumulative exposure to 2 formaldehyde for leukemia although an elevated RR was observed for the highest 3 category of highest category of average intensity of exposure  $\geq 1$  ppm) vs. the lowest 4 category (RR = 1.61, 95% CI = 0.76 to 3.39, 11 deaths,  $P_{\text{trend}} = 0.43$ ) (See Table 3.6a). 5 Deaths from Hodgkin's lymphoma were significantly elevated in the highest peak vs. the 6 lowest peak exposure group and the relative risks increased with increasing peak 7 exposure. (RR = 3.96, 95% CI = 1.31 to 12.02, 11 deaths,  $P_{\text{trend}} = 0.01$ ). RRs for 8 Hodgkin's lymphoma increased with increasing average intensity of exposure ( $P_{\text{trend}} =$ 9 (0.05) and cumulative exposure ( $P_{\text{trend}} = 0.08$ ). Elevated RRs were found for the highest 10 category of exposure vs. lowest category of exposure: RR = 2.48 (95% CI = 0.84 to 7.32, 11 6 deaths, for  $\geq$  1 ppm average intensity of exposure and RR = 1.30 (95 % CI = 0.40 to 12 4.19, 4 deaths for  $\geq$  5.5 ppm-yr cumulative exposure). Peak exposure was also associated 13 with deaths from multiple myeloma (RR = 2.04, 95% CI = 1.01 to 4.12, 21 deaths, P<sub>trend</sub> 14 = 0.08), but no association was found with average or cumulative exposure. Relative risks 15 were also computed for unexposed workers in comparison with the lowest exposure 16 groups for peak, average, and cumulative exposure, and subjects with no estimated 17 exposure to formaldehyde were found to be at significantly increased risk of multiple 18 myeloma compared with low exposed workers for peak and average exposure, but not for 19 cumulative exposure. For other lymphohematopoietic cancers, unexposed workers had 20 similar or lower risks in comparison with the lowest exposed group. Non-Hodgkin's 21 lymphoma was not associated with peak, average or cumulative exposure (See Table 3.2 22 and 3.6a in Section 3.4). 23 In general, the 2004 update confirmed the findings of the 1994 update; however, the

magnitude of the risks estimates for the highest category of peak exposure were higher in the 1994 update compared to the 2004 update, and some of the exposure response relationships were stronger in the earlier update(See Table 3-1). Analyses due to recoding of some of the lypmphohematopoietic cancers did not substantially affect the previously reported results. The 1994 update (Hauptmann *et al.* 2003) also reported findings by duration of exposure (not presented in the 2004 update), and found no statistically significant risk estimates by specific categories of exposure duration and no overall 1 trends with increasing duration.) (Note that if the cohort was censored at 1980, the date

2 after which exposures were assumed to be zero, the risk for myeloid leukemia was

3 increased, according to the authors. If exposure was considered to continue at 1980

4 levels, however, no changes in the results were seen for any of the lymphohematopoietic

5 sites.)

6 Controlling for duration of exposure or for 11 other co-exposures with possible

7 associations with lymphohematopoietic cancers did not alter the above findings, and

8 excluding 586 workers with possible exposure to benzene (a known leukemogen) did not

9 alter the results for lymphatic or myeloid leukemia and peak exposure (data not reported).

10 Similarly, adjusting for plant type did not substantively alter the results.

11 When time period analyses for trends in relative risk were examined, significant excesses 12 of myeloid leukemia in relation to peak exposure were observed up to 1994 and then 13 declined. Risks for all lymphohematopoietic cancers, leukemia, myeloid leukemia, and 14 Hodgkin's lymphoma were highest 15 to 25 years after first exposure. Beane Freeman et 15 al. (2009) concluded that evaluation of lymphohematopoietic risk over time was 16 consistent with the relatively short induction periods characteristic of leukemogenesis, 17 and suggest an association between lymphohematopoietic cancer and formaldehyde 18 exposure, particularly for myeloid leukemia and possibly Hodgkin's lymphoma and 19 multiple myeloma.

20 Solid cancers. Mortality from solid tumors was also lower than expected compared to

21 U.S. rates (SMR among unexposed = 0.78, 95% CI = 0.70 to 0.86, 341 deaths; SMR

22 among exposed = 0.91, 95% CI = 0.87 to 0.96, 1,580 deaths) (Hauptmann *et al.* 2004). A

23 statistically significant excess of mortality from nasopharyngeal cancer was observed

among the exposed group (SMR = 2.10, 95% CI = 1.05 to 4.21, 8 deaths). One death

25 from nasopharyngeal cancer was subsequently re-classified as oropharyngeal cancer and

26 excluded from internal analysis of average, peak, and cumulative exposure, however.

27 SMRs exceeding 1.0 were observed for cancers of the oral cavity (SMR = 1.01, 95% CI =

28 0.77 to 1.34, 49 deaths), nose and nasal cavity (SMR = 1.19, 95% CI = 0.38 to 3.68, 3

deaths) and bone (SMR = 1.57, 95% CI = 0.75 to 1.18, 7 deaths). Lung cancer was not

1 elevated among exposed workers (SMR = 0.97, 0.90 to 1.05, 641 deaths), although it was 2 slightly higher than among the unexposed workers (SMR = 0.79, 95% CI = 0.65 to 0.96, 3 103 deaths). Internal analysis of exposure-response relationships between average, peak, 4 cumulative and duration of exposure to formaldehyde and solid cancers, lagged by 15 5 years, the following results were conducted for nasopharyngeal cancers. 6 Cancer of the nasopharynx was elevated at the highest category of average exposure 7 intensity (RR = 1.67 for  $\geq$  1.0 ppm vs. > 0 to < 0.5 ppm (ref.), 6 deaths); the trend among 8 exposed workers was  $P_{\text{trend}} = 0.066$ , and across exposed and unexposed workers,  $P_{\text{trend}} =$ 9 0.126. For peak exposure, the RR was 1.83 at the maximum peak category of  $\geq$  4.0 ppm 10 (7 deaths), and the tests for trend were  $P_{\text{trend}} < 0.001$  among exposed workers and  $P_{\text{trend}} =$ 11 0.044 across exposed and unexposed workers. For cumulative exposure, the RR was 4.14 12 for the highest exposure category of  $\geq 5.5$  ppm-years, 3 deaths); the  $P_{\text{trend}}$  was 0.025 13 among exposed workers and  $P_{\text{trend}} = 0.029$  across exposed and unexposed workers. For 14 duration of exposure, the RR was 4.18 for the longest duration of  $\geq$  15 years (2 deaths), 15 and the trends were  $P_{\text{trend}} = 0.147$  and  $P_{\text{trend}} = 0.206$  respectively. Because five of the nine 16 nasopharyngeal cancer cases occurred at the Wallingford, CT plant, the authors 17 conducted analyses adjusted for plant and found increasing risks for peak exposure ( $P_{\text{trend}}$ ) 18 among exposed = 0.008), cumulative exposure ( $P_{\text{trend}}$  among exposed = 0.007), and 19 duration of exposure ( $P_{\text{trend}}$  among exposed = 0.043). Plant adjusted relative risks were 20 also higher among worker with higher average exposure (RR = 8.51 for workers exposed 21 to 0.5 < 1 ppm, and 23.54 for workers exposed to > 1 ppm), but the test for trend was not 22 statistically significant ( $P_{\text{trend}}$  among exposed = 0.404) Combining cancers of the upper respiratory tract (i.e. cancers of the salivary gland, 23 24 mouth, nasopharynx, nasal cavity, and larynx) yielded increasing relative risks with 25 increasing average intensity of exposure (RR = 1.69 for 0.5 to 1.0 ppm, 11 deaths; RR =

- 26 2.21 for  $\ge$  1.0 ppm, *P* < 0.05, 15 deaths, CI excluding 1.0; *P*<sub>trend</sub> = 0.122). Cancer of the
- 27 upper respiratory tract was also associated with peak exposure (RR = 1.24, 12 deaths, for
- 28 2.0 to 4.0 ppm; RR = 1.65, 18 deaths, for  $\ge$  4.0 ppm;  $P_{\text{trend}} = 0.142$ ) but not with
- 29 cumulative exposure or duration of exposure. No evidence was observed of a positive
- 30 association between lung cancer mortality and any of the exposure measures, except for a

statistically significant relative risk associated with peak exposure of 2.0 to 4.0 ppm (RR
= 1.45, 227 deaths). A statistically significant decrease in lung cancer risk was observed
for duration of exposure of 5 to 15 years (RR = 0.80, 123 deaths). [The only other
observed statistically significant elevation in risk was a RR of 161 for 42 deaths from
prostate cancer in association with a peak exposure of 2.0 to 4.0 ppm.]

6 The authors noted that RR estimates were not adjusted by plant because plants were

highly correlated with exposure. However, findings from repeated analyses where each
plant was selectively removed from the model one at a time were similar to those from
the analysis including all plants [data not presented].

10 *Re-analyses.* Marsh and Youk (2004) conducted a re-analysis of the updated cohort of 11 Hauptmann et al. (2003) to re-examine mortality risk from leukemia. Exposure-specific 12 SMRs using both local and national reference rates were calculated by highest peak 13 exposure, average intensity, cumulative exposure, duration, and by categorizing 14 formaldehyde exposure into tertiles based on the exposure distribution among all 15 leukemia deaths in exposed workers. Generally, the SMRs increased in magnitude with 16 increasing peak and average intensity of exposure for all leukemias combined and for 17 myeloid leukemia. An internal analysis that applied alternative regression modeling 18 yielded RRs similar to those observed by Hauptmann et al. (2003); a significant 19 exposure-response relationship was observed for all leukemias ( $P_{\text{trend}} = 0.001$ ) and 20 myeloid leukemia ( $P_{\text{trend}} = 0.003$ ) by peak exposure. Tests for trend by average intensity 21 for all leukemias ( $P_{\text{trend}} = 0.193$ ) or myeloid leukemias ( $P_{\text{trend}} = 0.086$ ) were not 22 statistically significant. Exposure tertiles were also examined in these models, and results 23 were similar to that of the NCI exposure categorization ( $P_{\text{trend}} = 0.145$  for all leukemia;  $P_{\text{trend}} = 0.092$  for myeloid leukemia). Duration of time worked in the highest category of 24 25 peak exposure was not associated with leukemia mortality.

In a re-analysis of nasopharyngeal cancer data from the Hauptmann *et al.* (2004) solid
cancer study, Marsh and Youk (2005) suggested that the observed relationship between
nasopharyngeal cancer mortality and formaldehyde was driven largely by one plant in
Wallingford, Connecticut, which had been independently studied by Marsh previously

1 (Marsh et al. 1996, Marsh and Youk 2005, Marsh et al. 2002), since five of the nine 2 nasopharyngeal cancer deaths in the NCI study had occurred among workers at this plant. 3 Marsh and Youk (2005) reported that when the SMR for nasopharyngeal cancers in 4 Plants 2 to 10 combined was re-calculated it was not elevated (SMR = 0.65, 95% CI 0.85 to 2.3, 4 deaths, in comparison with that of plant 1 alone (the Wallingford plant) (SMR = 6 10.3, 95% CI = 3.8 to 22.5, N = 6). Also see separate analyses of the Wallingford plant 7 by Marsh *et al*, 1996, 2002, 2007a, below). In a further re-analysis of the nasopharyngeal 8 cancers observed in the Hauptmann et al. (2004) study, Marsh et al. (2007b) further 9 examined the interaction between the plant and also peak exposures to formaldehyde, 10 since the elevated SMR for nasopharyngeal cancers in the NCI cohort was largely driven 11 by an association with peak (> 4 ppm) exposure to formaldehyde in the Wallingford 12 plant. By examining the interaction between a new 2-factor variable (Plant 1 vs. Plants 2– 13 10) and a continuous variable for peak exposure, Marsh *et al.* concluded that the observed 14 increase in risk of nasopharyngeal cancers in the NCI cohort could be attributable to the 15 effect of an association between peak exposure in Plant 1 and nasopharyngeal cancers 16 and was not generalizable within the entire NCI cohort. In addition, they pointed out that 17 the internal analysis of the NCI cohort was not robust (i.e., the risk estimates obtained 18 were subject to considerable instability depending on the addition of one or more 19 nasopharyngeal cancer death to the cohort) and did not warrant the conclusion of a causal 20 relationship between formaldehyde and nasopharyngeal cancer.

21 Related studies. Marsh et al. (1994a, 1994b, 1996) studied the plastics manufacturing 22 plant in Wallingford, Connecticut that was included in the NCI study; construction of the 23 cohort and exposure assessment at this facility was conducted independently of the NCI 24 study. Mortality in this cohort was updated through December 31, 1998 (Marsh et al. 25 2002) for 7,328 male workers (82% white) employed between 1941 and 1984. The 26 results presented below are from the 1998 update only (Marsh et al. 2002). 27 Approximately half of the individuals in the cohort were employed for less than one year. 28 Exposure estimation through 1995 was based on available sampling data (sporadic 29 measurements were taken between 1965 and 1987), job descriptions, and information 30 from plant personnel including the plant industrial hygienist. Exposure to formaldehyde 31 was estimated for each job and task, yielding measures of average intensity, cumulative

1 exposure, and duration of exposure. Though the exposure assessment for formaldehyde 2 was developed to maximize comparability with the NCI study, the authors noted that 3 exposure estimates were generally less than one tenth of the corresponding values 4 estimated for the same Wallingford workers in the NCI study. Analyses of mortality were 5 performed only for malignant neoplasms of the upper and lower respiratory tract; the 6 person-years method was used to estimate expected mortality rates using both U.S. and 7 local standard populations. A nested case-control study was formed to examine the 8 association between all pharyngeal cancer and exposure to formaldehyde. The 9 conditional logistic regression analysis included 22 cases (5 oropharynx, 7 nasopharynx, 10 3 hypopharynx, and 7 unspecified pharynx), which were matched on race, sex, age, and 11 year of birth (within 2 years) to four controls from the remaining living and deceased 12 members of the cohort. Information about smoking and other relevant exposures was 13 obtained through telephone interviews with study subjects or proxies (68% response 14 among cases, 76% among controls).

Compared with both national and local expected rates (local estimates subsequently 15 16 presented), SMRs were elevated for all cancers of the oral cavity and pharynx (SMR = 17 1.52, 95% CI = 1.03 to 2.15, 31 deaths) including all pharyngeal cancer (SMR = 2.23, 18 95% CI = 1.40 to 3.38, 22 deaths) and cancers of the oropharynx (SMR = 1.80, 95% CI = 19 0.58 to 4.19, 5 cases), nasopharynx (SMR = 5.00, 95% CI = 2.01 to 10.30, 7 deaths), and 20 hypopharynx (SMR = 1.52, 95% CI = 0.31 to 4.43, 3 deaths). Mortality from cancer of 21 the respiratory system was also greater than expected (SMR = 1.22, 95% CI = 1.08 to 22 1.38, 278 deaths), including cancers of the sinonasal cavity (SMR = 3.06, 95% CI = 0.6323 to 8.93, 3 deaths), larynx (SMR = 1.59, 95% CI = 0.84 to 2.71, 13 deaths), and bronchus, 24 trachea, and lung (SMR = 1.21, 95% CI = 1.06 to 1.36, 262 deaths). Standardized 25 mortality ratios for nasopharyngeal cancer increased monotonically with cumulative 26 exposure to formaldehyde. (As noted, no other SMR analyses were presented.) 27 In the nested case-control analysis of all pharyngeal cancers adjusted for smoking and 28 time since hire, the OR for ever being exposed to formaldehyde was 3.04 (95% CI = 0.36)

- 29 to 145.58, 20 deaths). Odds ratio estimates increased with duration of exposure,
- 30 particularly for duration of exposure at jobs with formaldehyde exposure greater than 0.2

1 ppm-years ( $P_{\text{trend}} = 0.163$ ), but did not increase significantly by cumulative exposure or 2 average intensity of exposure.

3 Marsh et al. (2007a) subsequently followed the Wallingford cohort through the end of 4 2003. Vital status was ascertained for 98% of the cohort, and cause of death was 5 determined for 95% of 2,872 deaths. Worker exposures to formaldehyde were 6 reconstructed and unlagged and lagged exposure metrics computed. New external (SMR) 7 analyses and a nested case-control analysis of nasopharyngeal cancers and all other 8 pharyngeal cancers (AOPC) were conducted, taking into account both demographic 9 variables and smoking as in the previous (2002) study, and also the external employment 10 of cases and controls before, during, and after employment at the Wallingford plant, 11 using various sources such as city directories, employment applications and genealogical 12 searches. Based on the frequency of external employment, three external occupational 13 groups were established: silver smithing; other metal work; and military service. No new 14 nasopharyngeal cancer cases were observed (compared with the 2002 analysis) and one 15 additional AOPC was observed, yielding SMRs of 4.43 (95% CI = 1.78 to 9.13, 7 16 nasopharyngeal cancer deaths) and SMR = 1.71 (95% CI = 1.01 to 2.72, 16 AOPC)17 deaths; both compared with local rates). In internal analyses, a statistically significant risk 18 of nasopharyngeal cancer (OR = 14.41, 95% CI = 1.30 to 757.8, 4 deaths), was observed 19 in association with ever working in silver smithing, and an OR of 7.31 (95% CI = 1.08 to 20 82.1, 5 deaths) for ever working in silver smithing and/or other metal work. No 21 association with external employment was observed for AOPC, with the exception of a 22 statistically nonsignificant increase in risk for workers with a history of employment in 23 other metal work (OR = 1.40, 95% CI = 0.31 to 5.1, 4 deaths). The risk of 24 nasopharyngeal cancer associated with formaldehyde exposure before adjustment for 25 smoking and external employment was 1.51 (95% CI = 0.20 to  $\infty$  (infinity), 7 deaths) and 26 after adjustment for smoking and silver smithing and/or metal working employment was 27 2.87 (0.21 to  $\infty$ ). An interaction model suggested that neither nasopharyngeal cancer nor 28 AOPC was associated with formaldehyde in the presence of these external occupations, 29 according to the authors.

1 There was no clear or statistically significant monotonic trend towards increasing 2 nasopharyngeal cancer risk with increasing duration, average intensity or cumulative 3 exposure to formaldehyde before and after adjustment for smoking and silver smithing 4 and/or other metal working employment, although some increase in risk was observed in 5 each exposure category both before and after adjustment. The authors concluded that the 6 observed association between formaldehyde exposure and nasopharyngeal cancer in this 7 cohort could be attributable to external employment in silver and other metal work rather 8 than to formaldehyde itself.

9 3.2.2 National Institute for Occupational Safety and Health (NIOSH) cohort: garment 10 industry

11 Study population and follow-up. Stayner and colleagues led a NIOSH-sponsored 12 investigation of formaldehyde exposure and cancer among garment workers at four shirt-13 manufacturing facilities located in Pennsylvania and Georgia where formaldehyde was 14 used to treat fabrics. The cohort was assembled to conduct a proportionate mortality 15 study (Stayner et al. 1985) and a retrospective cohort mortality study (Stayner et al. 16 1988). Vital status and death certificates were ascertained through December 31, 1982, 17 and cause of death was coded by a trained nosologist (Stayner et al. 1988). Workers 18 enrolled in death benefit insurance were included in the proportionate mortality study if 19 they met certain eligibility requirements, including having worked at least six months at 20 an exposed facility; 256 deaths were included in the proportionate mortality study.

21 Follow-up for vital status was later updated through December 31, 1998 (Pinkerton *et al.* 

22 2004). However, work histories were not updated and were truncated for approximately

23 11% of subjects. Eligible workers for the updated retrospective cohort study (N = 11,039;

24 82% female, 76% white) must have served as production workers for at least three

25 months at one of three facilities between the time formaldehyde was first introduced into

the facility (1955 or 1959, depending on the facility) and December 1977. Of 2,206 total

27 deaths observed in the updated retrospective cohort, 608 deaths were due to cancer

28 (Pinkerton *et al.* 2004).

*Exposure assessment*. Company personnel records were used to obtain information about
 demographics and occupational history for each worker. When available, union records

1 and Internal Revenue Service files were used to verify plant records. Virtually all 2 production workers in any facility were considered consistently exposed to formaldehyde 3 over the workshift. The median 8-hour TWA concentration of formaldehyde obtained 4 during air monitoring across all departments at three plants in 1981 and 1984 ranged from 5 0.09 to 0.20 ppm (mean = 0.15 ppm), and levels did not vary appreciably between 6 facilities. Previous exposures were assumed to be higher at every facility since 7 improvements in the resins have greatly reduced the amount of free formaldehyde 8 contained in the fabrics; formaldehyde levels at other garment factories in the 1970's and 9 earlier were estimated to be as high as 10 ppm (Stayner *et al.* 1988). The authors noted 10 that workers were not thought to be exposed to any other potentially carcinogenic agents 11 at the work site.

12 *Statistical methods*. Standardized mortality ratios using U.S. and state rates were

13 stratified by duration of exposure, time since first exposure, and year of first exposure.

14 Poisson regression was used to estimate age-adjusted rate ratios by exposure duration for

15 selected cancer sites including the upper respiratory tract, leukemia, and brain.

16 Proportionate mortality ratios (PMRs) were estimated based on U.S. rates (adjusted for

17 sex, race, age, and calendar time), and further stratified by duration of exposure, latency,

18 and facility. Proportionate cancer mortality ratios (PCMR) were also calculated to

19 address the potential for healthy worker bias.

20 *Results*. Results of the earlier proportionate cancer mortality analysis (Stayner *et al.* 

21 1985) showed a statistically significant excess of deaths from oral cavity (PCMR = 6.82,

22 90% CI = 1.85 to 17.58, 3 deaths) and lymphohematopoietic cancers excluding leukemia

23 (PCMR = 3.42, 90% CI = 1.17 to 7.82, 4 deaths). Other excess cancer mortalities

24 (PCMRs > 1.0) were noted including biliary passages and liver (PCMR = 2.74, 90% CI =

25 0.94 to 6.27, 4 deaths), unspecified liver (PCMR = 3.70, 90% CI = 0.66 to 11.66, 2

- deaths), skin (PCMR = 1.50, 90% CI = 0.78 to 2.44, 2 deaths), and pancreas (PCMR = 1.50, 90% CI = 0.78 to 2.44, 2 deaths), and pancreas (PCMR = 1.50, 90% CI = 0.78 to 2.44, 2 deaths), and pancreas (PCMR = 1.50, 90% CI = 0.78 to 2.44, 2 deaths), and pancreas (PCMR = 1.50, 90% CI = 0.78 to 2.44, 2 deaths), and pancreas (PCMR = 1.50, 90% CI = 0.78 to 2.44, 2 deaths), and pancreas (PCMR = 1.50, 90% CI = 0.78 to 2.44, 2 deaths), and pancreas (PCMR = 1.50, 90% CI = 0.78 to 2.44, 2 deaths), and pancreas (PCMR = 1.50, 90% CI = 0.78 to 2.44, 2 deaths), and pancreas (PCMR = 1.50, 90% CI = 0.78 to 2.44, 2 deaths), and pancreas (PCMR = 1.50, 90% CI = 0.78 to 2.44, 2 deaths), and pancreas (PCMR = 1.50, 90% CI = 0.78 to 2.44, 2 deaths), and pancreas (PCMR = 1.50, 90% CI = 0.78 to 2.44, 2 deaths), and pancreas (PCMR = 1.50, 90% CI = 0.78 to 2.44, 2 deaths), and pancreas (PCMR = 1.50, 90% CI = 0.78 to 2.44, 2 deaths), 90\%
- 27 1.07, 90% CI = 0.37 to 2.46, 4 deaths). In the updated retrospective cohort analysis
- 28 (Pinkerton et al. 2004), a statistically significant deficit in mortality from all cancers was
- observed (SMR = 0.89, 95% CI = 0.82 to 0.97, 608 deaths). Elevated SMRs were
- 30 observed for cancer of the oral cavity (SMR = 1.33, 95% CI = 0.36 to 3.41, 4 deaths),

1	leukemia (SMR = $1.09$ , $95\%$ CI = $0.70$ to $1.62$ , 24 deaths), and certain other tumor sites
2	that had imprecise estimates. [The magnitude of the risk estimates for the latter two
3	cancer sites was much lower than the PCMRs.] Further analysis showed that the largest
4	excess in leukemia was among myeloid leukemia (SMR = 1.44, 95% CI = $0.80$ to 2.37,
5	15 deaths), which was greatest among workers exposed presumably to higher levels of
6	formaldehyde in the earliest years of follow-up (before 1963) (SMR = $1.61, 95\%$ CI not
7	reported), with at least 10 years of exposure (SMR = $2.19$ , lower bound of 95% CI value
8	less than 1), and exposed at 20 years diagnosis (SMR = $1.91$ ; lower bound of 95% CI
9	greater than 1). Among workers with at least 10 years exposure and 20 or more years
10	since first exposure, multiple cause mortality from myeloid leukemia was significantly
11	elevated (SMR = 2.55; 95% CI = 1.10 to 5.03; 8 deaths). No deaths from cancers of the
12	nasopharynx (0.96 expected) or nose (0.16 expected) were observed in this cohort.

### 13 3.2.3 British Chemical Workers Study

14 Study population and follow-up. Acheson et al. (1984) assembled a large industry-based 15 cohort of approximately 14,000 male workers employed after 1937 at one of six factories 16 in the British chemical and plastics industry where formaldehyde had been manufactured 17 or used. The cohort was updated by Gardner et al. (1993). More recently, Coggon et al. 18 (2003) reported on an updated analysis of this cohort (which subsumed findings by 19 Gardner *et al.*), extending the original cohort with 11 additional years of follow-up. 20 Workers were followed for mortality and cancer incidence through December 31, 2000 21 using the National Health Service Central Register and National Insurance records.

22 *Exposure assessment.* Occupational histories extracted from employment records were 23 used to classify formaldehyde exposure for each job into five categories (background, 24 low, moderate, high, or unknown). Exposure measurements taken after 1970 as well as 25 workers' recall of irritant symptoms were used to estimate exposure levels for each 26 exposure category. According to Gardner et al. (1993), a total of 3,872 (27.6%) workers 27 were exposed to background levels of formaldehyde corresponding to time-weighted 28 average concentrations of less than 0.1 ppm; 3,815 (27.2%) were classified in the low 29 exposure category (0.1 to 0.5 ppm); 1362 (9.7%) in the moderate exposure category (0.6 30 to 2.0 ppm), and 3993 (28.5%) in the high exposure category (greater than 2.0 ppm). Job-

1 exposure matrices were constructed for each factory. Within each factory, each job was 2 assigned to the same exposure category for all time periods; however, jobs were not 3 necessarily assigned to the same exposure category across factories. Workers were 4 individually classified as having no, low, moderate, high or unknown exposure. For 5 workers with more than one job, their exposure classification was based on the job with 6 the highest exposure. In one factory, no worker was classified as highly exposed; the 7 portion of highly exposed workers in the other five factories ranged from 3% to 7%. Of 8 14,014 workers, 13,865 (99%) were successfully traced through the follow-up period: 9 5,185 (37%) had died (99% with a known cause of death), and 859 (6%) were lost to 10 follow-up.

11 Statistical methods. Person-year analysis was used to calculate SMRs; expected numbers 12 of deaths were obtained from national rates for England and Wales in 5-year age strata 13 for 5-year calendar periods. Adjustments for local geographic variations in mortality were 14 made by multiplying the expected numbers of deaths from national rates by the SMRs for 15 the localities in which each factory was located. [This method of adjustment may 16 underestimate the risk if rates are higher among workers, and these workers live in the 17 areas surrounding the factories.] Exposure-response trends were evaluated using Poisson 18 regression.

19 Results. (Coggon et al. 2003 update). Mortality from all cancers was somewhat elevated 20 in the cohort (SMR = 1.10, 95% CI = 1.04 to 1.16, 1.511 deaths), especially among 21 workers ever classified as highly exposed to formaldehyde (SMR = 1.31, 95% CI = 1.2122 to 1.42, 621 deaths). Statistically significant increases in the number of deaths from 23 stomach (SMR = 1.31, 95% CI = 1.11 to 1.54, 150 deaths) and lung cancer (SMR = 1.22, 24 95% CI = 1.12 to 1.32, 594 deaths) were observed among all workers. Standardized 25 mortality ratios were higher among workers with high exposure (SMR for stomach = 26 1.53, 95% CI = 1.17 to 1.95, 63 deaths; and SMR for lung = 1.58, 95% CI = 1.40 to 1.78, 27 272 deaths). A positive trend was noted for lung cancer by increasing exposure level 28  $(P_{\text{trend}} < 0.001)$ , though the trend was no longer statistically significant when adjusted for 29 geographic location. No exposure-response relationships by years of employment in 30 high-exposure jobs or years since first employment in a high-exposure job were

1 observed. However, lung cancer mortality was highest among workers who were highly

2 exposed before 1965 (SMR = 1.61, 95% CI = 1.41 to 1.82, 243 deaths); the authors noted

- 3 that during this time period, occupational exposures to formaldehyde would have been
- 4 higher.

5 Excess cancer mortality at several other tumor sites was also observed among highly

- 6 exposed workers, though estimates were not statistically significant. These tumor sites
- 7 included: lip (SMR = 5.62, 95% CI = 0.14 to 31.30, 1 death), tongue (SMR = 1.91, 95%
- 8 CI = 0.39 to 5.68, 3 deaths), mouth (SMR = 1.32, 95% CI = 0.16 to 4.75, 2 deaths),
- 9 pharynx (SMR = 1.91, 95% CI = 0.70 to 4.17, 6 deaths), esophagus (SMR = 1.28, 95%
- 10 CI = 0.81 to 1.92, 23 deaths), rectum (SMR = 1.30, 95% CI = 0.93 to 1.77, 40 deaths),
- 11 liver (SMR = 1.26, 95% CI = 0.82 to 1.84, 26 deaths), larynx (SMR = 1.56, 95% CI =
- 12 0.63 to 3.22, 7 deaths), bone (SMR = 3.38, 95% CI = 0.92 to 8.65, 4 deaths), genital
- 13 excluding breast, testis, and prostate (SMR = 1.42, 95% CI = 0.04 to 7.90, 1 death),
- 14 bladder (SMR = 1.25, 95% CI = 0.79 to 1.88, 23 deaths), kidney (SMR = 1.37, 95% CI =
- 15 0.73 to 2.35, 13 deaths), and multiple myeloma (SMR = 1.18, 95% CI = 0.48 to 2.44, 8
- 16 deaths). No deaths from cancer of the nose or nasal sinuses were observed among men
- 17 with high exposure (0.8 deaths expected), and two deaths were reported in the entire
- 18 cohort (2.3 expected).
- 19 3.2.4 Studies of fiberglass workers
- In this section, two studies of workers in the fiberglass industry are reviewed. Workers in this industry may be exposed to formaldehyde in addition to respirable fibers during the fiberglass manufacturing process. Evaluation of the association between formaldehyde exposure and cancer outcomes was not a primary objective of either study. Therefore, the description of the study methods and results are limited to formaldehyde-related analyses only.
- 3.2.4.1 United States: Nested case-control study of respiratory cancer in a historical
   cohort of 10 fiberglass manufacturing plants
- The following analyses draw from a large historical cohort study established in 1975 of production and maintenance workers from some of the largest and oldest fiberglass and
- 30 rock/slag wool manufacturing plants in the United States. Marsh *et al.* (2001) updated

5 [Note that the primary focus of these studies was the relationship between glass wool

6 exposure and cancer mortality, and specifically of respiratory (lung and laryngeal)

7 cancers.]

1

2

3

4

8 Study population and follow-up. Marsh et al. (2001) led an effort to expand this historical 9 cohort to capture female workers, workers employed after the original 1963 cohort end 10 date, and workers from additional manufacturing sites. The expanded cohort included 11 32,110 production or maintenance workers (84% white, 82% female) employed for at 12 least one year between 1945 and 1978 in any of the 10 facilities. Vital status was 13 ascertained through December 31, 1992, and the cause of death was determined for 14 nearly all deceased workers (98.8%) using the National Death Index or death certificates. 15 Using this updated cohort, Marsh et al. (2001) conducted a nested case-control analysis 16 to investigate occupational exposures at the fiberglass manufacturing plants and 17 respiratory system cancers (lung and larynx) among male workers. Cases were defined as 18 workers who died from respiratory system cancer between 1970 and 1992; 96% of cases 19 were diagnosed with cancer of the bronchus, trachea, or lung. Controls were eligible if 20 they were at risk during 1970 to 1992 as well as alive and at risk at the age when the case 21 died. Cases were matched to one control by date of birth (within one year). Smoking 22 information was collected as ever/never having used any form of tobacco via telephone 23 interview with the worker or proxy; the response rate was 88% for 716 eligible cases and 24 80% for 713 controls.

*Exposure assessment.* Potential exposures to known or suspected carcinogens, including
formaldehyde, were estimated from plant start-up until closing or the end of the study
period (Quinn *et al.* 2001). Exposure data developed by integrating industrial hygiene
data and epidemiologic methods were combined with worker histories to estimate
exposures over time for all unique production areas. A job-exposure matrix was used to
produce job location-weighted exposure measures and three summary exposure metrics:

duration, cumulative exposure, and average intensity. Exposure to formaldehyde was the
second most prevalent exposure (22.4% of total person-years) after respirable glass wool
or continuous glass filament fibers (28.5% of total person-years) among workers. The
median average intensity of exposure to formaldehyde was 0.066 ppm for all plants
(range = 0.030 to 0.130); the median cumulative exposure was 0.173 ppm (range = 0.063
to 0.469).

7 *Statistical methods and results*. Complete data were available for 502 of 713 matched

8 pairs, and unmatched cases and controls were combined with the matched set nearest in

9 age to form 516 matched pairs (631 cases and 570 controls) available for analysis.

10 Conditional logistic regression was used to estimate RRs adjusted for smoking. Marsh et

11 *al.* found that compared with unexposed workers, exposure to formaldehyde was

12 associated with a statistically significant increase in respiratory system cancer (RR =

13 1.92, 95% CI = 1.25 to 2.94, 591 exposed deaths, global test P value = 0.003)) which

14 remained after adjustment for estimated smoking (RR = 1.61, 95% CI = 1.02 to 2.57,

15 global test P value = 0.04). However, tests for trend by exposure duration, cumulative

16 exposure, and average intensity of exposure were not statistically significant.

17 Related analyses. Youk et al. (2001) analyzed the Marsh et al. nested case-control study 18 using exposure weighting as an alternative form of exposure characterization to explore a 19 possible exposure-response relationship between respiratory system cancer and 20 formaldehyde. Nine different configurations of exposure lag and window periods were 21 considered. The RR for exposed workers was 1.62 (95% CI = 1.04 to 2.54, 588 exposed)22 cases) with 5-year lag and 1.46 (95% CI = 0.96 to 2.23, 581 exposed cases), with 10-year 23 lag. Estimates from other combinations of lag and window periods were otherwise closer 24 to the null compared with the unweighted estimate (OR = 1.61, 95% CI = 1.02 to 2.56) 25 noted by Marsh et al. (2001). No clear trends with cumulative exposure or average 26 intensity of exposure were observed.

27 Stone *et al.* (2001) also analyzed data from the nested case-control study by further

28 adjusting conditional logistic regression models for exposure to respirable particles in

addition to smoking, and by considering exposure to formaldehyde as a continuous

1 quantitative term in piecewise linear functions (i.e., linear splines) with knots placed at 2 the deciles of the distribution of formaldehyde exposure among cases. Application of the 3 linear splines allowed for multiple exposure-response functional forms to be evaluated. 4 Cumulative exposure to formaldehyde was not significantly associated with an increased 5 risk of respiratory system cancer in any of the models. A positive association was 6 observed between relatively high average exposure intensity and respiratory system 7 cancer risk; the authors noted, however, that the dramatic increase in risk was only 8 predicted for the small number of workers with average exposure intensity at levels 9 above 0.4 ppm. [Estimated exposure to formaldehyde in this cohort of fiberglass 10 production workers was considerably below the current OSHA permissible exposure 11 limit of 0.75 ppm based on an 8-hour time-weighted average.] 12 Stone *et al.* (2004) performed an analysis of respiratory system cancer among the 4,008

13 female fiberglass workers included in the updated cohort of fiberglass workers followed 14 until 1992 (Marsh et al. 2001). [Previous analyses were restricted to male workers.] Fifty-15 three deaths due to respiratory cancer were observed. Estimated relative risks were 16 calculated for a 1 ppm-year increase in cumulative formaldehyde exposure score using 17 multiplicative models fit to the internal cohort cancer rates. Estimated RRs ranged from 18 1.10 to 1.21 depending on adjustment factors (e.g., fiberglass production group, year of 19 hire, duration of employment, or time since first employment.) The authors noted that 20 very few women had a cumulative exposure score greater than 3 ppm-years in this study.

3.2.4.2 South Carolina: Nested case-control study in a historical cohort of one fiberglass
 manufacturing plant

23 Study subjects and follow-up. Chiazze et al. (1997) conducted a nested case-control study 24 evaluating lung cancer mortality among continuous filament fiberglass manufacturing 25 workers at an Owens Corning facility in Anderson, South Carolina. [This plant was not 26 included among those studied by Marsh et al. (2001).] The cohort from which the 27 subjects were selected included 4,631 current and former employees (74% male; 87% 28 white) who had worked for at least one year between 1951 and 1991. Follow-up for vital 29 status was completed through 1991 (96% complete), and cause of death was obtained 30 from death certificates (96% complete). Cases (N = 47) included white male members of

the cohort for whom lung cancer was the underlying cause of death; controls (N = 122)
included any white male non-case cohort member and were matched to cases (case to
control ratio = 1:2) on year of birth (within 2 years) and survival to end of follow-up or
death (within 2 years).

5 *Exposure assessment.* Exposure to occupational substances including formaldehyde was 6 estimated by an exposure assessment committee composed of former and current 7 employees knowledgeable in industrial hygiene and plant processes (Chiazze et al. 1993). 8 For each process, one of four ranges of estimated potential exposure for each substance 9 was assigned based on 8-hour time weighted averages. Cumulative exposure was then 10 estimated for each employee based on the number of days spent performing each process; 11 cumulative exposure days for formaldehyde ranged from none to 2,585 days (only one 12 case and three controls had cumulative exposure greater than 1,000 days). In addition, a 13 telephone interview was used to obtain demographic information, lifetime residence 14 history, lifetime occupational history, smoking and alcohol use, and medical history.

15 Statistical methods and results. Conditional logistic regression was applied to estimate

16 the association between formaldehyde and lung cancer death, adjusted for smoking

17 (adjusted models used information from 33 cases and 82 controls who were smokers).

18 Compared to 11 workers with no exposure to formaldehyde, the unadjusted ORs for those

19 with 0.25 to 99.99 and 100 to 999 cumulative days of exposure were 0.94 (95% CI = 0.38

to 2.36, 14 cases) and 1.27 (95% CI = 0.50 to 3.21, 15 cases), respectively; the respective

estimates among smokers only were 0.92 (95% CI = 0.29 to 2.88, 10 cases) and 1.72

22 (95% CI = 0.17 to 25.5, 11 cases). Only one case (a smoker) was exposed for more than

23 1,000 days (OR = 2.07).

### 24 3.2.5 Studies of woodworking and related industries

25 In this section, the findings from smaller case-control and cohort studies of woodworking

and related industries are reviewed, including a nested case-control study of Finnish

- 27 workers (Partanen et al. 1990, Partanen et al. 1993, Partanen et al. 1985); and a cohort
- 28 (and nested case-control study) of workers from the entire United States as well as
- 29 American territories (Stellman et al. 1998). (See Section 3.3.1 for a discussion of case-
- 30 control studies in this industry.) Workers in these industries are commonly exposed to

wood dust, which is a known risk factor for sinonasal cancer and nasopharyngeal cancer.
This review will focus on study findings for formaldehyde exposure only, though other
occupational exposures such as wood dust were also evaluated. Industries related to
woodworking that were examined in these studies included sawmills, particleboard and
plywood manufacture, construction carpentry, and formaldehyde adhesive production for
furniture.

3.2.5.1 Finland: Nested case-control studies in a historical cohort of woodworkers from
 various industries

9 Partanen et al. (1985) assembled a retrospective cohort of 3,805 male workers at 19 10 particleboard, plywood, and formaldehyde glue factories and sawmills in Finland. This 11 cohort was later expanded (N = 7,303) with additional years of follow-up and additional 12 factories to re-evaluate the association between formaldehyde exposure, respiratory 13 cancer (Partanen et al. 1990), and lymphohematopoietic malignancies (Partanen et al. 14 1993) in a nested case-control study. Findings from the updated cohort subsumed the 15 3,805 workers included in the original analysis; therefore, this review will focus only on 16 the most recent findings (Partanen et al. 1990, 1993).

17 Study population. The Finnish woodworker cohort was expanded to include 7,307 18 workers from 35 Finnish factories employed for at least one year between January 1944 19 and December 1965 in various woodworking facilities. Approximately 9% of cohort 20 members worked at particleboard plants, 24% at plywood plants, 12% at construction 21 carpentry plants, 20% at furniture manufacturing plants, 35% at sawmills, and less than 22 1% at a glue manufacturing plant (Partanen et al. 1990). Cohort members were followed 23 for vital status from January 1957 to December 1982. In this study, respiratory cancer 24 was defined by the authors as primary malignant neoplasms of sites with which inhaled 25 formaldehyde was thought to come into direct epithelial contact, including: oral cavity, 26 pharynx, nasal and sinus cavities, larynx, lung, and trachea. Cases of respiratory cancer 27 (N = 136) and malignant lymphomas and leukemias (N = 24) were ascertained using the 28 Finnish Cancer Registry. For analyses of respiratory cancer, three controls were 29 randomly selected from the cohort and matched to each case by year of birth (N = 408).

1 For analyses of lymphohematopoietic malignancies, between one and eight non-cancer 2 controls (N = 152) were matched to each case by year of birth and vital status in 1983. 3 *Exposure assessment.* Job-exposure matrices were constructed by industrial hygienists for 4 each factory using factory records that included information on exposures, ventilation, 5 work procedures, and actual air quality monitoring data (Kauppinen and Partanen 1988). 6 The job-exposure matrices were linked with worker histories using factory registers, 7 interviews with factory personnel, and questionnaires conducted with cases, controls, or 8 their next of kin (control histories were obtained from company records only). For each 9 of the 73 uniquely classified jobs, exposure to formaldehyde and several other concurrent 10 agents was estimated by cumulative dose and level: unexposed, low (0.1 to 1 ppm-11 months), moderate (1 to 2 ppm-months), and heavy (> 2 ppm-months). Both exposure to 12 formaldehyde fumes and formaldehyde attached to wood dust was considered. Exposure 13 was also categorized dichotomously (ever/never) and lagged by 10 years to account for 14 latency. Workers were considered ever exposed to formaldehyde if their estimated

15 cumulative exposure reached 3 ppm-months.

16 Results for respiratory cancers (Partanen et al. 1990). Odds ratios and 90% CIs were 17 estimated using conditional logistic regression and, in most cases, adjusted for vital status 18 and smoking (< 35 years vs.  $\geq$  35 years). Comparing workers with at least 3 ppm-months 19 of exposure to formaldehyde with workers with less than 3 ppm-months, the OR for all 20 respiratory cancers combined was 1.11 (90% CI = 0.40 to 3.11, 11 exposed cases, 21 adjusted for vital status and smoking) with no latency period, and 1.39 (90% CI = 0.40 to 22 4.10, 9 exposed cases, adjusted for vital status and smoking) with a minimum latency 23 period of 10 years. Corresponding estimates were lower for lung cancer (OR = 0.69, 90%24 CI = 0.21 to 2.24, 9 cases, no latency, adjusted for vital status and smoking; and OR =25 0.89,90% CI = 0.26 to 3.00, 7 cases, 10-year latency, adjusted for vital status and 26 smoking), and higher for combined upper respiratory cancers only (OR = 2.38, 90% CI = 27 0.43 to 13.2, 2 cases, no latency, adjusted for vital status only, and OR = 2.40, 90% CI = 28 0.31 to 18.6, 2 cases, 10 year latency, adjusted for vital status only). Exposure to dust-29 borne formaldehyde (yes or no) was also estimated; ORs ranged from 1.33 to 1.42, 30 depending on the latency period, but none was statistically significant. No evidence of an

1 association was observed between peak exposure to formaldehyde and respiratory cancer, 2 nor was any evidence observed of an exposure-response relationship for any exposure 3 indicator including cumulative dose, duration of exposure to peak levels, and duration of 4 exposure to dust-borne formaldehyde. [The numbers of respiratory cancers was small and 5 only permitted analyses of all respiratory cancers combined in exposure-response 6 analyses. Adjustment for smoking substantially reduced the sample size and consequently 7 reduced statistical power for estimation of effects, because smoking history was unknown 8 for approximately 35% of workers in this study. Further, estimates were not adjusted for 9 wood dust or phenol exposure, both factors that the authors noted were correlated with 10 formaldehyde exposure in this study population.]

11 Results for lymphohematopoietic malignancies (Partanen et al. 1993). Odds ratios and

12 95% CIs were estimated using conditional logistic regression. For the

13 lymphohematopoietic cancers combined, the OR associated with at least 3 ppm-months

14 of formaldehyde was 2.49 (95% CI = 0.81 to 7.59, 7 exposed cases), which did not

15 change markedly after controlling for exposure to wood dust or solvents. Corresponding

16 (unadjusted ) ORs for specific lymphohematopoietic cancers were 1.40 (95% CI = 0.25 to)

17 7.91, 2 exposed cases) for leukemia, and 4.24 (95% CI = 0.68 to 26.6, 4 exposed cases)

18 for non-Hodgkin's lymphoma. An OR for Hodgkin's lymphoma alone could not be

19 estimated because only one case was considered exposed to formaldehyde. The OR for

20 all lymphomas combined (Hodgkin's and non-Hodgkin's lymphomas) was 4.02 (95% CI

21 = 0.87 to 18.6, 5 exposed cases). The authors noted that more sensitive exposure

22 assessment among cases than controls could have biased the observed effect estimates

away from the null. [Effect estimates in this study are imprecise since ORs were based on

24 a very small number of exposed cases.]

# 3.2.5.2 United States: American Cancer Society Cancer Prevention Study and nested case-control study

- 27 Stellman *et al.* (1998) studied the association between mortality and occupational
- 28 exposure to wood dust in the American Cancer Society's population-based Cancer
- 29 Prevention Study. The cohort consists of over half a million males from all 50 states,
- 30 Washington, D.C., and Puerto Rico enrolled in 1982 and who completed questionnaires

1 on demographic and lifestyle characteristics including smoking, medical history, and 2 occupational history. Exposure to 12 occupational substances including formaldehyde 3 was self-indicated on a check-list. The analysis included 11,541 woodworkers, of whom 4 305 reported exposure to both formaldehyde and wood dust, and 387 reported exposure 5 to formaldehyde only. Site-specific cancer mortality information was obtained from death 6 certificates during six years of follow-up (September 1982 to August 1988). Incidence 7 density ratios adjusted by age and smoking status were calculated for subjects reporting 8 formaldehyde exposure employed in any occupation, and for subjects reported 9 formaldehyde exposure employed in a wood-related occupation. The reference group for 10 all estimates consisted of subjects who did not report either employment in a wood-11 related occupation or regular exposure to wood dust. Woodworkers who reported regular 12 exposure to formaldehyde had a statistically significant increase in lung cancer mortality 13 (RR = 2.63, 95% CI = 1.25 to 5.51, 7 exposed cases) and leukemia (RR = 5.79, 95% CI = 14 1.44 to 23.25, 2 exposed cases). Effect estimates were elevated for rectal cancer (RR =15 5.77, 95% CI = 0.81 to 41.22) and non-Hodgkin's lymphoma (RR = 2.88, 95% CI = 0.40 16 to 20.50), though both estimates were based on only one exposed case and were not 17 statistically significant. Among non-woodworkers exposed to formaldehyde, increased 18 risk of cancer mortality was observed for stomach cancer (RR = 1.63, 95% CI = 0.94 to 19 2.86, 11 exposed cases) and all lymphohematopoietic cancers combined (RR = 1.22, 95%) 20 CI = 0.84 to 1.77, 28 exposed cases). [Results for cancers of the paranasal sinuses and 21 nasal cavity were not presented.]

22 Nested case control study within the American Cancer Society Cancer Prevention Study: 23 A population-based nested case-control study of 282 deaths from multiple myeloma 24 observed in the second stage of the American Cancer Society's Cancer Prevention 25 prospective cohort study and matched with up to 4 within-cohort controls was conducted 26 by Boffetta et al. (1989). The association between multiple myeloma, occupational 27 groups and selected exposures was examined, based on questionnaires completed by 28 enrollees and assignment of exposure status by the investigators. Using conditional 29 logistic regression, a statistically nonsignificant association between multiple myeloma 30 and formaldehyde exposure was observed (OR = 1.8, 95% CI = 0.6 to 5.7, 4 cases). [The 31 likelihood of misclassification of exposure in this study was high, however, and subjects

1 assigned to the high exposure group had lower OR than those in the low exposure group.

2 The power to detect effects of given agents in this study was also limited.]

 3.2.6 Miscellaneous studies: abrasive material manufacturing, Iron foundry, mixed industry and chrome leather tannery workers
 5 In this section, four historical studies examining the association between formaldehyde

6 exposure and cancer among abrasive material manufacturing, iron foundry, mixed

7 industry, and chrome leather tannery workers are summarized.

### 8 3.2.6.1 Sweden: Cohort mortality and incidence study of abrasive materials 9 manufacturing workers (Edling et al. 1987a)

10 Study population and methods. 911 workers (211 women) at a plant manufacturing

abrasive materials and employed between 1955 and 1983 for at least five years were

12 enrolled in the study. Workers were traced through the Swedish national death registry

13 (from 1958 to 1983) and the national cancer registry (from 1958 to 1981). Deaths

14 occurring at ages 74 and older were excluded, based on less reliable diagnostic validity.

15 Age-, sex- and calendar year-stratified expected rates were calculated using the person-

16 year method based on national data.

17 *Exposure assessment*. The plant manufactured grinding wheels from aluminum oxide and

18 silicon carbide as abrasives bound with clay or phenol formaldehyde resins. Industrial

19 hygiene measurements were available since the 1970s; during the manufacture of

20 formaldehyde resins, exposure to formaldehyde ranged from 0.1 to 1.0 mg/m<sup>3</sup>. According

21 to the authors, 59 workers had heavy intermittent exposure to peaks of 20 to  $30 \text{ mg/m}^3$  of

22 formaldehyde during the manufacture of abrasive belts. No exposure assessments were

23 conducted for individual workers.

24 *Results.* Findings were reported for 506 male "blue collar" workers only. No statistically

25 significant increases in mortality or incidence for all cancers combined

- 26 (observed/expected = 0.93, 95% CI = 0.5 to 1.5, 17 deaths; and observed/expected =
- 27 0.84, 95% CI = 0.5 to 11.3, 24 cases). Elevations in cancer incidence were observed for
- 28 pancreas (obs/exp = 1.8, 95% CI = 0.2 to 6.6, 2 cases), lymphoma (obs/exp = 2.0, 95% CI
- 29 = 0.2 to 7.2, 2 cases) and multiple myeloma (obs/exp = 4.0; 95% CI = 0.5 to 14.4, 2
- 30 cases). One case of nasopharyngeal cancer was observed in a worker with formaldehyde
- 31 exposure of  $< 1.0 \text{ mg/m}^3$  and less than 5 years of employment.

### 1 3.2.6.2 Michigan: Historical cohort of iron foundry workers

2 Mortality among workers at an iron foundry in Michigan was investigated in a 3 retrospective cohort study assembled by Andjelkovich *et al.* (1990). Workers (N = 8,147) 4 were employed at an automotive gray iron foundry for at least six months between 1950 5 and 1979. During the period of observation from 1950 to 1984, an excess of lung cancer 6 deaths among these workers was observed. Though the authors suspected that the excess 7 could have been in part explained by smoking, other hypotheses related to occupational 8 exposures at the plant were proposed, including exposure to formaldehyde. To further 9 evaluate these hypotheses, the investigators conducted a nested case-control study of lung 10 cancer in the entire cohort (Andjelkovich et al. 1994) as well as a standardized mortality 11 analysis of a subset of the cohort exposed to formaldehyde between 1960 and 1987 12 (Andjelkovich et al. 1995). A summary of the major methods and findings from these 13 two studies follows.

14 *Nested case-control study*. To investigate the potential association between lung cancer 15 and relevant exposures at the iron foundry, including silica and formaldehyde, a nested 16 case-control study was conducted with additional years of follow-up through December 17 1989 (Andjelkovich *et al.* 1994). Cases (N = 220, 51% white) were defined as primary 18 lung cancer deaths among men in the cohort between January 1, 1950 and December 31, 19 1989. For each case, 10 controls matched on race and attained age were selected from the 20 cohort using incidence density sampling (52% of controls were alive at the end of the 21 study period). Smoking information was obtained by questionnaire or records (including 22 plant records and death certificates) for 76% of cases and 69% of a random sample of 23 controls. Detailed work histories within the foundry were used to identify 107 unique 24 occupational titles, which were then characterized by an industrial hygienist according to 25 exposure to silica (high, medium, low) and formaldehyde (high, medium, low, none). For 26 analyses, exposure to formaldehyde was dichotomized (ever/never) because only 25% of 27 workers were considered ever exposed to formaldehyde (57 cases and 538 controls).

28 Conditional logistic regression was applied to estimate the effect of exposure to

- 29 formaldehyde on lung cancer mortality adjusting for smoking, birth cohort (< 1915  $vs. \ge$
- 30 1915), and silica exposure (quartiles). Using the subset of controls for which collection of

smoking information was attempted, the OR for exposure to formaldehyde was 1.31
(95% CI = 0.83 to 2.07, number of cases not specified). Effect estimates consistently
decreased in magnitude with increasing lag periods (10, 15, and 20 years) to 0.84 (95%
CI = 0.44 to 1.60) with a 20-year lag. Effect estimates were slightly higher and more
precise when all controls were included, though the same decrease in risk was observed
with increasing lag periods. No evidence was observed of an interaction between
smoking and formaldehyde.

8 Cohort mortality sub-analysis. A subsequent analysis examined mortality among a subset 9 of foundry workers (N = 3.929, 67% white) exposed to formaldehyde for 6 months or 10 more during core making operations between 1960 and 1987 (Andjelkovich et al. 1995). 11 An internal referent group included a sample of workers (N = 2,032) from the original 12 cohort who were unexposed to formaldehyde during the same time period. Cumulative 13 exposure to formaldehyde was estimated for each worker by an industrial hygienist based 14 on job-specific exposure levels (low = 0.05 ppm; medium = 0.55 ppm; and high = 1.515 ppm) and duration of exposure. Smoking information was obtained by questionnaire or 16 records (including plant records and death certificates) for 65% of exposed workers and 17 55% of the unexposed referent group.

18 Mortality among the exposed workers through December 1989 was compared with 19 mortality among the U.S. population; SMRs adjusted for sex, race, age, and calendar 20 period were obtained using the person-years method. To address the potential for healthy 21 worker bias, mortality among all the workers was compared with that of an occupational 22 referent population assembled by the NCI and NIOSH, using Poisson regression adjusted 23 for race, smoking, and silica exposure. Statistically non-significant excess mortality was 24 observed among the exposed workers for cancers of the oral cavity and pharynx (SMR =25 1.31; 95% CI = 0.48 to 2.86; 127 deaths), esophagus (SMR = 1.07, 95% CI = 0.39 to 26 2.33, 6 deaths), stomach (SMR = 1.64, 95% CI = 0.82 to 2.94, 11 deaths), large intestine 27 (SMR = 1.03, 95% CI = 0.49 to 1.90, 10 deaths), rectum (SMR = 1.17, 95% CI = 0.23 to 1.90, 10 deaths)28 3.41, 3 deaths), trachea, bronchus, and lung (SMR = 1.20, 95% CI = 0.89 to 1.58, 5129 deaths) and other and unspecified genital organs (SMR = 1.13, 95% CI = 0.23 to 3.31, 330 deaths). SMRs below 1.0 were reported for all other cancer sites, including but not

1 limited to larynx, (2 deaths), and all lymphohematopoietic cancers (7 deaths). Directly 2 adjusted relative risks (comparing exposed workers with unexposed workers) were 3 elevated for laryngeal cancer (RR = 1.50, 95% CI not reported,  $P \ge 0.05$ ) and cancer of 4 the trachea, bronchus, or lung (RR = 1.13, 95% CI not reported,  $P \ge 0.05$ ). The authors 5 report that the majority of SMRs increased when the NCI/NIOSH referent population was 6 applied (data not presented). In the Poisson regression analysis of men for whom 7 smoking status was known, cumulative exposure to formaldehyde (third and fourth 8 quartiles combined vs. unexposed) was not associated with cancers of the lung or oral 9 cavity and pharynx (data for other cancer sites not presented). One death from 10 nasopharyngeal cacner was reported for a man who had no recorded formaldehyde 11 exposure, according to the authors. (Deaths from sinonasal cancers were not presented.) 12 3.2.6.3 Denmark: Proportionate cancer incidence study of mixed industry workers 13 Study population and follow-up. Hansen and Olsen (1995) conducted a standardized 14 proportionate cancer incidence study of workers in Denmark born between 1897 and 15 1964 whose cancer was diagnosed between 1970 and 1984; eligible workers were 16 identified using the national Danish Cancer Registry and then linked with the compulsory 17 Supplementary Pension Fund to obtain employment history (N = 91,182 males). Using 18 the national Danish Product Register, 265 companies in which more than one kilogram of 19 formaldehyde was used or manufactured per employee per year since 1970 were 20 identified.

21 Exposure assessment. Workers considered exposed to formaldehyde were those whose

22 longest work experience since 1964 had started at one of the 265 companies at least 10

23 years prior to diagnosis (N = 2,041, 2.2% of study population). Based on job title,

24 exposed workers were further classified as having low (white-collar workers), high (blue-

25 collar workers), and unknown (no information on job title) exposure.

26 *Statistical methods and results.* Standardized proportionate incidence cancer ratios

27 (SPICR) adjusted for age (5-year strata) and calendar time (per year) were estimated

28 using all Danish workers in the study population as the referent group. (Results for

29 73,423 female workers for whom work history and exposure were concurrently obtained

30 were reported in a separate publication (Hansen and Olsen 1996).) Among the 2041 men,

1	who had worked in companies where formaldehyde was used, a statistically significant
2	excess in incidence was noted for tumors of the colon (SPICR = 1.2, 95% CI = 1.1 to 1.4,
3	166 exposed cases), nasal cavity (SPICR = $2.3$ , $95\%$ CI = $1.3$ to $4.0$ , $13$ cases), and
4	kidney (SPICR = 1.3, 95% CI = 1.0 to 1.6, 60 cases). Statistically non-significant
5	increases in cancer incidence (SPICRs $> 1.0$ ) were also observed among men for the
6	nasopharynx (SPICR = $1.3$ , $95\%$ CI = $0.3$ to $3.2$ , 4 exposed cases), liver (SPICR = $1.2$ ,
7	95% CI = 0.9 to 1.8, 29 exposed cases), rectum (SPICR = 1.1, 95% CI = 0.9 to 1.3, 117
8	cases), melanoma of the skin (SPICR = $1.1$ , 95% CI = $0.8$ to $1.5$ , 39 cases), brain (SPCIR
9	= 1.1, 95% CI = 0.9 to 1.5, 54 cases) and breast (SPICR = 2.2, 95% CI = 0.9 to 4.3, 8
10	exposed cases). Other sites had SPICRs of 1.0 or less. (Among lymphohematopoietic
11	cancers, data were reported only for non-Hodgkin's lymphoma (32 cases), Hodgkin's
12	lymphoma (12 cases) and leukemia (39 cases); no increase in risk was observed. Data
13	were also presented on selected cancers (nasal, colon, lung, breast, kidney, brain and
14	CNS, and leukemia) among workers with estimated exposure to low or high
15	formaldehyde, the latter with or without potential wood dust exposure. No differences by
16	estimated exposure category were observed, with the exception of nasal cavity cancers;
17	among those estimated to be more highly exposed to formaldehyde and unexposed to
18	wood dust (based on job industry and title), the SPICR was $3.0 (95\% \text{ CI} = 1.4 \text{ to } 5.7, 9$
19	cases), compared with 5.0 (95% $CI = 0.5$ to 13.4, 2 cases) for both higher formaldehyde
20	and wood dust exposure and 0.8 (95% $CI = 0.02$ to 4.4, 1 case) for low formaldehyde
21	exposure. Among women, an increase was found for nasal cancer (SPICR = $2.4$ , $95\%$ CI
22	= 0.6 to 6.0. 4 exposed cases).

#### 23 3.2.7 Studies of resin, chemical, and plastics manufacturing workers

In this section, historical cohort studies of workers in the formaldehyde-based resin
(Bertazzi *et al.* 1986), chemical (Bond *et al.* 1986, Ott *et al.* 1989), and plastics (Dell and
Teta 1995) manufacturing industries are reviewed. Bond *et al.* (1986) evaluated lung
cancer specifically, and Ott *et al.* (1989) evaluated lymphohematopoietic malignancies.
[Collectively, the studies reviewed in this section are limited by small numbers of study
participants exposed to formaldehyde. Note also that in these studies formaldehyde was
not the primary occupational exposure of interest. Workers in these cohorts were exposed

to various other agents such as asbestos, styrene, and solvents.] The following review will
 focus on study findings for formaldehyde only.

3.2.7.1 Italy: Historical cohort of formaldehyde-based resin production workers
4 Study population and follow-up. Bertazzi *et al.* (1986) studied mortality among male

5 workers at a resin manufacturing plant in Italy where formaldehyde-based resins 6 including urea- and melamine-formaldehyde resins were primarily produced since 1959. 7 A cohort of workers was assembled including 1,332 men ever employed in the plant for 8 at least 30 days between 1959 and 1980 (Bertazzi et al. 1986). Vital status was originally 9 ascertained as of December 31, 1980 through the local vital statistics offices, and death 10 certificates were obtained for cause of death (follow-up was complete for nearly 99% of 11 the cohort). In a subsequent analysis, vital status was updated through 1986 (Bertazzi et 12 al. 1989); however, the 1989 study was published in Italian and is not reported here).

13 *Exposure assessment*. Work histories for each worker were reconstructed using

14 incomplete plant employment records and interviews with current and retired workers as

15 well as foremen. Work histories were completed for over 80% of the cohort, and each

16 worker was assigned to one of three exposure categories based on their work history: (1)

17 exposed to formaldehyde, (2) exposed to other compounds (including styrene and

18 solvents), and (3) unknown exposure. Air sampling was conducted at the plant in 1974,

19 1978 and 1979; mean levels of formaldehyde ranged from 0.2 to  $3.8 \text{ mg/m}^3$  [0.13 to 2.53

20 ppm]. The authors noted that formaldehyde-based resins were produced in a separate area

21 from other resins, and also that job mobility was low, especially among workers engaged

22 in formaldehyde-based resin production [these factors reduce the potential for exposure

23 misclassification].

24 *Results*. Mortality in the cohort was compared with national and local rates using the

25 person-years method, adjusting for sex, age (5-year strata), and calendar time (5-year

26 intervals). Among workers "definitely" exposed to formaldehyde (5,731 person-years of

27 exposure), excess mortality was observed in the 1986 for cancers of the gastrointestinal

tract (SMR = 1.55 [95% CI not reported for any SMR], 8 observed deaths vs. 5.2

- expected), esophagus + stomach (SMR = 1.33, 4 observed deaths vs. 3 expected and,
- 30 lung (SMR = 1.36, 5 observed deaths vs. 3.7 expected) and lymphohematopoietic cancers

(SMR = 2.73, 3 observed deaths vs. 1.1 expected). [Note that only certain cancer sites
were reported in these studies.]

## 3.2.7.2 Texas: Nested case-control study in a historical cohort of chemical production workers

5 Study population and follow-up. A nested case-control study of workers was conducted to 6 investigate elevated lung cancer mortality rates at a chemical production facility (Dow 7 Chemical) in Texas (Bond et al. 1986). A retrospective cohort was assembled including 8 19,608 male workers hired between 1940 and 1980 and who had worked at the Texas 9 facility for at least one year. Vital status was ascertained for 97% of the cohort; death 10 certificates were obtained for 96% of the 3,444 deceased workers. Cases (N = 308) were 11 defined as former workers who had died before December 1980 and whose death 12 certificate listed cancer of the respiratory system as the underlying or contributing cause 13 of death. Two control series without lung cancer were randomly selected and individually 14 matched by race, year of birth (within 5 years), and year of hire (case to control ratio = 15 1:1). One series included workers alive when the matched case died of lung cancer, and 16 the other series included workers who had died of other causes within five years after the 17 matched case had died.

*Exposure assessment.* For each subject, exposure to 171 chemical and physical agents (yes/no), including formaldehyde, was estimated by an industrial hygienist blinded to case/control status using information from employee work history records about work areas, tasks, agents handled, and duration of employment. Information on potentially confounding variables such as smoking and vitamin A intake was obtained from interviews (82% response rate) conducted with subjects or their next-of-kin.

*Results.* Stratified analyses and conditional logistic regression were used to calculate ORs and 95% CIs. Reported risk estimates for formaldehyde were unadjusted for exposure to other agents and other potential confounders like smoking. The estimated OR between exposure to formaldehyde (9 exposed deaths) and lung cancer mortality was less than 1.0; the negative association remained after accounting for a 15-year latency period (4 exposed deaths). [Eligible controls included participants with cancers suspected to be

- 1 associated with formaldehyde exposure, which may have attenuated observed effect
- 2 estimates.]

## 3.2.7.3 West Virginia: Nested case-control study in a historical cohort of chemical manufacturing workers

*Study population and follow-up.* Ott *et al.* (1989) conducted a nested case-control study
of lymphohematopoietic carcinomas within a cohort of nearly 30,000 male workers

- 7 employed in two chemical manufacturing facilities and a research and development
- 8 center (Union Carbide Corporation). Cases of non-Hodgkin's lymphoma (N = 52),
- 9 multiple myeloma (N = 20), nonlymphocytic leukemia (N = 39), and lymphocytic
- 10 leukemia (N = 18) among workers in the cohort were identified by reviewing both
- 11 underlying and contributory causes of death noted on death certificates from 1940
- 12 through 1978; follow-up was complete for 96% of the cohort. Controls were selected
- 13 from the cohort using group-matched incidence density sampling so that controls were
- 14 first employed in the same decade and survived to at least the same 5-year period as cases
- 15 (case to control ratio = 1:5).
- 16 *Exposure assessment*. Work history information was used to link work areas and
- 17 assignments with records of departmental usage for each substance; a worker was
- 18 considered exposed to formaldehyde (ever/never) if he worked for at least one day with
- 19 the chemical or in a work area specified as exposed.
- 20 *Statistical methods and results.* Unadjusted ORs were obtained using unconditional
- 21 logistic regression. Elevated but statistically non-significant risks were found for non-
- 22 Hodgkin's lymphoma (OR = 2.0, 95% CI not reported, 2 exposed deaths),
- 23 nonlymphocytic leukemia (OR = 2.6, 2 exposed deaths), and lymphocytic leukemia (OR
- 24 = 2.6, 1 exposed death). The OR for multiple myeloma was 1.0 (1 exposed death). [Very
- 25 few workers were exposed to formaldehyde and workers with only one day of exposure
- 26 in their occupational lifetime were considered exposed.]
- 27 3.2.7.4 New Jersey: Historical cohort of plastics manufacturing workers
- 28 *Study population and follow-up.* Cancer mortality among male workers at a plastics
- 29 manufacturing plant (Union Carbide Corporation) in New Jersey was studied by Dell and

Teta (1995). [This plant is not included among those studied by Ott *et al.* (1989).] The
cohort included 5,932 male employees who worked more than six months between
January 1, 1946 and December 31, 1967. Vital status was ascertained through December
31, 1988 (94% complete) using company records, Social Security files, and information
from the National Death Index. Underlying causes of death were obtained from death
certificates (98% complete).

7 Exposure assessment and statistical methods. Exposure to asbestos, polyvinyl chloride,

8 and formaldehyde was assigned (yes/no) based on the major work department for each

9 worker. One hundred eleven (111) workers were assigned exposure to formaldehyde.

10 Mortality in the cohort was evaluated using person-years analysis, with age- and

11 calendar-year-specific mortality rates among white males for the U.S. (1940 to 1989) and

12 New Jersey (1950 to 1989) as the referents.

*Results.* An excess of lung cancer was noted among 57 workers exposed to formaldehyde
during hexamethylenetretamine production (4 observed cases vs. 1.1 expected, no risk
estimate reported). No cases of sinonasal or nasopharyngeal carcinoma were observed.
[As noted by the authors, the power of this study is limited with regard to formaldehyde
because of small sample size. Further, the potential effect of individual exposures cannot

18 be distinguished within each work area.]

19 3.2.8 Studies of health professionals, embalmers, and funeral directors

20 This section covers multiple studies of health professionals (e.g., anatomists,

21 pathologists, and medical lab technicians), embalmers, and funeral directors. These

22 occupations are known to involve exposure to formaldehyde, which is used as a human

tissue preservative (see Section 2.4.6 for more information on exposure levels). This

section is divided into studies of health professionals (Hall et al. 1991, Harrington and

- 25 Oakes 1984, Harrington and Shannon 1975, Jensen and Andersen 1982, Stroup et al.
- 26 1986) and studies of embalmers and funeral directors (Hayes *et al.* 1990, Levine *et al.*
- 27 1984, Walrath and Fraumeni 1983, 1984). One study of pathologists was excluded from
- 28 this review because its primary objective was to examine low-level ionizing radiation
- 29 among pathologists with membership in the Radiation Registry of Physicians (Logue et

al. 1986). A small case-control analysis of lung cancer among Danish physicians (Jensen
 and Andersen, 1982) is reported in Section 3.3.4.

Studies included in this section examined the association between occupational groups assumed to be exposed to formaldehyde and excess mortality from cancer (compared with cancer mortality among internal or external reference populations). None of these studies attempted to quantify or characterize exposure or estimate exposure-response relationships, but they examined cancer outcomes by occupation and occupational characteristics (e.g., duration of employment) only.

#### 9 3.2.8.1 Health professionals

10 Pathologists: United Kingdom. Harrington and Shannon (1975) studied mortality among 11 pathologists and medical laboratory technicians who were members of professional 12 organizations in the United Kingdom. Members of the Royal College of Pathologists and 13 the Pathological Society active at some time between January 1955 and December 1973 14 were enrolled (N = 2,079). Enrolled technicians (N = 12,944) included members of the 15 Council for Professions Supplementary to Medicine active between August 1963 and 16 December 1973. Death certificates were obtained for 97% of deaths among pathologists 17 (N = 156, 10 deaths among women) and all technicians (N = 154, 20 deaths among)18 women). Expected numbers of deaths were calculated using sex-, age- (5-year strata), and 19 calendar time- (5-year intervals) specific death rates from England, Wales, or Scotland. 20 Mortality was less than expected among pathologists and technicians for all causes of 21 death and for all neoplasms. A statistically significant excess mortality from 22 lymphohematopoietic cancers was observed among male pathologists in England (8) 23 observed deaths vs. 3.3 expected, P < 0.05); no increase in leukemia was found. No 24 increase was observed for other individual tumors.

Harrington and Oakes (1984) extended the previous study to include pathologists active
in the professional organizations from January 1974 through December 1980. This study
population included 2,307 male (110 deaths) and 413 female (16 deaths) pathologists;
medical laboratory technicians included in the original cohort (Harrington and Shannon
1975) were excluded from this study. SMRs were only reported for selected tumor sites.
Mortality from all causes and all cancers combined were significantly lower than

1	expected among men, and among women for all causes; the SMR for all cancers for
2	women was slightly elevated (SMR = $1.41$ , 90% CI = $0.66$ to $2.65$ , 7 deaths). In contrast
3	to the 1975 study, deaths from lymphohematopoietic cancers were not elevated in this
4	population: SMRs for male leukemia was 0.90 (90% $CI = 0.05$ to 4.29, 1 death) and for
5	female leukemia the SMR was 9.26 (90% $CI = 0.47$ to 43. 92, 1 death); for other
6	lymphohematopoietic cancers, the SMR was $0.54$ (90% CI = 0.03 to 2.54, 1 male death
7	only). An increase in brain cancer was observed among men (SMR = $3.31$ , $90\%$ CI =
8	1.13 to 7.58, 4 deaths); no cases were observed among women (0.11 expected). A
9	marginal increase in bladder cancer among men was observed (SMR = 1.07, 90% $CI =$
10	0.19 to 3.37, 2 deaths); no increases in lung cancer or gastrointestinal cancers was

11 observed.

12 Hall et al. (1991) further updated this cohort of British pathologists, adding new members 13 of the Pathological Society and extending follow-up to 1987; a total of 3,872 pathologists 14 were included (3,069 men, 803 women) after excluding 640 females from Northern 15 Ireland and Scotland for whom reference rates were unavailable. Sex-specific SMRs 16 adjusted for age (5 year strata) and calendar time (2 year intervals) were calculated based 17 on expected mortality rates from England, Wales, or Scotland (for males only). 18 Compared with national rates, mortality from all causes (SMR men = 0.43, 95% CI = 19 0.37 to 0.50; SMR women = 0.65, 95% CI = 0.38 to 1.03) and also from all cancers was 20 substantially less than expected. No statistically significant excesses were observed for 21 cancer at any site. However, increases in mortality were noted for lymphohematopoietic 22 cancer (SMR = 1.44, 95% CI = 0.69 to 2.65, 10 deaths) and leukemia (SMR = 1.52, 95% 23 CI = 0.41 to 3.89, 4 deaths) among all pathologists in England and Wales, brain cancer 24 (SMR = 2.40, 95% CI = 0.88 to 5.22, 6 deaths) among male pathologists from England 25 and Wales, prostate cancer (SMR = 3.30, 95% CI = 0.39 to 11.80, 2 deaths) among 26 pathologists from Scotland, and breast cancer (SMR = 1.61, 95% CI = 0.44 to 4.11, 427 deaths) among female pathologists from England and Wales. Among all pathologists, 28 non-statistically significant excesses were also observed for liver, Hodgkin's lymphoma 29 and tongue, each based on one death only. [Only nine deaths were observed among 30 Scottish pathologists.]

1 Anatomists: United States. Stroup et al. (1986) conducted a retrospective cohort study of 2 mortality among members of the American Association of Anatomists. Eligible subjects 3 included 2,317 male residents of the United States who joined the professional 4 organization between 1888 and 1969; each subject was followed from date of initial 5 membership through December 1979. Death certificates were obtained and coded by a 6 trained nosologist for underlying and contributing causes of death. Standardized mortality 7 ratios were obtained using 5-year age-specific and 5-year time-specific mortality rates 8 among U.S. white males from 1925 to 1979. A second referent group consisting of 5-year 9 age-specific mortality rates among 19,000 male members of the American Psychiatric 10 Association between 1900 and 1969 was also used to reduce any influence of the 11 "healthy-worker effect." Compared with the general population, this cohort of anatomists 12 experienced less-than-expected numbers of death from all causes (SMR = 0.65, 95% CI = 13 0.60 to 0.70, 738 deaths) and all cancers (SMR = 0.64, 95% CI = 0.53 to 0.76, 118 14 deaths). Despite these overall deficits, a statistically significant excess of brain cancer 15 was observed (SMR = 2.7, 95% CI = 1.3 to 5.0, 10 cases), and SMRs increased in 16 magnitude with duration of membership. Excess mortality was also noted for 17 lymphohematopoietic cancers (SMR = 1.2, 95% CI = 0.7 to 2.0, 18 deaths), including 18 leukemia (SMR = 1.5, 95% CI = 0.7 to 2.7, 10 deaths) and other lymphohematopoietic 19 cancer of other lymphatic tissues (SMR = 2.0, 95% CI = 0.7 to 4.4, 6 deaths). The authors 20 noted that of the 10 leukemia deaths, five were from myeloid leukemia, and the SMR for 21 chronic myeloid leukemia was statistically significantly elevated (SMR = 8.8, 95% CI = 22 1.8 to 25.5, 3 deaths) during the period from 1969 to 1979 when cell type-specific 23 mortality rates were available. Slight increases in cancers of the colon (SMR = 1.1, 95%24 CI = 0.7 to 1.7, 20 deaths) and pancreas (SMR = 1.1, 95% CI = 0.6 to 2.0, 11 deaths) 25 were also observed. Brain cancer was also statistically significantly elevated when 26 compared to the to the referent group of psychiatrists (SMR = 6.0, 95% CI = 2.3 to 15.6); 27 the SMR for leukemia was not elevated in comparison with the referent group of 28 psychiatrists, however (SMR = 0.8, 95% CI = 0.2 to 2.9, 3 deaths).

- 29 3.2.8.2 Embalmers and funeral directors
- 30 Embalmers: New York. Using records obtained from the New York Bureau of Funeral
- 31 Directing and Embalming, Walrath and Fraumeni (1983) assembled a cohort of all
1 embalmers licensed to practice in New York between 1902 and 1980 and known to have 2 died between 1925 and 1980. Death certificates were obtained for 1,263 eligible subjects 3 (75% of cohort), and the underlying cause of death was coded by a trained nosologist. 4 Deaths observed among the embalmers were compared with expected numbers calculated 5 by applying the age-, race-, and calendar-year-specific proportions of deaths for each 6 cause among the U.S. male population to the total number of deaths in the cohort by five-7 year age and calendar periods. Time since first licensure was used to approximate 8 duration of exposure. Results focused on findings from 1,132 white men (10 women and 9 42 men of unknown race were excluded). Among white male embalmers, a statistically 10 nonsignificant increase in PMR for all cancers was observed (PMR = 1.11, 243 observed 11 deaths vs. 218.9 expected). A statistically significant (P < 0.05) excess mortality was 12 observed for cancers of the colon (PMR = 1.43, 29 observed deaths vs. 20.3 expected) 13 and skin (PMR = 2.21, 8 observed deaths vs. 3.6 expected). Mortality was also greater 14 than expected for cancers of the kidney (PMR = 1.50, 8 observed deaths vs. 5.4 15 expected), brain (PMR = 1.56, 9 observed deaths vs. 5.8 expected), liver and gallbladder 16 (PMR = 1.06, 1.06, 5 observed deaths vs. 4.7 expected), pancreas (PMR = 1.05, 13)17 observed deaths vs. 12.3 expected), lung (PMR = 1.08, 72 observed deaths vs. 66.8 18 expected; 2 of these deaths were pleural cancers), oral cavity and pharynx (PMR = 1.13, 19 8 observed deaths vs. 7.1 expected), and lymphohematopoietic cancers (PMR = 1.21, 2520 observed deaths vs. 20.6 expected) including leukemia (PMR = 1.40; 12 observed deaths 21 vs. 8.5 expected). (PCMRs were calculated and were similar to PMRs in most cases, 22 although estimates were less stable for cancers with small numbers of deaths.) Analysis 23 by time since first licensure did not produce markedly different results, with the 24 exception of mortality from skin cancer (PMR<sub><35 years</sub> = 1.73, 4 deaths; PMR<sub> $\geq35$  years</sub> = 25 3.08, 35 deaths). Among non-white males (N = 79), the authors noted that significantly 26 higher mortality from cancers of the larynx (2 observed deaths) and 27 lymphohematopoietic system (3 observed deaths) was found (data not presented). 28 Stratification by type of license among the white male embalmers showed that cancer 29 mortality was generally more elevated among the 546 subjects who practiced only as 30 embalmers than among the 586 who practiced both as embalmers and funeral directors; 31 the authors considered embalmers more highly exposed to formaldehyde than funeral

1	directors. Among those that practiced only as embalmers, only oral cavity and pharyngeal
2	cancer were increased (PMR = $2.01$ , 7 observed deaths vs. $3.5$ expected) but not among
3	those licensed as both embalmers and funeral directors. Statistically significant excess
4	mortality was noted among those that practiced only as embalmers but not among dually
5	licensed subjects for cancers of the skin ( $PMR = 3.26$ , 5 observed cases vs. 1.5 expected,
6	P < 0.05), kidney (PMR = 2.47, 6 observed cases vs. 2.4 expected, $P < 0.05$ ) and brain
7	(PMR = 2.34, 6 observed cases vs. 2.6 expected, $P < 0.05$ ). Lymphohematopoietic
8	cancers (PMR = 1.39, 16 observed cases vs. 11.5 expected), bladder cancer (PMR = 1.32,
9	5 observed deaths vs. 3.8 expected) and gastrointestinal and gallbladder cancers (PMR =
10	1.33, 42 observed deaths vs. 31.7 expected) were elevated only among dually licensed
11	subjects, however.

12 Embalmers: California. The study design and analysis used by Walrath and Fraumeni 13 (1983) was replicated by Walrath and Fraumeni (1984) using a second cohort including 14 all embalmers licensed to practice in California between 1916 and 1978 and known to 15 have died between 1925 and 1980. Licensing records were obtained from the Bureau of 16 Funeral Directing and Embalming in Sacramento, California, and death certificates were 17 obtained for 1,109 eligible subjects (94% male, 96% white). Reported results excluded 63 18 women and 39 non-white men. Mortality from all malignant neoplasms was significantly 19 higher than expected in this cohort (PMR = 1.21, 205 observed deaths vs. 169.9 20 expected; P < 0.05). A statistically significant (P < 0.05) excess mortality was observed 21 for cancers of the colon (PMR = 1.87, 30 observed deaths vs. 16 expected), prostate 22 (PMR = 1.75, 23 observed deaths vs. 13.1 expected), brain and central nervous system 23 (PMR = 1.94, 9 observed deaths vs. 4.7 expected), and leukemia (PMR = 1.75, 12)24 observed deaths vs. 6.9 expected). The excess of leukemia cases was noted largely among 25 embalmers with greater than 20 years licensure (PMR = 2.21, 8 observed deaths; P <26 0.05). Statistically non-significant increases were also noted for cancers of the oral cavity 27 and pharynx (PMR = 1.31, 8 observed deaths vs. 6.1 expected), pancreas (PMR = 1.35, 28 12 observed cases vs. 8.9 expected), bladder (PMR = 1.38, 8 observed deaths vs. 5.8 29 expected), rectum (PMR = 1.02, 7 observed deaths vs. 6.9 expected), all 30 lymphohematopoietic cancers (PMR = 1.22, 19 observed deaths vs. 15.6 expected), and

31 other (unspecified) cancers (PMR = 1.37, 21 observed deaths vs. 15.3 expected).

*Embalmers: Canada.* Levine *et al.* (1984) assembled a cohort of 1,413 male embalmers
first licensed by the Ontario Board of Funeral Services between 1928 and 1957 and
known to have died between 1950 and 1977. Death certificates were obtained from the
Canadian Mortality Database and coded for underlying cause of death by trained
nosologists. Standardized mortality ratios were calculated using expected deaths
determined by applying age- and calendar-year-specific mortality rates among all males
in Ontario from 1950 to 1977. A statistically non-significant increase in deaths from all
lymphohematopoietic cancers was noted (SMR = 1.24, 8 observed cases vs. 6.5 expected,

9 including 4 leukemia deaths vs. 2.5 expected), [though this finding was based on small

10 numbers]. SMRs were less than 1.0 for all other major cancer sites reported, except for

11 sites for which numbers were too small to calculate ratios.

1

2

3

4

5

6

7

8

12 Embalmers and funeral directors: United States. Hayes et al. (1990) conducted a 13 proportionate mortality study of 4,046 (90% white) male embalmers and funeral directors 14 from multiple locations in the United States who had died between 1975 and 1985. 15 Information on occupation and cause of death was ascertained from death certificates, 16 licensing board, and state funeral directors association. Observed numbers of deaths by 17 cause were compared with expected numbers using sex-, race-, 5-year age- and calendar-18 year-specific proportions of deaths among the U.S. general population. Results were 19 stratified by race. A borderline statistically significant increase in all cancers combined 20 was observed among whites (PMR = 1.07, 95% CI = 1.01 to 1.15, 900 deaths) but not 21 among non-whites (PMR = 1.08, 95% CI = 0.87 to 1.31, 102 deaths). Colon cancer was 22 statistically significantly elevated among non-whites (PMR = 2.31, 95% CI = 1.32 to 23 3.76, 16 deaths) but not whites (PMR = 1.18, 95% CI = 0.95 to 1.44, 95 deaths), as were 24 lymphohematopoietic cancers among both whites (PMR = 1.31, 95% CI = 1.06 to 1.59, 25 100 deaths) and non-whites (PMR = 2.41, 95% CI = 1.35 to 3.97, 15 deaths). Mortality 26 from lymphohematopoietic cancers did not vary substantially between embalmers and 27 funeral directors. Among all subjects with lymphohematopoietic cancers, PMRs were 28 statistically significant for myeloid leukemia (PMR = 1.57, 95% CI = 1.01 to 2.34, 2429 deaths) and unspecified leukemias (PMR = 2.28, 95% CI = 1.39 to 3.52, 20 deaths); 30 statistically non-significant excesses were observed for several other histologic subtypes 31 including non-Hodgkin's lymphoma (PMR = 1.26, 95% CI = 0.87 to 1.76, 34 deaths) and

1	multiple myeloma (PMR = $1.37, 95\%$ CI = $0.84$ to $2.12, 20$ deaths). PMRs were non-
2	significantly elevated for several other cancer sites including the oral cavity and pharynx
3	(whites: PMR = 1.19, 95% CI = 0.78 to 1.74, 26 deaths; non-whites: PMR = 1.25, 95%
4	CI = 0.34 to 3.20, 4 deaths); nasopharynx (whites: PMR = 1.89, 95% $CI = 0.39$ to 5.48, 3
5	deaths; non-whites: $PMR = 4.00, 95\%$ CI = 0.10 to 22.29, 1 death); esophagus (whites:
6	PMR = 1.15, 95% CI = 0.72 to 1.73, 22 deaths; non-whites: PMR below 1.0); pancreas
7	(whites: PMR = 1.19, 95% CI = 0.89 to 1.57, 51 deaths; non-whites: PMR = 1.67, 95%
8	CI = 0.72 to 3.29, 8 deaths); skin (whites: PMR = 1.34, 95% $CI = 0.81$ to 2.09, 19 deaths;
9	non-whites: no observed deaths), breast (whites: $PMR = 2.00, 95\%$ CI = 0.24 to 7.22, 2
10	deaths; non-whites: no observed deaths); prostate (whites: $PMR = 1.06, 95\%$ CI = 0.84 to
11	1.32, 79 deaths; non-whites: PMR = 1.35, 95% CI = 0.82 to 2.12, 9 deaths); kidney
12	(whites: $PMR = 1.26, 95\%$ CI = 0.82 to 1.87, 25 deaths; non-whites: $PMR = 1.52, 95\%$
13	CI = 0.18 to 5.50, 2 deaths), eye (whites: PMR = 3.62, 95% $CI = 0.44$ to 13.08, 2 deaths;
14	non-whites: no observed deaths), brain and other central nervous system (whites: PMR =
15	1.23, 95% $CI = 0.80$ to 1.84, 24 deaths; non-whites: no observed deaths), and thyroid
16	(whites: $PMR = 2.37, 95\%$ CI = 0.49 to 6.93, 3 deaths; non-whites: no observed deaths).

## 17 3.2.8.3 U.S. Stern et al. (1987)

Study population. Stern et al. (1987) conducted a retrospective cohort mortality study of 9,365 workers employed from 1940 to June 1979 (Plant A) or May 1980 (Plant B) in two chrome leather tannery plants in the U.S. Approximately 75% of the cohort was male and approximately 80% were white. Vital status was ascertained for 95% of the cohort, using Social Security and National Death Index records. Death certificates were obtained for 96.8% of all deaths.

24 *Exposure assessment*. No exposure monitoring data were available from the plants.

25 Industrial hygiene surveys were conducted by the investigators and used to assess

26 exposures by process and department. Duration of employment was used as a surrogate

- 27 for cumulative exposure. Multiple potentially hazardous agents were used in the tannery
- 28 process, including nitrosamines, chromates, benzidine-based dyes, leather dust, and
- 29 organic solvents, as well as formaldehyde, which was used in the finishing process.
- 30 Ambient formaldehyde levels were measured in the finishing department at the time of

the study and ranged from 0.5 to 7 ppm (mean 2.45 ppm). (Other potential exposures at
 detectable levels in this department included acetone, toluene, methyl isobutyl ketone,

3 butyl cellusolve, and ambient leather fibers.)

4 Statistical methods and results. A modified life-table analysis was used to construct 5 person-years at risk from the start of employment to the end of 1982. A minimum latency 6 period of 15 years was used in some cancer analyses. Expected mortality rates were 7 computed from age-, sex-, race-, and calendar-year-specific rates in the two states in 8 which the plants were located. No statistically significant increases in SMRs for any site-9 specific cancers among the combined cohort were observed; for several sites, significant 10 decreases were observed. With respect to workers in the finishing department who were 11 potentially exposed to formaldehyde, a statistically nonsignificant increase in kidney 12 cancer (SMR = 1.02, 95% CI = 0.26 to 2.73, 3 deaths) and leukemia + aleukemia (SMR = 13 1.25, 95% CI = 0.50 to 2.58, 7 deaths) was observed. One death from squamous-cell 14 carcinoma of the nasal cavity was observed, however, in a man who had worked in the 15 finishing department for over 18 years and died 55 years after the start of employment; 16 the SMR was not estimated, but the annual incidence rate among white males in the 17 United States cited by the authors was approximately 8 in one million at the time of the 18 study). [It is not possible to distinguish a specific effect of formaldehyde in this study, 19 and the power is limited to detect an effect for rare cancers. In addition, there was 20 evidence of a healthy worker effect and a greater than expected number of deaths and 21 accidents.]

22 **3.3 Case-control studies** 

23 Over 40 case-control studies have examined the relationship between occupational 24 exposure to formaldehyde and various cancers. This section reviews epidemiological 25 case-control studies (and some cross-sectional studies) chronologically by major cancer 26 site. The review covers head and neck cancers, lung cancer, lymphohematopoietic 27 malignancies, and cancers at all other sites that have been studied in relation to 28 formaldehyde. Head and neck cancers are further divided into three distinct sections: 29 cancers of the paranasal sinuses and nasal cavity (i.e. sinonasal cancer), cancer of the 30 nasopharynx, and all other head and neck cancers. (See Section 3.1 for a brief orientation 31 to these cancer sites.) See Tables 3-3 to 3-8 for cancer specific tumor site findings.

Some studies evaluated cancer risk at more than one tumor site; results from these studies
 will be presented for each tumor site individually, though the study population and

3 methods will be described only at the first citation.

# 4 3.3.1 Cancers of the paranasal sinuses and nasal cavity

5 This section reviews seven case-control studies that examined the association between 6 formaldehyde and sinonasal carcinoma. Five studies were conducted in Europe (Olsen et 7 al. 1984, 1986; Hayes et al. 1986, Luce et al. 1993a, Pesch et al. 2008), and two in the 8 United States (Roush et al. 1987, Vaughan et al. 1986a). In addition, a cross sectional 9 studies evaluating the association between changes in the nasal mucos among 10 formadehyde exposed workers is discussed (Edling et al. 1987a, 1988). [In a number of 11 these studies, exposure to wood dust may have occurred in addition to formaldehyde. 12 Wood dust is a known human carcinogen with a strong association with sinonasal 13 cancers, predominantly of the adenocarcinoma type; some studies have also reported 14 associations with squamous-cell carcinomas (IARC 1995, NTP 2005a)] 15 3.3.1.1 Denmark: Olsen et al. (1984), Olsen and Asnaes (1986) 16 Study population. The association between occupational formaldehyde exposure and 17 sinonasal and nasopharyngeal cancers was explored in a population-based, case-control 18 study in Denmark (Olsen et al. 1984). Cases of non-sarcoma carcinomas of the sinonasal 19 cavity (N = 488, 66% male) and nasopharynx (N = 266, 68% male) diagnosed between 20 1970 and 1982 were identified using the Danish Cancer Registry (see Section 3.2.2 for 21 results on nasopharyngeal cancer). Eligible controls (N = 2,465) diagnosed with 22 colorectal, prostate, or breast cancer were also selected from the registry and matched to 23 cases (case to control ratio = 1:3) by sex, age (within 5 years), and year of diagnosis 24 (within 5 years). In 1986, Olsen and Asnaes performed a re-analysis after conducting 25 additional data collection to obtain histological information for each case included in 26 their original case-control study. Seven hundred fifty-nine (759) histologically verified 27 cancers of the nasal cavity (N = 287), paranasal sinuses (N = 179), and nasopharynx (N =

28 293) were included in the analysis.

29 Exposure assessment. Information on occupational history since 1964 was obtained by

30 linking subjects with national pension and population registries with information

including job title, industry, job description, company of employment, and period of
employment for each worker. These data, in addition to information about Danish
industries and occupations supplied by the national Labor Inspection Service, were used
by three industrial hygienists blinded to case/control status to classify each subject by
exposure (ever/never) to certain agents including formaldehyde. Each reported job was
further classified as unexposed, certainly exposed, probably exposed, or unknown.

7 Statistical methods and results. Odds ratios were estimated with tabular analysis and 8 Mantel-Haenszel summary estimates were calculated to assess confounding and 9 interaction with wood dust. Among controls, 4.2% of men and 0.1% of women were 10 considered exposed to formaldehyde (percentage of cases exposed not reported); further 11 analyses were thus restricted to men only. Olsen et al. (1984) reported that the RR for 12 sinonasal cancers among men considered certainly exposed to formaldehyde compared 13 with those unexposed was 2.8 (95% CI = 1.8 to 4.3, 33 exposed cases). When a lag time 14 was applied by excluding exposures within 10 years of diagnosis, the corresponding RR 15 increased to 3.1 (95% CI = 1.8 to 5.3, 23 exposed cases). Effect estimates among men 16 considered probably exposed were closer to the null. Exposure to wood dust was 17 evaluated both as a potential confounding factor and as an effect modifier. Among 18 subjects unexposed to wood dust, the RR for any formaldehyde exposure and sinonasal 19 cancers was 1.8 (95% CI = 0.7 to 4.9, 5 cases). Among those unexposed to formaldehyde, 20 the RR for any wood dust exposure and sinonasal cancers was 2.0 (95% CI = 1.1 to 3.7, 8 21 cases). The joint effect of exposure to both formaldehyde and wood dust was 3.5 (95% CI 22 = 2.2 to 5.6, 28 cases). The authors noted that workers with both exposures were at higher 23 risk of nasal cancer than workers with exposure to only one factor. Adjusting for wood 24 dust to evaluate whether the effect of formaldehyde alone was confounded by wood dust, 25 the pooled RR for any formaldehyde exposure was 1.6 (95% CI not reported;  $P \ge 0.05$ ). 26 When a 10-year exposure lag time was applied, the adjusted summary measure was 27 unchanged; however, the joint effect of both exposures increased to 4.1 (95% CI = 2.3 to 28 7.3, 20 cases). Effect estimates for formaldehyde did not markedly change after 29 adjustment by occupational exposure to paint, lacquer, and glue. The authors noted that 30 this study had 80% power to detect an OR of 2.0 for sinonasal cancer.

1 Olsen and Asnaes (1986) reported findings by histological type of cancer. For squamous-2 cell type sinonasal cancers, the RR among men ever exposed to formaldehyde was 2.3 3 (95% CI = 0.9 to 5.8, 13 exposed cases) after adjusting for exposure to wood dust. 4 Among those unexposed to wood dust, the RR was 2.0 (95% CI = 0.7 to 5.9, 4 exposed 5 cases). For adenocarcinoma of the sinonasal cavities, the RR among men exposed to 6 formaldehyde vs. unexposed was 2.2 (95% CI = 0.7 to 7.2, 17 exposed cases) after 7 adjusting for wood dust. Among those unexposed to wood dust, the RR was 7.0 (95% CI 8 = 1.1 to 43.9, 1 exposed case). Restricting exposures to those occurring at least 10 years 9 before diagnosis did not markedly change the magnitude of the effect of formaldehyde on 10 either histologic type of sinonasal cancers. [The difference in RRs adjusted for wood dust 11 and the RRs for only men unexposed to wood dust may reflect residual confounding by 12 wood dust and a loss of precision due to small numbers.]

# 13 3.3.1.2 The Netherlands: Hayes et al. (1986)

14 Study population. One hundred sixteen (116) male residents of the Netherlands aged 35 15 to 79 and diagnosed with histologically confirmed primary epithelial sinonasal cancers 16 between 1978 and 1981 were identified from six major cancer treatment centers in 1982 17 for a case-control study of occupational formaldehyde exposure and other environmental 18 risk factors for sinonasal cancers (Hayes et al. 1986). Sixty seven (67) of the cases (58%) 19 were squamous-cell carcinomas, 28 (24%) adenocarcinomas, and 21 (18%) of other 20 types, mostly undifferentiated. At the start of study implementation, 74 (64%) patients 21 were alive and 42 were deceased. Controls were frequency matched by age and randomly 22 selected from living resident males in 1982 (case to control ratio = 1:2 for living cases, 23 yielding 223 living controls), and from deceased resident males in 1980 (case to control 24 ratio = 1:1 for deceased cases, yielding 36 deceased controls).

*Exposure assessment:* Interviews were conducted in person or on the phone (10%) to
obtain occupational histories for all jobs held at least six months including information
such as year(s) of employment, industry and company, and type of work. Interviews were
completed for 91 cases and 195 controls. Each reported job was first classified by
industry and occupational title. Two industrial hygienists blinded to case status (IH<sub>A</sub> and
IH<sub>B</sub>) then independently classified each occupation and assigned scores of 0 (no

1 exposure) to 9 (highest exposure) based on the level and probability of exposure to

2 formaldehyde. Exposure to wood dust was similarly assessed by one hygienist.

3 Statistical methods and results. Relative risks were estimated along with 90% confidence 4 intervals, and exposure-response trends were evaluated using the Breslow-Day chi-square 5 test for trend. Of the 286 subjects, 65 (23%) were considered exposed to formaldehyde by 6  $IH_A$  and 125 (44%) by IH<sub>B</sub>. Among the 224 subjects considered unlikely to be exposed to 7 wood dust (scores 0 to 2), 15% and 30% were considered exposed to formaldehyde by 8 IH<sub>A</sub> and IH<sub>B</sub>, respectively. The age-adjusted RR for nasal cancer associated with any 9 formaldehyde exposure was 2.5 (90% CI = 1.5 to 4.3) for IH<sub>A</sub> and 1.9 (90% CI = 1.2 to 10 3.0) for IH<sub>B</sub>. These effect estimates did not change after adjustment for smoking or 11 alcohol use. Restricting this analysis to subjects with low exposure to wood dust (scores 0 12 to 2), the age-adjusted RRs for nasal cancer and different levels of exposure to 13 formaldehyde were as follows: (1) any exposure: 2.5 (90% CI = 1.2 to 5.0, 15 exposed 14 cases) for IH<sub>A</sub> and 1.6 (90% CI = 0.9 to 2.8, 24 exposed cases) for IH<sub>B</sub>; (2) low exposure 15 (scores 1 to 2): 2.2 (90% CI = 0.8 to 5.4, 8 exposed cases) for IH<sub>A</sub> and 1.0 (90% CI = 0.4 16 to 2.5, 7 exposed cases) for IH<sub>B</sub>: and (3) high exposure (scores 3 to 9): 3.0 (90% CI = 1.017 to 8.7, 7 exposed cases) for IH<sub>A</sub> and 2.1 (90% CI = 1.1 to 4.1, 17 exposed cases) for IH<sub>B</sub>. 18 Among subjects with low exposure to wood dust, elevated RRs for squamous cell nasal 19 carcinoma were also observed: (1) any exposure: 3.0 (90% CI = 1.3 to 6.4, 12 exposed)20 cases) for IH<sub>A</sub> and 1.9 (90% CI = 1.0 to 3.6, 19 exposed cases) for IH<sub>B</sub>; (2) high 21 exposure: 3.1 (90% CI = 0.9 to 10.0, 5 exposed cases) for IH<sub>A</sub> and 2.4 (90% CI = 1.1 to 22 5.1, 13 exposed cases) for  $IH_B$ . (There were insufficient numbers of cases of 23 adenocarcinomas with low wood dust exposure to permit a separate analysis of 24 formaldehyde exposure, according to the authors.) The authors noted that though exposure 25 assessment by IH<sub>A</sub> and IH<sub>B</sub> varied, all effect estimates were positive and thus suggested 26 an increased risk of sinonasal cancers associated with occupational exposure to 27 formaldehyde despite intra-rater variability.

- 28 3.3.1.3 Washington State: Vaughan et al. (1986a)
- 29 Study population. A population-based case-control study was conducted by Vaughan et
- 30 al. (1986a) to determine whether occupational exposure to formaldehyde in 13 counties

1 in Washington, USA was associated with sinonasal or pharyngeal cancer (see Sections 2 3.2.2 and 3.2.3 for results on the different types of pharyngeal cancer). Incident cases 3 were identified through a population-based cancer registry operated as part of the 4 Surveillance, Epidemiology and End Results (SEER) program of the National Cancer 5 Institute. Eligible cases were aged 20 to 74 years at enrollment, resided in the study area, 6 and were diagnosed during the period 1979 to 1983 for sinonasal cancer, and 1980 to 7 1983 for pharyngeal cancer. Controls from the study area were identified using random-8 digit dialing and frequency-matched to cases by age and sex. Information about medical, 9 smoking, alcohol, residential, and occupational histories was either self-reported or 10 reported by next-of-kin (for deceased cases) in a telephone interview. Two hundred 11 eighty-five cases (285) (69% of eligible cases) including 53 sinonasal, 27 12 nasopharyngeal, and 205 oro- or hypopharyngeal cases were included in the analysis; half 13 the case interviews were conducted with next-of-kin. Of 690 eligible controls, 552 (80%) 14 were included in the analysis. 15 Exposure assessment. Occupational formaldehyde exposure was assessed using a job-

16 exposure linkage system in which each unique job is identified by the 3-digit U.S. Census 17 occupation and industry codes. Estimates of the likelihood and intensity of formaldehyde 18 exposure for each job were combined to create a 4-level summary exposure metric: (1) 19 high = probable exposure to high levels. (2) medium = probable exposure to low levels. 20 (3) low = possible exposure at any level, and (4) background = no occupational exposure. 21 Exposure assignments were made blinded to case status. Individual estimates of exposure 22 to formaldehyde were then calculated for each subject including maximum lifetime 23 intensity, lifetime duration, and cumulative exposure.

Statistical methods and results. Unconditional logistic regression was used to produce
ORs adjusted for sex, age, smoking, alcohol use, and race. Over 90% of sinonasal cancers
occurred among subjects with cumulative exposure scores less than 5 because most cases
were classified as being unexposed (0 years lifetime exposure) and having a lifetime
maximum exposure intensity level of "background." Effect estimates were based on very
small numbers of exposed cases (12 cases exposed at any level, 3 cases exposed for at
least 10 years) and showed no increase in risk associated with formaldehyde exposure.

1 Cumulative exposure scores were also analyzed excluding jobs within 15 years of the 2 date of diagnosis to account for a latency period. For sinonasal cancers, this exposure 3 lagging resulted in only one case in the highest exposure category and did not produce 4 interpretable estimates. The authors noted some methodological limitations including low 5 statistical power, non-differential exposure misclassification, and bias due to recall error 6 by next-of-kin. This latter limitation was explored by examining data obtained from live 7 cases only; live cases reported a higher mean number of jobs than proxies, and most ORs 8 increased in magnitude when restricted to live cases only.

9 3.3.1.4 Connecticut: Roush et al. (1987)

10 *Study population.* From the Connecticut Tumor Registry, Roush *et al.* (1987) identified 11 198 cases of sinonasal cancer and 173 cases of nasopharyngeal cancer (see Section 3.2.2 12 for results on nasopharyngeal cancer) among male residents of Connecticut who had died 13 of any cause between 1935 and 1975. Controls (N = 605) were randomly selected without 14 stratification or matching from male residents who died during the same time period.

15 *Exposure assessment.* Occupational information including job title, industry, and year(s) 16 of employment was obtained from death certificates and from annual city directories; the 17 latter were examined for the years corresponding to 1, 10, 20, 25, 30, 40 and 50 years 18 before death (as long as the subject was  $\geq 20$  years old at each assessment). An industrial 19 hygienist blinded to case/control status classified each reported job by probability and 20 level of exposure to formaldehyde, and subsequently categorized each subject into 4 21 exposure groups: (1) probably exposed to some level for most of working life, (2) 22 probably exposed to some level for most of working life and probably exposed to some 23 level at 20+ years prior to death, (3) probably exposed to some level for most of working 24 life and probably exposed to high level in some year, and (4) probably exposed to some 25 level for most of working life and probably exposed to high level at 20+ years prior to 26 death. This latter exposure category was intended to capture short-term high exposures 27 and account for the latency period necessary for sinonasal cancers to develop.

Statistical methods and results. Logistic regression was applied to estimate ORs and 95%
confidence intervals. Approximately 47% of sinonasal cancer cases had occupational
information for three or more jobs; 11% of sinonasal cancer cases were categorized into

1 exposure level 1 (N = 21), 8% in level 2 (N = 16), 4.5% in level 3 (N = 9), and 3.5% in 2 level 4 (N = 7). No association between occupational exposure to formaldehyde and 3 sinonasal cancers was observed for levels 1 to 3. The OR for men who were probably 4 exposed to some level for most of their working life and probably exposed to high levels 5 at some point 20 years or more before death (level 4) was 1.5 (95% CI = 0.6 to 3.9, 7 6 exposed cases).

7 3.3.1.5 Sweden: Edling et al. (1987a, 1988)

8 Study population. In this small cross-sectional study of woodworkers in a Swedish plant, 9 histological changes in nasal mucosa among 38 woodworkers (35% of whom were ever 10 smokers) who were engaged in processing [laminate] were compared with 25 unexposed 11 men (48% ever smokers) working elsewhere in the same plant. Ninety-two percent (92%) 12 of the men exposed to formaldehyde agreed to be medically examined, with an average 13 length of exposure of 6 years (Edling *et al.* 1987a). In a follow-up to this preliminary 14 investigation (Edling et al. 1988), clinical and histological findings were described for a 15 total of 75 men who exposed to formaldehyde out of a possible 104 exposed workers at 16 three plants, two of which processed particle board and one, laminate (72% participation 17 rate). (This group of men presumably included all 38 studied previously in the laminate 18 plant.) Findings were compared to 25 unexposed workers.

*Exposure assessment.* Industrial hygiene measurements between 1975 and 1983 at the three plants indicated ambient exposures to formaldehyde ranging from 0.1 to 1.1 mg/m<sup>3</sup>, with peaks of up to 5 mg/m<sup>3</sup>. No exposure measurements were available prior to this date but were repsumed to have been higher. Wood dust levels in the two plants processing particle board ranged from approximately 0.6 to 1.1 mg/m<sup>3</sup>. Exposure histories for individual workers were not estimated. Workers in the laminate plant were not exposed to wood dust, according to the authors.

26 *Results.* In the initial study of the laminae workers, a significant difference (P < 0.05) in

- 27 the histological score for the presence of precancerous hyperplasia and squamous
- 28 metaplasia of the nasal mucosa was observed among exposed workers in comparison
- 29 with nonexposed workers. No clear relationship with duration of exposure was observed.
- 30 Ever smoking was associated with a statistically nonsignificant increase in abnormal

1 histology but did not explain the difference in scores between exposed and nonexposed 2 workers, although there was some evidence of a synergistic effect of smoking with 3 formaldehyde exposure, according to the authors. In the follow-up of all 75 4 formaldehyde-exposed workers, the average exposure duration ranged from 1 to 39 years 5 with a mean of 10.6 years. Thirty-three (33) of the exposed workers were smokers or ex-6 smokers compared to 16 of the unexposed group. Normal nasal mucosa were observed in 7 only three exposed men, and mild dysplasia, hyperplasia and squamous metaplasia of the 8 nasal mucosa was observed in the remainder of the exposed group; the average 9 histological score (2.9) was significantly higher than that for the unexposed workers (1.8, 10 P < 0.05). Among exposed workers, this score was not related to duration of exposure, 11 however; smokers had a somewhat higher but not statistically significantly different score 12 compared to non- and ex-smokers. No difference in histological scores was found when 13 workers in the particle board plants (also exposed to wood dust) were compared with 14 those in the lamina plant.

15 3.3.1.6 France: Luce et al. (1993)

16 Study population. Luce et al. (1993a) reported on a case-control study of primary 17 sinonasal cancer in France. Cases of sinonasal cancers (N = 303) diagnosed between 18 January 1986 and February 1988 among male and female residents of France were 19 identified at 27 hospitals; 207 (67%) cases were enrolled in the study. All but one case 20 was histologically confirmed. Two control series were enrolled. A hospital-based control 21 series included patients with cancers other than sinonasal cancers diagnosed during the 22 same time period as cases at the same or nearby hospitals; of 340 eligible hospital 23 controls, 323 (95%) were enrolled and frequency matched by age and sex (case to control 24 ratio = 2:3). A population-based control series was selected from lists of friends and 25 family provided by cases; of 103 eligible convenience controls, 86 (84%) were enrolled 26 and matched to cases by sex, age (within 10 years), and residence.

27 *Exposure assessment.* Interviews were conducted by trained physicians to elicit

28 information on socio-demographic characteristics, smoking and alcohol intake, medical

29 history and nasal diseases, and occupational history. An additional questionnaire was

30 administered to assess occupational exposure to a pre-determined list of substances

1 including formaldehyde. Exposure assessment was performed by an industrial hygienist

2 blinded to case/control status and involved classifying each subject according to

3 probability of exposure based on information from the questionnaires. Jobs considered

4 exposed to formaldehyde were further classified by exposure frequency, concentration,

5 and cumulative exposure.

6 Statistical methods and results. Multivariate logistic regression was used to estimate ORs 7 and 95% confidence intervals and to evaluate confounding by occupational and non-8 occupational factors. Odds ratios were stratified by histologic subtype and sex (regression 9 results were reported for men only), and adjusted by age and exposure to wood dust, 10 glues, and adhesives. The two control series were combined for analysis. [Eligible 11 controls included participants with cancers suspected to be associated with formaldehyde 12 exposure, which might have attenuated observed effect estimates.] Among cases, 36% of 13 males (N = 60) and 25% of females (N = 10) were exposed to formaldehyde; among 14 controls, 55% of males (N = 176) and 29% of females (N = 26) were exposed. Analyses 15 were based on 16 cases with probable or definite exposure and 81 controls. The 16 proportion of subjects with at least one probable or definite exposure was higher among 17 exposed cases than among exposed controls. However, regression results showed no 18 relationship between any formaldehyde exposure index and squamous-cell sinonasal 19 cancers among males. The OR for adenocarcinoma-type sinonasal cancers and any 20 exposure to formaldehyde was 8.1 (95% CI = 0.9 to 72.9, 4 exposed cases) among those 21 unexposed to wood dust and 692 (95% CI = 91.9 to 5,210, 71 exposed cases) among 22 those jointly exposed to wood dust and formaldehyde. [The association between 23 formaldehyde and adenocarcinoma-type sinonasal cancers independent of exposure to 24 wood dust could not be estimated with any precision in this study because the majority of 25 subjects with probable or definite exposure to formaldehyde were also exposed to wood 26 dust (97% of subjects were jointly exposed). Among subjects with cancers of "other" 27 histologies (7 esthesioneuromas, 3 sarcomas, 2 melanomas, 1 lymphoma, and 4 28 unspecified cases), a positive association was generally observed for subjects with 29 probable or definite exposure to formaldehyde. For the highest index exposure levels of 30 these other histologies, ORs ranged from 1.62 (exposure duration > 20 years) to 3.27 31 (date of first exposure  $\geq$  1955); only the latter estimate was statistically significant (95%)

1 CI = 1.15 to 9.33, 6 cases). The authors noted that adjustment by smoking and re-analysis

2 taking into account a 15-year induction period did not markedly change the reported

3 effect estimates.

4 3.3.1.7 Germany: Pesch et al. (2008)

5 Study population. Pesch et al. (2008) conducted a case-control study of workers in the

6 woodworking industry in Germany with histologically confirmed diagnosis of

7 adenocarcinoma of the nasal cavity or paranasal sinuses between 1994 and 2003. 86 cases

8 (57 survivors and 29 next of kin) agreed to participate and were matched with 204

9 frequency matched controls (including 69 next of kin).

10 *Exposure assessment.* A semi-quantitative job exposure matrix was constructed for each

11 subject based on occupational histories, job titles and types of materials used within the

12 woodworking industry, together with previously monitored wood dust exposure

13 measurements conducted within the industry. Potential exposures included wood dust

14 particulates, wood preservatives, stains, and varnishes, as well as formaldehyde.

Statistical methods and results. Logistic regression conditional on age and adjusted for smoking and other demographic variables was used to calculate odds ratios for low, medium and high levels of average and cumulative exposures, duration of exposure, and time since first exposure to select agents. Inhalable wood dust exposure was associated with a highly significant increase in the risk of ADCN, but formaldehyde exposure (either pre- or post 1985) adjusted for wood dust exposure was not associated with a significant

21 increase in risk (ORs were less than 1.0 and statistically nonsignificant).

22 3.3.2 Cancer of the nasopharynx

23 Section 3.2.2 reviews case-control studies that examined the association between

24 formaldehyde and nasopharyngeal cancer. Three studies were conducted in Asia

25 (Armstrong et al. 2000, Hildesheim et al. 2001, West et al. 1993), one in Europe (Olsen

and Asnaes 1986, Olsen et al. 1984) and three in the United States (Roush et al. 1987,

27 Vaughan et al. 2000, Vaughan et al. 1986a). Some of these studies were described

previously in Section 3.2.1 (Olsen and Asnaes 1986, Olsen et al. 1984, Roush et al. 1987,

29 Vaughan *et al.* 1986a).

#### 1 3.3.2.1 Denmark: Olsen et al. 1984, Olsen and Asnaes 1986

- 2 Olsen *et al.* (1984) also evaluated the association between formaldehyde exposure in the
- 3 workplace and risk of nasopharyngeal carcinoma (N = 266 cases, 2,465 controls) in a
- 4 population-based, case-control study in Denmark (see Section 3.2.1 for complete study
- 5 description). Among controls, 4.2% of men and 0.1% of women were considered exposed
- 6 to formaldehyde (percentage of cases exposed not reported). The RR for nasopharyngeal
- 7 carcinoma comparing those ever exposed vs. never exposed was 0.7 (95% CI = 0.3 to 1.7,
- 8 no. of exposed cases not reported) among men and 2.6 (95% CI = 0.3 to 21.9) among
- 9 women. Analysis of nasopharyngeal cancers by histologic subtype did not show any
- 10 association with either formaldehyde or wood dust (Olsen and Asnaes 1986).
- 11 3.3.2.2 Washington State: Vaughan et al. (1986a)

12 The association between nasopharyngeal cancers (N = 27) and occupational 13 formaldehyde exposure was also examined by Vaughan et al. (1986a) in the population-14 based, case-control study in Washington state (see Section 3.3.1 for complete study 15 description and results on sinonasal cancers; see Section 3.3.3 for results on oro- and 16 hypopharyngeal cancer). Approximately 60% of nasopharyngeal cancers occurred among 17 subjects classified as unexposed; cumulative exposure scores less than 5 represented over 18 75% of cases. Adjusting for race and smoking, the ORs for nasopharyngeal cancers for low and medium/high exposure were 1.2 (95% CI = 0.5 to 3.3, 7 exposed cases) and 1.4 19 20 (95% CI = 0.4 to 4.7, 4 exposed cases), respectively, compared with subjects with a 21 background level maximum lifetime exposure (unexposed). Compared with subjects with 22 zero years of lifetime exposure, the ORs for 1 to 9 years duration were 1.2 (95% CI = 0.523 to 3.1, 8 exposed cases) and for 10+ years 1.6 (95% CI = 0.4 to 5.8, 3 exposed cases). 24 Cumulative exposure estimates were 0.9 (95% CI = 0.2 to 3.23, 3 exposed cases) for 25 scores 5 to 19 and 2.1 (95% CI = 0.6 to 7.8, 3 exposed cases) for scores 20+ compared 26 with scores less than 5. Cumulative exposure scores were also analyzed excluding job 27 histories within 15 years of the date of diagnosis to account for a cancer latency period. 28 The OR for the 5 to 19 exposure score group was 1.7 (95% CI = 0.5 to 5.7, 4 exposed)29 cases); the point estimate for the 20+ group did not change.

- 1 3.3.2.3 *Connecticut: Roush* et al. (1987)
- 2 Occupational exposure to formaldehyde and mortality from nasopharyngeal cancers
- 3 among men (N = 173) was also investigated by Roush *et al.* (1987) in their population-
- 4 based, case-control study in Connecticut (see Section 3.2.1 for complete study
- 5 description). The OR for nasopharyngeal cancer mortality among men was 1.0 (95% CI = 1.0% CI)
- 6 0.6 to 1.7, 21 exposed cases) for level 1, 1.3 (95% CI = 0.7 to 2.4, 17 exposed cases) for
- 7 level 2, 1.4 (95% CI = 0.6 to 3.1, 9 exposed cases) for level 3, and 2.3 (95% CI = 0.9 to
- 8 6.0, 7 exposed cases) for level 4 exposure category.
- 9 3.3.2.4 Philippines: West et al. (1993)

10 Study population. West et al. (1993) investigated non-viral risk factors including

11 occupational exposure to formaldehyde for nasopharyngeal cancers in the Philippines.

12 This hospital-based, case-control study included 104 incident cases of histologically

13 confirmed nasopharyngeal cancers (100% participation rate, 73% male) recruited from

14 the Philippine General Hospital, and two control series: 104 hospital controls (100%

15 participation rate) matched to cases by sex, age, and hospital ward type (public vs.

16 private), and 101 community controls (77% participation rate) matched to cases by sex,

17 age, and neighborhood.

18 *Exposure assessment.* During interviews conducted with a trained nurse, information was 19 collected on socio-demographics, diet, smoking, occupational history, and use of herbal 20 medicines, betel nut, and anti-mosquito coils. Reported occupations were classified by a 21 industrial hygienist blinded to case/control status as likely or unlikely to involve exposure 22 to formaldehyde, solvents, wood dust and other dusts, and pesticides. This classification 23 was then combined with information from the complete occupational history to obtain for 24 each individual four estimates of exposure: (1) overall duration of exposure, (2) duration 25 excluding exposure in the 10 years preceding diagnosis (for cases) or interview (for 26 controls), (3) years since first exposure, and (4) age at first exposure.

Statistical methods and results. Conditional logistic regression was applied to estimate
ORs and 95% CIs. The authors reported that results of the occupational analyses were
similar for each control series and thus combined controls for analyses. Estimates of
association for formaldehyde and nasopharyngeal cancers were reduced toward the null

1 after adjusting for years since first exposure to dusts and/or exhaust fumes. Overall 2 duration of exposure was not clearly associated with nasopharyngeal cancers after 3 adjusting for exposure to dusts and/or exhaust; however, duration of exposure lagged by 4 10 years yielded an increased risk (RR (2.1, 95% CI = 0.70 to 6.2, 8 exposed cases) for 5 subjects with at least 15 years exposure. Statistically significant effects were observed for 6 formaldehyde with 25+ years since first exposure (RR = 2.9, 95% CI = 1.1 to 7.6, 14 7 cases) and among subjects who were < 25 years old at first exposure (RR 2.7, 95% CI = 8 1.1 to 6.6, 16 cases), adjusted for years since first exposure to dusts and/or exhaust 9 (unlagged estimates). The RR for subjects jointly exposed to both formaldehyde (25+ 10 years since first exposure) and dust/exhaust (35+ years since first exposure) compared 11 with subjects with neither exposure was 15.7 (95% CI = 2.7 to 91.2, no. exposed subjects 12 not reported). In further models, a statistically significantly increased risk of 13 nasopharyngeal cancers was also observed with increasing years since first exposure to 14 formaldehyde after adjusting for other confounding factors including education, exposure 15 to dust and exhaust, diet, smoking, and use of herbal medicines and anti-mosquito coils. 16 Compared with subjects never exposed to formaldehyde, the RRs were 1.2 (95% CI = 17 0.41 to 3.6, 12 exposed cases) for subjects first exposed less than 25 years before 18 diagnosis or interview, and 4.0 (95% CI = 1.3 to 12.3, 14 exposed cases) for subjects first 19 exposed 25 years or more ago.

20 3.3.2.5 Malaysia: Armstrong et al. (2000)

21 Study population. Histologically confirmed cases of nasopharyngeal cancers (all 22 squamous-cell carcinomas) diagnosed or treated in Kuala Lumpur and Selangor from 23 January 1987 to June 1992 were assembled for a case-control study of nasopharyngeal 24 cancers and work-site inhalation of dust and smoke particles, formaldehyde, and certain 25 aromatic hydrocarbons among Malaysian Chinese (Armstrong et al. 2000). Of 530 26 eligible cases who had lived in the study area for at least 5 years, 282 (53%) were 27 enrolled (31% female). Each case was matched by sex and age (within 3 years) to one 28 control with no history of head, neck, or respiratory system cancer; controls were selected 29 from the general population using a house-to-house multistage area sampling.

1 *Exposure assessment.* Data on residential history, occupational history, diet, and tobacco 2 and alcohol use were collected by trained interviewers during two in-home structured 3 interviews. Occupational history included information about job description, tasks, 4 workplace characteristics, use of industrial equipment and substances, and exposure to 5 dusts, smoke, gases, and chemicals at each job. Additional information about exposures 6 to industrial heat and 20 inhalants known to be deposited or absorbed in the nasopharynx 7 were collected by trade or profession, calendar time, frequency and duration. Jobs were 8 classified according to official Malaysian occupational codes, and exposure for each 9 occupational code was assigned by a study investigator blinded to case/control status and 10 familiar with Malaysian industry. Industries considered exposed to formaldehyde 11 included adhesives, foundries, latex processing, metalworking and welding, plywood 12 manufacturing, rubber tire manufacturing, sawmilling, shoe-making (glues), and textiles 13 (permanent press fabrics). Four categories of exposure to inhalants (never, low, medium, 14 high) were created based on job type, task, mode of exposure (inhalation and/or dermal), 15 interview data on exposure, years of exposure, frequency, and duration. To account for 16 latency, cumulative exposure was evaluated using 5 lag time periods: > 1, 5, 10, 15, and17 20 years prior to diagnosis. Exposure intensity was also assessed by categorizing 18 participants according to cumulative years exposed. The authors presented air monitoring 19 data for formaldehyde levels within 10 industries (42 worksites) reported by participants 20 in this study. Samples were taken in 1991 to 1992 and showed that formaldehyde levels exceeded the recommended limit  $(0.37 \text{ mg/m}^3)$  in the adhesives industry only, and the 21 22 range of levels for all other industries sampled was wide (mean 8-hour concentration 0.16 23 to  $0.35 \text{ mg/m}^3$ ).

24 Statistical methods and results. For analysis, Armstrong et al. examined exposure 25 dichotomously (ever/never) as well as by cumulative duration using conditional logistic 26 regression. Approximately 10% of cases were considered exposed to formaldehyde 27 compared with 8.2% of controls. The unadjusted OR for ever/never formaldehyde 28 exposure and nasopharyngeal cancers was 1.24 (95% CI = 0.67 to 2.32, cases not)29 specified); the diet and smoking-adjusted estimate was 0.71 (95% CI = 0.34 to 1.43). The 30 authors assessed dose-response in relation to a 10-fold increase in ratio of hours exposed; 31 no dose-response trend was observed with increasing duration of formaldehyde exposure. 1 No difference in effect estimates was observed in analyses by lag time or intensity. [The

2 participation rate among diagnosed cases was low (53%); according to the authors, the

3 possibility of prevalence-incidence or other forms of selection bias could not be

4 excluded. In addition, although some inhalants (wood dust, for example) were found to

5 be significantly associated with nasopharyngeal cancers in these data, these factors were

6 not evaluated as potential confounders when evaluating the relationship between

7 formaldehyde and the outcome.]

# 8 3.3.2.6 United States – SEER: Vaughan et al. (2000)

9 *Population.* To further investigate whether occupational exposures to formaldehyde and 10 wood dust increase the risk of nasopharyngeal cancers, Vaughan et al. (2000) conducted 11 a cancer registry-based population, case-control study that identified 294 nasopharyngeal 12 cancer cases (diagnosed between April 1987 and June 1993 among persons 18 to 74 years 13 of age) from five cancer registries (Connecticut, Detroit, Iowa, Utah, and Washington) in 14 the National Cancer Institute's Surveillance, Epidemiology and End Results (SEER) 15 program. This study focused on a subset of 196 interviewed cases (68% male) diagnosed with epithelial carcinoma including epithelial not-otherwise-specified (N = 24), 16 17 undifferentiated or non-keratinizing (N = 54), and differentiated squamous-cell types (N 18 = 118). Controls were identified from the same geographic locations using random digit 19 dialing, and were frequency matched to cases by age (within 5 years), sex, and cancer 20 registry. Of 2,885 households contacted, 244 of 324 eligible controls were successfully 21 enrolled and interviewed.

22 *Exposure assessment.* Structured telephone interviews were conducted with study 23 participants or proxies (44 case and 3 control interviews by proxy) collecting information 24 on demographics, personal and family medical history, tobacco and alcohol use, and 25 lifetime history of occupational and chemical exposure; information since diagnosis for 26 cases or since ascertainment for controls was excluded. Information collected about 27 occupational history for any job held at least 6 months included job title, tasks, industry 28 type, calendar dates, and exposure to specific chemicals or other agents including wood 29 dust and formaldehyde. Participants were also asked specifically about any jobs held in 30 particular industries including furniture manufacturing, construction, foundry, and

1	smelting. Industrial hygienists blinded to case/control status used these data combined
2	with estimates from both published and unpublished literature to assess exposure to
3	formaldehyde for each unique reported job. Each job was assigned a probability of
4	formaldehyde exposure based on the percentage of workers with a similar job profile
5	expected to be exposed: definitely not or unlikely (< 10%), possible (10% to < 50%),
6	probable (50% to < 90%), and definite ( $\geq$ 90%). Using information about frequency
7	(days/year) and duration (hours/day), jobs with potential exposure were further classified
8	by the estimated concentration of exposure representing an 8-hour time-weighted average
9	(TWA-8): low (< 0.10 ppm), moderate (0.10 to < 0.50 ppm), and high ( $\geq 0.50$ ppm).
10	Twenty-four (24) reported jobs (of 2,209 unique reported jobs) were considered to entail
11	exposure to formaldehyde; 19 were classified as definitely exposed (16 low-level and 3
12	moderate), 3 as probable (all low-level), and 2 as possible (1 low-level and 1 moderate).
13	Exposure to wood dust was assessed by identifying jobs in occupational or industry codes
14	considered exposed, and by using interview data of subjects self-reported as exposed to
15	wood dust; jobs were assigned total wood dust TWA-8 estimates. Using results from the
16	exposure assessment, exposure to formaldehyde and wood dust were coded using the
17	following variables: ever exposed, maximum concentration exposed, duration exposed,
18	and cumulative exposure. Duration and cumulative exposure were further evaluated with
19	a 10-year lag.

20 Statistical methods and results. Multivariate logistic regression was used to estimate the 21 association between nasopharyngeal cancers and exposure to formaldehyde and wood 22 dust. Confounding and effect measure modification by age, sex, race, SEER site, 23 smoking, alcohol intake, education, and proxy status were evaluated. Forty-three percent 24 (43%) of cases were potentially exposed to formaldehyde, compared with 32% of 25 controls. The adjusted (age, sex, race, SEER site, smoking, education, and proxy status) 26 OR for nasopharyngeal cancers comparing ever occupationally exposed to unexposed by 27 histological subtype was 1.3 (95% CI = 0.8 to 2.1, 79 exposed cases) for all epithelial, 0.9 28 (95% CI = 0.4 to 2.0, 18 exposed cases) for undifferentiated or non-keratinizing, 1.5 29 (95% CI = 0.8 to 2.7, 49 exposed cases) for differentiated squamous-cell, and 3.1 (95% 30 CI = 1.0 to 9.6, 12 exposed cases) for epithelial NOS [not otherwise specified]. No 31 consistent pattern of association or trend in risk was observed with maximum lifetime

1	exposure concentration. For lifetime duration of exposure and risk of nasopharyngeal
2	cancers, there was some evidence of an increased risk of nasopharyngeal cancers with
3	increasing lifetime duration of exposure among all subjects with any possibility of
4	exposure ( $P_{\text{trend}} = 0.014$ , 79 exposed cases); the OR for subjects who had worked at least
5	18 years in potentially exposed jobs was 2.7 (95% $CI = 1.2$ to 6.0, 25 exposed cases). A
6	trend was observed with increasing years of exposure ( $P_{\text{trend}} = 0.070$ ); the adjusted OR
7	for subjects who had worked at least 18 years in potentially exposed jobs was 2.1 (95%
8	CI = 1.0 to 4.5, 29 exposed cases). This trend was stronger for differentiated squamous
9	cell ( $P_{\text{trend}} = 0.033$ ) and epithelial NOS ( $P_{\text{trend}} = 0.036$ ) histologies than undifferentiated
10	or non-keratinizing types ( $P_{\text{trend}} = 0.820$ ). The adjusted ORs for 61 cases of
11	nasopharyngeal cancers (excluding undifferentiated or non-keratinizing type) for
12	estimated probability of formaldehyde exposure were $1.6 (95\% \text{ CI} = 1.0 \text{ to } 2.8, 61$
13	exposed cases) for ever having a job classified as possibly, probably, or definitely
14	exposed, 2.1 (95% $CI = 1.1$ to 4.2, 27 exposed cases) for probably or definitely exposed,
15	and 13.3 (95% $CI = 2.5$ to 70.0, 10 exposed cases) for definitely exposed. Again, among
16	the group of cases excluding undifferentiated and non-keratinizing types, there was some
17	evidence of an increased risk of nasopharyngeal cancers with increasing lifetime duration
18	of exposure among all subjects with any potential exposure ( $P_{\text{trend}} = 0.014$ ); the OR for
19	subjects who had worked at least 18 years in any potentially exposed jobs was 2.7 (95%
20	CI = 1.2 to 6.0, 25 exposed cases). The risk of nasopharyngeal cancers also increased
21	with increasing cumulative exposure ( $P_{\text{trend}} = 0.033$ ) among all potentially exposed
22	subjects. The OR for subjects in the highest category of cumulative exposure (> $1.10$
23	ppm-yrs) was 3.0 (95% CI = 1.3 to 6.6, 24 exposed cases). The authors reported that
24	estimates were similar when exposures were lagged by 10 years, and that adjustment by
25	exposure to wood dust did not affect results for exposure to formaldehyde. However,
26	some evidence of effect measure modification by smoking was observed; measures of
27	association as well as estimates of trend were generally stronger among current and
28	former smokers than non-smokers. [A strength of this study is its large sample size,
29	which improved the precision of the effect estimates and allowed for adjustment of the
30	effect estimates by a number of potentially confounding factors, after which a positive
31	association between formaldehyde exposure and nasopharyngeal cancers still remained.]

1 3.3.2.7 *Taiwan: Hildesheim* et al. (2001)

2 *Study population.* Hildesheim *et al.* (2001) conducted a population-based, case-control

3 study of nasopharyngeal cancers and occupational exposure to wood dusts,

4 formaldehyde, and solvents in Taipei, Taiwan. Incident cases of histologically confirmed

5 nasopharyngeal cancers diagnosed between July 1991 and December 1994 were

6 identified from two tertiary care hospitals in Taipei; eligible cases (N = 378) were

7 residents of Taipei city or county for at least six months, and were less than 75 years of

8 age. Ninety-nine percent (99%) of eligible cases (N = 375, 69% male) agreed to

9 participate. Over 90% of cases were diagnosed with non-kertainizing or undifferentiated

10 carcinomas and the remainder with squamous-cell carcinomas. Controls were identified

11 using a National Household Registration System and were individually matched to cases

12 (case to control ratio = 1:1) on age (within 5 years), sex, and area of residence. Eligible

13 controls (N = 376) lived in Taipei city or county for at least six months and had no

14 history of nasopharyngeal cancer; 87% (N = 327) agreed to participate.

15 *Exposure assessment.* Interviews administered to each participant by a trained nurse

16 collected information about occupational, medical, and residential histories,

17 demographics, diet, smoking and alcohol use. Occupational histories were collected for

18 all jobs held for at least one year and included information on job title, industry,

19 duties/activities, and tools/materials used on the job. Exposure assessment was conducted

20 by an industrial hygienist blinded to case/control status; jobs were first classified into

21 Standard Industry/Occupational Classification codes, and then each code was evaluated

22 for probability and intensity of exposure to formaldehyde, wood dusts, and solvents and

assigned a score of 0 (unexposed) to 9 (< 4 was considered low,  $\geq$  4 high). For each

subject, this score plus information about duration were combined to produce six

estimates of exposure: (1) years of exposure, (2) average intensity, (3) average

probability, (4) cumulative exposure, (5) age at first exposure, and (6) years since first

27 exposure. Duration of exposure was also calculated excluding exposures occurring within

- 28 10 years of diagnosis (for cases) or interview (for controls). Occupational data were
- available for 100% of cases and over 99% of controls. Of the 2,034 jobs reported by all
- 30 700 subjects, 156 (7.7%) were classified as exposed to formaldehyde; 74 cases and 41
- 31 controls were considered "ever" exposed. Some of the reported occupations considered

1	exposed to formaldehyde included farmers ( $N = 68$ ), barbers, hairdressers, and
2	cosmetologists (N = 15), carpenters (N = 14), and health professionals (N = 13).
3	Statistical methods and results. Unconditional logistic regression was used to estimate
4	ORs [reported as risk ratios] for the association between formaldehyde exposure and
5	nasopharyngeal cancers. Exposure-response trends were assessed by entering exposure
6	into the model as a continuous variable and testing the resulting ß-coefficient.
7	Stratification was used to examine effects by age, sex, Epstein-Barr virus (EBV)
8	seroprevalence (established as a risk factor for the development of nasopharyngeal
9	cancers), and histologic subtype. After adjustment by age, sex, education, and ethnicity,
10	the OR for subjects ever exposed to formaldehyde vs. never exposed was $1.4 (95\% \text{ CI} =$
11	0.93 to 2.2, 74 exposed cases). Risk increased with increasing duration of exposure ( $P_{\text{trend}}$
12	= 0.08) and increasing cumulative exposure ( $P_{\text{trend}} = 0.10$ ). The observed trend was lower
13	when a 10-year exposure lag was applied. Increased risks were were observed among
14	subjects with high average intensity or high probability of exposure compared with low
15	exposure intensity or probability. No clear pattern of risk was observed in analyses by
16	age at first exposure or years since first exposure. The authors noted that estimates were
17	unaffected by adjustment for wood dust or solvent exposure. The OR estimating the joint
18	effect of formaldehyde and wood dust was 1.8 (95% CI not reported). Among subjects
19	who were seropositive for EBV, the adjusted OR for ever exposure to formaldehyde
20	exposure was higher than among nonseropositive individuals ( $RR = 2.7, 95\%$ CI = 1.2 to
21	5.9, no. exposed cases not specified, but 360 of the total of 375 nasopharyngeal cancer
22	cases were EBV positive.) Results of stratified analysis suggested that the effect of
23	formaldehyde exposure was the same across age ranges and histologic subtype
24	(excluding squamous-cell type because sample size was too small for meaningful
25	analysis).

- 26 3.3.3 Other head and neck cancers
- 27 Section 3.3.3 reviews case-control studies that examined the association between
- 28 formaldehyde and head and neck cancer at sites including the oro- and/or hypopharynx
- 29 (OHPC) (Berrino et al. 2003, Laforest et al. 2000, Merletti et al. 1991, Vaughan et al.
- 30 1986a), the whole pharynx combined (Gustavsson et al. 1998, Tarvainen et al. 2008), the

oral cavity (Gustavsson *et al.* 1998, Merletti *et al.* 1991, Tarvainen *et al.* 2008), salivary
 glands (Wilson *et al.* 2004), and larynx (Berrino *et al.* 2003, Elci *et al.* 2003, Gustavsson

3 et al. 1998, Laforest et al. 2000, Wortley et al. 1992). Pharyngeal carcinomas can include

4 nasopharyngeal (see Section 3.3.2), oropharyngeal, and hypopharyngeal carcinomas. Six

5 studies were conducted in Europe (Merletti et al. 1991; Gustavsson et al. 1998; Laforest et

6 al.2000; Berrino et al.2003; Elci et al.2003, Tarvainen et al. 2008) and three in the

7 United States (Vaughan et al. 1986a, Wilson et al. 2004, Wortley et al. 1992). Most

8 studies evaluated more than one type of cancer. One study was described previously in

9 Section 3.3.1 (Vaughan *et al.* 1986a). In this section, studies are organized by tumor site.

10 3.3.3.1 Salivary gland: United States, Wilson et al. (2004)

11 Study Population. Wilson et al. (2004) reported on a case-control investigation of

12 occupational risk factors for salivary gland cancer mortality using mortality records

13 collected between 1984 and 1989 in 24 U.S. states. In this analysis, 2,505 cases aged 20

14 years or older were included whose death certificate listed cancer of the salivary gland as

15 the underlying cause of death (60% men, 7% black). Controls (N = 9,420) were randomly

16 selected from all deaths unrelated to infectious disease and frequency matched by age

17 (within 5 years), race, sex, and region (case to control ratio = 1:4).

18 *Exposure assessment*. Usual occupation and industry was obtained from death certificates

19 for 95% of white and 87% of black men, and for 45% of white and 31% of black women.

20 Jobs were coded according to the 1980 U.S. Census occupational classification scheme  $\$ 

21 and entered into a job-exposure matrix developed by the study industrial hygienist to

22 estimate the probability and intensity of exposure to several occupational substances

23 including formaldehyde. Subjects whose occupation was recorded as homemaker or

24 retired were excluded from the job-exposure matrix.

25 Statistical methods and results. Multiple logistic regression was used to calculate ORs

- 26 adjusted for age, marital status, and socio-economic status based on occupation. A
- 27 statistically significant exposure-response trend was observed for formaldehyde exposure
- probability combined with intensity among white men (P < 0.001) but not women:
- 29 Compared with unexposed subjects, the adjusted OR for white men with a mid-high
- 30 probability/low intensity of exposure was 2.4 (95% CI = 0.86 to 6.75, 6 exposed cases),

and 1.6 (1.30 to 2.00, 31 exposed cases) for mid-high probability/mid-high intensity. No
statistically significant ORs were observed for formaldehyde exposure and salivary gland
cancer among black subjects, though elevated ORs were observed among black women.
3.3.3.2 Oral cavity and oropharynx: Italy Merletti et al. (1991)
Population. All incident cases of oral (N = 74) and oropharyngeal carcinoma (N = 12)
diagnosed from July 1982 to December 1984 among male residents of Turin, Italy were

7 assembled for a population-based, case-control study to investigate whether occupational

8 factors have an etiologic role in these cancers (Merletti et al. 1991). Of 103 eligible cases,

9 86 (83%) agreed to participate. Of 689 eligible controls selected from a stratified random

10 sample of male Turin residents by age, 373 (55%) were enrolled.

11 *Exposure assessment.* Detailed occupational histories as well as history of smoking,

12 alcohol intake, and diet were obtained from standardized questionnaires conducted by

13 non-blinded, trained interviewers. For each job held since 1945 for at least six months,

subjects reported job title, activity of the plant, and type of production. The 1,150

15 reported jobs were classified by two industrial hygienists blinded to case status into 771

16 unique categories based on the International Standard Classification of Occupations of

17 the International Labor Office and the International Standard Industrial Classification. A

18 job-exposure matrix constructed by IARC for a study of laryngeal cancer was applied to

19 estimate the probability and intensity of exposure to 16 occupational substances including

20 formaldehyde and non-specific exposures (e.g., dust).

21 Results. Odds ratios for oral and oropharyngeal carcinoma combined were estimated 22 using unconditional logistic regression adjusting for age, education, birth place, smoking, 23 and alcohol consumption. Compared with subjects whose occupational exposure to 24 formaldehyde did not exceed that of the general population, the adjusted OR for subjects 25 with any excess exposure was 1.6 (95% CI = 0.9 to 2.8, 25 exposed cases) and the OR for 26 subjects with probable or definite exposure was 1.8 (95% CI = 0.6 to 5.5, 6 exposed)27 cases). The authors reported that inconsistent relationships were observed for duration of 28 exposure to formaldehyde, though effect estimates ranged from 1.4 to 2.1 (95% CIs not 29 reported). Separate results for oropharyngeal cancer (N = 12 cases) were not presented.

1 3.3.3.3 Oral cavity, tongue and pharynx: Finland, Tarvainen et al. 2008 2 Study population. The association between oral cavity, tongue, and pharyngeal cancers 3 and occupational exposures was investigated in a standardized incidence study by 4 Tarvainen et al. (2008), using all diagnosed cases identified among all Finnish men and 5 women, born between 1906 and 1945 and followed from 1971 to 1995, through the 6 Finnish Cancer Registry. A total of 46.8 million person-years were represented by the 7 cohort, and a total of 2,708 cases of oral cavity, tongue and pharyngeal cancers 8 (excluding nasopharyngeal cancers) were identified. 9 *Exposure assessment.* The occupation held the longest according to the 1970 census was 10 converted via a national job-exposure matrix to semi-quantitative (low, medium, and 11 high) estimates of cumulative exposure to 43 separate chemical agents.

12 Statistical methods and results. Standardized incidence ratios for combined oral, tongue,

13 and pharyngeal cancers were calculated based on national rates. Exposure to low,

14 medium, and high estimated cumulative levels of formaldehyde was associated with

15 statistically nonsignificant SIRs of 0.79 (95% CI = 0.6 to 1.03, 59 cases), 1.01 (95% CI =

16 0.43 to 1.98, 8 cases) and 0.73 (95% CI 0.27 to 1.59, 6 deaths), respectively.

17 3.3.3.4 Oro- and hypopharynx: Washington State, Vaughan et al. (1986a)

18 The association between oro- and hypopharyngeal cancer (OHPC) (N = 205) and

19 occupational formaldehyde exposure was also examined by Vaughan et al. (1986a) in the

20 population-based, case-control study (552 controls) in Washington state (see Section

21 3.2.1 for complete study description and results on sinonasal cancers; see section 3.2.2

22 for results on nasopharyngeal cancers). Approximately 72% of OHPC cases occurred

among subjects classified as unexposed. Odds ratios adjusted for age, sex, smoking, and

24 alcohol showed no association between maximum lifetime exposure to formaldehyde and

- 25 OHPC. Effect estimates for total number of years exposed and cumulative exposure
- 26 scores showed a modestly increased risk only for the longest exposure period or highest
- 27 cumulative exposure categories: OR = 1.3 (95% CI = 0.7 to 2.5, 26 exposed cases) for  $\geq$
- 28 10 years exposure, and OR = 1.5 (95% CI = 0.7 to 3.0, 21 exposed cases) for a
- 29 cumulative exposure score of  $\geq$  20. These estimates were higher when the analysis
- 30 excluded occupational data obtained from proxy interviews.

# 1 3.3.3.5 Hypopharynx and larynx: France, Laforest et al. (2000)

2 Study population. A hospital-based, case-control study was conducted in France to assess 3 possible associations between occupational exposures including formaldehyde and 4 histologically confirmed squamous-cell carcinomas of the hypopharynx and larynx 5 among men (Laforest et al. 2000). Cases were diagnosed at one of 15 French hospitals 6 between January 1989 and April 1991. Of 664 eligible living cases, 201 cases of 7 hypopharyngeal cancer and 296 cases of laryngeal cancer were included. Controls were 8 identified from the same medical catchment area as cases and were frequency matched to 9 cases by age and hospital. Controls were diagnosed between 1987 and 1991 with primary 10 cancers at other sites including colon/rectum, liver/gall bladder, pancreas, hematopoietic 11 system, bones/cartilage, skin, soft tissue, prostate/testis, bladder/urinary organs, 12 brain/nervous system, thyroid, and stomach. Of 355 eligible living controls, 296 (83%) 13 were enrolled.

14 Exposure assessment. Trained occupational physicians, who were not blinded to case

15 status, conducted interviews with subjects to collect information about demographic

16 characteristics, smoking and alcohol consumption, and lifetime occupational history. Jobs

17 were first coded by occupation and industry, and then occupational exposure to

18 formaldehyde and other agents and were evaluated using a job-exposure matrix. The

19 matrix estimated the probability and intensity of exposure for each job as well as lifetime

20 duration for each subject; subjects with an estimated probability of exposure to

21 formaldehyde less than 1% were considered unexposed. Three summary exposure indices

22 were constructed: maximum probability of exposure (3 levels), total duration of

exposure, and cumulative level of exposure (< 0.25 ppm, 0.25 to 1.00 ppm, > 1.00 ppm).

Statistical methods and results. Multivariate unconditional logistic regression was used to estimate ORs and 95% CIs adjusting for age, alcohol, and smoking. Other occupational exposures as well as education were considered as potential confounders. Subjects who were missing data on alcohol use or reported being non-drinkers (N = 33) were excluded for analysis. Further analyses were conducted excluding subjects with probability of exposure less than 10%, and excluding the 5, 10, and 15 years of exposure immediately preceding diagnosis to allow for a possible induction period. The adjusted (age, alcohol,

1	smoking, and exposure to coal dust and asbestos) OR for hypopharyngeal cancers for
2	men ever exposed to formaldehyde was 1.35 (95% $CI = 0.86$ to 2.14, 83 exposed cases).
3	This estimate was 1.74 (95% CI = 0.91 to 3.34, 41 exposed cases) after excluding
4	subjects with less than 10% probability of exposure. The OR comparing subjects with the
5	highest probability of exposure (> 50% probability) to those unexposed was 3.78 (95%
6	CI = 1.50 to 9.49, 26 exposed cases); increasing probability of exposure was significantly
7	associated with increasing risk of hypopharyngeal cancers ( $P_{\text{trend}} < 0.005$ ). Excluding
8	subjects with probability of exposure less than 10%, the OR for subjects with the highest
9	duration of exposure (> 20 years) was 2.70 (95% $CI = 1.08$ to 6.73, 16 exposed subjects).
10	The corresponding OR for subjects with the highest cumulative level of exposure was
11	1.92 (95% CI = 0.86 to 4.32, 25 exposed subjects). Evidence of a trend of increasing ORs
12	for hypopharyngeal cancers with increasing duration ( $P_{\text{trend}} < 0.04$ ) and cumulative level
13	of exposure ( $P_{\text{trend}} < 0.14$ ) to formaldehyde was observed.
14	Compared with unexposed subjects, the OR for laryngeal cancer among men ever
15	exposed to formal dehyde was $1.14$ (95% CI = 0.76 to 1.70, 102 exposed cases) after

exposed to formaldehyde was 1.14 (95% CI = 0.76 to 1.70, 102 exposed cases) after 15 16 adjustment for age, alcohol, smoking, and exposure to coal dust and asbestos. This 17 estimate did not change markedly after excluding subjects with probability of exposure 18 less than 10%. The authors noted that no indication of an exposure-response trend was 19 observed for any exposure index (data not presented). Among heavy drinkers (at least 5 20 glasses per day), the OR for laryngeal cancer associated with ever being exposed to 21 formaldehyde was 1.68 (95% CI = 0.97 to 2.89, no. of cases not specified). [An OR for 22 the association between alcohol consumption and laryngeal cancer independent of 23 formaldehyde exposure was not reported.] Elevated but statistically non-significant 24 associations were observed when cases were further stratified into laryngeal sub-sites. 25 The authors noted that introducing an induction time did not substantially change the 26 results for either hypopharyngeal cancer or laryngeal cancer (data not presented). 27 [Controls included subjects with primary cancers at sites that have suspected associations 28 with formaldehyde exposure (e.g., lymphohematopoietic malignancies). Such inclusion 29 could have biased the observed effect estimates toward the null.]

1 3.3.3.6 Hypopharynx and larynx: Europe, Berrino et al. (2003)

- 2 Study population. Berrino et al. (2003) used occupational data obtained from a previously
- 3 conducted case-control study by IARC of hypopharyngeal cancer and laryngeal
- 4 carcinoma to investigate the association between occupational exposure to formaldehyde
- 5 and cancer at these two sites. Cases of non-*in situ* cancer of the hypopharynx (N = 100)
- 6 and larynx (N = 213) were identified between 1979 and 1982 at six centers in four
- 7 southern European countries (France, Italy, Spain, and Switzerland). An age-stratified
- 8 random sample of controls (N = 819) was selected by each center.

9 *Exposure assessment*. Occupational histories and information on diet, alcohol, and

- 10 smoking were collected by interview in the hospital for cases and at home for controls.
- 11 Some interviews were conducted with next of kin (details not provided). The
- 12 occupational history questionnaire covered each job held at least one year after 1944 and
- 13 collected information about title, task, industry, calendar time of employment, and
- 14 potential exposure. A panel of occupational physicians, industrial hygienists, and
- 15 chemical engineers blinded to case status assessed the probability of exposure for each
- 16 job to 16 industrial chemicals including formaldehyde. A job-exposure matrix was then
- 17 created to estimate intensity and probability of exposure for each job as well as a
- 18 cumulative exposure index for each subject.

19 Statistical methods and results. Odds ratios and 95% confidence intervals were estimated

- 20 using unconditional logistic regression and adjusted for study center, age, smoking,
- 21 alcohol, socio-economic status, diet, and other occupational exposures. Results for
- 22 formaldehyde were presented from analyses restricted to subjects less than 55 years of
- 23 age in order to better estimate lifetime exposures, since occupational histories were only
- collected since 1945 (123 exposed cases and 196 exposed controls for hypopharyngeal
- and laryngeal carcinomas combined). No association between the probability of exposure
- 26 to formaldehyde and either hypopharyngeal or laryngeal cancer was observed.
- 27 Individuals with 10 to 19 years of exposure had an increased risk of laryngeal cancer (OR
- for 10 to 19 years = 2.2, 95% CI = 1.2 to 4.2, no. of exposed cases not reported), though a
- 29 clear exposure-response trend was not evident. [The ability to detect an effect was limited
- 30 by small numbers of exposed subjects and potential exposure misclassification.

- 1 Independent validations of the exposure classification used in this analysis found that
- 2 14% of jobs classified by the job-exposure matrix as unexposed were considered
- 3 definitely exposed according to the independent assessment.]
- 4 3.3.3.7 Larynx: Washington state, Wortley et al. (1992)

5 Study population. Incident cases of laryngeal cancer identified by a population-based 6 cancer registry in Seattle, Washington and diagnosed between September 1983 and 7 February 1987 among residents of three large counties in western Washington state aged 8 20 to 70 years were included in a population-based case-control study of occupational 9 risk factors for laryngeal cancer (Wortley *et al.* 1992). Of 291 eligible cases, 235 (81%) 10 participated in the study (79% males). Controls were identified by random-digit dialing 11 and frequency matched to cases by age and sex; the participation rate among eligible 12 controls was 8%, yielding 547 controls (65% males).

13 *Exposure assessment.* In-person interviews were conducted (7% of case interviews with 14 next-of-kin) to obtain information about lifetime occupational history, smoking, and 15 alcohol intake. Occupational questions related to job titles, tasks, and industry for each 16 job held at least six months; job title and industry were then coded according to the 1980 17 U.S. Census occupational codes. Exposure to six agents including formaldehyde was 18 assessed in greater detail by a panel of four industrial hygienists who constructed a job-19 exposure matrix for each agent; jobs were then classified into four levels of exposure 20 based on probability and intensity of exposure.

21 *Statistical methods and results*. Multivariate logistic regression was applied and a latency 22 effect was considered by excluding all exposures within 10 years of case diagnosis or 23 control selection. Ninety cases (90) (38%) and 154 controls (28%) were considered ever 24 exposed to formaldehyde. No statistically significant effect estimates were observed 25 between laryngeal cancer and exposure to formaldehyde estimated by peak exposure or 26 duration of exposure, adjusted for age, smoking, alcohol, and education. When low-level 27 exposures were excluded, the OR among workers with medium or high exposure for at 28 least 10 years duration compared with unexposed workers was 4.2 (95% CI = 0.9 to 19.4, 29 no. exposed cases not reported); the corresponding OR among workers with high 30 exposure was 4.3 (95% CI = 1.0 to 18.7). The authors noted that these estimates

1 increased slightly when the 10-year exposure lag was applied to account for a latency

2 period (data not presented).

3 3.3.3.8 *Larynx: Turkey, Elci* et al. (2003)

4 *Study population.* A hospital-based incident case-control study was conducted to

5 investigate occupational risk factors for laryngeal cancer among men in Turkey (Elci et

6 *al.* 2003). The case group included 951 confirmed cases of laryngeal cancer among men

7 presenting at an oncology treatment center at a hospital in Istanbul between 1979 and

8 1984. Controls (N = 1,519) were selected from hospital patients with other cancers

9 thought not to share similar etiologic factors with laryngeal cancer (including Hodgkin's

10 lymphoma, soft tissue sarcoma, and testicular cancer) and non-cancer diagnoses.

11 *Exposure assessment*. Upon admission to the hospital, all patients responded to a

12 questionnaire about occupational history, tobacco, and alcohol use; questionnaire data

13 was complete for 99% of cases and all controls. A job-exposure matrix was constructed

14 by an industrial hygienist blinded to case/control status and used to estimate for each

15 reported occupation and industry the probability and intensity of exposure to five

16 occupational substances, including formaldehyde.

17 Statistical methods and results. Unconditional logistic regression was applied to estimate

18 ORs adjusted by age, smoking, and alcohol use. No association between exposure to

19 formaldehyde and laryngeal cancer was observed by either probability or intensity of

20 exposure. The OR for laryngeal cancer among men considered ever exposed to

formaldehyde was 1.0 (95% CI = 0.8 to 1.3, 89 exposed cases).

22 3.3.3.9 Various head and neck cancers: Sweden, Gustavsson et al. (1998)

23 Study population. Occupational risk factors for squamous-cell carcinoma of the upper

24 gastrointestinal tract among men 40 to 70 years were investigated in an incident case-

control study in Sweden (Gustavsson *et al.* 1998). From weekly health care facility

26 reports and regional cancer registries, 605 cases of head and neck squamous-cell

27 carcinoma were identified. Ninety percent (90%) of cases (N = 545) were enrolled: 138

28 with pharyngeal cancer, 128 with oral cancer, 122 with esophageal cancer, and 157 with

29 laryngeal cancer. Controls (N = 756) were selected from the same study base by stratified

random sampling from population registries; 641 (85%) eligible controls were enrolled
 and frequency matched to cases by region and age.

3 *Exposure assessment.* Subjects were interviewed by one of two trained nurses about 4 lifestyle and environmental factors including oral hygiene, smoking, alcohol and snuff 5 use, and occupational history. Questions about occupational history covered all jobs ever 6 held for more than one year and included information about title, task, duration, industry, 7 and potential exposures. An industrial hygienist blinded to case/control status coded each 8 job according to the Swedish standard occupational classifications and then further 9 classified each occupation by probability and intensity of exposure to 17 specific agents 10 including formaldehyde (9.4% of controls were exposed to formaldehyde). For 11 formaldehyde, three primary measures of exposure were estimated: ever/never exposed, 12 duration of exposure, and cumulative exposure.

Statistical methods and results. Unconditional logistic regression was used to estimate
 ORs and 95% CIs. Formaldehyde effect estimates were adjusted for region, age, alcohol,

15 and smoking. Elevated estimates were observed for most cancer sites, though no

16 estimates achieved statistical significance. For cancers in all sites combined, the adjusted

17 OR comparing subjects ever exposed to formaldehyde to those unexposed was 1.42 (95%

18 CI = 0.94 to 2.15, 69 exposed cases). Adjusted odds ratios for individual sites were as

follows: 1.01 (95% CI = 0.49 to 2.07, 13 exposed cases) for pharyngeal cancer, 1.45

20 (95% CI = 0.83 to 2.51, 23 exposed cases) for laryngeal cancer, 1.90 (95% CI = 0.99 to

21 3.63, 19 exposed cases) for esophageal cancer, and 1.28 (95% CI = 0.64 to 2.54, 14

22 exposed cases) for cancers of the oral cavity. The authors reported that no dose-response

23 trend based on cumulative exposure or duration exposed was observed for any cancer site

24 (data not presented). [It is not clear whether other occupational exposures were

25 considered as confounders; reported effect estimates were not adjusted for other known

- 26 occupational exposures.]
- 27 3.3.4 Lung cancer

28 Section 3.3.4 reviews case-control studies that examined the association between

29 formaldehyde and lung cancer. These studies were conducted in Denmark (Jensen and

30 Anderson (1982), the United Kingdom (Coggon et al. 1984), Canada (Gérin et al. 1989),

1 the United States (Brownson *et al.* 1993), and Taiwan (Chen *et al.* 2008). Four nested

- 2 case-control studies of respiratory cancer are described in Sections 3.2.4 (Chiazze et al.
- 3 1997, Marsh et al. 2001), 3.2.5 (Partanen et al. 1990), 3.2.6 (Andjelkovich et al. 1994)
- 4 and 3.2.7 (Bond et al. 1986). Note that Coggon et al. (1984) included cancer of the
- 5 trachea in their analysis of respiratory cancers.
- 6 3.3.4.1 Denmark: Jensen and Anderson (1982)
- 7 Physicians: Denmark. Jensen and Andersen (1982) reported on a small case-control
- 8 series of 84 lung cancers (79 male, 5 female) among Danish physicians, identified from
- 9 the Danish Cancer Registry between 1943 and 1976 and 252 physician controls matched
- 10 on age, sex and survival (no details on the selection of controls or cases was given.). No
- 11 association with potential sources of formaldehyde exposure were reported. 8 cases and
- 12 23 controls had ever worked in anatomy, pathology or forensic medicine (RR = 1.0, 95%

13 
$$CI = 0.4$$
 to 2.4).

- 14 3.3.4.2 United Kingdom: Coggon et al. (1984)
- 15 *Study population.* Coggon *et al.* (1984) conducted a population-based, case-control study
- 16 using death certificates to obtain information about the occupations of all males under the
- 17 age of 40 years who died in England or Wales between 1975 and 1979 of epithelial
- 18 cancers of the lung, trachea, or bladder (see Section 3.2.6 for results on bladder cancer).
- 19 Cases of lung and tracheal carcinoma were combined and considered cancer of the
- 20 bronchus (N = 598). Controls (N = 1,180) that had died from any other cause during the
- same time period were individually matched to each case by sex, year of death (within 5
- 22 years), year of birth, and residential district. Of 598 cases, 582 (97%) were matched with
- 23 two controls; the remaining cases were matched with one control.
- 24 *Exposure assessment*. Occupations noted on the death certificates were coded using the
- 25 1970 Office of Population Census and Surveys Classification of Occupations scheme and
- 26 entered into a job-exposure matrix by a trained occupational hygienist. Using this matrix,
- each of the 233 uniquely classified occupations was then assigned an exposure score
- 28 (high/low/none) to nine known or suspected carcinogens, including formaldehyde.
- Among workers with carcinoma of the bronchus, 296 cases (50%) were considered
- 30 exposed to formaldehyde; 472 controls (40%) were considered exposed.

1 Statistical methods and results. Matched tabular analysis was used to calculate estimates 2 of the association between each carcinogen and carcinoma of the bronchus. For all 3 exposed occupations, the OR for formaldehyde was 1.5 (95% CI = 1.2 to 1.9, 296)4 exposed cases). Among occupations considered to have high exposure to formaldehyde, 5 the OR was 0.9 (95% CI = 0.6 to 1.4, 44 exposed cases). [The ability to detect an effect in 6 this study was limited by (1) the use of death certificates for occupational information, 7 thus limiting the construction of a complete job-exposure matrix and resulting in potential 8 non-differential exposure misclassification, (2) matching by pay class, which is likely to 9 be correlated with occupation, and (3) insufficient capture of long-term exposures and 10 insufficient follow-up to account for the relevant latency period of lung cancer, since 11 subjects in this study had died before 40 years of age.]

12 3.3.4.3 Canada: Gérin et al. (1989)

13 Study population. Gérin et al. (1989) investigated the association between exposure to 14 formaldehyde and subsequent risk of cancer at 14 primary sites of interest among males 15 aged 35 to 70 years, using data from a large multi-site case-control study in Montreal, 16 Canada of occupational exposures and cancer. Histologically confirmed primary incident 17 cases of cancer (N = 4,510) diagnosed between September 1979 and December 1985 18 were ascertained from all hospitals in the Montreal area. This analysis included 857 cases 19 of lung cancer (see Section 3.2.5 for results on lymphohematopoietic malignancies, and 20 Section 3.2.6 for results on other cancer sites). Sub-types of lung cancer were also 21 examined including oat-cell (N = 159) and squamous-cell cancers (N = 359), 22 adenocarcinomas (N = 162), and other histologic sub-types (N = 177). For each case 23 series, a cancer control group was selected from the case series that included patients 24 with tumors at any other site (some exceptions noted). In addition to the internal cancer 25 control series, 740 population-based controls frequency matched by age were selected 26 from electoral lists; 533 (72%) agreed to participate.

27 Exposure assessment. Trained interviewers collected information from each patient or

28 next-of-kin on demographic characteristics, medical history, diet, and a complete

- 29 occupational history including a semi-structured probing section designed to elicit
- 30 detailed descriptions of each job ever held in a working lifetime. Jobs were coded

according to standard Canadian classifications and then further classified by a team of
 chemists and hygienists by probability, frequency, and concentration of exposure to 300

- 3 occupational exposures including formaldehyde. Of 4,259 interviewed subjects, 971
- 4 (23%) subjects ever held at least one job classified as exposed to formaldehyde.

5 Statistical methods and results. Odds ratios and 95% CIs were estimated using logistic 6 regression. Both occupational and non-occupational factors were evaluated as potential 7 confounders using change-in-estimate methods whereby any factor that changes the 8 estimate of formaldehyde for the cancer site of interest by more than 10% is considered a 9 confounder. Models were further adjusted by five *a priori* variables including age, 10 ethnicity, income, smoking, and "dirtiness" (a semi-quantitative measure constructed by 11 the study chemists) of the jobs held. The OR for all lung cancer and any formaldehyde 12 exposure was 0.8 (95% CI = 0.6 to 1.0, 180 exposed cases) using the cancer control 13 series. Results using the population control series were not markedly different. [Some 14 controls had types of cancer potentially associated with formaldehyde; inclusion of these 15 controls could potentially attenuate true effects.] The OR for the highest exposure 16 category (i.e., greater than 10-years duration of exposure at high concentrations) was 1.5 17 (95% CI = 0.8 to 2.8, 24 exposed cases). In the analysis by histologic subtype, the largest 18 estimates in magnitude were observed for adenocarcinomas: the OR for subjects 19 classified into the highest exposure category was 2.3 (95% CI = 0.9 to 6.0, 7 exposed 20 cases) using the cancer control series.

21 3.3.4.4 *Missouri: Brownson* et al. (1993)

22 Study population. Brownson et al. (1993) conducted a population-based, case-control

- 23 study to investigate occupational risk factors for incident lung cancer among non-
- smoking women. Eligible cases included cases of primary lung cancer (N = 429)
- 25 identified by the Missouri Cancer Registry and diagnosed between 1986 and 1991 among
- 26 white women aged 30 to 84 years who were Missouri residents and either lifetime non-
- 27 smokers or ex-smokers who had stopped smoking at least 15 years prior to diagnosis or
- had smoked less than one pack-year. Controls (N = 1,021) were selected from state
- 29 driver's license files (for women less than 65 years of age) and from Medicare recipient
1 rosters (for women aged 65 or older); controls were frequency matched by age (case to

2 control ratio = 1:2).

*Exposure assessment*. In-person occupational history interviews were conducted with 429
cases (66% of eligible cases; 58% case interviews with next-of-kin) and 1,021 controls
(67% of eligible controls) to obtain information about job titles, calendar duration of
employment, and exposure to specific substances.

7 Statistical methods and results. Odds ratios were estimated using multivariate logistic

8 regression. All subjects who reported exposure to formaldehyde were also lifetime non-

9 smokers. The OR for lung cancer among all subjects ever exposed to formaldehyde was

10 0.9 (95% CI = 0.2 to 3.3, 3 exposed cases), adjusted for age and history of previous lung

11 disease. [Use of self-reported exposure to formaldehyde may have caused non-

12 differential exposure misclassification, which would likely bias observed ORs towards

13 the null.]

14 3.3.4.5 *Taiwan: Chen* et al. 2008.

15 *Study population.* Chen *et al.* (2008) conducted a hospital-based, case-control study of

16 147 incident cases of lung cancer and 400 controls from a population exposed to the

17 smoke from mosquito coils, which primarily contain pyrethrin insecticides, but also

18 release formaldehyde (which may form a reactive species of bischloromethyl ether) from

19 the active ingredient octachlorodipropyl, as well as dyes, oxidants, and other compounds.

20 Exposure assessment, statistical methods, and results. Frequency of exposure to

21 mosquito coils and other variables was ascertained by personal interview, and

22 unconditional logistic regression was used to calculate adjusted odds ratios. The authors

reported statistically significantly elevated ORs of 3.78 (95% CI = 1.55 to 6.90, 24 cases)

and 2.67 (95% CI = 1.60 to 4.50, 32 cases), adjusted for smoking and demographic

25 variables, in association with coil use more than 3 times per week or less than 3 times per

26 week, respectively, compared with those who did not use coils. [The independent effect

27 of formaldehyde exposure cannot be evaluated in this study.]

- 1 3.3.5 Lymphohematopoietic malignancies
- 2 Section 3.3.5 reviews case-control studies that examined the association between
- 3 formaldehyde and lymphohematopoietic malignancies (ICD codes 200-209) including
- 4 non-Hodgkin's and Hodgkin's lymphoma (Gérin et al. 1989, McDuffie et al. 2001,
- 5 Tatham et al. 1997, Wang et al. 2009), leukemia (Blair et al. 2001), multiple myeloma
- 6 (Boffetta *et al.* 1989, Heineman *et al.* 1992, Pottern *et al.* 1992) and myelodysplastic
- 7 syndrome (West et al. 1995). Two studies were conducted in Canada (Gérin et al. 1989,
- 8 McDuffie et al. 2001), three in Europe (Heineman et al. 1992, Pottern et al. 1992, West
- 9 et al. 1995), and four in the United States (Blair et al. 2001, Boffetta et al. 1989, Tatham
- 10 et al. 1997, Wang et al. 2009). Gérin et al. (1989) was described previously in Section
- 11 3.2.4. Two nested case-control studies of lymphohematopoietic malignancies are
- 12 described in Sections 3.2.5 (Partanen *et al.* 1993) and 3.2.7 (Ott *et al.* 1989).
- 13 3.3.5.1 Canada: Gérin et al. (1989)
- 14 Gérin *et al.* (1989) investigated the association between exposure to formaldehyde and
- 15 Hodgkin's (N = 53) and non-Hodgkin's lymphoma (N = 206) among males aged 35 to 70
- 16 years, using data from a large multi-site case-control study in Montreal, Canada (see
- 17 Section 3.2.4 for complete study description and results on cancer of the bronchus).
- 18 Controls consisted of various internal control groups selected from the case series, and
- 19 740 population controls. Using the cancer control series, the ORs (adjusted for age,
- 20 ethnicity, socioeconomic status, smoking, and "dirtiness" of jobs held) for non-Hodgkin's
- 21 and Hodgkin's lymphoma comparing ever exposed to never exposed was 0.9 (95% CI =
- 22 0.6 to 1.3, 47 exposed cases), and 0.5 (95% CI = 0.2 to 1.2, 8 exposed cases),
- 23 respectively. [Effect estimates did not change markedly using the population-based
- 24 control series.] Non-Hodgkin's lymphoma was further evaluated by exposure duration
- and concentration; effect estimates ranged from 0.7 to 1.3 (e.g., OR = 1.3, 95% CI = 0.7
- 26 to 2.4, for 15 cases exposed at low cumulative concentration for greater than 10 years).
- 27 3.3.5.2 United States: Boffetta et al. (1989)
- 28 Study population. A population-based nested case-control study of 282 deaths from
- 29 multiple myeloma observed in the second stage of the American Cancer Society's Cancer
- 30 Prevention prospective cohort study and matched with up to 4 within-cohort controls was

- 1 conducted by Boffetta et al. (1989). The association between multiple myeloma (MM),
- 2 occupational groups and selected exposures was examined, based on questionnaires
- 3 completed by enrollees and assignment of exposure status by the investigators.

4 Statistical methods and results. Using conditional logistic regression, a statistically

5 nonsignificant association between multiple myeloma and formaldehyde exposure was

6 observed (OR = 1.8, 95% CI = 0.6 to 5.7, 4 cases). [The likelihood of misclassification of

7 exposure in this study was high, however, and subjects assigned to the high-exposure

8 group had lower OR than those in the low exposure group. The power to detect effects of

9 given agents in this study was also limited.]

10 3.3.5.3 Denmark: Heineman et al. (1992) and Pottern et al. 1992

11 Study population. Heineman et al. (1992) and Pottern et al. (1992) conducted a

12 population-based, case-control study of the association between multiple myeloma

13 incidence in Danish men and women in relation to their occupation. The analysis of men

14 was conducted based on 1,098 incident cases for whom industrial occupational histories

15 could be constructed and diagnosed between 1970 and 1984. Cases were identified via

16 the Danish Cancer Registry and matched with age- and sex-matched controls. The

17 analysis of women was based on 363 cases and 1,517 controls diagnosed over the same

18 period who had a history of industrial employment and for whom exposure to one or

19 more of 47 chemical agents could be evaluated.

20 Exposure assessment. A job-exposure matrix was constructed by industrial hygienists

21 based on pension and tax records of employment history by industrial employment

22 history and most recent occupations. Among men. those recorded with more than 5 years

of employment (791 cases and 3,070 controls), potential exposure to one or more of 47

24 chemicals were evaluated. [The numbers of cases and controls for whom historical

25 industrial exposures could be established is not clearly stated.]

26 Statistical methods and results. Maximum likelihood odds ratios were calculated for each

27 occupation vs. all occupations combined. For analyses of specific exposures, comparison

- 28 between estimated exposed and never exposed subjects was conducted. Possible (144
- 29 cases) and probable (41 cases) exposure to formaldehyde was not associated with an

- 1 increased risk of multiple myeloma among men in this study (OR = 1.1, 95% CI = 0.7 to
- 2 1.6, 41 cases). Fifty-six (56) women with multiple myeloma were considered to have
- 3 possible exposure to formaldehyde and 4 probable exposure; in neither case were the
- 4 odds ratios significantly elevated in comparison with controls (ORs = 1.1, 95% CI = 0.8
- 5 to 1.6) and 1.6, 95% CI = 0.4 to 5.3), respectively.
- 6 3.3.5.4 United States: Tatham et al. (1997)

7 Study population. Occupational risk factors for subgroups of non-Hodgkin's lymphoma 8 were investigated in a population-based, case-control study of male cases born between 9 1929 and 1953, diagnosed between 1984 and 1988, and identified by population-based 10 cancer registries in Atlanta, Connecticut, Iowa, Kansas, Miami, San Francisco, Detroit, 11 and Seattle (Tatham et al. 1997). Only living cases were eligible, and diagnoses were 12 confirmed by a panel of pathologists. Living controls were identified using random-digit 13 dialing and frequency matched to cases by registry and date of birth (within 5 years). Of 14 2,354 identified cases and 1,910 controls, the final numbers of subjects available for 15 analysis were 1,048 cases (45%) and 1,659 controls (87%) after exclusions for a variety 16 of reasons including unconfirmed diagnosis and presence of comorbid medical 17 conditions. Three subgroups of non-Hodgkin's lymphoma were identified: small-cell 18 diffuse lymphoma (N = 185), follicular lymphoma (N = 268), and large-cell diffuse

19 lymphoma (N = 526).

20 Exposure assessment. All study subjects were interviewed by telephone to collect

21 information about demographic and lifestyle characteristics, medical and military

22 histories, and occupational history covering all jobs held for at least one year. The job

23 history included questions about job title, tasks, type of industry, and calendar duration as

24 well as information about exposure to specific substances including formaldehyde. Study

25 investigators classified exposure to formaldehyde and other substances using data from

26 the self-reported occupational histories.

27 Statistical methods and results. Conditional logistic regression was used to estimate ORs

and 95% CIs. Covariates considered potential confounders included age at diagnosis,

29 education, ethnicity, year of entry into the study, being Jewish, marital status, risk factors

30 for AIDS, military service, and smoking. Among all cases of non-Hodgkin's lymphoma

combined, 93 (8.9%) cases were exposed to formaldehyde; 130 (7.8%) controls were

- 2 considered exposed. The adjusted OR for all lymphomas combined associated with ever
- 3 being exposed to formaldehyde was 1.20 (95% CI = 0.86 to 1.50, 93 exposed cases). For
- 4 the specific subgroups, the corresponding ORs were 1.4 (95% CI = 0.87 to 2.40, 21
- 5 exposed cases) for small-cell diffuse lymphomas, 0.71 (95% CI = 0.41 to 1.20, 17
- 6 exposed cases) for follicular lymphomas, and 1.10 (95% CI = 0.79 to 1.70, 46 exposed)
- 7 cases) for large-cell diffuse lymphomas.

1

8 3.3.5.5 Canada: McDuffie et al. (2001)

9 Study population. A national multi-center population-based study of non-Hodgkin's 10 lymphoma incidence association with pesticide use among Canadian men was conducted 11 by McDuffie et al. (2001). Cases of NHL diagnosed between 1991 and 1994 and 12 identified via provincial cancer registries were eligible for the study; age-matched 13 controls were identified through health records, telephone directories and voter rolls from 14 the general population. Men who reported using pesticides for more than 10 hours per 15 year on a mailed questionnaire were selected for telephone interview to obtain detailed 16 pesticide exposure, demographic, and other risk factor data, together with a random 17 sample of 15% of other (mail) respondents. All cases and controls were alive at the time 18 of interview. The final analyses included 517 cases and 1,506 controls.

*Exposure assessment*. Exposure to specific pesticides, including both occupational and
 nonoccupational use, was ascertained by telephone questionnaire using a pre-designated
 list of pesticides.

22 Statistical methods and results. Conditional logistic regression was used to compute odds

ratios stratified by age and province of residence, and adjusted for medical and other

24 variables with significant associations in initial univariate analyses. The use of

- 25 formaldehyde-containing fungicides among 7 cases of NHL and 233 controls was not
- significantly associated with NHL (adjusted OR = 0.92, 95% CI = 0.37 to 2.29).
- 27 [Misclassification of exposure is likely in studies of this type; it is also not clear whether
- 28 past exposures were taken into account. In addition, no adjustments were made for co-
- 29 exposures, and few of the cases or controls were exposed to any given type of pesticide,
- 30 so that the power to detect effects is low.]

1 3.3.5.6 Connecticut, US: Wang et al. (2009)

Study population. Wang et al. (2009) conducted a population-based case-control study of
non-Hodgkin's lymphoma incidence among women residents aged 21 to 84 years old in
Connecticut, and solvent exposures. Seventy-two percent (72%) of the women (N = 601)
were available for in-person interviews and were included in the study, together with 71
controls identified through random-digit dialing (69% participation rate) or Medicare or
Medicare files (47% participation rate).

8 *Exposure assessment.* A job-exposure matrix developed by the National Cancer Institute 9 was used to construct exposure histories from occupation and industry histories provided 10 by respondents, who were assigned semi-quantitative estimates of solvent exposure by 11 intensity and probability (low, medium and high) according to combinations of industry 12 and occupation.

13 Statistical methods and results. Unconditional logistic regression models, adjusting for 14 age, family history of hematopoietic cancers, alcohol consumption, and race were used to 15 estimate odds ratios of the association between cumulative solvent exposures and risk of 16 NHL. (Adjustment for other variables including income, education, smoking, and 17 immune disease history did not affect observed associations and were excluded from final 18 models.) Polytomous models were used to evaluate the association between histological 19 subtypes of NHL and solvent exposure. Ever exposure was associated with a borderline 20 statistically significant increase in risk of NHL (OR = 1.3, 95% CI = 1.0 to 1.7, 203 21 cases; adjusted for age, family history of hematopoietic disease, race, and alcohol use). 22 However, results by level of intensity of estimated exposure and level of probability of 23 exposure were somewhat inconsistent: borderline statistically significant associations 24 were observed for low average intensity (OR = 1.4, 95% CI = 1.0 to 1.8, 129 exposed 25 cases) and low average probability (OR = 1.3, 95% CI = 1.0 to 1.7, 165 exposed cases) 26 but not medium or high intensities and probabilities ( $P_{\text{trend}} = 0.21$  and 0.11, respectively). 27 The risk of NHL appeared to be confined to large B-cell lymphomas, which were 28 associated with an OR of 1.9 (95% CI = 1.3 to 2.6, 80 exposed cases) among ever vs. 29 never exposed. A statistically significantly increased risk of this subtype was observed for formaldehyde exposure at low average intensity (OR = 2.1, 95% CI = 1.4 to 3.1, 54 30

1	exposed cases), but medium to high average intensity of exposure was associated with a
2	lower risk (OR = $1.5$ , $95\%$ CI = $0.9$ to $2.4$ , $26$ exposed cases). When exposure
3	probabilities were analyzed, a medium-high probability of formaldehyde exposure
4	yielded a risk of 2.6 (95% CI = 1.5 to 4.7, 20 exposed cases) for large B-cell lymphomas
5	( $P_{\text{trend}} < 0.01$ ). No association with follicular lymphoma, chronic lymphocytic
6	lymphoma/small lymphocytic lymphomas and formaldehyde were observed.
7	3.3.5.7 <i>Iowa,US: Blair</i> et al. (2001)
8	Study population. Blair et al. (2001) conducted a population-based, case-control study of
9	occupation and leukemia including all cases of histologically confirmed leukemia
10	diagnosed among white men at least 30 years of age identified from the Iowa State
11	Cancer Registry between 1981 and 1983 and all such cases from a surveillance network
12	of hospitals in Minnesota (97% coverage) between 1980 and 1982. Because the primary
13	purpose of the study was to evaluate agricultural risk factors, cases and controls residing
14	in the urban areas of Minneapolis, St. Paul, Duluth, and Rochester were excluded. Of 669
15	eligible cases, 578 (86%) participated in the study; interviews were conducted with 340
16	living cases and 238 surrogates for deceased or severely ill cases. Population-based
17	controls ( $N = 1,245$ ) were identified using random-digit dialing to obtain controls under
18	65 years of age (N = 474, 77% participation rate), from Health Care Financing
19	Administration records to obtain controls over 65 years of age ( $N = 519, 79\%$
20	participation rate), and from state death certificate records to obtain surrogate respondents
21	for deceased subjects ( $N = 550, 77\%$ participation rate). Controls were frequency
22	matched by 5-year age group, vital status at time of interview, and state of residence. Five
23	hundred thirteen (513) cases and 1,087 controls were used for analysis after excluding
24	subjects whose sole occupation was farming since the incidence of leukemia was
25	previously found to be significantly elevated among farmers in this study population.
26	Histologic subtypes included in this analysis were: chronic lymphocytic leukemia (N =
27	214), acute myeloid leukemia (N = 132), chronic myeloid leukemia (N = 46), acute
28	lymphocytic leukemia (N = 13), myelodysplasia (N = 58), and other miscellaneous
29	leukemia types (N = $50$ ).

1 *Exposure assessment.* Structured interviews were conducted between 1981 and 1984 to 2 collect information about occupational history for each job held for at least one year, 3 demographic characteristics, residential history, medical history and family history of 4 cancer, as well as smoking and alcohol use. The occupational history included questions 5 about job title, industry, and calendar duration of employment. A job-exposure matrix 6 was constructed for selected occupational exposures including formaldehyde, and 7 exposure assignment was made without knowledge of case status. Probability and 8 intensity of exposure were each classified into 4 scores (unexposed/low/moderate/high), 9 considering known changes in potential exposure probabilities by industry and calendar 10 decade.

11 Statistical methods and results. Unconditional logistic regression was used to estimate 12 ORs and 95% CIs for all leukemias and for individual histological subtypes, adjusting for 13 the matching factors as well as pesticide use, education, hair dye use, family history of 14 cancer, and smoking. Effect estimates for formaldehyde were generally close to the null 15 for all leukemias combined and by histologic subtype. Elevated effect estimates were 16 based on small sample sizes (e.g., the OR for chronic myeloid leukemia was 2.9 [95% CI 17 = 0.3 to 24.5, 1 exposed case]). [Small numbers of exposed cases and controls (e.g., 3) 18 highly exposed cases total and 9 highly exposed controls) limited the ability of this study 19 to detect an effect.]

20 3.3.5.8 United Kingdom: West et al. (1995)

21 Study population. West et al. (1995) conducted a population-based, case-control study of

22 incident cases of myelodysplastic syndrome (MDS) in residents over 15 years of age in

23 Southeast Wales, Wessex, and West Yorkshire to identify occupational and

- 24 environmental exposures potentially associated with myelodysplasia in the United
- 25 Kingdom. Of 635 eligible cases, 400 (63%) were available for analysis; 46% of the cases
- 26 were women. Non-cancer controls [approximately 400, actual no. not reported] were
- 27 selected from hospitals and outpatient clinics and individually matched to cases by age
- 28 (within 3 years), sex, area of residence, hospital, and year of diagnosis (within 2 years).
- 29 *Exposure assessment*. Lifetime exposure to over 70 potential risk factors for MDS
- 30 including formaldehyde was estimated using in-depth interviews that probed subjects

- 1 about duration and intensity of exposure from jobs held six months or more, relevant
- 2 hobbies, and medical therapies. Occupational exposure was estimated in consultation
- 3 with industrial chemists and occupational hygienists using the self-reported job histories
- 4 and then categorized by duration and intensity (low/medium/high).

5 *Statistical methods and results.* Odds ratios were obtained using matched pair analysis.

- 6 Confidence intervals were only reported if the lower 95% limit was greater than 0.80.
- 7 The ORs for formaldehyde were 1.17 (15 exposed cases, 13 exposed controls) for
- 8 subjects with at least 10 hours of lifetime exposure at any intensity, 2.33 (no. of exposed

9 cases and controls not reported) for subjects with at least 50 hours of lifetime exposure at

10 medium or high intensity, and 2.00 for subjects with at least 2,500 hours of lifetime

- 11 exposure at medium or high intensity.
- 12 3.3.6 Cancers at other sites
- 13 Section 3.2.6 reviews seven case-control studies that examined the association between
- 14 formaldehyde and several other tumor sites not reviewed in previous sections. Gérin *et al.*
- 15 (1989) (described previously in Section 3.3.4) reported results for various cancers. Tumor
- 16 sites examined in other investigations include bladder (Coggon *et al.* 1984, Siemiatycki *et*

17 al. 1994), breast (Cantor et al. 1995), pancreas (Kernan et al. 1999), rectum (Dumas et al.

- 18 2000), and eye (Holly *et al.* 1996). The studies in this section are organized by site.
- 19 3.3.6.1 Multiple tissue sites: Canada, Gérin et al. (1989)
- 20 Gérin *et al.* (1989) evaluated potential associations between occupational exposure
- among men to formaldehyde and cancers of the esophagus (N = 107), stomach (N = 250),
- 22 colorectum (N = 787), liver (N = 50), pancreas (N = 117), prostate (N = 452), bladder (N
- 23 = 486), kidney (N = 181), and melanoma of the skin (N = 121) in a large multi-site case-
- 24 control study in Montreal (see Section 3.2.4 for complete study description and results for
- 25 respiratory cancer; see Section 3.2.5 for results for lymphohematopoietic malignancies).
- 26 Controls consisted of various internal control groups selected from the case series and
- 27 740 population controls. No elevated ORs were observed for any of these cancers.

### 1 3.3.6.2 Bladder cancer: United Kingdom, Coggon et al. (1984)

- 2 Coggon et al. (1984) used death certificates in this population-based, case-control study
- 3 to obtain information about the occupations of all males under the age of 40 years who
- 4 died in England or Wales during 1975 to 1979 of epithelial bladder cancer (see Section
- 5 3.2.4 for complete study description and results for cancer of the bronchus). Two hundred
- 6 ninety-one (291) cases and 578 controls were included in the analysis. Exposure to
- 7 formaldehyde was determined using a job-exposure matrix. Among subjects with bladder
- 8 cancer, 132 cases (45%) were considered exposed to formaldehyde; 472 controls (40%)
- 9 were considered exposed. For all exposed occupations, the OR for formaldehyde was 1.0
- 10 (95% CI = 0.7 to 1.3, 132 exposed cases). Among occupations considered to have high
- 11 exposure to formaldehyde, the OR increased in magnitude to 1.5 (95% CI = 0.9 to 2.5, 30 cm)
- 12 exposed cases).
- 13 3.3.6.3 Bladder cancer: Canada, Siemiatycki et al. (1994)
- 14 Siemiatycki et al. (1994) investigated the association between exposure to formaldehyde
- 15 and bladder cancer using data from the large multi-site case-control study in Montreal,
- 16 Canada studied by Gérin et al. (1989) (see Section 3.2.4 for complete study description).
- 17 Included in this analysis were 484 men (ages 35 to 70 years) with primary, incident,
- 18 histologically confirmed bladder cancer (575 eligible cases, 84% participation rate). From
- 19 the parent study, 1,879 controls with cancer at other sites (excluding lung and kidney)
- 20 and 533 community controls (72% participation rate) were selected; control groups were
- 21 pooled for analysis. Adjusting for age, ethnicity, socioeconomic status, smoking, coffee
- 22 consumption, and interview type (self/proxy), the OR for bladder cancer was 1.2 (95% CI
- 23 = 0.9 to 1.6, 67 exposed cases) among men with non-substantial exposure to
- formaldehyde and 1.2 (95% CI = 0.7 to 2.0, 17 exposed cases) among men with
- 25 substantial exposure. Adjusting for additional exposure to several occupational
- 26 substances reduced effect estimates for men considered to have substantial formaldehyde
- exposure (OR = 0.9, 95% CI = 0.5 to 1.7), but did not alter the estimate for
- 28 nonsubstantial exposure.

1 3.3.6.4 Breast cancer: United States, Cantor et al. (1995)

2 Study population. A database of mortality records from 1984 to 1989 in 24 states in the 3 United States was assembled for a series of case-control studies designed to investigate 4 associations between occupational factors and cancer mortality. Cantor et al. (1995) 5 reported on their investigation of occupational risk factors for breast cancer mortality 6 among women. For this analysis, cases (N = 59,515) included white and black women 7 (10% black) whose death certificate listed breast cancer as the underlying cause of death. 8 Controls were randomly selected from all non-cancer deaths and frequency matched by 9 age (within 5 years) and race (case to control ratio = 1:4).

10 *Exposure assessment.* Usual occupation and industry were obtained from death

11 certificates and coded according to the 1980 U.S. Census occupational classification

12 scheme. Homemakers were excluded, leaving 29,387 white and 4,112 black breast cancer

13 cases, and 102,955 white and 14,839 black controls. The remaining occupational and

14 industry codes were then entered into a job-exposure matrix to estimate the probability

15 and level of exposure to 31 occupational exposures, including formaldehyde.

16 Statistical methods and results. Odds ratios were stratified by race and adjusted for age at

17 death and socio-economic status (based on occupation). The risk estimate for breast

18 cancer was elevated among black women with the highest category of exposure

- 19 probability (OR = 1.45, 95% CI 1.2 to 1.7, 311 exposed cases) and with the highest
- 20 exposure level (OR = 1.26, 95% CI = 1.0 to 1.5, 192 exposed cases). However, these
- trends were not observed among white women: ORs ranged from 0.93 to 1.19 (e.g., 1.19,
- 22 95% CI = 1.1 to 1.3 for 1,815 cases exposed at the highest level). Further analysis

23 excluded women considered to have a low probability of exposure. Among white women,

- 24 the ORs were 1.14 (P < 0.05), 0.93, and 1.20 (P < 0.05) for low, moderate, and high
- 25 intensity of exposure, respectively; among black women, the corresponding ORs were
- 26 1.38 (P < .05), 1.30 (P < 0.05), and 1.36 (P < 0.05). Confidence intervals were not
- 27 reported.
- 28 3.3.6.5 Pancreatic cancer: United States, Kernan et al. (1999)
- 29 Study population. Kernan et al. (1999) reported on a case-control investigation of
- 30 occupational risk factors for pancreatic cancer mortality using the mortality records

1 collected between 1984 and 1993 in 24 U.S. states (Cantor *et al.* 1995, reviewed in this

2 section, also used this database, though the study period was earlier). In this analysis,

3 63,097 cases were included whose death certificate listed pancreatic cancer as the

4 underlying cause of death. Controls (N = 252,368) were randomly selected from all non-

5 cancer deaths (excluding pancreatitis and other pancreatic diseases) and frequency

6 matched by age (within 5 years), race, sex, and state (case to control ratio = 1:4).

7 *Exposure assessment.* Usual occupation and industry were obtained from death

8 certificates, coded according to the 1980 U.S. Census occupational classification scheme,

9 and entered into a job-exposure matrix developed by industrial hygienists to estimate the

10 probability and intensity of exposure to formaldehyde, 11 chlorinated hydrocarbons, and

11 2 groups of solvents. Forty-eight percent (48%) of male cases (N = 30,389) and 51% of

12 female cases (N = 31,962) were considered exposed to formaldehyde.

13 Statistical methods and results. Logistic regression was applied to estimate ORs and 95% 14 CIs, stratified by race (black/white) and sex and adjusted for age at death, metropolitan 15 status, region of residence, and marital status. Analysis by exposure intensity yielded 16 ORs ranging from 1.0 to 1.4 for each race-sex combination, with some estimates 17 achieving statistical significance. [The large number of exposed cases in this study 18 increased the power to detect an effect.] Analysis by exposure probability yielded ORs 19 ranging from 0.8 to 1.5; again, some estimates were statistically significant. Analysis by 20 exposure intensity and probability combined showed that among the entire study sample, 21 the OR for those with both high exposure intensity and high exposure probability was 1.4 22 (95% CI = 1.0 to 1.8, 56 exposed cases). Among all subjects with high exposure 23 probability, the ORs were 2.8 (95% CI = 0.7 to 1.8, 3 exposed cases) for those with low 24 exposure intensity, and 1.4 (95% CI = 1.2 to 1.6, 546 exposed cases) for those with 25 medium intensity. Among all subjects with high exposure intensity, the ORs were 1.0 26 (95% CI = 0.9 to 1.3, 171 exposed cases) for those with low exposure probability and 1.2 27 (95% CI = 0.8 to 1.6, 47 exposed cases) for those with medium probability. Though an 28 exposure-response relationship was not observed with intensity of exposure, exposure-29 response relationships by probability of exposure were consistent for each level of 30 exposure intensity.

- 1 3.3.6.6 Rectal cancer: Canada, Dumas et al. (2000)
- 2 Study population. Dumas et al. (2000) evaluated the association between exposure to
- 3 formaldehyde and incident cases of rectal cancer among males aged 35 to 70 years, using
- 4 data from the large multi-site case-control study in Montreal, Canada studied by Gérin et
- 5 *al.* (1989) (see Section 3.2.4 for complete study description and exposure assessment).
- 6 For this analysis, 257 cases of primary rectal cancer (304 eligible cases; 85%
- 7 participation rate), 1,295 cancer controls (excluding lung and cancers at intestinal sites),
- 8 and 533 community controls (72% participation rate) were enrolled.
- 9 Statistical methods and results. Odds ratios were adjusted for age, education, interview
- 10 status (self/proxy), smoking, beer consumption, and body mass index, but not other
- 11 occupational exposures. Results were presented using the cancer control series as the
- 12 referent group. Among men considered to have any occupational exposure to
- 13 formaldehyde, the OR for rectal cancer was 1.2 (95% CI = 0.8 to 1.9, 36 exposed cases).
- 14 Among men with substantial exposure, the OR increased to 2.4 (95% CI = 1.2 to 4.7, 13)
- 15 exposed cases). The authors noted that the overall exposure-response pattern reflected an
- 16 increase in risk with increasing duration and concentration of exposure (data not shown).
- 17 [Use of a control group including subjects with cancers that other studies have suggested
- 18 are potentially associated with formaldehyde exposure (such as esophageal carcinoma,
- 19 bladder cancer, and lymphomas) may have attenuated the observed effect estimate.]
- 20 [Case reports of a possible association between prostate, rectal, or endometrial cancer and
- 21 topical application of formalin were investigated by Stern and Steinhagen (2007).
- 22 Patients receiving radiation therapy for prostate, rectal, or endometrial cancer developed
- 23 hemorraghic radiation proctitis and received 4% topical formalin solution as a treatment.
- 24 Two patients subsequently developed anorectal cancer. It is not possible to distinguish a
- 25 potential effect of formalin from the effects of radiation or other treatment of the primary
- 26 cancer, however.]
- 27 3.3.6.7 Uveal cancer: United States, Holly et al. (1996)
- 28 Study population. Holly et al. (1996) conducted a case-control study to evaluate whether
- 29 certain occupational exposures were associated with incident cases of uveal cancer (also
- 30 known as intraocular melanoma) among white males aged 20 to 74 years living in the

western United States. The case group (N = 121, 95% participation rate) comprised all
histologically confirmed cases of uveal carcinoma either diagnosed or treated between
January 1978 and February 1987 at the Ocular Oncology Unit of the University of San
Francisco. For each case, two controls were selected using random-digit dialing and
individually matched by area of residence and age (within 5 years); 447 controls were
enrolled (77% participation rate).

*Exposure assessment.* Telephone interviews were conducted to elicit information about demographic, medical, and phenotypic characteristics (i.e., eye color), occupational history and exposure to chemicals, and history of smoking, diet, residence, and sun exposure. Exposure to chemicals of interest including formaldehyde was determined by asking each participant whether they had ever worked with or been regularly exposed (at least three hours per week for at least six months) to each chemical at a job or while engaging in hobbies, recreational activities, or home maintenance.

14 Statistical methods and results. Odds ratios were estimated using unconditional logistic

15 regression adjusting for age, eye characteristics, and response type to sun exposure. The

16 OR for uveal carcinoma among men who reported ever being exposed to formaldehyde

17 either occupationally or recreationally was 2.9 (95% CI = 1.2 to 7.0, 13 exposed cases).

18 [Results of this study may be affected by recall bias since exposure assessment was based

- 19 entirely on a subject's personal recollection of formaldehyde exposure.]
- 20 3.3.6.8 Thyroid cancer: China, Wong et al. 2006

21 Study population. Wong et al. (2006) conducted a nested case-cohort study of thyroid

22 cancer among a cohort of 267,400 female textile workers in Shanghai, China, who had

23 been followed for cancer incidence from 1989 to 1998. One hundred thirty (130) incident

- 24 thyroid cases and 3,187 non-case controls randomly selected from the cohort of all
- 25 eligible textile workers and matched by year of birth in five-year strata were identified.
- 26 *Exposure assessment.* Historical exposures were estimated by industrial hygienists using
- 27 a job-exposure matrix constructed from individual job histories and production process
- 28 data.

Statistical methods and results. The stratified analysis was conducted using a weighting

2 scheme for the stratified case-cohort design. Age-adjusted hazard ratios (HR) were

3 calculated using Cox proportional hazards methods with robust variance estimation. The

4 HR for only 2 cases of thyroid cancer were considered to have exposure to formaldehyde

5 compared with 11 controls; the HR was 8.33 (95% CI = 1.16 to 60.0, 2 exposed cases),

 $6 \quad \text{with} > 10 \text{ years of exposure.}$ 

1

# 7 **3.4** Summary by tumor site

8 This section summarizes the findings for the cohort and case-control studies for each of 9 the major cancer sites. A number of the cohort studies, the majority of which have 10 studied workers in a variety of industries, relied on external (SMR and PMR) analyses; 11 relatively few conducted internal analyses of exposed and unexposed workers. Few 12 studies have either sufficient numbers of exposed individuals to enable exposure-13 response relationships to be assessed and have quantitative exposure measurements on 14 which to base the assignment of exposure categories. Since some of the tumor types 15 potentially related to formaldehyde exposure are rare (e.g. sinonasal cancers, 16 nasopharyngeal cancers) most of the cohort studies have limited statistical power to 17 detect statistically significant increases in risk in association with exposure to 18 formaldehyde, and the case-control studies of these and other endpoints often lack 19 adequate data on exposure to formaldehyde. Three cohort studies were available that had 20 relatively large numbers of formaldehyde exposed workers: (1) the NCI cohort of mixed 21 industry workers (Hauptmann et al. 2003, 2004, Beane Freeman et al. 2009), (2) the 22 cohort of British chemical workers (Coggen et al. 2003), and (3) the NIOSH cohort of 23 garment workers (Pinkerton et al. 2004). The NCI mixed industry combined cohort is the 24 only cohort study to date in which detailed exposure-response relationships were 25 examined according to peak, average, duration, and cumulative exposure. The other large 26 cohort study, of British chemical workers, also examined exposure-response relationships 27 by level, duration of exposure, and time since first exposure, in external SMR 28 comparisons for selected cancer sites. The NIOSH cohort of garment workers evaluated 29 mortality for selected cancer sites by duration of exposure, time since first exposure, and 30 time of first exposure (exposure was higher for earlier time periods). The other cohorts 31 (both industrial and professional health workers) were smaller, and in general only

reported mortality for ever exposed. [Note that not all cohort studies reported findings for
each cancer site. Where findings were reported but no deaths or cases were observed, as
specifically noted by the authors, the annotation "0 deaths" is used in the accompanying
tables. Studies in which no findings for a given site were specifically reported are noted
in the footnotes for that table.]

## 6 3.4.1 Cancers of the paranasal sinuses and nasal cavity

7 Sinonasal carcinoma is a rare cancer (the annual incidence is approximately 1 case per 8 100,000 in most countries), which limits the ability of even large occupational cohort 9 studies to achieve enough statistical power to detect significant associations. Further, 10 sinonasal carcinoma is thought to have a long latency period (at least 10 years, with some 11 estimates as high as 40 years), meaning that study designs must have a long enough 12 follow-up to capture exposed cases. Approximately 70% to 80% of primary sinonasal 13 carcinoma occurs in the paranasal sinuses rather than the nasal cavity, but most of the 14 available studies do not distinguish between sites when identifying cases of sinonasal 15 cancers [Hauptmann et al. (2004) is one exception].

16 The relationship between sinonasal cancers and occupational exposure to formaldehyde 17 has been investigated in cohort, nested case-control and population-based case-control 18 studies. The key findings are summarized in Table 3-3a and b. (See Section 3.1 for a 19 description of sinonasal cancers, and Section 3.3.1 for a detailed summary of case-control 20 studies that investigated sinonasal cancers.) The majority of cohort studies have low 21 statistical power to detect sinonasal cancers.

# 22 3.4.1.1 Cohort studies

23 Increases in the risk of sinonasal cancers were reported in two cohort studies of

24 formaldehyde-exposed workers: (1) a statistically significant increased incidence of

25 sinonasal cancers was observed among male Danish workers exposed to formaldehyde

- 26 (SPICR = 2.3, 95% CI = 1.3 to 4.0, 13 exposed cases and SPICR = 3.0, 95% CI = 1.4 to
- 27 5.7, 9 exposed cases for exposed male workers without exposure to wood dust); risks,
- 28 although not statistically significant, were also increased among women (SPICR = 2.4,
- 29 95% CI = 0.6–6.0; 4 exposed cases) (Hansen and Olsen 1995, 1996), and (2) a non-
- 30 significant increased risk in sinonasal cancer mortality among formaldehyde exposed

1 workers was observed in the NCI cohort (SMR = 1.19, 95% CI = 0.38 to 3.68, 3 deaths) 2 (Hauptmann *et al.* 2004). In the latter study, statistically nonsignificant elevated relative 3 risks were observed for some categories of average, peak and cumulative exposure; 4 [however, the small number of exposed cases limits the ability to evaluate exposure-5 response relationships]. One death from squamous-cell sinonasal cancer was reported 6 among formaldehyde-exposed workers in an industrial cohort study of tannery workers 7 by Stern et al. (1987) [SMR or expected numbers of cases not reported]. No association 8 with formaldehyde exposure was found in a standardized mortality analysis among 9 British chemical workers (Coggon et al. 2003), which was one of the larger cohort 10 studies. No cases of sinonasal cancers were identified in the NIOSH cohort (Pinkerton et 11 al. 2004) or in the very small cohort of Dell and Teta (1995). [No findings were 12 specifically reported for this site by Andjelkovich et al. (1995), Bertazzi et al. (1986), 13 Edling et al. (1987b), Stellman et al. (1998), and Hall et al. (1991)] Among the studies of 14 health professionals, embalmers, anatomists, and pathologists, no cases of sinonasal 15 cancers were observed (Hayes et al. (1990), Levine et al. (1984), Stroup et al. (1986), 16 and Walrath and Fraumeni (1983, 1984). [However, these were small cohorts with 17 limited power to detect rare cancers].

#### 18 3.4.1.2 Case-control studies

19 Six case control studies on sinonasal cancers were identified. Four of these studies 20 reported increased sinonasal cancer risk among formaldehyde-exposed workers (or 21 subsets of workers). Luce et al. (1993a) evaluated exposure to 14 substances including 22 formaldehyde in a case-control study of 207 male cases (75 adenocarcinoma, 82 23 squamous-cell carcinoma and 25 other histological types). Among males with probable 24 exposure to formaldehyde, risks increased with increasing exposure duration and 25 cumulative exposure. A substantial proportion of cases were exposed to both 26 formaldehyde and wood dust. The authors noted a statistically non-significant elevated 27 risk of formaldehyde exposure for adenocarcinoma (OR = 8.1, 95% CI = 0.9 to 72.9, 4 28 exposed cases with low or no wood dust exposure) but a statistically significant highly 29 elevated risk when both formaldehyde and wood dust exposure were present (OR = 692, 30 95% CI = 91.9 to 5,210, 71 exposed cases). Among cases of the squamous-cell type, 31 which were adjusted for wood dust, glue, and adhesive exposure, no consistent pattern of

1 risk with year of first exposure, duration of exposure, cumulative exposure, or age at first 2 exposure was observed although a statistically non-significant elevated risk was observed 3 among cases. Adjustment for smoking did not alter effect estimates in this study. 4 Hayes et al. (1986) reported elevated risks for all sinonasal cancer and high formaldehyde 5 exposure among subjects unlikely to be exposed to wood dust, although the risk estimates 6 varied somewhat between two independent industrial hygienists' assessments (RR = 3.0, 7 90% CI = 1.0 to 8.7, 7 exposed cases vs. RR = 2.1, 90% CI = 1.1 to 4.1, 17 cases). Most 8 of the cases were squamous-cell carcinomas, and similar risk estimates were reported for 9 these histological types of cancer (see Table 3-3b). Olsen and colleagues (1994, 1996) 10 found elevated risks for adenocarcinomas (RR = 2.2, 95% CI = 0.7 to 7.2, 17 ever-11 exposed cases), squamous-cell carcinomas (RR = 2.3, 95% CI = 0.9 to 5.8, 13 ever-12 exposed cases), and all sinonasal cancers (RR = 1.6, 95% CI NR, P > 0.05). A 13 significantly increased risk of all sinonasal cancer was observed among cases with 14 "certain exposure" to formaldehyde (RR = 2.8, 95% CI = 1.8 to 4.3, 33 deaths). When 15 only those cases with no wood dust exposure were considered, the observed risk for

16 squamous-cell carcinomas, and all sinonasal cancers was not altered, but a statistically

17 significant increase in the risk of formaldehyde exposure was observed among

18 adenocarcinoma cases (RR = 7.0, 95% CI = 1.1 to 43.9) based on only one exposed case,

19 however. Among all cases of sinonasal cancer cases with both wood dust and

formaldehyde exposure, the RR was 3.5 (95% CI = 2.2 to 5.6, 28 exposed cases).

21 [Known risk factors for sinonasal cancers include the human carcinogens nickel dust 22 (NTP 2005a) and wood dust, particularly in the latter case, for adenocarcinomas (IARC 23 1995, NTP 2005a). In some studies, e.g., including workers in the woodworking and 24 lamination industries, there may be a high degree of colinearity between formaldehyde 25 and wood dust exposure (for example, 97% of subjects considered to be probably or 26 definitely exposed to formaldehyde were also jointly exposed to wood dusts in a case-27 control study by Luce *et al.* [1993a], which could result in residual confounding.) Effect 28 modification by wood dust has also been observed, whereby concurrent exposure to 29 wood dust increases the independent risk of sinonasal cancers associated with exposure to 30 formaldehyde or wood dusts alone (Olsen et al. 1984).]

#### 1 3.4.1.3 Pooled and meta-analyses

2 A pooled analysis (Luce et al. 2002) combining 12 case-control studies from seven 3 countries was conducted to further evaluate the relationship between sinonasal cancers 4 and occupational exposure to formaldehyde. The studies were selected on the basis of 5 availability of information on histologic type, age, sex, smoking, and occupational 6 histories. They differed according to the source and vital status of cases and controls as 7 well as the method of interview. Exposures were independently assessed for each study 8 by the authors of the pooled analysis using a job-exposure matrix designed specifically 9 for the analysis, and industrial hygiene data were used to determine semi-quantitative 10 exposure indices (only 3 of the 12 studies had originally conducted exposure assessments 11 for formaldehyde). Logistic regression was applied to estimate ORs adjusted for age, 12 study, and additional occupational factors that were found to be confounders (smoking 13 was not found to be a confounder). Only 11 cases exposed to formaldehyde were 14 estimated to have never been exposed to wood dust. Among men, the ORs for 15 adenocarcinoma sinonasal cancers by cumulative exposure to formaldehyde (adjusted for 16 wood dust exposure) were 0.7 (95% CI = 0.3 to 1.9, 6 pooled exposed cases) for low 17 exposure, 2.4 (95% CI = 1.3 to 4.5, 31 pooled exposed cases) for medium exposure, and 18 3.0 (95% CI = 1.5 to 5.7, 91 pooled exposed cases) for high exposure. The estimates for 19 squamous-cell sinonasal cancers were 1.2 (95% CI = 0.8 to 1.8, 43 pooled exposed 20 cases), 1.1 (95% CI = 0.8 to 1.6, 40 pooled exposed cases), and 1.2 (95% CI = 0.8 to 1.8, 21 30 pooled exposed cases), respectively. Effect estimates among women were generally 22 higher. To investigate the potential for residual confounding by wood dust, the authors 23 repeated the analyses for adenocarcinoma including only subjects who had never been 24 exposed to wood or leather dusts; effect estimates were reduced though still elevated (OR 25 for high cumulative exposure = 1.9, 95% CI = 0.5 to 6.7).

Bosetti *et al.* (2008) conducted a pooled analysis of occupational cohort mortality studies
of formaldehyde exposure which included sinonasal cancers, and reported a
nonsignificantly elevated estimated RR (using weighted average SMRs) of 1.01 (95% CI
= 0.33 to 2.35, 5 deaths) among 8 cohorts of industrial workers (no deaths were reported
among 5 cohorts of medical workers).

1 Collins et al. (1997) conducted a meta-analysis to evaluate the association between 2 formaldehyde exposure and upper respiratory cancers, including sinonasal cancers. Nine 3 cohort and 11 case-control mortality studies that reported findings on sinonasal cancers 4 and in which formaldehyde exposure was analyzed separately were included. A total of 5 933 observed vs. 807.7 deaths were included. The estimated meta relative risk (mRR) for 6 the 9 cohort studies was 0.3 (95% CI = 0.9, 3 deaths); each of the 3 deaths occurred in the 7 3 industrial cohorts (with none reported in 6 other cohorts) and yielded a mRR of 0.6 8 (95% CI = 0.1 to 1.7). Among the 11 case-control studies, the estimated mRR was 1.89 (95% CI = 1.4 to 2.3, 933 deaths); there was substantial variation between the 5 U.S. 10 studies (mRR = 1.0 to 1.5, 351 deaths) and the 6 European studies (mRR = 2.9, 95% CI = 11 2.2 to to 4.0, 582 deaths), which the authors suggested might be due in part to wood dust

12 exposure in some of the latter studies.

Reference	Study population and follow up	Risk estima number of deaths	ate, 95% CI; observed cases or	Comments
Coggon <i>et al.</i> 2003 (update of Acheson <i>et al.</i> 1984)	British Chemical Workers Study, UK N = 14,014 1941–2000	SMR All High exp.	0.87 (0.11–3.34); 2 0 (0–4.66); 0	
Dell and Teta 1995	Workers employed at a Union Carbide plastics manufacturing plant in New Jersey, USA 111 formaldehyde exposed workers 1946–1988	NR, 0 deaths		Small numbers of formaldehyde exposed workers
Hansen and Olsen 1995, 1996	Denmark N = 2,041 men, 1,263 women 1970–84	SPICR Men Women No exposure Men Women	2.3 (1.3–4.0); 13 2.4 (0.6–6.0); 4 to wood dust 3.0 (1.4–5.7); 9 NR	SPICR adjusted for age and calendar time
Hauptmann <i>et al.</i> 2004 NCI combined cohort Marsh <i>et al.</i> 2007a Wallingford subcohort	NCI cohort, USA N = 25,619 Employed 1934–66 Follow-up 1966–94 Wallingford N = 7,345 Employed 1941–84 Follow-up 1945–2003	SMR NCI cohort Wallingford Exposure res RR; number $\underline{Mean intensi}$ > 0-< 0.5 0.5-< 1.0 $\geq$ 1.0 $P_{trend}$ $\underline{Peak exposur}$ > 0-< 2.0 2.0-< 4.0 $\geq$ 4.0 $P_{trend}$ $\underline{Cumulative e}$ > 0-< 1.5 1.5-< 5.5 $\geq$ 5.5 $P_{trend}$	1.19 (0.38–3.68); 3 2.64 (0.54–7.71); 3 sponse analysis (NCI) of exposed deaths $\frac{ty (ppm)}{1.00}$ 1.48; 1 NA; 0 -0.802 <sup>a</sup> re (ppm) 1.00 1.55; 1 1.47; 1 0.414 exposure (ppm-yrs) 1.00 1.32; 1 NA; 0 -0.855 <sup>a</sup>	Endpoint cannot be defined as SNC since paranasal sinuses are excluded Adjusted by calendar year, age, sex, race, and pay category; exposure was calculated with a 15- year lag interval
Pinkerton <i>et al.</i> 2004 (update of Stayner <i>et al.</i> 1985 (PMR study), 1988 (SMR study))	NIOSH cohort of garment workers, USA N = 11,039 1955–98	0 deaths, 0.10	6 expected	

Table 3-3a. Summary of cohort studies of formaldehyde exposure and cancer of the
sinus and nasal cavities (SNC)

Reference	Study population and follow up	Risk estimate, 95% CI; number of observed cases or deaths	Comments
Stern <i>et al.</i> 1987	Workers employed in two chrome leather tannery plants, USA N = 9,365 1940–79 or 1980	SMR NR; 1 death in finishing department	Formaldehyde- exposed workers in the finishing department (N not stated)
Studies on health pro	ofessional workers		
Hayes et al. 1990	Deceased embalmers and funeral directors identified using licensing board records, death certificates, and other sources, USA N = 4,046 1975–85	0 deaths, 1.7 expected	Small cohort
Levine et al. 1984	Licensed embalmers in Ontario, Canada N = 1,413	0 deaths, 0.2 expected	Small cohort
Stroup et al. 1986	Anatomists who were members of the American Association of Anatomists, USA N = 2,317 1888–1979	0 deaths, 0.5 expected	Small cohort
Walrath and Fraumeni 1983	All licensed embalmers in New York, USA N = 1,263 1902–80	0 deaths, 0.5 expected	Small cohort
Walrath and Fraumeni 1984	All licensed embalmers in California, USA N = 1,109 1916–80	0 deaths, 0.6 expected	Small cohort

Findings for SNC cancers were not reported by Andjelkovich *et al.* (1995), Bertazzi *et al.* (1986), Stellman *et al.* (1998), Hall *et al.* (1991), Edling *et al.* (1987b).

NR = not reported; SMR = standardized mortality ratio; SPICR = standardized proportionate incidence cancer ratios.

<sup>a</sup> The  $P_{\text{trend}}$  value reported was based on only 2 values for trend.

Reference/Study geographic location	Study population	Exposure assessment	OR or RR (95% CI); exposed cases/controls	Comments
Olsen and Asnaes 1986, Olsen <i>et al.</i> 1984 Denmark	Population-based study 1970–82 Cases: 466 (67% men) identified by Danish Cancer Registry Controls: 2,465 men and women identified from registry with cancer of the colon, rectum, breast, or prostate and matched to cases for age, sex and yr. of diagnosis	Employment histories obtained from national pension and population registries and exposure classified by job description and industry	Analysis only on men <sup>a</sup> Certainly exposed (not adjusted)         SNC       2.8 (1.8–4.3); 33         Ever exposed (adj. for wood dust exposure)         ADC       2.2 (0.7–7.2); 17/10         SCC       2.3 (0.9–5.8); 13/11         SNC       1.6 (NR)         Ever exposed, not exposed to wood dust         ADC       7.0 (1.1–43.9); 1/8         SCC       2.0 (0.7–5.9); 4/113         SNC       1.8 (0.7–3.9); 5         Exposed to both formaldehyde and wood dust         SNC       3.5 (2.2–5.6); 28         With 10-year lag	80% power to detect an OR of 2.0 for SNC Lagging exposure by 10 years did not alter results 3 <i>t</i>
Hayes <i>et al.</i> 1986 The Netherlands	Population-based study 1978–81 Cases: 91 men (deceased and alive) with confirmed SNC, identified from cancer treatment center records Controls: 195 age-matched (frequency) men randomly selected from the population (both living and deceased)	Occupational histories obtained by interview and exposure classified by job description and industry by two independent industrial hygienists (IH <sub>A</sub> and IH <sub>B</sub> )	Subjects with little or no exposure to w $All SNC$ Any exposure/IH <sub>A</sub> 2.5 (1.2–5.0); 15/18         Any exposure/IH <sub>B</sub> 1.6 (0.9–2.8); 24/44         High exposure/IH <sub>A</sub> 3.0 (1.0–8.7); 7/7         High exposure/IH <sub>B</sub> 2.1 (1.1–4.1); 17/24         SCC         Any exposure/IH <sub>A</sub> 3.0 (1.3–6.4); 12/18	ood       No adjustment, but effect estimates did not change after adjustment for smoking or alcohol use         4         4         3

Table 3-3b. Summary of case-control studies investigating formaldehyde exposure and sinonasal cancer

Reference/Study geographic location	Study population	Exposure assessment	OR or RR (95% CI); exposed cases/controls	Comments
			Any exposure/IH <sub>B</sub> 1.9 (1.0–3.6); 19/44 High exposure/IH <sub>A</sub> 3.1 (0.9–10.0); 5/7 High exposure/IH <sub>B</sub> 2.4 (1.1–5.1); 13/24	
Vaughan <i>et al.</i> 1986 Washington, United States	Population-based study 1979–83 Cases: 53 incident cases identified using the SEER registry Controls: 552 frequency matched, and identified from random-digit dialing	Occupational histories and other information obtained by interview (present and proxy) and exposure classified using a JEM	12 exposed cases at any level, 3 exposed for at least 10 years $ORs \le 1.0$ [all CIs included 1.0] for all exposure estimates including: Maximum exposure level (low and medium or high) Number of yr exposed (1–9, 10+) Exposure scores (5–19 and 20+)	Adjusted for sex, age, smoking, and alcohol Only 12 exposed cases at any level Recall error due to next of kin interviews for the deceased subjects
Roush <i>et al.</i> 1987 Connecticut, United States	Population-based study 1935–75 Cases: 198 men who died with SNC identified using the Connecticut Tumor Registry Controls: 605 randomly selected men who died during the same time period	Occupational histories obtained from death certificates and city directories, and exposure classified by job title and industry High exposure ≥ 1 ppm	Probably exposed: level/lag time           Any/none         0.8 (0.5–1.3); 21/79           Any/20-yr         1.0 (0.5–1.8); 16/51           High <sup>c</sup> 1.0 (0.5–2.2); 9/27           High <sup>c</sup> /20 yr         1.5 (0.6–3.9); 7/14	Adjusted for age and calendar period
Luce <i>et al.</i> 1993 France	Hospital-based study 1986–98 Cases: 207 male cases (75 adenocarcinoma- 7 unexposed, 6 with possible exposure, 69 with probable or definite exposure; 82 squamous-cell carcinoma- 36 unexposed, 7 with possible exposure, 16 with probably or definite exposure; and 27 histological types) identified	Occupational histories and other information obtained by interview and exposure classified by job title and industry	Possible exposure among menSCC0.96 (0.38–2.42); 7/36ADC1.28 (0.16–10.42); 4/3SCC: Probable or definite exposure to formaldehyde among menCases/controls16 (27.1%)/81 (25.3%)No relationship <sup>d</sup> between SCC risk and exposure variables for average and cumulative level, duration of exposure, age of first exposure	Adjusted for age and exposure to wood dust ( <i>squamous-cell type only</i> ), glues, and adhesives; 97 % of ADC cases were also exposed to wood dust (which is a risk factor for ADC)

Reference/Study geographic location	Study population	Exposure assessment	OR or RR cas	(95% CI); exposed ses/controls	Comments
	from area hospital records		Date of first exp	<u>osure</u>	
	<i>Controls:</i> (1) Hospital-based series – 323 patients with		≤1944 ≥1945	1.47 (0.58–3.71); NR 0.66 (0.27–1.64); NR	
	cancers other than SNC and frequency matched by age and sex; (2) population-based series		ADC: Probable formaldehyde ar exposure to woo	or definite exposure to 1d with medium or high d dust among men	
	(N = 80) = 11515 of filends and family provided by cases and		Average level		
	matched by sex, age and residence		≤ 2 > 2	4.15 (0.96–17.84); 24/8 5.33 (1.28–22.20); 43/9	
			Duration (yr)		
			≤ 20 > 20	1.03 (0.18–5.77); 10/7 6.86 (1.69–27.80); 57/10	
			Cumulative leve	<u>.1</u>	
			$\leq 30$ 30-60 > 60	1.13 (0.19–6.90); 8/5 2.66 (0.38–18.70); 7/3 6.91 (1.69–28.23); 52/9	
			Date of first exp	osure	
			≤ 1944	6.02 (1.18–30.69); 26/6	
			≥ 1955	4.26 (1.06–17.20); 41/11	
			ADC: Combined among men	l effects with wood dust	
			Formaldehyde o	nly 8.1 (0.9–72.9); 4	
			Wood dust only	130 (14.2–1,191); 6	
			Both exposures	692 (91.9–5,210); 71	

Reference/Study geographic location	Study population	Exposure assessment	OR or RR (95% CI); exposed cases/controls	Comments
Pesch <i>et al.</i> 2008 Germany	Industry-wide case-control study woodworking industry 2003–05 Cases: 129 men [86 (57 living plus 29 next of kin) participated] identified through industry insurance records with Sinonasal adenocarcinomas (ADC) Controls: frequency matched (4 accident cases per case) 204 participants, including 69 next of kin	Occupational exposure assessed by interview and job exposure matrix	Formaldehyde exposure           Never         1.0. ref. 39/92           < 1985	Adjusted for age, region, smoking, interview status and average exposure to wood dust. Wood dust exposure; wood dust exposure associated with highly significant elevations of risk in this population

ADC = adenocarcinoma; NR = not reported; OR = odds ratio; PMR = proportionate mortality ratio; RR = risk ratio; SMR = standardized mortality ratio; SNC = sinonasal cancer, SCC = squamous-cell carcinoma.

<sup>a</sup> Women excluded from analysis since only 0.1% of controls were exposed; 4.2% of control men were exposed.

<sup>b</sup> Confidence intervals are 90% instead of 95%.

<sup>c</sup> High exposure in some year of working life; only 10 individuals were exposed to high exposure for most of their working lives. <sup>d</sup> ORs for all categories below 1.1 (except cumulative exposure < 30, OR = 1.26), and 95% CIs included 1.0.

## 1 3.4.2 Cancer of the nasopharynx

2 Nasopharyngeal carcinoma is a rare cancer, with an annual incidence rate less than 1 per 3 100,000 in most populations. WHO has classified nasopharyngeal cancers into three 4 major types: I) squamous-cell carcinomas with keratinizing potential, II) squamous-cell 5 carcinomas without keratinizing potential, and III) undifferentiated carcinomas or 6 lymphoepitheliomas) (Barnes et al. 2005). The etiology of these subtypes appears to be 7 distinct, and appears to have viral, genetic, and environmental etiology. Only Type I 8 nasopharyngeal carcinomas have been associated with potential exposure to chemical 9 agents including formaldehyde, alcohol, or smoking (Bray et al. 2008). The majority of 10 cohort studies have low statistical power to detect nasopharyngeal cancers. As in the case 11 of sinonasal cancers, findings for this site are not specifically reported in a number of 12 studies; these are noted in a footnote to the table. In other studies, the authors reported 13 specifically that no deaths from this site were observed, indicated by the note "0 deaths 14 observed" in the tables.

15 The relationship between nasopharyngeal cancers and occupational exposure to

16 formaldehyde has been investigated in cohort, nested case-control and population-based

17 case-control studies, and the key findings are summarized in Table 3-4a and b. (See

18 Section 3.1 for a description of nasopharyngeal cancers, and Section 3.3.2 for a detailed

19 summary of case-control studies investigating nasopharyngeal cancers.) [Note that in

20 several studies, findings for nasopharyngeal cancers have not been reported separately,

21 and only pharyngeal cancers combined or buccal cavity and pharyngeal cancers

22 combined are reported. Findings for these sites are reported in the section that follows.]

# 23 3.4.2.1 Cohort studies

24 Three cohort studies reported an increased risk of nasopharyngeal cancers among

25 formaldehyde-exposed workers: (1) a statistically significant increase in the risk of

26 nasopharyngeal cancers mortality in the NCI cohort (SMR = 2.10, 95% CI = 1.05 to 4.21,

- 27 8 exposed cases) (Hauptmann *et al.* 2004), (2) statistically non-significant increases in
- 28 mortality among white and non-white embalmers from the United States (Hayes *et al.*
- 29 1990), and (3) a non-significant increased incidence of nasopharyngeal cancers among
- 30 male Danish workers exposed to formaldehyde (SPICR = 1.3, 95% CI = 0.3 to 3.2.4

1 exposed cases) (Hansen and Olsen 1995, 1996). Edling et al. (1987b) reported one 2 incident case among formaldehyde exposed workers in the abrasive material industry. 3 and Coggon et al. (2003) reported one death from nasopharyngeal cancer among exposed 4 British chemical workers. Risk estimates (or expected numbers) were not provided in 5 these three studies. No deaths from nasopharyngeal cancers were reported in a very small 6 study of formaldehyde-exposed plastics manufacturing workers (Dell and Teta 1995), 7 among women in the Danish cohort (Hansen and Olsen 1996), in a study of 8 formaldehyde-exposed iron foundry workers (Andjelkovich et al. 1995), in the NIOSH 9 cohort (0 observed vs. 0.16 expected deaths; Pinkerton et al. 2004), and in two studies of 10 professionals (Stroup et al. 1986, Walrath and Fraumeni 1983). [Six studies did not report 11 findings for nasopharyngeal cancers, see Table 3-4a.]

12 Exposure-response relationships between formaldehyde exposure and nasopharyngeal 13 cancer risk were evaluated in the large NCI-sponsored historical cohort study in mixed 14 industries. In the follow-up of this cohort to December 1994, Hauptmann et al. (2004) 15 found 8 nasopharyngeal cancer deaths exposed to formaldehyde and 2 unexposed (SMR 16 = 2.10,95% CI = 0.91 to 4.14, 8 deaths). One exposed death was subsequently 17 reclassified as oropharyngeal based on secondary information not on the death certificate. 18 In internal analyses, exposure-response relationships were analyzed using the lowest 19 exposure group as the referent group. Two exposure trends were reported; one among the 20 exposed group only and one for the combined exposed and unexposed group. Relative 21 risks of nasopharyngeal cancers increased with peak exposure ( $P_{\text{trend}} < 0.001$  among 22 exposed and  $P_{\text{trend}} = 0.044$  for combined exposed and unexposed workers), average 23 exposure ( $P_{\text{trend}} = 0.066$  among exposed and  $P_{\text{trend}} = 0.126$  among combined exposed and 24 non-exposed workers), cumulative exposure ( $P_{\text{trend}} = 0.025$  among exposed and  $P_{\text{trend}} =$ 25 0.029 among combined exposed and unexposed workers). The trends for duration of 26 exposure were  $P_{\text{trend}} = 0.147$  and 0.206, respectively. All seven of the exposed deaths 27 occurred among workers with the highest peak exposure (> 4 ppm), and six of the 28 exposed deaths were among workers with average exposures of > 1.0 ppm. Because five 29 of the nine nasopharyngeal cancer cases occurred in one plant (Wallingford, 30 Connecticut), the authors conducted analyses adjusting for plant and found similar 31 exposure-response relationships with peak (adjusted  $P_{\text{trend}}$  among exposed = 0.008),

1 average (adjusted  $P_{\text{trend}}$  among exposed = 0.404), and cumulative exposure ( $P_{\text{trend}}$  among 2 exposed = 0.007), and also found a significant trend for exposure duration ( $P_{\text{trend}}$  among 3 exposed = 0.043). Marsh *et al.* (2002, 2007a) reported findings on the Wallingford cohort 4 (follow-up was to 1998 in the 2002 report and 2003 in the 2007 report), and found a 5 significant excess of nasopharyngeal cancers in both (SMR = 4.23, 95% CI = 1.78 to 6 9.13, 7 deaths for the 2007 follow-up). The authors reported that for five of the seven 7 formaldehyde-exposed nasopharyngeal cancer deaths, external employment in metal 8 working occupations was observed. In a case-control analysis of these deaths, and after 9 adjustment for metal working and smoking, the OR for exposure to formaldehyde was 10 2.87 but no longer robust. A trend toward increasing risk with increasing duration and 11 cumulative, but not average, exposure to formaldehyde was still observed. When 12 interaction modeling was applied, the OR for the five cases with both formaldehyde 13 exposure and metal-working employment and 12 controls was 9.20 (95% CI = 0.91 to 14 436.5, adjusted for smoking). Marsh et al. (2007b) also re-analyzed the findings of the 15 NCI cohort for nasopharyngeal cancers and peak formaldehyde exposure and concluded 16 that their models did not take into account the observed effect of plant type.

#### 17 3.4.2.2 Case-control studies

The relationship between formaldehyde exposure and nasopharyngeal cancer risk was evaluated in seven case-control studies (see Table 3-4b), six of which reported elevated risks for nasopharyngeal cancers among the formaldehyde-exposed subgroup of workers. Olsen *et al.* (1984) reported no increase in nasopharyngeal cancers among men ever exposed to formaldehyde (RR = 0.7, 95% CI = 0.3 to 1.7, no. of exposed cases not reported), although a statistically nonsignificant increase was observed among women (RR = 2.6, 95% CI = 0.3 to 21.9; no. of exposed cases not reported).

- 25 Hildesheim et al. (2001) and Vaughan et al. (2000) reported exposure-response trends in
- their analyses. The risk of nasopharyngeal cancers was found to increase linearly in both
- studies with duration of exposure to formaldehyde ( $P_{\text{trend}} = 0.08$ ,  $P_{\text{trend}} = 0.01$ ,
- respectively) and cumulative exposure ( $P_{\text{trend}} = 0.10$ ,  $P_{\text{trend}} = 0.03$ , respectively). In
- addition to the two studies with larger sample sizes (Hildesheim et al. 2001, Vaughan et
- 30 al. 2000), three other case-control studies examined semi-quantitative exposure indices

1 and found elevated odds ratios among workers with longer latencies, duration of 2 exposure or exposure categories (Table 3-3b). For example, West *et al.* 1993 reported 3 higher risks among workers exposed before the age of 25 (OR of 2.7, 95% CI = 1.1 to 4 6.6, 16 exposed cases) and with greater than 25 years since first exposure (OR = 2.7, 95%5 CI = 1.1 to 6.6, 16, exposed cases) in models adjusted for exposure wood dust and 6 exhaust fumes; Roush et al. 1987 reported an OR of 2.3 (95% CI = 0.9 to 6.0, 7 exposed 7 cases) for subjects with high probability of exposure and 20 years' lag time; and Vaughan 8 et al. (1986) reported an OR of 2.1 (95% CI = 0.6 to 7.8, 3 exposed cases) for their 9 highest exposure category. However, Armstrong et al. (2000) did not find an association 10 between nasopharyngeal cancers and ever being exposed to formaldehyde (OR = 0.71, 11 95% CI = 0.34 to 1.43, no. of cases not reported) after adjustment for smoking and diet, 12 and the authors reported that no exposure-response relationship was observed for a 10-13 fold increase in ratio of hours exposed [quantitative data not presented]. 14 Risk factors for nasopharyngeal cancers include wood dust, Epstein-Barr virus (EPV) 15 seroprevalence, and some dietary factors. Smoking might also be a confounder (for 16 example, Armstrong et al. (2000) reported, for subjects with nasopharyngeal cancers, a 17 statistically significant 2 to 3 fold increase in risk associated with > 6 months of active 18 smoking, and also for parental smoking among nonsmokers). Four of the seven studies of 19 formaldehyde exposure and nasopharyngeal cancers evaluated concurrent exposure to 20 wood dust as a potential confounder, and three of these four studies concluded that wood 21 dust was not a confounding factor (Hildesheim et al. 2001, Olsen et al. 1984, Vaughan et 22 al. 2000). Smoking, however, was considered as a potential confounder in several 23 studies, but an increase in risk of nasopharyngeal cancers associated with exposure to 24 formaldehyde was still observed after controlling for smoking (Vaughan et al. 2000, 25 Vaughan et al. 1986a, West et al. 1993). Hildesheim et al. (2001) did not observe a 26 confounding effect of smoking in their study, and also reported a statistically 27 nonsignificant association between ever exposure to formaldehyde and nasopharyngeal 28 cancers (OR = 1.4, 95% CI = 0.93 to 2.2, 74 exposed cases, adjusted for age, sex, 29 education, and ethnicity). (EBV seroprevalence and wood exposure were also 30 investigated in this study; the risk of nasopharyngeal cancers was associated with an OR

- of 2.3 (95% CI = 1.2 to 5.9) for EBV-seropositive subjects and with an OR of 1.7 (95%
  CI = 1.0 to 3.0) for ever exposure to wood dust).
- 3 3.4.2.3 Pooled analysis

Bosetti *et al.* (2008) conducted a pooled analysis of 3 cohort mortality studies of
formaldehyde exposure among industrial workers which included nasopharyngeal
cancers, and reported a nonsignificantly elevated estimated SMR for nasopharyngeal
cancers of 1.33 (95% CI = 0.61 to 2.53, 9 deaths). (Note that studies by Bertazzi *et al.*(1986), Edling *et al.* (1987a), and Andjelkovich *et al.* (1995) were excluded as they did
not report expected deaths).

11 of formaldehyde exposure and upper respiratory tract cancers, including nasopharyngeal 12 cancers. Fourteen cohort studies (6 of industrial workers, 4 of pathologists and 4 of 13 embalmers), together with 4 nested and 11 non-nested case-control studies, were included 14 in the meta-analysis. A statistically significant increase in the risk of nasopharyngeal 15 cancers across all studies combined was observed (mRR = 1.3, 95% CI = 1.2 to 1.5, 45516 deaths). The mRR for the cohort studies alone was not elevated, however (mRR = 1.0; 17 95% CI = 0.5 to 1.8, 10 deaths), and the mRRs for the case-control studies was elevated 18 but not statistically significant (mRR = 1.3, 95% CI = 0.9 to 2.1, 445 deaths). The authors 19 concluded that there was insufficient evidence of a causal relationship between

20 formaldehyde and nasopharyngeal cancers.

Reference	Study population and follow up	Risk estimate, 95% Cl; number of observed cases or deaths	Comments
Andjelkovich <i>et al.</i> 1995	Iron foundry workers, Michigan, USA	NR, 0 deaths	SMR – formaldehyde exposed subcohort
	N = 3,929 1959–89		Small cohort to detect rare cancers
Coggon <i>et al.</i> 2003 (update of Acheson <i>et al.</i> 1984)	British Chemical Workers Study, UK N = 14,014 1941–2000	NR, 1 death. 2 expected	
Dell and Teta 1995	5,923 workers employed at a Union Carbide plastics manufacturing plant in New Jersey, USA 1946– 67 111 formaldehyde exposed workers Follow-up 1946–88	NR, 0 deaths	Small numbers of formaldehyde exposed workers
Edling <i>et al.</i> 1987b	Swedish abrasive materials industry N = 506 male blue collar workers Mortality 1958–83 Incidence 1958–81	NR, 1 incident case	Small cohort Case had exposure <0.1 mg/m <sup>3</sup> and <5 years exposure to formaldehyde
Hansen and Olsen 1995, 1996	Denmark N = 2,041 men, 1,263 women 1970–84	SPICR analysisMen1.3 (0.3–3.2); 4WomenNR; 0 vs. 0.8expected	SPICR adjusted for age and calendar time

Table 3-4a. Summary of cohort studies of formaldehyde exposure and	d
nasopharyngeal cancers	

Reference	Study population and follow up	Risk estimate, 95% CI; number of observed cases or deaths	Comments
Hauptmann <i>et al.</i> 2004, NCI combined cohort Marsh <i>et al.</i> 2007a, Wallingford subcohort	NCI cohort, USA N = 25,619 Employed 1934– 66 Follow-up 1966–94 Wallingford N = 7,345 Employed 1941–84 Follow-up 1945– 2003	SMR NCI cohort 2.10 (1.05–4.21); 8 Wallingford 4.23 (1.78–9.13); 7 Exposure response analyses (NCI) (RR, number of exposed deaths) Average intensity (ppm) 0 (ref.) 1.00; 2 > 0–< 0.5 NA; 0 0.5–< 1.0 0.38; 1 $\geq$ 1.0 1.67; 6 Purend 0.126 Peak exposure (ppm) 0 ppm (ref.) 1.00; 2 > 0–< 2.0 NA; 0 2.0–< 4.0 NA; 0 $\geq$ 4.0 1.83; 7 Purend 0.044 <sup>c</sup> Cumulative exposure (ppm-yr) 0 ppm 2.40; 2 > 0–< 1.5 (ref) 1.00; 3 1.5–< 5.5 1.19; 1 $\geq$ 5.5 4.14; 3 Purend 0.029 Wallingford plant (Marsh 2007a) Formaldehyde exposure – nested case-control analysis Unadj. 1.41 (0.2 to $\infty$ ); 7 No increasing trends with increasing duration, average or cumulative exposure after adjusting for smoking and external employment	Hauptmann et al. Adjusted by calendar year, age, sex, race, and pay category; exposure was calculated with a 15- year lag interval 10 total deaths (8 exposed) from cancer of the nasopharynx; one death subsequently re-classified as oropharynx and excluded from internal analysis (6 of the 10 deaths occurred in Wallingford plant) Marsh et al. 2007a Adjusted for smoking and external employment (silver smithing or other metal work) Reanalysis by Marsh et al. 2004, see Section 3. 2
Pinkerton <i>et al.</i> 2004 (update of Stayner <i>et al.</i> 1985 (PMR study), 1988 (SMR study)	NIOSH cohort of garment workers, USA (N = 11,039) External analysis SMR 1955–98 PMR 1959–82	NR, 0 deaths vs. 0.16 expected	

Reference	Study population and follow up	Risk estimate, 95% CI; number of observed cases or deaths	Comments
Hayes <i>et al.</i> 1990	Deceased embalmers and funeral directors identified using licensing board records, death certificates, and other sources, USA N = 4,046 1975–85	PMR Whites 1.89 (0.39–5.48); 3 Non-whites 4.00 (0.10–22.29); 1	Small cohort
Stroup et al. 1986	Anatomists, members of the American Association of Anatomists, USA N = 2,317 1888–1979	NR, 0 deaths	Small cohort
Walrath and Fraumeni 1983	All licensed embalmers in New York, USA N = 1,263 1902–80	NR, 0 deaths	Small cohort

Results for NPC not reported individually by Bertazzi *et al.* 1986, Stellman *et al.* 1998, Stern *et al.* 1987, Hall *et al.* 1991, Levine *et al.* 1984, and Walrath and Fraumeni 1984.

SPICR = standardized proportionate incidence cancer ratios, PMR = proportionate mortality ratio, SMR = standardized mortality ratio, NR = not reported.

<sup>a</sup>  $P_{\text{trend}}$  across exposed.

<sup>b</sup>  $P_{\text{trend}}$  across exposed and non-exposed.

<sup>c</sup>[The  $P_{\text{trend}}$  value reported was based on only 2 values for trend.]

# Table 3-4b. Summary of case-control studies (including nested case-control studies) and cancer registry studies of formaldehyde exposure and nasopharyngeal cancer.

Reference	Study population	Exposure assessment	OR or RR (95% CI); exposed cases/controls	Comments
Olsen <i>et al.</i> 1984 Denmark	Population based study 1970–82 Cases: 293 men with NPC identified using Danish Cancer Registry; 266 used in analysis of NPC (excluding sarcomas) <i>Controls</i> : 2,465 men and women identified from registry with cancer of the colon, breast, or prostate and matched to cases for age, sex and yr. of diagnosis	Employment histories obtained from national pension and population registries and exposure classified by job title and industry	Ever exposed Men 0.7 (0.3–1.7); NR Women 2.6 (0.3–21.9); NR	No adjustment 4.2% of male and 0.1% of female controls considered exposed, number of cases not given
Vaughan <i>et al.</i> 1986 Washington, United States	Population based study 1979–83 Cases: 27 incident cases identified using the SEER registry Controls: 552 frequency matched, and identified from random-digit dialing	Occupational histories and other information obtained by interview and exposure classified using a JEM	Maximum exposure level           Low         1.2 (0.5–3.3); 7/121           Med. or high         1.4 (0.4–4.7); 4/50           Exposure duration (yr)         1–9           1–9         1.2 (0.5–3.1); 8/127           10+         1.6 (0.4–5.8); 3/44           Exposure score (weighted sum of duration and exposure level)         Low           Low         0.9 (0.2–3.2); 3/59           High         2.1 (0.6–7.8); 3/29	Adjusted for smoking and race Low = exposure score of 5– 19 High = exposure score of 20+

Reference	Study population	Exposure assessment	OR or RR (95% CI); exposed cases/controls	Comments
Roush <i>et al.</i> 1987 Connecticut, United States	Population-based study 1935–75 Cases: 173 men who died with SNC identified using the Connecticut Tumor Registry Controls: 605 randomly selected men who died during the same time period	Occupational histories obtained from death certificates and city directories, and exposure classified by job title and industry High exposure ≥ 1 ppm	Probably exposed: level/lag time         Any/none       1.0 (0.6–1.7); 21/79         Any/20-yr       1.3 (0.7–2.4); 17/51         High/none       1.4 (0.6–3.1); 9/27         High/20 yr       2.3 (0.9–6.0); 7/14	Adjusted for age and calendar period
West <i>et al.</i> 1993 Philippines	Hospital-based study (period of case ascertainment is unclear) <i>Cases:</i> 104 incident cases of NPC identified at Philippines General Hospital <i>Controls:</i> (1) 104 matched (sex, age, and ward type) hospital controls; and (2) 101 matched (sex, age, and neighborhood) community controls	Occupational histories and other information obtained by interview and exposure classified by job description and industry	Adjusted for wood and exhaust fumesDuration of exposure $(yr)/lag (yr)$ < 15/0	Risk estimate calculated using all controls Two models: (1) Adjusted for years since first exposure to wood and exhaust fumes; analysis of years since first exposure (2) final model - further adjusted for education, consumption of processed meats and fresh fish, smoking, and use of mosquito coils and herbal medicines

\_\_\_\_\_
Reference	Study population	Exposure assessment	OR or RR (95% CI); exposed cases/controls	Comments
Armstrong <i>et al</i> . 2000 Malaysia	Population-based study 1987–92 Cases: 282 NPC cases identified from health center records in Kuala Lumpur and Selangor among Malaysian Chinese Controls: 282 matched (sex and age) controls	Occupational histories and other information obtained by interview and classified by job description and industry Range of exposures – TWA = 0.16  to  0.35 mg/m <sup>3</sup> (except adhesives industry, $\ge 0.37 \text{ mg/m}^3$ )	Ever exposed 0.71 (0.34–1.43) <sup>a</sup> No exposure response relation with increasing duration, lag time or intensity No. exposed cases not specified; 9.9% of total cases exposed to formaldehyde and 49 pairs (at least one exposed to formaldehyde) included in analyses	Adjusted for smoking and diet Controls selected by house to house sampling
Vaughan <i>et al.</i> 2000 United States (Connecticut, Iowa, Utah, Washington, and Detroit)	Population based study 1987–93 Cases: 196 NPC identified from SEER registries Controls: 244 frequency matched (age, sex, and registry) controls in the same locations identified from random digit dialing	Occupational histories and other information obtained by interview (participant and proxy) and classified by job description and industry <i>Exposure groups: TWA-8</i> h (ppm) Low < 0.10 Moderate $\geq$ 0.10–< 0.50 High $\geq$ 50	Histological type and ever exposed         Undifferentiated and         non-keratinising       0.9 (0.4–2.0); 18/79         Differentiated squamous         cell       1.5 (0.8–2.7); 49/79         Epithelial       3.1 (1.0–9.6); 12/79         Analysis excluding undifferentiated and non-keratinizing histologies         Possible, probable, or definite exposure         Ever exposed       1.6 (1.0–2.8); 61/79         Duration (yrs)         1–5       0.9 (0.4–2.1); 16/41         6–17       1.9 (0.9–4.4); 20/19         ≥ 18       2.7 (1.2–6.0); 25/19         Ptrend       0.014         Cumulative exposure (ppm-yrs)         0.05–0.40       0.9 (0.4–2.0); 15/40         > 0.4–1.10       1.8 (0.8–4.1); 22/20         > 1.10       3.0 (1.3–6.6); 24/19         Ptrend       0.033	Adjusted for age, sex, region, smoking, proxy status, and education Exposure to wood dust did not increase the risk of NPC in this study

Reference	Study population	Exposure assessment	OR or RR (s case	95% CI); exposed es/controls	Comments
Kelerence Hildesheim <i>et al.</i> 2001 Taipei, Taiwan	Population based study 1991–94 Cases: 375 NPC cases identified at 2 tertiary care hospitals Controls: 325 individually matched (sex, age, residence) controls with no history of NPC identified using a National Household Registration system	Occupational histories and other information obtained by interview and classified by job title and industry	CaseProbable or definitEverDuration, $P_{trend}$ Definite exposureEver exposedDuration, $P_{trend}$ Ever exposedDuration, $P_{trend}$ Ever exposedCumulative, $P_{trend}$ EverCumulative exposed> 25 $\geq 25$ $P_{trend}$ Exposure durationAll subjects $\leq 10$ > 10 $P_{trend}$ Subjects without exposure $\leq 10$ > 10 $P_{trend}$ Risk estimates (~2with high averageof exposure but not	<b>isymposize</b> 2.1 (1.1-4.2); 27/30         0.069         0.13         13.3 (2.5-70); 10/2         < 0.001	Adjusted for age, sex, ethnicity, and education Exposure to wood dust was associated with an increased risk of NPC in this study Correlation between wood and formaldehyde exposure in the control population ranged from 0.26 to 0.35
			exposure were obs	duration or cumulative served	

<sup>a</sup>Only 8 individuals were exposed for > 10 years outside the 10 year latency period.

\_\_\_\_\_

### 1 3.4.3 Other head and neck cancers

2 This section summarizes studies of head and neck cancers other than sinonasal cancers 3 and nasopharyngeal cancers, including combined cancers of the upper respiratory system, 4 and cancers of the oral or buccal cavity, pharynx, the oro- and/or hypopharynx (OHPC), 5 salivary glands, and larynx. See Section 3.1 for a description of these head and neck 6 cancers, and Section 3.3.3 for a detailed summary of corresponding case-control studies 7 and Tables 3-5a and 3-5b for a summary of the site-specific risk estimates. Note that no 8 results were reported for other head and neck cancer in studies conducted by Edling et al. 9 1987b, Dell and Teta 1995, Bertazzi et al. 1986, Stellman et al. 1998, and Hall et al. 10 1991.

Known risk factors for cancers of the upper respiratory system include smoking and
alcohol use, though these factors contribute more heavily to some cancer sites than
others. All of the case-control studies reviewed in this section adjusted for smoking, with
the exception of Wilson *et al.* (2004).

## 15 3.4.3.1 Upper respiratory cancer

16 One large nested case-control study (Partanen et al. 1990) (see Table 3-5b) and one 17 cohort of mixed industries (Hauptmann et al. 2004) (see Table 3-5a) examined all upper 18 respiratory tract cancers combined; Partanen *et al.* (1990) found an increase in cancer risk 19 in relation to formaldehyde exposure (OR = 2.38, 95% CI = 0.43 to 13.2, deaths adjusted 20 for vital status, but this was based on only 2 deaths) and Hauptmann et al. (2004) 21 reported some evidence of increasing risk with increasing average, peak, and exposure in 22 the NCI cohort study, although no statistically significant trends were observed (see 23 Table 3-5b). [Hauptmann et al. 2004 did not control for smoking in the cohort because, 24 according to the authors, the prevalence of smoking did not differ by formaldehyde 25 exposure.]

- 26 3.4.3.2 Buccal cavity and pharyngeal cancer
- 27 Elevated (although not statistically significant) risks for cancers of the mouth, buccal
- 28 cavity, or buccal cavity combined with the pharynx were observed in several cohort
- 29 studies including iron foundry workers exposed to formaldehyde (SMR = 1.31, 95 % CI

1	= 0.48 to 2.86, 6 deaths) (Andjelkovich <i>et al.</i> 1995), male and female garment workers
2	with potential exposure to formaldehyde (SMR = $1.33$ , $95\%$ CI = $0.36$ to $3.41$ , 4 deaths)
3	(Pinkerton <i>et al.</i> 2004), British chemical workers (SMR for mouth = 1.28, 0.47 to 2.78; 6
4	deaths, SMR = 1.55, 95% CI = 0.87–2.56; 15 deaths), (Coggon et al. 2003) and
5	embalmers from the United States (PMR for whites = $1.19 (0.78 \text{ to } 1.74)$ ; 26 deaths, and
6	PMR for non-whites = 1.25 (0.34 to 3.2, 4 deaths) (Hayes <i>et al.</i> 1990), New York (PMR
7	= 1.13, 8 deaths) (Walrath and Fraumeni 1983), and California (PMR = 1.3 8 deaths)
8	(Walrath and Fraumeni 1984). Hansel and Olsen (1996) reported a SPICR of 1.1 (95% CI
9	= 0.7 to 1.7; 23 cases) among male Danish workers, and 1 death from buccal cavity
10	cancer was reported among formaldehyde-exposed tannery workers (Stern et al. 1987).
11	No association with formaldehyde exposure and cancer of the buccal cavity or buccal
12	cavity and pharynx cancers (combined) was found in the NCI cohort study (Hauptmann
13	et al. 2004), the Danish cohort (women) (Hansel and Olsen (1996), and in two studies of
14	health professionals (Levine et al. 1984, and Stroup et al. 1986) (see Tables 3-5a and 3-
15	5b).

16 In the standardized incidence study of Finnish men and women by Tarvainen et al. 17 (2008), no association was found between formaldehyde exposure and combined oral 18 cavity, tongue, and pharyngeal cancer (SIRs range from 0.73 to 1.01). Two population-19 based case-control studies found non-significant increases for cancer of the oral cavity or 20 oral cavity and pharynx combined and any exposure to formaldehyde: OR for oral cavity 21 and oropharynx combined = 1.6 (95% CI = 0.9 to 2.8, 25 cases) (Merletti *et al.* 1991) and 22 OR for oral cavity = 1.28 (95% CI = 0.64 to 2.54, 14 cases) (Gustavsson *et al.* 1998) 23 (Table 3-5b). In the only study of salivary gland cancer (Wilson et al. 2004) found that 24 risks increased with increasing higher probability and intensity of exposure (combined) 25 was associated with cancer ( $P_{\text{trend}} < 0.001$ , in analyses including low-level exposures). 26 Though this case-control study was quite large, no adjustment was made for smoking 27 status.

28 Laforest *et al.* (2000) found a positive association between formaldehyde and

29 hypopharyngeal squamous-cell carcinoma; this study also noted a strong exposure-

30 response trend with increasing probability ( $P_{\text{trend}} < 0.005$ ), duration ( $P_{\text{trend}} < 0.04$ ), and

1 cumulative exposure ( $P_{\text{trend}} < 0.14$ ) to formaldehyde. Berrino *et al.* (2003) reported 2 increased risks of hypopharyngeal cancer among workers with > 10 years duration of 3 exposure although risk estimates did not increase with increasing duration of exposure or 4 probability of exposure; this study included a validation analysis which suggested that the 5 exposure assessment was not sensitive to formaldehyde. Vaugan et al. 1986 found a 6 statistically non-significant increased risk for oro-and hypopharynx cancers (combined) 7 among subjects with high exposure scores or longer exposure duration. In a nested-case 8 control study among workers in the Wallingford plant of the NCI study, Marsh et al. 9 (2002) found that risk of pharyngeal cancer (including 5 cases of nasopharyngeal cancer) 10 increased with increasing duration of exposure (OR for 10+ years exposure duration = 11 2.23, 95% CI = 0.34 to 14.97, 5 cases), but not with cumulative, average intensity of 12 exposure.

### 13 3.4.3.3 Laryngeal cancer

14 With respect to laryngeal cancer, none of the cohort studies reported an association with

15 laryngeal cancer except for a statistically non-significant increase among highly exposed

16 British chemical workers (SMR = 1.6, 95% CI = 0.63-3.22; 7 deaths) (see Table 3-5a)

17 (Coggon *et al.* 2003). In internal analyses, Hauptmann *et al.* (2004) observed an

18 increased risk (OR = 2.02, 95% CI not reported) for the highest category of exposure

19 intensity only.

20 Among three case-control studies that focused on cancer of the larynx, Wortley et al.

21 (1992) found elevated risks at the highest levels of peak exposure with greater than 10

22 years of exposure (OR = 4.3, 95% CI = 1.0 to 18.7, cases not reported), but no exposure

23 response relationship was observed with duration, peak, or level of exposure. Gustavsson

*et al.* (1998) observed an elevated though statistically non-significant risk ratio for any

exposure and squamous-cell type laryngeal cancer (OR = 1.45, 95% CI = 0.83 to 2.51, 23

26 cases). However, other effect estimates were generally close to the null. No association

27 between formaldehyde exposure and laryngeal cancer was found in a hospital based case-

control study (Elci *et al.* 2003).

# 1 3.4.3.4 Pooled analysis.

- 2 In a pooled analysis of 10 occupational cohort mortality studies which included analyses
- 3 of oral cavity and pharyngeal cancers, Bosetti et al. (2008) calculated a combined
- 4 estimated RR (using a weighted average of SMRs and/or PMRs) of 1.09 (95% CI = 0.88
- 5 to 1.34, 88 deaths) among industrial workers and 0.96 (95% CI = 0.75 to 1.24, 61 deaths)
- 6 among medical workers exposed to formaldehyde.

Reference	Study population and follow up	Risk estimate exposed case	, 95% CI; number of es or deaths	Comments
Andjelkovich et al. 1995	Iron foundry workers, MI USA N = 3,929 1960–89	Buccal cavity/pl SMR Internal analysi unexposed quartiles of estin	harynx 1.31 (0.48–2.86); 6 s; 6 exposed, 5 nated cumulative	SMR – formaldehyde exposed subcohort Internal analyses using unexposed workers as reference were adjusted for race smoking and exposure
		Ever Q3+Q4 (vs. never) Larynx SMR	0.59 (0.14–2.93) 1.16 (0.20–6.51) 0.98 (0.11–3.53); 2	to silica
Coggon <i>et al.</i> 2003 (update of Acheson <i>et al.</i> 1984)	British Chemical Workers Study, UK N = 14,014 1941–2000	SMR analysis Mouth Pharynx Larynx High exposed w Mouth Pharynx	1.28 (0.47–2.78); 6 1.55 (0.87–2.56); 15 1.07 (0.58–1.79); 14 vorkers 1.32 (0.16–4.75); 2 1.91 (0.78–4.17); 6 1.56 (0.63–3.22); 7	
Hansen and Olsen 1995, 1996	Denmark N = 2,041 men 1,263 women 1970–84	SPICR analysis Buccal cavity/pl Men Women Larynx Men Women	$harynx^{a}$ 1.1 (0.7–1.7); 23 0.8 (0.3–1.7); 6 0.9 (0.6–1.2); 32 0.6 (0.1–1.7); 3	SPICR adjusted for age and calendar time Workers had 10 or more years of formaldehyde exposure before diagnosis
Hauptmann <i>et</i> <i>al.</i> 2004, NCI combined cohort Marsh <i>et al.</i> 2007a, Wallingford subcohort	NCI cohort, USA N = 25,619 Employed: 1934–66 Follow-up: 1966–94 Wallingford N = 7,345 Employed: 1941–84 Follow-up: 1945– 2003	SMR analyses NCI cohort Buccal cavity Larynx Wallingford Pla Lip Tongue Salivary gland. Mouth floor Other oral Larynx <u>Pharynx</u> All (not NPC) Oropharynx Hypopharynx	1.01 (0.77–1.34); 49 0.95 (0.63–1.43); 23 <i>int (Marsh 2007a)</i> 7.08 (0.18–39.45); 1 0.92 (0.30–2.78); 5 0.66 (0.02–3.65); 1 1.41 (0.17–5.07); 2 1.18 (0.32–3.02); 4 1.51 (0.85–2.50);15 1.71 (1.01–2.72); 16 1.71 (0.56–4.00); 5 1.43 (0.29–4.17); 3	Adjusted by calendar year, age, sex, race, and pay category; exposure was calculated with a 15-year lag interval

Table 3-5a. Summary of cohort studies of formaldehyde exposure and cancers of the oral cavity, pharynx, and larynx

Reference	Study population and follow up	Risk estimate exposed case	, 95% Cl; number of s or deaths	Comments
		Other	1.88 (0.81-3.70);16	
		Internal analysis	s RR, cases	
		NCI Cohort		
		Upper respirator	y tract	
		Mean intensity (	, mag	
		0	1.47:11	
		> 0-< 0.5	1.00; 18	
		0.5-< 1.0	1.69; 11	
		$\geq 1.0$	2.21*; 15	
		P <sub>trend</sub>	0.158	
		Peak exposure (	<u>opm)</u>	
		0	1.32; 11	
		> 0-< 2.0	1.00; 14	
		2.0-< 4.0	1.24; 12	
		$\geq$ 4.0	1.65; 18	
		P <sub>trend</sub>	0.302	
		Cumulative expo	osure (ppm-yrs)	
		0	1.24; 11	
		> 0-< 1.5	1.00; 23	
		1.5-< 5.5	1.92; 15	
		$\geq$ 5.5	0.86; 6	
		$P_{\rm trend}$	0.744	
		Buccal cavity		
		Mean intensity (	<u>ppm)</u>	
		0	2.42*; 13	
		>0-<0.5	1.00; 18	
		0.5 - < 1.0	2.41*; 16	
		$\geq 1.0$	1.89, 15	
		Peak exposure (1	0.791 mm)	
		0	2.08:13	
		> 0-< 2.0	1.00: 15	
		2.0-< 4.0	1.07; 11	
		$\geq$ 4.0	1.83; 23	
		$P_{\mathrm{trend}}$	0.433	
		Cumulative expo	osure (ppm-yrs)	
		0	1.98; 13	
		> 0-< 1.5	1.00; 25	
		1.5-< 5.5	1.59; 12	
		$\geq 3.3$	1.74;12	
		P trend	0.422	
		Mean intensity (	(mag	
		$\frac{1}{0}$	1.09; 6	
		> 0-< 0.5	1.00; 11	
		0.5-< 1.0	1.00; 4	
		$\geq 1.0$	2.02; 8	
		P <sub>trend</sub>	0.284	
		Peak exposure (	<u>opm)</u>	
		0	0.86; 6	
		> 0-< 1.5	1.00; 10	

Reference	Study population and follow up	Risk estimate exposed case	, 95% CI; number of s or deaths	Comments
		$\begin{array}{c} 1.5 < 5.5 \\ \geq 5.5 \\ P_{\text{trend}} \\ \hline Cumulative expended \\ 0 \\ > 0 < 1.5 \\ 1.5 < 5.5 \\ \geq 5.5 \\ P_{\text{trend}} \end{array}$	1.19; 8 0.64; 5 -0.645 <u>osure (ppm-yrs)</u> 0.97; 6 1.00; 13 1.81; 9 0.84; 1 -0.043	
Pinkerton <i>et</i> <i>al.</i> 2004 (update of Stayner <i>et al.</i> 1985 (PMR study), 1988 (SMR study))	NIOSH cohort of garment workers, USA (N = 11,039) External analysis SMR 1955–98 PMR 1959–82	SMR study Buccal cavity Pharynx Larnyx PCMR study Buccal cavity	1.33 (0.36–3.4); 4 0.64 (0.13–1.86); 3 0.88 0.18–2.59); 3 6.82 (1.85–17.58) <sup>b</sup> ; 3	
Stern <i>et al.</i> 1987	Workers employed in two chrome leather tannery plants, USA N = 9,365 1940–1982	SMR Buccal cavity/ Pharynx Larynx	NR, 1 death NR	Formaldehyde-exposed workers in the finishing department (N not stated)
Studies on hea	lth professional worker	ŝ		
Hayes <i>et al.</i> 1990	Deceased embalmers and funeral directors identified using licensing board records, death certificates, and other sources, USA N = 4,046 1975–85	PMR analysis Buccal cavity/ph Whites Non-whites Larynx Whites Non-whites	harynx 1.19 (0.78–1.74); 26 1.25 (0.34–3.2); 4 0.64 (0.26–1.33); 7 0 death vs. 1.6 exp.	Small cohort
Levine <i>et al.</i> 1984	Licensed embalmers in Ontario, Canada N = 1,413	SMR analysis Buccal cavity/ pharynx Larynx	1 death vs. 2.1 exp. 1 death vs. 1 exp.	Small cohort
Stroup <i>et al.</i> 1986	Anatomists, members of the American Association of Anatomists, USA N = 2,317 1888–1979	SMR analysis Buccal cavity/ pharynx Larynx	0.2 (0.00–1.71); 2 0.4 (0.0–2.0); 1	Small cohort

Reference	Study population and follow up	Risk estimate exposed case	, 95% Cl; number of s or deaths	Comments
Walrath and Fraumeni 1983	All licensed embalmers and funeral directors in New York, USA N = 1,263 1902–80	PMR analysis of Buccal cavity ar All whites Embalmers only Larynx Whites Non-whites	n males nd pharynx 1.13; 8 2.01; 7 2 vs. 3.4 exp. 2 deaths, <i>P</i> < 0.05	Small cohort
Walrath and Fraumeni 1984	All licensed embalmers in California, USA N = 1,109 1916–80	PMR study on v Buccal cavity/ pharynx Larynx	white males 1.31; 8, <i>P</i> > 0.05 2 vs. 2.6 exp.	Small cohort

\* P < 0.05.

Results for oral cavity, pharynx and larynx cancers were not reported by Edling *et al.* 1987b, Dell and Teta, 1995, Bertazzi *et al.* 1986, Stellman *et al.* 1998, and Hall *et al.* 1991.

NPC = nasopharyngeal cancer; NR = not reported; PCMR = proportionate cancer mortality ratio; PMR = proportionate mortality ratio; Q = quartile, SMR = standardized mortality ratio; SPICR = standardized proportionate incidence cancer ratio.

<sup>a</sup> Excluding nasopharynx.

<sup>b</sup> 90% CI.

# Table 3-5b. Summary of case-control studies (including nested case-control studies) and cancer registry studies of formaldehyde exposure and cancers of the oral cavity, pharynx, and larynx

Reference	Study population	Exposure assessment	OR or RR (95% CI); exposed cases and controls	Comments
Partanen <i>et al.</i> 1990; (update of Partanen <i>et al.</i> 1985) Finland	Nested case-control study Cohort: particleboard, plywood, or formaldehyde glue factory workers, 1957–80 <i>Cases:</i> 136 cases of all respiratory system cancer including tongue, pharynx, larynx, epiglottis, trachea and lung <i>Controls:</i> 408 controls randomly selected from cohort; 3:1 ratio, matched on year of birth and alive at date of case diagnosis	Occupational histories obtained using plant records and classified using factory-specific JEMs	Upper respiratory only ≥ 3 ppm-months 2.38 (0.43–13.2); 2 With 10-yr lag 2.40 (0.31–18.6); 2	Adjusted for vital status and smoking
Tarvainen <i>et al.</i> 2008 Finland	Cancer registry-based standardized incidence study All oral cavity, tongue and pharyngeal cancers (excluding nasopharynx) in Finnish Cancer Registry, from 1971 to 1995, males and females born 1906–45	1970 census data used to construct national job exposure matrix based on longest-held occupation	SIR (95% CI); no. observed cases         Formaldehyde, estimated cumulative         exposure, ppm-years:         Low       0.79 (0.6–1.03); 59         Medium       1.01 (0.43–1.98); 8         High       0.73 (0.27–1.59); 6	Adjusted for age, calendar period and socioeconomic status. Exposures lagged for ten years.

Reference	Study population	Exposure assessment	OR or RR (95% CI); exposed cases and controls	Comments
Merletti <i>et al</i> . 1991 Turin, Italy	Population-based study Jul. 1982–Sep. 1984 <i>Cases:</i> All male Turin residents diagnosed with cancer of the oral cavity and oropharynx (103 eligible cases) 86 agreed to interview <i>Controls:</i> random sample of 679 age and sex matched controls: 385 agreed to interview (371 with complete occupational history)	Occupational histories obtained by interview and classified using a JEM	Oral cavity and oropharynx combined <i>Exposure to formaldehyde</i> Any 1.6 (0.9–2.8); 25/79 Probable or definite 1.8 (0.6–5.5); 6/13 No exposure-response relationships observed but elevated ORs observed for most exposure categories	Adjusted for age, education, area of birth, smoking, and alcohol
Gustavsson <i>et al.</i> 1998 Sweden	Population-based, case- control studies various cancers Jan. 1988–Jan. 1991 Cases: identified from health care records and cancer registries Oral cavity (N = 128) Pharynx (N = 138) Larynx (N = 157) Controls: 641 selected from population registries and matched by region and age	Occupational histories, lifestyle and environmental information obtained by interview and exposure classified by job title and industry	Ever exposed Oral cavity 1.28 (0.64–2.54); 14 Pharynx 1.01 (0.49–2.07); 13 Larynx squamous cell type 1.45 (0.83–2.51); 23 No exposure relationship with cumulative exposure or duration	Adjusted for age, region, smoking, and alcohol

Reference	Study population	Exposure assessment	OR or RR (95% CI); exposed cases and controls	Comments
Marsh <i>et al.</i> 2002 United States	Nested case-control study within the Wallingford plant cohort (N = 7,328); 1941–98 <i>Cases:</i> 22 cases of pharyngeal cancer (including 7 nasopharynx) <i>Controls:</i> 88 members of the cohort matched on race, sex, age and year of birth	Occupational histories obtained from employment and sampling records	All pharyngeal cancers (inc. nasopharynx)         Non-exp       1.0 (ref); 2         Ever       3.04 (0.36–145.58); 20         < 0.2 ppm	Adjusted for smoking and year of hire Wallingford plant is a plant in the NCI cohort Smoking data available on 15 cases and 77 controls
Wilson <i>et al.</i> 2004 United States (24 states)	Death certificate-based study 1984–89 Cases: 2,505 cases of salivary gland carcinoma (60% men, 7% black) identified by mortality records Controls: 9,420 frequency matched (age, race, sex and region) randomly selected from deaths not related to infectious disease	Occupational histories were obtained from death certificates and classified using a JEM	White men: Salivary gland         Probability/intensity of exposure         Low/low $0.9 (0.70-1.15)$ Low/mid-high $0.7 (0.35-1.26)$ Mid-high/low $2.4 (0.86-6.75)$ Mid-high/mid-high $1.6 (1.30-2.0)$ $P_{trend}$ $< 0.001$	Adjusted for age, marital status, and socioeconomic status
Vaughan <i>et al.</i> 1986 Washington, United States	Population-based study, 1980–83 Cases: 205 cases of oro- and hypopharynx cancer	Occupational histories obtained by interview and classified using a JEM	Oro- and hypopharynx <i>Exposure scores</i> Low 0.6 (0.3–1.2); 14/59	Adjusted for sex, age, smoking, and alcohol For exposure scores: Low =

Deference	Study population		OR or RR (95%	6 CI); exposed cases	Commonto
Reference	Study population	Exposure assessment	and	a controis	Comments
	identified by SEER		High	1.5 (0.7–3.0); 21/29	5-19 and High = $20+$
	registry		Exposure Duration	n (yrs)	
	Controls: 552 frequency		1–9	0.6 (0.3–1.0); 32/127	
	matched, and identified		> 10	1.3 (0.7–2.5); 26/44	
	by random-digit dialing		Maximum exposur	re level	
			OR < 1.0 for all gr	roups and CIs included	
			1.0.		
Laforest et al. 2000	Hospital based study	Occupational histories	Hypopharynx - SC	CC	Adjusted for age, smoking,
France	Jan. 1989–Apr. 1991	and other information	Probability of exp	osure (%)	alcohol, and exposure to
	Cases: 201 men with	exposure to formaldehyde	< 10	1.08 (0.62–1.88); 42/50	subjects matched by age
	confirmed SCC of the	classified using a JEM	10–50	1.01 (0.44–2.31); 15/20	Controls included subjects
	from 15 French hospitals	C	> 50	3.78 (1.50–9.49); 26/15	with primary cancers at sites
	(from 644 eligible cases		P <sub>trend</sub>	< 0.005	that have suspected
	of laryngeal and		For probability of	$exposure \ge 10\%$ :	associations with
	pharyngeal cancers and		Ever exposed	1.74 (0.91–3.34); 41/35	formaldehyde exposure
	80% participation rate)		Exposure Duration	<u>n (yr)</u>	Also studied laryngeal
	Controls: 355 controls		< 7	0.74 (0.20–2.68); 3/2	cancer (see below)
	matched (frequency) by		7–20	1.65 (0.67–4.08); 13/11	
	age and hospital with		20+	2.70 (1.08-6.73); 16/16	
	sites: 296 interviewed and		$P_{\rm trend}$	< 0.04	
	included in analyses		Cumulative level		
	2		< 0.02	0.78 (0.11–5.45); 3/2	
			0.02-0.09	1.77 (0.65–4.78); 13/11	
			> 0.09	1.92 (0.86–4.32); 25/22	
			P <sub>Prend</sub>	< 0.14	
Berrino et al. 2003	Population based study	Occupational histories	Individuals less th	an 55	Adjusted for age, sex,
Europe: France, Italy,	1979–82	and other information	Hypopharynx/lary	nx	smoking, alcohol, diet, SES,
Spain, Switzerland	Cases: 315 <sup>a</sup> men under	obtained by interview and	Ever exposed	1.3 (0.8–2.0); 113/192	center, and exposure to
	55 with hypopharyngeal/	exposure to formaldehyde	Probability of exp	osure:	asbestos, PAH, Cr, As,
		was classified using a			wood dust, solvents, and

Reference	Study population	Exposure assessment	OR or RR (95% CI); exposed cases and controls	Comments
	larvngeal cancer (213	IEM Some interviews	Possible $1.5 (0.9-2.4) \cdot 90/146$	other dusts and gases
	endolarynx and 100 HPC + epilarynx) identified from 6 health care centers <i>Controls:</i> 819 men under 55 identified from a random sample (age and sex stratified) of the population from each center	with next of kin	Probable       1.3 (0.9–2.4), 90/140         Probable       0.9 (0.4–1.9); 23/50         Exposure duration (yr)         < 10	Independent validation of JEM classified 14% of the unexposed jobs as definitely exposed. No significant associations found in analysis of individuals (695 cases and 1,357 controls) over 55
	113 exposed cases and 192 unexposed cases; 196 exposed controls and 623 unexposed controls		Endolarynx           Possible         1.4 (0.8–2.7)           Probable         1.0 (0.4–2.3)           Hypopharnx (includes epilarynx)           Possible         1.3 (0.6–2.6)           Probable         0.5 (0.1–1.8)	(numbers for formaldehyde not given)
Wortley <i>et al.</i> 1992 Washington,	Population-based, case- control study Sep. 1983–Feb. 1987 Cases: identified from population-based cancer registry in Seattle (with phones) Larynx (N = 235) Controls: 547 identified by random digit dialing, matched 2:1 with cases on age and sex Exposed cases and controls 58/124	Occupational histories and other information obtained by phone interview and exposure to formaldehyde classified using a JEM 7% of case interviews with next-of-kin	Larynx Highest exposure score 4.3 (1.0–18.7) > 10 yr exp. 4.2 (0.9–19.4) Analyses excluded low-level exposure In analyses that included low-level exposure, no exposure-response relationship was seen with duration, peak, or level of exposure	Adjusted for age, smoking, alcohol, and education; subjects matched by age and sex
Elci et al. 2003	Hospital-based, case	Occupational histories	Larynx	Adjusted for age, smoking,

Reference	Study population	Exposure assessment	OR or RR (95% and	% CI); exposed cases d controls	Comments
Turkey	control study	and lifestyle information	Ever	1.0 (0.8–1.3)	and alcohol
	1979–84	obtained by interview and	Exposure intensity	<u>v</u>	
<i>Cases:</i> 951 men with confirmed cases of laryngeal cancer presenting <i>Controls:</i> 1,519 hospital patients (non-cancer)	exposure classified using	Low	1.1 (0.8–1.5)		
	confirmed cases of	aJEM	Medium	0.5 (0.2–1.3)	
	laryngeal cancer		High	0.7 (0.1–7.1)	
	presenting		Exposure probabi	lity	
	controls: 1,519 nospital		Low	1.0 (0.7–1.4)	
	patients (non-cancer)		Medium	1.1 (0.6–2.2)	
			High	1.0 (0.1–11.2)	

\* P < 0.05

•

<sup>a</sup> Original study included 1,010 cases and 2,176 controls. Complete lifetime occupational histories were only available for subjects under 55, so analysis was restricted to this age group.

## 1 3.4.4 Respiratory cancers or lung cancer

- 2 The relationship between occupational exposure to formaldehyde and lung or respiratory
- 3 system cancers has been investigated in a large number of cohort, nested case-control,
- 4 and population-based case-control studies. The key findings are summarized in Table 3-
- 5 6a and b. (See Section 3.3.4 for a detailed summary of case-control studies investigating
- 6 lung cancer.)

# 7 3.4.4.1 Cohort studies

- 8 Increased risks for lung or respiratory cancer were reported in five of the industrial
- 9 cohorts, two of which were statistically significant or border-line significant
- 10 (Andjelkovich et al. 1995, Bertazzi et al. 1986, Coggon et al. (2003), Dell and Teta
- 11 (1995), Hansen and Olsen (1995, 1996) (women only). (See below for a discussion of the
- 12 nested case-control study of the iron foundry workers reported by Andelkovich *et al.*
- 13 [1994].) Coggon et al. (2003) reported a statistically significant increase in the risk of
- 14 lung cancer among highly exposed (> 2 ppm) British chemical workers (SMR = 1.58,
- 15 95% CI = 1.40 to 1.78, 272 deaths). Risks increased with increasing exposure level (low,
- 16 medium, high,  $P_{\text{trend}} < 0.001$ ), but not with duration of exposure. Increased lung cancer
- 17 risks were found in internal analyses of formaldehyde-exposed workers in some exposure
- 18 categories in the NCI cohort (Hauptmann et al. 2004), but no clear trends with average,
- 19 peak, or cumulative exposure were observed; no increase was observed in external
- 20 analysis of the combined cohort, although a statistically significant increase was found in
- 21 one of the constituent cohorts (Marsh *et al.* 2007a). No increases were observed in the
- 22 NIOSH garment workers cohort (Pinkerton *et al.* 2004), the Danish mixed industry
- cohort (men) (Hansen and Olsen 1995, 1996), the abrasive material industry (Edling *et al.*
- 24 1987b), among tannery workers exposed to formaldehyde (Stern et al. 1987), or among
- 25 most of the studies of health professional workers (see Table 3-6a). Stellman *et al.* (1998)
- 26 reported a significant risk for woodworkers exposed to formaldehyde (SMR = 2.63 (95%
- 27 CI = 1.25-5.51, 7 deaths) but not among workers only exposed to formaldehyde (SMR =
- 28 0.93, 95% CI = 0.73 to 1.18, 104 deaths).

### 1 3.4.4.2 Case-control studies

2 Ten case-control (including nested case-control) studies have evaluated the relationship 3 between exposure to formaldehyde and lung or respiratory cancer; two studies reported 4 on respiratory system cancers and eight studies on lung cancer independently. Marsh et 5 al. (2001) reported a statistically significant risk of respiratory (lung and larynx) cancers 6 associated with formaldehyde exposure in their nested case-control study within an 7 industrial cohort of glass wool manufacturing workers (OR = 1.61, 95% CI = 1.02 to 8 2.57, 591 ever-exposed cases, adjusted for smoking but not other exposures). Partanen et 9 al. (1990, 1985) noted elevated but statistically non-significant risks in combined mouth, 10 tongue, nose and sinuses, pharynx, larynx, trachea, epiglottis, and lung cancer associated 11 with formaldehyde exposure; in their updated analysis (Partanen et al. 1990), the OR for 12 cumulative exposure of at least three ppm-months with a 10-year lag was 1.39 (95% CI =13 0.40 to 4.10). Risk estimates were higher for cancers of the upper respiratory system 14 only.

Several studies reported increased risks (both statistically significant and non-significant
 risk) for lung cancer. Increased risks were found in nested case-control studies among

17 male and female glass wool workers exposed to formaldehyde (RR = 1.61, 95% CI =

18 1.02 to 2.57, 91 deaths for men, and 1.24, 95% CI = 1.24, 95% CI = 0.74 to 2.09, 39

19 deaths for women) (Marsh et al. 2001, Stone et al. 2004), glass wool workers

20 (independent study) with 100 to 999 cumulative days of exposure to formaldehyde (RR =

21 1.27, 95% CI = 0.50 to 3.21, 15 deaths) (Chiazze *et al.* 1997), and iron foundry workers

exposed to formaldehyde (OR of 1.31. 95% CI = 0.38 to 2.07) (Andjelkovich *et al.* 

23 (1994); however, risks decreased in exposure-response analyses by lag or duration of

24 exposure. Increased risks were also observed in two population-based case-control

studies. Gérin *et al.* (1989) reported an OR of 1.5 (95% CI = 0.8 to 2.8) for high-level

26 formaldehyde exposure only with at least 10-years duration, but no adjustment was made

27 for smoking. Chen et al. (2008) reported a statistically significant association between

28 lung cancer and the burning of mosquito coils (a practice common in Taiwan), after

adjustment for smoking and other variables, which may involve exposure to a range of

30 particulates and chemicals including formaldehyde as a combustion product.No increased

31 risks of lung cancer were reported in a nested case-control study of Dow Chemical

1 workers (Bond *et al.* 1986), a small cancer registry study of physicians (Jensen and

2 Anderson 1982), and a population-based case-control study of women (Brownson *et al.* 

3 1993).

4 For lung cancer and any respiratory system cancer, smoking is the principal potential

5 confounder; occupational exposure to dusts, synthetic vitreous fibers and other ambient

6 exposures may also be of concern. Several studies have attempted to make some

7 adjustment for smoking status (exceptions include Coggon et al. 1984, Bond et al. 1986,

8 Gérin et al. 1989, Chiazze et al. 1997 and Hauptmann et al. 2004), though in most cases

9 estimates of smoking are limited to a sample of subjects, to proxy data, or to ever-never

10 smoking status.

11 3.4.4.3 Pooled analysis.

12 In a pooled analysis of 14 occupational cohort mortality studies, which included deaths

13 from lung cancer, Bosetti et al. (2008) calculated combined estimated RRs (using

14 weighted SMRs and/or PMRs) of 1.06 (95% CI = 0.92 to 1.23, 1,459 deaths) among

15 industrial workers and 0.63 (95% CI = 0.47 to 0.84, 562 deaths) among medical workers

16 in association with formaldehyde exposure.

Reference	Study population and follow up	Risk estimate, 95% CI, number of exposed cases or deaths	Comments
Andjelkovich <i>et al.</i> 1995	Iron foundry workers, MI USA N = 3,929 1960–87	Lung cancerSMR $1.20 (0.89-1.58); 51$ RR $1.13, NR, P > 0.05$ Internal analysis (quartiles of cumulative exposure compared with. never)Ever $0.71 (0.43-1.21)$ Q3 + Q4 $0.59 (0.28-1.20)$	SMR – formaldehyde exposed subcohort See Table 3-6b for related nested case-control of larger cohort Internal analyses using unexposed workers as reference were adjusted for race, smoking, and exposure to silica
Bertazzi et al. 1986	Resin manufacturing workers, Italy N = 1,332 men 1959–80, 1986	SMR analysis Lung 1.36 (NR); 5 No increased risk with increasing years since first exposure	No quantitative exposure assessment; 28% person- years assigned to definite exposure to formaldehyde
Coggon <i>et al.</i> 2003 (update of Acheson <i>et al.</i> 1984)	British Chemical Workers Study, UK N = 14,014 1941–2000	Lung cancer (SMR analysis)All $1.22 (1.12-1.32)$ ; 594high exposed $1.58 (1.40-1.78)$ ; 272Exposure response for lung cancerIncreasing risk with increasingexposure level (low, medium, high), $P_{trend} < 0.001$ Inverse trend with duration ofexposure	
Dell and Teta 1995	Workers employed at a Union Carbide plastics manufacturing plant in New Jersey, USA 57 formaldehyde exposed workers in hexamethylenetetramine production 111 workers (total) exposed to formaldehyde 1946–88	Lung cancer (SMR) Hexamethylenetetramine production workers 4 deaths vs. 1.1 exp. All formaldehyde exposed workers NR	Small numbers of formaldehyde exposed workers Lung cancer risk elevated in whole cohort
Edling <i>et al.</i> 1987b	Abrasive materials industry, Sweden N = 506 male blue collar workers Mortality 1958–83 Incidence 1958–81	Lung cancer SMR NR SIR 0.57 (0.07–2.06); 2	

Table 3-6a. Summary of cohort studies of formaldehyde exposure and cancers of the lung

Reference	Study population and follow up	Risk estimate, 95% Cl, number of exposed cases or deaths	Comments
Hansen and Olsen 1995, 1996	Danish formaldehyde exposed worker N = 2,041 men, 1,263 women 1970–84	SPICR lung           Men         1.0 (0.9–1.1); 410           Women <sup>a</sup> 1.2 (0.96–1.4); 108           Men, formaldehyde no wood dust         1.0 (0.9–1.1); 250           Women         NR	SPICR adjusted for age and calendar time Workers had 10 or more years exposure to formaldehyde before diagnosis
Hauptmann <i>et al.</i> 2004, NCI combined cohort Marsh <i>et al.</i> 2007a, Wallingford subcohort	NCI cohort, USA N = 25,619 Employed 1934–66 Follow-up 1966–94 Wallingford N = 7,345 Employed 1941–84 Follow-up 1945–2003	Lung cancerSMRNCI cohort $0.97 (0.90-1.05); 641$ Wallingford $1.18 (1.05-1.32); 322$ NCI internal analysis (RR, number of cases):Average exposure (ppm)> $0.0-< 0.5$ $1.0 (ref.); 348$ > $0.5-<1.0$ $1.51; 146$ $\geq 1.0$ $1.16; 160$ Peak exposure (ppm)> $0.0-<2.0$ $1.0 (ref.); 237$ $2.0-<4.0$ $1.45; 227 (P < 0.01)$ $\geq 4.0$ $0.94; 177$ All RRs for cumulative exposure < 1	Internal analysis adjusted by calendar year, age, sex, race, and pay category; exposure was calculated with a 15-year lag interval Average, cumulative, and peak exposures compared to lowest exposed category
Pinkerton <i>et al.</i> 2004 (update of Stayner <i>et al.</i> 1985, 1988 – PMR and SMR studies respectively)	NIOSH cohort of garment workers, USA N = 11,039 SMR 1955–98 PMR 1959–82	Lung cancerSMR0.98 (0.82–1.15); 147PCMR0.88 (0.49–1.45) <sup>ab</sup> 11SMR did not increase with increasing duration, time since first exposure, or earlier start dates	Standardized mortality and PMR study
Stellman <i>et al.</i> 1998 50 U.S. states, District of Columbia, Puerto Rico	Woodworkers: American Cancer Society Cancer Prevention Study N = 362,823; 43,339 in woodworking occupations	Internal analysis (RR) for lung FOR only 0.93 (0.73–1.18); 104 FOR + wood 2.63 (1.25–5.51); 7	Internal analysis using non-woodworkers or workers without exposure to wood dust Adjusted for age and smoking
Stern <i>et al</i> . 1987	Workers employed in two chrome leather tannery plants, USA (N = 9,365) Employed 1940–79 or 1980	Lung SMR 0.70 (0.45–1.05); 24	Formaldehyde-exposed workers in the finishing department (N not stated)

Reference	Study population and follow up	Risk estimate, 95% CI, number of exposed cases or deaths	Comments
Studies of health pro	fessional workers	·	
Hall <i>et al.</i> 1991; (update of Harrington and Shannon 1975 and Harrington and Oakes 1984)	Pathologists, members of professional organizations in the UK N = 3,872 1974–87	Lung cancer (England & Wales) SMR 0.19 (0.09–0.36); 9	
Hayes <i>et al.</i> 1990	Deceased embalmers and funeral directors identified using licensing board records, death certificates, and other sources, USA N = 4,046 1975–85	Lung (PMR) Whites 0.97 (0.86–1.09); 285 Non-whites 0.75 (0.47–1.13); 23	
Levine et al. 1984	Licensed embalmers in Ontario, Canada (N = 1,413)	Lung SMR 0.94 (NR); 19	
Stroup et al. 1986	Anatomists, members of the American Association of Anatomists, USA N = 2,317 1888–79	Lung SMR 0.3 (0.1–0.5); 12	
Walrath and Fraumeni 1983	All licensed embalmers and funeral directors in NY, USA N = 1,263 1902–80	Lung (white males) PCMR 1.1 (NR); 70 Lung and pleura PMR 1.08 (NR); 72	
Walrath and Fraumeni 1984	All licensed embalmers in CA, USA N = 1,109 1916–80	Lung and pleura (white males) PMR 0.87 (NR); 41	

\* P < 0.05.

FOR = formaldehyde; NR = not reported; PMR = proportionate mortality ratio; PCMR = proportionate cancer mortality ratio; SMR = standardized mortality ratio; SPICR = standardized proportionate incidence cancer ratio.

<sup>a</sup> As reviewed by IARC 2006. <sup>b</sup>90 % CI.

Reference	Study population	Exposure assessment	OR or RR (95% CI); exposed cases/controls	Comments
Jensen and Andersen 1982 Denmark	Cancer registry-based case- control study of physicians 1943–76 Cases: 84 incident lung cancers Controls: physicians matched on age, sex and survival to date of diagnosis	Medical specialization and place of work for cases were compared with controls to assess the potential for increased relative exposure levels.	Ever worked in pathology, forensic medicine, anatomy : RR 1.0 (0.4–2.4); 8/23	Small no. cases No increase in risk among other physician specialties
Coggon <i>et al.</i> 1984 United Kingdom	Population-based study 1975–79 Cases: 598 men under 40 identified from death certificates with cancer of the trachea, bronchus or lung Controls: 1,180 men who died from other causes and matched to cases by sex, year of birth and death, and residence	Occupational histories obtained from death certificates, exposure classified by JEM	Ever-exposed 1.5 (1.2–1.8); 296/472 Occupations with high exposure 0.9 (0.6–1.4); 44/90	Matched tabular analysis, including matching for pay class
Bond <i>et al.</i> 1986 Texas, United States (cohort of Bond <i>et</i>	Nested case-control of Dow Chemical workers (Bond et al. 1985) 1940–80	Occupational histories obtained from company employment records and classified by job	Ever exposed         0.62 (0.29–1.34); 9/27           15-yr lag         0.31 (0.11–0.86); 4/24	

# Table 3-6b. Summary of case-control studies (including nested case-control) investigating formaldehyde exposure and lung or respiratory cancer

Reference	Study population	Exposure assessment	OR or RR (95% CI); exposed cases/controls	Comments
al. 1985)	Cases: 308 men identified using death certificates Controls: matched by race, years of birth and hire	task		
Gérin <i>et al.</i> 1989 Montreal, Canada	Multi-site study 1979–85 Cases: 857 men; incident cases identified from all hospitals Controls: (1) cancer controls, internal controls with tumors at other sites and (2) 740 population based controls matched by age	Occupational histories obtained by interview; exposure classified by job description and industry <i>Estimated exposure</i> <i>index</i> Low < 0.1 ppm Med.0.1 - 1 ppm High $\geq$ 1 ppm	Exposure duration (yrs)/exposure index (cancer controls) <sup>a</sup> Lung cancer (all)< 10/any	Adjusted for 1) age, 2) ethnicity, 3) cigarette smoking, 4) self-reported income, 5) jobs held and other occupational factors; highest OR observed 
Partanen <i>et al.</i> 1990 (update of Partanen <i>et al.</i> 1985) Finland	Nested case-control of plywood, particleboard, and formaldehyde glue factory workers (N = 7,303) 1957–82 Cases: 136 respiratory cancer cases including tongue, pharynx, larynx, trachea, epiglottis, and lung identified using the Finnish Cancer Registry Controls: 408 controls selected randomly from cohort and matched (3:1) by year of birth	Occupational histories obtained using plant records and classified using factory-specific JEMs	Workers with $\geq$ 3 ppm-months vs/ < 3	Adjusted for vital status and smoking
Brownson <i>et al.</i> 1993 Missouri, United States	Population-based study 1986–91 Cases: 429 women identified from	Occupational histories obtained by interview; exposure classified by job description	Ever-exposed 0.9 (0.2–3.3); 3/10	Adjusted for age, previous history of lung disease and smoking

\_\_\_\_\_

Reference	Study population	Exposure assessment	OR or RR (95% CI); exposed cases/controls	Comments
	the Missouri Cancer Registry <i>Controls</i> : 1,021 age-matched, selected from Medicare records			
Andjelkovich <i>et al.</i> 1994 Michigan, United States	Nested case-control study of iron foundry workers ( $N = 8,147$ ) (update of Andjelkovich <i>et al.</i> 1990) 1950–89 <i>Cases</i> : 220 lung cancer <i>Controls</i> : matched on race from cohort (10:1) using incidence density sampling	Occupational histories obtained from employment records and classified using a JEM	Ever exposed 1.31 (0.83–2.07); NR Effects decreased with increasing lag periods	Adjusted for smoking, birth cohort, and exposure to silica Analysis using subset of controls with smoking information
Chiazze <i>et al.</i> 1997 South Carolina, United States	Nested case-control of fiberglass manufacturing plant workers (N = 4,631); 1951–91 Cases: 47 white men with lung cancer Controls: 122 white men matched on year of birth and survival to end of follow-up or death	Occupational histories obtained by interview and a historical exposure reconstruction; exposure was classified by a committee of experts	Cumulative days of exposure           0.2 < 100	Unadjusted
Marsh 2001, Youk et al. 2001 Stone et al. 2004 United States	Marsh <i>et al.</i> 2001: Nested case-control study of male and female fiberglass workers (N = 32,110) 1970–92 <i>Cases:</i> 874 respiratory system cancers combined including larynx, bronchus, trachea, and lung <i>Controls:</i> alive when case died	Occupational histories obtained from company employment records and relevant industrial hygienic literature; exposure estimated using job location- weighted measures	All respiratory system combined         RR for men ever exposed to         formaldehyde         1.61 (1.02–2.57); 591         lag (yr)         5       1.62 (1.04–2.54); 588/503         10       1.46 (0.96–2.23); 581/498         20       1.17 (0.82–1.67); 537/458         No clear trends with cumulative or average exposure	MenAdjusted for smokingAnalysis on 516 pairs (631cases and 570 controls)Women37.6 person-years exposed toformaldehydeNo adjustment for smoking;models with formaldehyde andglass wool were similar to

Reference	Study population	Exposure assessment	OR or RR (95% CI); exposed cases/controls	Comments
	and matched by date of birth Stone et al. 2004: N = 4,008 women; 1970–92 3,563 included in analysis 53 respiratory-system cancer cases		RR for cumulative exposure to formaldehyde Women 1.24 (0.74–2.09); 39	univariate analysis
Chen <i>et al.</i> 2008 Taiwan	Hospital-based study of mosquito coil exposure Jul. 2002–Feb. 2004 Cases: 148: All new diagnoses of lung cancer in three medical centers; one refusal Controls: 889 age, sex-matched non-cancer hospital controls recruited, 400 participated	Exposure to mosquito coils, occupation, and lifestyle factors obtained by interview; occupational exposure classified by job and industry description	Use of mosquito coils (times per week) >3 3.78 (1.55–6.90); 24 < 3 2.67 (1.60–45.0); 32	Adjusted for age, marital status, smoking and tuberculosis No direct estimate of formaldehyde exposure available

<sup>a</sup> ORs calculated using hospital controls; similar estimates using population-based controls.

## 1 3.4.5 Lymphohematopoietic cancers

- 2 The relationship between occupational exposure to formaldehyde and
- 3 lymphohematopoietic cancer has been investigated in several cohort, nested case-control,
- 4 and population-based case-control studies. The key findings are summarized in Table 3-
- 5 7a and b. (See Section 3.3.5 for a detailed summary of case-control studies investigating
- 6 lymphohematopoietic cancer.)

## 7 3.4.5.1 Cohort studies

- 8 Eight cohort studies (including all six studies of health profession workers) have reported
  9 increased mortality of all lymphohematopoietic cancers combined although most of the
- 10 increases were not statistically significant (Bertazzi *et al.* 1986, Stellman *et al.* 1988 [the
- 11 increase was strongest among woodworkers exposed to formaldehyde], Hall et al. 1991,
- 12 Hayes *et al.* 1990, Stroup *et al.*, 1986, Levine *et al.* 1984 and Walrath and Fraumeni
- 13 1983, 1984). (See Table 3-7a for risk estimates). No increased risk of
- 14 lymphohematopoietic cancers was observed among garment workers in the NIOSH
- 15 cohort (Pinkerton et al. 2004) and among formaldehyde-exposed workers in the iron
- 16 foundry industry (Andjelkovich et al., 1995). Risk estimates (or number of deaths) were
- 17 not reported by Coggon et al. (2003), Edling et al. (1987b), Hansen and Olsen (1995,
- 18 1996), Stern *et al.* (1987), and Dell and Teta (1995). Although no increase in all
- 19 lymphohematopoietic cancers combined was observed in the external analysis in the
- 20 large NCI cohort, a statistically significant trend for all lymphohematopoietic cancers was
- 21 observed with peak, but not average or cumulative exposure in the internal analysis
- 22 (SMR = 0.94, 95% CI = 0.84 to 1.06, 286 cases) (Beane Freeman *et al.* 2009). Peak
- exposures exceeding 4 ppm (compared with peaks of > 0.0 to 1.9 ppm) were associated
- 24 with a statistically significant increase in all lymphohematopoietic cancers (OR = 1.37,

25 95% CI = 1.03 to 1.81, 108 deaths).

26 Most studies (except for Dell and Teta 1995, Edling *et al.* 1987b and Bertazzi *et al.* 1986)

- 27 reported results for leukemia. Similar to the findings for all lymphohematopoietic
- 28 cancers, all six studies of health professionals reported increased risks (SMR or PMR) for
- 29 leukemia, although most findings were not statistically significant;. In general, most
- 30 studies reported the highest risks for myeloid leukemia: statistically significant increased

1 mortality for myeloid leukemia was found among white embalmers (PMR = 1.61, 95%) 2 CI = 1.02 to 2.41, 23 deaths) (Hayes *et al.* 1990) and U.S. anatomists (SMR = 8.8, 95%) 3 CI = 1.8 to 25.5, 3 deaths) (Stroup et al. 1986). In the industrial cohort studies, 4 statistically non-significant increased risks for leukemia were found among garment 5 workers in the NIOSH cohort (Pinkerton et al. 2004), U.S. formaldehyde-exposed 6 workers in the NCI cohort (Beane Freeman et al. 2009), Danish women (Hansen and 7 Olsen 1995, 1996), and the subset of tannery workers exposed to formaldehyde (Stern et 8 al. 1987). (See Table 3.7a for risk estimates).

9 A few studies evaluated risk by exposure duration, date of first exposure, or time since 10 first exposure. In the NIOSH cohort (Pinkerton et al. 2004), risks for leukemia, myeloid 11 leukemia, and acute myeloid leukemia were higher among workers with longer duration 12 of exposure (> 10 yrs), longer time since first exposure (> 20 years), and who were 13 exposed prior to 1963 (when formaldehyde exposure was thought to be higher). An 14 excess of mortality for myeloid leukemia among workers with both 10 years or more of 15 exposure and with 20 years since first exposure was 2.55 (95% CI = 1.10 to 5.03, 8) 16 deaths). A statistically significant PMR was found among white embalmers who were 17 licensed greater than 20 years (PMR = 2.21). The NCI cohort study provided the most 18 extensive exposure-response relationship analyses (Beane Freeman et al. 2009). In 19 internal analyses, statistically significant trends were observed for all leukemias ( $P_{\text{trend}} =$ 20 0.02), with peak exposures  $\geq$  4.0 ppm compared with > 0.0 to 1.9 ppm (associated with a 21 relative risk of 1.42 (95% CI = 0.92 to 2.18, 48 deaths); the trend for myeloid leukemia 22 was ( $P_{\text{trend}} = 0.07$ ). No statistically significant trends for leuekemia were observed for 23 average or cumulative exposure. Leukemias observed in the early update by Hauptmann 24 et al. (2003) were re-analyzed by Marsh and Youk (2004) using different exposure 25 assessment methods; effect estimates and exposure-response trends were slightly reduced toward the null and were no longer statistically significant, though risk ratios remained 26 27 elevated for both myeloid leukemia and all leukemias combined.

No increased risks for leukemia were reported in the large cohort of British chemical
workers (Coggon *et al.* 2003), woodworkers in the American Cancer Society Cancer

1 Prevention study (Stellman *et al.* 1998), and iron foundry workers (Andjelkovich *et al.* 

2 1995).

3 Fewer cohort studies reported findings for other types of lymphohematopoietic cancers. 4 [The majority of studies were too small to be able to evaluate these cancers or did not 5 report findings by each subtype.] With respect to Hodgkin's lymphoma, Beane Freeman 6 et al. (2009) reported an increased risk for Hodgkin's lymphoma in their external analysis 7 (SMR = 1.42, 95% CI = 0.96 to 2.10, 25 deaths); in internal analyses, risks increased 8 with increasing peak exposure ( $P_{\text{trend}} = 0.004$ ), and average exposure ( $P_{\text{trend}} = 0.03$ ), but 9 not with cumulative exposure. Statistically significant risks were observed among 10 workers with peak exposure of 2.0 to 3.9 ppm (RR = 3.30, 95% CI = 1.04 to 10.50; 8 11 deaths), peak exposures  $\geq 4.0$  ppm (RR = 3.96, 95% CI = CI = 1.31 to 12.02, 11 deaths), 12 and average exposure for 0.5 to 0.9 ppm (RR = 3.62, 95% CI = 1.41 to 9.31, 9 deaths). 13 Hall et al. (1991) reported a SMR of 1.21 (95% CI = 0.03 to 6.71) based on one observed 14 death among U.K. pathologists. One death was reported among the foundry workers 15 (Andjelkovich et al. 1995). No excess in mortality of Hodgkin's lymphoma was found 16 among the British Chemical workers (Coggon et al. 2003), U.S. garment workers 17 (Pinkerton et al. 2004), Danish workers (Hansen and Olsen et al. 1995, 1996), or in most 18 of the studies of professional workers (Hayes et al. 1990, Stroup et al. 1986, and Walrath 19 and Fraumeni (1983, 1984). [The numbers of exposed cases were small in these studies.] 20 For NHL and other lymphomas, no excess risks were found in most studies (Beane 21 Freeman et al. 2009, Coggon et al. 2003, Hansen and Olsen 1995, 1996, Stellman et al. 22 1998, Stern et al. 1987, Stroup et al. 1986, Walrath and Fraumeni 1983, 1984) with the 23 exception of Hayes et al. (1990), who reported a non-significantly increased PMR for 24 NHL (PMR = 1.26, 95% CI = 0.87 to 1.76, 34 deaths) and Edling *et al.* (1987b), who 25 found 2 cases of lymphoma (vs.1 expected) among workers in the abrasive material 26 industry. Non-significantly increased risks for multiple myeloma were found among 27 highly exposed British chemical workers (SMR = 1.18, 95% CI = 0.48 to 2.44, 7 deaths) 28 (Coggon et al. 2003); abrasive material workers (4 observed vs. 2 expected) (Edling et al. 29 1987b) and U.S. embalmers (PMR = 1.37, 95% CI = 0.84 to 2.12, 20 deaths) (Hayes et 30 al. 1990). In the NCI cohort, relative risk increased with increasing peak exposure, but

1 the trend was not significant, and statistically significant increased risks were also found

2 among workers with peak exposures  $\geq$  4.0 ppm No increased risk was found in the

3 American Cancer Society Cancer Prevention Study (Stellman et al. 1998) (see below for

4 a discussion of the nested-case control study from this cohort conducted by Boffetta *et al.* 

5 1989).

6 3.4.5.2 Case-control studies

7 Ten case-control studies (including three nested case-control studies) were identified that

8 evaluated exposure to formaldehyde and lymphohematopoietic cancers: three studies

9 reported on leukemia, six studies on NHL, one study on Hodgkin's lymphoma, two

10 studies on multiple myeloma, and one study on myelodysplasia (see Table 3-7b). (Some

11 studies evaluated more than one type of lymphohematopoietic cancers.)

12 In a cancer registry-based study of leukemias, Blair et al. (2001) noted an elevated risk

13 for chronic myeloid leukemia (OR = 2.9, 95% CI = 0.3 to 24.5, based on one highly

14 exposed case, and for chornic myeloid leukemia and low-medium exposure to

15 formaldehyde, but not for other histologic subtypes of leukemia, and all leukemia.

16 Partanen *et al.* (1993) found an increase in leukemia among woodworking industry

17 workers (OR = 1.40, 95% CI = 0.25 to 7.01), and Ott *et al.* (1989) reported ORs in excess

18 of 2 for leukemia in association with 3 formaldehyde-exposed deaths.

19 Four population-based studies and two nested case-control studies evaluated

20 formaldehyde exposure and NHL risk, and one study evaluated Hodgkin's lymphoma.

21 Tatham et al. (1997) found slightly elevated but non-significant associations with

formaldehyde exposure and NHL (OR = 1.20, 95% CI = 0.86 to 1.50, 93 cases). Wang *et* 

23 al. (2009) investigate 601 incident cases of NHL among Conecticut women in association

24 with potential occupational exposure to organic solvents, and found a borderline

25 statistically significant association between potential exposure to formaldehyde and NHL

26 (OR = 1.3, 95% CI = 1.1 to 1.7, 203 exposed cases). Risks increased with increasing

27 probability and intensity (combined) of exposure ( $P_{trend} < 0.01$ ). In two U.S. population-

28 based case-control studies, Gérin *et al.* (1989) did not observe a relationship between

29 NHL and estimated duration of exposure to formaldehyde or Hodgkin's lymphoma and

30 ever exposure to formaldehyde in a population-based study in Montreal. In industry-

based studies, Ott *et al.* (1989) reported a 2-fold increase in NHL among ever-exposed workers based on 2 cases, and Partanen *et al.* (1993) found a 4-fold increase in NHL among workers exposed to  $\geq$  3 ppm-months of formaldehyde (OR = 4.24, 95% CI = 0.68 to 26.6, 4 exposed cases). McDuffie *et al.* (2001) did not find increases in the risk of NHL among a subset of individuals in the woodworking industry from a large prospective cancer cohort study in the U.S. and among users of formaldehyde-containing fungicides, respectively. [No quantitative measures of formaldehyde exposure were

8 available in these studies.]

9 Boffetta et al. (1989) reported results for 128 cases of multiple myeloma incidence in a 10 case-control study nested within a large prospective cohort assembled by the American 11 Cancer Society (Stellman et al. 1998). Formaldehyde exposure was estimated for four 12 cases and nine controls, yielding an OR of 1.8 (95% CI = 0.6 to 5.7). Two parallel studies 13 of cases of multiple myeloma were conducted among 835 men (Heineman et al. 1992) 14 and 607 women (Pottern et al. 1992) drawn from all cases reported to the Danish Cancer 15 Registry between 1970 and 1984 for whom occupational data were available from 16 government records. A borderline elevation in risk was observed with probable exposure 17 to formaldehyde (OR = 1.1, 95% CI = 0.7 to 1.6, 41 cases) but not with possible exposure 18 in men; in women, the observed risk was 1.1 (95% CI = 0.8 to 1.6, 56 exposed cases), 19 and 1.6 (95% CI = 0.4 to 5.3, 4 exposed cases) for probable exposure. West *et al.* (1995) 20 noted elevated but stastistically non-significant associations between myelodysplastic 21 syndrome and formaldehyde (ORs ranged from 1.17 to 2.33, 95% CIs not reported); 22 effect estimates tended to increase with increasing cumulative exposure, but no clear 23 exposure-response pattern was observed.

### 24 3.4.5.3 Pooled and meta-analyses

25 Bosetti et al. (2008) conducted a pooled analysis of 12 cohort mortality studies that

analyzed lymphohematopoietic cancers. With respect to all lymphohematopoietic

- 27 cancers, the authors calculated a pooled estimated RR (computed as a weighted average
- of the SMRs and/or PMRs) of 0.85 (95% CI = 0.74 to 0.96, 234 deaths) for industrial
- workers and 1.31 (95% CI = 1.16 to 1.48, 263 deaths) for medical workers. The

1	corresponding pooled RRs for leukemia were 0.90 (95% $CI = 0.75$ to 1.07, 122 deaths)
2	and 1.39 (95% CI = 1.15 to 1.68, 106 deaths), respectively.

3 Two recent meta-analyses have been undertaken to summarize findings across studies 4 investigating occupational exposure to formaldehyde and lymphohematopoietic cancers 5 or leukemia and are reviewed here (Collins and Lineker 2004, Zhang et al. 2009a). (One 6 recent comprehensive review of available studies (Blair *et al.* 2007) is also briefly noted. 7 The meta-analysis conducted by Collins and Lineker included 12 cohort studies 8 (including Hauptmann et al. 2003), four proportionate mortality studies, and two case-9 control studies. Fixed-effects models were used to obtain meta-relative risk values (mRR) 10 and 95% confidence intervals, and random effects models were used to evaluate 11 heterogeneity across studies as a potential indicator of bias, unmeasured confounding, 12 effect modification, or different exposure levels across studies. The meta-analysis found 13 no consistent support for the relationship between formaldehyde exposure and leukemia 14 risk. The mRR across all studies was 1.1 (95% CI = 1.0 to 1.2), and estimates varied by 15 type of study, country of study population, type of industry, year of publication, and 16 study size. Generally, only weak or null mRRs were found for cohort studies (vs. case-17 control), industry-based studies (vs. embalmers and pathologists), studies published after 18 1995, and studies with at least 40 expected cases of leukemia.

19 Zhang et al. (2009a) conducted a meta-analysis of 26 peer-reviewed cohort and/or case-20 control studies that provide data on relative risk estimates and confidence intervals for 21 lymphohematopoietic cancers and formaldehyde exposure, focusing on 15 studies of 22 leukemia. [Note that 6 studies included in either the Collins and Lineker (2004) or Bosetti 23 et al. (2008) meta-analyses were excluded as they either did not include leukemia cases, 24 or had no clear exposed group, or did not report relative risks and/or confidence intervals, 25 or were not peer-reviewed publications]. The meta-analyses were confined to data from 26 occupations known to have high formaldehyde exposure. In addition, results were 27 grouped by subtype of leukemia where possible [Six of the leukemia studies reviewed by 28 the authors reported results by subtype.] Summary risk estimates were calculated using 29 both a fixed effects inverse variance weighting method and a random effects methods; 30 heterogeneity was assessed using a general variance-based method. The results below are

reported for the fixed effects models, which was applied to analyses of each of the types
 of lymphohematopoietic cancers. [Results for random effects models (leukemia only) did
 not differ substantially from those for fixed effects models.]
 The calculated summary mRR for all lymphohematopoietic cancers (19 studies) was 1.25

5 (95% CI = 1.09 to 1.43, *P* value not stated); for Hodgkin's lymphoma (8 studies) the 6 mRR = 1.23 (95% CI = 0.67 to 2.29, *P* not significant); for non-Hodgkin's lymphoma (11 7 studies) mRR = 1.08 (95% CI = 0.86 to 1.35, *P* not significant), and for multiple 8 myeloma (9 studies) mRR = 1.31 (95% CI = 1.02 to 1.67, *P* = 0.02). With respect to 9 leukemia in the 15 studies reviewed, the mRR was significantly elevated at 1.54 (95% CI

10 = 1.18 to 2.00; P < 0.001). The highest risk was observed in association with myeloid

11 leukemia in the 6 studies where subtypes were reported: mRR = 1.90 (95% CI = 1.31 to

12 2.76, P = 0.001) (all 6 studies reported RRs of 1.4 or higher). The authors noted that 51%

13 of the leukemias observed in these studies of formaldehyde exposure were of the myeloid

14 type, of which 64% were acute myeloid leukemia (AML), 19% are of the lymphocytic

15 type, with others of unspecified type. They concluded that the meta-analysis results

16 suggest a causal relationship between formaldehyde and leukemia, and specifically of the

17 myeloid subtype of leukemia.

Blair *et al.* (2007) conducted a comprehensive review of epidemiological studies of the association between chemical exposures and lymphohematopoietic cancers, particularly chronic lymphocytic leukemia (CLL), and concluded that there was some evidence of an association between formaldehyde exposure and leukemia, particularly of the myeloid subtype, but no clear evidence for an association between formaldehyde exposure and CLL, non-Hodgkin's lymphoma, or multiple myeloma.

Reference	Study population and follow up	Risk estimate, 95 exposed cases o	i% CI, number of r deaths	Comments
Andjelkovich et al. 1995	Iron foundry workers, MI, USA N = 3,929 1960–1987	SMR LH Leukemia reticulosarcoma/ lymphsarcoma Hodgkin's diseases	0.59 (0.23–1.21); 7 0.43 (0.05–1.57); 2 0.57 (0.01–3.15); 1 0.72 (0.01–4.00); 1	SMR – formaldehyde exposed subcohort based on national rates
Beane Freeman <i>et al.</i> 2009 (update of Hauptmann <i>et al.</i> 2003)	NCI cohort, USA N = 25,619 Entire cohort 1966- 2004	SMR All LH Hodgkin's NHL All leukemia Lymphatic leukemia Lymphatic leukemia Internal analysis (R All LH malignancie Peak exposure 0.1-1.9  ppm 2.0-3.9  ppm 2.4.0  ppm $P_{\text{trend}}$ Average intensity 0.1-0.4  ppm 0.5-0.9  ppm 2 1.0  ppm $P_{\text{trend}}$ Non-Hodgkin's lymphom Peak exposure 0.1-1.9  ppm 2.0-3.9  ppm 2.0-3.9  ppm 2.0-3.9  ppm 2.0-3.9  ppm 2.0-3.9  ppm 2.0-3.9  ppm 2.0-3.9  ppm 2.1.0  ppm $P_{\text{trend}}$ Average intensity 0.1-0.4  ppm 0.5-0.9  ppm 2.1.0  ppm $P_{\text{trend}}$ Multiple myeloma Peak exposure 0.1-1.9  ppm 2.0-3.9  pp	$\begin{array}{c} 0.94 \ (0.84-1.06); 286\\ 1.42 \ (0.96-2.10); 25\\ 0.85 \ (0.70-1.05); 94\\ 1.02 \ (0.85-1.22); 116\\ 0.90 \ (0.67-1.21); 44\\ a \ 1.15 \ (0.83-1.59); 36\\ R, number of cases)\\ s\\ \hline 1.00; 103\\ 1.17 \ (0.86-1.59); 75\\ 1.37 \ (1.03-1.81); 108\\ 0.04\\ \hline 1.00; 164\\ 1.29 \ (0.97-1.73); 67\\ 1.07 \ (0.78-1.47); 55\\ > 0.50\\ \hline \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ $	Internal analysis adjusted by calendar year, age, sex, race, and pay category; exposure was calculated with a 15-year lag interval No association with cumulative exposure Reanalysis of Hauptmann et al. (2003) data by Marsh and Youk (2004) found significant exposure response relationship for all leukemia and myeloid leukemia for peak exposure, see Section 3.2

Table 3-7a. Summary of cohort studies of formaldehyde exposure and lymphohematopoietic cancers

Reference	Study population and follow up	Risk estimate, 95 exposed cases o	5% CI, number of or deaths	Comments
		$\frac{\text{Average intensity}}{0.1-0.4 \text{ ppm}}$ $0.5-0.9 \text{ ppm}$ $\geq 1.0 \text{ ppm}$ $P_{\text{trend}}$	1.00; 25 1.40 (0.68–2.86); 11 1.49 (0.73–3.04); 12 > 0.50	
		$\begin{array}{l} All \ leukemia\\ \underline{Peak\ exposure}\\ 0.1-1.9\ ppm\\ 2.0-3.9\ ppm\\ \geq 4.0\ ppm\\ P_{trend} \end{array}$	1.00; 41 0.98 (0.60–1.62); 27 1.42 (0.92–2.18); 48 0.020	
		$\frac{\text{Average intensity}}{0.1-0.4 \text{ ppm}}$ $0.5-0.9 \text{ ppm}$ $\geq 1.0 \text{ ppm}$ $P_{\text{trend}}$	1.00; 67 1.13 (0.71–1.79); 25 1.10 (0.68–1.78); 24 0.50	
		$\begin{array}{l} Myeloid \ leukemia\\ \underline{Peak\ exposure}\\ 0.1-1.9\ ppm\\ 2.0-3.9\ ppm\\ \geq 4.0\ ppm\\ P_{trend} \end{array}$	1.00; 14 1.30 (0.58–2.92); 11 1.78 (0.87–3.64); 19 0.07	
		$\frac{\text{Average intensity}}{0.1-0.4 \text{ ppm}}$ $0.5-0.9 \text{ ppm}$ $\geq 1.0 \text{ ppm}$ $P_{\text{trend}}$	1.00; 24 1.21 (0.56–2.62); 9 1.61 (0.76–3.39); 11 0.40	
		<i>Lymphatic leukemic</i> No association with exposure	a peak or average	
Bertazzi <i>et al.</i> 1986	Resin manufacturing plant in Italy N = 1,332 1959-1986	SMR analysis LH Leukemia	2.73 (0.71–3.64); 3 NR	
Coggon <i>et al.</i> 2003 (update of Acheson <i>et al.</i> 1984)	British Chemical Workers Study, UK N = 14,014 1941-2000	SMR analysis Entire cohort LH Multiple myeloma leukemia Hodgkin's disease NHL	NR 0.86 (0.48–1.40); 15 0.91 (0.62–1.29); 31 0.70 (0.26–1.53); 6 0.98 (0.67–1.39); 31	
		Highly exposed Multiple myeloma Leukemia Hodgkin's disease NHL	1.18 (0.48–2.44); 7 0.71 (0.31–1.39); 8 0.36 (0.01–2.01); 1 0.89 (0.41–1.70); 9	

September 3, 2009

Reference	Study population and follow up	Risk estimate, 95 exposed cases of	5% CI, number of or deaths	Comments
Edling et al.	Abrasive materials	Observed/expected.		Small cohort
1987b	industry	LH	NR	
	N = 421 male	Leukemia	NR	
	workers	Lymphoma	2.0 (0.2–7.2); 2	
		Multiple myeloma	4.0 (0.5–14.4); 2	
Hansen and	Danish formaldehyde	SPICR analysis		SPICR adjusted for age
Olsen 1995,	exposed worker	LH	NR	and calendar time
1996	N = 2,041 men, 1,263	Leukemia		
	women	Men	0.8 (0.6–1.6); 39	
	1970–84	Women	1.2 (0.7–1.8); 21	
		NHL	0.0 (0.6, 1.0), 20	
		Men Womon	0.9(0.6-1.2); 32 1.0(0.6, 1.6); 30	
			1.0 (0.0–1.0), 59	
		Hodgkin's disease	$10(05-17) \cdot 12$	
		Women	1.0(0.3-1.7), 12 1.1(0.3-2.7); 4	
Pinkerton <i>et</i>	NIOSH cohort of	SMR analysis		Standardized mortality
al. 2004	garment workers,	LH	0.97 (0.74–1.26): 59	and PMR study
(update of	USA	Leukemia	1.09 (0.70–1.62): 24	
Stayner et al.	N = 11,039	Myeloid leukemia	1.44 (0.80–2.37): 15	
1985, 1988	SMR 1955–98	Hodgkin's disease	0.55 (0.07–1.98); 2	
– PMR and	PMR 1959-82	Reticulosarcoma/		
SMR studies,	11111 1909 02	lymphosarcoma	0.85 (0.28–1.99); 5	
respectively)		Other LH	0.97 (0.64–1.40); 28	
		Exposure duration:	10 + years	
		Leukemia	1.53 (NR); 12	
		Myeloid leukemia	2.19 (NR); 8	
		Acute myeloid		
		leukemia	2.02 (NR); 5	
		Time since first exp	osure: 20+ yrs	
		Leukemia	1.31 (NR); 19	
		Myeloid leukemia	1.91* (NR); 13	
		Acute myeloid		
		leukemia	1.93 (NR); 9	
		10+ yrs duration, 2	<u>0+ yr since first</u>	
		<u>exposure</u>	1 02 (1 09 2 17).15	
		Leukemia Muoloid loukomia	1.92(1.08-3.17);13	
		wyeioid ieukemia	2.33 (1.10-3.03); 8	
		PCMR analyses (90	0% CI)	
		LH	1.44 (0.78–2.44); 10	
		Leukemia & aleuke	mia $1.52(0.52(2.47))$ , 4	
		Other I H	1.32 (0.32 - 3.47); 4	
		Ouler LH	3.42 (1.1/-/.82); 4	
Reference	Study population and follow up	Risk estimate, 95 exposed cases o	5% CI, number of or deaths	Comments
---	---	---	---	--
Stellman <i>et</i> <i>al.</i> 1998	Woodworkers: American Cancer Society Cancer Prevention Study 50 U.S. states, District of Columbia, Puerto Rico N = 362,823 (total cohort); 43, 339 in woodworking activities 1982–86	Formaldehyde only LH Leukemia NHL Multiple myeloma Formaldehyde and LH Leukemia NHL Multiple myeloma	1.22 (0.84–1.77); 28 0.96 (0.54–1.71); 12 0.92 (0.50–1.68); 11 0.74 (0.27–2.02); 4 woodworker 3.44 (1.11–10.68); 3 5.79 (1.44–23.25); 2 2.88 (0.40–20.5); 1 0	Internal analysis using non-woodworkers or workers without exposure to wood dust Adjusted for age and smoking Number of formaldhyde exposed workers not reported See Table 3.3b for nested case-control on multiple myeloma
Stern <i>et al.</i> 1987	Workers employed in two chrome leather tannery plants, USA N = 9,365 1940–79 or 1980	SMR Leukemia and aleukemia Lymphomas	1.25 (0.50–8.58); 7 0.92 (0.37–1.90); 7	Formaldehyde-exposed workers in the finishing department (N not stated)
SMR and PMI	R studies on professiona	l workers (patholog	ists, anatomists, and emb	oalmers)
Hall <i>et al.</i> 1991; (update of Harrington and Shannon 1975, and Harrington and Oakes 1984)	Pathologists, members of professional organizations in the UK N = 3,872 1974–87	SMR analyses (mal and Wales) LH Leukemia Hodgkin's disease	e and female in England 1.44 (0.69–2.65); 10 1.52 (0.41–3.89); 4 1.21 (0.03–6.71); 1	Small cohort
Hayes <i>et al.</i> 1990	Deceased embalmers and funeral directors identified using licensing board records, death certificates, and other sources, USA N = 4,046 1975–85	PMR analyses All subjects LH Hodgkin's disease NHL Multiple myeloma Myeloid leukemia Unspec. leukemia	1.39 (1.15–1.67); 15 0.72 (0.15–2.10); 3 1.26 (0.87–1.76); 34 1.37 (0.84–2.12); 20 1.57 (1.01–2.34); 24 2.28 (1.39–3.52); 20	Small cohort
Levine <i>et al.</i> 1984	Licensed embalmers in Ontario, Canada N = 1,413	SMR analyses LH Leukemia	1.24 [0.53–2.43] <sup>a</sup> ; 8 [1.60] [0.44–4.10]; 4	Small cohort
Stroup <i>et al.</i> 1986	Anatomists, members of the American Association of Anatomists, USA N = 2,317 1888–1979	SMR analyses LH Lymphoma Hodgkin's disease Leukemia Chronic myeloid leukemia	1.2 (0.7–2.0); 18 0.7 (0.1–2.5); 2 0 deaths 1.5 (0.7–2.7); 10 8.8 (1.8–25.5); 3	Small cohort Chronic myeloid leukemia is for 1969– 1979 when subtype data was available

Reference	Study population and follow up	Risk estimate, 95 exposed cases o	i% CI, number of r deaths	Comments
Walrath and Fraumeni 1983	All licensed embalmers and funeral directors in NY, USA N = 1263 1902–80	PMR analyses for w LH Lymphomas Hodgkin's disease Leukemia Myeloid leukemia PMR for non-white Leukemia	white males 1.21 (NR); 25 1.08 (NR); 5 2 vs. 2.3 exp. 1.40 (NR); 12 [1.5] <sup>a</sup> (NR); 6 males NR*; 3 cases	Small cohort
Walrath and and Fraumeni 1984	All licensed embalmers in CA, USA N = 1,109 1916–80	PMR analyses for w LH Lymphomas Hodgkin's disease Leukemia Myeloid leukemia <i>Length of licensure</i> < 20 yrs > 20 yrs	white males 1.22 (NR); 19 [1.0] (NR); 3 0 vs. 2.5 exp. 1.75 (NR); 12 [1.5] <sup>a</sup> (NR); 6 and leukemia 1.24 (NR); 4 2.21* (NR); 8	Small cohort

\* *P* < 0.05.

Results not reported for formaldehyde exposed workers in Dell and Teta (1995).

FOR = formaldehyde; NR = not reported; PMR = proportionate mortality ratio, SMR = standardized mortality ratio SPICR = standardized proportionate incidence cancer ratio. <sup>a</sup>As reviewed by IARC 2006.

Reference	Study population	Exposure assessment	OR or RR (95% cases/controls	CI); exposed	Comments
Gérin <i>et al.</i> 1989 Montreal, Quebec	Multi-site study 1979–85 Cases: men, 206 Non-Hodgkin's lymphoma, 53 Hodgkin's disease, incident cases identified from all hospitals Controls: (1) cancer controls, internal controls with tumors at other sites and (2) 740 population based controls (men) matched by age	Occupational histories and other information obtained by interview; exposure classified by job description and industry <i>Estimated exposure index</i> Low < 0.1 ppm Med. 0.1–1 ppm High $\geq$ 1 ppm	Exposure duration (cancer controls) <sup>a</sup> Non-Hodgkin's lyn < 10 yr/any ≥ 10 yr/ low med. high Hodgkin's disease Ever exposed	(yrs)/exposure index <i>mphoma</i> 0.8 (0.4–1.5); 13/NR 1.3 (0.7–2.4); 15/NR 0.8 (0.5–1.5); 14/NR 0.7 (0.3–1.9); 5/NR 0.5 (0.2–1.2); 8/NR	Adjusted for age, ethnicity, self-reported income, jobs held, and other occupational factors
Ott <i>et al.</i> 1989 United States	Nested case-control of workers chemical manufacturing workers (N = 29,139) 1940–78 Cases: 129 LH (52 NHL, 20 multiple myeloma, 30 non- lymphocytic leukemia, and 18 lymphocytic leukemia) Controls: group matched incidence density sampling by decade first employed and survival	Occupational histories obtained from company employment records and classified using a job exposure matrix	OR for ever exposi NHL Lymphocytic leukemia Non-lymphocytic leukemia	ed 2.0 (NR); 2 2.6 (NR); 1 2.6 (NR); 2	Unadjusted Very few workers exposed to formaldehyde

Table 3-7b. Summary of case-control studies (including nested case-control) investigating formaldehyde exposure and lymphohematopoietic cancers

Reference	Study population	Exposure assessment	OR or RR (95% CI); exposed cases/controls	Comments
Boffetta <i>et al.</i> 1989 United States	Nested case-control study, American Cancer Society Cancer Prevention Study (1982 enrollment) Follow-ups 1982–1986 Cases: 128 incident cases of multiple myeloma Controls: 512 randomly selected incident controls matched on age, ACS region, sex, ethnicity(4:1)	Occupational exposures obtained by questionnaire	OR for history of exposure Multiple myeloma 1.8 (0.6–5.7); 4/9	
Heineman <i>et al.</i> 1992; Pottern <i>et al.</i> 1992 Denmark	Nation-wide cancer registry- based population study 1970–84 Eligible cases: All 1,222 men and 1,010 women with multiple myeloma in Denmark reported to Danish Cancer Registry (1,098 men and 607 women included in study based on availability of occupational data) <i>Controls:</i> 4,888 age-matched men and 4,040 women from state pension fund records (4,169 men and 2,596 women included in study)	Exposures classified by job exposure matrix based on occupational and industry codes	Possible exposure to formaldehyde vs.           never exposed           Men         1.0 (0.8–1.3); 144/527           Women         1.1 (0.8–1.6); 56/235           Probable exposure to formaldehyde vs.           never exposed:           Men         1.1 (0.7–1.6); 41/142           Women         1.6 (0.4–5.3); 4/12	Adjusted for age

Reference	Study population	Exposure assessment	OR or RR (95% CI); exposed cases/controls	Comments
Partanen <i>et al.</i> 1993 Finland	Nested case-control of plywood, particleboard, and formaldehyde glue factory workers (N = 7,303) 1957–1982 Cases: 204 LH cases (NHL, Hodgkin's disease, and leukemia) identified using the Finnish Cancer Registry Controls: 152 controls selected randomly from cohort and matched by year of birth and	Occupational histories obtained from company employment records and classified using plant- specific job exposure matrices	Non-Hodgkin's lymphoma $< 3 \text{ ppm-months}$ $1.00$ $\geq 3 \text{ ppm-months}$ $4.24 (0.68-26.6); 4$ Leukemia $< 3 \text{ ppm-months}$ $1.00$ $\geq 3 \text{ ppm-months}$ $1.40 (0.25-7.91); 2$	Wood dust and solvents not found to be confounders OR for Hodgkin's disease could not be calculated due to small numbers
West <i>et al.</i> 1995 United Kingdom (South East Wales, Wessex, and West Yorkshire)	vital status in 1983 Population-based study, case ascertainment is unclear Cases: 400 cases of myelodysplastic syndrome (> 15 years old) identified from health care records Controls: 400 matched (age, sex, residence, hospital and yr of diagnosis) non-cancer controls selected from out and inpatient clinics	Occupational histories and other information obtained by interview; exposure classified by job description, exposure to a list of specific chemicals, and industry	Hours of lifetime exposure/exposure intensity (low, med., high) Myelodysplasia $\geq 10/any$ 1.17 (NR); 15/13 $\geq 50/>$ med. 2.33 (NR); NR $\geq 2,500/>$ med. 2.00 (NR): NR	Matched pair analysis

Reference	Study population	Exposure assessment	OR or RR (95% CI); expose cases/controls	ed	Comments
Tatham <i>et al.</i> 1997 United States (Atlanta, CT, IA, KS, Miami, San Francisco, Detroit, and Seattle)	Population based study 1984–88 Cases: 1,048 living cases of non- Hodgkin's lymphoma identified using population-based cancer registries Controls: 1,659 frequency matched (registry and date of birth) identified by random digit dialing	Occupational histories and other information obtained by interview; exposure classified by job description and industry	Ever exposedAll NHL1.20 (0.86-Small-cell diffuse1.40 (0.87-Follicular type0.71 (0.41-Large cell diffuse1.10 (0.79-	-1.50); 93 -2.40); 21 -1.20); 17 -1.70); 46	Adjusted for age at diagnosis, ethnicity, education, smoking, marital status, and other factors
Blair <i>et al.</i> 2001 Iowa, Minnesota, United States	Population-based study 1980–84 Cases: 513 leukemia cases (669 eligible cases of leukemia in white men >30 yrs old identified from the Iowa Cancer Registry and hospitals in Minnesota; men with farming as sole occupation excluded; 86% response rate) Controls: 1,087 frequency- matched controls (age, vital status, and residence), identified by random digit dialing, Health Care Financing Administration records, and death certificates. (1,245 eligible)	Occupational histories and other data obtained by interview (present or proxy); exposure classified using a JEM	Exposure intensity         All Leukemia         Low $1.0 (0.7-1.)$ High $0.7 (0.2-2.)$ Acute myeloid leukemia         Low $0.9 (0.5-1.)$ High       NA         Chronic myeloid leukemia         Low $1.3 (0.6-3.)$ High $2.9 (0.3-24)$ Chronic lymphocytic leukemia         Low $1.2 (0.7-1.)$ High $0.6 (0.1-5.)$ Myelodysplasia         Low $0.8 (0.3-1.)$ High       NA	4); 61/128 6); 3/9 6); 14/128 1); 7/128 4.5); 1/9 8); 29/128 3); 1/9 9); 6/128	Adjusted by family history, education, smoking, and hair dye use Urban residents excluded from selection of subjects and farmers excluded from analysis due to higher risk of leukemia

Reference	Study population	Exposure assessment	OR or RR (95% CI); exposed cases/controls	Comments
McDuffie <i>et al.</i> 2001 Canada	Multi-center cancer registry - based incident study of men reporting >10 hr. pesticide use/year Cases: 517 cases of non- Hodgkin's lymphoma for men ≥ 19 years old from six Canadian provinces, identified from cancer registries Controls: 15% of random sample reporting >10 hr pesticide use/yr., identified though mail questionnaire	Occupational histories and other data obtained by mailed and telephone interviews	Exposure to formaldehyde-containing fungicides: OR 0.92 (0.37–2.29); 7/255	Adjusted for age, province, and medical history
Wang <i>et al.</i> 2009 Connecticut, United States	Population-based incident study 1996–2000 Cases: 832 women with histologically confirmed non- Hodgkin's lymphoma diagnosed in Connecticut 21-84 years old, no previous cancer (601 participated) Controls: 717 frequency – matched random digit dialing plus Medicare/Medicaid record sample	Exposures classified using a job exposure matrix based on occupational and industry data obtained from in-person interviews	Ever exposed to formaldehyde:OR $1.3 (1.0-1.7); 203/201$ IntensityLow $1.4 (1.0-1.8); 129/120$ Med-high $1.2 (0.8-1.7); 74/81$ $P_{trend}$ $0.21$ ProbabilityLow $1.3 (1.0-1.7); 165/166$ Med-high $1.4 (0.9-2.3); 38/35$ $P_{trend}$ $0.11$ Probability/intensityMed-high/Med-high $1.6 (0.9-3.1); 24/19$ Large cell-typeever exposed $1.9 (1.3-2.6)$ med-high prob. $2.6 (1.5-4.7); 20$ $P_{trend}$ $< 0.01$	69% of telephone controls and 47% of Medicare/Medicaid sample participated. Matched on age, sex, and Connecticut residence

<sup>a</sup> ORs calculated using cancer controls; similar estimates using population-based controls.

~ . .

1

Concern of the brain and control new rous overtain

1	3.4.6 Cancers of the brain and central hervous system
2	Several cohort mortality studies of health professionals including pathologists,
3	anatomists, and embalmers have reported excess mortality from brain and central nervous
4	system malignancies (Hall et al. 1991, Hayes et al. 1990, Levine et al. 1984, Stroup et al.
5	1986, Walrath and Fraumeni 1983, 1984) (see Section 3.2.8 and Table 3.8). Statistically
6	significant increases were observed among anatomists in the United States (SMR = $2.7$ ,
7	95% CI = 1.3 to 5.0, 10 deaths, compared with U.S. population, and 6.0, 95% CI = 2.3 to
8	15.6 using psychiatrists as a reference) (Stroup et al. 1983), and white male embalmers in
9	New York (SMR = 2.34, 6 deaths) (Walrath and Fraumeni 1983) and California (PMR =
10	1.94, 9 deaths) (Walrath and Fraumeni 1984). Some studies of health professionals
11	reported that longer exposure (as assessed by length of licensure or professional
12	membership) may be associated with brain cancer mortality: higher risks were found
13	among anatomists with professional membership of 40 to 69 years (SMR = 7.0, 95% CI =
14	0.9 to 26.8) for 40 to 60 years vs. between 2 and 2.8 for 1 to 19, and 20 to 39 years).
15	PMRs were also higher among New York embalmers who were> 30 years old (2.94, 5
16	deaths, $P < 0.05$ for > 30 years vs. 0.98, 4 deaths for < 30 yrs) at first license and who
17	had only an embalmers license (PMR = 2.34, $P < 0.50$ for embalmer only vs. 0.93 for
18	embalmer and funeral directors); embalmers are thought to have higher exposure to
19	formaldehyde (Walrath and Fraumeni 1983). All of the brain cancers among anatomists
20	occurred among subjects performing gross or microanatomy.
21	Hauptmann et al. (2004) found no increase in brain and CNS cancers in their external
22	SMR analysis of the NCI cohort; when these cancers were analyzed in internal analyses
23	by average, peak, cumulative, and duration of exposure, no trends with exposure category
24	were observed, and relative risks were generally at or below the reference category (in

25 this study, the lowest exposure group). In general, other cohort studies found no increases

- 26 for brain cancer except small statistically non-significant increases were found in the
- 27 NIOSH and Danish cohorts. In the NIOSH cohort, SMRs were higher (but not
- 28 statistically significant) among workers exposed 20 years since first exposure (SMR =
- 1.20, 13 deaths) and workers whose first exposure was prior to 1963 (SMR = 1.17, 14)
- 30 deaths), but not among workers with the longest duration of exposure (10+ years)
- 31 (Pinkerton *et al.* 2004). No case-control studies evaluating exposure to formaldehyde and

- 1 brain cancer were identified. Bosetti *et al.* (2008) analyzed pooled data from a total of 11
- 2 cohorts that included deaths from brain cancer and calculated a pooled estimated RR of
- 3 0.92 (95% CI = 0.75 to 1.13, 94 deaths) among industrial workers and 1.56 (95% CI =
- 4 1.24 to 1.96, 74 deaths) among health professional workers. [Note that the findings for
- 5 separate studies of health professional workers were significantly heterogeneous,
- 6 according to the authors.]

Reference	Study population and follow up	Risk estimate, 95% CI, number of exposed cases or deaths	Comments
Andjelkovich <i>et al.</i> 1995	Iron foundry workers, MI, USA N = 3,929 1960–87	SMR analysis Brain & CNS 0.62 (0.07–2.23); 2	SMR – formaldehyde exposed subcohort based on national rates
Coggon <i>et al.</i> 2003 (update of Acheson <i>et al.</i> 1984)	British Chemical Workers Study, UK N = 14,014 1941–2000	SMR analysis for brain & CNS           Entire cohort         0.85 (0.57–1.21); 30           High exp.         0.63 (0.25–1.29); 7	
Hansen and Olsen 1995, 1996	Danish formaldehyde exposed workers N = 2,041 men, 1,263 women 1970–84	SPICR analysis for brain and CNS Men 1.1 (0.9–1.5); 54 Women 1.2 (0.8–1.6); 39 Formaldehyde, no exposure to wood dust 1.3 (0.8–1.8); 30	SPICR adjusted for age and calendar time
Hauptmann <i>et al.</i> 2004	NCI cohort, USA N = 25,619 Entire cohort 1966– 94	SMR analysis Brain & CNS 0.92 (0.68–1.23); 43 RR did not increase with increasing peak, average and cumulative exposure, and exposure duration	
Pinkerton <i>et al.</i> 2004	NIOSH cohort of garment workers, USA N = 11,039 SMR 1955–98 PMR 1959–82	SMR analysis for brain & CNS All 1.09 (0.66–1.71); 19 Time since first exposure: 20 + yrs 1.20 (NR); 13 Year of first exposure: prior to 1963 1.17 (NR); 14 No increase risk with increasing duration	Standardized mortality and PMR study
Studies on health p	orofessional workers		
Hall <i>et al</i> . 1991	Pathologists, members of professional organizations in the UK N = 3,872 1974–87	SMR analyses for male and females in England and Wales Brain & CNS 2.18 (0.83–4.75); 6 (all six cases in males)	

Table 3-8. Summary of industrial SMR and PMR studies of formaldehyde exposure and brain and CNS cancers

Reference	Study population and follow up	Risk estimate, 95% CI, number of exposed cases or deaths	Comments
Hayes <i>et al.</i> 1990	Deceased embalmers and funeral directors identified using licensing board records, death certificates, and other sources, USA N = 4,046 1975–85	PMR analyses for brain & CNSWhite1.23 (0.80–1.84); 24Non-whiteNR; 0PMRs were similar between embalmersand funeral directors	Small cohort
Levine <i>et al.</i> 1984	Licensed embalmers in Ontario, Canada N = 1,413	SMR analyses Brain & CNS [1.15] [0.24–3.37] <sup>a</sup> ; 3	Small cohort
Stroup <i>et al</i> . 1986	Anatomists, members of the American Association of Anatomists, USA N = 2,317 1988–79	SMR analyses for brain & CNSReference groupU.S.2.7 (1.3–5.0); 10Psychiatrists6.0 (2.3–15.6); 10Increasing SMRs (U.S. reference) with increasing duration of membership40-49 yr7.0 (0.9–26.8); 2	Small cohort All brain cancers were gliomas
Walrath and Fraumeni 1983	All licensed embalmers and funeral directors in NY, USA N = 1,263 1902–1980	PMR analyses (white males)Brain & CNSAll1.56 (NR); 9Embalmers2.34* (NR); 6Embalmers & funeral directors0.93 (NR); 3Age at first license< 30 yrs	Small cohort
Walrath and Fraumeni 1984	All licensed embalmers in CA, USA N = 1,109 1916–80	PMR analyses for white males Brain 1.94* (NR); 9 > 20 years length of licensure 1.89 (NR); 4	Small cohort

\* *P* < 0.05.

Results not reported for Bertazzi *et al.* (1986), Dell and Teta (1995), Edling *et al.* 1987b, Stellman *et al.* (1998), and Stern *et al.* 1987.

SPICR = standardized proportionate incidence cancer ratio, PMR = proportionate mortality ratio, NR = not reported.

<sup>a</sup>Calculated by IARC.

### 1 3.4.7 Cancer at other sites

2 The association between formaldehyde exposure and cancers of sites other than the head 3 and neck, the respiratory and lymphohematopoietic system, and brain and central nervous 4 system has been examined in both historical cohort and case-control studies. These 5 cancer sites include (but are not limited to): urinary bladder, brain, breast, colo-rectum, 6 esophagus, kidney, liver, oral cavity, pancreas, prostate gland, salivary gland, stomach, 7 and skin or dermis as well as intraocular melanoma. In general, reported estimates were 8 null or slightly elevated but statistically non-significant, and studies have not consistently 9 reported an elevated risk in cancer associated with formaldehyde exposure at any of these 10 sites. The following review primarily focuses on findings of elevated risk for specific 11 solid cancer sites reported in at least two case-control or cohort studies, in addition to 12 statistically significant findings. [Not all cohort studies report findings for all cancer sites, 13 or do not report confidence intervals or p-values. Most of the cohort and case-control 14 studies are of male workers, so that associations between formaldehyde and cancers 15 among women and of the female reproductive system are underrepresented.]

16 Cancers of the gastrointestinal system and associated organs. Several studies have 17 reported small but consistent increases in stomach cancer. Bertazzi et al. (1989, 1986) 18 reported an increase in risk of gastrointestinal cancers in a cohort of resin production 19 workers exposed to formaldehyde (SMR = 1.34, 11 deaths), with stomach cancer risk of 20 1.64 (3 deaths). Coggon et al. (2003) reported a statistically significant increase in the 21 risk of stomach cancer in a large cohort study of plastics and chemical manufacturing 22 workers exposed to formaldehyde (SMR = 1.31, 95% CI = 1.11 to 1.54, 150 deaths), and 23 Stellman et al. (1998) found an elevated risk of stomach cancer among a group estimated 24 to have potential exposure to formaldehyde in an internal analysis of a population-based 25 cohort (RR = 1.69, 95% CI = 0.94 to 2.86, 11 deaths). In addition, Andjelkovich et al. 26 (1995) reported a small increase in stomach cancer in association with formaldehyde 27 exposure in a cohort study of iron foundry workers (SMR = 1.64, 95% CI = 0.82 to 2.94, 28 11 deaths), together with borderline elevations in cancers of the esophagus, large 29 intestine, and rectum. Walrath and Fraumeni (1984) reported an excess of colon cancer 30 among embalmers in California (PMR = 1.87, 30 observed vs. 16 expected deaths, p < 31 (0.05), and in a previous study of embalmers in New York (PMR = 1.43, 29 observed vs.

1	20.3 expected deaths, $P < 0.05$ ) (Walrath and Fraumeni, 1983). Hayes <i>et al.</i> (1990) also
2	reported increases in gastrointestinal cancers combined, including rectum (PMR = $2.31$ ,
3	95% CI = 0.64 to 6.00, 4 deaths) and colon (PMR = 2.31, 95% CI = 1.32 to 3.76, 16
4	deaths), among non-white embalmers; in white, embalmers, non-significant increases
5	were observed. Hansen and Olsen (1995) also reported a statistically significant increase
6	in the risk of colon cancer in association with occupational formaldehyde exposure
7	(standardized proportionate incidence ratio (SPIR) = $1.2$ , $95\%$ CI = $1.1$ to $1.4$ , $166$ cases)
8	in a population-wide study of the Danish Cancer Registry. A subsequent analysis, taking
9	a subgroup of "blue collar" workers with estimated formaldehyde exposure but no wood
10	dust exposure, slightly reduced this risk (SPIR = $1.1$ , 95% CI = $0.9$ to $1.4$ , 73 cases)
11	(Hansen and Olsen 1996). In a population-based case-control study of rectal cancer in
12	men, Dumas et al. (2000) reported a statistically significant increase in this endpoint in
13	association with "substantial" exposure to formal dehyde (OR = 2.4, 95% CI = 1.2 to 1.6,
14	36 deaths). Marginal but statistically nonsignificant increases in this cancer have been
15	noted only in the cohort studies of Walrath and Fraumeni (1984) and Andjelkovich et al.
16	(1990).

An increase in the risk of liver cancer was noted in the population studied by Hansen and Olsen (1996) (SPIR = 1.2, 95% CI = 0.9 to 1.8, 29 cases). Bertazzi *et al.* (1986) reported an increase in the risk of alimentary tract cancer in a cohort of resin production workers exposed to formaldehye (SMR = 1.55, 8 cases), with stomach and esophageal cancer risk of 1.33 (4 cases).

22 *Meta-analyses.* Two meta-analyses have been published summarizing data from multiple 23 studies of pancreatic cancer (Collins et al. 2001a, Ojajärvi et al. 2000). Ojajarvi et al. 24 consolidated epidemiologic data on formaldehyde exposure and pancreatic cancer 25 estimates from two analytic studies and three proportionate mortality studies; the 26 resulting mRR was 0.8 (95% CI = 0.5 to 1.0). Collins et al. reported a similar mRR of 1.1 27 (95% CI = 1.0 to 1.3) using data from 14 studies of workers exposed to formaldehyde 28 where pancreatic cancer rates were reported. The small increase in risk was attributable to 29 embalmers (mRR = 1.3, 95% CI = 1.0 to 1.6) and pathologists and anatomists (mRR = 30 1.3, 95% CI = 1.0 to 1.7). For industrial workers with the highest exposure levels on

average, no increased risk in pancreatic cancer was observed (mRR = 0.9, 95% CI = 0.8
to 1.1). In Section 3.3.6, a case-control study of pancreatic cancer is summarized (Kernan *et al.* 1999) in which some evidence of an increased risk was observed with higher levels
of formaldehyde exposure probability and intensity. The biologic mechanism by which
exposure to formaldehyde could cause pancreatic cancer is unknown (Collins *et al.*2001a).

7 *Cancers of the genitourinary system.* Small but generally statistically non-significant 8 excesses of kidney cancers have been reported in a number of cohort studies. No case-9 control studies of this endpoint have been conducted. In a study of tannery workers, Stern 10 et al. (1987) found only a slight excess of kidney cancers among workers in one 11 department where formaldehyde was used for finishing (SMR = 1.02, 95% CI = 0.26 to 12 2.73, 3 deaths). Hansen and Olsen (1995) reported a borderline statistically significant 13 increase in kidney cancer (SPIR = 1.3, 95% CI = 1.0 to 1.6, 60 cases) among a population 14 with potential occupational formaldehyde exposure in a population-wide Danish Cancer 15 Registry study, and Walrath and Fraumeni (1983, 1984) found an increase in kidney 16 cancers among white male embalmers in New York (PMR = 2.47, 6 observed vs. 2.4 17 expected deaths, P < 0.05) but not among embalmers in California (PMR = 1.00, 4) 18 observed vs. 4 expected deaths).

19 With respect to urinary bladder cancer, cohort studies have not reported excess of this 20 site. Two case-control studies of bladder cancer have been conducted. In a population-21 based study by Siemiatycki et al. (1994) the authors found a marginal increase in bladder 22 cancer in association with "nonsubstantial" exposure to formal dehyde (OR = 1.2, 95% CI 23 = 0.9 to 1.6, 67 exposed cases, adjusted for demographic and lifestyle variables and other 24 occupational exposures) but not with "substantial" exposure (adjusted OR = 0.9, 95% CI 25 = 0.5 to 1.7, 17 exposed cases). In a population-based case-control mortality study of 26 bladder cancers among all male deaths under the age of 50 in the U.K from 1975 to 1979 27 (Coggon *et al.* 1984), no association with occupations with any potential for exposure to 28 formaldehyde was observed (OR = 1.0, 95% CI = 0.7 to 1.3, 132 exposed deaths), and a 29 borderline association with occupations with a high probability of formaldehyde exposure 30 (OR = 1.5, 95% CI = 0.9 to 2.8, 30 deaths).

1 Other cancers. Few other cancers have been reported in excess in the cohort studies. [In a 2 number of studies, the all cause mortality is decreased, suggesting the possibility of a 3 healthy worker effect, which would tend to bias rates based on external population 4 comparisons toward the null.] Walrath and Fraumeni (1983) found a statistically 5 significant increase in skin cancer among white male embalmers in New York state 6 (PMR = 3.26, 5 observed vs. 1.5 expected deaths, p < 0.05); among those who practiced7 both as embalmers and funeral directors, the risk was reduced (PMR 1.44, 3 observed vs. 8 2.1 expected deaths). This finding was not replicated in a subsequent study of white male 9 Californian embalmers (2 observed vs. 3.4 expected deaths (Walrath and Fraumeni 1984) 10 and increases in this cancer risk have not been reported in other studies of embalmers, 11 pathologists or anatomists. Small excesses of prostate cancers were reported in a study of 12 pathologists (Hall et al. 1991) (SMR = 3.30, 95% CI = 0.39 to 11.8, 2 deaths) and in 13 study of embalmers by Hayes *et al.* (1990) (PMR = 1.06, 95% CI = 0.84 to 1.32, 79 14 deaths, white males, and PMR = 1.35, 95% CI = 0.82 to 2.12, 19 deaths, non-white 15 males) but not in other studies of embalmers and anatomists or men in other occupations. 16 Cantor et al. (1995) conducted a population-based case-control study of breast cancer 17 among women in the U.S. using death certificates from 24 states from 1984 to 1989, and 18 coded occupations by probability and intensity of exposure to formaldehyde and other 19 agents. Statistically significant excess of breast cancer were noted among black women 20 with a high probability of exposure (OR = 1.45, 95% CI = 1.2 to 1.7, 311 deaths) or all 21 levels of intensity of exposure (ORs from 1.11 to 1.31, all CIs 1.0 or above); among 22 white women, breast cancer was statistically significantly associated with high intensity 23 of exposure (OR = 1.19, 95% CI = 1.1 to 1.3, 1815 deaths) only.

Finally, a single case-control study of uveal (eye) cancer among white men by Holly *et al.* (1996) reported a statistically significant association with any possible formaldehyde exposure (estimated only by personal interview with subjects) (OR = 2.9, 95% CI 1.2 to 7.0, 3 exposed cases) and a nested case-control study of thyroid gland cancer among female textile workers (Wong *et al.* 2006) found a statistically significant association for 10 or more years of estimated formaldehyde exposure (hazard ratio = 8.33, 95% CI = 1.16 to 6.60, 2 exposed cases). Excesses of thyroid gland cancer have not been reported in other cohort studies, with the exception of a statistically nonsignificant increase in the
cohort study of garment workers by Pinkerton *et al.* (2004) (SMR = 1.16, 95% CI = 0.14
to 4.18, based on only 2 deaths).

#### 4 3.5 Summary

5 A large number of epidemiological studies have evaluated the relationship between formaldehyde exposure and carcinogenicity in humans. The studies fall into the following 6 7 main groups: (1) historical cohort studies and nested case-control studies of workers in a 8 variety of industries that manufacture or use formaldehyde, including the chemical, 9 plastics, fiberglass, resins, and woodworking industries, as well as construction, garment, 10 iron foundry, and tannery workers; (2) historical cohort studies of health professionals, 11 including physicians, pathologists, anatomists, embalmers, and funeral directors; and (3) 12 population-based or occupationally-based case-control incidence or mortality studies of 13 specific cancer endpoints. In addition, several studies have re-analyzed data from specific 14 cohort or case-control studies or have conducted pooled analyses or meta-analyses for 15 specific cancer endpoints

16 The largest study available to date is the combined cohort mortality study of mixed 17 industries conducted by the National Cancer Institute (NCI). This cohort includes 26,561 18 male and female workers, enrolled from ten different formaldehyde-producing or using 19 industries, employed before 1966 and followed most recently to 1994 and 2004, most of 20 the workers were exposed to formaldehyde (Hauptmann et al. 2003, 2004 and Beane 21 Freeman et al. 2009). Quantitative exposure data were used to construct job exposure 22 matrices for individual workers, some of whom experienced peak exposures to 23 formaldehyde > 4 ppm. This cohort is the only study in which exposure-response 24 relationships for peak, average, cumulative, and duration of exposures and mortality for 25 multiple cancer sites were investigated. Two other large cohort studies are available: (1) a 26 large multi-plant cohort study (N = 14,014) of workers in six chemical manufacturing plants in the United Kingdom (Coggon et al. 2003), which calculated SMRs among ever-27 exposed and highly exposed workers for formaldehyde, and (2) a NIOSH cohort of 28 29 garment workers (N = 11,039), which evaluated mortality for duration of exposure, time 30 since first exposure, and year of first exposure to formaldehyde for selected cancer sites.

1 The other cohorts (both for industrial and professional health workers) were smaller, and 2 in general only reported mortality or incidence for ever-exposed workers in external 3 (SMR or PMR) analyses, although some of the studies of professional health workers 4 attempted indirect measures of exposure (such as length in a professional membership) as 5 a proxy for exposure duration. In general, the majority of the nested case-control and 6 other studies attempted to look at exposure-response relationships, but most were semi-7 quantitative. Since most of the cohorts have relatively low statistical power to evaluate 8 rare cancers such as sinonasal and nasopharyngeal cancer, case-control studies are 9 generally more informative for these outcomes. Findings across studies for cancer sites

that have been the principal focus of investigation are summarized below.

### 11 3.5.1 Sinonasal cancers

10

12 There are two major histological types of sinonasal cancer (adenocarcinomas and 13 squamous-cell carcinomas). Sinonasal cancers are rare, and the majority of cohort studies 14 have insufficient numbers of exposed workers to be informative; many of the cohort 15 studies did not report findings or did not observe any deaths for this specific endpoint. 16 Increased risks of sinonasal cancers were observed among male (SPICR = 2.3, 95% CI = 1.3 to 4.0, 13 exposed cases) and female (SPICR = 2.4, 95% CI = 0.6 to 6.0, 4 exposed 17 18 cases) Danish workers exposed to formaldehyde (Hansen and Olsen 1995, 1996) and 19 among formaldehyde-exposed workers in the NCI cohort (SMR = 1.19, 95% CI = 0.38 to 20 3.68, 3 deaths) (Hauptmann et al. 2004). No increase in risk was found among 21 formaldehyde-exposed workers in the large cohort of British chemical workers, based on 22 two observed deaths (Coggon et al. 2003). Of the six case-control studies reviewed, four 23 (Olsen et al. 1994, Olsen and Asnaes 1986, Hayes et al. 1986, Roush et al. 1987, and 24 Luce et al. 1993) reported an association between sinonasal cancers and formaldehyde 25 exposure; statistically significant risks were found in three studies (for ever exposed or 26 individuals with higher measures of exposure) (Olsen et al. 1994, Hayes et al. 1986, Luce 27 et al. 1993). Stronger associations were found for adenocarcinomas, and higher risks of 28 adenocarcinomas were found among individuals with higher average and cumulative 29 exposure, duration of exposure, and earlier dates of first exposure (Luce *et al.* 1993). 30 Wood dust is an established cause of sinonasal cancer, particularly adenocarcinomas 31 (NTP 2005) and is a possible confounder in studies of woodworking industry workers;

1	however, elevated risks for formaldehyde exposure were found among workers with low
2	or no exposure to wood dust (Hayes et al. 1986, Olsen et al. 1994, Olsen and Asnaes
3	1986, Luce et al. 1993) and a possible synergistic effect was suggested in the latter two
4	studies. A pooled analysis of 12 case-control studies of sinonasal cancer from seven
5	countries (Luce et al. 2002) found an increase in adenocarcinomas among formaldehyde-
6	exposed cases, adjusted for wood dust exposure, with increasing level of estimated
7	exposure (OR = $3.0, 95\%$ CI = $1.5$ to $5.7, 91$ exposed cases for men and OR = $1.5, 95\%$
8	CI = 0.6 to 3.8, 6 exposed cases for women; both in the highest exposure groups). For
9	squamous-cell carcinomas, the association with formaldehyde exposure was weaker,
10	except among men with 30 or more years of exposure (OR = 1.4, 95% CI = $0.9$ to 2.3,
11	number of cases not specified; not adjusted for wood dust exposure).

12 3.5.2 Nasopharyngeal cancers

13 As in the case of sinonasal cancer, nasopharyngeal cancers are rare, and the majority of 14 cohort studies have insufficient numbers of exposed workers to be informative. Several 15 cohort studies did not report findings for nasopharyngeal cancer, or observed one or no 16 cases or deaths, for this tumor site. A statistically significant increase in mortality from 17 nasopharyngeal cancer was observed in the large NCI cohort (SMR = 2.10, 95% CI = 18 1.05 to 4.21, 8 exposed cases, one subsequently reclassified as oropharygneal cancer) 19 (Hauptmann et al. 2004). Statistically non-significantly elevated risks were observed 20 among white embalmers from the United States (SMR = 1.89, 95% CI = 0.39 to 5.48, 321 deaths) (Hayes et al. 1990), and among male Danish workers exposed to formaldehyde 22 (SPICR = 1.3, 95% CI = 0.3 to 3.2, 4 exposed cases) (Hansen and Olsen 1995, 1996). In 23 the British chemical workers cohort, one death was observed (SMR not reported) 24 (Coggon *et al.* 2003).

25 Exposure-response relationships between formaldehyde exposure and nasopharyngeal

26 cancers risk were evaluated in the large NCI cohort study. Among seven exposed deaths,

- 27 relative risks of nasopharyngeal cancers increased with peak exposure ( $P_{\text{trend}} < 0.001$ ),
- average exposure ( $P_{\text{trend}} = 0.066$ ) and cumulative exposure ( $P_{\text{trend}} = 0.025$ ); tests for trend
- among combined, exposed, and unexposed workers were  $P_{\text{trend}} = 0.044, 0.126, \text{ and } 0.029,$
- 30 respectively. Adjustment for duration of exposure to a number of potentially confounding

substances did not substantively alter the findings. An analysis adjusted for plant type
found statistically significant trends among exposed workers for peak and cumulative
exposure and duration of exposure. Marsh and colleagues studied one of the plants, in
which five of the nasopharyngeal cancers deaths had occurred, separately (Marsh *et al.*2002, 2007a). These authors also reanalyzed the nasopharyngeal cancers cancer findings
in the NCI cohort (Marsh *et al.* 2007b) and concluded that external employment in metal
working may have partly explained the findings for nasopharyngeal cancers in this

8 cohort.

9 Six of the seven available case-control studies reported increases in nasopharyngeal

10 cancers in association with probable exposure to formaldehyde or at higher levels or

11 duration of estimated exposure (Olsen et al. 1984 [women only], Vaughan et al. 1986,

12 Roush et al. 1987, West et al. 1993, Vaughan et al. 2000, and Hildesheim et al. 2001).

13 Risks of nasopharyngeal cancers increased with exposure duration and cumulative

14 exposure in two population based case-control studies (Vaughan et al. 2000, Hildesheim

15 et al. 2001). In a meta-analysis of case-control and cohort studies (Collins et al. 1997), a

16 statistically significant increased risk for nasopharyngeal cancers and formaldehyde

17 exposure was estimated (mRR = 1.3, 95% CI = 1.2 to 1.5), and a pooled analysis of

18 SMRs from three cohort mortality studies (Bosetti et al. 2008) reported an overall

19 increase in the SMR of 1.33 (95% CI = 0.61 to 2.53, 9 deaths).

20 3.5.3 Other head and neck cancers, and respiratory cancer

21 Most cohort studies reported risk estimates for cancers of the buccal cavity, pharynx,

22 larynx, and lung or combinations of these cancers. Most of these studies, including two of

the three larger cohorts (Pinkerton et al. 2004 and Coggon et al. 2003), three of the

24 professional health worker studies (Hayes *et al.* 1990, Walrath and Freumeni 1983 and

25 1984), and two of the smaller industrial cohorts (Anjelkovich et al. 1995 and Hansen and

Olsen 1995, 1996) found elevated (between approximately 10% and 30%) but

27 statistically non-significant risks for cancers of the buccal cavity or buccal cavity and

28 pharynx combined; risk estimates were usually based on small numbers of deaths or

29 cases. In the NCI cohort, no association between buccal cavity and formaldehyde

30 exposure was observed; however, a statistically significant increased risk for all upper

1 respiratory cancers combined was found among workers with the highest average 2 exposure (> 1 ppm) compared with the lowest exposure group (RR = 2.21, 15 deaths) 3 (Hauptmann et al. 2004). Relative risks increased somewhat with increasing average and 4 peak (but not cumulative) exposure, but the trends were not statistically significant. Most 5 of the case-control studies that reported on head and neck cancers found elevated (usually 6 statistically non-significant) risks for formaldehyde exposure and cancers of the buccal 7 cavity and pharynx (or parts of the pharynx) (Vaughan et al. 1986, Merletti et al. 1991, 8 Gustavsson et al. 1998, Laforest et al. 2000, Marsh et al. 2002, Wilson et al. 2004). 9 Positive exposure-response relationships with probability and duration of exposure for 10 cancers of the hypopharynx and larynx combined were reported by Laforest et al. (2000) 11 and for combined probability and intensity of exposure and salivary cancer by Wilson et 12 al. (2004). No clear association between formaldehyde exposure and hypopharyngeal or 13 laryngeal cancer was observed by Berrino et al. (2003) or for combined head and neck 14 cancers by Tarvainen et al. (2008). Most of the cohort studies and two of the three 15 available case-control studies found no association between formaldehyde exposure and 16 laryngeal cancer. Bosetti et al. (2008) calculated a combined estimated RR (using a 17 weighted average of SMRs and/or PMRs) for combined buccal cavity and pharynx of 18 1.09 (95% CI = 0.88 to 1.34, 88 deaths) among industrial workers and 0.96 (95% CI = 19 0.75 to 1.24, 61 deaths) among health professional workers exposed to formaldehyde in a 20 pooled analysis of 10 occupational cohort mortality studies.

21 Five of the industrial cohort studies reported increases in the risk of lung or respiratory 22 system cancers (Andjelkovich et al. 1995, Bertazzi et al. 1986, Dell and Teta 1995, 23 Hansen and Olsen 1996 [women only]) including the large cohort of British chemical 24 workers, which reported a statistically significant increased risk (SMR = 1.22, 95% CI = 25 1.12 to 1.32, 594 deaths, all workers) (Coggon et al. 2003). In this study, risks increased 26 with increasing exposure level ( $P_{\text{trend}} < 0.001$ ) but not with duration of exposure. No 27 association was observed in the other two large cohorts (Pinkerton et al. 2004, 28 Hauptmann et al. 2004), in several of the smaller occupational cohorts (Hansen and 29 Olsen 1995, 1996 [in men, although a small increase was seen in women], Edling et al. 30 1987b, Stellman et al. 1998, Stern et al. 1987), or in the six studies of health professional 31 workers. Findings from case-control studies were also mixed: statistically significant

1 increased risks were found among fiberglass manufacturing workers who were ever 2 exposed to formaldehyde (OR = 1.61, 95% CI = 1.02 to 2.57, 591 cases) (Marsh *et al.* 3 2001) and among formaldehyde-exposed individuals in a population-based case-control 4 study (Coggon et al. 1984), although risks were not increased among workers with higher 5 exposure. Three studies reported statistically non-significant elevated risks for lung 6 cancer, but no clear exposure response patterns were observed (Gerin et al. 1989, 7 Andjelkovich et al. 1994, Chiazze et al. 1997). No association of lung cancer with 8 formaldehyde exposure was reported in three other occupational case-control studies and 9 one population-based study (Bond et al. 1986, Jensen and Andersen 1982, Partanen et al. 10 1990, Brownson et al. 1993). In a pooled analysis of 14 occupational mortality studies of 11 formaldehyde exposure, which included an analysis of lung cancers, Bosetti et al. (2008) 12 calculated a combined RR of 1.06 (95% CI = 0.92 to 1.23, 1,459 deaths) among 13 industrial workers and 0.63 (95% CI = 0.47 to 0.84, 562 deaths) among health

14 professional workers.

## 15 3.5.4 Lymphohematopoietic cancers

16 Among workers in the NCI cohort study, peak exposure to formaldehyde was associated 17 with increased mortality for several types of lymphohematopoietic cancer (Beane 18 Freeman et al. 2009). With respect to all lymphohematopoietic cancers combined and 19 leukemias, relative risks increased with increasing peak exposure and statistically 20 significant increased risks were found among workers with the highest peak exposure ( $\geq$ 21 4ppm) vs. the lowest exposed category for all lymphohematopoietic cancers (OR = 1.37, 22 95% CI = 1.03 to 1.81, 108 deaths,  $P_{\text{trend}} = 0.02$ ) and statistically non-significant increases 23 in risk were observed for all leukemia and peak exposure  $\geq$  4ppm (RR = 1.42, 95% CI = 24 0.92 to 2.18, 48 deaths,  $P_{\text{trend}} = 0.02$ ) and for myeloid leukemia (RR = 1.78, 95% CI = 25 0.87 to 3.64, 19 deaths,  $P_{\text{trend}} = 0.13$ ). No association was found with cumulative or 26 average exposure. Leukemias observed in the earlier (1984) NCI follow-up (Hauptmann 27 et al. 2003) were re-analyzed by Marsh and Youk (2004) using different exposure 28 assessments; these authors reported no statistically significant trends with exposure, 29 although risks remained elevated for all leukemias (combined) and myeloid leukemia.

1 Increases in all lymphohematopoietic cancers were also observed in other studies. Each 2 of the studies of health professionals found elevated mortality for all 3 lymphohematopoietic cancers combined and for leukemia (Hall et al. 1991, Hayes et al. 4 1990, Stroup et al. 1986, Levine et al. 1984 and Walrath and Fraumeni 1983, 1984). 5 Most estimates were statistically non-significant, except for those of Hayes et al. (1990), 6 and Stroup et al. (1986), where statistically significant excess mortality was found for all 7 leukemia or myeloid leukemia. An excess of leukemia, especially myeloid leukemia, was 8 also found among garment workers in the large NIOSH cohort (Pinkerton et al. 2004), 9 but not in the British chemical workers cohort (Coggon et al. 2003). In the NIOSH 10 cohort, risks for leukemia, myeloid leukemia, and acute myeloid leukemia were higher 11 among workers with longer duration of exposure (> 10 yrs), longer time since first 12 exposure (> 20 years), and among those exposed prior to 1963 (when formaldehyde 13 exposure was thought to be higher). In the smaller industrial cohort studies, some studies 14 reported excesses for lymphohematopoietic cancers combined (Bertazzi et al. 1986, 15 Stellman et al. 1998) or leukemia (Hansen and Olsen 1995, 1996, Stern et al. 1987), but 16 others observed no associations among formaldehyde-exposed workers for all 17 lymphohematopoietic cancers (Pinkerton et al. 2004, Andjelkovich et al. 1995) or 18 leukemia (Stellman et al. 1998). Of the three available case-control studies, a population-19 based study found no association between leukemia and exposure to formaldehyde (Blair 20 et al. 2001), and two nested case control studies reported statistically non-significant 21 increases in risk based on small numbers of exposed cases (Partanen et al. 1993, and Ott 22 et al. 1989).

23 Few cohort studies reported findings for other types of lymphohematopoietic cancers. 24 Most of the cohort studies had relatively low power to detect effects, and either did not 25 report findings or did not evaluate exposure-response relationships. The NCI study was 26 the only cohort that observed an association between formaldehyde exposure and 27 Hodgkin's lymphoma (Beane Freeman et al. 2009). Among exposed workers, relative 28 risks increased with increasing peak ( $P_{\text{trend}} = 0.01$ ) and average exposure ( $P_{\text{trend}} = 0.05$ ), 29 but not with cumulative exposure; statistically significant risks were found for the highest 30 peak ( $\geq$  4.0 ppm) vs. lowest formaldehyde exposure category (RR = 3.96, 95% CI = 1.31 31 to 12.02, 11 deaths). In external analyses, a statistically non-significant elevation in

1	mortality was observed (SMR = 1.4, 95% CI = 0.96 to 2.10, 25 deaths). For non-
2	Hodgkin's lymphoma (NHL), almost all the cohort studies that reported results observed
3	no increases in mortality or incidence. Two nested case-control studies (Partanen et al.
4	1993, Ott et al. 1989) reported increases in NHL risk, but these studies had very small
5	numbers of exposed cases. In the population case-control studies, the risk of NHL
6	increased with increasing probability and intensity combined ( $P < 0.001$ ) in a large U.S.
7	study (Wang et al. 2008), but most of the other studies found no clear association (Gerin
8	et al. 1989, McDuffie et al. 2001, Tatham et al. 1997). For multiple myeloma, peak
9	exposure was associated with a statistically significant increase in risk in the NCI cohort
10	(RR= 2.04, 95% CI = 1.01 to 4.12, 21 deaths, $P_{\text{trend}} = 0.08$ ) (Beane Freeman <i>et al.</i> 2009),
11	and increased risks were seen among British chemical workers (Coggon et al. 2003),
12	abrasive materials workers (Edling et al. 1987b), and U.S. embalmers (Hayes et al.
13	1990). Other studies did not find associations. Small but non-significant increases in risks
14	were also observed in three case-control studies (Boffetta et al. 1989, Heineman et al.
15	1992, Pottern et al. 1992).

16 Bosetti et al. 2008 conducted a pooled analysis of 12 cohort mortality studies and 17 reported a pooled estimated RR for all lymphohematopoietic cancers of 0.85 (95% CI = 18 0.74 to 0.96, 234 deaths) for industrial workers and 1.31 (95% CI = 1.16 to 1.48, 263 19 deaths) for health professional workers. The corresponding pooled RRs for leukemia 20 were 0.90 (95% CI = 0.75 to 1.07, 122 deaths) and 1.39 (95% CI = 1.15 to 1.68, 106 21 deaths), respectively. A meta-analysis by Collins and Lineker (2004) of leukemia and 22 formaldehyde exposure among 12 cohort and case-control studies reported an mRR of 23 1.1 (95% CI = 1.0 to 1.2). Zhang et al. (2009a) conducted a meta-analysis of data from 26 24 studies of occupations with known high formaldehyde exposures, and found an mRR of 25 1.25 (95% CI = 1.09 to 1.43) for all lymphohematopoietic cancers (19 studies), an mRR 26 of 1.31 (95% CI = 1.02 to 1.67, P = 0.02, 9 studies) for multiple myeloma, and an mRR 27 of 1.54 (95% CI =1.18 to 2.00, P < 0.001, 15 studies) for leukemia in association with 28 formaldehyde exposure. The highest risk in the latter group was among myeloid 29 leukemias (mRR = 1.90, 95% CI = 1.31 to 2.76, P = 0.001, 6 studies).

## 1 3.5.5 Other cancer sites

2 In general, few of the cohort studies reported consistently elevated risks for cancers at 3 other sites. [Not all studies reported findings for all cancer sites and few studies included 4 women.] Few case-control studies of other cancer endpoints have been conducted. An 5 excess of mortality from brain and central nervous system cancers have been reported in 6 all six of the cohort studies of health professionals; statistically significant SMR/PMRs 7 (1.68 to 2.7) were reported in three studies (Stroup et al. 1986, Walrath and Fraumeni 8 1983, 1984). Higher risks were found among workers with longer employment as 9 estimated by length of professional membership (Stroup et al. 1986). No increases in 10 these cancers have been observed in the industrial cohort studies that have reported 11 findings, although a small increased risk was reported among garment workers exposed 12 20 years since first exposure (SMR = 1.20, CI not reported, 13 deaths), and among those 13 whose first exposure was prior to 1963 (Pinkerton et al. 2004). A pooled analysis of 14 cohorts by Bosetti et al. (2008) found an increase of 1.56 (95% CI = 1.24 to 1.96, 74 15 deaths) among professional health workers but not among industrial cohorts. 16 Several industrial studies have reported increases in stomach, colon, rectal, and kidney 17 cancers, and a case-control study of pancreatic cancer (Kernan et al. 1999) suggested an 18 increase in this endpoint at higher levels of formaldehyde exposure. Two meta-analyses 19 of pancreatic cancer (Ojajarvi et al. 2000, Collins et al. 2001) showed no consistent 20 increase in risk across studies, with the possible exception of a statistically significant 21 increase among pathologists, anatomists and embalmers.

This Page Intentionally Left Blank

# **4** Studies of Cancer in Experimental Animals

2 The carcinogenic effects of formaldehyde have been investigated in mice (inhalation and 3 dermal administration), rats (inhalation and oral administration), and hamsters (inhalation 4 administration). Although no chronic studies of formaldehyde exposure in primates were 5 found, the effects of formaldehyde on monkeys exposed by inhalation for 1 to 26 weeks 6 have been reported. Several studies also have investigated the interactions or promoting 7 effects of formaldehyde in rodents when administered with other substances. IARC 8 (1995, 2006) reviewed the available data on formaldehyde and concluded that there was 9 sufficient evidence of carcinogenicity in experimental animals. This section is organized 10 by route of administration and species and then discusses the effects of co-exposure with 11 other substances.

## 12 4.1 Inhalation

13 Chronic and subchronic inhalation studies have been conducted in mice, rats, and 14 hamsters. In addition, subacute and subchronic inhalation studies have been conducted in 15 monkeys. All studies were conducted in inhalation chambers (i.e., whole-body rather than 16 nose-only exposure), and formaldehyde vapor usually was generated by heating of 17 paraformaldehyde (see Section 1). Exposure concentrations were reported as parts per 18 million or milligrams per cubic meter of air by the study authors. All tables in this section 19 report concentrations in parts per million. For formaldehyde in air, 1 ppm is equivalent to 20 about 1.23 mg/m<sup>3</sup>.

21 Because of the complexity of nasal anatomy, inhalation studies typically examine

22 multiple transverse sections from four or more anatomical levels of the nasal turbinates in

23 order to determine the location and distribution of lesions. The anatomical levels, nasal

turbinates, and a few other features of the rat nose are illustrated in Figure 4-1. The

25 mouse nose has a similar anatomic structure.



Figure 4-1. Midsagittal section of the rat nose showing the anatomical levels typically examined in inhalation studies.

The Roman numerals identify the positions of the various anatomical levels. The curved dashed lines indicate the junction of the squamous/transitional and respiratory epithelia (anterior line) and the respiratory and olfactory epithelia (posterior line). N = nasoturbinates, M = maxilloturbinates, E = ethmoturbinates, ID = incisive duct, NPD = nasopharyngeal duct, OB = olfactory bulb, 2PR = second palatal ridge.

Source: adapted from Kerns *et al.* 1983 and Mery *et al.* 1994. (Illustration prepared by Donna Jeanne Corcoran, ImageAssociates.)

- 1 4.1.1 Mice
- 2 Horton et al. (1963) conducted a series of experiments in C3H mice to determine whether
- 3 repeated inhalation of formaldehyde would cause bronchiogenic carcinoma and whether
- 4 exposure to formaldehyde would make the mice more susceptible to pulmonary
- 5 carcinoma from subsequent exposure to coal-tar aerosols. Results from the formaldehyde
- 6 experiment are reported here, and results from the formaldehyde plus coal tar experiment
- 7 are discussed in Section 4.3. Groups of 42 to 60 mice [sex and age not reported] were
- 8 exposed to formaldehyde vapor (produced by heating a 2:1 mixture of paraformaldehyde
- 9 and white mineral oil) at a concentration of 0, 50, 100, or 200 mg/m<sup>3</sup> [about 41, 82, or
- 10 163 ppm] for 1 hour/day, 3 days/week, for up to 35 weeks. The low- and medium-
- 11 exposure groups tolerated formaldehyde reasonably well; normal weight gain throughout
- 12 the 35-week exposure period was reported for these groups. However, high mortality was

1 observed in the high-exposure group after the second week. Exposure was discontinued 2 in this group after the eleventh exposure, with only 45 of the 60 original mice surviving. 3 Some mice died of pneumonia, but the authors did not report specific mortality data for 4 each exposure group. No pathological examination of the nasal epithelium was 5 performed. Histological changes in the lungs of all mice that died or were killed during 6 the first 35 weeks are shown in Table 4-1. No statistical analyses were reported. The 7 remaining mice were used in the second experiment (see Section 4.3). No tumors were 8 observed; however, incidences of basal-cell hyperplasia, epithelial stratification, 9 squamous metaplasia, and atypical metaplasia in the trachea and major bronchi were 10 higher in the exposed mice than in the controls. IARC (2006) noted that this study had 11 several limitations, including high doses, short exposure interval, short study duration,

12 and no pathological examination of the nose.

 Table 4-1. Histologic changes in the lungs of C3H mice exposed to formaldehyde by inhalation for up to 35 weeks

		Ν	Incidence [%]				
Conc. (ppm)	Initial	Examined	Basal-cell hyperplasia	Epithelial stratification	Squamous metaplasia	Atypical metaplasia	Lung tumors
0	59	26	0	4 [15]	3 [12]	0	0
[40.8]	60	23	6 [26]	9 [39]	0	0	0
[81.5]	60	34	10 [29]	14 [41]	6 [18]	0	0
[163]	42	35	4 [11]	8 [23]	16 [46]	5 [14]	0

Source: Horton et al. 1963.

13 Kerns et al. (1983) conducted a two-year inhalation study using groups of 119 to 121

14 male and female B6C3F<sub>1</sub> mice and F344 rats (results for the rats are discussed in Section

15 4.1.2.2). Beginning at 6 weeks of age, mice were exposed to formaldehyde at a

16 concentration of 0, 2.0, 5.6, or 14.3 ppm for 6 hours/day, 5 days/week, for up to 24

17 months. After 24 months of exposure, the mice were observed for an additional 6 months

18 without further exposure. Mice were killed at 6, 12, 18, 24, 27, and 30 months for gross

- 19 pathological examinations, hematology, serum chemistry, and urinalysis. Ten animals of
- 20 each sex and group were selected at random for each scheduled sacrifice. All major
- 21 tissues from animals in the control and the high-exposure groups were given thorough
- 22 histological examinations, and multiple sections of nasal turbinates were evaluated in all

groups. Cumulative tumor rates and survival curves were calculated from life-table data
 by the method of Kaplan and Meier. Both unadjusted and adjusted data were analyzed.
 [Data were adjusted to account for differences in time to tumor and survival among the
 groups.] For unadjusted data, exposure groups were compared with Fisher's exact test.
 Overall and pairwise comparisons of adjusted data were made by the methods of Cox and
 Tarone.

7 Female mice in the high-exposure group showed a trend toward lower body weight than 8 the controls after 72 weeks, but body weights returned to normal after exposure stopped. 9 No clear exposure-related effect on body weight was seen in male mice. Survival in the 10 exposed groups was not significantly different from that of the controls; however, 11 survival was slightly lower for exposed male mice from 6 to 24 months. Survival was 12 lower in all groups of males than females, as a result of fighting and infections of the 13 genitourinary tract. The numbers of mice surviving for at least 18 months were 41, 33, 14 32, and 25 males and 89, 83, 92, and 88 females in the control, 2.0-, 5.6-, and 14.3-ppm 15 exposure groups, respectively. Nasal lesions, including inflammation, squamous-cell 16 hyperplasia, metaplasia, and dysplasia, were described as "common" in the nasal mucosa 17 of mice exposed to formaldehyde; however, no incidence data were reported. These nasal 18 lesions were first detected at 12 months in the high-exposure group; by 24 months, more 19 than 90% of mice in this group were affected. The onset, distribution, and severity of 20 these lesions were concentration-dependent. Nasal lesions in the low-exposure group 21 were limited to minimal squamous-cell hyperplasia in a few mice at 24 months. 22 Squamous-cell carcinoma of the nasal cavity occurred in 2 of 17 male mice killed at 24 23 months in the high-exposure group but not in any of the other groups. The authors 24 believed that the carcinoma was caused by formaldehyde exposure, because the 25 spontaneous incidence of these tumors is very low in mice and because the lesions were 26 similar to those observed in rats.

27 4.1.2 Rats

The carcinogenicity of formaldehyde has been studied more extensively in rats than in mice, in four subchronic (4 to 26 weeks) and seven chronic ( $\geq$  1 year) studies. Two of 1 these studies also evaluated the effects in rats of concomitant or sequential exposure to

2 formaldehyde and other substances (discussed in Section 4.3).

3 4.1.2.1 Subchronic studies

4 Rusch et al. (1983) conducted 26-week inhalation studies in monkeys, rats, and hamsters. 5 Results from experiments with monkeys and hamsters are presented in Sections 4.1.3 and 6 4.1.4, respectively. Groups of 20 male and 20 female F344 rats, 7 weeks of age, were 7 exposed to formaldehyde at an average concentration of 0, 0.19, 0.98, or 2.95 ppm for 8 22 hours/day, 7 days/week, for 26 weeks. The target concentrations of 0.20, 1.00, and 9 3.00 ppm were selected to represent environmental exposures to the general public. 10 However, after the first six weeks, the initial high-exposure group was terminated 11 because of uncertainty associated with measurements of exposure concentrations. The 12 high-exposure group was replaced with a new group exposed to a target concentration of 13 3.00 ppm and a corresponding control group. The nasal turbinates, lungs, trachea, and all 14 gross lesions were examined microscopically. No exposure-related effects were seen in 15 the low- and medium-exposure groups. Rats in the high-exposure group showed lower 16 body-weight gain and liver weight than the controls. Incidences of squamous metaplasia 17 and hyperplasia and basal-cell hyperplasia were higher in the high-exposure group than in 18 the controls. No tumors were observed.

19 Groups of 10 male and 10 female albino Wistar rats [age not reported] were exposed to 20 formaldehyde at a concentration of 0, 1, 10, or 20 ppm for 6 hours/day, 5 days/week, for 21 13 weeks (Woutersen *et al.* 1987). Growth retardation was evident in the high-exposure 22 groups of both sexes. Formaldehyde exposure caused an exposure-related increase in the 23 incidences and severity of proliferative lesions in the nasal respiratory and olfactory 24 epithelium, including squamous metaplasia and keratinization.

Feron *et al.* (1988) exposed groups of 45 male Wistar rats [age not reported] to

formaldehyde at a concentration of 0, 10, or 20 ppm for 6 hours/day, 5 days/week, for 4,

- 27 8, or 13 weeks. The primary purpose of this study was to examine the long-term effects
- 28 following relatively short-term exposure to cytotoxic concentrations of formaldehyde.
- 29 Five rats per group were killed at the end of the 4- and 8-week-exposure periods, and 10
- 30 rats per group were killed at the end of the 13-week exposure period. The remaining rats

1 were necropsied when found moribund or dead or were killed at the end of the 2 observation period, during week 131. All rats were examined for gross pathological 3 changes, and six standard cross sections of the nose were examined by light microscopy. 4 Body weight was significantly lower in the high-exposure group than in the controls 5 during the exposure period but returned to normal after about 8, 40, and 100 weeks in 6 groups exposed for 4, 8, and 13 weeks, respectively. Mortality was not significantly 7 different in the formaldehyde-exposed groups than in the controls. Non-neoplastic 8 changes observed in the high-exposure groups included slight to severe hyperplasia and 9 squamous metaplasia of the respiratory epithelium, moderate to severe rhinitis, and 10 varying degrees of squamous metaplasia in the olfactory epithelium. Similar but more 11 focal and less pronounced lesions were observed in the low-exposure group. A total of 14 12 nasal tumors were reported, most occurring in the high-exposure groups (Table 4-2). 13 Although the authors did not report *P*-values for pairwise comparisons, they did consider 14 2 polypoid adenomas, 3 squamous-cell carcinomas, and 1 carcinoma *in situ* observed in 15 groups exposed to 20 ppm for 4 to 13 weeks to be related to formaldehyde exposure. 16 Thus, the incidence of tumors attributed to formaldehyde exposure was 4.5% (6 of 132). 17 IARC (2006) reported that this was significantly higher than the incidence in the controls 18 (P = 0.01, Fisher's exact test) and noted that the positive results occurred even though the 19 exposure duration was short.

Exposure			Incidence [%]				
Duration (wk)	Conc. (ppm)	N	Squamous-cell carcinoma	Polypoid adenoma	Other tumors		
4	0	44	0	0	0		
	10	44	0	0	0		
	20	45	1 [2.2]	1 [2.2] <sup>b</sup>	0		
8	0	45	2 [4.4]	0	0		
	10	44	1 [2.3]	0	0		
	20	43	1 [2.3]	1 [2.3] <sup>b</sup>	0		
13	0	45	0	0	0		
	10	44	1 [2.3]	0	0		
	20	44	3 [6.8] <sup>b</sup>	0	3 [6.8] <sup>c</sup>		

Table 4-2. Neoplastic responses in the nasal cavity of male Wistar rats exposed to formaldehyde by inhalation for 4 to 13 weeks<sup>a</sup>

Source: Feron et al. 1988.

<sup>a</sup>Tumor incidence data are for rats killed immediately after the exposure period, rats that died during the observation period, and rats killed during week 131 at the end of the experiment.

<sup>b</sup>Tumors considered to be associated with formaldehyde exposure.

<sup>c</sup> Tumors included 1 cystic squamous-cell carcinoma, 1 carcinoma *in situ*, and 1 ameloblastoma. The authors considered the carcinoma *in situ* to be related to formaldehyde exposure.

### 1 4.1.2.2 Chronic studies

2 Groups of 120 male and 120 female F344 rats, 7 weeks of age, were exposed to

3 formaldehyde at a concentration of 0, 2.0, 5.6, or 14.3 ppm for 6 hours/day, 5 days/week,

4 for up to 24 months (Kerns *et al.* 1983, Swenberg *et al.* 1980b, Swenberg *et al.* 1980a).

5 Interim sacrifices and histopathological examinations were conducted as described in

6 Section 4.1.1 for B6C3F<sub>1</sub> mice. After 24 months of exposure, the rats were observed for

7 an additional 6 months without further exposure. Swenberg et al. (1980a,b) reported

8 interim results after 18 months of the study, and Kerns et al. (1983) reported the complete

9 results. Statistical analyses were conducted as described above for mice. Compared with

10 the controls, body-weight gain was significantly lower from week 3 to week 103 in both

11 sexes in the medium- and high-exposure groups. Mortality of male and female rats was

12 significantly higher in the high-exposure group than in the controls (P < 0.001). Rhinitis,

- 13 epithelial dysplasia, and squamous metaplasia occurred in all exposed groups, and the
- 14 distribution and severity of these lesions were concentration-dependent. Lesions were
- 15 confined to the nasal cavity and proximal trachea. Neoplastic lesions of the nasal cavity
- 16 were first observed on day 358 in females and day 432 in males. Incidences of neoplastic
- 17 lesions in the nasal cavity are shown in Table 4-3. The incidence of squamous-cell

- 1 carcinoma was significantly higher in the high-exposure groups than in the controls.
- 2 There also was a significant exposure-dependent trend for increased incidence of
- 3 polypoid adenoma in male rats after adjustment for survival differences among groups (P

4 < 0.05).

Table 4-3. Nasal tumors in F344 rats exposed to formaldehyde by inhalation for up to 24 months

			Incidence [%]			
Sex	Exposure (ppm)	N	Squamous-cell carcinoma	Nasal carcinoma	Polypoid adenoma	Other tumors <sup>a</sup>
Male	0	118	0	0	1 [1] <sup>b</sup>	1 [1]
	2.0	118	0	0	4 [3]	0
	5.6	119	1 [1]	0	6 [5]	0
	14.3	117	51 [44]*** <sup>c</sup>	1 [1]	4 [3]	3 [3]
Female	0	114	0	0	0 [0]	0
	2.0	118	0	0	4 [3]	0
	5.6	116	1 [1]	0	0 [0]	0
	14.3	115	52 [45]*** <sup>d</sup>	1 [1]	1 [1]	0

Source: Kerns et al. 1983.

\*\*\*P < 0.001 (compared with controls, Fishers's exact test).

<sup>a</sup>Osteochondroma (controls); 2 undifferentiated carcinomas or sarcomas and 1 carcinosarcoma (highexposure group).

<sup>b</sup>Significant dose-related trend (P < 0.05) after adjustment for survival.

<sup>c</sup>After adjustment for survival, incidence at 24 months was 67%.

<sup>d</sup>After adjustment for survival, incidence at 24 months was 87%.

5 Morgan *et al.* (1986b) reexamined histologic sections from the nasal passages of the rats

6 from the Kerns *et al.* (1983) study to determine the point of origin of the neoplasms. This

7 study showed that the squamous-cell carcinomas developed from the surface epithelium

8 rather than the underlying glandular epithelium. The apparent sites of origin are shown in

9 Table 4-4. The results were assigned accuracy ratings (low or high) based on the degree

10 of confidence assigned by the pathologists. It was more difficult to determine the point of

11 origin of the large tumors that had extensively invaded the nasal cavity than of smaller

- 12 tumors. More than half (57%) of the tumors were found on the anterior portion of the
- 13 lateral aspect of the nasoturbinate and adjacent lateral wall (Levels I and II, see
- 14 Figure 4-1), and 26% were found on the midventral nasal septum (Levels II and III).
- 15 Polypoid adenomas occurred only in a small region of the anterior nasal cavity and were
- 16 restricted to the nasoturbinate, maxilloturbinate, and lateral wall. One of the nasal

- 1 carcinomas was considered a malignant counterpart of the polypoid adenoma and
- 2 originated on the dorsal margin of the maxilloturbinate at Level II. Some neoplasms were
- 3 too large or too poorly preserved to determine their site of origin. All of the apparent sites
- 4 of origin are normally lined by respiratory epithelium.

 Table 4-4. Apparent sites of origin of squamous-cell carcinomas in the nasal

 passages of F344 rats exposed to formaldehyde by inhalation for up to 24 months

	Accuracy rating	Total tumors	% of total carcinomas by area of orgin				
Sex			Area I	Area II	Area III	Area IV	
Male	high low <sup>a</sup>	36 25	56 56	28 20	14 8	3 0	
Female	high	45	62	27	7	4	
	low <sup>b</sup>	15	47	33	13	0	
Total		121	57	26	10	3	

Source: Morgan et al. 1986b.

Area I = lateral aspect of the nasoturbinate and adjacent lateral wall (Levels I and II, see Figure 4-1). Area II = midventral septum (Levels II and III).

Area III = dorsal septum and roof of dorsal meatus (Levels I, II, and III).

Area IV = dorsal and lateral aspect of the maxilloturbinate (Levels II and III).

<sup>a</sup>Unable to determine the site of origin for 4 tumors (16%).

<sup>b</sup>Unable to determine the site of origin for 1 tumor (7%).

5 Appelman *et al.* (1988) conducted a one-year study to determine the role of cytotoxic

6 damage in formaldehyde-induced carcinogenesis in rats. This was followed by a 28-

7 month study of the same design (Woutersen *et al.* 1989). These authors also tested the

8 hypothesis that damage to the nasal mucosa (induced by bilateral electrocoagulation)

9 with subsequent regenerative hyperplasia might enhance the carcinogenic response

10 following exposure to subcytotoxic concentrations of formaldehyde (see Section 5.7.6).

11 These studies are discussed below.

12 Appelman *et al.* (1988) conducted a one-year inhalation study in male albino Wistar rats

13 [age not reported] to study whether damage to the nasal mucosa affected the carcinogenic

14 response to subcytotoxic concentrations of formaldehyde. The anterior third of the nasal

15 mucosa of half of the rats was damaged by electrocoagulation, and after 20 to 26 hours,

16 these rats received their first exposure to formaldehyde. Groups of 10 rats with either

17 damaged or undamaged nasal mucosa were exposed to formaldehyde at a concentration

18 of 0, 0.1, 1, or 10 ppm for 6 hours/day, 5 days/week, for 52 weeks. The exposure

1 concentrations were selected based on 13-week studies showing that formaldehyde was 2 noncytotoxic at a concentration of 2 ppm or lower, slightly cytotoxic at a concentration of 3 3 to 4 ppm, and highly cytotoxic at a concentration of 10 ppm or higher. Some common 4 irreversible lesions associated with electrocoagulation included loss of turbinates and 5 perforation of the nasal septum. Rhinitis and basal-cell hyperplasia and squamous 6 metaplasia of the respiratory epithelium were visible after 13 weeks, but after 52 weeks, 7 effects from electrocoagulation were limited to slight basal-cell hyperplasia and rhinitis. 8 The primary effects of formaldehyde in rats with damaged nasal mucosa included basal-9 cell hyperplasia, squamous metaplasia, and damage to the olfactory epithelium at 10 ppm 10 and focal squamous metaplasia of nasal respiratory epithelium at 0.1 and 1 ppm. No 11 adverse effects were seen in groups of rats with undamaged nasal mucosa exposed to 12 formaldehyde at the two lower concentrations. Rats with undamaged noses in the high-13 dose formaldehyde group had increased incidences of rhinitis, basal-cell hyperplasia, and 14 squamous metaplasia. The authors concluded that rats with damaged noses were more 15 susceptible to the cytotoxic action of formaldehyde.

16 Woutersen et al. (1989) conducted a follow-up of the Appelman et al. (1988) study. A 17 total of 720 male rats [age not reported] were used in the experiment. Half of the animals 18 were exposed to formaldehyde at a concentration of 0, 0.1, 1, or 10 ppm for 3 months and 19 allowed to recover for 25 months, and the other half were exposed for 28 months. Each 20 exposure group included 30 rats with undamaged noses and 60 rats with damaged noses. 21 [The authors did not report why they used unequal numbers of animals in these groups.] 22 All surviving rats were killed at 29 months and examined for gross lesions. Histological 23 examination was limited to six cross sections of the nose. Rats with undamaged noses 24 exposed to formaldehyde at 10 ppm for 28 months had increased incidences of 25 degenerative, inflammatory, and hyperplastic changes of the nasal respiratory and 26 olfactory mucosa, but no tumors. Rats with damaged noses had higher incidences of 27 formaldehyde-induced lesions than did rats with undamaged noses, and the group 28 exposed to formaldehyde at 10 ppm for 28 months had a significantly higher incidence of 29 nasal tumors than the control group (P < 0.001). [The authors did not report P-values; 30 this *P*-value is based on Fisher's exact test conducted by NTP.] Very few tumors

- 1 occurred in the other groups (Table 4-5). The authors concluded that severe damage to
- 2 the nasal mucosa can contribute to formaldehyde carcinogenicity.

Table 4-5. Neoplastic responses in the nasal cavity of male albino Wistar rats, with and without damaged nasal mucosa, exposed to formaldehyde by inhalation for 3 or 28 months

Exposure				Incidence [%]		
Duration (mo)	Group	Conc. (ppm)	N	Squamous-cell carcinoma	Polypoid adenoma	Other tumors
3	undamaged	0	26	0	0	0
		0.1	30	0	0	0
		1	29	0	0	0
		10	26	1 [3.8]	1 [3.8]	0
	damaged	0	57	0	0	0
		0.1	57	2 [3.5]	0	0
		1	53	2 [3.8]	0	0
		10	54	1 [1.9]	0	1 [1.9] <sup>a</sup>
28	undamaged	0	26	0	0	0
		0.1	26	1 [3.8]	0	0
		1	28	1 [3.6]	0	0
		10	26	1 [3.8]	0	0
	damaged	0	54	1 [1.9]	0	0
		0.1	58	1 [1.7]	0	0
		1	56	0	0	0
		10	58	15 [25.9***]	0	2 [3.4] <sup>b</sup>

Source: Woutersen et al. 1989.

\*\*\*[P < 0.001 (compared with controls, Fisher's exact test conducted by NTP)].

<sup>a</sup>Carcinoma *in situ*.

<sup>b</sup>1 adenosquamous carcinoma and 1 adenocarcinoma.

3 Sellakumar et al. (1985) exposed groups of 99 or 100 9-week-old male Sprague-Dawley

4 rats to formaldehyde at a concentration of 15 ppm for 6 hours/day, 5 days/week, for life.

5 This study also investigated the effects of a mixture of formaldehyde and hydrogen

6 chloride [gas] (see Section 4.3.2). A complete necropsy was performed on each animal,

7 with particular attention to the respiratory tract. Multiple cross sections spaced 1.5 to 2

8 mm apart were taken beginning just behind the nostrils and extending back to the orbits.

9 Histologic sections also were prepared from the lungs, trachea, larynx, liver, kidneys,

10 testes, and other organs where gross pathology was observed. After 16 weeks, rats

11 exposed to formaldehyde had markedly lower body weight than controls; however,

12 mortality was not significantly affected by formaldehyde exposure. Nasal tumors, arising
1 from the anterior portion of the nasal cavity, included polyps or papillomas (10 of 100 2 animals examined) and squamous-cell carcinomas (38 of 100 animals examined) in 3 formaldehyde-exposed rats. One fibrosarcoma and one mixed carcinoma also occurred in 4 the exposed group. No nasal tumors were observed in controls. The authors did not 5 statistically compare tumor incidences between these groups; however, IARC (2006) 6 reported that incidences of squamous-cell papilloma and carcinoma were significantly 7 higher than in controls when compared with Fisher's exact test (P = 0.001). No tumors 8 were observed in the trachea or lungs, and tumor incidences in organs outside the 9 respiratory tract did not differ significantly between the exposed and control groups.

10 In a chronic inhalation study conducted by Holmstrom *et al.* (1989a), groups of 16 female 11 Sprague-Dawley rats, 11 weeks of age, were exposed to formaldehyde at a concentration 12 of 0 or 12.4 ppm for 6 hours/day, 5 days/week, for 104 weeks. This study also 13 investigated the effects of combined exposure to formaldehyde and wood dust (see 14 Section 4.3.2). All rats in the formaldehyde-exposed group survived until the end of the 15 study. Body weight did not differ significantly between the two groups. Histological 16 examinations of the nose (five cross sections from the vestibulum of the nose to the 17 posterior ethmoturbinates) and lungs were conducted. Pathological findings in the nasal 18 cavity included pronounced metaplasia or dysplasia in 10 of 16 rats [62.5%] exposed to 19 formaldehyde and none in the control group. One rat in the formaldehyde-exposed group 20 developed squamous-cell carcinoma. Because this type of tumor is not known to occur 21 spontaneously in rats, the authors concluded that it was related to formaldehyde exposure. 22 Pulmonary epithelial histology did not differ significantly between the exposed and 23 control groups. Non-respiratory-tract tumors, primarily mammary-gland tumors, were 24 common in all groups (46% to 53%). Neither the incidence nor the latency period of the 25 non-respiratory-tract tumors was affected by formaldehyde exposure. [IARC (2006) 26 noted the small number of animals used in this study.]

27 Monticello et al. (1996) examined the correlation of cell-proliferation indices with sites

of formaldehyde-induced nasal tumors in male F344 rats. Groups of 90 to 147 rats, 6 to 7

29 weeks of age, were exposed to formaldehyde at a concentration of 0, 0.7, 2, 6, 10, or

30 15 ppm for 6 hours/day, 5 days/week, for up to 24 months. Six rats per group were

1 anesthetized five days before interim sacrifice at 3, 6, 12, and 18 months, and an osmotic 2 pump was surgically implanted subcutaneously over the dorsal thoracolumbar area. Each pump contained 2 mCi of [methyl-<sup>3</sup>H]thymidine, which was administered continuously 3 until sacrifice. Cell proliferation was expressed as the number of <sup>3</sup>H-labeled cell profiles 4 per millimeter of basement membrane and was determined for seven locations in the 5 nasal passages (anterior lateral meatus, posterior lateral meatus, anterior mid-septum, 6 7 posterior mid-septum, anterior dorsal septum, anterior medial maxilloturbinate, and 8 maxillary sinus). Cross-sectional blocks of the nasal cavity were prepared at six levels 9 and processed for histopathology. The distribution of nasal tumors was recorded. 10 Compared with the controls, survival was significantly reduced in the high-exposure 11 group (P < 0.001), but was similar or slightly higher in the three lower-exposure groups. 12 Non-neoplastic lesions (including epithelial hypertrophy and hyperplasia, squamous 13 metaplasia, mixed inflammatory cell infiltrate, nasal turbinate adhesions, and olfactory 14 degeneration) were generally confined to the transitional and respiratory epithelia of the 15 anterior nasal passages and were most severe at the two highest concentrations. The 16 authors stated the tumor response to formaldehyde exposure was highly nonlinear, 17 showing a sharp increase at the two highest exposure levels. A clear exposure-response 18 relationship was observed for squamous-cell carcinoma and polypoid adenoma (Table 4-19 6) [statistics not reported by authors]. Squamous-cell carcinoma was the primary tumor 20 type and occurred most frequently in the lateral meatus and mid-septum. However, many 21 of the tumors were too large for their site of origin to be determined. Other tumors 22 thought to be related to formaldehyde exposure included two nasal rhabdomyosarcomas 23 and two adenocarcinomas which occurred in the two highest dose groups [specific 24 locations not reported]. The population-weighted unit length labeling index (*i.e.*, S-phase 25 nuclei per millimeter of basement membrane × total number of epithelial cells in the site) 26 showed a good correlation ( $r^2 = 0.88$ ) with regional tumor incidence. The authors 27 concluded that target-cell population size, cell proliferation, and local dosimetry are the 28 primary determinants of formaldehyde carcinogenicity.

		In	cidence [%]		Tumor location <sup>b</sup>					
Conc. (ppm)	N	Squamous- cell carcinoma	Polypoid adenoma	Other tumors <sup>a</sup>	Im	ms	amm	ads	unk	
0	90	0	0	0	0	0	0	0	0	
0.7	90	0	0	0	0	0	0	0	0	
2	96	0	0	0	0	0	0	0	0	
6	90	1 [1]	0	0	1	0	0	0	0	
10	90	20 [22.2***]	5 [5.6*]	2 [2.2]	14	0	0	0	6	
15	147	69 [46.9***]	14 [9.5***]	2 [1.4]	26	9	4	3	27	

Table 4-6. Neoplastic responses in the nasal cavity of male F344 rats exposed to formaldehyde by inhalation for up to 24 months

Source: Monticello et al. 1996.

lm = anterior and posterior lateral meatus, ms = anterior and posterior mid-septum, amm = anterior medial maxilloturbinate, ads = anterior dorsal septum, unk = unknown.

\*[P < 0.05 (compared with controls, Fisher's exact test conducted by NTP)].

\*\*\*[P < 0.001 (compared with controls, Fisher's exact test conducted by NTP)].

<sup>a</sup>Rhabdomyosarcoma and adenocarcinoma.

<sup>b</sup>For squamous-cell carcinoma only.

1 Kamata et al. (1997) exposed groups of 32 male F344 rats, 5 weeks of age, to

2 formaldehyde at a concentration of 0, 0.3, 2, or 15 ppm for 6 hours/day, 5 days/week, for

3 up to 28 months. A control group was exposed to methanol at a concentration 4.2 ppm,

4 because the formalin solution used to generate the formaldehyde vapor contained 10%

5 methanol as an antipolymerization agent. An additional room control group was included.

6 Five animals per group were killed at the end of months 12, 18, and 24 for hematological,

7 biochemical, and pathological examination. All animals found dead or moribund were

8 necropsied, and all surviving animals were killed at 28 months. Histopathological

9 examinations were performed on five cross sections of the nasal turbinates and most

10 major organs and tissues. Mortality rates at 28 months were 45.5% and 59.6% in the two

11 control groups, compared with 31.8% in the low-exposure, 55.9% in the medium-

12 exposure, and 88.3% in the high-exposure group. Mortality in the high-exposure group

13 was significantly higher than in the control groups. In addition, the high-exposure group

14 had significantly lower body weight, liver weight, and food consumption than the

15 controls. No lesions related to formaldehyde exposure were observed outside the nasal

- 16 cavity. Incidences of proliferative lesions in the nasal cavity are shown in Table 4-7.
- 17 Epithelial-cell hyperplasia with squamous-cell metaplasia occurred in all groups exposed
- 18 to formaldehyde, and its incidence was significantly higher in the medium- and high-

- 1 exposure groups than in the controls. These lesions did not appear until month 21 in the
- 2 low-exposure group, but appeared as early as month 6 in the high-exposure group.
- 3 Incidences of epithelial-cell hyperkeratosis and squamous-cell carcinoma also were
- 4 significantly elevated in the high-exposure group. Neoplastic lesions were observed only
- 5 in the high-exposure group.

Table 4-7. Proliferative lesions and neoplastic responses in the nasal cavity of male
F344 rats exposed to formaldehyde by inhalation for up to 28 months

		Incidence [%]								
Group (ppm)	N	Epithelial-cell hyperplasia with squamous-cell metaplasia	Epithelial-cell hyperkeratosis	Papillary hyperplasia	Squamous- cell papilloma	Squamous- cell carcinoma				
Controls:										
Methanol	32	0	0	0	0	0				
Room	32	0	0	0	0	0				
0.3	32	4 [12.5]	0	0	0	0				
2	32	7 [21.9]**	1 [3.1]	0	0	0				
15	32	29 [90.6]**	26 [81.3]**	2 [6.3]	3 [9.4]	13 [40.6]**				

Source: Kamata et al. 1997.

\*\* P < 0.01 (compared with methanol control group, Fisher's exact test).

#### 6 4.1.3 Hamsters

7 Two inhalation studies in hamsters, one subchronic and one chronic, were identified. In

- 8 the subchronic study, groups of 10 male and 10 female Syrian golden hamsters, 6 weeks
- 9 of age, were exposed to formaldehyde at an average concentration of 0, 0.19, 0.98, and
- 10 2.95 ppm for 22 hours/day, 7 days/week, for 26 weeks (Rusch *et al.* 1983). All animals
- 11 were killed at 26 weeks. The lungs, nasal turbinates, and trachea were fixed and
- 12 sectioned. No exposure-related mortality or significant toxic effects were seen in any
- 13 exposure group. The formaldehyde-exposed groups showed slightly higher incidences of
- 14 rales, nasal discharge, and lacrimation. None of the hamsters developed tumors.

15 Dalbey (1982) exposed a group of 88 male Syrian golden hamsters [age not reported] to

- 16 formaldehyde at a concentration of 10 ppm for 5 hours/day, 5 days/week, for life. The
- 17 non-exposed control group included 132 hamsters. A second experiment was conducted
- 18 to examine the effect of formaldehyde on diethylnitrosamine (DEN) carcinogenesis (see
- 19 Section 4.3.3). The second experiment also included a group of 50 male hamsters

1 exposed to formaldehyde at 30 ppm once per week, 5 hours/day, for life. Two transverse 2 sections of the nasal turbinates, longitudinal sections of the larynx and trachea, and all 3 lung lobes were examined. Survival time was significantly lower in the 10-ppm group 4 than in the controls (P < 0.05); however, there was very little evidence of toxicity. 5 [Effects on body-weight gain were not reported.] Rhinitis was observed in 31% of the 6 controls, compared with 24% of the 10-ppm exposure group. Hyperplastic and 7 metaplastic lesions of the nasal epithelium occurred in 5% of the 10-ppm group but were 8 not observed in the controls. Weekly exposures to formaldehyde at 30 ppm did not affect 9 mortality. No tumors occurred in either the 10-ppm or 30-ppm exposure group.

#### 10 4.1.4 Monkeys

Rusch *et al.* (1983) exposed six male Cynomolgus monkeys (*Macaca fascicularis*) [age not reported] to formaldehyde for 26 weeks using the same exposure protocol and dose levels as reported above for rats and hamsters. Body weight was not affected by formaldehyde exposure. Squamous metaplasia and hyperplasia was evident in the nasal turbinates of all animals in the high-exposure group. Hoarseness and congestion also occurred in this group. No tumors occurred in the lungs, trachea, or nasal turbinates in any exposure group.

18 Monticello *et al.* (1989) investigated the effects of acute or subacute exposure to 19 formaldehyde on the respiratory tract of rhesus monkeys. Nine young adult male rhesus 20 monkeys (*Macaca mulatta*), aged 4 to 5 years, were randomly divided into three groups. 21 Group 1 (control) was sham exposed to biologically filtered air for 6 hours/day, 22 5 days/week, for 6 weeks. Groups 2 and 3 were exposed to formaldehyde at a 23 concentration of 6 ppm for 1 and 6 weeks, respectively. All animals were tranquilized 18 hours after the last scheduled exposure, injected with  $[^{3}H]$  thymidine (1  $\mu$ Ci/g b.w.), and 24 25 killed 2 hours later. A series of transverse sections of the nose, cross sections of the 26 larynx and mid-trachea, a frontal section of the carina of the trachea, and sections from 27 all lung lobes were examined. In addition, tissues were collected from bone marrow, 28 eyes, adrenal glands, duodenum, esophagus, gall bladder, heart, kidneys, liver, lymph 29 nodes, pancreas, stomach, spleen, and tongue and examined by light microscopy. Five 30 transverse sections from the nasal passages and sections of the larynx, trachea, carina

1 tracheae, lung, and duodenum were processed for histoautoradiography to determine the 2 cell-proliferation rate. Formaldehyde exposure did not significantly affect body weight. 3 Eye irritation and lacrimation were observed in the formaldehyde-exposed groups. 4 Exposure-related effects were observed in the respiratory tract only. Lesions within the 5 respiratory tract were characterized by mild degeneration and squamous metaplasia 6 confined to the transitional and respiratory epithelia of the nasal passages and the 7 respiratory epithelia of the trachea and major bronchi. Although there was little 8 progression of histologic changes from 1 to 6 weeks of exposure, the percent of nasal 9 surface area affected was significantly greater at 6 weeks. Cell-proliferation rates in the 10 formaldehyde-exposed groups were up to 18 times the rates in the control group, with the 11 greatest increase in the anterior nasal cavity. Based on a comparison of the extent of 12 lesions and the cell-proliferation rates observed in this study with those seen in previous studies in rats, the authors concluded that monkeys appeared to be more sensitive than 13 14 rats to the acute and subacute effects of formaldehyde at 6 ppm.

- 15 4.1.5 Summary of inhalation studies
- 16 This section reviewed two inhalation studies in mice, eleven in rats, two in hamsters, and
- 17 two in monkeys. Nasal tumors (primarily squamous-cell carcinoma) were the only
- 18 exposure-related tumors reported. Results from these studies are summarized in
- 19 Table 4-8.

	Exp	osure		Tumor in	ncidence <sup>a</sup>		
Animals	h/d (d/wk)	Duration (wk)	Conc. (ppm)	Male	Female	Results and comments	Reference
Mice (subchro	onic and chi	ronic)					
СЗН	1 (3)	35	0 41 82 166	0, 0, 0, 0,	/26 /23 /34 /35	[Sex and age not reported, examined lung tissue and did not examine nasal tissue], short duration, short exposure time, high mortality in high-exposure group	Horton <i>et al.</i> 1963
B6C3F1	6 (5)	104	0 2.0 5.6 14.3	0/120 0/120 0/120 2/120	0/120 0/120 0/120 0/120	All groups initially contained 119 to 121 animals [number of mice in each group not specifically reported]. Interim sacrifices at 6, 12, 18, 24, and 30 mo. The only tumors occurred in 17 males sacrificed at 24 mo.	Kerns <i>et al.</i> 1983
Rats (subchro	onic)						
F344	22 (7)	26	0 0.19 0.98 2.95	0/20 0/20 0/20 0/20	0/20 0/20 0/20 0/20	[Short duration], increase in squamous metaplasia and basal- cell hyperplasia in high-exposure groups	Rusch <i>et al.</i> 1983
Wistar	6 (5)	13	0 1 10 20	0/10 0/10 0/10 0/10	0/10 0/10 0/10 0/10	[Short duration], exposure-related increase in proliferative lesions of the nasal respiratory and olfactory epithelia, including squamous metaplasia and keratinization	Woutersen <i>et al.</i> 1987
Wistar	6 (5)	13	0 10 20	0/45 1/44 3/44	NT	[Short duration], 1 carcinoma <i>in situ</i> also detected in high- exposure group and thought to be exposure-related	Feron <i>et al.</i> 1988
Wistar	8 (5) 8 (5) 8 (5) 4 <sup>b</sup> (5) 4 <sup>b</sup> (5)	13 13 13 13 13 13	0 1 2 2 4	0/25 0/25 0/25 0/25 0/25	NT	[Short duration], exposure-related effects observed only in high-exposure group and included hyperplasia and squamous metaplasia of the respiratory epithelium	Wilmer <i>et al.</i> 1989

### Table 4-8 Summary of inhalation studies of formaldehyde in experimental animals

	Exp	osure		Tumor i	ncidence <sup>a</sup>		
Animals	h/d (d/wk)	Duration (wk)	Conc. (ppm)	Male	Female	Results and comments	Reference
Rats (chronic)	)						
F344	6 (5)	104	0 2.0 5.6 14.3	0/118 0/118 1/119 51/117	0/114 0/118 1/116 52/115	Nasal carcinoma observed in 1 rat of each sex in the high- exposure groups; polypoid adenoma observed in all groups except female controls and medium-exposure group; undifferentiated carcinoma or sarcoma and carcinosarcoma observed in high-exposure males	Kerns <i>et al.</i> 1983
Wistar	6 (5)	52	0 0.1 1.0 10	0/10 0/10 0/10 0/10	NT	Reported that rats with damaged nasal mucosa were more susceptible to the cytotoxic action of formaldehyde	Appelman <i>et al.</i> 1988
Wistar	6 (5)	117	0 0.1 1.0 10	1/54 1/58 0/56 15/58	NT	Results reported for groups with damaged noses; 1 or 2 nasal tumors also occurred in groups with undamaged noses or in groups exposed for only 3 months	Woutersen <i>et al.</i> 1989
Sprague- Dawley	6 (5)	life	0 15	0/99 38/100	NT	Squamous papilloma observed in 10 rats; mixed carcinoma and fibrosarcoma observed in 1 rat each	Sellakumar <i>et al.</i> 1985
Sprague- Dawley	6 (5)	104	0 12.4	NT	0/15 1/16	[Small number of animals.] Pronounced squamous-cell metaplasia or dysplasia reported in 10 of the exposed rats and none of the controls	Holmström <i>et al.</i> 1989a
F344	6 (5)	104	0 0.7 2 6 10 15	0/90 0/90 0/96 1/90 20/90 69/147	NT	Polypoid adenoma, rhabdomyosarcoma, and adenocarcinoma also observed in the two highest exposure groups. The population-weighted unit length labeling index was correlated with regional tumor incidence.	Monticello <i>et al.</i> 1996
F344	6 (5)	117	0 0.3 2 15	0/32 0/32 0/32 13/32	NT	Squamous-cell papilloma also observed in 3 rats in the high- exposure group	Kamata <i>et al.</i> 1997

	Ехро	osure		Tumor i	ncidence <sup>a</sup>		
Animals	h/d (d/wk)	Duration (wk)	Conc. (ppm)	Male	Female	Results and comments	Reference
Hamsters (subchronic and chronic)							
Syrian golden	22 (7)	26	0 0.19 0.98 2.95	0/10 0/10 0/10 0/10	0/10 0/10 0/10 0/10	[Short exposure duration], no significant responses reported	Rusch <i>et al.</i> 1983
Syrian golden	5 (5) 5 (1)	life life	0 10 30	0/132 0/88 0/50	NT	Minimal increase in hyperplastic and metaplastic areas in the nasal epithelium of exposed animals.	Dalbey 1982
Monkeys (sub	acute and su	ubchronic)					
Cynomolgus	22 (7)	26	0 0.19 0.98 2.95	0/6 0/9 0/6 0/6	NT	[Short exposure duration], squamous metaplasia in the nasal turbinates in the high-dose group	Rusch <i>et al.</i> 1983
Rhesus	6 (5)	6	0 6	0/3 0/3	NT	[Short exposure duration and small number of animals], increased cell-proliferation rates and squamous metaplasia of the transitional and respiratory epithelia of the nasal passages and respiratory epithelia of the trachea and major bronchi	Monticello <i>et al.</i> 1989

NT = not tested.

<sup>a</sup>All tumors are nasal squamous-cell carcinomas unless otherwise noted. <sup>b</sup>Exposed in 30-minute intervals, 8 times/day, separated by 30-minute non-exposure periods.

### 1 **4.2** Oral and dermal administration

2 Formaldehyde was administered to rats via their drinking water in five studies (Soffritti et

3 *al.* 2002a, Soffritti *et al.* 1989, Takahashi *et al.* 1986, Til *et al.* 1989, Tobe *et al.* 1989)

4 and by skin application in one study (Iversen 1986).

## 5 4.2.1 Drinking-water studies

6 Takahashi et al. (1986) investigated the tumor-promoting activity of orally administered

7 formaldehyde on stomach carcinogenesis in 7-week-old male Wistar rats (see Section

8 4.3.2 for a complete description). One group of 10 rats was exposed to formaldehyde in

9 drinking water (0.5% formalin [5,000 mg/L]) from weeks 8 to 40, and a control group of

10 10 rats was given tap-water only. Of 10 formaldehyde-exposed rats, 8 developed

11 squamous-cell papilloma of the forestomach. No tumors occurred in the control group.

12 Til *et al.* (1989) administered formaldehyde (obtained as paraformaldehyde) in drinking

13 water to groups of 70 male and 70 female Wistar rats, aged 5 weeks, for up to 24 months.

14 Target doses were 5, 25, and 125 mg/kg of body weight (b.w.) for both sexes. Average

15 formaldehyde concentrations in the drinking water were 20, 260, and 1,900 mg/L. Based

16 on water consumption, the average daily doses were 0, 1.2, 15, or 82 mg/kg b.w. for

17 males and 0, 1.8, 21, or 109 mg/kg b.w. for females. Subgroups of 10 male and 10 female

18 rats were killed after 12 and 18 months. Formaldehyde exposure did not affect mortality.

19 The high-exposure group of each sex had lower body weight and food intake than the

20 controls, and liquid consumption was about 40% less than in the controls. The high-

21 exposure groups also had severe damage to the gastric mucosa and significantly increased

22 incidences of epithelial hyperplasia and hyperkeratosis of the forestomach and

23 hyperplasia of the glandular stomach (Table 4-9). No tumors were reported at any

24 exposure level.

			Forest	Glandular stomach	
Sex	Dose (mg/kg)	N	Epithelial hyperplasia	Focal hyperkeratosis	Hyperplasia
Male	0	47	1	2	0
	1.2	45	2	6	1
	15	44	1	4	0
	82	47	45***	24***	20***
Female	0	48	1	3	0
	1.8	49	0	5	0
	21	47	2	3	0
	109	48	45***	33***	13***

 Table 4-9. Non-neoplastic responses in Wistar rats given formaldehyde in drinking water for 24 months

Source: Til et al. 1989.

\*\*\*P < 0.001 (compared with controls, Fisher's exact test).

1 Tobe *et al.* (1989) exposed groups of 20 male and 20 female Wistar rats [age not

2 reported] to formaldehyde (obtained as paraformaldehyde) in drinking water for 24

3 months at a concentration of 0, 200, 1,000, or 5,000 mg/L. Based on water consumption,

4 the estimated average daily formaldehyde intakes were 0, 10, 50, and 300 mg/kg b.w.

5 Food intake, water intake, and body weight were significantly lower in the high-exposure

6 groups of both sexes than in the controls. Mortality was 100% in the high-exposure

7 groups by 24 months, occurring as early as 9 days after the beginning of exposure. For

8 males and females, respectively, mortality at 24 months in the other groups was 12.5%

9 and 28.6% in the controls, 46.9% and 33.7% in the low-exposure group, and 0% and

10 14.3% in the medium-exposure group. Non-neoplastic lesions associated with

11 formaldehyde exposure (primarily in the high-exposure group) included erosions, ulcers,

12 hyperkeratosis, basal-cell hyperplasia, and hyperplasia of the squamous epithelium in the

13 forestomach. Similar lesions were observed in the glandular stomach and included

14 erosions and/or ulcers accompanied by submucosal inflammatory-cell infiltrates and

15 glandular hyperplasia. Only a few lesions of the gastrointestinal tract were seen in the

16 medium-exposure groups, and no toxicological effects were observed in the low-

17 exposure groups. Incidences of non-neoplastic lesions were reported only for 6 animals

18 per group at 12 months. All tumors observed (*i.e.*, of the pituitary gland, thyroid gland,

19 testes, adrenal glands, mammary glands, and skin) were the typical spontaneously

1 occurring tumors for this strain. The incidences of these tumors did not differ

2 significantly between the formaldehyde-exposed groups and the controls.

3 Soffritti et al. (1989, 2002a) examined the carcinogenicity of formaldehyde in male and 4 female Sprague-Dawley rats when administered in the drinking water for two years. Oral 5 administration was selected (1) because humans are exposed to formaldehyde in foods 6 and (2) to determine whether formaldehyde might prove to be a multipotential carcinogen 7 (*i.e.*, causing more than one tumor type by various routes of administration). One study examined the effects of age at the start of the experiment (Soffritti et al. 1989). This study 8 9 included two groups of 18 to 20 male and female breeder rats (25 weeks old) exposed to 10 formaldehyde at a concentration of 0 or 2,500 mg/L for up to 104 weeks, and their 11 offspring, initially exposed to formaldehyde *in utero* beginning on gestation day 13. 12 Postnatally, the offspring were exposed to formaldehyde via drinking water at 0 or 13 2,500 mg/L for up to 104 weeks. Survival rates were similar in the exposed and control 14 groups. All animals were necropsied and given a thorough histopathological examination.

15 No exposure-related, non-neoplastic effects were reported for either experiment.

16 Soffritti et al. (1989) reported that formaldehyde exposure was associated with a slight 17 increase in hemolymphoreticular neoplasms in male and female breeder rats (Table 4-10). 18 Gastrointestinal-tract tumors occurred in two breeder rats but were more prevalent in 19 their offspring. These included both benign tumors (adenoma, papilloma, and acanthoma) 20 and malignant tumors (adenocarcinoma and leiomyosarcoma). Leiomyosarcoma was the 21 most frequent malignant tumor. The authors noted that these gastrointestinal tumors were 22 very rare in the historical controls from the colony used in these experiments and that 23 none of these tumors were observed in the concurrent controls. [No statistical analyses 24 were reported for these results.] IARC's (2006) review of this study reported that the 25 incidence of leiomyosarcoma in the intestine was significantly increased in the exposed female offspring alone and in exposed female and male offspring combined ( $P \le 0.01, \chi^2$ 26 27 test) and that the incidence of malignant intestinal tumors in the female offspring was 28 significantly higher than in controls (pairwise comparisons with Fisher's exact test).

			Incidence (%)					
	Conc.		Hemolympho-	Sto	Stomach		Intestine	
Sex	(mg/L)	Ν	reticular	Benign	Malignant	Benign	Malignant	
М	0	20	0 (0)	0 (0)	0 (0)	0 (0)	0 (0)	
Μ	2,500	18	2 (11.1)	0 (0)	1 (5.6)	0 (0)	0 (0)	
F	0	20	1 (5)	0 (0)	0 (0)	0 (0)	0 (0)	
F	2,500	18	2 (11.1)	1 (5.6)	0 (0)	0 (0)	0 (0)	
М	0	59	3 (5.1)	0 (0)	0 (0)	0 (0)	0 (0)	
Μ	2,500	36	4 (11.1)	1 (2.8)	2 (5.6)	1 (2.8)	1 (2.8)	
F	0	49	3 (6.1)	0 (0)	0 (0)	0 (0)	0 (0)	
F	2,500	37	0 (0)	0 (0)	2 (5.4)	0 (0)	6 (16.2)**	
	Sex M M F F M M F F F	Sex         Conc. (mg/L)           M         0           M         2,500           F         0           F         2,500           M         0           F         0           F         2,500           M         2,500           F         0           F         0           F         0           F         0           F         0           F         0           F         0           F         0           F         2,500	SexConc. (mg/L)NM020M2,50018F020F2,50018M059M2,50036F049F2,50037	Sex         Conc. (mg/L)         N         Hemolympho- reticular           M         0         20         0 (0)           M         2,500         18         2 (11.1)           F         0         20         1 (5)           F         2,500         18         2 (11.1)           M         0         59         3 (5.1)           M         2,500         36         4 (11.1)           F         0         49         3 (6.1)           F         2,500         37         0 (0)	Sex         Conc. (mg/L)         N         Hemolympho- reticular         Benign           M         0         20         0 (0)         0 (0)           M         2,500         18         2 (11.1)         0 (0)           F         0         20         1 (5)         0 (0)           F         2,500         18         2 (11.1)         1 (5.6)           M         0         59         3 (5.1)         0 (0)           M         2,500         36         4 (11.1)         1 (2.8)           F         0         49         3 (6.1)         0 (0)           F         2,500         37         0 (0)         0 (0)	Sex $N$ $Hemolympho-reticularIncidence (%)M0200(0)BenignMalignantM0200(0)0(0)0(0)M2,500182(11.1)0(0)1(5.6)F0201(5)0(0)0(0)F2,500182(11.1)1(5.6)0(0)M0593(5.1)0(0)0(0)M2,500364(11.1)1(2.8)2(5.6)F0493(6.1)0(0)0(0)F2,500370(0)0(0)2(5.4)$	Sex         Conc. (mg/L)         N         Hemolympho- reticular         Incidence (%)           M         0         20 $0(0)$ $Benign$ Malignant         Benign           M         0         20 $0(0)$ $0(0)$ $0(0)$ $0(0)$ $0(0)$ M         2,500         18 $2(11.1)$ $0(0)$ $1(5.6)$ $0(0)$ F         0         20 $1(5)$ $0(0)$ $0(0)$ $0(0)$ F         2,500         18 $2(11.1)$ $1(5.6)$ $0(0)$ $0(0)$ M         0         59 $3(5.1)$ $0(0)$ $0(0)$ $0(0)$ M         2,500         36 $4(11.1)$ $1(2.8)$ $2(5.6)$ $1(2.8)$ F         0         49 $3(6.1)$ $0(0)$ $0(0)$ $0(0)$ F         2,500 $37$ $0(0)$ $0(0)$ $2(5.4)$ $0(0)$	

Table 4-10. Tumor incidences in Sprague-Dawley rats exposed to formaldehyde in drinking water at two different ages for up to 104 weeks

Source: Soffritti et al. 1989.

\*\*P < 0.01 (compared with controls,  $\chi^2$  test conducted by IARC 2006); [no statistical analyses were reported by the study authors.]

<sup>a</sup>Transplacental exposure beginning on gestational day 13, then postnatal exposure continued via drinking water.

1 In the second experiment conducted by Soffritti and co-workers, groups of 50 male and

2 50 female rats, 7 weeks of age, were exposed to formaldehyde at a concentration of 10,

3 50, 100, 500, 1,000, or 1,500 mg/L for 104 weeks and then observed for life (Soffritti *et* 

4 *al.* 1989, 2002a). The formalin solution used to prepare the test solutions contained 30%

5 formaldehyde and 0.3% methanol. All animals died by week 163. Additional groups of

6 50 male and 50 female rats were exposed to methanol at a concentration of 15 mg/L,

7 because methanol was used in the formaldehyde solution as a stabilizer. [Based on a

8 concentration of 0.3% methanol in the stock solution, the concentrations of methanol in

9 the formaldehyde test solutions ranged from about 0.1 to 15 mg/L.] The control group

10 included 100 male and 100 female rats given tap water only.

11 No exposure-related non-neoplastic effects were reported. Tumor incidences were

12 analyzed with the  $\chi^2$  test, and dose-response relationships with the Cochrane-Armitage

13 test for trend. The authors did not report statistical comparisons between the

14 formaldehyde-exposed groups and the methanol group; however, IARC (2006) conducted

15 statistical analyses for trend and incidence between these groups (results presented

16 below). The incidence of total malignant tumors was significantly higher in male rats

17 exposed to formaldehyde at 1,500 mg/L than in the unexposed controls. The total number

1 of malignant tumors per 100 animals was significantly increased in males at 500 or

- 2 1,500 mg/L and in females at 100, 1,000, or 1,500 mg/L (Table 4-11). [The NTP
- questioned the appropriateness of applying a  $\chi^2$  test (which is designed for dichotomous 3
- response data) to tumor counts such as total number of tumors per 100 animals. There is 4
- also concern that the authors'  $\chi^2$  test considered the individual tumor rather than the 5
- animal as the experimental unit and did not take into account the variability in tumor 6
- 7 response among animals.]

Sex	Concentration (mg/L)	Ν	Tumor-bearing animals (%)	Total no. tumors (per 100 animals) <sup>ª</sup>
Male	0	100	38 (38)	50 (50)
	methanol only	50	21 (42)	29 (58)
	10	50	14 (28)	19 (38)
	50	50	12 (24)	15 (30)
	100	50	22 (44)	23 (46)
	500	50	24 (48)	36 (72)*
	1,000	50	23 (46)	30 (60)
	1,500	50	36 (72)**	56 (112)**
Female	0	100	43 (43)	49 (49)
	methanol only	50	23 (46)	32 (64)
	10	50	20 (40)	22 (44)
	50	50	20 (40)	26 (52)
	100	50	25 (50)	41 (82)**
	500	50	19 (38)	25 (50)
	1,000	50	29 (58)	39 (78)**
	1,500	50	27 (54)	48 (96)**

Table 4-11. Total malignant tumors in Sprague-Dawley rats exposed to formaldehyde in drinking water for up to 104 weeks

Source: Soffritti et al. 2002a.

\*P < 0.05, \*\*P < 0.01 (compared with controls,  $\chi^2$  test). <sup>a</sup> [The NTP questioned the validity of the  $\chi^2$  test for these data (see text).]

8 An exposure-related increase in the incidence of hemolymphoreticular neoplasms

9 (including lymphoblastic leukemia and lymphosarcoma, immunoblastic lymphosarcoma,

- 10 other leukemias, and hemolymphoreticular sarcoma) was reported in male and female
- 11 rats exposed to formaldehyde (Soffritti et al. 2002a). The incidence of
- 12 hemolymphoreticular neoplasms was significantly increased in males at concentrations of
- 13 100 mg/L or higher and in females at the two highest concentrations (Table 4-12a). The
- 14 incidence of hemolymphoreticular neoplasms was higher in males exposed to methanol

1	only than in the control group, but the difference was not reported as statistically
2	significant. IARC (2006) also reported a significant increase in total malignant
3	mammary-gland tumors (adenocarcinoma, fibrosarcoma, liposarcoma, and
4	angiosarcoma) in females (100, 1,000, and 1,500 mg/L) and testicular interstitial-cell
5	adenoma in males (500, 1,000, and 1,500 mg/L) (Table 4-12a). Most of the mammary-
6	gland tumors in female rats were adenocarcinomas. Several stomach and intestinal
7	tumors, including a few of the very rare leiomyomas or leiomyosarcomas, were observed
8	in some of the formaldehyde-exposed groups but not in the methanol or control groups
9	(Table 4-12b). IARC (2006) statistical analyses showed that when compared with the
10	methanol-only group, the formaldehyde-exposed rats had significantly higher total
11	numbers of tumor-bearing animals, incidence of hemolymphoreticular tissue tumors in
12	high-exposure males, and incidence of testicular interstitial-cell adenoma in the medium-
13	exposure males ( $P < 0.01$ ). A significant exposure-response relationship also was found
14	for the increased incidences of hemolymphoreticular tumors in males. IARC noted the
15	pooling of lymphoma and leukemia as hemolymphoreticular neoplasia, the lack of
16	reporting of non-neoplastic lesions, and the absence of information on incidences of
17	hemolymphoreticular tumors in historical controls in this study.

			Incidence (%)						
			Mammary gland						
Sex	Conc. (mg/L)	N	Adeno- carcinoma	Fibro- sarcoma	Lipo- sarcoma	Total <sup>ª</sup>	Testes	Hemolymphoreticular	
Male	control	100	1 (1)	0	0	1 (1)	10 (10)	8 (8)	
	methanol	50	0	1 (2)	0	1 (2)	3 (6)	10 (20)	
	10	50	0	0	0	0	3 (6)	4 (8)	
	50	50	0	0	0	0	6 (12)	10 (20)	
	100	50	0	0	1 (2)	1 (2)	6 (12)	13 (26)**	
	500	50	0	0	0	$1(2)^{b}$	10 (20)*	12 (24)*	
	1,000	50	0	0	0	0	12 (24)* <sup>c</sup>	11 (22)*	
	1,500	50	1 (2)	0	0	1 (2)	9 (18)*	23 (46)** <sup>d</sup>	
Female	control	100	11 (11)	0	0	11 (11)	_	7 (7)	
	methanol	50	7 (14)	0	1 (2)	8 (16)	_	5 (10)	
	10	50	2 (4)	1 (2)	0	3 (6)	_	5 (10)	
	50	50	4 (8)	0	1 (2)	5 (10)	_	7 (14)	
	100	50	8 (16)*	2 (4)	0	10 (20)	_	8 (16)	
	500	50	3 (6)	1 (2)	2 (4)	6 (12)	_	7 (14)	
	1,000	50	9 (18)*	1 (2)	0	10 (20)	_	11 (22)*	
	1,500	50	11 (22)*	0	1 (2)	12 (24)* <sup>e</sup>	_	10 (20)*	

Table 4-12a. Incidences of mammary, testicular, and hemolymphoreticular tumors in Sprague-Dawley rats exposed to formaldehyde in drinking water for up to 104 weeks

Source: Soffritti et al. 2002a, IARC 2006.

\*P < 0.05, \*\* P < 0.01 (compared with controls,  $\chi^2$  test).

<sup>a</sup>IARC noted that this category is an aggregate of tumors of different cellular origins.

<sup>b</sup>Angiosarcoma also reported in 1 rat.

<sup>c</sup>Significantly different from the methanol control group (P < 0.01, 2-tailed Fisher's exact test conducted by IARC).

<sup>d</sup>Significantly different from the methanol control group (P < 0.01,  $\chi^2$  test conducted by IARC).

			Incidence (%)						
	Conc.		Stomach- lei	iomyosarcomaª	Intestine				
Sex	(mg/L)	Ν	Forestomach	Glandular stomach	Leiomyoma <sup>a</sup>	Leiomyosarcoma <sup>a</sup>			
Male	control	100	0	0	0	0			
	methanol	50	0	0	0	0			
	10	50	1 (2)	0	0	0			
	50	50	0	0	0	0			
	100	50	0	0	0	0			
	500	50	0	0	0	0			
	1,000	50	0	1 (2)	0	0			
	1,500	50	0	0	0	2 (4)			
Female	control	100	0	0	0	0			
	methanol	50	0	0	0	0			
	10	50	0	0	2 (4) <sup>b</sup>	0			
	50	50	0	0	1 (2)	1 (2)			
	100	50	0	0	0	0			
	500	50	0	0	0	0			
	1,000	50	0	0	0	0			
	1,500	50	0	0	3 (6)	0			

Table 4-12b. Incidences of stomach and intestinal tumors in Sprague-Dawley rats exposed to formaldehyde in drinking water for up to 104 weeks

Source: Soffritti et al. 2002a, IARC 2006.

<sup>a</sup>Statistical analyses were not provided for these tumors, which were reported as being very rare in Sprague-Dawley rats [not significantly different from controls, Fisher's exact test conducted by NTP].

<sup>b</sup>IARC 2006 reported only 1 tumor (2%) for this group, without an explanation.

#### 1 4.2.2 Skin application

2 Formaldehyde is widely used in laboratories as a fixative for tissue; therefore, researchers 3 and technicians may be chronically exposed by skin contact. Iversen (1986) conducted 4 skin-painting experiments with hairless Oslo mice to test the potential carcinogenic 5 potency of formaldehyde at concentrations typically used in pathology laboratories. Two 6 groups of 16 male and 16 female mice [age not reported] received two weekly topical 7 applications of 200 µL of aqueous solutions of 1% or 10% formaldehyde for up to 60 8 weeks. Formaldehyde was also tested as a skin-tumor promoter (see Section 4.3.1). 9 Mortality was not increased in groups exposed to 1% or 10% formaldehyde. No lesions 10 were observed in the mice exposed to 1% formaldehyde, while mice in the 10% 11 formaldehyde group had slight hyperplasia of the epidermis. The author concluded that 12 1% or 10% formaldehyde applied to the skin of hairless mice did not have an observable 13 carcinogenic effect. IARC (2006) noted that there was no water-only control group. [This 14 study is also limited by the small number of animals and less-than-lifetime exposure 15 duration.]

16 4.2.3 Summary of oral and dermal exposure studies

17 Five drinking-water studies and one skin-painting study of the carcinogenicity of

18 formaldehyde were reviewed. Ingestion of formaldehyde at high concentrations was

19 associated with gastrointestinal-tract tumors in two studies in rats. One study reported

20 increased incidences of total malignant tumors, testicular tumors, malignant mammary-

21 gland tumors, and hemolymphoreticular tumors. No tumors were observed in the skin-

22 painting study in mice. Results from these studies are summarized in Table 4-13.

Exposure			Gastrointestinal tumor incidence			
	Duration (wk)	ration wk)         Conc. (mg/L)           32         0 5,000           04         0	Male	Female	Results and comments	Reference
	32	0 5,000	0/10 8/10	NT	Forestomach papilloma	Takahashi <i>et al.</i> 1986
	104	0 20 260 1,900	0/70 0/70 0/70 0/70	0/70 0/70 0/70 0/70	Rats in the high-concentration groups had extensive damage to the gastric mucosa and an increase in proliferative lesions of the forestomach and glandular stomach.	Til <i>et al.</i> 1989
	104	0 200 1,000 5,000	0/20 0/20 0/20 0/20	0/20 0/20 0/20 0/20	No exposure-related tumors. Increased proliferative lesions and ulcers of the forestomach and glandular stomach in high-concentration group. High mortality in high- concentration groups.	Tobe <i>et al.</i> 1989
	104	0 2,500	0/20 1/18	0/20 1/18	Two hemolymphoreticular tumors in each exposed group; one in female controls.	Soffritti <i>et al.</i> 1989
	104	0 2,500	0/59 5/36 <sup>c</sup>	0/49 8/37 <sup>c</sup>	Three hemolymphoreticular tumors in each control group; four in the male exposed group.	1
	104	0	0/100 $2/50^{c}$	0/100 2/50	Males: increased numbers of tumor-bearing animals (high concentration), testicular tumors (3 highest concentrations),	Soffritti <i>et al.</i> 2002a

and hemolymphoreticular tumors (4 highest

and hemolymphoreticular tumors (2 highest

concentrations).

less-than-lifetime exposure.]

concentrations). Females: increased incidence of mammary-

gland tumors (2 highest concentrations and at 100 mg/L)

[No water-only control group, small number of animals,

T 11 4 13 C	· · · · · · · · · · · · · · · · · · ·	••••	4 1' 66	111 1. •	· · · · · · · · · · · · · · · · · · ·
Tahla /L_I & Nummar	ry at arel end derme	l corcinacanicity	etudiae at tarma	Idahvda in av	norimontal animale
I ADIC T-IS. NUIIIIIAI	i v vi vi ai anu uci ma	i taitinustintiti i	SLUUICS VI IVI IIIA	лиспуис пі сл	Dei mientai ammais

NT = not tested.

Oslo hairless

mice

Animals

Wistar rats

Wistar rats

Wistar rats

rats

rats

Sprague-Dawley

Sprague-Dawley

Sprague-Dawley

rats (offspring)

<sup>a</sup>Given two weekly applications of 200 µL of test solution.

Route

oral

oral

oral

oral

oral

dermal

in utero

and oral<sup>b</sup>

<sup>b</sup>Offspring exposed *in utero* from gestation day 13; postnatal exposure via drinking water.

60

<sup>c</sup>Total number of stomach and intestinal tumors (benign and malignant). See Tables 4-10 and 4-12b.

10

50

100

500

1,000

1,500

 $1\%^{a}$ 

 $10\%^{a}$ 

 $2/50^{\circ}$ 

0/50

0/50

0/50

1/50

6/50<sup>c</sup>

0/16

0/16

2/50

 $3/50^{\circ}$ 

0/50

0/50

0/50

5/50<sup>c</sup>

0/16

0/16

Iversen 1986

#### 1 **4.3** Co-exposure with other substances

This section reviews studies of various designs that investigated the carcinogenic effects
in mice, rats, and hamsters following concurrent or sequential exposure to formaldehyde
and other substances. In some cases, the primary purpose was to determine whether
formaldehyde exposure enhanced or promoted the carcinogenicity of another substance.
In other cases, the primary purpose was to determine whether co-exposure to other
substances enhanced the carcinogenicity of formaldehyde.

#### 8 4.3.1 Mice

9 One of the objectives of the Horton *et al.* (1963) study (discussed in Section 4.1.1) was to 10 determine whether exposure to formaldehyde increased susceptibility to the carcinogenic 11 effects of coal tar. A group of 60 C3H mice [sex and age not reported] was exposed to formaldehyde vapor at a concentration of 100 mg/m<sup>3</sup> for 1 hour/day, 3 days/week, for 35 12 weeks and then exposed to a coal-tar aerosol at a concentration of  $300 \text{ mg/m}^3$  for 13 14 2 hours/day, 3 days/week, for up to 36 weeks. Another group of 59 mice was exposed 15 only to coal tar starting after week 35 and continuing for up to 36 weeks. A third group of 60 mice was exposed to formaldehyde at 50 mg/m<sup>3</sup> for 1 hour/day, 3 days/week, for 35 16 weeks and then exposed to formaldehyde at 150 mg/m<sup>3</sup> for 1 hour/day, 3 days/week, for 17 18 an additional 29 weeks. The control group consisted of 30 unexposed mice that were 19 killed at 82 weeks. Incidences of lung tumors in these mice are shown in Table 4-14. 20 There was no evidence that exposure to formaldehyde increased susceptibility to lung 21 tumors in mice exposed to coal-tar aerosol. No squamous-cell lung tumors were observed 22 in mice exposed to formaldehyde for up to 64 weeks.

	E	kposure (mg/m				
N	Formaldehyde wk 1–35	Coal tar Formaldehyde wk 36–71 wk 36–64		No. examined	Tumor incidence [%]	
30	0	0	0	30	0	
59	0	300	0	33	5 [15]	
60	100	300	0	26	1 [4]	
60	50	0	150	36	0	

 Table 4-14. Incidences of squamous-cell lung tumors in C3H mice exposed to formaldehyde and coal tar by inhalation

Source: Horton et al. 1963.

1 Iversen (1986) tested the potential promoting effect of formaldehyde on skin 2 carcinogenesis in hairless Oslo mice initiated with dimethylbenz(a)anthracene (DMBA). 3 Solutions were applied to the skin of the back. Two groups of 16 male and 16 female 4 mice [age not reported] were given two weekly applications of 200  $\mu$ L of an aqueous 5 solution of 1% or 10% formaldehyde for up to 60 weeks (results reported in 6 Section 4.2.2). A third group of 16 male and 16 female mice received an initial topical 7 application of 51.2  $\mu$ g of DMBA in 100  $\mu$ L of reagent-grade acetone and, beginning 8 9 days later, two weekly applications of 200  $\mu$ L of 10% formaldehyde, for up to 9 60 weeks. The positive control group of 16 male and 16 female mice received DMBA 10 followed by two weekly applications of 17 nmol 12-O-tetradecanoylphorbol 13-acetate 11 (TPA [vehicle not reported]). An additional group of 176 mice [sex not reported] 12 received a single application of 51.2  $\mu$ g of DMBA and was observed for 80 weeks. One 13 accidental death of a mouse exposed to DMBA + formaldehyde occurred at week 26. 14 Lesions observed in this group included epidermal hyperplasia in 1 mouse, lung adenomas in 3 mice, and skin tumors in 11 mice (3 squamous-cell carcinomas and 22 15 16 papillomas). The authors did not consider the lung adenoma to be exposure-related; they 17 reported an incidence of about 1 in 30 in unexposed mice from unpublished data. The 18 first skin tumors occurred at week 10 in mice given DMBA + formaldehyde. In the 19 positive-control group (DMBA + TPA), survival at 20 weeks was 80%, and the 20 experiment was terminated at week 46 with only 11 of 32 mice still alive. Tumors first 21 appeared in the DMBA + TPA group after 5 weeks, and all mice that survived until week 22 20 had skin papillomas; however, no carcinoma or sarcoma was observed. Most of the 23 mice in the DMBA-only group survived until the end of the experiment, and 225 skin 24 tumors (primarily papilloma) occurred in 85 mice; the first tumors in this group appeared 25 after 20 weeks.

The authors reported there was no difference in tumor yields between groups given DMBA + formaldehyde and mice given DMBA only. The final tumor yield (the total number of tumors as a function of time) was evaluated according to the method of Gail *et al.* (1980). The final tumor rate (the percentage of tumor-bearing mice in relation to the number of mice alive at the appearance of the first tumor) was not significantly higher in

- 1 mice given DMBA + formaldehyde than in mice given DMBA only; however, the time to
- 2 appearance of the first tumor and the mean latency period were significantly reduced (P =
- 3 0.01, Peto's test). Tumor incidence and the total number of reported tumors are shown in
- 4 Table 4-15. The authors concluded that 10% formaldehyde applied twice a week to the
- 5 skin of Oslo hairless mice following one application of DMBA did not increase the total
- 6 number of tumors but significantly reduced the mean latency period for tumor formation.
- 7 This effect was much weaker than that observed with TPA.

Table 4-15. Skin tumor promotion study of formaldehyde in Oslo hairless mice

			Time to		Total number of tumors			
Group	Study length (wk)	N	first tumor (wk)	incidence [%] <sup>ª</sup>	Papilloma	Carcinoma	Total	
DMBA	80	176	[22] <sup>b</sup>	85 [48]	219	6	225	
DMBA + HCHO	60	32	10	11 [34]	22	3	25	
DMBA + TPA	46	32	[8] <sup>b</sup>	26 [100] <sup>c</sup>	NR	0	NR	

Source: Iversen 1986.

DMBA = dimethylbenz(a)anthracene, HCHO = formaldehyde, TPA = 12-O-tetradecanoylphorbol 13-acetate, NR = not reported.

<sup>a</sup>Tumor incidences cannot be compared directly because of the differing study lengths and because they are not adjusted for survival differences.

<sup>b</sup>Estimated from a figure.

<sup>c</sup>Six mice died before week 20 and were not included in the analysis.

#### 8 4.3.2 Rats

- 9 Albert et al. (1982) and Sellakumar et al. (1985) investigated the carcinogenicity of a
- 10 mixture of formaldehyde and hydrogen chloride (HCl) in rats. Previous studies had
- 11 shown that low levels of bis(chloromethyl)ether (BCME), which is highly carcinogenic in
- 12 the respiratory tract of rats and is a known human carcinogen, could form from the gas-
- 13 phase reaction of formaldehyde and hydrogen chloride. In the first study (Albert *et al.*
- 14 1982), 8-week-old male Sprague-Dawley rats were divided into three groups of 50
- 15 unexposed colony controls, 50 controls sham-exposed to air, and 99 rats exposed to a
- 16 mixture of approximately 14 ppm formaldehyde and 10 ppm HCl (the gases were
- 17 premixed at high concentrations before introduction into the inhalation chamber, to
- 18 maximize formation of BCME). Exposures were for 6 hours/day, 5 days/week, for life. A
- 19 complete necropsy was performed on each animal. Formation of BCME was monitored
- 20 by gas chromatography. BCME levels in the mixing vessel ranged from 8 to 179 ppb

1 (mean = 75 ppb); however, BCME concentrations in the exposure chamber were less than 2 the detection limit [not identified by study authors] and were estimated to be no greater 3 than 1 ppb, based on a 75-fold dilution factor. The exposed group had substantially lower 4 body-weight gain and higher mortality than the controls. Early deaths in the exposed 5 group and controls were attributed to bronchopneumonia. The exposed group showed 6 high incidences of squamous metaplasia of the nasal cavity and epithelial hyperplasia 7 with and without atypia. Nasal tumors (3 squamous-cell papillomas and 25 squamous-cell 8 carcinomas) were observed in the exposed group but not in the controls (Table 4-16). 9 Incidences of non-respiratory-tract tumors were higher in the control groups (23 of 100) 10 than in the exposed rats (7 of 99). These tumors included lymphoma, pituitary gland and 11 adrenal cortical adenoma, subcutaneous fibrosarcoma, and 1 splenic hemangioma. No 12 statistical analyses were reported by the study authors. However, the IARC (2006) 13 evaluation of this study reported that the incidence of squamous-cell carcinoma was 14 significantly higher in the exposed group than in the controls (P < 0.001, Fisher's exact 15 test).

16 Sellakumar *et al.* (1985) conducted a follow-up of the Albert *et al.* (1982) study to

17 examine the carcinogenic effects of formaldehyde and HCl when administered alone or in

18 combination. Groups of 99 or 100 male Sprague-Dawley rats, 9 weeks of age, were

19 randomly assigned to six treatment groups: (1) colony controls, (2) controls sham-

20 exposed to air, (3) exposed to formaldehyde at a target concentration of 15 ppm and HCl

21 at a target concentration of 10 ppm, premixed before being introduced into the inhalation

chamber, (4) exposed to formaldehyde (15 ppm) and HCl (10 ppm) introduced separately

23 into the exposure chamber, (5) exposed to formaldehyde alone (15 ppm), and (6) exposed

to HCl alone (10 ppm). Rats were exposed for 6 hours/day, 5 days/week, for life.

25 Formation of BCME by the premixed formaldehyde and HCl was again monitored by gas

chromatography. BCME concentrations in the mixing vessel ranged from 3.6 to 33.7 ppb,

and the calculated concentrations in the inhalation chamber ranged from 0.1 to 0.4 ppb.

- 28 Complete necropsies were performed, with particular attention to the respiratory tract.
- 29 Histologic sections were prepared from the lungs, trachea, larynx, liver, kidneys, testes,
- 30 and any other organs with gross pathology. After 16 weeks, groups exposed to
- 31 formaldehyde alone or formaldehyde plus HCl had lower body weights than the controls.

1 Mortality rates among all the groups were similar up to 32 weeks. After 32 weeks, the 2 group exposed to premixed formaldehyde plus HCl showed a higher mortality rate than 3 the other groups. Nasal tumors occurred only in groups exposed to formaldehyde alone or 4 in combination with HCl (Table 4-16). No tumors developed in the trachea or lungs. The 5 total number of non-respiratory-tract tumors did not differ between the exposed and 6 control groups. The authors reported that the incidence of nasal tumors was significantly 7 higher in the group exposed to premixed formaldehyde plus HCl than in the formaldehyde-only group (P < 0.025,  $\chi^2$  test). IARC's (2006) review of this study also 8 9 reported that the incidence of squamous-cell carcinoma and papilloma combined was 10 significantly higher in the formal dehyde-only group than in the controls (P < 0.001, 11 Fisher's exact test). [In statistical analysis conducted by NTP, the incidences of 12 squamous-cell carcinoma in the groups exposed to formaldehyde only, premixed 13 formaldehyde plus HCl, and non-premixed formaldehyde plus HCl were significantly 14 higher than in the controls (P < 0.001, Fisher's exact test).] The authors noted that the 15 higher incidences in the group exposed to premixed formaldehyde plus HCl could have 16 been due to traces of alkylating agents other than BCME formed by the interaction of 17 formaldehyde and HCl. Nevertheless, the authors concluded that HCl had little to no 18 effect on the carcinogenicity of formaldehyde and that formaldehyde accounted for most, 19 if not all, of the carcinogenic activity of the mixture.

	Nasal-cavity lesion [%]								
Group	N	Epithelial hyperplasia	Squamous metaplasia	Squamous- cell papilloma or polyps	Squamous- cell carcinoma	Other <sup>a</sup>			
Study 1									
Colony controls	50	8 [16]	0	0	0	NR			
Sham air	50	NR	NR	NR	NR	NR			
HCl + HCHO	99	71 [72]	64 [65]	3 [3]	25 [25***]	NR			
Study 2									
Colony controls	99	45 [45]	6 [6]	0	0	0			
Sham air	99	51 [52]	5 [5]	0	0	0			
HC1	99	62 [63]	9 [9]	0	0	0			
HCHO <sup>b</sup>	100	57 [57]	60 [60]	10 [10]	38 [38***]	2 [2]			
Premixed HCl + HCHO <sup>c</sup>	100	54 [54]	64 [64]	13 [13]	45 [45***]	3 [3]			
Non-premixed HCl + HCHO	100	53 [53]	68 [68]	11 [11]	27 [27***]	2 [2]			

 

 Table 4-16. Proliferative and neoplastic lesions in the nasal cavity of male Sprague-Dawley rats exposed to formaldehyde and hydrogen chloride

Source: Albert et al. 1982, Sellakumar et al. 1985, IARC 2006.

HCl = hydrogen chloride, HCHO = formaldehyde, NR = not reported.

\*\*\*P < 0.001 (compared with controls, Fisher's exact test conducted by IARC 2006 or NTP).

<sup>a</sup>Includes adenocarcinoma, mixed carcinoma, fibrosarcoma, or esthesioneuroepithelioma of the nasal mucosa.

<sup>b</sup>IARC reported that the incidence of squamous-cell carcinoma and papilloma combined was significantly higher in this group than in the controls (P < 0.001, Fisher's exact test).

<sup>c</sup> The study authors reported a significantly higher incidence of nasal cancer in this group than in the formaldehyde-only group (P < 0.025,  $\chi^2$  test).

- 1 Homma et al. (1986) investigated whether repeated intravesical instillation of formalin
- 2 would promote urinary-bladder carcinogenesis in male F344 rats. Heterotopically
- 3 transplanted bladders were used, because transient generalized hyperplasia can be readily
- 4 and repeatedly induced by intravesical instillation of formalin without the risk of
- 5 infection or calculus formation, which are unavoidable when homotopic bladders are
- 6 used. The rats were randomly divided into four groups of 35 animals each. Four weeks
- 7 after bladder transplant, three groups received 0.25 mg of *N*-methyl-*N*-nitrosourea
- 8 (MNU) in 0.9% saline to initiate bladder carcinogenesis. At week 5, group 1 was given
- 9 an intravesical instillation of 0.5 mL of 0.3% formalin, followed by instillation of 0.5 mL
- 10 of normal rat urine 24 hours later and 0.5 mL of 2.1% sodium chloride (NaCl) solution
- 11 1 week after the urine instillation. Group 2 was treated similarly to group 1 except that
- 12 the order of the urine and salt solution instillation was reversed. Group 3 received 0.9%
- 13 NaCl solution at week 5 instead of formalin, then 2.1% NaCl 24 hours later and rat urine

1 week later. Group 4 was treated the same as group 1 but without MNU initiation. The
 alternating instillation schedule was repeated every 2 weeks for 15 cycles in each group,
 and the experiment was terminated at week 34. The heterotopically transplanted bladders
 were inflated with Bouin's solution, fixed overnight, and examined for gross tumors. In
 addition, longitudinal strips were examined microscopically. Repeated formalin exposure
 did not enhance bladder carcinogenesis.

7 Takahashi et al. (1986) tested formaldehyde and other compounds for tumor-promoting 8 activity in a two-stage stomach carcinogenicity study. Stomach tumors were initiated by 9 giving two groups of 7-week-old male Wistar rats N-methyl-N'-nitro-N-nitrosoguanidine 10 (MNNG) in drinking water at a concentration of 100 mg/L and a diet supplemented with 11 10% sodium chloride for 8 weeks. Thereafter, one group of 30 rats received no further 12 treatment (*i.e.*, no exposure to a promoter), and one group of 20 rats received 0.5% 13 formalin in drinking water from week 8 to 40. Two additional groups of 10 rats received 14 no MNNG; one of these groups was exposed only to formaldehyde from week 8 to 40, 15 and a control group received no treatment. All animals that survived beyond week 30 16 were included in the analysis; 3 rats in the MNNG plus formaldehyde group died early 17 and were not included in the analysis. For the first 8 weeks, the two groups that received 18 MNNG showed lower body-weight gain than the groups that did not receive MNNG; 19 however, their weight gain increased after week 8. Throughout the study, growth 20 retardation was most marked in the group that received MNNG plus formaldehyde. 21 Formaldehyde showed possible tumor-promoting effects in the pylorus of the glandular 22 stomach, and the incidence of squamous-cell papilloma of the forestomach was 23 significantly increased in groups exposed to formaldehyde with or without initiation. In 24 addition, the incidence of adenomatous hyperplasia of the fundus was significantly higher 25 in the MNNG plus formaldehyde group than in the MNNG-only group (88.2% vs. 0). 26 Results are summarized in Table 4-17.

		Forestomach	Glandular stomach adenocarcinomas (%)					
Group	Ν	papilloma (%)	Fundus	Pylorus	Duodenum			
Control	10	0	0	0	0			
MNNG only	30	0	0	1 (3.3)	3 (10)			
MNNG + HCHO	17	15 (88.2)**	0	4 (23.5)* <sup>,a</sup>	1 (5.9)			
HCHO only	10	8 (80)**	0	0	0			

 Table 4-17. Effects of formaldehyde on gastric carcinogenesis in male Wistar rats

 initiated with MNNG

Source: Takahashi et al. 1986.

HCHO = formaldehyde, MNNG = *N*-methyl-*N'*-nitro-*N*-nitrosoguanidine. \*P < 0.05; \*\* P < 0.01 (compared with MNNG group, Fisher's exact test).

<sup>a</sup>[P = 0.051, Fisher's exact test conducted by NTP].

1 Holmstrom et al. (1989a) investigated the cocarcinogenic effects of formaldehyde 2 (average concentration of 12.4 to 12.7 ppm) and wood dust. Concurrent exposure to 3 formaldehyde and wood dust is common, particularly in the furniture industry. Groups of 4 16 female Sprague-Dawley rats, 11 weeks of age, were exposed to formaldehyde alone (results reported in Section 4.1.2), wood dust alone  $(25 \text{ mg/m}^3)$ , or formaldehyde plus 5 6 wood dust for 104 weeks. No nasal or lung tumors occurred in the wood-dust or 7 formaldehyde plus wood-dust exposure groups. One squamous-cell carcinoma of the 8 nasal mucosa occurred in the group exposed to formaldehyde only. Squamous-cell 9 metaplasia with dysplasia was most common in the group exposed to both formaldehyde 10 and wood dust. Pulmonary emphysema was most common in the group exposed only to 11 wood dust. The authors considered that the most important finding of this study was the 12 additive deleterious effect of combined exposure to formaldehyde and wood dust. The 13 IARC (2006) evaluation of this paper noted that a small number of animals was used in 14 this study.

IARC (2006) also reviewed a study published in Russian (Yanysheva *et al.* 1998) that
investigated the promoting effects formaldehyde administered by inhalation at low
concentrations. Groups of 50 white non-inbred female rats [age and strain not reported],
including a control group, were exposed to formaldehyde at a concentration of 0.003,
0.03, or 0.3 mg/m<sup>3</sup> [0.002, 0.02, and 0.24 ppm] either alone or in combination with and
benzo[*a*]pyrene. Benzo[*a*]pyrene was administered by intratracheal injection once every
weeks for 20 weeks (for a total dose of 0.02, 1, or 5 mg). Formaldehyde was

1 administered by inhalation for 7 hours/day, 5 days/week, for 1 year. Animals were held 2 until natural death. Tumors were observed in all groups. Two rats in the control group 3 developed reticulosarcoma of the lung, and two others developed fibroadenoma of the 4 mammary gland. Similar incidences of these tumors were observed in the three 5 formaldehyde-only exposure groups. In rats given only benzo[a] pyrene, the total 6 incidence of tumors ranged from 13% to 28%, and incidence of lung tumors ranged from 7 9% to 19%. A dose-dependent tumor response was observed in groups exposed to both 8 benzo[a]pyrene and formaldehyde. The most significant effect was an increase in lung 9 tumors (43%) and total tumors (69%) in the group exposed to the highest levels of 10 benzo[a]pyrene and formaldehyde. Tumors also developed earlier in this group and had 11 greater multiplicity than in the other groups. The authors concluded that combined 12 exposure to benzo[*a*]pyrene and formaldehyde enhanced the tumor response in rats.

#### 13 4.3.3 Hamsters

14 Although inhalation exposure to formaldehyde alone did not induce respiratory-tract 15 tumors in male Syrian golden hamsters (see Section 4.1.3), there was evidence that it 16 could be a cofactor in the induction of respiratory-tract tumors by DEN (Dalbey 1982). A 17 group of 50 male hamsters [age not reported] was exposed to formaldehyde at a 18 concentration of 30 ppm for 5 hours/day, 1 day/week, for life (also reported in Section 19 4.1.3). Two additional groups of hamsters were exposed to formaldehyde at 30 ppm; one 20 of these groups also received weekly injections of 0.5 mg of DEN 48 hours after the 21 weekly formaldehyde exposure for the first 10 weeks, and the other group received 10 22 weekly DEN injections before beginning formaldehyde exposure. An unexposed control 23 group consisted of 50 hamsters, and a DEN-only control group consisted of 100 24 hamsters. The lungs, trachea, larynx, nasal turbinates, and lower jaw were examined for tumors. Tumor incidence data were analyzed with a  $\chi^2$  test [the statistical method used to 25 26 analyze tumor multiplicity was not identified]. Mortality was not affected by exposure to 27 formaldehyde but was significantly increased in the DEN-only group and both DEN plus 28 formaldehyde groups. Because of mortality due to an exposure accident at 48 weeks, the 29 sizes of the DEN plus formaldehyde groups were reduced to 27 and 23. No tumors 30 occurred in the unexposed controls or in the formaldehyde-only group. The tumor 31 incidence (primarily tracheal tumors) was 77% in the DEN-exposed group and was not

- 1 significantly higher than this in the DEN plus formaldehyde groups (the incidences were
- 2 not reported). However, tumor multiplicity (tumors per tumor-bearing animal) was
- 3 significantly higher in the group that received DEN plus formaldehyde simultaneously
- 4 than in the DEN-only group (Table 4-18). All tumors were adenomas. Nasal tumor
- 5 incidence was only 2% in the DEN-only group and the group exposed to DEN plus
- 6 formaldehyde sequentially, but no nasal tumors occurred in the other three groups.

Table 4-18. Effects of formaldehyde on induction of respiratory-tract tumors byDEN in male Syrian hamsters

		Tumor	[Tumors/t	g animal] <sup>a</sup>	
Group	N (%)		Larynx	Trachea	Lung
Unexposed control	50	0	0	0	0
HCHO only	50	0	0	0	0
DEN only	100	77	1	1.6	1.4
HCHO + DEN, then HCHO	27	NR	1	2.8*	1.0
DEN, then HCHO	23	NR	1	1.7	2.0

Source: Dalbey 1982.

DEN = diethylnitrosamine, HCHO = formaldehyde, NR = not reported; however, the authors stated that the incidence was not significantly different from that of the DEN-exposed group.

\*P < 0.05 (compared with the DEN-only group, statistical test not identified).

<sup>a</sup>Values were estimated from Figure 3 in Dalbey 1982.

#### 7 4.3.4 Summary of promotion and cocarcinogenicity studies

8 Several studies investigated the promoting or cocarcinogenic effects of formaldehyde.

9 Formaldehyde did not enhance lung carcinogenesis in mice exposed to coal tar but did

10 reduce the latency period for skin tumors in mice initiated with DMBA. Studies in rats

- 11 indicated that formaldehyde exhibited possible tumor-promoting effects in stomach and
- 12 lung but not in the urinary bladder. In another study, hydrogen chloride had little or no
- 13 effect on the carcinogenicity of formaldehyde. One study in hamsters indicated possible
- 14 tumor-promoting effects in the respiratory tract. Results from all co-exposure studies of
- 15 formaldehyde and other substances are summarized in Table 4-19.

-		ë •	•	-		
Species and strain		Exposure				
(sex) <sup>a</sup>	Route	Exposure (concentration)	Duration (wk)	Results	Reference	
C3H mice	inhalation	HCHO $(100 \text{ mg/m}^3) + \text{coal tar}$ (300 mg/m <sup>3</sup> )	35 + 33	Did not enhance induction of lung tumors	Horton et al. 1963	
Oslo mice	skin	DMBA (51.2 µg) + HCHO (10%)	$1^{b} + 60$	Tumor latency was decreased; no effect on tumor incidence	Iversen 1986	
Sprague-Dawley rats (male)	inhalation	HCHO (14 ppm) + HCl (10 ppm)	life <sup>c</sup>	Increased nasal tumor incidence, compared with colony controls	Albert <i>et al.</i> 1982	
Sprague-Dawley rats (male)	inhalation	HCHO (15 ppm) + HCl (10 ppm)	life <sup>c</sup>	HCl had little effect on induction of nasal tumors by formaldehyde	Sellakumar et al. 1985	
F344 rats (male)	intravesical	MNU (0.25 mg) + HCHO (3,000 ppm)	1 <sup>b</sup> + 34	Did not promote urinary bladder carcinogenesis	Homma <i>et al</i> . 1986	
Wistar rats (male)	drinking water	MNNG (100 ppm) + HCHO (5,000 ppm)	8 + 32	Possible weak promotion effect for adenocarcinoma in the glandular stomach	Takahashi <i>et al.</i> 1986	
Sprague-Dawley rats (female)	inhalation	HCHO (12.7 ppm) + wood dust (25 mg/m <sup>3</sup> )	104	One squamous-cell carcinoma in formaldehyde-only group; squamous-cell metaplasia with dysplasia increased in combined exposure group	Holmström <i>et al.</i> 1989a	
White non-inbred rats (female)	inhalation	HCHO $(0.3 \text{mg/m}^3) + \text{B}[a]\text{P} (5 \text{ mg})$	52 <sup>d</sup>	Combined exposure enhanced induction of lung and total tumors	Yanysheva <i>et al.</i> 1998 (cited in IARC 2006)	
Syrian golden hamsters (female)	inhalation	DEN $(0.5 \text{ mg})$ + HCHO $(30 \text{ ppm})$	$10 + life^{e}$	Tumor multiplicity was increased	Dalbey 1982	

Table 4-19. Co-exposure carcinogenicity studies of formaldehyde and other substances in experimental animals

BaP = benzo[a]pyrene, DEN = diethylnitrosamine, DMBA = dimethylbenz(a)anthracene, HCHO = formaldehyde, HCl = hydrogen chloride, MNNG = N-methyl-N'-nitrosoguanidine, MNU = N-methyl-N-nitrosoguanidine.

<sup>a</sup>When only one sex was used.

<sup>b</sup>Single application of the initiator.

<sup>c</sup>Exposed to a mixture of formaldehyde and hydrogen chloride.

<sup>d</sup>Exposed for one year and observed until death.

<sup>e</sup>DEN given in 10 weekly injections either before or concurrently with formaldehyde exposure.

#### 1 4.4 Summary

2 Formaldehyde has been tested for carcinogenicity in mice, rats, and hamsters

3 (Table 4-20). Studies reviewed include chronic and subchronic inhalation studies in mice,

4 rats, and hamsters; chronic and subchronic drinking-water studies in rats; and one chronic

5 skin-application study in mice. No chronic studies in primates were found, but one

6 subchronic inhalation study and one acute/subacute inhalation study in monkeys were

7 reviewed.

8 Formaldehyde exposure resulted in nasal tumors (primarily squamous-cell carcinoma) in

9 rats when administered chronically by inhalation (Kerns *et al.* 1983, Appelman *et al.* 

10 1988, Woutersen et al. 1989, Sellakumar et al. 1985, Monticello et al. 1996, Kamala et

11 al. 1997). Only two inhalation studies in mice or hamsters were found. No tumors were

12 reported in C3H mice exposed to formaldehyde at 200 mg/m<sup>3</sup> for 1 hour/day, 3

13 days/week, for 35 weeks (Horton et al. 1963), but squamous-cell carcinoma of the nasal

14 cavity occurred in 2 of 120  $B6C3F_1$  male mice exposed at 14 ppm for 6 hours/day, 5

15 days/week, for 104 weeks (Kerns et al. 1983). The authors concluded that the tumors

16 were exposure-related, although the increase was not statistically significant. No tumors

17 were reported in Syrian golden hamsters exposed at 10 ppm for life (Dalbey 1982) or

18 2.95 ppm for 26 weeks (Rusch *et al.* 1983). No tumors occurred in monkeys exposed at

19 2.95 ppm for 26 weeks (Rusch *et al.* 1983) or 6 ppm for 6 weeks (Monticello *et al.* 1989);

20 however, squamous metaplasia and hyperplasia in the nasal passages and respiratory

21 epithelia of the trachea and major bronchi occurred.

22 Male rats administered formaldehyde in drinking water at 5,000 ppm for 32 weeks 23 developed forestomach tumors (squamous-cell papillomas) in one study (Takahashi et al. 24 1986); however, in two other drinking-water studies, no tumors were reported in either 25 male or female rats administered formaldehyde at concentrations ranging from 20 to 26 5,000 ppm for two years (Til et al. 1989, Tobe et al. 1989). In another study, male and 27 female breeder rats administered formaldehyde at 2,500 ppm in drinking water had 28 slightly increased incidences of hemolymphoreticular neoplasms (Soffritti et al. 1989). 29 Offspring of these breeder rats exposed transplacentally beginning on gestation day 13 30 and postnatally via drinking water for life showed increased incidences of benign and

1 malignant tumors of the gastrotinestinal tract, particularly intestinal leiomyosarcoma. 2 Male rats administered formaldehyde at concentrations up to 1,500 ppm showed 3 increased incidences (compared with control groups given tap water or tap water 4 containing 15 mg/L methanol) of the number of animals bearing malignant tumors, 5 hemolymphoreticular neoplasms (leukemia and lymphoma combined), and testicular 6 tumors (interstitial-cell adenoma) (Soffritti et al. 2002a). Female rats showed higher 7 incidences of mammary-gland adenocarcinoma and hemolymphoreticular neoplasms than 8 the tap-water control group; however, the incidences were not significantly higher than in 9 the tap-water-plus-methanol control group. In addition, some rare stomach and intestinal 10 tumors occurred in a few male and female rats in the exposed groups but not in the 11 control groups. 12 Other studies examined the promoting effects of formaldehyde when administered after 13 initiation with DBMA, DEN, MNU, or MNNG or cocarcinogenic effects when 14 administered with coal tar, benzo[a]pyrene, wood dust, and hydrogen chloride. Some of 15 these studies did not show an enhanced tumor response. However, a few studies, 16 including a skin-painting study in mice (Iverson et al. 1986), a drinking-water study in 17 rats (Takahashi et al. 1986), and inhalation studies in rats (Albert et al. 1982, Holmstorm

18 et al. 1989a) and hamsters (Dalbey et al. 1986), indicated that formaldehyde could act as

19 a tumor promoter or act as a cocarcinogen when administered with other substances.

		B6C3F <sub>1</sub> Mouse F344 F		44 Rat	Wis	tar Rat	Sprague- Dawley Rat	
Organ or system	Tumor type	Male	Male	Female	Male	Female	Male	Female
Inhalation studies								
Nasal epithelium	squamous-cell carcinoma	×	+	+	×		+	×
	papilloma or polyps						+	
	polypoid adenoma		$+^{t}$	×	×			
	carcinoma in situ				×			
	rhabdomyosarcoma		×					
	adenocarcinoma		×					
	combined tumor types				+ <sup>a</sup>			
Ingestion studies								
Gastrointestinal	forestomach papilloma				+			
	adenoma, papilloma, acanthoma						×	×
	adenocarcinoma						×	×
	leiomyosarcoma						× <sup>b,c</sup>	+°
	leiomyoma							×
Hemolymphoreticular	leukemia and lymphoma						+	$+^{d}$
Mammary-gland	total malignant (primarily adenocarcinoma)							+ <sup>d</sup>
Testicular	interstitial-cell adenoma						+	

# Table 4-20. Summary of neoplasms associated with formaldehyde exposure in experimental animals

+ = Statistically significant increase in tumor incidence reported.

+<sup>t=</sup> Statistically significant dose-related trend.

 $\times$  = Statistical results were not reported or were not significant, but study authors reported the effect to be exposure-related.

<sup>a</sup>Incidence of formaldehyde-related tumors (squamous-cell carcinoma, carcinoma *in situ*, and polypoid adenoma) (incidence = 4.5%; 6 tumors/132 rats) reported as significant (P = 0.01, Fisher's exact test) by IARC 2006.

<sup>b</sup>Significant when combined with female rats.

<sup>c</sup>Transplacental exposure beginning on gestation day 13 and postnatal exposure via drinking water for life. <sup>d</sup>Not significant when compared with the control group given methanol at 15 mg/L in tap water.

# 1 5 Other Relevant Data

Other data that are relevant for evaluating the carcinogenicity of formaldehyde are
reviewed in this section. This includes absorption, distribution, metabolism and excretion,
general toxic effects, carcinogenicity data for metabolites and analogues, genetic and
related effects, and potential mechanisms of action.

# 6 5.1 Absorption, distribution, and excretion

As discussed in Section 2, formaldehyde exposure occurs from both endogenous and exogenous sources. Formaldehyde is an essential metabolic intermediate used in the biosynthesis of purines, thymidine, and some amino acids. Metabolically it is produced from serine, glycine, methionine, and choline, and from the demethylation of *N*-, *O*-, and *S*-methyl compounds (IARC 2006). The endogenous concentrations of formaldehyde in human blood are about 2 to 3  $\mu$ g/g of blood and are similar to concentrations measured in the blood of monkeys and rats (Casanova *et al.* 1988, Heck *et al.* 1985).

14 Formaldehyde is rapidly hydrated when dissolved in water and forms methylene glycol

15 (Fox et al. 1985). The equilibrium lies far in favor of methylene glycol. In tissues,

16 formaldehyde in solution reacts readily with macromolecules (e.g., proteins,

- 17 glycoproteins, nucleic acids, and polysaccharides) resulting in more formaldehyde
- 18 forming from dissociation of methylene glycol. The equilibrium between formaldehyde
- 19 and methylene glycol helps explain why formaldehyde penetrates rapidly (as methylene

20 glycol) and fixes slowly (as carbonyl formaldehyde).

21 The metabolic pathways for formaldehyde are the same in all tissues of the body.

- 22 Formaldehyde is rapidly metabolized to formic acid (formate  $+ H^+$ ) (see Section 5.3) at
- 23 the site of contact and by erythrocytes in the blood, or is incorporated into serum proteins
- 24 and other macromolecules via the one-carbon metabolic pool. The reported half-life of
- 25 formaldehyde in the plasma of rats and monkeys is about 1 to 1.5 minutes (IARC 2006,
- 26 McMartin et al. 1979). Burkhart (1990) reported an apparent plasma half-life of formate

and formaldehyde of 3.1 hours and 3.3 hours, respectively, in a 58-year-old man that
 committed suicide by ingesting 4 ounces of formaldehyde.

3 5.1.1 In vitro studies

4 Loden et al. (1986b) investigated the skin permeability of formaldehyde and other chemicals using excised human skin in a flow-through diffusion cell. <sup>14</sup>C-Formaldehyde 5 was diluted in either concentrated formalin (37% formaldehyde in water containing 10% 6 7 to 15% methanol) or a 10% v/v solution of formalin in 0.1 M phosphate buffer and 8 applied to full thickness skin mounted in Teflon® flow-through diffusion cells. 9 Phosphate-buffered saline (pH 7.4) was used as the receptor medium. The rates of resorption (*i.e.*, the uptake by the receptor fluid beneath the skin) of formaldehyde in 10 concentrated formalin and 10% formalin were 319  $\mu$ g/cm<sup>2</sup> per hour and 16.7  $\mu$ g/cm<sup>2</sup> per 11 hour, respectively. The total amount absorbed (i.e., the amount in the skin and the 12 receptor medium) at steady state was 6.02 mg/cm<sup>2</sup> (concentrated formalin) and 0.48 13  $mg/cm^2$  (10% formalin). The effect of methanol on the uptake of formaldehyde was not 14 determined. Up to approximately half the radioactivity absorbed was retained in the skin. 15

#### 16 5.1.2 In vivo studies

17 Formaldehyde is rapidly and almost completely absorbed from the respiratory and 18 gastrointestinal tracts but is poorly absorbed from the skin (ATSDR 1999, IARC 1995, 19 2006). In addition, Myers *et al.* (1997) reported rapid absorption of formalin following 20 rectal instillation in dogs. In rats, almost all inhaled formaldehyde is absorbed in the nasal 21 passages, while in primates, although almost all is absorbed in the nasal passages, some 22 absorption occurs in the trachea and proximal regions of the major bronchi (Casanova et 23 al. 1991, Chang et al. 1983, Heck Hd et al. 1983, Monticello et al. 1989). Nasal anatomy, 24 which is highly variable among species, and breathing patterns are the primary factors 25 associated with the efficiency and specific location of formaldehyde absorption.

- 26 5.1.2.1 Inhalation exposure
- 27 Formaldehyde concentrations and air flow patterns in the nasal passages of rodents,
- 28 monkeys, and humans have been correlated with the location of nasal lesions and levels
- 29 of DNA-protein crosslinks (IARC 2006). One important physiological difference is that
- 30 rats are obligate nose breathers while monkeys and humans are oronasal breathers. Thus,

1 during oronasal breathing, a significant amount of the inhaled formaldehyde would 2 bypass the nose and deposit directly into the lower respiratory tract of humans. Overton 3 et al. (2001) conducted dosimetry modeling of inhaled formaldehyde in the respiratory 4 tract of humans at four activity levels. The respiratory tract was divided into segments or 5 generations beginning at nose and mouth and ending at the alveolar sacs. These authors 6 predicted that for each activity state, the respiratory tract would retain over 95% of 7 inhaled formaldehyde and that the rate of mass flow across a unit area of the respiratory 8 tract (i.e., flux) in the first few tracheobronchial model generations would be more than 9 1,000 times higher than in the first pulmonary region, with no flux to the alveolar region. 10 Egle (1972) reported similar findings in dogs exposed to formaldehyde at concentrations 11 of 0.15 to 0.35 µg/mL [122 to 285 ppm]. Uptake of formaldehyde by the upper 12 respiratory tract was near 100% regardless of the concentration.

13 Heck *et al.* (1982) exposed male F344 rats to 6 ppm formaldehyde for 6 hours/day for 10

14 days. The rats were killed within 10 minutes of exposure termination. Formaldehyde

15 concentrations in the nasal mucosa of exposed rats ( $0.39 \pm 0.12 \ \mu mol/g$ ) were not

16 significantly different from controls  $(0.42 \pm 0.09 \ \mu mol/g)$ .

17 Heck et al. (1983) conducted several experiments in groups of four male F344 rats to investigate the distribution, elimination, and pharmacokinetics of <sup>14</sup>C-formaldehyde 18 19 following inhalation exposure (head only). [There were no unexposed control groups in 20 this study.] Total radioactivity in the nasal mucosa, trachea, and plasma were measured immediately after a 6-hour exposure to 5, 10, 15, or 24 ppm <sup>14</sup>C-formaldehyde. 21 22 Concentrations were highest in the nasal mucosa and ranged from about 0.5 to 2.3 µmole 23 equivalents/g tissue and were related to dose. Concentrations in the trachea (about 0.3 24  $\mu$ mole equivalents/g) and plasma (about 0.1  $\mu$ mole equivalents/g) were not affected by 25 dose, which indicates that absorption occurs primarily in the upper respiratory tract. The ratio of levels of <sup>14</sup>C (total radioactivity) in internal organs to that in plasma ranged from 26 27 0.31 in the testes to 4.94 in the esophagus and was not affected by dose. The higher 28 concentrations in the esophagus were thought to reflect mucociliary clearance from the 29 upper respiratory tract. Values for other organs declined in the order of kidney, liver, 30 intestine, lung, spleen, heart, and brain. Another experiment examined the effects of pre-
exposure to formaldehyde on tissue concentrations. One group was pre-exposed to 15
 ppm formaldehyde 6 hours/day for 9 day while the other group was not pre-exposed to

- 3 formaldehyde (naïve animals). On the tenth day, both groups were exposed (head-only)
- 4 to <sup>14</sup>C-formaldehyde at 14.9 ppm for 6 hours. There were no differences in tissue
- 5 concentrations between these groups, thus, pre-exposure to formaldehyde did not

6 influence either the absorption or distribution to plasma.

7 Other groups of male F344 rats were exposed to 0.63 or 13.1 ppm <sup>14</sup>C-formaldehyde for

- 8 6 hours (Heck *et al.* 1983). Following exposure, the rats were placed in metabolism cages
- 9 for 70 hours and then sacrificed. Radioactivity in urine, feces, expired air, and the carcass

10 was measured. The dose did not affect the proportion recovered from the various

11 elimination pathways (Table 5-1). Exhalation accounted for about 40% of the total dose.

12 The authors noted that exhalation of  ${}^{14}CO_2$  was biphasic, with a rapid decline over the

13 first 12 hours followed by a more gradual decline. About 17.5% was eliminated in the

14 urine and 4% to 5% was eliminated in the feces. The amount of radioactivity remaining

- 15 in the carcass was 38.9% (low dose) and 35.2% (high dose). The authors noted that since
- 16 formaldehyde is a precursor for many biological compounds, the high levels of
- 17 radioactivity remaining in the carcass were probably due to metabolic incorporation.

	Exposure concentration (ppm)			
Source of Radioactivity	0.63	13.1		
Expired air	39.4 ± 1.5	$41.9\pm0.8$		
Urine	$17.6 \pm 1.2$	$17.3 \pm 0.6$		
Feces	$4.2 \pm 1.5$	$5.3 \pm 1.3$		
Tissues and carcass	$38.9\pm1.2$	$35.2\pm0.5$		

Table 5-1. Disposition of inhaled  $^{14}\mathrm{C}$  -formaldehyde in male F344 rats (% radioactivity  $\pm$  SD)

Source: Heck et al. 1983.

- 18 Heck et al. (1983) also investigated the uptake and disappearance of radioactivity from
- 19 the blood of male F344 rats following exposure to formaldehyde by inhalation (6 hours,
- 20 head only) or a single i.v. injection of formaldehyde or formate. Blood samples were
- 21 collected during and after exposure through a cannula implanted in the jugular vein. The
- 22 concentrations of radioactivity in plasma increased during the exposure period, peaked at
- 23 approximately the time of removal from the exposure chamber, and then gradually

1 declined over a period of several days. The terminal half-life of radioactivity in plasma 2 was approximately 55 hours; however, the authors stated that the radioactivity most 3 likely indicated incorporation into serum proteins because the half-life of these proteins is 4 about 2.9 days in the rat and the half-life for free formaldehyde in rat plasma is 5 approximately 1 minute (Rietbrock 1965, as cited in IARC 2006). Radioactivity in the 6 packed cell fraction of the blood showed a multiphasic profile that increased during 7 exposure but rapidly declined within the first post-exposure hour. This was followed by 8 an increase that peaked at about 35 hours post-exposure. The terminal phase showed a 9 slow decline that was consistent with incorporation into the erythrocytes. The kinetic 10 profiles following i.v. injection of formaldehyde or formate were similar and exhibited 11 the same characteristics as described above following inhalation exposure. There was a 12 rapid decline in radioactivity in both the plasma and the packed-cell fraction following 13 i.v. administration of formaldehyde or formate. Plasma concentrations then gradually 14 declined. Concentrations in the packed-cell fraction increased after the initial decline, 15 peaked after about 35 h, and then slowly declined just as was observed following 16 inhalation exposure.

17 Chang et al. (1983) investigated nasal cavity deposition and toxicity of formaldehyde in 18 male F344 rats and B6C3F<sub>1</sub> mice. Groups of naïve and pretreated rats and mice (whole 19 body exposure to 6- or 15-ppm formaldehyde, 6 hours/day for 4 days or 5 days were exposed (head only) to <sup>14</sup>C-formaldehyde at 15 ppm for 6 hours. The amounts of 20 21 radioactivity deposited in the nasal cavity of pretreated and naïve male F344 rats were 22 similar, while naïve male  $B6C3F_1$  mice had more radioactivity in the nasal cavity than 23 pretreated mice. In both rats and mice, pretreated animals had less visceral radioactivity 24 than naïve animals. This was attributed to decreased grooming and impaired mucociliary 25 clearance in pretreated animals.

The concentrations of formaldehyde in the blood of rats, monkeys, and humans did not increase after inhalation exposure to formaldehyde. Heck *et al.* (1985) investigated the effect of formaldehyde exposure on the concentrations in blood of rats and humans. Eight male F344 rats were exposed by inhalation to 14 ppm formaldehyde for 2 hours, and blood samples were collected immediately after exposure. The mean concentration of

- 1 formaldehyde in the exposed group was  $2.25 \pm 0.07 \,\mu$ g/g of blood compared to  $2.24 \pm$
- $2 \quad 0.07 \,\mu\text{g/g}$  in eight unexposed rats. Formaldehyde concentrations in human blood were
- 3 measured in six volunteers before and after exposure to 1.9 ppm for 40 minutes. Mean
- 4 formaldehyde concentrations before exposure were  $2.61 \pm 0.14 \,\mu\text{g/g}$  compared with 2.77
- 5  $\pm 0.28 \ \mu g/g$  after exposure and were not significantly different. However, there was
- 6 considerable interindividual variation with both increases and decreases observed after
- 7 exposure (Table 5-2).

	Concentration (µg/g of blood)			
Subject (gender)	Before exposure	After exposure		
1 (female)	$3.09\pm0.41$	$2.18\pm0.09$		
2 (female)	$2.56\pm0.10$	$3.31\pm0.34$		
3 (male)	$2.66\pm0.17$	$3.74\pm0.13$		
4 (male)	$2.61\pm0.34$	$1.93\pm0.05$		
5 (male)	$2.05\pm0.16$	$2.76\pm0.21$		
6 (male)	$2.73\pm0.14$	$2.72\pm0.31$		
Mean	$2.61 \pm 0.14$	$2.77 \pm 0.28$		

 Table 5-2. Concentrations of formaldehyde in human blood before and after exposure to 1.9 ppm for 40 minutes

Source: Heck et al. 1985

8 Formaldehyde concentrations in the blood of three rhesus monkeys were measured 9 immediately after exposure to 6 ppm for 6 hours/day, 5 days/week, for 4 weeks and 10 compared to unexposed controls (Casanova et al. 1988). The average concentration of 11 formaldehyde in the exposed group was  $1.84 \pm 0.15 \,\mu\text{g/g}$  of blood and did not change significantly over the next 45 hours without further exposure  $(2.04 \pm 0.40 \ \mu g/g)$ . The 12 13 average concentration in the blood of unexposed controls was  $2.42 \pm 0.09 \,\mu\text{g/g}$ , which 14 indicates that subchronic exposure to formaldehyde did not have a significant effect on 15 formaldehyde concentrations in the blood of monkeys. McMartin et al. (1979) slowly infused a dose of 1 mmol/kg <sup>14</sup>C-formaldehyde into the femoral vein of two Cynomolgus 16 17 monkeys over a 3- to 4-minute period and collected blood samples from the femoral 18 artery on the same side. The specific activity of the solution was 1,500 dpm/µmol for one 19 monkey and 115,000 dpm/µmol for the other. Formaldehyde was detected for about 5 20 minutes after infusion with the lower specific activity solution, but was detected for up to 1 60 minutes when the higher specific activity solution was used. In both cases, the

2 elimination half-life from the blood was about 1.5 minutes.

3 5.1.2.2 Oral exposure

Feeding studies in rats, mice, rabbits, and livestock (described below) show that
formaldehyde is readily absorbed from the gastrointestinal tract (Barry and Tomé 1991,
Buckley *et al.* 1988, Galli *et al.* 1983, Nishi *et al.* 1988); however, no studies specifically
reporting absorption and distribution of radiolabeled formaldehyde were identified. In
addition, several cases of formaldehyde poisoning by ingestion in humans have been
described (ATSDR 1999). These studies show that formic acid rapidly accumulates in the
blood following formaldehyde ingestion.

Galli et al. (1983) fed grana cheese that contained <sup>14</sup>C-formaldehyde to groups of male 11 12 Sprague Dawley rats and male Swiss albino mice. Commercial grana cheese is normally 13 made with milk that has formaldehyde added as a bacteriostatic agent. In this experiment, unlabeled and <sup>14</sup>C-labeled formaldehyde were added to the milk to obtain a final 14 15 concentration of 35 to 40 ppm, and grana cheese was made following the usual process. 16 Animals were placed individually in metabolism cages and fed 2.2 g (rats) or 0.5 g (mice) 17 of radiolabeled cheese. Controls were fed unlabeled cheese. Rats were killed at 4, 8, 16, 18 32, or 64 hours, and mice were killed after 2, 4, 8, 16, 32, 64, and 96 hours, and 8 and 12 19 days after the end of treatment. The decay of radioactivity was measured in the plasma, 20 liver, gastrointestinal tract, kidneys, spleen, testes, brain, muscle, adipose tissues, urine 21 and feces. The toxicokinetic profile was similar in rats and mice. The half-lives of the 22 elimination phase were 27.8 hours in mice and 26.4 hours in rats. Excretion of 23 radioactivity was essentially complete after 32 hours in both species with about 64% to 24 67% eliminated in the urine and feces and 24% to 28% eliminated as expired CO<sub>2</sub>. In 25 rats, maximum radioactivity in the tissues occurred at 16 hours while maximum activity 26 in the blood reached about 0.08% of the dose after 8 hours. In mice, peak concentrations 27 in the tissues occurred at 4 hours. The highest concentration measured in the blood was about 0.03% of the dose and occurred after 2 hours. However, the authors noted that  $^{14}C$ -28 29 activity did not accumulate in the tissues of rats or mice, and that the low levels of

radioactivity still present 32 hours after administration were likely due to residues of
 labeled fractions in milk proteins that had not been completely metabolized.

3 Buckley et al. (1988) measured the levels of formaldehyde in milk and blood of Holstein 4 dairy cows fed diets that included formalin-preserved whey. The experiment was divided 5 into three trials lasting 35 days each with a 2-week interval between trials. Six cows were 6 fed untreated whey, and six cows were fed whey treated with 0.05% (0.0185%) 7 formaldehyde) (trial 1), 0.1% (0.037% formaldehyde) (trial 2), or 0.15% (0.0555% 8 formaldehyde) (trial 3) formalin. Morning milk samples were collected 3 days prior to 9 beginning each trial, on days 2 through 6, 13, 27, and 34 of each trial, and 46 hours after 10 the end of trial 3. Blood samples were collected 3 days prior to the beginning of trial 3, 11 and on days 9, and 33 of that trial. Levels of formaldehyde in milk samples from the 12 control group were below the detection limit of 0.026 mg/kg. Formaldehyde was detected 13 in milk samples collected in the treatment groups at average concentrations of 0.034, 14 0.095, and 0.208 mg/kg in the three trials; however, levels were below the detection limit 15 prior to beginning each trial and at 46 hours after the end of trial 3. During the first trial, 16 formaldehyde was detected in milk samples from only 3 of the 6 cows. Formaldehyde 17 concentrations did not increase over time and there was no significant effect due to day of 18 milk collection during any of the trials. Concentrations in blood were significantly higher 19 (P < 0.01) in the treatment group at day 33 of trial 3 compared with the control group. In 20 another experiment, bull calves were fed diets containing 0, 0.05%, or 0.1% formalin and 21 sacrificed at days 81, 88, and 95. Formaldehyde concentrations were measured in blood, 22 muscle, kidney, liver, and heart tissue. Formaldehyde concentrations were higher in the 23 muscle tissue of the high-dose group but did not differ among treatment groups in the 24 other tissues. About 0.0038% to 0.0067% of ingested formaldehyde was eliminated in the 25 milk. Barry and Tome (1991) also reported a dose-related increase in formaldehyde 26 concentrations in milk from goats fed 0, 0.63, or 1.26 g of formaldehyde daily in soybean 27 oil-meal. Approximately 0.02% of the ingested formaldehyde was excreted in the milk. 28 Nishi *et al.* (1988) published a case report of a 52-year-old man that had committed

suicide by ingesting formalin. There was an obvious odor of formaldehyde in the stomachand air passages. Formaldehyde and formic acid were detected in the serum, brain, heart,

September 3, 2009

- 1 lungs, liver, spleen, pancreas, kidneys, and gastric contents (Table 5-3). Formic acid is
- 2 the primary metabolite of formaldehyde (see Section 5.3). The urine also contained
- 3 formic acid. These authors also conducted a study in two male rabbits that were
- 4 administered an oral dose of 15 mL/kg of formalin. These animals died after 12 minutes.
- 5 Formaldehyde, methyl alcohol, and formic acid were detected in serum, brain, heart,
- 6 lungs, liver, spleen, and kidneys (Table 5-3).

	Concentration (µmol/g)							
	Hun	nan <sup>a</sup>	Rabbits <sup>b</sup>					
Tissue/body fluid	Formaldehyde	Formic acid	Formaldehyde	Formic acid				
Brain	1.5	5.39	4.33-6.63	3.60-5.12				
Heart	1.63	11.60	1.70-1.87	9.42-10.59				
Lungs	0.77	13.99	0.40-0.53	14.19–14.68				
Liver	5.63	16.44	10.76-23.48	21.39-24.71				
Spleen	6.89	11.48	1.80 - 2.00	5.80-5.93				
Pancreas	11.09	14.42	NR	NR				
Kidneys	1.4	11.54	5.71-5.86	14.82–15.53				
Gastric contents	233.10	ND	NR	NR				
Serum	1.10	11.79	6.39–7.03	9.75-11.48				
Urine	ND	ND	NR	NR				

 Table 5-3. Formaldehyde and formic acid concentrations detected in body fluids

 and tissues following formaldehyde ingestion

Source: Nishi et al. 1988.

NR = not reported, ND = not detected.

<sup>a</sup> 52-year-old male suicide case.

<sup>b</sup>Range for two rabbits.

# 7 5.1.2.3 Dermal exposure

8 Very few studies have investigated absorption and distribution of formaldehyde

9 following dermal exposure, but the available data indicate that formaldehyde is poorly

10 absorbed from the skin. However, Maibach (1983) noted that if some amount of

11 formaldehyde or its metabolites did not penetrate, allergic contact dermatitis could not

12 occur (see Section 5.4.2.2). Jeffcoat et al. (1983) administered 10 µL of an aqueous

- 13 solution containing 0.1 mg of <sup>14</sup>C-formaldehyde or 40  $\mu$ L containing 11.2 mg of <sup>14</sup>C-
- 14 formaldehyde to the skin of F344 rats or Dunkin-Hartley guinea-pigs (5 to 6 males and
- 15 females per group), and 2 mg in 200 µL to three Cynomolgus monkeys. Urine, feces,
- 16 expired air, and evaporation products were collected. Blood samples were collected from
- 17 a catheter implanted in the carotid artery at 1, 2, 3, 4, 7, and 24 hours after dosing.

1 Animals were sacrificed 72 hours after dosing, and tissue samples from the heart, liver, 2 lung, spleen, kidney, leg, brain, gonads, skin at the application site, distant skin, and the remaining carcass were analyzed for <sup>14</sup>C content. The mean values of recovered <sup>14</sup>C are 3 shown in Table 5-4. There was no accumulation of <sup>14</sup>C in any tissue in any species. Blood 4 5 concentrations were stable throughout the experiment, averaging about 0.015% of the 6 administered dose in monkeys and about 0.1% of the dose in rats and guinea-pigs. In rats 7 and guinea pigs, about 4.5% to 8.3% of the applied radioactivity was detected in the 8 urine, 0.7% to 1.5% in the feces, and 21.4% to 28.3% in the air traps; 22.2% to 28.4% 9 remained in the carcass. Almost the entire air-trapped radioactivity was due to evaporation from the skin because less than 3% was <sup>14</sup>CO<sub>2</sub>. The amount of radioactivity 10 11 remaining in the skin ranged from 3.8% to 15.6% in guinea-pigs and 3.4% to 16.2% in 12 rats. Although the percentage of the dose remaining in the skin was lower for the high 13 dose, the actual amount of radioactivity was still higher compared with the low dose. In 14 monkeys, about 0.24% of the applied dose was excreted in the urine, 0.2% was excreted 15 in the feces, 0.37% was exhaled, and about 9.5% remained in the skin at the site of 16 application. Data were not reported for the amount remaining in the carcass of monkeys. 17 The authors concluded that the skin of the monkey was much less permeable to 18 formaldehyde than that of rodents, and that the large majority of applied radiolabel was 19 lost to evaporation.

			=		=	-	=	
Species	Dose (mg)	Air traps	Urine	Feces	Skin (application site)	Carcass	Total <sup>14</sup> C recovered	Mean blood content
Rat	0.1	$28.3\pm2.4$	$5.0\pm0.6$	$1.5\pm0.5$	$16.2 \pm 1.4$	$22.2\pm1.2$	$73.4\pm3.1$	$0.12\pm0.01$
Guinea-pig	0.1	$21.4\pm1.6$	$4.5\pm1.0$	$1.4\pm0.2$	$15.6\pm2.5$	$27.1 \pm 1.7$	$70.0\pm3.7$	$0.10\pm0.02$
Rat	11.2	$22.1\pm2.6$	$8.3\pm1.0$	$0.7\pm0.1$	$3.4 \pm 0.4$	$25.9 \pm 1.9$	$60.4\pm2.6$	$0.13\pm0.01$
Guinea-pig	11.2	$23.8\pm3.1$	$6.8\pm1.1$	$1.2\pm0.4$	$3.8\pm0.5$	$28.4 \pm 1.6$	$63.6\pm2.6$	$0.09\pm0.01$

 $9.49 \pm 3.9$ 

 $0.2 \pm 0.12$ 

Table 5-4. Distribution of <sup>14</sup>C-labelled formaldehyde in rodents and monkeys during the first 72 h after topical administration<sup>a</sup>

Source: Jeffcoat et al. 1983.

NA = not analyzed.

Monkey

<sup>a</sup> Data are reported as % of administered dose  $\pm$  SE.

 $0.37\pm0.17$ 

 $0.24 \pm 0.1$ 

2.0

[~10]

NA

 $0.015 \pm 0.0006$ 

1 Bartnik *et al.* (1985) applied <sup>14</sup>C-formaldehyde and non-labeled formaldehyde mixed into

2 a cream at a concentration of 0.1% to the clipped backs of male and female rats.

3 Radioactivity was measured in feces, urine, expired air, carcass, and treated skin.

4 Between 60% and 70% of the radioactivity remained in the skin. Levels in the urine

5 ranged from about 1.2% to 3.5% of the applied radioactivity. Feces contained 0.2% to

6 0.8%, and the expired air contained 0.8% to 1.3% of the applied radioactivity.

7 Iverson *et al.* (1986) tested the possible carcinogenic potency of formaldehyde when

8 applied to the skin of Oslo hairless mice (see Section 4.2.2). Mice received topical

9 applications of 200 µg of 1% or 10% formaldehyde on the back skin twice a week and

10 were observed for 60 weeks. [No blood or tissue samples were examined for the presence

11 of formaldehyde or its metabolites.] Animals that received 1% formaldehyde had no skin

12 lesions. Slight hyperplasia of the epidermis was reported for animals treated with 10%

13 formaldehyde. A few animals had small skin ulcers or scratches and two animals had

14 small nonspecific granulomas in the lungs. No lesions were reported in the brain or other

15 tissues.

16 5.1.2.4 Parenteral and transplacental exposure

17 Keefer *et al.* (1987) injected <sup>14</sup>C-labeled formaldehyde and sodium formate (i.p.) into

18 male Sprague-Dawley rats and measured the cumulative excretion of carbon dioxide.

19 Approximately 70% of the administered dose was excreted as carbon dioxide within the

20 first 12 hours. The data showed that excretion was biexponential with half-lives of

21 approximately 0.4 hours and 3 hours for the two phases.

22 Katakura *et al.* (1993) administered <sup>14</sup>C-formaldehyde i.v. to pregnant mice and measured

23 the distribution in maternal and fetal tissues and blood. Radioactivity was found

24 immediately after injection and showed strong accumulation and retention 3 hours after

- 25 injection. Maternal liver, intestinal mucosa, bone marrow, kidneys, and salivary glands
- 26 showed the highest activity. Radioactivity was found in the fetus 6 hours after injection at
- 27 concentrations similar to those in maternal tissues. Elimination of radioactivity from the
- 28 placenta and fetus was slower than from maternal tissues.

Thrasher and Kilburn (2001) also investigated the distribution of <sup>14</sup>C-labeled 1 2 formaldehyde in maternal and fetal tissues. Pregnant ICR mice were injected with 0.05 3 mL of a 1% formalin solution that contained 3.5 mg of labeled compound via the tail vein 4 on the 16th day of gestation. The animals were killed at intervals from 5 minutes up to 48 5 hours. There was a rapid uptake of radioactivity into maternal liver, lung, heart, salivary 6 glands, gall bladder, spleen, kidney, bone marrow, nasal mucosa, uterus, placenta, and 7 fetal tissues. The placenta, uterus, and fetal tissues had the highest concentrations, and the 8 fetal brain had twice the concentration of radioactivity that was observed in the maternal 9 brain. Radioactivity appeared in urine and feces up to 6 h after treatment. The DNA 10 fraction from maternal and fetal liver contained 20% and 50% of the total radioactivity, 11 respectively after 6 hours. These values showed little change at 24 hours. Elimination 12 was slower from fetal tissues than maternal tissues.

#### 13 5.2 Airway deposition models

14 Morgan and Monticello (1990) reviewed the literature on the site specificity of nasal 15 lesions induced by exposure to inhaled gases with special reference to nasal airflow and 16 effects of formaldehyde. These authors reported that the distribution of nasal lesions is 17 influenced by the regional deposition of inhaled material, local tissue susceptibility, or a 18 combination of these factors. Nasal airflow patterns are particularly important in 19 determining lesion distribution for highly water-soluble or reactive gases such as 20 formaldehyde. Their review suggested that differences in nasal airflow patterns in rats 21 and monkeys were likely responsible for the characteristic differences in the distribution 22 of nasal lesions induced by formaldehyde in these species. This hypothesis has since been 23 investigated by several researchers using three-dimensional, anatomically accurate, 24 computational fluid dynamics (CFD) models.

It is very difficult to determine formaldehyde uptake patterns in nasal passages of experimental animals because of its rapid metabolism and reactivity, and because of the low resolution of dissection techniques used to obtain tissues samples from different locations in the rat nasal epithelium (Kimbell *et al.* 2001a). Therefore, CFD models of the nasal passages of the rat, monkey, and human have been developed (1) to determine the primary factors affecting nasal uptake, (2) to make interspecies dosimetric comparisons,

1 (3) to provide detailed anatomical information for the nasal passages of these species, and 2 (4) to provide estimates of regional air-phase mass transport coefficients (a measure of 3 the resistance to gas transport from inhaled air to airway walls) in the nasal passages 4 (Kimbell and Subramaniam 2001). These models allow investigators to examine the 5 relationship between the delivered dose at various sites in the respiratory tract to 6 biomarkers of dose or effect (*e.g.*, DNA-protein crosslinks or regional cell proliferation) 7 (Kimbell et al. 2001a). This section provides a brief review of these models. Section 8 5.7.5.1 discusses how these models have been used to predict crosslink and tumor 9 formation in rats, monkeys, and humans.

10 CFD models have been developed for the F344 rat (Kimbell et al. 1993, 1997), rhesus 11 monkey (Kepler et al. 1998), and human (Subramaniam et al. 1998) with the primary 12 objective of improving human health risk assessment. These models were developed in 13 three stages: (1) computer reconstructions of the nasal passages using sequential cross-14 sectional data, (2) simulation of steady-state inspiratory airflow for several volumetric flow rates (predicted flow streams and velocities from the simulations were compared 15 16 with observations and measurements made in hollow molds), and (3) simulation of 17 regional formaldehyde flux resulting from inspiratory airflow patterns and absorption 18 characteristics of the gas (Kimbell and Subramaniam 2001). The models were calibrated 19 by comparing predicted uptake data with actual measurements of formaldehyde uptake 20 and comparing predicted DNA-protein crosslink yield with measured crosslink yield and 21 adjusting model parameters accordingly.

22 CFD models use mathematical descriptions to simulate movement of inspired air in 23 respiratory air spaces and movement of inhaled chemical within air spaces via airflow 24 and diffusion (Kimbell et al. 1993). The concentrations of a chemical of interest that are 25 distributed throughout the respiratory tract are simulated by solving these equations. The 26 method involves dividing the nasal cavity into geometrically simple three-dimensional 27 elements to obtain a wire-frame grid of the nasal passage. The mass transport equations 28 are solved in each element and the elements are reassembled to produce simulated flow 29 and transport throughout the entire grid. Air-phase delivery is calculated as the mass flux of inhaled chemical at specific sites within the airway and incorporates airflow patterns
 and air-phase diffusion.

The CFD models have been used to test the hypothesis that the distribution of
formaldehyde-induced lesions can be attributed to species-specific patterns in
formaldehyde flux to various regions of the upper respiratory tract (Kimbell and
Subramaniam 2001). These studies show a strong correspondence between simulated
airflow-dependent transport patterns and local nasal lesion sites (see Section 5.7.5.1).

### 8 5.3 Metabolism

9 As discussed above, inhaled formaldehyde is rapidly absorbed by the epithelial cells of 10 the nasal mucosa of mammalian species. Once inside the epithelial layer, formaldehyde 11 binds rapidly and reversibly to glutathione and forms S-hydroxymethylglutathione 12 (Franks 2005). The nasal cavity has a substantial amount of enzyme activity, including 13 aldehyde dehydrogenases, cytochrome P-450 monooxygenases, glutathione transferases, 14 epoxide hydrolases, and carboxyl esterases; however, the two main enzymes responsible 15 for the rapid metabolism of formaldehyde are formaldehyde dehydrogenase (FDH) and S-16 formylglutathione hydrolase. FDH (which is also known as alcohol dehydrogenase 3 17 [ADH3]) oxidizes S-hydroxymethylglutathione to S-formylglutathione; S-18 formylglutathione is hydrolyzed by S-formylglutathione hydrolase to form reduced 19 glutathione and formic acid (Figure 5-1). FDH is a ubiquitous enzyme in mammals and is 20 widely distributed in various tissues (e.g., respiratory tract, liver, kidney, brain, muscle, 21 and erythrocytes). Therefore, formaldehyde metabolism occurs throughout the body 22 (ATSDR 1999). Øvrebø et al. (2002) demonstrated that cultured human bronchial 23 epithelial cells have formaldehyde biotransforming activity similar to that of hepatocytes 24 and are capable of oxidizing formaldehyde at a relatively fast rate at concentrations up to 25 3 mM. Casanova-Schmitz et al. (1984b) tentatively identified both FDH and aldehyde 26 dehydrogenase in nasal mucosal tissues from the rat nose and showed that homogenates 27 from both respiratory and olfactory epithelia efficiently oxidized formaldehyde. Other 28 enzymes that may catalyze the oxidation of formaldehyde to formate include catalase, 29 aldeyhde dehydrogenase, xanthinoxidase, peroxidase, aldehyde oxidase, and 30 glycerinaldehyde-3-phosphate dehydrogenase (WHO 1989). The contribution of

1 aldehyde dehydrogenases (ALDHs) increases with increasing concentrations of

2 formaldehyde (IARC 2006).

3 Formate, the primary metabolite of formaldehyde, enters the one-carbon pool, and can 4 either be excreted in the urine as the sodium salt, or be further oxidized to carbon dioxide 5 and exhaled (ATSDR 1999). Elimination of formate shows intra- and interspecies 6 variability, but elimination is generally slower than its formation. The plasma half-life of 7 formate in mammals ranges from about 1 to 90 minutes, with humans near the middle of 8 the range (WHO 1989). Øvrebø et al. (2002) investigated the capacity of human 9 bronchial epithelial cells and rat hepatocytes to metabolize formaldehyde to formate. 10 Normal human bronchial explants, primary bronchial epithelial cells, and rat hepatocytes 11 were grown in medium containing 0.5 to 5 mM formaldehyde for up to 48 hours. Human 12 bronchial explants and epithelial cells were shown to metabolize formaldehyde to 13 formate at a relatively fast rate, which was comparable with that measured for rat 14 hepatocytes. 15 Unmetabolized formaldehyde also may react non-enzymatically with sulfhydryl groups

16 or urea, form protein-protein crosslinks, or form protein-DNA crosslinks (single-stranded

17 DNA only) or form nucleic acid-nucleic acid crosslinks (single-stranded nucleic acids

18 only) (Figure 5-2). Formate can combine with tetrahydrofolate enzymatically and enter

19 the single-carbon intermediary metabolic pool. The availability of tetrahydrofolate,

20 derived from folic acid in the diet, determines the rate of formate metabolism.



Figure 5-1. Metabolism and fate of formaldehyde Adapted from IARC (2006).



Figure 5-2. Biological reactions of formaldehyde

Adapted from Bolt 1987: cys = cysteine,  $C_1$  = single carbon pool,  $TH^4$  = tetrahydrofolate.

### 1 5.4 Toxic effects

2 The toxicity of formaldehyde has been extensively reviewed (ATSDR 1999, WHO 2002, 3 IARC 2006); however, the exact mechanisms are not completely understood. Although 4 formaldehyde is a normal intermediary cellular metabolite, it is cytotoxic at high 5 concentrations ( $\geq$  6 ppm in the rat and rhesus monkey) (Casanova *et al.* 1994, Chang *et* 6 al. 1983, Monticello et al. 1991, Monticello et al. 1996). The carbonyl atom of 7 formaldehyde is electrophilic; thus, it readily reacts with nucleophilic sites on cell 8 membranes and in body tissues such as amino groups in protein and DNA (ATSDR 9 1999). This section provides an overview of the toxic effects reported from *in vitro* 10 studies, humans, and experimental animals. The following discussion summarizes the 11 findings from the IARC (2006) and other reviews, as well as relevant studies published 12 after the IARC review.

13 5.4.1 In vitro toxicity studies

14 In vitro studies conducted with human and animal cells demonstrate that formaldehyde is

15 cytotoxic, and affects cell proliferation, gene expression, apoptosis, and the mucociliary16 apparatus (IARC 2006).

17 Schäfer *et al.* (1999) showed a reduced frequency of ciliary beat in cultured human nasal 18 epithelial cells exposed to  $5 \text{ mg/m}^3$  [4 ppm] for 2 hours but no effect when exposed to 5 19 mg/m<sup>3</sup> for 1 hour or 0.5 mg/m<sup>3</sup> for 2 hours.

20 Lovschall et al. (2002) investigated the cytotoxic effects of formaldehyde in human 21 dental pulp fibroblasts, human buccal epithelial cells, and HeLa cervical cancer cells. The 22 purpose of this study was to compare the relative sensitivity of human target tissue cells 23 with that of an established human cancer cell line. Dose-response relationships and  $TC_{50}$ 24 values were determined using three different assays: bromodeoxyuridine (BrdU) 25 incorporation, neutral red uptake, and methylthiazole tetrazolium (MTT) conversion. Cell 26 cultures were exposed for 24 hours to graded formal dehyde dilutions based on  $TC_{50}$ 27 estimates obtained in pilot studies for each cell type. Dental pulp fibroblasts and buccal 28 epithelial cells had significantly lower TC<sub>50</sub> values in both the BrdU and neutral red

29 assays compared with HeLa cells. There were no statistically significant differences

1 among the cell types with the MTT assay. Overall dental pulp fibroblasts and buccal

2 epithelial cells appeared to be more sensitive to formaldehyde toxicity than HeLa cells.

Other *in vitro* studies reported effects on glutathione levels and oxidative stress. These
studies are discussed in Section 5.7.2.

5 5.4.2 Toxic effects in humans

6 A wide range of health effects have been associated with exposure to formaldehyde in 7 both residential and occupational settings. These effects are summarized below and are 8 route dependent. The most common effects include irritation at the point of contact 9 following inhalation (upper respiratory tract and eyes), oral (mouth and gastrointestinal 10 tract), or dermal exposure (skin and eyes). Other effects include allergic contact 11 dermatitis, histopathological abnormalities (e.g., hyperplasia, squamous metaplasia and 12 mild dysplasia) of the nasal mucosa, occupational asthma, reduced lung function, 13 neurophysiological disorders (e.g., insomnia, memory loss, mood alterations, and loss of 14 appetite), and altered immune response. Formaldehyde concentrations associated with 15 reported effects in humans show wide interindividual variation as illustrated in Table 5-5. 16 Although some symptoms have been reported at concentrations as low as 0.05 ppm 17 [primarily sensory irritation], they occur only rarely at concentrations below 0.5 ppm 18 (IARC 2006). Paustenbach et al. (1997) reviewed approximately 150 articles in order to 19 recommend an occupational exposure limit for formaldehyde based on irritation. They 20 reported that eye irritation did not occur in most people at concentrations < 1 ppm, and 21 that moderate to severe irritation did not occur until airborne concentrations exceeded 2 22 to 3 ppm. Persons exposed to 0.3 ppm for 4 to 6 hours in chamber studies reported eye 23 irritation at a rate similar to that reported by persons exposed to clean air. Arts et al. 24 (2006) also reviewed data on respiratory irritation of formaldehyde and reported that 25 mild/slight eye irritation was observed at levels  $\geq 1$  ppm, and mild/slight respiratory tract 26 irritation at levels  $\geq 2$  ppm.

Reported effects	Formaldehyde concentration (ppm)
Neurophysiological effects	0.05–1.05
Odor threshold	0.05–1.0
Eye irritation	0.05–2.0
Upper airway irritation	0.1–25
Lower airway and pulmonary effects	5.0–30
Pulmonary edema, inflammation, pneumonia	50-100
Death	$\geq 100$

 Table 5-5. Formaldehyde concentrations associated with various health effects

Source: Newell 1983.

### 1 5.4.2.1 Inhalation exposure

2 Inhalation is an important exposure pathway for formaldehyde in occupational, domestic,

3 and environmental settings. In addition to the epidemiologic studies and case reports, a

4 number of controlled studies of human exposure to formaldehyde have been conducted.

5 The most common and consistently reported effects include sensory and airway irritation.

6 Some studies indicate an association with occupational asthma. Effects associated with

7 acute and chronic exposures are discussed. Studies that indicate an association with

8 occupational asthma are reviewed briefly in a separate section.

# 9 Acute exposure

10 Ballenger (1984) reported that nasal mucous membranes may begin to swell at formaldehyde concentrations of 0.16 ppm  $[0.2 \text{ mg/m}^3]$ , and chest tightness and coughing 11 occur at about 1.2 ppm [1.5 mg/m<sup>3</sup>]. IARC reviewed 10 controlled experimental studies 12 13 of acute inhalation exposure to formaldehyde (Table 5-6). These studies included healthy 14 individuals, asthmatics, and individuals with allergic symptoms due to exposure to 15 formaldehyde. These individuals were exposed to 0.4 to 3 ppm  $[0.49 \text{ to } 3.7 \text{ mg/m}^3]$ 16 formaldehyde for 30 minutes to 3 hopurs. Reported effects included eye, nose, and throat 17 irritation; nasal itching; congestion; and sneezing. One study evaluated dose-response 18 effects and reported that eye irritation increased linearly at doses from 0.5 to 3 ppm [0.62] to 3.7 mg/m<sup>3</sup>]; no effects were observed at 0.5 ppm. Exposure to 3 ppm for 1 hour while 19 20 exercising resulted in moderate to severe eye irritation in 27% of healthy subjects and 21 19% of asthmatics. Moderate to severe nose and throat irritation occurred in 32% of the 22 healthy subjects and 31% of asthmatics. IARC (2006) also cited a review by Bender et al. 23 (2002) who reviewed 9 controlled chamber studies of asthmatic subjects. Exposure to 2

- 1 to 3 ppm  $[2.5 \text{ to } 3.7 \text{ mg/m}^3]$  for up to 3 hours did not provoke asthma in unsensitized
- 2 asthmatics, and exposure to 0.1 to 3 ppm  $[0.12 \text{ to } 3.7 \text{ mg/m}^3]$  did not provoke asthma in
- 3 men or women who reported chest tightness, cough, and wheeze when exposed to
- 4 formaldehyde at home or work.

Subjects (no.)	Exposure (mg/m <sup>3</sup> )	Results	References (as cited in IARC 2006)
Healthy (22) Asthmatics (16)	3.7 (1 h)	Moderate to severe symptoms in both groups Eye (27%), nose/throat (32%) (healthy) Eye (19%), nose/throat (31%) (asthmatics)	Green et al. 1987
Healthy (10) Asthmatics <sup>a</sup> (10)	0.5 (2 h)	Krakowiak <i>et al</i> . 1998	
Healthy (19)	0.6–3.7 (3 h) Eye irritation increased linearly with dose; mild nose and throat irritation threshold at 1 ppm [1.2 mg/m <sup>3</sup> ]		Kulle 1993, Kulle <i>et al.</i> 1987
Healthy (11) Contact dermatitis (9)	0.5 (2 h)	Mean nasal score (sneezes, itching and congestion) of 4 at 10 minutes in both groups	Pazdrak et al. 1993
Healthy (9)	3.7 (3 h)	Increase in mean symptom scores for eyes, nose and throat irritation after exposure	Sauder et al. 1986
Asthmatics (9)	3.7 (3 h)	Eye and nose irritation after 2 min	Sauder et al. 1987
Healthy (15)	2.5 (40 min)	Odor (80%), sore throat and nasal irritation (0%), eye irritation (47%)	Schachter et al. 1987
Asthmatics (15)	2.5 (40 min)	Odor (100%), sore throat (33%), nasal irritation (47%), eye irritation (73%)	Witek et al. 1987
Healthy (9) Asthmatics <sup>b</sup> (9)	3.7 (2 h) 1.2 (90 min) 2 (30 min) <sup>c</sup>	Eye (83%), nose (39%) and throat (28%) irritation; no significant differences between groups.	Day <i>et al.</i> 1984

Table 5-6. Irritant effects of formaldehyde following acute inhalation exposures

Adapted from IARC 2006.

<sup>a</sup> Subjects had allergic symptoms due to formaldehyde exposure.

<sup>b</sup> Subjects with urea-formaldehyde foam insulation symptoms.

<sup>c</sup> Exposure to urea-formaldehyde foam insulation.

5 Nasal lavage studies of workers who had skin hypersensitivity (positive patch test) to

- 6 formaldehyde and healthy men with a negative patch test showed similar responses
- 7 following a 2-hour exposure to 0.5 mg/m<sup>3</sup> [0.41 ppm] formaldehyde (Pazdrak *et al.*
- 8 1993). In both groups, eosinophils peaked shortly after exposure and were still elevated
- 9 after 18 hours, while the percentage of epithelial cells was reduced. Albumin levels also
- 10 were increased. The authors concluded that a non-specific, non-allergic pro-inflammatory
- 11 effect occurred from exposure to low concentrations  $(0.5 \text{ mg/m}^3)$  of formaldehyde.

1 Lang et al. (2008) conducted a controlled study in Germany of sensory irritation in 21 2 healthy volunteers (11 males and 10 females) exposed to formaldehyde. Each subject was 3 exposed for 4 hours to each of 10 selected exposure conditions on 10 consecutive 4 working days. The 2-week exposure sequences were randomized. Formaldehyde 5 concentrations ranged from 0 to 0.5 ppm. During three of the exposures, the 6 concentration of formaldehyde was doubled to generate intermittent exposure to peak 7 concentrations four times during the exposure period. Once the peak concentration was 8 reached, forced ventilation of the exposure chamber was used to reduce the concentration 9 back to the desired base level. During 4 of the 10 exposures, ethyl acetate at 12 to 16 ppm 10 was used as a masking agent for formaldehyde. Measurements included conjunctival 11 redness, blinking frequency, nasal flow and resistance, pulmonary function, and reaction 12 times. There were no significant treatment effects on nasal flow and resistance, 13 pulmonary function, and reaction times. Blinking frequency and conjunctival redness 14 were significantly increased by short-term peak exposures of 1 ppm. Subjective ratings 15 indicated eye and olfactory symptoms at concentrations as low as 0.3 ppm. Eye irritation 16 was the most sensitive parameter. All increased symptom scores returned to normal 17 levels 16 hours after the end of the exposures.

Tang *et al.* (2009) reported that 17 employees at a pharmaceutical company in China who were continuously exposed to formaldehyde vapors showed symptoms of eye irritation, tearing, sneezing, coughing, chest congestion, fever, heartburn, lethargy, and loss of appetite. Some of the workers also experienced vomiting, abdominal pain, and tachycardia.

### 23 Chronic exposure

24 IARC (2006) reviewed six occupational studies and one residential study that

25 investigated the effects of chronic inhalation exposure to formaldehyde on the nasal

26 mucosa (Table 5-7). The average length of employment ranged from 10 to 20 years in the

- 27 occupational studies. Time-weighted average exposure levels ranged from 0.007 to 2.4
- 28 ppm with a peak concentration as high as 18.5 ppm. The most common effects on the
- 29 nasal mucosa in the exposed groups were loss of cilia, goblet-cell hyperplasia, and
- 30 squamous metaplasia. Irritation of the upper respiratory tract and eyes was also common

1 among the exposed groups. Histological scores, based on severity of effect, were 2 significantly higher in the exposed group compared with matched controls in most of the 3 studies; however, there was not always a clear association with exposure to formaldehyde 4 [i.e., no concentration-response relationship or no correlation between histological score 5 and duration of exposure]. Two of the studies did not show significant differences 6 between the exposed and control groups. Atypical squamous metaplasia was associated 7 with age in at least one study. The residential study reported that the prevalence of 8 squamous metaplasia was significantly increased in occupants of urea-formaldehyde 9 foam-insulated homes compared with subjects who lived in homes without this type of 10 insulation.

11 IARC (2006) also reviewed three studies (Akbar-Khanzadeh and Mlynek 1997, Akbar-12 Khanzadeh et al. 1994, Kriebel et al. 1993) that investigated the effects of formaldehyde 13 exposure on lung function in groups of physical therapy or medical students and their 14 instructors. Pulmonary function (peak expiratory flow or forced expiratory volume in 1 15 second) was measured before and after completing laboratory sessions, or was compared 16 with a group of unexposed controls. Formaldehyde concentrations ranged from about 0.07 to 2.94 ppm [0.09 to 3.6 mg/m<sup>3</sup>]. These studies included 24 to 50 subjects that were 17 18 exposed to formaldehyde during anatomy classes. Eye nose and throat irritation were 19 common in the exposed groups. Formaldehyde exposure was associated with lung 20 function decrements in all three studies.

21 In a review on occupational formaldehyde exposure in China, Tang *et al.* (2009)

22 identified six reports of pulmonary disorders in factory workers chronically exposed to

formaldehyde. One study reported that workers exposed to  $3.07 \pm 5.83 \text{ mg/m}^3$  had

24 decreased pulmonary ventilation compared with a control group. Another study reported

25 that chronic exposure to a lower concentration (1.3 mg/m<sup>3</sup>) significantly decreased mid-

26 expiratory airflow and forced vital capacity values [data not reported]. Other studies

27 showed exposure-related increases in pulmonary damage over time, more abnormalities

28 in the small airways, and higher resistance to pulmonary ventilation.

- 1 Lyapina *et al.* (2004) reported a statistically significant (P = 0.02) predominance of
- 2 subjective symptoms and clinical findings of chronic upper respiratory tract inflammation
- 3 among 29 workers (13 men and 16 women) occupationally exposed to formaldehyde for
- 4 an average of 12.7 years. Results were compared with 21 non-exposed, age- and gender-
- 5 matched controls. Further details of this study are provided in Section 5.4.2.4.

Exposure setting	Concentration <sup>a</sup> (mg/m <sup>3</sup> )	No.	Histological score <sup>b</sup>	Comments	Reference
Laminate plant	0 0.5–1.1	25 38	1.8 2.8*	Smoking had a slight modifying effect; no correlation of histological score and exposure duration; four cases of mild dysplasia in the exposed group	Edling <i>et al.</i> 1987a
Particle board or laminate plant	0 0.1–1.1 (peaks to 5)	25 75	1.8 2.9*	Some exposure to wood dust, but no dose-response relationship; no differences between workers exposed only to formaldehyde compared with those exposed to formaldehyde and wood dust; six exposed men had mild dysplasia	Edling et al. 1988
Phenol-formaldehyde resins used in paper processing	0 0.2–2.4 (peaks to 11–18.5)	38 42	NR	Higher prevalence of mucosal irritation was reported in non-smoking exposed workers compared with controls ( $P = 0.04$ ); however, cytologic exams did not show a statistical relationship to formaldehyde exposure	Berke 1987
Formaldehyde and formaldehyde resins production plant	0 0.5-> 2.0	37 37	1.4 1.9	Incidence of subjective nasal complaints was significantly higher ( $P < 0.01$ ) in the exposed group, mild dysplasia in 3 exposed workers	Boysen et al. 1990
Formaldehyde resin or particle board production	0 0.05–0.5 0.2–0.3 <sup>b</sup>	32 62 89 <sup>b</sup>	1.56 2.16* 2.07 <sup>c</sup>	No correlation between duration of exposure and histological changes, 2 cases of dysplasia among particle board workers who ground wood for $> 4$ h/day	Holmström <i>et al.</i> 1989b
Plywood factory and warehouse	0 0.1–0.39	15 15	1.6 2.3**	Co-exposure to wood dust, significantly higher ( $P < 0.01$ ) incidence of micronuclei in exposed workers, one case of mild dysplasia in the exposed group	Ballarin <i>et al</i> . 1992
Residential (homes with and without urea-formaldehyde foam insulation)	0.007–0.14 0.009–0.28	720 1,726	NR	Positive relationships between level of exposure and the presence of symptoms, a number of exposure-response relationships were enhanced by urea-formaldehyde, small but significant increase in incidence of squamous-metaplasia in occupants of urea- formaldehyde insulated homes	Broder <i>et al.</i> 1991, 1988

 Table 5-7. Effects on the nasal mucosa from chronic exposure to formaldehyde

Adapted from IARC 2006.

\* P < 0.05; \*\* P < 0.01.

NR = not reported

<sup>a</sup>Time-weighted average concentrations for occupational settings.

<sup>b</sup>Several different scales were used by the authors. Edling *et al.* 1987, 1988 and Holmstrom *et al.* 1989b used an 8-point scale (0 = normal to 8 = carcinoma); Boysen *et al.* 1990 used a 5-point scale (0 = pseudostratified columnar epithelium to <math>5 = dysplasia), and Ballarin *et al.* 1992 used a 6-point scale (1 = normal cellularity to 6 = malignant cells). <sup>c</sup>Co-exposed to wood dust.

### 1 Occupational asthma

- 2 Inhalation exposure to formaldehyde has also been identified as an occasional cause of
- 3 occupational asthma. IARC (2006) reviewed eight studies (some were case reports) of
- 4 occupational asthma in workers (Table 5-8). Hypersensitivity is thought to be the likely
- 5 mechanism because the reactions were often delayed and unsensitized asthmatics did not
- 6 react to the same concentrations. Asthmatic reactions may also be caused by an irritant
- 7 mechanism at high concentrations. Tang *et al.* (2009) reported that the likelihood of
- 8 developing allergic asthma increases proportionately with indoor formaldehyde
- 9 concentrations, especially at concentrations  $> 0.12 \text{ mg/m}^3$ .

Study population (no.)	Sex	Concentration (mg/m <sup>3</sup> )	Duration	Results	References	
Workers (NR)	NR	NR	NR	Immediate and late reaction in 2 workers	Popa <i>et al.</i> 1969 (cited in IARC)	
Neurology resident (1)	Male	NR	2 h	Acute pneumonitis; breath smelled of formaldehyde, resolved in 5 wk	Porter 1975 (cited in IARC)	
Nurse (1)		[6.1]	15 min	Late asthmatic reaction	Hendrick and Lane 1975,	
Pathologist (1)	Female	[6.1]	1 h	No reaction	1977, Hendrick et al. 1982	
NR (1)		[3.7]	5 min	Late asthmatic reaction	(cited in IARC)	
	Both	2.3	30 min	One late asthmatic reaction		
W. 1 (15)		4.8	30 min	Two immediate and late asthmatic reactions	Burge <i>et al.</i> 1985 (cited in IARC)	
workers (15)		4.8	30 min	No reaction in unsensitized asthmatics		
		31	7 min	One irritant asthmatic reaction		
Washerry (220)	Deth	1.2	30 min	One early reaction	Nordman et al. 1985 (cited in	
workers (250)	Both	2.5	30 min	Five early and six late reactions	IARC)	
		[0.07]	6 mo	Asthma		
Worker (1)	Male	0.01	20 min	None	Kim <i>et al.</i> 2001 (cited in	
		0.6	20 min	Late asthmatic reaction, IgE negative	IARC)	
Residential				There was a significant relationship		
Controls (41)	Both	0.017	NR	between formaldehyde concentrations and	Norbäck et al. 1995	
Asthmatics (47)		0.029		asthma-like symptoms		

 Table 5-8. Studies of occupational asthma and formaldehyde exposure

Adapted from IARC 2006. NR = not reported.

### 1 5.4.2.2 Dermal exposure

2 Although formaldehyde is recognized as a skin irritant, very few quantitative data are 3 available. Maibach (1983) reported that it is likely that formulations containing formalin 4 at 300 ppm or greater would induce clinical irritation. Unlike contact dermatitis 5 (discussed below) skin irritation is non-immunologic (ConsensusWorkshop 1984). 6 Sensory irritation may be caused by nucleophilic addition, disulfide bond cleavage, and 7 physical interaction. Nucleophilic addition at -SH or  $-NH_2$  groups on proteins is probably 8 the most important mechanism for formaldehyde. Approximately 5% of subjects exposed 9 to a single application of 1% formalin in water with occlusion will develop skin irritation.

10 Formaldehyde is a primary skin sensitizing agent and has been associated with both 11 immediate, anaphylactic reactions (Type I allergy) and contact dermatitis (Type IV 12 allergy) (ConsensusWorkshop 1984). More quantitative data were available for contact 13 dermatitis than for skin irritation. The Consensus Workshop reported that the threshold 14 level for induction of contact dermatitis in humans is less than 5% formalin in water. 15 Approximate thresholds for elicitation of allergic contact dermatitis in sensitized subjects 16 range from about 30 ppm for patch testing to 60 ppm for actual use concentrations of 17 formalin. Flyvholm et al. (1997) conducted patch tests with formaldehyde solutions 18 ranging from 25 to 10,000 ppm in 20 formaldehyde-sensitive individuals and 20 healthy 19 controls and reported a threshold concentration of 250 ppm. No positive reactions were 20 observed in the control group. Maibach (1983) reported rates of allergic contact 21 dermatitis (patch test responders) ranging from about 3.5% to more than 6%. More recent 22 results indicated positive reaction rates of 7.9% in 1,324 patients at the Mayo Clinic and 23 9.2% from 5,830 patients tested by the North American Contact Dermatitis Group 24 (Wetter et al. 2005). Warshaw et al. (2007) reported that formaldehyde was the second 25 most common allergen associated with contact dermatitis of the hands in a cross-sectional 26 analysis of more than 22,000 patients patch tested between 1994 and 2004 in North 27 America. Zug et al. (2008) conducted a retrospective cross-sectional analysis of North 28 American contact dermatitis data from 2001 to 2004. Formaldehyde was the fourth most 29 frequently positive allergen (positive patch test in 170 of 1,496) among patients with a 30 scattered generalized distribution of dermatitis.

1 There are several case reports that document contact dermatitis from exposure to 2 formaldehyde in clothing. Formaldehyde resins were added to clothing to make 3 permanent creases, to make the garments wrinkle resistant, to preserve their new 4 appearance, for mothproofing, and to reduce shrinking. O'Quinn and Kennedy (1965) 5 and Shellow and Altman (1966) reported cases of intermittent or persistent dermatitis that 6 had lasted for years and typically involved the neck, shoulders, upper arms, lower legs, 7 feet, hands, and peripheral areas of the axillae. The patients also had positive patch tests 8 when exposed to 2% or 5% formaldehyde solutions, or when exposed to some samples of 9 clothing that contained formaldehyde. Fowler (2003) also reported a case of urticaria that 10 was associated with formaldehyde use in leather dresses in Finland, and a case of shoe 11 dermatitis in a woman who wore formaldehyde-treated leather shoes. Carlson et al. 12 (2004) conducted patch tests on 852 patients in the University Hospitals of Cleveland 13 Environmental and Occupational Dermatitis Clinic from August 1999 to April 2004. 14 Reactions to formaldehyde and to several formaldehyde textile resins were recorded. 15 Positive reactions to a 1% aqueous solution of formaldehyde were reported for 61 16 patients (7.2%), while 17 patients had a positive reaction to an ethylene urea/melamine 17 formaldehyde resin. Donovan and Skotnicki-Grant (2007) reported a case of severe 18 contact dermatitis in a 49-year-old pediatrician that was caused by contact with 19 formaldehyde textile resins in her hospital "greens" (or "scrubs") and mask. Patch testing 20 revealed a very strong reaction to melamine formaldehyde and milder reactions to urea 21 formaldehyde and ethylene urea/melamine formaldehyde.

22 De Groot *et al.* (1988) investigated the relationship between allergic contact dermatitis to 23 formaldehyde and patch test reactions to dimethyloldimethyl hydantoin [a formaldehyde 24 donor used as a preservative in cosmetic products]. Patients that had positive patch tests 25 to 0.1% or 0.3% formaldehyde tended to have a higher incidence of positive patch tests to the preservative than those who reacted only to 1% formaldehyde. Takahashi et al. 26 27 (2007) reported that 2 of 60 medical students had a positive patch test to 1% 28 formaldehyde at the end of a human anatomy class. None of the students had a positive 29 patch test prior to taking the anatomy class. Ravis et al. (2003) reported a 2% incidence 30 of formaldehyde-induced allergic contact dermatitis among 101 dental hygienists or 31 dental assistants. The incidence in 51 control subjects also was 2%.

1 Kiec-Swierczynska (1996) reported incidences of occupational allergic contact dermatitis 2 among 1,619 patients in Poland that were examined over a five-year period (1990 to 3 1994). A total of 332 patients were diagnosed with contact dermatitis. Medical histories 4 and occupational exposure data were obtained, and all patients were patch-tested with the 5 standard Polish series of allergens. Sixty individuals had a positive patch test to 6 formaldehyde. Geier et al. (2008) also reported positive patch tests to several 7 formaldehyde releasers in a 39-year-old metalworker with work-related dermatitis of the 8 hands and lower arms. Formaldehyde releasers were used as a biocide in the water-based 9 metalworking fluid used by this worker.

10 Tang *et al.* (2009) reported cases of contact dermatitis in 4 of 10 operators of chemical 11 melting devices in a phenol-formaldehyde factory and two thirds of the workers on a 12 mushroom farm that were exposed to formaldehyde developed dermatitis on their arms 13 and forearms. Symptoms included red spots, swelling, irritation, pain, and a burning 14 sensation.

#### 15 5.4.2.3 Oral exposure

16 Formaldehyde ingestion is rare because it is a strong irritant and has an unpleasant odor. 17 Only 11 cases of formalin ingestion (usually suicidal or homicidal attempts) have been 18 reported in the English literature since 1950. At least 15 cases have been published in the 19 Japanese literature (Yanagawa et al. 2007), and other cases have been reported in China 20 (Tang et al. 2009). These cases suggest that the fatal oral dose of formaldehyde is 60 to 21 90 mL (Bartone et al. 1968, Yanagawa et al. 2007). In addition to severe corrosive 22 damage to the gastrointestinal tract, other effects may include central nervous system 23 (CNS) depression, myocardial depression, circulatory collapse, multiple organ failure, 24 kidney and liver damage, and metabolic acidosis. The primary late complication for 25 survivors is cicatrical stricture of the stomach which may require a gastrectomy 26 (Yanagawa et al. 2007).

Köppel *et al* (1990) presented case reports of two patients (a 55-year-old female and a
34-year-old male) that died after ingesting an unknown quantity of formaldehyde. Both
patients survived the initial gastrointestinal necrosis and renal failure, but died several
weeks later from respiratory distress and cardiac failure. Autopsy findings in one of the

1 patients included burns of the entire digestive tract, including the colon, with extensive 2 hemorrhagic jejunitis, ileitis, and colitis. Plasma levels of formic acid were elevated in 3 both patients, but no free formaldehyde was detected in blood or plasma. These authors 4 speculated that formaldehyde may exert systemic toxicity in the form of its labile Schiff's 5 base with proteins, but not as free formaldehyde. One patient died 28 hours after 6 ingesting 120 mL of a formaldehyde/methanol solution (Eells et al. 1981). Plasma 7 methanol, formaldehyde, and formate levels were measured in a 50-year-old male who 8 was found unconscious and unresponsive at a meat packing plant after drinking about 4 9 ounces of a formaldehyde solution (Burkhart et al. 1990). The clinical course included an 10 initial CNS depression followed by abdominal pain, retching, seizures, hypotension, and 11 cardiac arrest. The patient died 13 hours after exposure. Methanol levels increased 12 throughout the 13-hour course, while formate and formaldehyde levels increased until 13 bicarbonate and ethanol therapy were instituted after 6 hours. Hilbert et al. (1997) 14 reported a case of fatal poisoning in a 46-year-old woman who deliberately ingested 50 to 15 100 mL of formalin. She was admitted to the intensive care unit 2 hours later and presented with metabolic acidosis, gastric ulceration, and circulatory shock. The patient 16 17 died 44 hours after ingesting the formalin from multiple organ failure, including severe ventricular failure. 18

19 Two cases of nonfatal poisoning were reviewed (Bartone et al. 1968, Yanagawa et al. 20 2007). Bartone et al. (1968) reported that a 46-year-old woman drank an estimated 120 21 mL of a 10% formaldehyde solution and experienced shock and severe abdominal pain, 22 and developed diffuse ulceration, fibrosis, and contracture of most of the stomach. She 23 was admitted to the hospital 3 months after the incident after experiencing frequent 24 episodes of weakness, loss of appetite, weight loss, nausea and vomiting. The lesion 25 culminated in an almost complete, high gastric obstruction and required a total 26 gastrectomy. A 28-year-old man also survived after reportedly ingesting 150 mL of a 27 40% formalin solution in an attempted suicide (Yanagawa *et al.* 2007). This patient was 28 admitted to the hospital 2 hours after ingesting the formalin. Endoscopy on hospital day 4 29 showed esophageal erosion and diffuse corrosive gastric ulcers. By day 6, ascites with 30 multiple spotty hemorrhages on the gastric serosa and omentum had developed. Further 31 complications included bacterial pneumonia, sepsis, enteritis, toxic epidermal necrolysis,

and gastric outlet obstruction. The patient was discharged on day 73. Gastroscopy was
 repeated on day 132 and showed that the stomach surface was covered by a regenerated

mucosa with scattered linear scars. The gastric outlet obstruction had improved by day
148.

5 In two separate incidences in China, 60 and 38 middle-school students reported

6 symptoms of nausea, vomiting, and dizziness 30 minutes to 2 hours after eating fish

7 illegally preserved in formaldehyde [no further information provided] (Tang *et al.* 2009).

8 5.4.2.4 Hematological and immunological effects

9 Intravascular coagulopathy was described in a 58-year-old man who swallowed 4 ounces

10 of formalin (Burkhart *et al.* 1990). This patient died shortly thereafter from cardiac arrest.

11 Kuo et al. (1997) investigated the possible effects of formaldehyde exposure in 50 12 hemodialysis nurses in four teaching hospitals in Taiwan. The control group included 71 13 ward nurses who did not work in the hemodialysis unit. A questionnaire was used to 14 gather information on health history, demographic data, exposure to formaldehyde, and 15 symptoms. Symptoms included itching, dizziness, nausea and vomiting, fatigue, impaired 16 concentration, tearing, nasal discharge, cough, and difficulty breathing and were scored 17 from 0 to 3 corresponding to never, seldom, occasionally, and frequently. The values for 18 the symptoms were totaled to derive a total symptom score. The control group was 19 younger, less likely to be married, and more likely to have allergic rhinitis than the 20 exposure group. There was a significant positive correlation between airborne 21 formaldehyde concentrations and total symptom score. Multiple regression analysis 22 indicated that the exposure group's white blood cell count was significantly lower than 23 the control group.

Most of the studies on the immunologic effects of formaldehyde have focused on the
allergic reactions (i.e., contact dermatitis and occupational asthma); however, several
studies have reported that formaldehyde exposure may affect immunological parameters.
These studies cover acute, subchronic, and chronic exposures and include workers,
medical students, residents, and children.

1 Madison et al. (1991) studied a group of residents who experienced acute symptoms 2 following exposure to formaldehyde and exothermic byproducts of an urea-formaldehyde 3 spill. Three years after the accident, the exposed group was compared with an unexposed 4 group selected from a nearby community. Immunological parameters included white 5 blood cell count, total lymphocyte count, percent and total lymphocyte subsets (CD4, 6 CD5, CD8, CD19, CD25, and CD26 cells), prevalence of autoantibodies, and antibodies 7 to formaldehyde-human serum albumin conjugate. Data were adjusted for age, gender, 8 smoking, mobile home residency, and use of wood stoves. White blood cell, lymphocyte, 9 and T-cell counts were not affected; however, significant differences were reported for 10 elevated percent and absolute numbers of CD26 cells, autoantibodies, and greater titers of 11 isotypes IgG and IgM to formaldehyde-human serum albumin conjugate. The authors 12 concluded that the exposed subjects had an activated immune system in addition to 13 increased autoantibodies.

14 Vargovà et al. (1992) investigated the immunological and cytogenetic effects (see 15 Section 5.6.4.3) of formaldehyde in a group of 20 workers (10 male and 10 female) who 16 had been occupationally exposed for 5 to more than 16 years. They were compared with a 17 matching control group (similar habits and social status) of 19 individuals from the same 18 plant who had no known exposure to formaldehyde. There were no significant 19 differences between the exposed group and controls in values of natural cellular or 20 specific humoral immunity; however, there were differences in the values of mitogen-21 induced proliferation of lymphocytes. The authors concluded that formaldehyde exposure 22 interfered with the immune system, but not enough to show changes in the classical 23 clinical-immunological responses.

Ying *et al.* (1999) examined both genetic and immunological parameters to investigate the effects of formaldehyde exposure on peripheral lymphocytes in 23 non-smoking medical students (11 males and 12 females). The study was conducted during an 8-week anatomy laboratory. Students were exposed three times per week for 3 hours per class. Formaldehyde concentrations were measured in the laboratories and in the students' dormitories. Blood samples were collected from each student at the beginning of the anatomy laboratory and after completing the laboratory. Lymphocyte subsets were 1 stained by mouse antihuman monoclonal antibodies CD3 (total T cells), CD4 (T helper-

- 2 inducer cells), CD8 (T cytotoxic-suppressor), and CD19 (B lymphocytes) surface
- 3 markers within 24 hours after collecting the blood samples. Genetic effects are discussed
- 4 in Section 5.6.4.3. Formaldehyde concentrations ranged from 0.071 to  $1.28 \text{ mg/m}^3$  in the
- 5 laboratories and 0.011 to 0.016  $mg/m^3$  in the dormitories. The time-weighted average
- 6 concentration in the laboratories was  $0.508 \pm 0.299 \text{ mg/m}^3$ . The results observed in the
- 7 study were determined to be similar for both males and females; therefore, the data were
- 8 pooled. The percentage of lymphocyte subsets did show significant changes at the end of
- 9 the study (Table 5-9). There was a significant increase in B cells, and a significant
- 10 decrease in total T cells, T-helper-inducer cells, and T-cytotoxic-suppressor cells. There
- 11 also was a higher ratio of T-helper-inducer cells to T-cytotoxic-suppressor cells.

 Table 5-9. Effects of formaldehyde exposure on peripheral lymphocyte subsets in anatomy students

Subset	Before exposure (%)	After exposure (%)
B cells	$16.87 \pm 1.52$	$23.98 \pm 4.52^{***}$
Total T cells	$72.63\pm2.90$	$65.46 \pm 4.65^{***}$
T-helper-inducer cells (T <sub>4</sub> )	$48.87 \pm 4.20$	$44.68 \pm 4.36 **$
T-cytotoxic-suppressor cells (T <sub>8</sub> )	$29.18\pm3.94$	$20.14 \pm 3.04 ***$
$T_{4}/T_{8}$	$1.71 \pm 0.34$	$2.25 \pm 0.44$ ***

Source: Ying et al. 1999.

\*\* P < 0.01 (t-test); \*\*\* P < 0.001.

12 Lyapina et al. (2004) reported that their previous studies demonstrated that the 13 immunotoxic action of formaldehyde resulted in delayed type skin sensitization and 14 reduced resistance to infections (recurrent rhinitis, upper respiratory tract infections and 15 pneumonitis) in exposed workers and suggested that formaldehyde exposure may result 16 in functional changes in neutrophils. Therefore, they examined the effects of 17 formaldehyde exposure on neutrophil respiratory burst activity in 29 workers exposed to 18 formaldehyde. The exposed group was further divided into 12 individuals (group 1a) with 19 a history of frequent viral or bacterial inflammatory relapses of the upper respiratory tract 20 and clinical observations of hypertrophy or atrophy of the upper respiratory mucous 21 membranes, chronic pharyngitis, rhinitis, rhinosinusitis and rhinopharyngitis. Group 1b 22 included the other 17 exposed workers, 12 of whom had no history or clinical findings of 23 upper respiratory tract infections, and 5 who had a history of rare, short, predominantly

1 acute, inflammatory relapses of viral etiology in the upper respiratory tract. The control 2 group included 21 non-exposed, age- and gender-matched healthy individuals. 3 Formaldehyde concentrations measured in the workplace of the exposed group ranged from 0.64 mg/m<sup>3</sup> to 1.92 mg/m<sup>3</sup> with a mean of  $0.87 \pm 0.39$  mg/m<sup>3</sup>. Although routine 4 5 hematological tests did not show any differences between the exposed and control 6 groups, there was a statistically significant negative correlation between the duration of 7 exposure and erythrocyte count and hematocrit level. Exposed workers had a statistically 8 significant decreased resistance to infection. Neutrophils generate reactive oxygen 9 species (the respiratory burst) in response to tissue damage or local invasion of 10 microorganisms. Although there were no significant differences in the spontaneous or 11 stimulated neutrophil respiratory burst activity between the exposed group and the 12 control group, there was a decrease of spontaneous neutrophil respiratory burst activity in 13 workers with a history and clinical findings of frequent and long-lasting relapses of 14 chronic inflammation of the upper respiratory tract (group 1a). Therefore, functional 15 changes in polymorphonuclear neutrophil granulocytes could serve as an early indicator 16 of an impact of formaldehyde on neutrophil respiratory burst activity. 17 Erdei et al. (2003) examined the relationship between immune biomarkers and indoor air 18 quality in 176 school children aged 9 to 11 years. These children had immunologically 19 related respiratory diseases and lived in Hungarian cities. Nitrogen dioxide, 20 formaldehyde, benzene, xylene, and toluene were measured in indoor air of the homes of 21 these children. Higher indoor formaldehyde concentrations were associated with 22 significantly increased monocyte concentrations and bacterial-specific IgGs.

Ye *et al.* (2005) examined two populations of formaldehyde-exposed workers in China. One group of 18 workers was exposed in a formaldehyde manufacturing facility while a second study group included 16 waiters who were exposed to low levels of formaldehyde while working in a newly fitted ballroom for 12 weeks. The control group included 23 college students. All study participants were nonsmokers. There was a significantly increased percentage of B cells accompanied by significantly decreased percentages of total T cells (CD3) and T-cytotoxic-suppressor cells (CD8) in the manufacturing workers compared with the student controls. T-suppressor (CD4) cells were unchanged. These
 authors also investigated clastogenic effects in these workers (see Section 5.6.4.3).

3 Veraldi et al. (2006) evaluated the immunotoxic effects of 20 chemicals (including 4 formaldehyde) that are widely used in the work environment. The primary purpose of this 5 study was to document the evidence and to construct a matrix that can be used to estimate 6 the relative risk of the chemicals. This evaluation consisted of three primary steps: (1) 7 conduct a systematic literature search and review the data on immunotoxicity testing and 8 testing schemes, (2) document the evidence (type of immunotoxicity, strength of 9 evidence, and power) in summary tables for each chemical, and (3) assign an index 10 (strong, intermediate, weak, or nil) based on the evidence of toxicity and the type of 11 effect (immunosuppression, autoimmunity, hypersensitivity). The evaluation included 12 both human and experimental animal studies. Based on the overall evidence, these 13 authors placed formaldehyde in the "weak" category. The main immunotoxic effect of 14 formaldehyde was hypersensitivity.

15 Sasaki et al. (2009) obtained peripheral blood mononuclear cells from nonatopic healthy

16 donors. T cells were isolated and stimulated with anti-CD3/anti-CD28 monoclonal

17 antibodies. Pretreatment with formaldehyde selectively suppressed interferon- $\gamma$  and

18 interleukin-10 mRNA expression and protein production in stimulated T cells.

19 Formaldehyde also suppressed nuclear factor kappa B (NF-κB) signaling and activated

20 mitogen-activated protein kinases (MAPKs). The authors reported that formaldehyde had

both transcriptional and nontranscriptional effects on T cell signaling that promoted a T

22 helper type 2-skewed immune response.

Tang *et al.* (2009) summarized eight reports of formaldehyde-induced hematotoxicity from Chinese studies (Table 5-10). In general, these studies showed a significant decrease in total white blood cell counts [leucopenia] in exposed workers when compared with controls. Two studies had evidence of pancytopenia [reduced white blood cells, platelets, and red blood cells]. They also presented a case report of pancytopenia in a previously apparently healthy woman after she lived 3 months in a newly remodeled apartment [data

- 1 not reported]. Formaldehyde air concentrations were 4-fold above the indoor exposure
- 2 standard, whereas benzene and toluene were within indoor concentration limits.

Subject <sup>a</sup>		Concentration					Deference (ee eited
Group	N	(mg/m <sup>3</sup> )	WBC (× 10 <sup>9</sup> /L)	Plt (× 10 <sup>9</sup> /L)	Hb (g/L)b	Notes	in Tang <i>et al.</i> 2009)
Exposed Control	65 70	N/A	$5.42 \pm 2.04^{***}$ $6.61 \pm 1.66$	$\begin{array}{c} 172.48 \pm 87.57^{***} \\ 243.10 \pm 84.08 \end{array}$	$\begin{array}{c} 125.66 \pm 21.83 \\ 128.59 \pm 13.11 \end{array}$	WBC and Plt counts decreased with increasing work years	Tong et al. 2007
Exposed Control1	239 200	0.022–0.044	33/239 (14%)** <sup>b</sup> 8/200 (4%)	26/239 (11%)** <sup>b</sup> 2/200 (1%)	77/239 (32%)** 43/200 (21.5%)	All counts decreased with increasing work years	Yang 2007a
Exposed Control	72 150	0.24–0.93	10/72 (14%)* <sup>b</sup> 8/150 (5%)	N/A	N/A		Cheng et al. 2004
Exposed Control	110 120	N/A	$4.91 \pm 1.17$ $5.92 \pm 1.51$	N/A	N/A	WBC count decreased with increasing work years	Tang and Zhang 2003
Exposed Control	50 71	0.184	NR	NR	NR	Significant correlation of decreased WBC count with increased [FA]	Kuo et al. 1997
Exposed Control	55 41	N/A	5.39*** 6.22	N/A	N/A	Reported increase in IgM, IgA, and eosinophil counts	Qian <i>et al.</i> 1988
Exposed Control	10 10	0.44–6.84	$5.74 \pm 1.35$ $6.48 \pm 2.15$	$122.46 \pm 32.87 \\ 118.84 \pm 22.52$	$119.77 \pm 11$ $120 \pm 10$	WBC counts decreased, but NS	Xu et al. 2007b
Exposed Control	104 68	0.7–19.2	NS	N/A	NS	Original data not provided	Feng et al. 1996

Table 5-10. Summary of blood cell counts in Chinese workers with formaldehyde exposure reported by Tang et al. (2009)

\*P < 0.05, \*\*P < 0.01, \*\*\*P < 0.001.

WBC = white blood cell, Plt = platelet, Hb = hemoglobin, N/A = not available, NR = not reported, NS = not significant, [FA], formaldehyde concentration.

<sup>a</sup> Most exposed subjects are industrial workers, with the exception of pathologists in the Cheng *et al.* 2004 study, and nurses in the Kuo *et al.* 1997 study.

<sup>b</sup> Numbers of subjects with decreased blood cell counts are given. Percentage (%) is calculated from subjects with abnormal counts among total subjects.
### 1 5.4.2.5 Neurophysiological effects

2 Neurobehavioral effects have been reported to be related to exposure to formaldehyde in 3 histology technicians (Kilburn et al. 1985a, Kilburn and Warsaw 1992, Kilburn et al. 4 1987) and fiberglass manufacturing workers (Kilburn 2001, Kilburn et al. 1985a); these 5 effects include lack of concentration and loss of memory, disturbed sleep, impaired 6 balance, variations in mood, alterations of appetite, indigestion, nausea, headache, and 7 fatigue. Many of these studies were reviewed by WHO (2002), and the conclusion of that 8 review was that there was little convincing evidence that formaldehyde is neurotoxic in 9 occupationally exposed populations. Other studies that reported neurobehavioral effects 10 in relation to exposure to formaldehyde include individuals living in homes insulated 11 with urea-formaldehyde foam (Harris et al. 1981, Thun et al. 1982) and in manufactured 12 homes or conventional homes (Kilburn 2000, Loomis 1979, Main and Hogan 1983, 13 Ritchie and Lehnen 1987). Although Ritchie and Lehnen (1987) reported that headaches 14 increased with the formaldehyde concentration in the home in a study of 2,000 residents 15 of nearly 900 mobile and conventional homes in Minnesota, other studies, such as Thun 16 et al. (1982) did not find any significant differences for headache, insomnia, or dizziness.

Kuo *et al.* (1997) (also discussed above under hematological and immunological effects) reported that incidences of dizziness, nausea, difficulty concentrating, tearing, nasal discharge, cough, and difficulty breathing were higher in a group of 50 hemodialysis nurses from four teaching hospitals in Taiwan compared with a control group of 71 ward nurses who did not work in the hemodialysis unit. Dizziness, nausea, fatigue, and difficulty concentrating were associated with formaldehyde exposure, while other symptoms may have been related to sodium perchlorate exposure.

### 24 5.4.2.6 Reproductive effects

Epidemiological studies have investigated the reproductive effects of occupational
exposures to formaldehyde; however, most of the available studies were not designed
specifically for formaldehyde and are confounded by co-exposures to other chemicals
(IARC 2006). The reproductive effects examined in these studies included spontaneous
abortion, congenital malformations, birth weight, infertility, and sperm abnormalities.
IARC reviewed five case control studies and one meta-risk analysis study that included

1 11 studies. Another study, (Saurel-Cubizolles et al. 1994) that was not included in the

- 2 IARC (2006) review investigated pregnancy outcome among operating room nurses. This
- 3 study surveyed 17 hospitals in Paris as part of mandatory annual occupational practitioner
- 4 visit; analyses were adjusted for age, number and outcome of previous pregnancies, and
- 5 tobacco use. Controls were selected from hospital employees that did not work in the
- 6 operating room and were matched by hospital, age, and duration of employment. These
- 7 studies showed inconsistent reports of higher rates of spontaneous abortion, birth defects,
- 8 and low birth weights in women occupationally exposed to formaldehyde. Results are
- 9 summarized in Table 5-11.

Subjects	Endpoint	Results	References
Hospital staff	Spontaneous abortion	No correlation when adjusted for age, parity, decade of pregnancy, tobacco, and alcohol use	Hemminki <i>et al.</i> 1982 (as cited in IARC 2006)
Nurses	Spontaneous abortion Congenital defects	No correlation with spontaneous abortion, OR of 1.74 (95% CI = $0.39-7.7$ ) for malformations based on 8 exposed subjects	Hemminki <i>et al.</i> 1985 (as cited in IARC 2006)
Laboratory workers	Spontaneous abortion Congenital defects Birth weight	OR of 3.5 (95% CI = $1.1-11.2$ ) for spontaneous abortion in women exposed to formalin at least 3 days/wk. No association with congenital malformations.	Taskinen <i>et al.</i> 1994 (as cited in IARC 2006)
Woodworkers	Time to pregnancy Spontaneous abortion	Significant association with delayed conception density and spontaneous abortion.	Taskinen <i>et al.</i> 1999 (as cited in IARC 2006)
Meta-risk analysis	Spontaneous abortion Birth weight	Four studies had higher rates of spontaneous abortion while 5 studies did not. No association with birth weights	Collins <i>et al.</i> 2001b (as cited in IARC 2006)
Autopsy service workers	Sperm abnormality	No significant differences between the exposed and control groups	Ward <i>et al.</i> 1984 (as cited in IARC 2006)
Nurses	Spontaneous abortion Birth defects	Significant increase ( $P < 0.05$ ) in spontaneous abortion and all birth defects combined in operating room nurses. No significant difference for major birth defects.	Saurel-Cubizolles <i>et al.</i> 1994

 Table 5-11. Reproductive effects of formaldehyde in humans

CI = confidence interval; OR = odds ratio.

- 10 Tang et al. (2009) noted two Chinese studies on formaldehyde exposure and menstrual
- 11 disorders. In a food additive factory, 70% of women exposed to formaldehyde through
- 12 inhalation (0.82 to 5.96 mg/m<sup>3</sup>) reported abnormal menstrual cycles, whereas 17%

reported menstrual abnormalities in the control group. In a separate study, anatomy
teachers exposed to over 0.5 mg/m<sup>3</sup> formaldehyde reported menstrual disorders and, in
some cases, dysmenorrhea [data not reported].

### 4 5.4.3 Toxic effects in experimental animals

5 The acute and chronic toxicity of formaldehyde has been extensively studied in 6 experimental animals and recently reviewed by IARC (2006). Acute effects include 7 irritation, pulmonary hyperreactivity, and cytotoxicity and cell proliferation in the nose 8 and upper respiratory tract. Mice are more sensitive than rats to respiratory depression. 9 The primary chronic effects also include cytotoxicity and cell proliferation in the upper 10 respiratory tract, gastrointestinal irritation and ulceration, and skin sensitization. 11 Developmental toxicity studies have been conducted on pregnant dams and generally 12 have not shown a developmental effect at exposure levels that were not maternally toxic. 13 Other effects reported include oxidative stress, neurotoxicity, immunotoxicity, and 14 decreased thyroid gland, liver and testis weights. Testicular toxicity has been reported in 15 rats, mice, and birds. However, effects on male reproductive performance were not 16 tested.

17 5.4.3.1 Irritation, sensitization, and respiratory effects

18 The irritant effects of formaldehyde in experimental animals range from mild irritation to 19 severe ulceration (IARC 2006). Skin contact sensitization has been reported in mice and 20 guinea-pigs. Formaldehyde is a potent respiratory tract irritant in rodents, causing slow 21 and shallow breathing, and histopathological lesions in the nose and upper respiratory 22 tract. B6C3F<sub>1</sub> mice exposed to 4.9 ppm and F344 rats exposed to 31.7 ppm had a 50% 23 reduction in respiratory rate. Pulmonary hyper-reactivity and bronchoconstriction were 24 reported in guinea-pigs exposed to 0.3 ppm for 8 hours or > 9 ppm for 2 hours. Ingestion 25 of 82 to 109 mg/kg formaldehyde for 2 years caused severe damage to the gastric mucosa 26 in male and female Wistar rats (Til et al. 1989).

27 Both acute and chronic inhalation exposures to formaldehyde can cause cytotoxicity and

cell proliferation in the nasal mucosa and upper respiratory tract of rodents (IARC 2006).

29 These studies generally show that formaldehyde increases cell proliferation and cell

30 turnover, inhibits mucociliary function, and causes histopathological changes in the nasal

1 mucosa in a concentration- and site-specific manner. Histopathological changes include 2 squamous metaplasia, epithelial erosion, epithelial hyperplasia, degeneration of the 3 respiratory and olfactory epithelium, and necrosis. Rats are more susceptible than mice, 4 presumably because mice reduce their minute ventilation more than rats when exposed to 5 high concentrations (Chang et al. 1983, Swenberg et al. 1983a). Furthermore, Swenberg 6 et al. (1983a) and Wilmar et al. (1987) reported that the severity of cytotoxic effects was 7 more dependent upon formaldehyde concentration than the cumulative dose in their 8 studies. Liteplo and Meek (2003) reviewed short-term, subchronic, and chronic studies of 9 the effects of formaldehyde on cell proliferation within the respiratory epithelium of rats 10 and reported that histopathological lesions and a sustained increase in proliferation of 11 nasal epithelial cells were not observed at concentrations of 2 ppm or less. More 12 information on respiratory tract cytotoxicity and cell proliferation is presented in Section 13 5.7.5.2 as it relates to mechanistic considerations for cancer.

14 Hilton et al. (1996) conducted a series of tests to study the sensitizing properties of 15 formaldehyde. These included the guinea-pig maximization test, the occluded patch test, 16 the murine local lymph node assay, and the mouse IgE test. The mouse IgE test was used 17 to determine the potential for sensitization of the respiratory tract. Chemicals known to 18 cause respiratory allergy in humans stimulate a significant increase in serum IgE 19 concentrations, while contact allergens do not. Female BALB/c mice and albino Dunkin-20 Hartley guinea-pigs were used. Formaldehyde elicited strong positive responses in the 21 guinea-pig maximization test, the occluded patch test, and the murine local lymph node 22 assay. The mouse IgE test was negative. The authors concluded that these data indicate 23 that formaldehyde is a potent contact allergen but did not cause sensitization of the 24 respiratory tract.

Lino dos Santos Franco (2006) investigated the mechanisms underlying rat lung injury and airway reactivity changes caused by formaldehyde exposure. Male Wistar rats were exposed to a 1% formaldehyde solution [air concentrations generated from the solution were not reported] for 30, 60, or 90 minutes/day for four days. Methanol (0.32%) was added to the solution to prevent polymerization. Both a non-exposed and a methanolexposed control groups were included. Animals were killed one day after the final

1 exposure. The reactivity of isolated trachea and intrapulmonary bronchi were assessed in 2 dose-response curves to methacholine. Local and systemic inflammatory responses were 3 evaluated by counting leukocytes in bronchoalveolar lavage fluid, blood, bone marrow 4 lavage, and spleen. Tracheal reactivity was not affected by formaldehyde exposure, but 5 there was a significant bronchial hyporesponsiveness in exposed rats. Formaldehyde 6 exposure was associated with a significant increase in the total cell numbers in 7 bronchoalveolar lavage fluid, peripheral blood and spleen, but not in bone marrow. The 8 effect was time-dependent in bronchoalveolar fluid with the maximum response observed 9 after 90 minutes exposure. Leukocytes in the bronchoalveolar fluid were composed 10 mainly of mononuclear cells in rats exposed for 30 or 60 minutes, but both mononuclear 11 cells and neutrophils were observed in rats exposed for 90 minutes. The authors proposed 12 that formaldehyde exposure may affect lung resident cells, including macrophages and 13 mast cells that could mediate the lung inflammatory response and the systemic release of 14 inflammatory mediators. The inflammatory mediators may trigger systemic immune 15 responses and be implicated in the increased number of cells in the spleen.

16 Lino dos Santos Franco et al. (2009) further investigated the lung allergic response in 17 male Wistar rats exposed to formaldehyde vapors produced from a 1% aqueous solution 18 for 90 minutes daily on three consecutive days. The rats were subsequently sensitized 19 with ovalbumin and aluminum hydroxide by i.p. injection. Two weeks later, the rats were 20 challenged with aerosolized ovalbumin. Rats treated with formaldehyde had a lower-21 intensity lung inflammation response (i.e., reduced number of inflammatory cells in 22 bronchoalveolar lavage) compared with rats that were not treated with formaldehyde. 23 Furthermore, the formaldehyde-treated rats had a reduced number of bone marrow cells 24 and blood leukocytes suggesting that the effects were not localized just to the airways. 25 The authors concluded that formaldehyde may impair the lung cell recruitment after an 26 allergic stimuls, thereby leading to a nonresponsive condition against inflammatory 27 stimuli.

#### 28 5.4.3.2 Cytotoxicity

- 29 Wilmer *et al.* (1989) compared the effects of intermittent versus continuous
- 30 formaldehyde exposures in male Wistar rats [age not reported]. Groups of 25 rats were

1 exposed to formaldehyde at a concentration of 0, 1, or 2 ppm for 8 hours or to a 2 concentration of 2 or 4 ppm during eight 30-minute intervals separated by 30-minute 3 non-exposure periods. These concentrations were selected to represent marginally 4 cytotoxic levels as determined from previous studies. Exposures were carried out 5 5 days/week for 13 weeks. For examination of cell proliferation, 5 rats from each group were given a single dose (74 kBq/g) of  $[^{3}\text{H}]$ thymidine 18 hours after the third day of 6 7 exposure and were killed 2 hours later. The cell-proliferation procedure was repeated in 5 8 additional rats from each group after 13 weeks. At the end of the study, the animals were 9 necropsied and examined for gross pathology. Six standard cross sections of the nasal 10 cavity were processed and examined by light microscopy. Body weight did not differ 11 between any exposure group and the controls. Exposure-related effects in the nasal cavity 12 were seen only in the rats exposed to formaldehyde intermittently at 4 ppm. Increased 13 degrees and incidences of disarrangement, hyperplasia, and squamous metaplasia with or 14 without keratinization of the respiratory epithelium were reported. The cell-proliferation 15 study indicated that after 13 weeks, the cell-turnover rate of the nasal respiratory 16 epithelium was three times as high in the 4-ppm group as in the controls. The cell-17 proliferation rates in the other groups were comparable to control values. The authors 18 concluded that the severity of the cytotoxic effects was determined by the exposure 19 concentration rather than total dose (concentration × exposure time).

20 5.4.3.3 Neurotoxicity

21 IARC (2006) reviewed two animal studies by Pitten et al. (2000) and Malek et al. (2003) 22 that reported possible neurobehavioral effects of formaldehyde. Pitten et al. (2000) 23 reported that exposure to formaldehyde by inhalation at either 2.6 or 4.6 ppm 24 significantly increased the time required to find food and the number of mistakes made 25 during the trials, and these effects increased with the length of the exposure period. 26 However, the IARC Working Group concluded that there was no evidence that the 27 changes seen in this study were due to formaldehyde-induced neurotoxicity and 28 suggested that loss of olfactory capacity and visual difficulties with irritant effects to the 29 cornea, changes that would have improved after treatment was stopped, could explain the 30 results. The study by Malek *et al.* reported the effects of exposure to formaldehyde on the 31 performance of male and female Lewis rats in a water maze. The formaldehyde-exposed

rats (0.5 and 5.4 ppm) required significantly longer swimming periods to reach the finish
and made significantly more errors that the control animals. Although the authors
concluded that formaldehyde affected the learning behavior and memory of rats, IARC
noted that complications of blurry vision and loss of olfactory cues were not controlled
for, and the Working Group suggested that the treatment-related response was not due to
a CNS effect.

7 A number of other studies of neurobehavioral effects in rats or mice exposed to

8 formaldehyde have been published. Malek *et al.* (2003) reported that a single exposure to

9 formaldehyde significantly affected the locomotor and explorative behavior of rats, but

10 the effects did not show any linear trends with respect to the formaldehyde concentrations

11 (1, 2.5, or 5 ppm). Malek *et al.* (2004) also exposed male AB mice to 1.1-, 2.3-, or 5.2-

12 ppm formaldehyde vapor for 2 hours, and locomotion and explorative activity in the open

13 field were significantly affected at both 2 and 24 hours after exposure. Usanmaz *et al.* 

14 (2002) reported that low concentrations (1.8 ppm) of formaldehyde increased the

15 excitability of the CNS in male and female BALB/c mice but, as the concentration

16 increased (up to 14.8 ppm), a general depressant effect on the CNS became more

17 pronounced.

18 Cellular and biochemical changes in the brains of rats and mice have also been proposed 19 to be related to exposure to formaldehyde. These studies involved measurements of cell 20 number or protein expression in the hippocampus, a region of the brain related to 21 memory and learning. Songur et al. (2003) reported increases in heat shock protein 70 22 kDa (Hsp70)-positive neurons in the hippocampus of formaldehyde-exposed Wistar rats 23 (0-, 6-, or 12-ppm formaldehyde). The number of pyknotic neurons also increased in the 24 exposed groups. Gurel et al. (2005) reported that male Wistar rats that received i.p. 25 injections of formaldehyde for 10 days had degenerated neurons with pyknotic nuclei and 26 fewer neurons in the frontal cortex and hippocampus compared with controls. Aslan et al. 27 (2006) and Sarsilmaz et al. (2007) reported that male Wistar rats exposed neontally to 0-, 28 6-, or 12-ppm formaldehyde for 30 days had significantly increased numbers of granule 29 cells in the hippocampal formation in both low- and high-dose groups (Aslan et al.) and

significantly fewer pyramidal cells in the hippocampus in the high-dose group (Sarsilmaz
 *et al.*).

3 Other reports of changes in the hippocampus were published in a series of studies of 4 formaldehyde exposure to ovalbumin-immunized mice by Fujimaki et al. (2004), 5 Tsukuhara et al. (2006), and Ahmed et al. (2007). Exposure to 400-ppb formaldehyde 6 significantly increased brain nerve growth factor (NGF) levels and NGF mRNA in 7 immunized mice (Fujimaki et al.). Exposure to 400-ppb formaldehyde in immunized 8 mice also significantly increased the ratio of Bcl-2 to Bax protein, which the authors 9 concluded would exert a protective effect against cell death by apoptosis (Tsukuhara et 10 al.). In the third paper, Ahmed et al. reported that formaldehyde exposure upregulated 11 expression of hippocampal genes (NR2A, D1 and D2 receptors, and CREB-1) known to 12 play an essential role in the hippocampal synaptic plasticity underlying learning and 13 memory in immunologically sensitized mice.

Lu *et al.* (2008b) reported that inhaled formaldehyde negatively affected learning and memory in Kun Ming mice (an outbred stock of Swiss albino mice). Mice exposed 6 hours/day to 3 mg/m<sup>3</sup> formaldehyde for 1 week had decreased water maze performance and lower dismutase superoxide activity and glutathione levels compared with a control group. Malondialdehyde content and NR1 and NR2B expression increased. Mice exposed to 1 mg/m<sup>3</sup> formaldehyde were not affected. Oxidative stress-induced neuron damage to the brain was identified as a possible mechanism.

21 5.4.3.4 Immunologic and other effects

22 IARC (2006) reviewed several studies that investigated immunologic effects of 23 formaldehyde in mice and rats. B6C3F<sub>1</sub> mice exposed to 15-ppm formaldehyde 6 24 hours/day, 5 days/week for 3 weeks did not have any significant changes in immune 25 function except for an increase in host resistance to Listeria monocytogenes infection. In 26 other studies in mice, formaldehyde exposure did not alter the number or impair the 27 function of resident peritoneal macrophages. BALB/c mice exposed to 2 mg/m<sup>3</sup> [1.6 28 ppm] 6 hours/day for 10 days had enhanced anti-ovalbumin IgE titer; however, in another 29 study, the IgG1 response of ICR mice to a mite allergen in the respiratory tract was not 30 enhanced after exposure to a 0.5% formaldehyde aerosol. There was no evidence that

1 long-term exposure to high concentrations (12.6 ppm) of formaldehyde impaired B-cell

2 function.

3 Vargová et al. (1993) evaluated immune function in male Wistar rats administered 4 formaldehyde by gastric lavage 5 days per week for 4 weeks at doses of 0, 20, 40, or 80 5 mg/kg. Other routine parameters, including hematology, clinical chemistry, and body and 6 organ weights also were examined. Immune system parameters evaluated included cell-7 mediated immunity, humoral-mediated immunity, and immunopathology. Lymph node 8 weights were significantly increased in the dosed groups, but the cellularity of lymphoid 9 organs was not affected. The percentage of monocytes was significantly increased, but 10 the percentage of lymphocytes was significantly reduced. There was a dose-dependent 11 decrease in antibody response (IgG + IgM), but there was no significant reduction in the 12 number of antibody-producing (IgM) cells in the spleen. There was a non-significant 13 reduction in microbicidal activity of blood phagocytes (measured by interaction with the 14 yeast *Candida albicans*). Phagocytic activity (measured by adhesion of hydrophilic 15 synthetic microspheric particles to leukocytes) was significantly reduced only at the 40 16 mg/kg dose for polymorphonuclear leukocytes and monocytes combined.

17 Patel et al. (2003) exposed groups of 10 male albino rats to 5, 10, or 15 mg/kg perday for 18 30 days by i.p. injection. A control group was injected with saline for 30 d. Animals were 19 killed on the 31st day. Rats exposed to 10 or 15 mg/kg had a significantly lower thyroid 20 gland weight, follicular regression, decreased triiodothyronine  $(T_3)$  and thyroxine  $(T_4)$ , 21 and enhanced thyroid stimulating hormone (TSH). Rats in the low-dose group had 22 significantly decreased  $T_3$  and enhanced TSH. Histological examination showed 23 follicular degeneration in the mid-dose group and follicular atrophy in the high-dose 24 group.

Long-term exposure to formaldehyde vapor induced differential immunogenic and
neurogenic inflammatory responses in female C3H/He mice (Fujimaki *et al.* 2004). Mice
were exposed to 0, 80, 400, or 2,000 ppb 16 hours/day, 5 days/week for 12 weeks. Some
mice were given i.p. injections of ovalbumin (OVA) before exposure to formaldehyde.
These mice also were exposed to aerosolized OVA on weeks 3, 6, 9, and 11 for 6 minutes

1 as a booster. Mice were killed the day after the final formaldehyde exposure. There were 2 no significant increases in various types of inflammatory cells in bronchoalveolar lavage 3 fluid in non-immunized mice, but in the high-dose OVA-immunized group, there was a 4 significant increase in the number of bronchoalveolar cells, macrophages, and 5 eosinophils. There was no histological evidence that formaldehyde caused impairment of 6 the epithelial cells in the lung of any of the exposed groups. Formaldehyde-exposed 7 immunized mice had significantly lower production of IL-1ß compared to controls, but 8 TNF-α, IL-6, and granulocyte/macrophage colony stimulating factor remained at control 9 levels. Nerve growth factor in non-immunized mice increased in a dose-dependent 10 manner. Spleen cells, stimulated with lipopolysaccharide to induce cell proliferation, 11 produced significantly higher levels of interferon (IFN- $\gamma$ ) in the high-dose nonimmunized 12 group. Immunized mice exposed to 400 or 2,000 ppb had a significant increase in the 13 production of monocyte chemoattractant protein from spleen cells cultured for 24 hours 14 with OVA. Antigen-specific antibody titers in plasma did not show any significant 15 differences in anti-OVA IgE, total IgE, or anti-OVA IgG2a production. Anti-OVA IgG1 16 and anti-OVA IgG3 production were significantly decreased in the 400-ppb exposure 17 group. There was a dose-dependent increase in substance P levels in the plasma of 18 nonimmunized mice but not in OVA-immunized mice. The authors noted that if the 19 decreased nerve growth factor is related to modulation of sensory neurons and immune 20 abnormalities, these associations may provide an explanation for the multi-organ 21 symptoms in patients with chemical sensitivities.

22 Beall and Ulsamer (1984) reviewed the hepatotoxic effects of formaldehyde. They 23 reported that formaldehyde appeared to be associated with hepatotoxicity in mice, rats, 24 hamsters, guinea-pigs, rabbits, dogs, and humans following injection, ingestion, or 25 inhalation. Effects included alterations in weight, centrilobular vacuolization, focal 26 cellular necrosis, and increased alkaline phosphatase concentrations. The hepatic changes 27 were generally not extensive, and were reversible following acute exposure, but the 28 authors believed that the effects could become progressively more serious with repeated 29 exposures. Quantification of dose-response relationships was not possible because the 30 chemical purity, exposure concentrations, and measurement methods were not always

reported. Possible mechanisms, depending on the route of exposure) suggested by the
authors included direct effects on hepatocytes, indirect effects through the circulatory and
immune systems, and possible additive effects with hepatotoxic chemicals due to
glutathione depletion. Some of the effects were probably caused by secondary
mechanisms such as passive hepatic congestion, serum pH fluctuations, or tissue damage
at other sites.

7 Woutersen et al. (1987) conducted a 13-week inhalation toxicity study in rats exposed to 8 formaldehyde at 0, 1, 10, or 20 ppm for 6 hours/day, 5 days/weeks. At the high dose, 9 uncoordinated locomotion and excitation was observed during the first 30 minutes of 10 each exposure. Other effects included yellowing of the fur, growth retardation, decreased 11 plasma protein levels, and squamous metaplasia of the nasal epithelium, and increased 12 activities of plasma aspartate amino transferase, alanine amino transferase, and alkaline 13 phosphatase (males only). At 100 ppm, the only effects were yellowing of the fur and 14 squamous metaplasia of the nasal epithelium. There was no histopathological evidence of 15 hepatotoxicity in any treatment group.

16 Golalipour *et al.* (2008) reported that exposure to formaldehyde vapor caused

17 morphometric changes in the spleen of albino Wistar rats. A total of 28 rats were divided

18 into 4 groups, including a control group which received no formaldehyde exposure. The

19 treatment groups were exposed to 1.5-ppm formaldehyde for 2 hours/day on 2

20 days/week; 2 hours/day on 4 days/week, or 4 hours/day on 4 days/week for 18 weeks.

21 The germinal center diameter, germinal center area, and marginal zone diameter were

22 increased by formaldehyde exposure, while the mantle layer diameter was decreased.

23 5.4.3.5 Reproductive and developmental effects

24 The reproductive and developmental toxicity of formaldehyde by various routes of

25 exposure has been investigated in rats, mice, hamsters, rabbits, and dogs (IARC 2006).

26 Reported effects included prolongation of pregnancy, changes in fetal organ weights, and

27 various clinical and biochemical changes in the spleen, liver, kidney, thymus, and

28 lymphocytes. There was no evidence of embryolethal or developmental effects when

29 pregnant Sprague-Dawley rats were exposed to 0, 5, 10, 20, or 40 ppm for 6 hours/day

30 from gestational day 6 to 20. IARC (2006) noted that 20 ppm would be considered a

1 toxic dose. Another study in Sprague-Dawley rats reported reduced ossification in 2 offspring at 5 and 10 ppm, but none of the reproductive parameters were affected. At 10 3 ppm, there was a significant decrease in food consumption and weight gain. 4 Formaldehyde was applied dermally to the shaved backs of anesthetized pregnant Syrian 5 hamsters for a 2 hours period on days 8 to 11 of gestation. The incidence of resorptions 6 increased, but no malformations were reported. The authors noted that the increased 7 resorptions might have been caused by the stress of anesthesia. Female Wistar rats exposed to 0.5 or 1.5 mg/m<sup>3</sup>, 4 hours/day for up to 4 months were mated with untreated 8 9 males. There was a significant increase in the number of degenerating embryos 10 (attributed to structural impairment in blastomeres) in the high-dose group. 11 Thrasher and Kilburn (2001) reviewed the embryo toxicity and developmental toxicity of 12 formaldehyde. Depending upon the exposure period of the dam, the available studies

13 resulted in increased embryo mortality, increased fetal anomalies, decreased

14 concentrations of ascorbic acid, and abnormalities in lysosomal, mitochondrial and

15 endoplasmic reticulular enzymes. Rats exposed before mating had increased embryo

16 mortality while those exposed during mating had increased fetal anomalies. They also

17 reported that <sup>14</sup>C-labelled formaldehyde (tail-vein injection) crosses the placenta and that

18 concentrations in fetal brain and liver were higher than in maternal tissues. Using a

19 similar protocol, Katakura et al. (1993) also studied the distribution of radioactivity from

20 <sup>14</sup>C-labelled formaldehyde in pregnant ICR mice. They reported formaldehyde or its

21 metabolites are rapidly transported to the fetus and that elimination of radioactivity is

slower in fetal tissues than in maternal tissues, especially in the fetal brain and liver.

23 5.4.3.6 Testicular toxicity

Ten studies (seven in rats, one in mice, and two in birds) were located that investigated
the effect of formaldehyde exposure on the testis and are briefly discussed below. After
formaldehyde exposure, decreased testis weights, decreased seminiferous tubule
diameters, and abnormal spermatogenesis and sperm morphologies were reported.

Exposure to formaldehyde vapor caused morphometric changes in the seminiferous
epithelium of Wistar rats (Golalipour *et al.* 2007). A total of 28 rats were divided into 4

30 groups. The treatment groups were exposed to 1.5 ppm formaldehyde for 2 hours/day on

- 1 2 days/week (E3); 2 hours/day on 4 days/week (E2), or 4 hours/day on 4 days/week (E1)
- 2 for 18 weeks. The mean seminiferous tubular diameter and seminiferous epithelial height
- 3 showed a significant decrease with increasing duration of exposure (Table 5-12). The
- 4 authors also reported a decrease in germ cells in E1 and E2 exposure groups, disruption
- 5 of the association between Sertoli cells and germinal cells in the E3 exposure group, and
- 6 arrested spermatogenesis in the E1 exposure group [no quantitative data provided].

 Table 5-12. Seminiferous tubular diameter and height in Wistar rats

		Treatment group					
Effect	Control, mean ± SD	E1 <sup>ª</sup> , mean ± SD	E2 <sup>b</sup> , mean ± SD	E3 <sup>°</sup> , mean ± SD			
Seminiferous tubular diameter (µm)	$252.12 \pm 4.82$	204.55 ± 3.29*	232.45 ± 2.42*	238.94 ± 4.37*			
Seminiferous epithelial height (µm)	82.77 ± 2.00	65.26 ± 1.43*	69.46 ± 1.78*	72.80 ± 2.03*			

Source: Golalipour et al. 2007. \* *P*<0.05 (compared with controls)

<sup>a</sup> Exposed 4 h/d, 4 d/wk.

<sup>b</sup>Exposed 2 h/d, 4 d/wk. <sup>c</sup> Exposed 2 h/d, 2 d/wk.

7 Özen et al. (2005) also reported decreases in seminiferous tubule diameter and serum

8 testosterone levels and a concomitant increase in immunochemical staining for Hsp 70 in

9 Wistar rats with increasing inhalation exposure to formaldehyde over a 13-week period

(Table 5-13). 10

Treatment (ppm)	Tubule diameter, mean ± SEM (μm) N = 100	Serum testosterone, mean ± SEM (ng/dL) N = 6
Control	$259.22 \pm 16.18$	$406.54 \pm 16.82$
5	236.17 ± 13.09***	244.01 ± 23.86***
10	233.24 ± 10.13***	$141.30 \pm 8.56 ***$

Table 5-13. Mean seminiferous tubular diameters and testosterone serum levelsafter 13-week exposure to formaldehyde by inhalation in rats

Source: Özen *et al.* 2005. \*\*\* *P* < 0.001.

1 In a separate study, Özen *et al.* (2002) measured trace element concentrations in the testis 2 after subacute (4-weeks) and subchronic (13-weeks) formaldehyde exposures for 8 3 hours/day and 5 days/week. Both copper and zinc tissue concentrations decreased (P <4 0.001) with subacute and subchronic exposure; however, iron levels increased with both 5 exposure durations. The authors noted that decrease in zinc and copper concentrations 6 might affect the functions of some antioxidant metalloenzymes that require these 7 cofactors, such as superoxide dismutase. 8 Ozen *et al.* (2008) investigated the effect of formaldehyde exposure on antioxidant

9 enzymes in the testis. Adult Wistar rats (7 per group) were injected with formaldehyde 10 (10 mg/kg b.w., i.p. every other day for one month). Glutathione peroxidase, superoxide 11 dismutase and malondialdehyde testicular enzyme levels were determined; the levels of 12 superoxide dismutase and glutathione peroxidase decreased significantly (P < 0.001) with 13 formaldehyde exposure, whereas, the level of malondialdehyde increased significantly (P14 < 0.001) compared to control values. Co-treatment with melatonin (25 mg/kg-bw, i.p.)

15 inhibited these effects.

16 A significant dose-related increase in rat sperm-head abnormalities 3 weeks after i.p.

17 injection of formaldehyde for five days (0.125, 0.250, and 0.50 mg/kg b.w. per day) was

18 reported by Odeigah (1997). There was a lower frequency of fertile matings within the

19 first two weeks after treatment, but not after 3 weeks. [IARC (2006) questioned the

20 biological significance of these findings because of the reactivity of formaldehyde and

21 the parenteral route of exposure.]

- 1 Majumder and Kumar (1995) treated adult male Wistar rats with i.p. injections of
- 2 formaldehyde (10 mg/kg b.w. per day) for 30 days. Animals were sacrificed on the 31st
- 3 day and testis, prostate, seminal vesicles, and epididymis were removed. Significant
- 4 decreases were noted in sperm counts, viability, and motility in the treated group (Table
- 5 5-14). Protein and DNA content were measured in these tissues. Significant decreases in
- 6 DNA content of the testis ( $9.8 \pm 1.01$  vs.  $4.6 \pm 0.37$  µg/mg tissue, P < 0.001) and prostate
- 7 (6.1 ± 1.39 vs. 1.2 ± 0.49 µg/mg tissue, P < 0.001) were reported for the treated group.

Table 5-14. In vivo effect of formaldehyde on spermatozoa

Parameters	Control, mean ± SEM (N = 10)	Treated, mean ± SEM (N = 8)
Sperm count (10 <sup>6</sup> /mL)	$46.30\pm5.01$	20.40 ± 2.01***
Sperm viability (%)	$87.10\pm0.83$	72.60 ± 2.32***
Sperm motility (%)	$75.00 \pm 10.90$	$22.00 \pm 6.40^{***}$

Majumder and Kumar 1995.

\*\*\* P < 0.001 (compared with controls).

8 Chowdhury *et al* (1992) treated Charles Foster rats with formaldehyde at i.p. doses of 5,

9 10, and 15 mg/kg body weight over 30 days. A significant decrease in testicular 3- $\beta$ ,- $\Delta^5$ -

10 hydroxy steroid dehydrogenase (determined by histochemical reaction intensity) and

11 serum testosterone (420, 200, 195, 150 ng/dL for control and increasing dose groups,

12 respectively, P < 0.01) was reported for formaldehyde-exposed groups. Leydig cell

13 nuclear diameter and cell number/cm<sup>2</sup> decreased.

14 Ward *et al.* (1984) investigated the effect of oral administration of 100 mg/kg formalin

15 solution (37% formaldehyde, 10% methanol in water) by giving 5 daily doses to B6C3F1

16 mice. Animals were sacrificed 5 weeks after treatment and sperm morphology analyzed.

17 A non-significant increase in the percentage of abnormal sperm was reported for the

18 formalin-exposed group as compared with the water-exposed control group  $(1.49 \pm 0.90$ 

19 vs.  $1.12 \pm 0.39$  %).

- 20 Two studies in birds examined testicular pathology after oral administration of
- 21 formaldehyde. Japanese quail (Anwar et al. 2001) were fed formalin-containing feed (20,
- 22 10, 5, 2.5, and 0 mL/kg feed) for 8 weeks; relative testis weights and seminiferous tubule
- 23 diameters were decreased significantly at the three highest doses ( $P \le 0.05$ ). In a separate

1 study (Khan et al. 2003), formalin was either mixed in feed (2.5, 5, 10 mL of 37% w/w

2 formalin/kg feed) or a 3% solution administered into the crops of White Leghorn

- 3 cockerels (5, 10, 15, 20 mL/d). All of the groups given formalin had significantly smaller
- 4 diameter seminiferous tubules than the control birds ( $P \le 0.05$ ). Further, testes absolute

5 and relative mass and volumes were significantly decreased in the groups administered

6 3% formalin in the crop at 15 and 20 mL/day ( $P \le 0.05$ )

## 7 **5.5** Carcinogenicity studies of metabolites and analogues

8 Formic acid has not been evaluated for carcinogenicity. Acetaldehyde and glutaraldehyde 9 are analogues of formaldehyde that have been tested for carcinogenicity by the NTP, as 10 has the aromatic aldehyde benzaldehyde (see Section 1 for structures of the formaldehyde 11 analogues). Other simple aldehydes, propionaldehyde, butyraldehyde, and n-pentanal, 12 have not been tested in 2-year bioassays by the NTP, but no information on other chronic 13 assays were identified.

14 Acetaldehyde is currently listed in NTP's Report on Carcinogens as *reasonably* 

15 anticipated to be a human carcinogen. Rats exposed by inhalation to acetaldehyde 16 developed respiratory tract tumors (primarily adenocarcinoma and squamous-cell 17 carcinoma of the nasal mucosa), while hamsters developed laryngeal carcinoma (IARC 18 1999). IARC also noted that human data are limited but indicate a possible increase in 19 oral, esophageal, pharyngeal, and laryngeal cancers in people who have a genetic 20 polymorphism leading to higher internal levels of acetaldehyde following heavy alcohol 21 intake. In addition, there have been case reports of bronchial and oral cavity tumors 22 among chemical workers exposed to various aldehydes. Glutaraldehyde was tested for 23 carcinogenicity in F344 rats and B6C3F<sub>1</sub> mice (NTP 1999). Rats were exposed to 0, 250, 24 500, or 750 ppb, and mice were exposed to 0, 62.5, 125, or 250 ppb glutaraldehyde vapor 25 6 hours/day, 5 days/week for 104 weeks. The NTP concluded that there was no evidence 26 of carcinogenic activity of glutaraldehyde in either rats or mice. Hester et al. (2005) 27 concluded that glutaraldeyde's lack of carcinogenicity may be due to a combination of its 28 greater toxicity from lack of DNA-repair, greater mitochondrial damage, and increased 29 apoptosis compared with formaldehyde (see Section 5.6.5). Benzaldehyde in corn oil was 30 administered by gavage 5 days/week to F344 male and female rats at 0, 200, or 400

1 mg/kg b.w. for 103 weeks, to male  $B6C3F_1$  mice at 0, 200, 400 mg/kg b.w. for 104

2 weeks, and to female  $B6C3F_1$  mice at 0, 300 or 600 mg/kg b.w. for 103 weeks (NTP

3 1990). The NTP concluded that there was no evidence of carcinogenic activity of

4 benzaldehyde for male and female rats and some evidence of carcinogenic activity for

5 male and female mice as indicated by increased incidences of squamous-cell papillomas

6 and hyperplasia of the forestomach.

# 7 **5.6 Genetic and related effects**

8 The genetic toxicology of formaldehyde has been investigated in a variety of *in vitro* and 9 *in vivo* assays and has been reviewed (ATSDR 1999, Conaway *et al.* 1996, IARC 1995, 10 2006, Liteplo and Meek 2003, WHO 1989). This section summarizes the genetic effects 11 in prokaryotes, non-mammalian eukaryotes, *in vitro* studies with mammalian and human 12 cells, and *in vivo* studies in experimental animals. The genetic effects of formaldehyde in 13 exposed humans are described in more detail in Section 5.6.4.

# 14 5.6.1 Prokaryotes

15 The studies summarized in this section include those reviewed by Conaway et al. (1996)

16 and IARC (2006) (Table 5-15). Only one additional study published after IARC (2006)

17 was identified (see discussion below).

18 All of the studies with *Salmonella typhimurium* strains TA102 and TA104 were positive 19 for base-pair mutations in the presence or absence of metabolic activation. Most (67%) of 20 the studies with TA100 were positive and all studies with TA1535 were negative. Results 21 were mixed for frameshift mutations with S. typhimurium strains TA97, TA98, TA1537, 22 and TA1538. One study with TA97 was positive without metabolic activation. Only two 23 of seven studies with TA98 were positive without metabolic activation, but three studies 24 with this strain were weakly positive with metabolic activation. All studies with TA1537 25 or TA1538 were negative, with or without metabolic activation. Ma and Harris (1988) 26 reported that about 75% of the reverse mutation studies in S. typhimurium strains were 27 positive. These authors noted that, in general, the mutation efficiency was higher in 28 studies that used the preincubation protocol (a test tube containing a suspension of the 29 tester strain plus S9 mix or plain buffer without S9 is incubated for 20 minutes with the

1 test chemical before adding agar and pouring into Petri dishes containing bacterial culture

2 medium) compared with studies that used the plate incorporation protocol (no

3 preincubation step prior to plating in Petri dishes).

4 Studies with *Escherichia coli* were positive for forward or reverse mutations without

5 metabolic activation (Table 5-15) (Conaway *et al.* 1996, IARC 2006). The mutational

6 spectrum in *E. coli* varied with concentration (Liteplo and Meek 2003). At 4 mmol/L,

7 formaldehyde induced 41% large insertions, 18% large deletions, and 41% point

8 mutations. Most of the point mutations were transversions at GC base pairs. However, at

9 40 mmol/L, point mutations (primarily transitions at a single AT base pair) accounted for

10 92% of the genetic alterations. In addition, formaldehyde caused differential toxicity,

11 DNA strand breaks, DNA-protein crosslinks, and related DNA damage in E. coli (Table

12 5-15).

13 A study by Wang et al. (2007) reported that formaldehyde treatment of E. coli resulted in

14 a dose-dependent microsatellite instability. Their results showed that with 2.5 mM

15 formaldehyde treatment, the complementary dinucleotide repeat microsatellites (GpT)<sub>n</sub>

16 and  $(ApC)_n$  were induced at different frequencies (13 to 24-fold vs. 2 to 3-fold higher

17 than controls, respectively). The authors postulated that this could be due to the

18 unprotected syn position of the guanosine nucleotides in the DNA; this may specifically

19 involve the formation of a Z-DNA structure, which is a conformation that is more

20 difficult for DNA repair enzymes to repair. They further hypothesized that the mutagenic

21 mechanism of formaldehyde and the formation of Z-DNA might account for the observed

22 microsatellite instability.

		Results <sup>a</sup>			
Test system	Without S9	With S9			
S. typhimurium	Forward mutation	+(1/1)	+ (1/1)		
[strains not reported]	Reverse mutation	- (0/1)	- (0/1)		
S. typhimurium					
TA100		(+) (8/12)	± (6/9)		
TA102	Powerse mutation (base noir)	+(5/5)	+(1/1)		
TA104	Reverse mutation (base-pair)	+ (3/3)	+(1/1)		
TA1535		- (0/5)	- (0/5)		
TA7005		+ (1/1)	NT		
S. typhimurium					
TA97		+(1/1)	NT		
TA98	Reverse mutation (frameshift)	- (2/7)	± (3/6)		
TA1537		- (0/5)	- (0/5)		
TA1538		- (0/4)	- (0/3)		
E. coli	Forward mutation	+ (3/3)	NT		
	Reverse mutation	+ (13/13)	NT		
	Strand breaks, crosslinks, related damage	+(2/2)	NT		
	Differential toxicity	+ (2/2)	NT		
E. coli	Instability of induced microsatellites	+ (1/1)	NT		

Table 5-15. Genetic effects of formaldehyde in bacteria

Source: Conaway *et al.* 1996, IARC 2006, Wang *et al.* 2007. + = positive studies, - = negative studies, (+) = mostly positive, (-) = mostly negative,  $\pm$  = at least 2 positive and 2 negative studies; NT = not tested.

<sup>a</sup> Number of positive studies/total number of studies reviewed shown in parentheses.

### 1 5.6.2 Non-mammalian eukaryotes

2 I official de maaded matalons, D1 (11 damage, strand ereans, eressinnes, and other	2	Formaldehy	yde induced	mutations,	DNA	damage,	strand	breaks,	crosslinks,	and other
--	---	------------	-------------	------------	-----	---------	--------	---------	-------------	-----------

3 genetic effects (Table 5-16) in all studies in yeast, fungi, plants, insects, and nematodes

- 4 (IARC 2006). A micronucleus study in newt larvae was negative. All of these studies
- 5 were conducted in the absence of metabolic activation. Several of these studies compared
- 6 effects in wild type and DNA repair-deficient organisms. For example, Magaña-
- 7 Schwencke et al. (1978) reported that Saccharomyces cerevisiae strains that were
- 8 deficient in excision repair were more susceptible to the lethal effects of formaldehyde
- 9 and had a reduced capacity to undergo single-strand breaks compared with the wild type.
- 10 The authors concluded that this indicates that single-strand breaks may be a step in the
- 11 repair process for formaldehyde-induced lesions. The mutagenic effects of formaldehyde
- 12 were also different in DNA repair-proficient and repair-deficient strains of *Neurospora*

- 1 crassa (de Serres and Brockman 1999). The mutant frequencies in the repair-deficient
- 2 strain were higher than in the repair-proficient strain.

Test system	Effect	Results <sup>a</sup> (without S9)
Saccharomyces cerevisiae	Gene conversion	+(1/1)
	Strand breaks, crosslinks, related damage	+(2/2)
	Homozygosis	+(1/1)
Neurospora crassa	Forward mutation	+ (4/4)
	Reverse mutation	± (1/3)
Tradescantia pallida	Micronucleus	+ (1/1)
Various plants	Mutation	+ (1/1)
	DNA damage	+ (1/1)
Drosophila melanogaster	Genetic cross-over or recombination	+ (3/3)
	Sex-linked recessive lethal mutations	+(8/8)
	Dominant lethal mutations	+(2/2)
	Heritable translocation	+ (2/2)
	Gene mutation	+ (1/1)
Caenorhabditis elegans	Recessive lethal mutation	+ (1/1)
Pleurodeles waltl (newt larvae)	Micronucleus	- (0/1)

 Table 5-16. Genetic effects of formaldehyde in non-mammalian eukaryotes

Sources: IARC 2006, Conaway et al. 1996.

+ = all studies were positive,  $\pm =$  both positive and negative studies, - = negative study.

<sup>a</sup>Number of positive studies/total number of studies reviewed shown in parentheses.

### 3 5.6.3 Mammalian systems

- 4 Data are reported here for genetic tests in mammalian cells, including human cells, and in
- 5 experimental animals. The reported effects of formaldehyde in mammalian systems
- 6 include DNA adducts, DNA-protein crosslinks, strand breaks, clastogenic effects,
- 7 mutations, unscheduled DNA synthesis, inhibition of DNA repair, and cell
- 8 transformation. Section 5.6.5 discusses effects on gene expression in humans.
- 9 5.6.3.1 DNA adducts, DNA-protein crosslinks, and DNA damage
- 10 Findings from studies that evaluated exposure to formaldehyde and DNA adducts, DNA-
- 11 protein crosslinks and DNA strand breaks are summarized in Tables 5-17 (in vitro
- 12 studies) and 5-18 (*in vivo* studies).
- 13 In vitro studies

1 Formaldehyde has been shown to react with mammalian cell DNA to form 2 hydroxymethyl derivatives. Formaldehyde induced DNA adducts when reacted with 3 deoxyribonucleosides (Cheng et al. 2008), calf thymus DNA (Beland et al. 1984, Von 4 Hippel and Wong 1971), Chinese hamster ovary cells (Beland et al. 1984), human 5 placental DNA (Zhong and Hee 2004a) and human nasal epithelial cells (Speit et al. 6 2008b, Zhong and Que Hee 2004b) (Table 5-17). Cheng et al. (2008) demonstrated that 7 nitrosamines that generate formaldehyde during metabolism also form formaldehyde 8 adducts when reacted with calf thymus DNA and deoxyribonucleosides. Using HPLC 9 and NMR analysis, hydroxymethyl derivatives at the exocyclic amines of 10 deoxyadenosine, deoxycytidine, and deoxyguanosine were identified after formaldehyde 11 exposure of calf thymus DNA, and hydroxymethyl deoxythymidine derivatives were 12 detected after exposure of Chinese hamster ovary cells (Beland et al. 1984). Zhong and Que Hee (2004b) showed that formaldehyde (in solution, but not in air) caused  $N^6$ -dA, 13 N<sup>2</sup>-dG, and N<sup>4</sup>-dC adducts in human epithelial cells. Formaldehyde-treated DNA and 14 15 RNA have also yielded methylene-bridged crosslinks connecting exocyclic amino groups 16 between nucleosides (Chaw et al. 1980).

17 Recently Lu et al. (2009) demonstrated that formaldehyde crosslinks DNA and glutathione to form  $S-[1-(N^2-deoxyguanosinyl)methyl]glutathione. The intermediate in$ 18 19 this reaction, S-hydroxymethylglutathione, is involved in formaldehyde detoxification 20 and is highly reactive. However, the authors noted that the adduct formed is reasonably 21 stable and may be useful in biomarker studies of exogenous formaldehyde exposure. 22 Numerous *in vitro* studies have shown that formaldehyde exposure (concentrations 23 ranging from 0.01 mM to 62.5 mM) causes DNA-protein crosslinks in human cells 24 (EBV-Burkitt's lymphoma cells, fibroblasts, lymphocytes, gastric mucosa cells,

25 lung/bronchial epithelial cells, skin keratinocytes, Jurkat E6-1 cells, HeLa cells, and

26 whole blood) and rodent cells (Chinese hamster ovary cells, Chinese hamster V79 cells,

- 27 mouse hepatocytes, mouse leukemia L1210 cells, rat Yoshida lymphosarcoma cells, rat
- 28 tracheal epithelial cells, and rat hepatocytes) (Table 5-17). Formaldehyde readily reacts
- 29 with hydrogens of amino groups forming stable methylene-bridged crosslinks between
- 30 the amines of proteins and nucleic acids (Conaway et al. 1996). This reaction is specific

1 for single-stranded DNA because hydrogen bonding with the opposite strand in double-

2 stranded DNA hinders the reactivity. DNA-protein crosslinks can lead to other genotoxic

3 effects through subsequent DNA replication errors (Casanova *et al.* 1989, Liteplo and

4 Meek 2003).

5 The reported removal half-times for these lesions in *in vitro* studies ranged from about 2

6 to 4 hours (Conaway et al. 1996, Cosma and Marchok 1988, Grafström et al. 1983,

7 1984). Craft *et al.* (1987) reported complete removal of DNA-protein crosslinks from

8 human lymphoblasts within 24 hours. Liu et al. (2006) reported that DNA-protein

9 crosslinks were significantly repaired in HeLa cells within 18 hours after removal of

10 formaldehyde compared with a group without formaldehyde removal. In addition, single-

11 strand breaks were significantly repaired within 30 minutes and were almost completely

12 repaired within 90 minutes. Schmid and Speit (2007) treated human blood cultures with

13 formaldehyde concentrations of up to 300 µM. DNA-protein crosslinks were significantly

14 increased by concentrations  $\ge 25 \ \mu$ M. Crosslinks induced by 100  $\mu$ M formaldehyde were

15 completely removed within 8 hours; however, at higher concentrations (200 or 300 μM),

16 some crosslinks remained after 24 hours.

17 Formaldehyde exposure (concentrations ranged from 0.001 to 0.8 mM) also caused

18 single-strand breaks in human cells (fibroblasts, lymphocytes, lung/bronchial epithelial

19 cells, and HeLa cells, but not skin keratinocytes) and rodent cells (mouse leukemia

20 L1210, rat Yoshida lymphosarcoma cells, rat tracheal epithelial cells, and rat hepatocytes,

21 but not Chinese hamster V79 cells) (Table 5-17).

Using the alkaline comet assay, Speit *et al.* (2008b) compared the human cell response to formaldehyde in an established cell line (A549 lung cells) with that of primary cultured cells (human nasal epithelial) under various treatment conditions. They reported no fundamental differences in response between these cells, e.g., observing non-significant decreases in tail moment for both cell cultures at 0.1 mM formaldehyde treatment but a significant (1% level for Dunnett test) effect after a 4-hour treatment with 0.2 mM formaldehyde.

1 Ridpath et al. (2007) noted that although DNA-protein crosslinks likely play an important 2 role in the genotoxicity and carcinogenicity of formaldehyde, little is known about which 3 DNA damage-response pathways are involved in repairing formaldehyde damage. In 4 patients with diseases such as Fanconi anemia (FANC; an inherited blood disorder that 5 leads to bone marrow failure), DNA damage cannot be repaired due to the presence of an 6 abnormal gene in the cells that prevents DNA repair. Ridpath et al. investigated the DNA 7 response pathways by measuring the reduction of cell survival in several repair-deficient 8 mutants in two different cell types. Chicken DT40 cells with targeted mutations in 9 various DNA repair genes were used to assess levels of DNA damage response to 10 formaldehyde. DT40 mutants deficient in the BRCA/FANC pathway, homologous 11 recombination, and translesion DNA synthesis were shown to be hypersensitive (i.e., 12 resulted in reduced cell survival) to formaldehyde. Similar results were observed for the 13 human colorectal cancer (RKO) cell line. Specifically, RKO cells deficient in the FANCC 14 and FANCG genes showed a dose-dependent hypersensitivity to formaldehyde. These 15 results suggest that the BRCA/FANC response pathway in mammalian cells is important 16 in the prevention of DNA damage from formaldehyde. In a review by Zhang *et al.* (2009b), the possible roles of formaldehyde, both endogenous

In a review by Zhang *et al.* (2009b), the possible roles of formaldehyde, both endogenous and exogenous, on the etiology of leukemia in FANC patients is discussed. The authors hypothesized that endogenous exposure might induce DNA-protein crosslinks, which could play a critical role in the initiation of bone marrow failure or in increasing tumor succeptibility in FANC patients. They suggest that subsequent exogenous exposure to formaldehyde may then result in genotoxic levels of induced DNA-protein crosslinks; however, this assumes that formaldehyde actually reaches the bone marrow cells, which has not yet been demonstrated.

Test system	Concentration (LEC or HIC)	Effect	Results	References
Deoxyribonucleosides	0.1 mM	Adducts	+	Cheng et al. 2008
Calf thymus DNA	[0.166 mM] 200 mM	Adducts	++	Beland <i>et al.</i> 1984 Von Hippel and Wong 1971
Chinese hamster ovary cells	1 mM	Adducts	+	Beland et al. 1984
Human nasal epithelial cells	0.33 mM	Adducts	+	Zhong and Hee 2004b
_	0.20 mM	DPX	+	Speit et al. 2008b
Chinese hamster ovary cells	0.20 mM	DPX	+	Zhitkovich and Costa 1992
	0.25 mM		+	Olin et al. 1996
	0.125 mM		+	Garcia et al. 2009
Chinese hamster V79 cells	0.12 mM	DPX	+	Swenberg and al. 1983b
	0.01 mM		$+^{a}$	Speit et al. 2007a
	0.125 mM		+	Merk and Speit 1998
	0.2 mM		_ <sup>b</sup>	Speit et al. 2007a
	62.5 mM		+	Merk and Speit 1999
Mouse hepatocytes	0.5 mM	DPX	+	Casanova and Heck 1997
	0.5 mM		+	Casanova et al. 1997
Mouse leukemia L1210 cells	0.125 mM	DPX	+	Ross et al. 1981
	0.2 mM		+	Ross and Shipley 1980
Rat Yoshida lymphosarcoma cells	0.25 mM	DPX	+	O'Connor and Fox 1987
Rat tracheal epithelial cells	0.05 mM	DPX	+	Cosma <i>et al</i> . 1988a
Rat hepatocytes	0.5 mM	DPX	+	Casanova and Heck 1997
Human EBV-Burkitt's lymphoma cells	0.003%	DPX	+	Costa et al. 1997
Human fibroblasts (skin or	0.1 mM	DPX	+	Snyder and Van Houten 1986
bronchus)	0.2 mM		+	Grafström et al. 1984
	0.25 mM		+	Olin <i>et al.</i> 1996
Human lymphocytes	0.05 mM	DPX	+	Craft <i>et al</i> . 1987
	0.05 mM		+	Liu et al. 2006
	0.1 mM		+	Shaham et al. 1996a
	0.1 mM		+	Andersson et al. 2003
Human gastric mucosa cells	1 mM	DPX	+	Blasiak et al. 2000
Human lung/bronchial	0.1 mM	DPX	+	Saladino et al. 1985
epithelial cells	0.2 mM		+	Grafström et al. 1984
	0.2 mM		+	Grafström et al. 1986
	0.2 mM		+	Speit <i>et al.</i> 2008b
	0.4 mM		+	Grafström 1990
	0.8 mM		+	Fornace et al. 1982
Human skin keratinocytes and fibroblasts	0.025 mM	DPX	+	Emri <i>et al.</i> 2004

Table 5-17. *In vitro* studies of DNA adducts, DNA-protein crosslinks and strand breaks in mammalian systems

<b>-</b>	Concentration	Effect.	Develo	D. farmers
Test system	(LEC or HIC)	Effect	Results	References
Human Jurkat E6-1 cells	1 mM	DPX	+	Saito <i>et al.</i> 2005
Hela cells	0.05 mM	DPX	+	Liu et al. 2006
Human whole blood	0.025 mM	DPX	+	Schmid and Speit 2007
Mouse leukemia L1210 cells	0.125 mM	SB	_	Ross et al. 1981
	0.2 mM		+	Ross and Shipley 1980
Rat Yoshida lymphosarcoma cells	0.25 mM	SB	+	O'Connor and Fox 1987
Rat tracheal epithelial cells	0.2 mM	SB	+	Cosma <i>et al.</i> 1988a
Rat hepatocytes	0.75 mM	SB	+	Demkowicz-Dobrzanski and Castonguay 1992
Chinese hamster V79 cells	0.2 mM	SB	-	Speit et al. 2007a
Human fibroblasts (skin or	0.1 mM	SB	+	Grafström et al. 1984
bronchus)	0.1 mM		+	Snyder and Van Houten 1986
Human lymphocytes	0.005 mM	SB	+	Liu et al. 2006
Human lung/bronchial	0.1 mM	SB	+	Saladino et al. 1985
epithelial cells	0.3 mM		+	Grafström et al. 1984
	0.4 mM		+	Grafström 1990
	0.8 mM		+	Fornace et al. 1982
	1 mM		+	Vock et al. 1999
Human skin keratinocytes and fibroblasts	0.1 mM	SB	_	Emri <i>et al.</i> 2004
Hela cells	0.005 mM	SB	+	Liu et al. 2006

+ = positive result for indicated effect, - = negative result for the indicated effect.

LEC = lowest effective concentration, HIC = highest ineffective concentration, DPX = DNA-protein crosslinks, SB = DNA strand breaks (most were single-strand breaks).

<sup>a</sup>Extended electrophoresis time.

<sup>b</sup> Standard conditions.

1 In vivo

2 No *in vivo* studies were identified that evaluated DNA adducts in experimental animals

3 directly exposed to formaldehyde, but one study reported induction of DNA adducts of

4 formaldehyde in rats treated with carcinogenic nitrosamines. Several studies reported

5 DNA-protein crosslinks and strand breaks (Table 5-18) in animals exposed directly to

6 formaldehyde. Inhalation exposure to formaldehyde caused DNA-protein cross links (0.3

7 ppm to 6 ppm) in rodents (nasal mucosa but not bone marrow) and rhesus monkeys

8 (nasal turbinates, nasopharynx, trachea, and bronchi), and strand breaks (5 ppm) in rats

9 (lymphocytes and liver). Instillation of formaldehyde into rat tracheal implants also

10 caused DNA-protein crosslinks. Transplacental exposure to formaldehyde caused both

DNA protein crosslinks and single-strand breaks in the rat fetal liver. These findings are
 discussed in greater detail below.

Wang *et al.* (2007) demonstrated that formaldehyde-based DNA adducts were formed in the lung and liver of rats treated s.c. with two *N*-nitrosomethyl carcinogens, which both metabolize to formaldehyde. The authors provide qualitative and quantitative [statistical significance not given] evidence for *in vivo* formaldehyde DNA adduct formation for both compounds and suggest that the formaldehyde released by the metabolism of the carcinogens contributes to adduct formation and may, therefore, play a role in the carcinogenic process.

10 Crosslink formation is an important indicator of tissue and DNA exposure; however, the 11 shape of the concentration-response curve is highly non-linear, showing a sharp increase 12 in the nasal epithelium of rats at concentrations greater than 2 ppm, and without 13 accumulation on repeated exposure (Casanova-Schmitz et al. 1984a, Casanova et al. 14 1989, Casanova et al. 1994). Casanova-Schmitz et al. (1984a) exposed male F344 rats for 15 6 hours to formaldehyde concentrations of 0.3, 2, 6, 10, or 15 ppm. Covalent binding of 16 formaldehyde to respiratory mucosal DNA occurred at concentrations  $\geq 2$  ppm; however, 17 the concentration bound to DNA at 6 ppm was 10.5-fold higher than at 2 ppm. Casanova 18 et al. (1989) exposed groups of F344 rats to formaldehyde concentrations of 0.3, 0.7, 2, 6, 19 or 10 ppm for 6 hours. DNA-protein crosslinks occurred at all concentrations, but the 20 slope of the concentration-response curve at 10 ppm was 7.3-fold greater than at 0.3 ppm. 21 Casanova et al. (1994) compared the yield of crosslinks between groups of pre-exposed 22 and naïve male F344 rats. Groups were pre-exposed to 0.7, 2, 6, or 15 ppm in one 23 experiment and 6 or 10 ppm in another experiment (6 hours/day, 5 days/week) for 11 24 weeks and 4 days while naïve rats were exposed to room air. On the fifth day of the 25 twelfth week animals were simultaneously exposed (3 hours) to the same concentrations 26 used in pre-exposure. Crosslink yields increased nonlinearly in a concentration-dependent 27 manner in both pre-exposed and naïve groups, but the yields were smaller in pre-exposed 28 rats, suggesting that accumulation of crosslinks did not occur. At low concentrations ( $\leq 2$ 29 ppm) crosslink yields were similar in pre-exposed and naïve rats, but at higher 30 concentrations, crosslink yields were greater in naïve than pre-exposed rats.

1 Cosma et al. (1988b) used an open-ended, flow-through rat tracheal implant model to 2 investigate DNA-protein crosslinks caused by benzo[a]pyrene and formaldehyde. Two 3 tracheas from male F344 rats were implanted s.c. in the retroscapsular region of 4 syngeneic recipients. After 4 weeks, both ends of the tracheal implants were connected to 5 the surface by two terminal tracheostomies. The tracheas were exposed twice weekly for 6 2, 4, or 8 weeks to gelatin pellets containing 0.005, 0.01, 0.05, or 2% formaldehyde. 7 There was a dose-dependent increase in crosslinks in the tracheal epithelium. The authors 8 also compared the induction and removal of crosslinks following single and multiple 9 exposures. The response was virtually identical for exposure either once or 5-times twice weekly to 0.2% formaldehyde when measured 3 hours after the last exposure. The 10 11 removal of crosslinks following 1 or 4 exposures demonstrated nearly complete repair in 12 either case by 72 hours.

13 DNA-protein crosslink yields were about six-fold higher in the lateral meatus (an area of 14 high tumor yield) than in the medial or posterior meatures (areas with low tumor yield) of 15 the rat nose (Casanova et al. 1994). In male rhesus monkeys, crosslink concentrations in 16 the nose were highest in the middle turbinates while lower concentrations occurred in the 17 anterior lateral wall, septum, and nasopharynx (Casanova et al. 1991, Heck et al. 1989). 18 Low, but statistically significant concentrations of crosslinks were found in the larynx, 19 trachea, carina, or in the proximal portions of the major bronchi in monkeys exposed to 2 20 or 6 ppm but not to 0.7 ppm. No crosslinks were found in the maxillary sinuses or lung 21 parenchyma in any of the nine monkeys tested.

22 Crosslinks and strand breaks in tissues other than the upper respiratory tract also have 23 been reported in rodents. Wang and Liu (2006) [reported in an English abstract] 24 investigated developmental and maternal toxicity in mice. Pregnant mice were injected 25 with 0.2 to 20 mg/kg per day from gestation day 6 to 19. Single-cell gel electrophoresis 26 was used to test for DNA damage (crosslinks and breaks) in maternal and fetal liver cells. 27 There was no DNA damage in the livers of fetal mice in the low-dose group; however, 28 increased DNA breakage was observed in the group exposed to  $\geq 1 \text{ mg/kg}$  per day, and 29 increased DNA-protein crosslinks occurred at 2 to 20 mg/kg per day. DNA damage

1 increased with dose in the dams, beginning at 0.2 mg/kg per day, but no increase in

2 DNA-protein crosslinks was observed.

3 Im et al. (2006) evaluated the genotoxic effects of formaldehyde exposure in rat 4 lymphocytes and liver. Male Sprague-Dawley rats (10 per group) were exposed to 0-, 5-, 5 or 10-ppm formaldehyde 6 hours/day, 5 days/week for 2 weeks in an inhalation chamber. 6 The comet assay was used to evaluate DNA single-strand breaks. Exposure to 5- or 10-7 ppm formaldehyde resulted in a significant, and dose-dependent, increase in single-strand 8 breaks in both lymphocytes and liver. Speit (2006) criticized this study and stated that 9 formaldehyde-induced DNA-protein crosslinks would be expected to reduce DNA 10 migration as measured by the comet assay. One study did not find crosslinks in bone 11 marrow of rats exposed to 15-ppm formaldehyde for 6 hours (Casanova-Schmitz et al. 12 1984a).

13 Lutz (1986) evaluated the levels of DNA-protein crosslinks produced from endogenous 14 formaldehyde generation. This author determined the level of DNA-protein crosslinks in 15 rat liver under conditions of maximum intracellular formaldehyde generation and 16 compared the results with positive control data from *in vitro* incubations of liver 17 homogenate with formaldehyde and methanol and with literature data on crosslinks in the 18 rat nasal epithelium. Since endogenous formaldehyde is generated by oxidation of 19 methanol (primarily in the liver), male Sprague-Dawley rats were given 1 g methanol per 20 kg body weight by gavage. Another group also received 0.6 g/kg disulfiram, an inhibitor 21 of acetaldehyde oxidation, under the assumption that higher steady-state levels of 22 formaldehyde might be achieved. After 4 hours, the rats were given ethanol by gavage to 23 inhibit further methanol oxidation, and were killed to isolate the chromatin fraction from 24 the liver. The levels of endogenous formaldehyde formed in the liver did not cause an 25 increase in DNA-protein crosslinks.

Test system	Concentration (LEC or HIC) <sup>a</sup>	Effect	Results	References	
Rat (nasal mucosa)	0.3 ppm		+	Casanova et al. 1989	
	0.7 ppm <sup>b</sup>		+	Casanova et al. 1994	
	2 ppm	DDV	+	Casanova-Schmitz et al. 1984a	
	2 ppm	DFA	+	Heck et al. 1986	
	2 ppm		+	Casanova and Heck Hd 1987	
	6 ppm		+	Lam <i>et al</i> . 1985	
Rat (bone marrow, olfactory mucosa)	15 ppm	DPX	_	Casanova-Schmitz et al. 1984a	
Rat (tracheal implant)	0.005% <sup>c</sup>	DPX	+	Cosma <i>et al.</i> 1988b	
Rat (fetal liver)	0.2 mg/kg <sup>d</sup>	DPX	+	Wang and Liu 2006	
Rhesus monkey (nasal	0.7 ppm		+	Heck et al. 1989	
turbinates)	0.7 ppm	0.7 ppm		Casanova et al. 1991	
Rhesus monkey (larynx, trachea, carina, bronchi)	2 ppm DPX		+		
Rhesus monkey (maxillary sinuses, lung)	maxillary 6 ppm		_	Casanova <i>et al.</i> 1991	
Rat (lymphocytes) 5 ppm <sup>e</sup>		SB	+	Im stal 2006	
Rat (liver)	Rat (liver) 5 ppm <sup>e</sup>		+	IIII <i>et al.</i> 2000	
Rat (maternal liver)	0.2 mg/kg <sup>d</sup>	SB	+	Wang and Lin 2006	
Rat (fetal liver)	1 mg/kg <sup>d</sup>	SB	+	wang anu Liu 2000	

 Table 5-18. In vivo studies of DNA-protein crosslinks and strand breaks in mammalian systems

+ = positive result for indicated effect; - = negative result for indicated effect.

LEC = lowest effective concentration; HIC = highest ineffective concentration; DPX = DNA-protein crosslinks; SB = DNA strand breaks (most were single-strand breaks).

<sup>a</sup> Single inhalation exposure (3-6 h) unless otherwise noted.

<sup>b</sup> Included pre-exposed groups (6 h/day, 5 d/wk, 11 wk + 4 d).

<sup>c</sup> Instillation exposure twice weekly for 2, 4, or 8 wk.

<sup>d</sup> Intraperitoneal injection to pregnant mice on gestation days 6 to 19.

<sup>e</sup> 5 d/wk for 2 wk.

## 1 5.6.3.2 Cytogenetic effects

2 Studies evaluating cytogenetic effects (SCE, micronucleus formation, and chromosomal

3 aberrations) due to formaldehyde exposure are described below and summarized in

4 Tables 5-19 and 5-20.

- 5 In vitro studies
- 6 In human and animal cells formaldehyde exposure (0.03 to 2 mM) caused SCE (Chinese
- 7 hamster ovary cells, Chinese hamster V79 lung fibroblast cells, human lymphocytes, and
- 8 human whole blood), chromosomal aberrations (Chinese hamster ovary cells, Syrian
- 9 hamster embryo cells, human lymphocytes and human fibroblasts), and micronuclei

1 (Chinese hamster V79 cells, human MRC5CV cells, and human whole blood) (Table 5-

2 19). All of the reported studies showed a positive correlation between formaldehyde

3 treatment and observed effect, although the lowest effective concentration varied with

4 different test systems, as well as for the same cell assay under similar or modified

5 conditions.

6 Recent studies have characterized the cytogenetic effects in more detail. Speit *et al.* 

7 (2000) reported that the frequency of micronuclei was increased [statistics not reported]

8 in two different DNA repair-deficient cell lines (xeroderma pigmentosum and Fanconi

9 anemia) compared with human cell lines with normal repair. Micronucleus frequency was

10 increased [statistics not reported] in Chinese hamster V79 cell cultures receiving repeated

11 treatments (3 treatments with time intervals of 3 hours) compared with cultures receiving 12 a single treatment, but not when the repeated treatment interval was increased to 24 hours

a single treatment, but not when the repeated treatment interval was increased to 24 hours
(Speit *et al.* 2007a). Schmid and Speit (2007) reported that exposure to formaldehyde

14 only increased micronucleus formation in human blood cultures using protocols in which

15 formaldehyde was added 44 hours after the start of culture (i.e., the last cell cycle before

16 preparation). In their study, 81% of micronuclei were centromere negative, compared

17 with 55% centromere-negative micronuclei in controls.

18 Characterization of the genotoxic action of formaldehyde was investigated in a study

19 utilizing the SCE assay in two mammalian cell lines, Chinese hamster V79 lung

20 fibroblasts and human A549 lung cells (Neuss and Speit 2008). For each of these cell

21 lines, formaldehyde treatment with 0.1 mM for 1 hour, then growth in the presence of 5-

22 bromodeoxyuridine (BrdU) for two cell cycles, resulted in statistically significant (P <

23 0.01) SCE induction. When the V79 cells were treated with formaldehyde for 1 hour then

24 cultured with BrdU 4 hours later, the effective concentration was increased to 0.2 mM,

- 25 suggesting DNA repair. Further, when the A549 cells were treated with 0.05 mM
- 26 formaldehyde for 1 hour then co-cultured with V79 cells immediately, there was enough
- formaldehyde still present to significantly (P < 0.05) induce SCE in the V79 cells. When
- 28 the A549 cells were treated at a maximum dose of 0.3 mM, then washed before co-
- 29 cultivating with V79 cells, there was no SCE induction in the V79 cells. The authors

1 suggested that this lack of response indicated that the formaldehyde was bound and/or

2 inactivated in the A549 cells.

3 Although most of these *in vitro* studies did not report any cytotoxicity findings, in five of 4 the studies cytotoxic effects were observed in cells treated with doses at which significant 5 cytogenetic effects were also reported. In 1986, Schmid et al. noted that 0.25 and 0.5 mM 6 formaldehyde treatments had a marked effect on cultured human lymphocytes and that 7 there was no cell proliferation at all in cells treated with 1.0 mM formaldehyde. Merk and 8 Speit (1998) evaluated cytotoxicity in V79 cells using relative cloning efficiency as a 9 measure of long-term survival. In this study, treatment of cells with 0.125 mM 10 formaldehyde significantly (P < 0.05) reduced the clonal growth of the cells to about 11 72% of controls. Treatments of clearly genotoxic doses of 0.25 and 0.5 mM 12 formaldehyde reduced the relative cloning efficiency in these cells to 40% and less than 13 10%, respectively.

14 According to Schmid and Speit (2007), the cytotoxic effect of formaldehyde appears to

15 be concurrent with, or may even precede, the genotoxic response. Specifically, they noted

16 a reduction in the proliferation index (i.e., increased cytotoxicity) of the blood cultures

17 treated with 0.2 mM formaldehyde, a dose at which SCE were significantly induced.

18 Further, there was a non-significant cytotoxic effect noted at 0.1 mM formaldehyde

19 treatment, which also showed an increased, although not statistically significant,

20 induction in SCE. Interestingly, in a different paper but using V79 Chinese hamster cells,

21 the same authors (Speit *et al.* 2007a), reported that SCE was significantly (P < 0.01)

22 induced at 0.1 mM formaldehyde treatment; however, in these cells the proliferation

23 index was not reduced, but was equivalent to the control value.

24 Cytotoxic effects of formaldehyde were evaluated in the human A549 cell line by Speit et

25 *al.* (2008b) by measuring colony-forming ability and cell growth inhibition. With

26 continuous two-week exposure to 0.02 mM formaldehyde, colony-forming ability was

27 significantly reduced to approximately 40% of controls; cell growth was reduced to less

28 than 20% with a continuous 48-hour treatment with 0.2 mM formaldehyde (significance

29 for both determined using Dunnett test, 1% level). Also reported was a non-significant

- 1 reduction (about 80% of controls) in cell growth measured after a one-hour treatment
- 2 with up to 0.5 mM formaldehyde.

Effect	Test system	Lowest ef concentr treatment	ffective ation <sup>a</sup> , duration	Result	Cytotoxicity or RTG (% survival)	References
SCE	Chinese hamster ovary	[0.03 mM]	24 h	+	ND	Obe and Beek 1979
	cells	[0.2 mM]	2 h	+	ND	Natarajan <i>et al</i> . 1983
		[0.04 mM]	26 h	+	NA	Galloway et al. 1985
		0.15 mM	1 h	+	ND	Garcia et al. 2009
	Chinese hamster V79	0.067 mM	28 h	+	ND	Basler et al. 1985
	cells	0.13 mM	2 h	+	ND	Basler et al. 1985
		0.1 mM	2 h	+	100 <sup>b</sup>	Speit <i>et al</i> . 2007a
		0.125 mM	4 h	+	72°,92°	Merk and Speit 1998, 1999
	Co-cultivation study <sup>d</sup>					
	A549 Human lung			+	ND	Neuss and Speit 2008
	cells	0.1 mM	1 h	+	ND	
	V79 cells (4 h	0.2 mM	1 h	$+^{c}$	ND	
	recovery)	0.05 mM	1 h			
	V79 cells + A549 cells					
	Human lymphocytes	0.125 mM	1 h	+	67 <sup>c,e</sup>	Schmid <i>et al</i> . 1986
		[0.167 mM]	24 h	+	ND	Obe and Beck 1979
		[0.167 mM]	72 h	+	20	Kreiger and Garry 1983
	Human whole blood	0.2 mM	72 h	+	75 <sup>°</sup>	Schmid and Speit 2007
CA	Chinese hamster ovary	[0.53mM]	8–12 h	+	NA	Galloway <i>et al</i> . 1985
	cells	[0.2 mM]	2 h	+	ND	Natarajan <i>et al</i> . 1983
		0.15 mM	2 h	+	ND	Garcia <i>et al</i> . 2009
	Chinese hamster lung fibroblasts	[0.6 mM]	24 h	+	ND	Ishidate Jr <i>et al</i> . 1981
	Syrian hamster embryo	0 033 mM	24 h	+	94	Hikiba et al. 2005
	cells	$0.33 \text{ mM}^{\text{f}}$	24 h	+	91	Hagiwara <i>et al.</i> 2005
	Human lymphocytes	0.5 mM	1 h	$+^{c}$	$0^{c,e}$	Schmid <i>et al.</i> 1986
		0.33 mM	NA	+ <sup>g</sup>	NA	Miretskaya and Shvartsman
		0.125 mM	1 h	$+^{h}$	ND	1982
						Dresp and Bauchinger 1988
	Human fibroblasts	2 mM	0.25 h	+	ND	Levy et al. 1983

 Table 5-19. In vitro studies of cytogenetic effects of formaldehyde in mammalian cells

Effect.	<b>T</b> 4	Lowest ef	ffective ation <sup>a</sup> ,	Decult	Cytotoxicity or RTG	Deferences
Effect	lest system	treatment	duration	Result	(% survival)	References
MN	Chinese hamster V79	0.075 mM	2 h	+	ND	Speit <i>et al</i> . 2007a
	cells	0.125 mM	4 h	+	72 <sup>°</sup>	Merk and Speit 1998
	Human MRC5CV (normal) XP cell line (repair deficient) FA cell line (repair deficient)	0.125 mM	2 h	+ <sup>i</sup>	ND	Speit <i>et al.</i> 2000
	Human whole blood	0.3 mM	72 h	+ <sup>j</sup>	77 <sup>c</sup>	Schmid and Speit 2007

+ = positive result for indicated effect, - = negative result for indicated effect.

CA = chromosomal aberration; FA = Fanconi anemia; MN = micronucleus; NA = not available; ND = not done; RTG = relative total growth; SCE = sister chromatid exchange; XP = xeroderma pigmentosum.

<sup>a</sup> Units in brackets [] were converted to mM from reported exposure data to facilitate comparison.

<sup>b</sup> Cytotoxicity measured by calculating proliferation index, which was equal to control (estimated from graph) at this dose. <sup>c</sup> Estimated data from graph.

<sup>d</sup> Treated A549 cells 1 h then co-cultivated with V79 showed induction in V79; however, changing media after treatment, then adding V79 cells, resulted in no SCE induction.

<sup>e</sup>Cytotoxicity was based on third cycle metaphase measured, as compared with control.

<sup>f</sup> Treatment substance was formocresol, potential confounding effect due to formaldehyde component.

<sup>g</sup> As cited by IARC 2006.

<sup>h</sup> Dose was negative with standard method, but positive in modified (premature chromosome condensation) technique. <sup>i</sup> The effect was enhanced in the repair-deficient cell lines compared to the normal cell line.

<sup>j</sup>Modified protocol: cells were cultured 44 hours before treatment; treatments at 0 and 24 hours were negative at this dose.

#### 1 In vivo studies

2 Formaldehyde did not cause micronucleus formation in bone marrow or chromosomal

3 aberrations in bone marrow, spleen, or spermatocytes of mice exposed to formaldehyde

4 by i.p. injection; no inhalation studies were available in mice. *In vivo studies in rats gave* 

5 mixed results. Kligerman *et al.* (1984) did not find SCE or chromosomal aberrations in

6 lymphocytes of F344 rats exposed to 15-ppm formaldehyde 6 hours/day for 5 days.

7 Increasing the duration of the 15-ppm formaldehyde treatment to 4 weeks did not yield

8 SCE or chromosomal aberrations in peripheral blood of F344 male rats (Speit *et al.* 

9 2009). When administered in a single oral dose of 200 mg/kg to Sprague-Dawley rats,

10 formaldehyde induced micronuclei in the gastrointestinal tract (Migliore *et al.* 1989).

11 Dallas et al. (1992) investigated chromosomal aberrations in pulmonary lavage cells and

- 12 bone marrow of male Sprague-Dawley rats exposed to 0-, 0.5-, 3-, or 15-ppm
- 13 formaldehyde for 6 hour/day, 5 days/week, for 1 to 8 weeks. There was no significant
- 14 increase in chromosomal aberrations in bone marrow, but there was a statistically
- 15 significant increase in chromosomal aberrations in pulmonary lavage cells in the
- 16 high-dose group. Kitaeva et al. (1990) investigated cytogenetic effects of inhaled

- 1 formaldehyde in the bone marrow of female Wistar rats exposed to 0.5 or 1.5 mg/m<sup>3</sup> [0.4
- 2 ppm or 1.2 ppm] for 4 hours/day (except weekends and holidays) for 4 months. Bone
- 3 marrow was collected within 48 to 72 hours after exposure was stopped. There was a
- 4 statistically significant increase in the number of bone marrow cells with chromosomal
- 5 aberrations at both dose levels compared with controls.

Effect	Test system	Concentration	Result	References
SCE	F344 rat (lymphocytes, inh., 6 h/d, 5 d)	15 ppm	-	Kligerman <i>et al.</i> 1984
	F344 male rat (peripheral blood, inh., 6h/d, 5 d/wk, 4 wk)	15 ppm	_	Speit <i>et al.</i> 2009
CA	F344 rat (lymphocytes, inh., 6 h/d, 5 d)	15 ppm	_	Kligerman <i>et al</i> . 1984
	F344 male rat (peripheral blood, inh, 6h/d, 5 d/wk, 4 wk)	15 ppm	_	Speit <i>et al.</i> 2009
	Sprague-Dawley rat (bone marrow, inh., 6 h/d, 1–8 wk)	15 ppm	_	-Dallas <i>et al.</i> 1992
	Sprague-Dawley rat (pulmonary lavage cells, inh., 6 h/d, 1–8 wk)	15 ppm	+	
	Wistar rat (bone marrow, inh., 4 h/d, 4 mo)	0.4 ppm	+	Kitaeva et al. 1990
	Mouse (bone marrow, i.p.)	25 mg/kg	-	Natarajan <i>et al.</i> 1983
	Mouse (spleen, i.p.)	25 mg/kg	_	Natarajan <i>et al.</i> 1983
	Mouse (spermatocytes, i.p.)	50 mg/kg	_	Fontignie-Houbrechts 1981
MN	Sprague-Dawley rat (G.I., p.o.)	200 mg/kg	+	Migliore et al. 1989
	Mouse (bone marrow, i.p.)	30 mg/kg	_	Gocke <i>et al.</i> 1981

 Table 5-20. Cytogenetic effects of formaldehyde in mammals in vivo

CA = chromosomal aberration; FA = Fanconi anemia; HIC = highest ineffective concentration; inh. = inhlation; i.p. = intraperitoneal; LEC = lowest effective concentration; MN = micronucleus; p.o. = *per os* (by mouth); SCE = sister chromatid exchange; XP = xeroderma pigmentosum.

+ = positive result for indicated effect, - = negative result for indicated effect.

<sup>a</sup> The effect was enhanced in the repair-deficient cell lines compared to the normal cell line.

- 6 5.6.3.3 *Mutations*
- 7 Formaldehyde exposure has caused mutations in mammalian cells in vitro and dominant
- 8 lethal mutations in mice and rats (Table 5-21). All but one of the *in vitro* studies was
- 9 positive. Two i.p. injection studies reported negative results for dominant lethal mutations
- 10 in mice, while one study (given a higher dose) reported a weak positive response.
- 11 Dominant lethal mutations were observed in rats exposed to formaldehyde by inhalation
- 12 and i.p. injection.

1 Heritable mutations in mice were reported in a study by Liu et al. (2009b) exposing male

- 2 specific-pathogen-free ICR mice to 2 to 200 mg/m<sup>3</sup> formaldehyde [formalin vapor] for 2
- 3 hours. After a 6-week recovery, the mice were bred and sperm DNA was extracted from
- 4 the male mice. Somatic DNA for analysis was extracted from tail tissue of both parents as
- 5 well as from offspring. Utilizing three expanded simple tandem repeats (ESTR) probes,
- 6 mutation rates were quantitatively and qualitatively evaluated to be both dose dependent
- 7 and mainly inherited from the paternal germ line. The authors speculated that
- 8 ramifications of this altered DNA, and subsequent abnormal protein expression, could
- 9 result in malformations in the offspring.

	Concentration					
Test system	LEC/HIC	Result	References			
In vitro						
Chinese hamster V79 cells (Hprt	0.3 mM	+	Grafström et al. 1993			
locus)	0.5 mM	-	Merk and Speit 1998, 1999			
Mouse lymphoma L5178Y cells ( $Tk^{+/-}$	0.8 mM	+	Mackerer et al. 1996			
locus)	> 0.067 mM	+	Speit and Merk 2002			
	0.13 mM	+	Goldmacher and Thilly 1983			
Human lymphoblast (TK6)	0.03 mM	+	Craft <i>et al</i> . 1987			
Tuman Tymphoblast (TK0)	0.15 mM	+	Crosby et al. 1988			
	0.15 mM	+	Liber <i>et al.</i> 1989			
Human bronchial fibroblasts and	0.1 mM	+	Grafström et al. 1985			
epithelial cells (HPRT locus)	0.1 mM	+	Grafström 1990			
In vivo						
Mouse (dominant lethal, i.p.)	20 mg/kg	-	Epstein and Shafner 1968			
	20 mg/kg	_	Epstein et al. 1972			
	50 mg/kg	(+)	Fontignie-Houbrechts 1981			
Rat (dominant lethal, inh., 4 h/d, 4 mo)	1.2 ppm	(+)	Kitaeva et al. 1990			
Rat (dominant lethal, i.p.)	0.125 mg/kg	+	Odeigah 1997			
Mouse (heritable mutation, inh.)	200 mg/m <sup>3</sup>	+	Liu et al. 2009b			

 Table 5-21. Mutagenic effects of formaldehyde in mammalian systems

+ = positive study, (+) = weak positive study, - = negative study.

inh. = inhalation; i.p. = intraperitoneal; LEC = lowest effective concentration; HIC = highest ineffective concentration; SCC = squamous cell carcinoma.

## 10 5.6.3.4 Other effects

- 11 Other genetic and related effects reported in mammalian *in vitro* studies include
- 12 unscheduled DNA synthesis (UDS), inhibition of DNA repair, and cell transformation
- 1 (Table 5-22). UDS was observed in rat hepatocytes (Williams et al. 1989), human HeLa
- 2 cells (Martin et al. 1978), and Syrian hamster embryo cells (Hamaguchi and Tsutsui
- 3 2000), but not in human bronchial epithelial cells (Doolittle et al. 1985). Other studies
- 4 indicate that formaldehyde can inhibit DNA repair processes and induce cell
- 5 transformation. Emri et al. (2004) investigated the interactions of low concentrations of
- 6 formaldehyde and UV radiation in human skin cells. Keratinocytes and fibroblasts
- 7 exposed to 10 μM formaldehyde prior to UV irradiation inhibited DNA repair kinetics
- 8 after UVB and UVC, but not after UVA irradiation. Single-strand breaks that were
- 9 repaired within 3 to 6 hours following exposure to UVB or UVC radiation, were still
- 10 present at these time points in the presence of formaldehyde. UVC-induced chromosomal
- 11 damage was also increased in the presence of formaldehyde at a concentration (12.5  $\mu$ M)
- 12 that did not cause micronuclei. These authors concluded that environmental exposure to
- 13 formaldehyde might contribute to UV-induced skin carcinogenesis.

Test system	Concentration LEC/HIC	Effect	Result	References
In vitro				
Rat hepatocytes	400 mM	UDS	+	Williams et al. 1989
Syrian hamster embryo cells	0.1 mM	UDS	+	Hamaguchi and Tsutsui 2000
Human HeLa cells	10 <sup>-5</sup> mM	UDS	+	Martin et al. 1978
Human bronchial epithelial cells	0.1 mM	UDS	-	Doolittle et al. 1985
Human bronchial epithelial cells and fibroblasts and skin fibroblast	0.2 mM	DNA repair (inhibition)	+	Grafström <i>et al.</i> 1984
Human MRC5CV normal cells XP cell line (repair defcient) FA cell line (repair deficient)	0.125 mM	DNA repair (inhibition)	+	Speit <i>et al.</i> 2000
Human skin fibroblasts and keratinocytes	10 mM	DNA repair (inhibition)	+	Emri <i>et al.</i> 2004
C3H10T1/2 mouse cells	0.017 mM	Cell transformation	$+^{a}$	Ragan and Boreiko 1981

Table 5-22. Other genetic effects of formaldehyde in mammalian systems

+ = positive study; - = negative study.

<sup>a</sup> Positive only in the presence of 12-*O*-tetradecanoylphorbol 13-acetate.

LEC = lowest effective concentration; HIC = highest ineffective concentration; UDS = unscheduled DNA synthesis.

### 1 5.6.4 Human in vivo studies

2 The genetic effects of formaldehyde have been investigated in humans that were exposed

- 3 in a number of settings (e.g., hospitals, pathology and anatomy laboratories,
- 4 woodworking facilities, formaldehyde manufacturing facilities, mortuaries, and
- 5 residences) and are described below. Most of these studies were reviewed by WHO
- 6 (1989), Conaway et al. (1996), IARC (1995, 2006), or Liteplo and Meek (2003).

### 7 5.6.4.1 DNA-protein crosslinks and strand breaks

8 Shaham *et al.* (1996a, 1997) conducted a pilot study to investigate the use of DNA-

- 9 protein crosslinks as a biomarker of formaldehyde exposure in humans. DNA-protein
- 10 crosslinks were measured in white blood cells from 12 exposed workers (physicians and
- 11 technicians at the Pathology Institute) and 8 controls. The workers had been exposed to
- 12 formaldehyde from 2 to 31 years with a mean of 13 years. Formaldehyde concentrations
- 13 were measured in the room air and by personal samples. Concentrations ranged from
- 14 about 1.4 to 3.1 ppm. There was a significant difference (P = 0.03, *t*-test) between the
- 15 levels of crosslinks in exposed workers and controls, and a significant difference (P <
- 16 0.05) between the most-exposed workers (technicians) and less-exposed workers
- 17 (physicians) (Table 5-23). Furthermore, there was a linear relationship between the years
- 18 of exposure and levels of crosslinks. Smoking did not influence the results. This was the
- 19 first study to measure DNA-protein crosslinks in humans exposed to formaldehyde.

20 Shaham *et al.* (2003) conducted a follow-up study of the relationship of occupational

- 21 exposure to formaldehyde and DNA-protein crosslinks. This study also investigated
- 22 effects on p53 protein expression. The workers included physicians, laboratory assistants
- and technicians, and hospital orderlies from 14 hospital pathology departments that had a
- 24 mean exposure period of 15.9 years (range 1 to 51 years). Fifty-nine (59) men and 127
- 25 women were included in the exposed group and were further divided into subgroups
- based on low and high exposures. The low-exposure group (0.04 to 0.7 ppm) included
- 27 laboratory assistants and technicians, while the high-exposure group (0.72 to 5.6 ppm)
- 28 included physicians and orderlies. [No explanation was given for physicians being in the
- 29 less highly exposed group in the 1996-97 study but in the highly exposed group in the
- 30 2003 study.] The control group included 213 administrative workers (127 men and 86

1	women) from the same hospitals. There were significant differences in the age
2	distribution, sex, origin, and education between the exposed and control group.
3	Therefore, the data were adjusted for these variables. DNA-protein crosslinks were
4	measured in the mononuclear cell fraction of peripheral blood. Also, p53 proteins,
5	including pantropic p53 (wild type and mutant) and mutant p53, were measured in serum.
6	The adjusted means of crosslinks between the exposed and unexposed groups were
7	compared by analysis of variance, the comparison between the two levels of exposure
8	was evaluated by the Mann-Whitney U test, and the Chi square test was used to compare
9	prevalence of high p53 levels. The adjusted mean amount of crosslinks was significantly
10	higher ( $P < 0.01$ ) in the total exposed group compared with the control group (Table 5-
11	23). Age, smoking habits, years of education, and origin were not significant
12	confounders. The mean amount of crosslinks did not show significant differences based
13	on level of exposure or median years of exposure ( $\leq 16$ versus > 16). Formaldehyde
14	exposure was associated with an increased risk of having a higher level of pantropic p53
15	protein above 150 pg/mL. A significantly higher proportion of exposed workers with
16	DNA-protein crosslink levels above the median level of 0.187 had elevated pantropic p53
17	protein levels compared with exposed workers with crosslink levels less than 0.187.

Group	N	DNA-protein crosslinks/total DNA	Pantropic p53 > 150 pg/mL (%)	Reference
Control	8	$0.23\pm0.067^a$	NT	Shaham et al.
Exposed (total)	12	$0.28 \pm 0.055 *$		1996a, 1997
Low exposure	6	$0.26 \pm 0.044$		
High exposure	6	$0.32 \pm 0.043^{*b}$		
Control	213	$0.14\pm0.006^{\rm c}$	36.3	Shaham et al.
Exposed	186	$0.21 \pm 0.006^{**}$	44.1	2003
Low exposure	NR	$\leq 0.187$	33.3 <sup>d</sup>	
High exposure	NR	> 0.187	55.7** <sup>b,d</sup>	

 Table 5-23. DNA-protein crosslinks and pantropic p53 protein levels in medical workers exposed to formaldehyde

\* P < 0.05; \*\* P < 0.01 (compared with controls, unless otherwise noted, see text for method).

NR = not reported, NT = not tested

 $a^{\pm} \pm SD.$ 

<sup>b</sup> Compared with low-exposure group.

 $^{c} \pm SE.$ 

<sup>d</sup> Low and high exposure groups based on DNA-protein crosslink levels above or below the median value of 0.187.

1 Costa et al. (2008) compared DNA damage in 30 pathology anatomy laboratory workers 2 in four hospitals in Portugal with 30 matched controls (age, sex, lifestyle factors, and 3 smoking habits) selected from administrative staff in the same hospitals. This study also 4 examined SCE and micronuclei (discussed below) and the association between 5 biomarkers and polymorphic genes of xenobiotic metabolizing and DNA repair enzymes. 6 The exposed group had been employed for 5 months to 27 years (mean 11 years). The 7 mean level of exposure measured at the breathing zone of the subjects was 0.44 ppm 8 (range 0.04 to 1.58 ppm). The subjects began work at 9 a.m. and blood samples were 9 collected between 10 and 11 a.m. The alkaline version of the comet assay was used to 10 evaluate DNA damage in lymphocytes. There was a significant increase (P < 0.05) in 11 comet tail length in exposed workers compared with controls, and a positive association 12 was found between formaldehyde exposure level and comet tail length. The 13 polymorphisms, age, and smoking status examined did not have a significant effect on 14 DNA damage. DNA damage was significantly increased in exposed females compared 15 with exposed males, but no effect on gender was observed in controls. Age and smoking 16 status did not affect DNA damage.

Genotoxicity studies published on peripheral lymphocytes of Chinese workers exposed to
formaldehyde were reviewed by Tang *et al.* (2009). Increases in DNA damage to

19 lymphocytes (comet assay) were reported in three studies in exposed workers (Jiang *et al.* 

20 2006, Tong et al. 2006, Yu et al. 2005).

21 5.6.4.2 DNA repair and mutations

22 Three studies were reviewed that examined the effects of formaldehyde exposure on

23 DNA repair (Hayes et al. 1997, Orsiere et al. 2006, Schlink et al. 1999). The study

24 populations included medical or mortuary science students and anatomy laboratory

25 workers. One study investigated the mutagenicity of urine samples collected from

- 26 medical workers (Connor *et al.* 1985a).
- 27 Hayes et al. (1997) examined the effects of formaldehyde exposure on DNA repair
- 28 capacity in mortuary science students.  $O^6$ -alkylguanine DNA alkyltransferase (AGT)
- 29 activity was measured in peripheral blood lymphocytes of 23 students (16 males and 7
- 30 females) before and after a 9-week course in embalming techniques. Personal

1 formaldehyde exposure was measured at the breathing zone during embalming, and 2 short-term (peak) exposure was measured with a continuous reading instrument. 3 Cumulative formaldehyde exposure was measured as ppm-hours formaldehyde for each 4 subject. The average air concentration of formaldehyde during embalming was about 1.5 5 ppm, but short-term monitoring during some embalmings showed that peak exposures 6 were 3 to 9 times higher than the time-weighted average concentration. Most students 7 performed between five and nine embalmings during the class. However, 15 students 8 reported prior exposure to formaldehyde during embalming procedures conducted within 9 90 days of the class. Differences in AGT activity were assessed by the Wilcoxon signed 10 rank test and by analysis of variance. Baseline AGT activity was somewhat lower (P =11 0.08) in students who reported a prior history of embalming. There were no significant 12 differences in baseline AGT activity based on gender, age, or current tobacco use. At the 13 end of the study, AGT activity decreased in 17 students and increased in 6 students 14 compared with baseline values (P < 0.05). Among the eight students with no previous 15 embalming experience, AGT activity decreased in all but one. Although post-exposure 16 AGT activity tended to decrease, no clear link was established between formaldehyde 17 exposure and AGT activity. The authors noted several study limitations. These included a 18 small number of subjects, many of which had prior exposure to formaldehyde, and the 19 study did not allow for a detailed temporal association between formaldehyde exposure 20 and AGT activity.

21 In a subsequent study by the same group of researchers, Schlink et al. (1999) measured AGT (also known as  $O^6$ -methylguanine DNA methyltransferase [MGMT]) activity in 22 23 mononuclear blood cells in 57 medical students before and after taking an anatomy 24 course. The students were exposed to an average formaldehyde concentration of 0.2 25 mg/m<sup>3</sup> [0.16 ppm] for 6 hours/week for about 16 weeks. Age, sex, cigarette smoking, 26 alcohol consumption, and allergic disease did not significantly affect MGMT activity. 27 The mean MGMT activity after 111 days of exposure was  $128.2 \text{ fmol}/10^6$  cells, which was not significantly different from the baseline value of  $133.2 \text{ fmol}/10^6 \text{ cells}$ . There also 28 29 was no significant difference in MGMT activity in a second group of 16 medical students 30 with mean formaldehyde exposure of 0.8 mg/m<sup>3</sup> [0.64 ppm] compared with a group of 51

1 students without formaldehyde exposure. Thus, formaldehyde did not affect MGMT

2 activity in mononuclear blood cells in medical students.

3 Orsière *et al.* (2006) examined the genotoxic effects of formaldehyde in 59 pathology and 4 anatomy laboratory workers from five hospitals. Personal air sampling was conducted for 5 short-term (15 minutes) and long-term (8 hours) intervals. The mean formaldehyde 6 concentrations were 2 ppm (range < 0.1 to 20.4 ppm) and 0.1 ppm (range < 0.1 to 0.7 7 ppm) in the short-term and long-term air samples. The highest formaldehyde 8 concentrations were recorded during macroscopic examination of formaldehyde-9 preserved specimens. Blood samples were collected from each worker in the morning 10 before beginning work and at the end of the work day. The chemiluminescence 11 microplate assay was used to measure primary DNA damage (ex vivo base or nucleotide 12 excision-repair activity) in peripheral lymphocytes. Data were expressed in relative light 13 units (RLU) per ng of DNA. Chromosomal damage was determined using the 14 cytokinesis-blocked micronucleus assay (see Section 5.6.4.3 for a description of these 15 results). There was no difference in DNA damage at the beginning of the work day 16 compared with the end of the work day. The mean pre-shift RLU was  $3.9 \pm 0.5$  compared 17 with the post-shift value of  $3.6 \pm 0.5$ . There was no correlation of DNA damage with 18 work practices or with personal air sampling data.

19 Connor et al. (1985a) tested the mutagenicity of urine samples from 19 autopsy service 20 and pathology department workers at the University of Texas medical school. The control 21 group included 20 individuals selected from the staff, faculty, and student populations 22 and were matched to the exposure group based on sex, age, and alcohol, tobacco, and 23 marijuana use. Medical history, past use of medications, exposure to industrial chemicals, 24 and other factors that could possibly affect the outcome of the study were considered in 25 the analysis. Urine samples were collected three times at 2-month intervals and were 26 tested for mutagenicity in S. typhimurium strains TA98 and TA100 with and without S9 27 metabolic activation. Formaldehyde concentrations ranged from 0.1 ppm (detection limit) 28 outside the immediate work area to 5.8 ppm in the work area. The estimated time-29 weighted average formaldehyde concentrations in the work areas ranged from 0.61 to 30 1.32 ppm. Urine concentrates were tested at 50 and 100  $\mu$ L per plate. There was no

1	difference in mutagenicity between the autopsy service workers and the control group.
2	The only samples that demonstrated substantial levels of mutagenicity were from two
3	individuals in the control group. One of these had received metronidazole therapy during
4	the study and was not included in the final analysis. The other individual was a heavy
5	smoker (2 packs a day). Urine samples from this individual contained the mutagenic
6	compound 2-naphthylamine. In addition, urine from two individuals in the exposed group
7	(both smokers) showed slight mutagenic responses when assayed in strain TA98 with the
8	addition of S9. However, there was a significant difference (P value was not reported) in
9	the number of urine samples from the exposed group (13) that were toxic compared with
10	the control group (4) (Table 5-24). Toxicity (determined by plates with a partial or
11	complete absence of a background lawn) was reduced in the presence of S9, and when
12	the urine samples were tested at lower concentrations, no mutagenicity was observed.
13	Analyses of the toxic samples showed that most of them contained a compound identified

14 as a glucuronide conjugate that did not appear to be related to formaldehyde exposure.

 Table 5-24. Distribution of autopsy service and pathology department workers with mutagenic or toxic urine samples

Experimental group	Non- mutagenic or non-toxic	Mutagenic	Toxic	Totals
Control	$16 (42)^{a}$	1 (3) <sup>b</sup>	2 (4)	19 (49)
Exposed	11 (27)	$2(5)^{b,c}$	5 (13)	18 (45)
Total	27 (69)	3 (8)	8 (17)	37 (94)

Source: Connor et al. 1985a.

<sup>a</sup> The number in parentheses is the total number of samples in each category.

<sup>b</sup> All mutagenic samples are from smokers.

<sup>c</sup> Both individuals were smokers; urine from both was slightly mutagenic in strain TA98, but only with S9 metabolic activation; urine was not mutagenic in strain TA100 with or without S9.

## 15 5.6.4.3 Cytogenetic effects

- 16 A number of studies have examined the cytogenetic effects of formaldehyde exposure in
- 17 peripheral blood lymphocytes or nasal mucosa in humans exposed to formaldehyde. The
- 18 findings are discussed below and summarized in Table 5-25 (chromosomal aberrations)
- 19 Table 5-26 (SCE) and Table 5-27 (micronuclei).
- 20 Genotoxicity studies published on peripheral lymphocytes in Chinese workers exposed to
- 21 formaldehyde were reviewed by Tang *et al.* (2009). Increases in micronucleus

frequencies in lymphocytes were reported for exposures over 1 year (Wang *et al.* 1997,
Yu *et al.* 2005) and in nasal epithelial cells after 8 weeks exposure to high levels (0.508
to 0.985 mg/m<sup>3</sup>) of formaldehyde (Cheng *et al.* 1995). Also, multiple chromosome
aberrations were reported in workers exposed to an average of 2.51 mg/m<sup>3</sup> of
formaldehyde for 10.5 years (Jin and Zhu 1992). In contrast, two studies reported no
increase in SCE in lymphocytes from formaldehyde-exposed workers (Jin and Zhu 1992,
Li *et al.* 1988, Ye *et al.* 2005). [These findings are not discussed in detail in the text or

8 the tables since the information comes from a secondary source.]

9 Fleig et al. (1982) conducted a cytogenetic analysis of 15 employees at a formaldehyde 10 manufacturing and processing facility in Germany. The workers had been employed for 11 23 to 35 years. The control group included 15 administrative or office staff employees at 12 the same facility who were matched by age and sex with the exposed group. Personal air 13 samplers were used to determine 8-hour time-weighted average formaldehyde exposures 14 for each individual. Mean formaldehyde concentrations at the work areas did not exceed 15 the maximum workplace concentrations (MAK value). MAK values were 5 ppm before 16 1971 and 1 ppm after 1971. Chromosomal aberrations were measured in peripheral blood 17 lymphocytes. One hundred (100) cells per individual were scored. There was no 18 difference in the incidences of aberrant cells including gaps (all types of aberrations with 19 both chromatid and isochromatid gaps between the exposed (3.07%) and control group 20 (3.33%). The mean incidence of aberrant cells excluding gaps (breaks, fragments, 21 deletions, chromatid exchanges, rings, and dicentric chromosomes) was greater in the 22 exposed group than in the controls (1.67 % versus 1.07%); however, this difference was 23 not statistically significant. There was no correlation between formaldehyde exposure and 24 the number of aberrant metaphases. The authors reported that chromosomal aberrations 25 were not increased among smokers.

26 Suskov and Sasanova (1982) examined peripheral lymphocytes from 31 persons,

27 including individuals of both sexes, exposed to formaldehyde in the air at 0.5 mg/m<sup>3</sup>

28 [0.41 ppm], the average concentration in an area in which phenolformaldehyde resin was

29 produced. The control group included 74 healthy individuals that had no occupational

30 contact with synthetic resins. The control group was matched for sex, smoking, alcohol

1 consumption, and medication. The average frequency of metaphases with chromosomal

2 aberrations was 5.0% for the exposed workers and 2.4% for the control group, which was

3 significant at P < 0.001 by  $\chi^2$  test. No difference in the average frequency of

4 chromosome breaks per chromosome was found.

5 Thomson *et al.* (1984) examined incidences of chromosomal aberrations and SCE [results

6 for SCE reported below] in the peripheral blood lymphocytes of six pathology workers

7 and five unexposed controls. Smoking history was obtained for each individual. The

8 pathology workers had been employed for 4 to 11 years and were exposed to

9 formaldehyde for 2 to 4 hours/day, 2 to 3 days/week. Time-weighted average

10 formaldehyde concentrations ranged from 1.14 to  $6.93 \text{ mg/m}^3$  [0.93 to 5.65 ppm]. One

11 hundred (100) first-division metaphases from each 48-hour culture were scored for

12 chromosomal aberrations for each individual. There were no significant differences in the

13 incidences of chromosomal aberrations between the exposed and control groups. The

14 most common chromosomal aberrations were aneuploid cells (36 in the exposed group

15 and 15 in the controls) and chromatid aberrations (8 in the exposed group and 6 in the

16 controls). Only one dicentric chromosome was observed, and this was from the control

17 group. [Although smoking history data were collected, there was no discussion of how

18 these data were used.]

19 Bauchinger and Schmid (1985) investigated the clastogenic effects of formaldehyde in 20 paper factory workers. Chromosomes were analyzed in peripheral blood lymphocytes 21 from 20 male papermakers who had occupational exposure to formaldehyde for 2 to 30 22 years. The control group included 20 male workers from the same factory that were not 23 exposed to formaldehyde. The exposed and control groups were matched for age, 24 smoking history, and social environment. The mean accumulated exposure time was 25 estimated to be about 45 to 90 minutes per 8-hour shift. Formaldehyde concentrations in 26 workroom air did not exceed 0.2 ppm; however, workers were required to enter the paper 27 machine for short periods to take samples or change the paper type, and formaldehyde 28 concentrations as high as 3 ppm were encountered. Five hundred (500) cells per 29 individual were scored for chromosomal aberrations, and 50 cells per individual were 30 scored for SCEs from 54-hour cultures [results for SCE are reported below]. The Mann1 Whitney rank *U* test was used to compare incidences of chromosomal changes.

2 Incidences of dicentrics or dicentrics and ring chromosomes were significantly higher

3 than in controls; however, there were no significant differences in structural chromosome

4 changes, acentric fragments, chromatid-type aberrations, or gaps. Stratified analyses by

5 supervisors and operators showed that only supervisors (mean occupational exposure 2.5

6 times higher than operators) had significantly higher incidence of dicentrics and dicentric

7 and ring chromosomes.

8 Chebotarev *et al.*  $(1986)^2$  reported a significantly higher level of chromosomal

9 aberrations in lymphocytes from 40 woodworkers (2.76%) compared with 22 control

10 workers (1.64%). The incidence of chromosomal breakage was also significantly higher

11 in woodworkers compared with controls (2.95% vs. 1.64%).

12 Vargová *et al.* (1992) compared chromosomal aberrations in peripheral blood

13 lymphocytes from 20 workers (10 men and 10 women) exposed to formaldehyde in a

14 wood-product manufacturing facility with 19 matched non-exposed workers from the

15 same factory. The control and exposed groups had similar habits and a similar social

16 status. The exposed workers had been employed at the facility for 5 to more than 16 years

and were exposed to time-weighted average formaldehyde concentrations of 0.55 to

18  $10.36 \text{ mg/m}^3$  (0.46 to 8.6 ppm). There were no significant differences between the

19 exposed workers and controls for chromatid and chromosome gaps, breaks, exchanges,

20 breaks per cell, or percentage of cells with aberrations. The exposed workers had 3.08%

aberrant cells and 0.045 breaks per cell compared with 3.6% aberrant cells and 0.08

22 breaks per cell in the control group. The authors noted that the frequency of aberrations in

the control group was higher than reported in the general population (1.2% to 2%) and

24 noted that smoking and alcohol consumption may have been a factor. The authors

25 concluded that both the exposed and control groups had a potential increased genotoxic

26 risk, but they had no explanation for the increased levels of chromosomal aberrations in

- 27 the control group. Both controls and the exposed groups had increased numbers of
- 28 inactive lymphocytes and decreased lymphoblast frequency, and exposed groups had a

<sup>&</sup>lt;sup>2</sup> Russian publication, information based on the English summary.

1 significant decrease in the mitotic index. Significant differences in immunological effects

2 were also found between the exposed group and the matched controls and the matched

3 controls and background controls (see Section 5.4.2).

4 Kitaeva et al. (1996) reported a significant increase in the frequency of chromosomal

5 aberrations in peripheral blood lymphocytes of workers at a nitrogen fertilizer

6 manufacturing plant who were exposed to formaldehyde concentrations above the

7 maximum permissible occupational limits (see Table 15-25).

8 Vasudeva and Anand (1996) compared chromosomal aberrations in peripheral blood
9 lymphocytes from 30 female medical students, who were exposed to formaldehyde for 15
10 months during an anatomy laboratory, to 30 age-matched, unexposed controls (non11 medical students). All participants were healthy, had unremarkable medical histories, and

12 had received no or insignificant radiation exposure. The average exposure concentration

13 was less than 1 ppm. The incidences of chromosomal aberrations were not significantly

14 different between the exposed and control groups.

15 He et al. (1998) examined the clastogenic effects of formaldehyde exposure in 13 16 students during a 12-week anatomy class. The control group included 10 students from 17 the same school who were not exposed to formaldehyde. All participants were 18 nonsmokers, and the sex and age of the two groups were similar. Breathing-zone air 19 samples were collected during dissection procedures and showed a mean formaldehyde 20 concentration of 2.37 ppm. Lymphocytes were examined for chromosomal aberrations, 21 SCE, and micronuclei. [Results for SCE and micronuclei are reported below.] 22 Chromosomal aberrations occurred at a significantly higher frequency in the exposed 23 group than in the controls (P < 0.01, [statistical method not identified]). The authors also 24 reported a correlation between micronuclei and chromosomal aberrations. 25 Lazutka et al. (1999) evaluated chromosomal aberrations among 97 (34 male and 63 female) plasticware workers who were exposed to formaldehyde (0.5 to  $0.9 \text{ mg/m}^3$ ), 26 styrene (4.4 to 6.2 mg/m<sup>3</sup>), and phenol (0.5 to 0.75 mg/m<sup>3</sup>) for 2 months to 25 years. 27 28 Non-exposed donors were used as controls (64 male and 26 females) and were matched

29 by age and similar smoking habits as the exposed workers. The mean frequency of

chromosomal aberrations was significantly higher in the exposed workers than controls.
 Significant increases in chromosomal aberrations were observed among workers with
 short and long exposures; however, the frequency of chromosomal aberrations induced
 did not increase with exposure duration. The study was not able to identify which
 exposure caused the chromosomal aberrations; however, the authors noted that styrene
 has been reported to cause chromosomal aberrations.

7 Neri et al. (2006) addressed some of the critical issues of environmental research in 8 pediatric populations. Data from several field studies that were focused on various 9 exposures in children were reviewed. One of these studies evaluated the frequency of 10 chromosomal aberrations in pre-school children (boys and girls, aged 5 to 6 years) and 11 elementary school boys (aged 8 to 12 years) from 1984 to 1986. These children were 12 exposed to elevated levels of formaldehyde from an adhesive that was used to secure 13 pressboard panels in prefabricated schools in Czechoslovakia in the 1980s. Formaldehyde concentrations in the elementary school were  $0.32 \text{ mg/m}^3$  [0.26 ppm] in 1984, 0.13 14  $mg/m^{3}$  [0.11 ppm] in 1985, and 0.037  $mg/m^{3}$  [0.03 ppm] in 1986. Formaldehyde 15 16 concentrations in the pre-school were reported as 0.21 to 0.36 mg/m<sup>3</sup> [0.17 to 0.29 ppm] in 1984. Chromosomal aberrations were determined in lymphocytes from 20 elementary 17 18 school children in 1984, 16 in 1985, and 18 in 1986 and in 13 pre-school children in 19 1984. The control groups included 17 elementary school children in 1984 and 1985 and 20 24 pre-school children in 1984. There were significantly increased percentages of 21 aberrant cells in 1984 and 1985 in the elementary school children compared with the 22 controls (P < 0.01, [statistical method not reported]).

		No. cells	Ex	posure	Aberrant			
Study population	Ν	person	ppm	duration	cells (%)	Comments	Reference	
Matched controls					3.33	Controls matched for age and sex		
Formaldehyde workers	15	100	0	23–35 vr	$(1.07)^{a}$	CA not increased for smokers	Fleig et al. 1982	
	15	100	< 5		3.07			
					(1.67)			
Matched controls	74	93	0		2.4	Controls matched for sex, smoking,	Suskov and Sazonova	
Phenolformaldehyde resin workers	31	104	0.41	0.33–30 yr	5.0***	alconol consumption and medication	1982	
Controls	_		0		b	Controls consisted of 3 females and 2 males, mean age 27.8; exposed consisted		
Controls	5	100	0.9–	4–11 yr	[4.6] <sup>b</sup>	of 2 females and 4 males, mean age 33.5.	Thomson et al. 1984	
Pathology workers	6	100	>9	-	[/./]*	Smoking histories collected but analyses (if any) not reported		
						Controls from the same factory were matched for age, smoking history and social environment.		
Matched controls	20	500	0	<b>a</b> aa	0.0005°	Stratified analyses by supervisors and	Bauchinger and	
Papermakers	20	500	0.2–3	2-30 yr	0.0013* <sup>c</sup>	operators showed that only supervisors (mean occucptional exposure 2.5 times higher than operators) had significantly higher incidence of dicentrics and dicentric and ring chromosomes.	Schmid 1985	
Controls	22	100	NR <sup>d</sup>	NR <sup>d</sup>	1.64		Chebotarev at al. 1086	
Woodworkers	40	100	1111		2.76*			

## Table 5-25. Chromosomal aberrations in peripheral blood lymphocytes from humans exposed to formaldehyde

		No. cells	Ex	posure	Aborrant		
Study population	N	person	ppm	duration	cells (%)	Comments	Reference
Matched controls	19	100	0	5 > 16 yr	3.60 <sup>d</sup>	Controls from same plant with similar habits and social status Authors stated that smoking and alcohol may have influenced findings, but no data	Vargova et al. 1992
wood-splinter product workers	20	100	0.46– 8.6	<i>J=&gt;</i> 10 yr	3.08	CA frequency in controls exceed the general population, and immunological effects were observed in both control and exposed groups.	Vargova <i>et al</i> . 1992
Controls Nitrogen fertilizer workers	6 8	NR NR	0 1.2- 2.4 ml/m 3	10 yr	1.8 5.4*	Controls were 6 individuals and workers 5 women and 10 men; groups were combined because there was no correlation between exposure and age, sex or length of service.	Kitaeva <i>et al</i> . 1996
						62% of aberrations were chromosomal.	
Matched controls Medical students	30 30	100 100	0 <1	15 mo	0.9 1.2	Controls were non-medical students matched on age.	Vasudeva and Anand 1996
Controls Anatomy class students	10 13	100 100	0 2.37	12 wk	3.4 5.9**	All students were non-smokers and had similar sex and age distributions.	He et al. 1998
Controls (donors) Plasticware workers	90 97	100 100	0.5– 0.9 mg/m	2 mo to 25 yr	1.68 4.2*	Controls matched on age, and had similar smoking habits; however most of the workers were females and most of the controls were males. Workers also exposed to styrene and phenol CA frequency did not increased with increasing duration of exposure	Lazutka <i>et al</i> . 1999

		No. cells	Ex	posure	Aborrant		
Study population	N	person	ppm	duration	cells (%)	Comments	Reference
Controls (1984) School children (1984) School children (1985) School children (1986)	17 20 16 18	100 100 100 100	0 0.26 0.11 0.03	1–3 yr	1.37 4.71** 2.83** 2.06	Children were exposed to formaldehyde from adhesive used to secure pressboard panels in prefabricated schools.	Neri <i>et al.</i> 2006
Controls (preschool, 1984) Preschool children (1984)	24 13	100 100	0 0.17- 0.3		1.12 2.40		

\* = P < 0.05, \*\* = P < 0.01, \*\*\* = P < 0.001.

CA = chromosomal aberrations, NR = not reported, NS = not significant compared with controls.

<sup>a</sup>Data reported for aberrant cells including gaps and excluding gaps (in parenthesis). <sup>b</sup>Frequencies were calculated from the totals for aneuploid cells, Cs cells, acentrics, dicentrics, rings, and chromatid aberrations. <sup>c</sup>Data are mean frequencies of dicentrics/cell. The frequency of dicentrics combined with ring chromosomes was also significantly different from controls. No significant differences were observed for structural chromosome changes, acentric fragments, gaps/cells, or chromatid-type aberrations.

<sup>d</sup> Exceeded the frequency of aberrations (1.2% to 2%) reported in the general population.

1 Occupational exposure to formaldehyde and SCE were evaluated in 11 studies. Three of 2 the earliest published studies (discussed above) did not find increased incidences of SCEs 3 among workers exposed to formaldehyde (Thomson et al. 1984, Bauchinger and Schmid 4 1985, Chebotarev et al. 1986). Thompson et al. examined incidences of SCE in the 5 peripheral blood lymphocytes of six pathology workers and five unexposed controls. 6 Bauchinger and Schmid (1985) studied 20 male paper factory workers who were 7 occupationally exposed to formaldehyde for 2 to 30 years, and Chebotarev et al. studied 8 40 woodworkers.

9 Yager et al. (1986) measured SCEs in the peripheral lymphocytes of eight non-smokers 10 exposed to formaldehyde embalming solution during a 10-week anatomy class. The 11 embalming fluid contained 5.6% formalin (37% formaldehyde and 15% methanol), 12 22.4% ethanol, 10% phenol, and 62% water. The class met two afternoons per week, but 13 students had free access to the laboratory throughout the week. None of the participants 14 had any known exposure to formaldehyde during the preceding year. Blood samples were 15 collected before, and at the end of the class. The mean concentration of formaldehyde in 16 the classroom air was 0.33 ppm, while the mean concentration from breathing zone 17 samples collected during dissection procedures was 1.2 ppm. The mean number of SCEs per cell increased from  $6.39 \pm 0.11$  before taking the class to  $7.2 \pm 0.33$  at the end of the 18 19 class. The increase was statistically significant (P = 0.02, paired *t*-test).

20 Suruda *et al.* (1993) examined SCEs in lymphocytes in mortuary science students

21 following low-level formaldehyde exposure during an embalming class. The students

22 performed an average of 6.9 embalmings (range 2 to 15) during the 85-day study period.

23 However, several of the students lived at funeral homes or had part-time jobs in funeral

24 homes, and participated in embalmings outside the class. Mean formaldehyde

concentrations measured during embalming ranged from 0.15 to 4.3 ppm with peak

26 concentrations as high as 6.6 ppm. The calculated 8-hour time-weighted average

formaldehyde concentration ranged from 0.1 to 0.96 ppm with an overall mean of 0.33

28 ppm. Furthermore, air sample measurements indicated little to no exposure to chemicals

29 other than formaldehyde. SCE frequency showed a significant decrease (7.5%, P < 0.05,

1 Student's *t*-test) compared with baseline values. No association was observed with

2 cumulative exposure to formaldehyde and SCE frequency.

3 Shaham et al. (1997) evaluated the frequency of SCE in peripheral blood lymphocytes in 4 13 workers (6 physicians and 7 technicians) at the Pathology Institute who were 5 occupationally exposed to formaldehyde compared with 20 unexposed, age-matched 6 controls [sex not reported]. There were 3 smokers in the exposed group (23%) and 6 7 smokers in the control group (30%). The workers had been occupationally exposed to 8 formaldehyde for 2 to 25 years (mean of 13 years). No past exposures to other mutagenic 9 agents were identified. Formaldehyde concentrations were measured in ambient air at 10 various periods throughout the day and ranged from 1.4 to 1.6 ppm in the rooms of the 11 Pathology Institute. Personal samples collected while work was in progress resulted in 12 slightly higher concentrations (2.8 to 3.1 ppm). There was a significant difference in the 13 mean number of SCEs per chromosome in the exposed workers compared with controls 14  $(0.212 \pm 0.039 \text{ [mean \pm SD]} \text{ vs. } 0.188 \pm 0.035; P = 0.05, t-\text{test})$ . Significant differences 15 remained after adjustment for smoking. There was a linear relationship between years of 16 exposure and the number of SCE.

17 Ying et al. (1999) examined SCE frequency in lymphocytes of 23 students (11 males and 18 12 females) enrolled in an anatomy class for 8 weeks. Each student served as their own 19 control and none of the students were smokers. Formaldehyde concentrations were 20 measured in the anatomy laboratory as well as the student's dormitories. The 3-hour time-weighted average formaldehyde concentrations were  $0.51 \pm 0.3$  mg/m<sup>3</sup> [0.41 ± 0.24] 21 ppm] in the anatomy laboratory and  $0.012 \pm 0.0025 \text{ mg/m}^3 [0.01 \pm 0.002 \text{ ppm}]$  in the 22 23 dormitories. There was no significant difference in SCE frequency in lymphocytes before 24 and after completing the 8-week anatomy course. (See Section 5.4.2.4) for lymphocyte 25 subset analyses)

He *et al.* (1998) reported that there was a statistically significant increase (P < 0.05,

27 [statistical method not identified]) in SCE frequency in 13 students exposed to

formaldehyde during a 12-week anatomy class compared with a control group of 10

29 students from the same school who were not exposed to formaldehyde. All participants

1	were nonsmokers, and the sex and age of the two groups were similar. Breathing-zone air
2	samples were collected during dissection procedures and showed a mean formaldehyde
3	concentration of 2.37 ppm. (This study also evaluated chromosomal aberrations.)
4	Shaham et al. (2002) investigated the mean number of SCEs per chromosome and the
5	proportion of high frequency cells (HFC, i.e., cells with more than eight SCEs) in the
6	peripheral lymphocytes of 90 workers (25 males and 65 females, mean age $44.2 \pm 8.5$
7	years) from 14 hospital pathology departments in Israel. The control group included 52
8	unexposed workers (44 males and 8 females, mean age $41.7 \pm 11.4$ ) from the
9	administrative staff of the same hospitals. The percent of active smokers was somewhat
10	higher ( $P > 0.05$ ) in the control group (46.9%) than the exposed group (34.4%).
11	Differences between the controls and exposed groups were (1) sex, higher percentage of
12	females in the exposed ( $P < 0.01$ ), (2) origin, higher number of workers with
13	European/American origin in the exposed ( $P < 0.05$ ) and (3) education, higher level of
14	education in the exposed ( $P = 0.06$ ). The mean exposure period was 15.4 years (range 1
15	to 39 years). No one in the exposed group was known to have been occupationally
16	exposed to other genotoxic substances, and no one in the control group was known to
17	have ever been occupationally exposed to formaldehyde. The exposed group was further
18	divided into a low-exposure group (formaldehyde concentrations of 0.04 to 0.7 ppm) and
19	a high-exposure group (formaldehyde concentrations of 0.72 to 5.6 ppm) based on
20	personal and field samples of ambient air in the pathology departments at various times
21	during the typical work day. The low-exposure group primarily included laboratory
22	assistants and technicians and the high-exposure group primarily included physicians and
23	hospital orderlies. Adjustments were made for sex, smoking habits, education, and
24	national origin (age was introduced in the model but it did not correlate with SCE
25	measures). Both measures of SCEs (SCE per chromosome and proportion of HFC) were
26	significantly higher in the exposed compared with the control group ( $P < 0.01$ , Mann-
27	Whitney test), and were significantly higher among workers with 15 years of exposure
28	compared with workers with less than 15 years of exposure ( $P < 0.05$ ). There were no
29	significant differences between the low- and high-exposure groups; however, among
30	smokers, both variables of SCE were higher in the high-exposure subgroup.

1 Ye et al. (2005) examined nasal mucosa cells and lymphocytes in two populations of 2 formaldehyde-exposed workers in China. One group of 18 workers (11 males and 7 3 females) was exposed in a formaldehyde manufacturing facility. The mean length of 4 employment was 8.5 years (range 1 to 15 years). The second group included 16 waiters 5 (4 males and 12 females) who worked in a newly fitted ballroom for 12 weeks and were 6 exposed to low levels of formaldehyde from building material, tobacco smoke and 7 furniture. The control group included 23 college students (12 males and 11 females). The 8 average ages in each of the groups were: manufacturing workers, 29 years (range 19 to 9 39); waiters, 22 years (range 19 to 27); and students, 19 years (range 18 to 23). The 8-10 hour time-weighted average formaldehyde concentration in the formaldehyde factory was 11  $0.99 \text{ mg/m}^3$  [0.8 ppm]. The 5-hour time-weighted average concentration measured in the ballroom was 0.11 mg/m<sup>3</sup> [0.09 ppm]. A background indoor air concentration of 0.011 12 13  $mg/m^3$  [0.009 ppm] was measured in the student dormitories. All study participants were 14 nonsmokers. The workers, but not the waiters, had a significantly increased frequency of 15 SCEs in lymphocytes compared with the controls (P < 0.05, one-way ANOVA). (See 16 Section 5.4.2.4 for lymphocyte subset analyses).

17 Costa et al. (2008) investigated DNA damage (see Section 5.6.4.2), SCE, and 18 micronuclei (results reported below) in 30 workers exposed to formaldehyde in four 19 hospital pathology anatomy laboratories in Portugal. Thirty non-exposed hospital 20 employees (matched by age, gender, lifestyle, and smoking) served as the control group. 21 Formaldehyde concentrations measured in the breathing zone of the laboratory workers 22 averaged 0.44 ppm. SCE values were significantly higher in the exposed group (P < 0.05) 23 compared with the control group. There was no association between SCE values and 24 genetic polymorphisms in genes involved with xenobiotic metabolism or DNA repair or 25 with duration of exposure. SCE frequency was higher among control smokers than non-26 smokers but no differences were observed in the exposed groups. Age and sex did not 27 affect the observed SCE frequency.

		No. cells	Exp	osure	SCE frequency/cell		
Study population	N	examined/ person	ppm	duration	(± SE)	Comments	Reference
Controls Pathology workers	5 6	50	0 0.9–>9	4–11 yr	$6.44 \pm 0.38$ $6.78 \pm 0.31$	Controls consisted of 3 females and 2 males, mean age 27.8 and exposed consisted of 2 females and 4 males, mean age 33.5. Smoking histories collected but analyses (if any) not reported	Thomson <i>et al.</i> 1984
Matched controls Papermakers	20 20	50	0 0.2–3	2–30 yr	$9.53 \pm 0.35 \\ 8.87 \pm 0.24$	Controls from the same factory and were matched for age, smoking history and social environment.	Bauchinger and Schmid 1985
Controls Woodworkers	22 40	NR <sup>a</sup>	NR <sup>a</sup>	NR <sup>a</sup>	$\begin{array}{c} 8.24 \pm 0.37 \\ 8.01 \pm 0.24 \end{array}$		Chebotarev <i>et al.</i> 1986
Anatomy class students Pre-exposure Post-exposure	8	80	1.2	10 wk	$6.39 \pm 0.11$ $7.20 \pm 0.33^*$	All students were non-smokers	Yager <i>et al.</i> 1986
Mortuary science students Pre-exposure Post-exposure	29 <sup>b</sup>	50	0.1– 0.96	85 d	$\begin{array}{c} 7.72 \pm 0.13 \\ 7.14 \pm 0.89^{b} \end{array}$	Several students had part time jobs involving formaldehyde exposure No association was observed with cumulative exposure to formaldehyde	Suruda <i>et al.</i> 1993
Matched controls Physicians and technicians	20 13	32 28	0 1.4–3.1	13 yr	$\begin{array}{c} 0.186 \pm 0.035^{c} \\ 0.212 \pm 0.039^{*c} \end{array}$	Controls matched on age; 3 (23%) smokers in exposed group, and 6 (30%) in control Significant differences remained after adjustment for smoking Linear relationship between years of exposure and SCE	Shaham <i>et al.</i> 1997
Anatomy class students	23 <sup>b</sup>	30	0.01-	8 wk	$6.38 \pm 0.41$	All students were non-smokers without	Ying et al. 1999

# Table 5-26. Sister chromatid exchange in peripheral blood lymphocytes from humans exposed to formaldehyde

\_\_\_\_\_

		No. cells	Exposure		SCE frequency/cell		
Study population	N	examined/ person	ppm	duration	(± SE)	Comments	Reference
Pre-exposure Post-exposure			0.4		$6.61\pm0.79$	exposure to x-ray (6 months)	
Controls Anatomy class students	10 13	25	0 2.37	12 wk	$5.26 \pm 0.51$ $5.91 \pm 0.71*$	All students were non-smokers and control and exposed groups had similar sex and age distributions	He et al. 1998
Controls Hospital pathology staff	52 90	30-31 30-32	0 0.04– 5.6	1–39 yr	$0.19 \pm 0.004$ $0.27 \pm 0.003*$	Controls were similar in age, but signfiicant differences were found for sex, and level of education. Non- signficant differences were found for active smokers and place of origin. Analyses were adjusted for smoking, sex, education, and origin. Higher SCE were found among those with longer exposure duration but not among workers with higher level of exposure	Shaham <i>et al.</i> 2002
Controls Formaldehyde factory workers Waiters	23 18 16	30	0.009 0.8 0.09	1–15 yr	$6.38 \pm 0.41$ $8.24 \pm 0.89*$ $\sim 6^{d}$	All subjects were non-smokers and had similar ages (average ages were 19 for controls, 22 for waiters and 29 for formaldehyde workers).	Ye <i>et al.</i> 2005
Matched controls Pathology/anatomy lab workers	30 30	50	0 0.44	0.5–27 yr	$4.49 \pm 0.16$ $6.13 \pm 0.29*$	Controls were matched by age, sex, lifestyle factors and smoking habits. Age and sex did not effect SCE; higher SCE were seen in control unexposed smokers than control unexposed non-smokers.	Costa <i>et al.</i> 2008

		No. cells	Exp	osure	SCE frequency/cell		
Study population	N	examined/ person	ppm	duration	(± SE)	Comments	Reference
						No association was observed with exposure duration	

\* = P < 0.05, \*\* = P < 0.01, \*\*\* = P < 0.001 compared with controls. <sup>a</sup> Not reported in the English summary of a Russian publication. <sup>b</sup> Significant decrease in post-exposure samples compared to baseline values. <sup>c</sup> Data are SCEs per chromosome ± SD. <sup>e</sup> Value was estimated from a figure (exact value was not provided by the study authors).

1 Ballarin et al. (1992) reported an increase in micronuclei in plywood factory workers 2 compared with an age- and sex-matched control group, who were university or hospital 3 workers. All subjects were non-smokers. The exposed group included 15 workers 4 employed at the plywood factory for 1.5 to 19 years (mean 6.8 years), 7 of which worked 5 in the warehouse, 6 in the shearing-pressing department, and 2 in the sawmill. The timeweighted average formaldehyde concentrations were about  $0.1 \text{ mg/m}^3$  [0.08 ppm] in the 6 7 sawmill and shearing press and 0.39 mg/m<sup>3</sup> [0.32 ppm] in the warehouse. The highest concentration of 0.6 mg/m<sup>3</sup> [0.5 ppm] was recorded in the warehouse. Wood dust levels 8 also were measured and ranged from about 0.23 mg/m<sup>3</sup> to 0.73 mg/m<sup>3</sup>. Respiratory nasal 9 10 mucosa cells were scraped from the inner turbinates and examined for micronuclei. No 11 fewer than 6,000 cells were counted for each slide. The frequency of micronucleated cells 12 was significantly higher in the exposed group compared with controls  $(0.90 \pm 0.47 \text{ vs.})$ 13  $0.25 \pm 0.22$ , P < 0.01, Mann-Whitney U test). No significant difference in micronuclei 14 frequency wasfound between workers in the warehouse  $(0.97 \pm 0.39)$  and the sawmill and 15 shearing-pressing departments  $(0.74 \pm 0.53)$ .

16 Two studies (Suruda et al. 1993, Titenko-Holland et al. 1996) examined micronuclei in 17 buccal cells, nasal epithelial cells, and/or lymphocytes in mortuary science students 18 following low-level formaldehyde exposure during an embalming class. Titenko-Holland 19 et al. (1996) used previously unstained and unanalyzed slides collected from participants 20 in the Suruda et al. (1993) study, and used fluorescence in situ hybridization (FISH) 21 rather than a staining method to detect micronuclei. The results of the two studies were 22 similar. Suruda et al. reported that post-exposure micronucleus frequencies increased 23 significantly in buccal epithelial cells and lymphocytes compared with baseline values (P 24 < 0.05, Wilcoxon sign-rank test). A significant dose-response relationship was reported 25 for increases in buccal micronuclei (but not nasal or lymphocyte micronulei) in the 22 26 male subjects but not in the 7 female subjects. There was a non-significant increase in 27 nasal epithelial micronucleus frequency. Titenko-Holland et al. (1996) reported that there 28 was a significant increase in micronucleus frequency in buccal cells (P = 0.007, 29 Wilcoxon sign-rank test) but not in nasal epithelial cells. Total buccal micronuclei were 30 weakly associated (r = 0.44, P = 0.06) with cumulative exposure to embalming fluid (90)

1 days). In both tissues, a higher increase in centromere-negative micronuclei (9-fold, P =2 0.005 for buccal cells; 2-fold, P = 0.03 for nasal cells) was found than for centromere-3 positive micronuclei (> 2-fold, P = 0.08 for buccal cells; no change, P = 0.31 for nasal 4 cells), suggesting that the primary mechanism of micronucleus formation appeared to be 5 chromosome breakage.

Kitaeva *et al.* (1996) reported a higher sensitivity to formaldehyde exposure for females
than males in a study of micronucleus induction in buccal epithelium. There was an

8 increased frequency (P < 0.05) of micronuclei reported in buccal mucosa cells collected

9 from 8 female but not from 5 male anatomy workers. However, there were significant

10 increases in both female (P < 0.01) and male (P < 0.05) students (6 female and 6 male)

11 exposed for 40 minutes. The number of micronucleated cells detected in the students

12 remained elevated 48 hours after the class.

13 Ying et al. (1997) examined the changes in the frequency of micronuclei in the nasal 14 mucosa, oral mucosa, and lymphocytes of 25 students (13 males and 12 females) enrolled 15 in an anatomy class for 8 weeks. Each student served as their own control; none of the 16 students were smokers, or had a history of drug use in the last 3 weeks or X-rays in the 17 last 6 months. Formaldehyde concentrations were measured in the anatomy laboratory as 18 well as the student's dormitories. The 3-hour time-weighted average formaldehyde concentrations were  $0.51 \pm 0.3 \text{ mg/m}^3$  [0.41 ± 0.24 ppm] in the anatomy laboratory and 19  $0.012 \pm 0.0025 \text{ mg/m}^3$  [0.01  $\pm 0.002 \text{ ppm}$ ] in the dormitories. There was a significantly 20 21 higher frequency of micronuclei in nasal and oral mucosal cells after exposure to 22 formaldehyde (P < 0.001, paired *t*-test). There was no significant difference in the 23 frequency of micronuclei in lymphocytes.

He *et al.* (1998) examined the frequency of chromosomal aberrations, SCE (see above), and micronuclei in peripheral blood lymphocytes in 13 students during a 12-week anatomy class. The control group included 10 students from the same school who were not exposed to formaldehyde. All participants were nonsmokers, and the sex and age of the two groups were similar. Micronuclei occurred at a significantly higher frequency in

1 the exposed group than in the controls (P < 0.01, [statistical method not identified]). The 2 authors also reported a correlation between micronuclei and chromosomal aberrations. 3 Burgaz et al. (2001, 2002) reported the frequency of micronuclei in nasal and buccal 4 mucosa cells in individuals exposed to formaldehyde in pathology and anatomy 5 laboratories. The first study examined cells from the nasal mucosa and included 23 6 pathology or anatomy department staff (11 females and 12 males) and a control group of 7 25 healthy males selected from university and hospital staff. The numbers of smokers 8 was much higher in the control group (19/25, 75%) compared with the exposed groups. 9 (9/23, 39%), but the workers had similar ages, dietary habits and use of medicine. The 10 second study examined cells from the buccal mucosa and included 28 subjects (15 males 11 and 13 females) who worked in pathology and anatomy laboratories and 18 male 12 volunteer controls who were university staff. Some of the subjects were apparently used 13 in both studies; however, details of the overlap were not provided. None of the referents 14 had been occupationally exposed to genotoxic materials. Workers and controls in the 15 second study reported similar diets, alcohol consumption, smoking habits and use of 16 medications. The formaldehyde concentrations in the laboratories ranged between 2 and 4 17 ppm. Formaldehyde exposure was associated with a statistically significant increase in 18 micronuclei frequency in nasal (P < 0.01, non-parametric statistics) and buccal (P < 0.05, 19 Student's t-test and Mann-Whitney test) mucosa cells. Nasal mucosa micronucleus 20 frequency was significantly higher in exposed smokers compared with control smokers. 21 There was no significant effect of age, sex, smoking status, or exposure duration. 22 Ye et al. (2005) (see discussion under SCE for details) also examined micronucleus 23 formation in nasal mucosa cells from workers at a formaldehyde manufacturing facility 24 and in a group of waiters who worked in a newly fitted ballroom and were exposed to 25 low levels of formaldehyde from building material, tobacco smoke, and furniture. All

27 increased frequency of micronuclei in nasal mucosa cells compared with the controls (*P* 

study participants were nonsmokers. The workers, but not the waiters, had a significantly

28 < 0.05, one-way ANOVA).

26

1 Orsière et al. (2006) also evaluated the effects of formaldehyde on micronucleus 2 formation in lymphocytes in the study of 59 pathology and anatomy laboratory workers 3 and 37 controls described above (see Section 5.6.4.2). Both the control and exposed 4 workers were matched for age, gender, and smoking habits. Chromosomal damage was 5 assessed with the cytokinesis-blocked micronucleus assay. Samples of whole blood were 6 cultured and prepared, then smeared on microscope slides and air dried. The frequency of 7 micronuclei was expressed per 1,000 cells. Micronuclei were measured using the 8 cytokinesis-blocked micronucleus (CMBN) assay. The binucleated micronucleated cell 9 rate (BMCR) was significantly higher in the lymphocytes of exposed workers compared 10 with controls (see Table 5-27). BMCR was correlated with exposure duration in 11 unadjusted analyses, but was no longer significant after controlling for age. Age and 12 gender, but not smoking and drinking habits, were associated with BMCR. 13 The presence of centromeres in the micronuclei was determined using fluorescent 14 hybridization (FISH) and a pan-centromeric DNA probe in combination with the CMBN 15 assay on 18 exposed and 18 controls randomized from the initial population. 16 Micronucleated cells were classified as centromere positive or negative. Centromere-17 positive cells were further classified based on the presence of a single centromere or 18 multiple centromeres. BMCR was statistically higher in the exposed group compared 19 with the controls, and the frequencies of micronuclei and centromere-positive 20 micronuclei were higher (but not statistically significant) in the exposed subjects, 21 however, no increased frequency was found for centromere-negative micronuclei. 22 Monocentromeric micronuclei frequency was significantly higher in the exposed group 23  $(11.0\% \pm 6.2 \text{ versus } 3.1\% \pm 2.4; P < 0.001)$ , but the frequency of micronuclei containing 24 more than one centromere was similar in controls and exposed groups. 25 Iarmarcovai et al. (2007) pooled data from three biomonitoring studies of untreated 26 cancer patients, welders, and the subset of 18 pathologists/anatomists who were exposed 27 to formaldehyde and 18 unexposed controls from the study population reported by 28 Orsière *et al.* (2006). In addition to the findings reported above, they reported the results 29 of multivariate regression analysis that adjusted for age, sex, cigarette smoking, and 30 alcohol consumption, and was weighted for the number of scored cells.

1 Pathologists/anatomists had significantly higher frequency ratios (FR) of centromere-2 positive micronuclei (FR = 1.65, 95% CI = 1.05 to 2.59), and monocentromeric 3 micronuclei (FR = 3.29 (95% CI = 2.04 to 5.30) compared with the controls. In the 4 pooled studies, alcohol drinking and gender affected endpoints measuring aneuploidy 5 (centromere positive micronuclei frequency and monocentromeric micronuclei 6 frequency), and total micronuclei whereas age only affected total micronuclei frequency. 7 Micronuclei were not induced in buccal mucosa cells in a study of healthy volunteers 8 exposed to formaldehyde vapors. In this study by Speit et al. (2007b), 10 women and 11 9 men were divided into 5 groups and exposed to formaldehyde in test chambers 4 hours 10 per day for 10 days. For each group, exposure varied from one day to the next from a 11 constant 0.15 ppp throughout the day, to 0.5 ppp with four peaks of 1.0 ppm for 15 12 minutes each. Exposure also varied daily across groups. The exposure scenarios resulted

prior to the exposure to formaldehyde. Treatment buccal smears were taken following the
10-day exposure and 7, 14 and 21 days afterwards. The authors noted that these results

17 demonstrated that formaldehyde vapors in the range of current Occupational Exposure

in cumulative exposures of 13.5 ppm-hours over the 10 working days. Control buccal

smears were prepared for each subject one week prior to treatment as well as immediately

18 Limits (e.g., 0.5 ppm in Germany and 2.0 ppm in the United Kingdom) did not induce

19 micronuclei in buccal mucosa cells.

13

14

20 Costa *et al.* (2008) reported a significantly higher frequency (P = 0.003) of micronuclei in

21 30 workers exposed to formaldehyde in four hospital pathology anatomy laboratories in

22 Portugal compared with matched controls. Heparinized whole blood was used to establish

23 duplicate lymphocyte cultures for evaluation by the cytokinesis-blocked micronucleus

24 test. Micronuclei were significantly higher in the exposed group compared with the

controls (see Table 5-27), and a positive correlation was found between formaldehyde

26 exposure levels and micronuclei frequency (r = 0.384, P = 0.001). Genetic

27 polymorphisms of xenobiotic metabolizing or DNA repair genes did not show a

28 significant effect. Age, gender and smoking habits were not significantly associated with

29 micronucleus frequency. [This study also evaluated DNA damage and SCE.]

			No colle	Exp	osure	Micronuclei		
Study population	N	Cell type	examined/ person	ppm	duration	00 cells (± SD)	Comments	Reference
Matched controls Plywood factory workers	15 15	Nasal epithelium	6,000	0.07- 0.32	1.5–19 yr	$\begin{array}{c} 0.25 \pm 0.22 \\ 0.90 \pm 0.47^{**} \end{array}$	All subjects were non- smokers. Controls matched for age and sex	Ballarin <i>et al.</i> 1992
Mortuary science students (Pre-exposure and	29	Nasal epithelium	1,500	0.1–0.96	85 d	$\begin{array}{c} 0.41 \pm 0.52 \\ 0.50 \pm 0.67 \end{array}$	Several students had part time jobs involving formaldehyde exposure.	Suruda et al. 1993
post-exposure measurements)		Buccal epithelium	1,500			$\begin{array}{c} 0.046 \pm 0.17 \\ 0.60 \pm 1.27 * \end{array}$	Cumulative exposure to formaldehyde was associated with buccal MN	
		Lymphocytes	2,000			$\begin{array}{c} 4.95 \pm 1.72 \\ 6.36 \pm 2.03 * \end{array}$	among male (22) subjects (r = $0.5$ , $P < 0.01$ ); no association was observed with nasal or lymphocyte MN.	
Mortuary science students (Same participants as	13 <sup>a</sup>	Nasal epithelium	187–5,000	0.1–0.96	90 d	$2 \pm 1.3$ $2.5 \pm 1.3^{b}$	Cumulative exposure and buccal MN ( $r = 0.44, P = 0.06$ )	Titenko-Holland et al. 1996
Suruda <i>et al.</i> 1993)	19 <sup>a</sup>	Buccal epithelium	503–4,113			$0.6 \pm 0.5$ $2.0 \pm 2.0^{**}$		
Anatomy lab workers Controls (all female) Females Males	7 8 5	Buccal epithelium	> 2000	NR°	17 yr	0.64 2.94** 1.18	Controls for students were pre-exposure measures	Kitaeva <i>et al.</i> 1996
Anatomy class students Females (pre-exp.)	6				40 min	0.58		

\_\_\_\_\_

			No colle	Exposure		Micronuclei		
Study population	N	Cell type	examined/ person	ppm	duration	00 cells (± SD)	Comments	Reference
exposed Males (pre-exp.) exposed	6 6 6					2.50** 0.77 2.02*		
Anatomy class students (Pre-exposure and post-exposure measurements)	25 25 <sup>a</sup>	Nasal epithelium Oral epithelium	2,870 2,962 3,167 3,088	0.01–0.4	8 wk	$1.20 \pm 0.0.68$ 3.84 ± 1.5*** 0.57 ± 0.32 0.86 ±	All students were non- smokers, and did not have a history of drug use (3 weeks) or X rays (6 months).	Ying et al. 1997`
	23 <sup>a</sup>	Lymphocytes	4,000 4,000			$0.56^{**}$ $0.91 \pm 0.39$ $1.11 \pm 0.54$		
Controls Anatomy class students	10 13	Lymphocytes	1,000	2.37	12 wk	$\begin{array}{c} 3.15 \pm 0.146 \\ 6.38 \pm 2.5 ** \end{array}$	All students were non- smokers and control and exposed groups had similar sex and age distributions.	He et al. 1998
Controls Pathology/anatomy lab workers	25 23	Nasal epithelium	3,000	2-4	1–13 yr	$\begin{array}{c} 0.61 \pm 0.27 \\ 1.01 \pm \\ 0.62^{**} \end{array}$	Controls and exposed group reported similar ages, dietary habits and medicine use; however, there was a greater number of smokers in the control than in the exposed group.	Burgaz <i>et al.</i> 2001
Controls Pathology/anatomy lab	18 28	Buccal epithelium	3,000	2–4	1–13 yr	$\begin{array}{c} 0.33 \pm 0.30 \\ 0.71 \pm 0.56 * \end{array}$	Control and exposed reported similar diets, alcohol consumption,	Burgaz <i>et al.</i> 2002

			No colle	Exp	osure	Micronuclei		
Study population	N	Cell type	examined/ person	ppm	duration	00 cells (± SD)	Comments	Reference
workers [Study populaton may overlap with that of Burgaz <i>et al.</i> 2001]							smoking habits, and use of medications.	
Controls Formaldehyde factory workers Waiters	23 18 16	Nasal epithelium	3,000	0.009 0.8 0.09	1–15 yr	$\begin{array}{c} 1.25 \pm 0.65 \\ 2.70 \pm 1.50^{*} \\ \sim \!\! 1.9 \pm 1^{d} \end{array}$	smokers and had similar ages (average ages were 19 for controls, 22 for waiters and 29 for formaldehyde workers).	Ye <i>et al.</i> 2005
Matched controls Pathology/anatomy lab workers	37 59	Lymphocytes	1,000	< 0.1– 20.4	0.5–34 yr	11.1 ± 6.0 16.9 ± 9.3*** <sup>ef</sup>	Controls matched for age, sex, and smoking habits Micronuclei were correlated with age and gender but not smoking or drinking habits.	Orsiere <i>et al.</i> 2006
Controls Pathologists/anatomists (randomly chosen from the 37 controls and 59 exposed workers described above)	18 18	Lymphocytes	1,000	0.4–7	NR	11.9 ± 5.6 19.1 ± 10.1*	Controls matched for age, sex, and smoking habits	Orsiere <i>et al.</i> 2006 Iarmarcovai <i>et al.</i> 2007
Controls Volunteer subjects (10 women and 11 men)	21 18	Buccal epithelium	2,000	1.0 peak (with daily variation) max 13.5 ppm-h cum. exp.	10 d	$0.86 \pm 0.84$ $1.33 \pm 1.45$	Subjects served as own controls, measured before first exposure.	Speit <i>et al.</i> 2007b
Controls Pathology/anatomy lab	30 30	Lymphocytes	1,000	0 0.44	0.5–27 yr	$3.27 \pm 0.69 \\ 5.47 \pm$	Controls were matched by age, gender, lifestyle factors	Costa <i>et al</i> . 2008

422

\_\_\_\_\_

			No colle	Ехр	osure	Micronuclei		
Study population	N	Cell type	examined/ person	ppm	duration	00 cells (± SD)	Comments	Reference
workers						0.76**	and smoking habits.	
							MN frequency was significantly associated with formaldehyde exposure levels ( $r = 0.384$ , $P = 0.001$ ) Age, gender and smoking did not affect MN	

\* = P < 0.05; \*\* = P < 0.01; \*\*\* = P < 0.001.

MN = micronuclei; NR = not reported; NS = not significant compared to controls.
 <sup>a</sup> There was a total of 28 subjects in the study but only 19 with complete data for buccal mucosa and 13 with complete data for nasal mucosa were included in the analyses.
 <sup>b</sup> There was a significant increase in centromere-negative micronuclei.
 <sup>c</sup> Exposure considered long-term for workers but no measurements reported for them or for anatomy students.
 <sup>d</sup> Value estimated from a figure.

<sup>e</sup> Binucleated micronuleated cell rate.

<sup>f</sup>Significant increase in centromere-positive micronuclei and monocentromeric micronuclei frequencies.

### 1 5.6.5 Gene expression

2 Kim et al. (2002) investigated the possible role of formaldehyde in sick-building 3 syndrome. These authors reported that formaldehyde increased the surface expressions of 4 intercellular adhesion molecule-1 (ICAM-1) and vascular cell adhesion molecule-1 5 (VCAM-1) on human mucosal microvascular endothelial cells (HMMECs), and 6 enhanced the adhesiveness between these cells and eosinophils. HMMECs were 7 incubated with formal dehyde at concentrations ranging from 1 ng/mL to 1  $\mu$ g/mL for 24 8 hours. There was a statistically significant up-regulation of both ICAM-1 and VCAM-1 9 at 0.1 and 1.0  $\mu$ g/mL. The authors concluded that induction of ICAM-1 and VCAM-1 by 10 formaldehyde might play an important role in allergic inflammation associated with sick 11 building syndrome.

12 Parfett et al. (2003) measured changes in proliferin mRNA over 1 to 3 days in response 13 to various promoters (including formaldehyde) of morphological transformation of 14 C3H/10T1/2 cells. Members of the proliferin protein family are known to influence 15 aspects of cell differentiation or proliferation. Cell cultures were seeded and grown for 2 16 to 4 d before treatment with test compounds. Formaldehyde was added to the cell cultures 17 at 50, 100, or 200  $\mu$ M and incubated for 18 to 20 hours. At 50  $\mu$ M, proliferin mRNA 18 levels were between 5- and 10-fold higher than controls but increased to 40-fold higher 19 than control levels at 100  $\mu$ M. Formaldehyde was thought to be toxic to the cell cultures 20 at 200 µM because induction was reduced to four-fold above control levels.

21 Hester *et al.* (2003) investigated gene expression in the rat nasal respiratory epithelium 22 after exposure to formaldehyde. Groups of male F344 rats received either 40  $\mu$ L of 23 distilled water or 400 mM formaldehyde instilled into each nostril. The rats were killed 24 24 hours later and the nasal epithelium was removed and examined for gene expression. 25 The analysis revealed that 24 of 1,185 genes queried were significantly upregulated and 26 22 genes were downregulated. The identified genes belonged to the functional categories 27 involved in xenobiotic metabolism, cell-cycle control, apoptosis, and DNA repair. Thus 28 multiple pathways are dysregulated by formaldehyde exposure, including those involved 29 in DNA synthesis and repair and regulation of cell proliferation.

1 Hester et al. (2005) compared the effects of formaldehyde and glutaraldehyde in male 2 F344 rats. Groups of rats were exposed to formaldehyde (400 mM) or glutaraldehyde (20 3 mM) by nasal instillation for 1, 5, or 28 days. Animals were killed at the end of the 4 experiments, and the nasal respiratory epithelium was removed for gene expression 5 analysis. Both compounds induce similar acute and subchronic histopathology 6 characterized by inflammation, hyperplasia, and squamous metaplasia; however, 7 glutaraldehyde does not cause nasal tumors in rats. Differences in the gene expression 8 profiles in rats exposed to formaldehyde and glutaraldehyde help explain the different 9 cancer response from these two aldehydes. Acute exposures generated alterations in gene 10 profiles associated with cellular proliferation, stress, and xenobiotic metabolism; 11 however, longer exposures induced a different subset of genes. Apoptosis gene 12 expression was increased by exposure to formaldehyde compared with controls but was 13 less than observed in glutaraldehyde-exposed rats. In addition, formaldehyde exposure 14 induced a greater increased expression of DNA repair genes than glutaraldehyde.

15 Decreased DNA repair could stimulate apoptosis, while increased DNA repair following

16 formaldehyde exposure could increase DNA misrepair. Misrepaired cells could persist

17 and pass on genetic damage.

18 Sul et al. (2007) investigated the effects of formaldehyde exposure on mRNA expression

19 in rat lung tissues. Male Sprague-Dawley rats were exposed to 0-, 5-, or 100-ppm

20 formaldehyde 6 hours/day, 5 days/week for 2 weeks. Cytotoxic effects were determined

21 by the malondialdehyde lipid peroxidation and the carbonyl protein oxidation assays and

22 showed that the cytotoxic effects increased with exposure. Gene expression analysis

23 indicated that there were 2 up-regulated and 19 down-regulated genes. Nine of these

24 genes were confirmed by real time PCR and included cytochrome P450,

25 hydroxymethylbilane synthase, glutathione reductase, carbonic anhydrase 2, natriuretic

26 peptide receptor 3, lysosomal-associated protein transmembrane 5, regulator of G-protein

- 27 signaling 3, olfactomedin-related ER-localized protein, and poly (ADP-ribose)
- 28 polymerase-1. These genes are involved in apoptosis, immunity, metabolism, signal

29 transduction, transportation, coagulation, and oncogenesis.

1 Andersen *et al.* (2008) investigated the relationship between histopathological changes in 2 nasal tissues and changes in gene expression in rats exposed to 0-, 0.7-, 2-, and 6-ppm 3 formaldehyde by inhalation, 5 days/week for up to 3 weeks. In addition, other groups of 4 rats were exposed to 15 ppm for 6 hours or to 40  $\mu$ L (400 mM) formaldehyde instilled in 5 the nostrils just inside the nares. Unequivocal treatment-related lesions were evident only 6 in the 6-ppm group. In this group, cell proliferation increased at day 5 but was not 7 increased at the end of day 15. Squamous metaplasia occurred at day 5 and epithelial 8 hyperplasia occurred at day 5 and day 15. Lesions were observed primarily in the 9 transitional and respiratory epithelium and displayed an anterior to posterior gradient. 10 The microarray analysis indicated that about 100 genes showed altered expression across 11 all time points and doses. No significant gene expression changes were observed in the 12 0.7-ppm group at any time point. One gene showed increased expression in the 2-ppm 13 group on day 1, while on day 5, 1 gene was decreased and 14 were increased. No gene 14 expression changes occurred in the 2-ppm group on days 6 or 15. The majority of gene 15 expression changes were seen in the 6-ppm group (day 1, 24 genes increased and 18 16 decreased; day 5, 24 increased and 4 decreased; day 6, 9 increased and 0 decreased; day 17 15, 23 increased and 31 decreased). In the acute studies, inhalation of 15 ppm or 18 instillation of 400 mM formaldehyde altered many more genes than were affected at 6 19 ppm, and instillation altered more than three times as many genes as the 15-ppm 20 exposure. U-shaped dose-response curves were observed in the acute study for many 21 genes that were also altered at 2 ppm on day 5. Many of the genes that showed increased 22 expression were involved in response to wounding, control and induction of apoptosis, 23 inflammation pathways, and receptor tyrosine kinase signaling.

24

### 5.7 Mechanistic considerations

Although the biological mechanisms associated with formaldehyde-induced cancer are
not completely understood, it is important to recognize that chemicals can act through
multiple toxicity pathways and mechanisms to induce cancer or other health effects
(Guyton *et al.* 2009). These authors identified at least 15 key events representing diverse
carcinogenic modes of action, the relative importance of which may vary with life stage,
genetic background, and dose. These events include DNA reactivity (covalent binding),

1	gene mutation, chromosomal breakage, aneuploidy, enzyme-mediated effects on DNA
2	damage or repair, epigenetic effects, cell signaling (nuclear-receptor mediated or other
3	than nuclear-receptor mediated), immune response modulation, inflammation,
4	cytotoxicity and compensatory cell proliferation, mitogenicity, chronic metabolic or
5	physiologic overload, nutrient deficiency, and interference with intercellular
6	communication (e.g., gap junctions). Nine of these (DNA reactivity, gene mutation,
7	chromosomal breakage, aneuploidy, enzyme-mediated DNA damage/repair, cell
8	signaling other than nuclear-receptor mediated, immune response modulation,
9	inflammation, and cytotoxicity) were listed as key events for formaldehyde. Although
10	epigenetic effects were not listed as a key event for formaldehyde, a recent study (Lu et
11	al. 2008a) indicates that formaldehyde may alter epigenetic regulation. This section
12	discusses the evidence for genotoxic and cytotoxic modes of action in formaldehyde
13	carcinogenesis and the mutational spectra of these tumors. Most of the literature has
14	focused on upper respiratory tract cancer; however, several investigators have discussed
15	possible modes of action for systemic cancers (i.e., leukemia).

### 16 5.7.1 Genotoxicity

Formaldehyde is highly reactive and can induce a number of genotoxic effects (see
Section 5.6), including DNA-protein crosslinks, strand breaks, mutations, cell
transformation, sister chromatid exchange, and micronuclei from both aneugenic and
clastogenic effects.

21 DNA-protein crosslinks, in particular, have been identified as a marker of formaldehyde-22 induced genotoxicity and have frequently been used as a surrogate for formaldehyde 23 exposure in dose-response modeling. Crosslinks have been detected in many in vitro 24 studies with a number of human and experimental animal cell types, and in vivo in 25 experimental animals and humans. The *in vitro* studies also showed consistent dose-26 response relationships, with crosslinks forming at doses that have low cytotoxicity (up to 27 75% cell survival). DNA-protein crosslinks were not repaired as efficiently in human 28 peripheral blood lymphocytes as in established cell lines. Formaldehyde might interfere 29 with DNA repair by inhibiting repair enzymes, inhibiting removal of DNA lesions, or 30 altering gene expression. Merk and Speit (1998) and Speit et al. (2000) reported that

1 formaldehyde-induced DNA-protein crosslinks are related to chromosomal effects (SCE

2 and micronuclei), but not directly to gene mutations.

3 In vivo studies with rats indicated that inhalation of formaldehyde vapors does results in 4 crosslinks in their nasal mucosa. Furthermore, crosslink yields were highest in the area of 5 the nose (lateral meatus) where tumor yields are the highest. Several studies have 6 examined dose-response relationships for the formation of these crosslinks in nasal 7 tissues of experimental animals and compared these results with nasal tumor data 8 (Casanova-Schmitz et al. 1984a, Casanova et al. 1989, Casanova et al. 1994, Casanova et 9 al. 1991, Heck et al. 1986, Heck et al. 1989). The dose-response curves for DNA-protein 10 crosslink formation and nasal tumor formation in rats showed a similar pattern (Liteplo 11 and Meek 2003). They are nonlinear, with the slope increasing sharply at concentrations 12 above 2 ppm (Table 5-28). This biphasic dose-response curve suggests protective 13 mechanisms, which may become saturated at high concentrations. Two protective 14 mechanisms have been identified: the mucous layer lining the nasal epithelium and 15 glutathione-mediated oxidation of formaldehyde to formate (Conaway et al. 1996). 16 Casanova et al. (1994) reported that the yield in pre-exposed versus naïve rats was about 17 the same. Crosslinks were not detected in rat bone marrow in rats [only one study 18 reviewed] or in the olfactory mucosa or bone marrow of mice exposed to formaldehyde..

Fxposure	DNA-protein crossli	Tumor incidence (%)	
(ppm)	High tumor region <sup>a</sup>		
0	0	0	0/90
0.7	5	5	0/90
2	8	8	0/96
6	30	10	1/90 (1.1)
10	nd	nd	20/90 (22.2)
15	150	60	69/147 (46.9)

 Table 5-28. Formaldehyde exposure, DNA-protein crosslinks, and nasal tumor

 incidence

Adapted from Liteplo and Meek 2003.

nd = no data.

<sup>a</sup> Includes the complete lateral meatus.

<sup>b</sup> Includes medial aspects of naso- and maxilloturbinates, posterior lateral wall, posterior dorsal septum (excluding olfactory region), and nasopharyngeal meatuses.
1 In monkeys, crosslink yields were highest in the middle turbinates. Casanova *et al.* 2 (1991) reported that the level of DNA-protein crosslinks in rhesus monkeys declined in 3 the order: middle turbinates > anterior lateral wall-septum > nasopharynx, which is 4 consistent with the location and severity of proliferative lesions reported in another study 5 (Monticello et al. 1989) in monkeys exposed to 6-ppm formaldehyde for up to 6 weeks. 6 Low levels of crosslinks also were found in the trachea and carina of some monkeys. The 7 yield of crosslinks in monkeys was about an order of magnitude lower than observed in 8 rats, which is primarily attributed to differences in minute volume and quantity of DNA 9 in the nasal mucosa (Casanova et al. 1991). These authors used the crosslink data from 10 rats and monkeys to extrapolate crosslink concentrations in humans and predicted that 11 adult men would have significantly lower rates than rats and slightly lower rates than 12 monkeys.

DNA-protein crosslinks were detected in peripheral lymphocytes of health professionals
 (physicians, laboratory assistants and orderlies from pathology departments) exposed to
 formaldehyde. (see Section 5.6.4). There was a linear relationship between years of
 exposure and DNA-protein crosslinks.

17 Other genotoxic endpoints have been examined in in vitro and in vivo studies. DNA 18 damage (single-strand breaks) was detected in Saccharomyces cerevisiae and in 19 mammalian cells in vitro, including human cells such as fibroblasts, lymphocytes, and 20 lung/bronchial epithelial cells. Strand breaks were also reported in rat lymphocytes 21 (inhalation exposure), and in maternal and fetal liver following i.p. injection on gestation 22 days 6 to 19. DNA damage, as assessed by the alkaline comet assay, increased in 23 lymphocytes from pathology laboratory workers exposed to formaldehyde compared with 24 unexposed controls (reviewed in Section 5.6.4.1): comet tail length for lymphocytes was 25 positively associated with formaldehyde exposure levels. 26 In prokaryotes, formaldehyde induced mainly base-pair mutations, in either the presence

- 27 or absence of metabolic activation at 100% frequency in certain *S. typhimurium* strains
- 28 (TA102, TA104, and TA7005), at a lower rate in TA100, and not at all in TA1535.
- 29 Mutations were induced in mammalian cells in vitro by exposure to formaldehyde, and
- 30 dominant lethal mutations were reported in multiple studies in both rats and mice. No

reports of mutations in humans were identified, and three studies of health professionals
 were negative for effects of formaldehyde on DNA repair.

3 Chromosomal aberrations were positive in both animal and human cells in vitro in all 4 studies summarized in Table 5-19. However, studies in mice with i.p. injection were 5 negative for chromosomal aberrations in bone marrow, spleen, and sperm. Exposure of 6 rats by inhalation caused chromosomal aberrations in pulmonary lavage cells at the 7 highest dose (15 ppm) tested. One study reported chromosomal aberrations in rat bone 8 marrow following inhalation exposure to 0.4 ppm formaldehyde for 4 months, but 9 another study did not find an increase in chromosomal aberrations in rat bone marrow 10 when exposed to 15 ppm for up to 8 weeks (see Section 5.6.3.2). The frequency of 11 chromosomal aberrations was increased in studies of lymphocytes from humans (mainly 12 workers) exposed to formaldehyde were found in 6 of the 11 reviewed in Table 5-25 and 13 one additional positive study published in Chinese and reviewed by Tang et al. (2009). 14 Of the five negative studies reported in Table 5-25, Thompson et al. (1984) reported on 15 small numbers of workers (six exposed and five controls) and Vargová et al. (1992) 16 noted that the frequency of chromosomal aberrations in the controls in their study was 17 higher than that reported in the general population. The results for chromosomal 18 aberrations are potentially of greater interest than other endpoints because of the report 19 by Bonassi et al. (2000) that high levels of chromosomal aberrations were associated with 20 increased risk of cancer in otherwise healthy individuals. 21 Sister chromatid exchange was positive in all studies in animal and human cells in vitro

summarized in Table 5-19, but negative results were reported for two studies in rats in

Table 5-10. Slightly more than half (i.e., 6) of the 11 studies of lymphocytes from

humans exposed to formaldehyde summarized in Table 5-26 were positive. Of the five

negative studies, the study by Thompson *et al.* (1984) was based on small numbers of

subjects, and there were two additional negative studies from the Chinese literature

27 reviewed by Tang *et al.* (2009).

28 Micronuclei were induced in all *in vitro* animal studies and studies of formaldehyde

29 exposed workers or subjects summarized in Table 5-19, but results were mixed for *in* 

30 vivo rat studies, with one oral study positive for the GI tract and one i.p. study negative

1 for bone marrow cells. Speit *et al.* reported that micronucleus formation was enhanced in 2 repair-deficient cell lines, particularly in xeroderma pigmentosum cells, which are 3 deficient in nucleotide excision repair. Loss of glutathione (i.e., GSH) did not affect 4 repair rates. Studies of workers or medical staff or students exposed to formaldehyde 5 measured micronuclei frequency in buccal or oral epithelium, nasal epithelium, and 6 lymphocytes: increased incidences of micronuclei were found in lymphocytes in 5 of 6 7 available studies, buccal or oral epithelium in 4 of the 5 available studies, and nasal 8 epithelium in 4 of the 6 available studies (see Table 5-27). In addition, a review of the 9 Chinese literature by Tang *et al.* 2009 of studies of humans exposed to formaldehyde 10 exposed reported increased micronuclei frequency in nasal epithelial cells in one study, 11 and in lymphocyte in three studies of long-term (> 1 year) formaldehyde exposure. 12 Micronuclei may form from clastogenic or aneugenic events. Titenko-Holland et al. 13 (1996) reported a greater increase of centromere-negative micronuclei in buccal and nasal 14 mucosa cells from mortuary science students and concluded that chromosome breakage 15 was the primary mechanism responsible for these effects. In contrast, Orsière *et al.* 16 (2006) and Iarmarcovai et al. (2007) reported greater increases in centromere-positive 17 micronuclei (evidence of aneugenic effects) in peripheral lymphocytes of untreated 18 cancer patients, welders, and pathologists/anatomists exposed to formaldehyde. Shaham 19 et al. (2003) reported an association between DNA-protein crosslinks in formaldehyde-20 exposed workers and increased serum p53 protein. Furthermore, a positive correlation 21 was found between increased p53 and mutant p53 protein, indicating a possible causal 22 relationship between crosslinks and p53 mutations that may represent steps in 23 formaldehyde carcinogenesis.

24 5.7.2 Glutathione depletion and oxidative stress

25 5.7.2.1 In vitro studies

26 Ku and Billings (1984) reported that the metabolism and toxicity of formaldehyde in

27 isolated rat hepatocytes was dependent upon the intracellular glutathione concentration.

- 28 Hepatocytes depleted of glutathione were more susceptible to formaldehyde toxicity (loss
- 29 of membrane integrity and lipid peroxidation). Cells treated with L-methionine had
- 30 increased concentrations of glutathione and were protected from formaldehyde toxicity.
- 31 Cells treated with antioxidants also showed a dose-related protection against toxicity

suggesting that formaldehyde toxicity in glutathione-depleted cells may be mediated by a
 free radical mechanism.

3 Grafström (1990) studied the ability of formaldehyde and acrolein to cause various 4 effects associated with carcinogenesis in cultured human bronchial cells. These included 5 cell viability, differentiation and growth, membrane integrity, thiol and ion homeostasis, 6 and genetic damage. Concentrations of formaldehyde associated with 50% inhibition 7 were as follows: 0.4 mM (colony-forming efficiency), 0.2 mM (clonal growth rate), and 2 8 mM (membrane integrity measured by trypan blue exclusion). Free cytosolic  $Ca^{2+}$  in bronchial fibroblasts was increased by 50% at 0.5 mM. In addition, 0.2 mM 9 10 formaldehyde decreased glutathione content to 80% of controls and increased the 11 percentage of crosslinked envelopes, a marker for squamous differentiation, to 12% 12 compared with 2% for controls. Grafström et al. (1996) also reported toxic effects of 13 formaldehyde in cultured human bronchial epithelial cells under defined serum- and 14 thiol-free exposure conditions. Formaldehyde was associated with the formation of thiohemiacetal, but not with overt oxidative stress; however, active re-reduction of 15 16 oxidized glutathione by glutathione reductase may have masked an oxidant effect. Loss 17 of membrane integrity coincided with extensive loss of intracellular glutathione. 18 Formaldehyde-induced growth inhibition may be explained by decreased glutathione 19 levels because decreased glutathione levels are known to inhibit cell growth. These 20 authors also noted that genetic damage may be responsible for some of the cytotoxic 21 action of formaldehyde because inhibition of DNA repair occurred in bronchial cells 22 exposed to 0.1 to 0.3 mM formaldehyde. Thus, loss of enzyme function (particularly 23 enzymes that carry a thiol moiety in their active site) might be an essential aspect of 24 formaldehyde toxicity.

Nilsson *et al.* (1998) investigated the role of exogenous and endogenous thiols in
formaldehyde toxicity in human oral fibroblasts and epithelial cells. Formaldehyde
decreased the colony-forming efficiency of both cell types in a concentration-dependent
manner, but was more toxic to fibroblasts than epithelial cells. The difference in toxicity
was attributed to the comparatively lower cellular levels of thiols (glutathione and
cysteine) in fibroblasts.

Teng *et al.* (2001) also investigated the cytotoxic effects of formaldehyde in isolated rat
hepatocytes. Hepatocytes were treated with 2, 4, or 10 mM formaldehyde. Dosedependent effects included a decrease in mitochondrial membrane potential, inhibition of
mitochondrial respiration that was accompanied by formation of reactive oxygen species,
glutathione depletion, and lipid peroxidation. Cells depleted of glutathione were much
more susceptible to the cytotoxic effects of formaldehyde. Cytotoxicity was associated
with a decrease in metabolism and an increase in lipid peroxidation.

8 Tyihák et al. (2001) exposed human HT-29 colon carcinoma and HUV-EC-C endothelial 9 cell cultures to formaldehyde concentrations of 0.1 to 10 mM. Cultures were evaluated at 10 24, 48, and 72 hours after treatment. The cell cultures exposed to the high dose were 11 completely eradicated. At 1 mM, enhanced apoptosis and reduced mitosis were observed 12 in cultures of both cell types, while at the low dose (0.1 mM), enhanced cell proliferation 13 and decreased apoptotic activity occurred. Tumor cells were more responsive than 14 endothelial cells at the low-dose level. The authors proposed that low doses of exogenous 15 or intrinsic formaldehyde may increase cell proliferation and inhibit apoptosis leading to 16 neoplasia, whereas at high doses, formaldehyde may cause damage to endothelial, 17 epithelial, or other cells by inducing apoptosis, and inhibiting repair.

18 Saito *et al.* (2005) investigated the cytotoxic effects exerted by formaldehyde in the 19 presence or absence of reactive oxygen species. Jurkat E6-1 cells from a human T-20 leukemia cell line were cultured with variable concentrations of formaldehyde (< 1 to 100 21 mM) for 3 hours. There was a concentration-dependent decrease in cell viability with 22 significant decreases at concentrations greater than 1 mM. Cells cultured with the water-23 soluble radical initiator, 2,2'-azobis-[2-(2-imidazolin-2-yl)propane] dihydrochloride 24 (AIPH) at concentrations up to 8 mM showed no decrease in viability. However, cell 25 viability was significantly decreased at AIPH concentrations of more than 3 mM in the 26 presence of 1 mM formaldehyde. Further analysis indicated that cell death resulted from 27 necrosis rather than apoptosis. Cell death was preceded by a significantly increased 28 cellular level of reactive oxygen species. Total cellular glutathione was reduced to about 29 60% of the control value in cells treated with 1 mM formaldehyde for 2 hours, while 6 30 mM AIPH reduced glutathione levels to about 5% of the control value. Glutathione was

completely depleted in cell cultures treated with both formaldehyde and AIPH. These
 results indicate a synergistic interaction of formaldehyde and free radicals.
 5.7.2.2 In vivo studies
 *In vitro* studies (discussed above) indicated that formaldehyde exposure resulted in the
 formation of reactive oxygen species, glutathione depletion, and lipid peroxidation and

6 that antioxidants had a protective effect (Ku and Billings 1984, Teng *et al.* 2001). Several

7 *in vivo* studies have examined oxidative stress in rats exposed to formaldehyde. These

8 studies show that formaldehyde exposure can cause oxidative stress in the rat liver,

9 plasma, lymphocytes, heart, and brain.

10 Söğüt *et al.* (2004) investigated the oxidant/antioxidant status of albino Wistar rats 11 exposed to 0-, 10-, or 200-ppm formaldehyde 8 hours/day, 5 days/week for 4 weeks. 12 Glutathione levels in liver tissues were significantly reduced at both exposure levels. 13 Xanthine oxidase levels were reduced in the high-dose group. There were no significant 14 changes in malondialdehyde or nitric oxide levels. Thus, the authors suggested that the 15 antioxidant system of liver tissue is moderately impaired by excessive formaldehyde 16 exposure. The authors also concluded that glutathione depletion from subacute exposures 17 to formaldehyde may increase susceptibility to oxidative damage.

18 Gurel et al. (2005) investigated the biochemical and histopathological changes occurring 19 in the frontal cortex and hippocampal tissue of the rat brain after formaldehyde exposure. 20 Male Wistar rats were divided into three groups of six rats each. One group received i.p. 21 injections of 10 mg/kg formaldehyde (37% solution) for 10 days. The second group 22 received i.p. injections of formaldehyde and vitamin E, and the third group was untreated 23 (controls). The animals were killed at the end of the treatment period, and the frontal 24 cortex and hippocampal tissues were removed. Malondialdehyde and protein carbonyl 25 levels were significantly increased in these tissues, while superoxide dismutase and 26 catalase enzyme activities were decreased in the formaldehyde-only treatment group 27 compared with controls. Rats treated with both formaldehyde and vitamin E showed 28 lower malondialdehyde and protein carbonyl levels with no inhibition of superoxide 29 dismutase or catalase. The authors concluded that formaldehyde caused oxidative damage 30 to tissues in the brain, which was likely mediated through the production of free radicals.

1 Gülec et al. (2006) evaluated the oxidant/antioxidant status and lipid peroxidation in the 2 hearts of rats exposed to formaldehyde. Groups of 10 adult Wistar rats [sex was not 3 identified] were placed in inhalation chambers and exposed to 0-, 10-, or 20-ppm 4 formaldehyde 8 hours/day, 5 days/week for 4 or 13 weeks. The animals were checked 5 daily and body weights were recorded weekly. At the end of the experiment, the animals 6 were necropsied, examined grossly for pathological changes, and heart tissues were 7 prepared for biochemical analysis. Superoxide dismutase levels were increased in all 8 exposed groups compared with controls. Catalase activity was significantly decreased at 9 both exposure levels in groups exposed for 4 weeks. Thiobarbituric acid-reactant 10 substances were measured as an index of lipid peroxidation and were slightly increased in 11 exposed groups compared with controls but the differences were not significant. Nitric 12 oxide levels were not affected. The authors concluded that subacute and subchronic 13 exposure to formaldehyde might stimulate oxidative stress in cardiac cells and tissues. 14 The increased superoxide dismutase activity was thought to be secondary to decreased 15 catalase activity, as a compensatory mechanism, thus protecting heart tissue from 16 damage.

17 Im et al. (2006) evaluated the effects of formaldehyde exposure on rat plasma proteins. 18 Male Sprague-Dawley rats (10 per group) were exposed to 0-, 5-, or 10-ppm 19 formaldehyde 6 hours/day, 5 days/week for 2 weeks in an inhalation chamber. Lipid 20 peroxidation and protein oxidation levels in plasma, lymphocytes, and liver were 21 determined using the malondialdehyde assay and carbonyl spectrometric assay. The 22 comet assay was used to evaluate DNA damage (see Section 5.6.3.1). Lipid peroxidation 23 and protein oxidation were dose-dependently increased in plasma, lymphocytes, and liver 24 of exposed rats. In addition, a proteomic analysis identified 19 up-regulated and 13 25 down-regulated proteins as biomarkers of formaldehyde exposure. These included 26 proteins involved in apoptosis, transportation, signaling, energy metabolism, and cell 27 structure and motility.

Kum *et al.* (2007a) measured oxidative stress in the adult and developing rat liver after
inhalation exposure to formaldehyde and xylene. Four age groups (embryonic day 1, 1day-old, 4-weeks-old, and adults), each containing 24 female Sprague-Dawley rats were

1 used. Each age group was further divided into four experimental groups of six rats each. 2 In addition to the control group, rats were exposed to 6-ppm formaldehyde, 300-ppm 3 xylene, or xylene + formaldehyde for 8 hours/day for 6 weeks. Body and liver weights were measured, and superoxide dismutase, catalase, glutathione, and malondialdehyde 4 5 levels were determined. Body and liver weights were decreased in all exposure groups 6 compared with controls in the embryonic day 1 group compared with controls. Body and 7 liver weights were significantly decreased in the xylene + formaldehyde exposure groups 8 of 1-day-old rats, but not in the xylene + formaldehyde combined exposure group. Liver 9 weights were significantly higher in the xylene and xylene + formaldehyde combined 10 exposure groups of 4-week-old rats. There were no significant differences in body or 11 liver weights in the adult rat exposure groups compared with controls. Superoxide 12 dismutase levels were significantly decreased in the formaldehyde-exposed group of 4-13 week-old rats. Glutathione levels were significantly decreased in the xylene and xylene + 14 formaldehyde combined exposure groups of 1-day-old rats. Malondialdehyde levels were 15 not significantly different from controls in any of the formaldehyde or xylene + 16 formaldehyde combined exposure groups. Catalase activity was slightly increased in the 17 xylene + formaldehyde combined exposure group of embryonic rats. The authors 18 concluded that these data suggested that the developing rat liver is more susceptible to the 19 toxic effects of formaldehyde and xylene than the adult rat liver.

20 5.7.3 Mutational spectra

21 Recio (1997) reviewed the literature on oncogene and tumor-suppressor gene alterations 22 in rodent nasal tumors. Molecular genetic studies on nasal squamous-cell carcinomas in 23 rats indicated that *p53* mutations occur at a high frequency. This finding combined with 24 the high prevalence of p53 mutations among human squamous-cell carcinomas suggests 25 that a common molecular alteration is shared between human and rodent squamous-cell 26 carcinomas. The *HPRT* mutational spectra in formaldehyde-exposed human lymphoblasts 27 show about 50% deletions and 50% point mutations, with the majority of point mutations 28 occurring at A:T base pairs (Liber et al. 1989). However, this finding is inconsistent with 29 the G:C base-pair mutations observed in formaldehyde-induced nasal squamous-cell 30 carcinomas in rats (Recio et al. 1992). Recio (1997) concluded that the lack of p53 point 31 mutations at A:T base pairs in formaldehyde-induced squamous-cell carcinomas

1 suggested an indirect mechanism of genotoxicity rather than a direct effect of 2 formaldehyde on the cellular genome. The origin of the point mutations in p53 observed 3 in formaldehyde-induced nasal squamous-cell carcinomas in rats is unknown, but 4 inflammation and regenerative cell proliferation are thought to be important factors. 5 Recio et al. (1992) examined the complementary DNA of the tumor-suppressor gene p53 6 from 11 primary nasal squamous-cell tumors taken from rats exposed to formaldehyde. 7 Point mutations at G:C base pairs were found in *p53* in 5 of 11 tumors analyzed. All of 8 the mutated p53 codons found in rat tumors have also been identified in a variety of 9 human cancers. In particular, one of the mutations that occurred at rat codon 271 10 (analogous to human codon 273), is a known *p53* mutational hot spot in human cancers. 11 In addition, Wolf et al. (1995) used an immunohistochemical technique to measure p53 12 protein, proliferating cell nuclear antigen (PCNA), and tumor growth factor- $\alpha$  in these 13 tumors. These authors observed p53-positive immunostaining and preneoplastic 14 hyperkeratotic plaques in the tumors but not in normal nasal mucosa. There was a 15 correlation between both the pattern and distribution of immunostaining of proliferating 16 cell nuclear antigen and p53. Four cell lines were established from these squamous-cell 17 carcinomas (Bermudez et al. 1994). All the cell lines were aneuploid and overexpressed 18 keratin, transforming growth factor- $\alpha$ , epidermal growth factor receptors, and p53. 19 Expression of transforming growth factor- $\alpha$  and epidermal growth factor is a common 20 feature of squamous-cell carcinoma and is frequently found in human tumors. When 21 injected into nude mice, the two cell lines that contained a p53 mutation were 22 tumorigenic, but the two cell lines that had wild-type *p53* were not.

23 5.7.4 Epigenetic effects

Lu et al. (2008a) reported that formaldehyde induced histone modifications in vitro.

- 25 Lysine residues on histones are subject to post-translational modifications (e.g.,
- 26 methylation, phosphorylation, and acetylation) which impact gene expression. DNA-
- 27 protein crosslinks involve all the major histones and are a dominant form of
- formaldehyde-induced DNA damage (Quievryn and Zhitkovich 2000). Lu et al. (2008a)
- 29 isolated histone 4 with post-translational modification from calf thymus tissues.
- 30 Unmodified human recombinant histone 4 was purified after expression in *E. coli* cells.

1 Both proteins had identical sequences. Formaldehyde was reacted with histone 4 and 2 analyzed by liquid chromatography-mass spectrometry. All the lysine residues located in 3 both the histone N-terminal tail and the globular fold domain were identified as binding 4 sites for formaldehyde. Formaldehyde could only bind to lysine residues without post-5 translational modification, thus, post-translational modification of lysine blocks the 6 reaction with formaldehyde. However, formaldehyde reactions with unmodified lysine 7 residues resulted in the formation of methylol groups followed by the formation of Schiff 8 bases. Formaldehyde-induced Schiff bases inhibited post-translational modifications of 9 lysine *in vitro*. Therefore, formaldehyde could alter epigenetic regulation by impairing 10 the post-translational modification pattern and possibly disturb subsequent protein 11 recruitment and trigger a series of abnormal cascade effects. Furthermore, the balance 12 between histone acetylation and deacetylation (which is important for normal cell 13 growth) could be disturbed. An imbalance of acetylation in promoter regions could 14 induce the deregulation of gene expression and affect carcinogenesis and cancer 15 progression. The authors noted that they used a simplified *in vitro* model and that further 16 testing in cells or tissues would be needed to demonstrate that such effects would occur in 17 vivo.

#### 18 5.7.5 Nasal tumors

19 Increased incidences of nasal tumors were found in studies in experimental animals (see 20 Section 4). In addition, oral administration of formaldehyde to rats resulted in increased 21 incidences of gastrointestinal tract cancers. There is considerable evidence that airway 22 deposition, genotoxicity, cytotoxicity, and cell proliferation are important factors in nasal 23 tumor formation (IARC 2006). A number of studies have investigated the underlying 24 mechanisms of the nasal tumor response (reviewed by Heck et al. 1990, Morgan 1997). 25 In parallel with the mechanistic studies, anatomically accurate three-dimensional 26 computation fluid dynamics (CFD) models have been developed to provide high 27 resolution predictions of nasal air flow and regional flux of inhaled formaldehyde (see 28 Section 5.2) into adjacent nasal tissue. CFD models also have been used to predict 29 crosslink formation, and, when combined with a two-stage clonal growth model, to link 30 crosslink and regenerative cellular proliferation with tumor formation (Conolly et al. 31 2003, 2004, Conolly et al. 2000).

## 1 5.7.5.1 Airway deposition models and predictions

2 Morgan (1997) considered that although the nasal passages of rats and humans are 3 fundamentally identical biological target organs, minor differences could be critically 4 important. Regional deposition of inhaled gases and tissue susceptibility are the two 5 major factors that influence the distribution of lesions in the respiratory tract. Tissue 6 susceptibility is frequently related to differences in local enzyme-mediated 7 biotransformation to a toxic species or to local doses that exceed detoxification 8 thresholds. Keller et al. (1990) conducted a histochemical analysis of formaldehyde 9 dehydrogenase (the primary metabolizing enzyme for formaldehyde) and reported that 10 regional differences were insufficient to account for the localized toxicity of 11 formaldehyde in the rat nose, which would indicate that nasal airflow and intranasal 12 uptake patterns of formaldehyde were important. CFD models have allowed researchers 13 to investigate interspecies differences in airflow patterns, formaldehyde flux and 14 absorption, and effects on the upper respiratory tract, and to gain a better understanding 15 of mechanisms and modes of action.

16 Studies with formaldehyde-exposed rats and rhesus monkeys show site- and species-17 specific patterns for both carcinogenic and noncarcinogenic lesions in the upper 18 respiratory tract (Casanova et al. 1994, Kimbell et al. 1997, Monticello et al. 1996). The 19 nasal vestibule in rats, monkeys, and humans is lined with squamous epithelium; 20 however, areas posterior to the nasal vestibule are lined with respiratory, transitional, and 21 olfactory epithelia (Kimbell et al. 1997). Inhaled formaldehyde does not result in lesions 22 in the nasal vestibule, but a common response in other epithelia is conversion to the 23 squamous form (i.e., squamous metaplasia). This observation suggests that squamous 24 epithelium is resistant to formaldehyde toxicity and that squamous metaplasia may be an 25 adaptive response. Further, squamous epithelium may be protective by absorbing less 26 formaldehyde than other epithelial types. Kimbell et al. (1997) compared CFD model 27 predictions and observed squamous metaplasia incidence in the area of the rat nose 28 (lateral meatus and mid-septum) where squamous-cell carcinoma occurred in chronic 29 inhalation studies (Figure 5-3). Regional formaldehyde flux was correlated with the 30 distribution of formaldehyde-induced squamous metaplasia in rats exposed to 10- or 15-

- 1 ppm formaldehyde. Kepler *et al.* (1998) conducted a similar study in the rhesus monkey.
- 2 Simulated airflow patterns showed good agreement with experimental observations.



Figure 5-3. Sagital (A) and cross-section (B) through the rat nose.

Source: adapted from Kerns *et al.* 1983 and Mery *et al.* 1994. (Illustration prepared by Donna Jeanne Corcoran, ImageAssociates.)

A) Sagital section through the rat nose. The curved dashed lines indicate the junction of the squamous/transitional and respiratory epithelia (anterior line) and the respiratory and olfactory epithelia (posterior line). N = nasoturbinates, M = maxilloturbinates, E = ethmoturbinates, ID = incisive duct, NPD = nasopharyngeal duct, OB = olfactory bulb, 2PR = second palatal ridge.

B) Cross section through the rat nose at the level indicated by the slanted line in panel A.

1 Kimbell et al. (2001a) predicted formaldehyde flux in the entire nasal passages of rats, 2 monkeys, and humans, estimated flux in specific sites for correlation with formaldehyde-3 induced cell proliferation data, and compared the flux values predicted for the three 4 species. Regions of the nasal passages in rats and monkeys that had similar cell 5 proliferation rates also had similar predicted flux values with a rat to monkey ratio of 6 0.98 for the highest site-specific flux values. Simulations using the human CFD model 7 predicted that flux values in an anterior portion of the human nose were similar to fluxes 8 predicted in a region of high tumor incidence in the rat nose. The authors concluded that 9 proliferative and carcinogenic responses could be expected to occur in humans under 10 conditions similar to those inducing these effects in rats and monkeys. Kimbell et al. 11 (2001b) further refined the CFD models to obtain quantitative descriptions of nasal 12 uptake patterns. Their simulations indicated a decreasing gradient of flux values from 13 anterior to posterior regions of the nasal cavity in all three species with steeper gradients 14 in rats and monkeys than in humans. Nasal flux patterns in humans shifted posteriorly, 15 and the overall nasal uptake decreased as inspiratory flow rate increased. The authors 16 noted that these results are consistent with an increased airflow pushing inhaled gas 17 further into the respiratory tract.

18 Cohen-Hubal et al. (1997) conducted the first quantitative demonstration of the role of 19 site-specific formaldehyde flux and crosslink formation. These authors used a CFD 20 model to link dosimetry predictions with measured tissue deposition. Crosslink 21 predictions compared well with experimentally measured data. Conolly et al. (2000) 22 expanded on the work of Cohen-Hubal et al. and used an improved CFD model to predict 23 regional flux of formaldehyde and crosslink formation in the respiratory and olfactory 24 mucosa of the rat, monkey, and human. Simulated formaldehyde concentrations ranged 25 from 0.1 to 20 ppm over a 3-hour exposure. Good fits to the rat and monkey crosslink 26 data were obtained. Differences in the predictions between regions of the nasal mucosa 27 were accounted for by site-specific tissue thickness and flux estimates. The predicted 28 crosslink dose response for the human case was compared with the rat and monkey and 29 was similar for all three species even though there were significant interspecies 30 differences in nasal anatomy, breathing rates, and parameter estimates.

1 Conolly et al. (2003) described biologically motivated quantitative modeling of the 2 exposure-tumor response continuum in the rat using a CFD model linked with a two-3 stage clonal growth model. Regenerative cell proliferation was used as a surrogate for 4 cytolethality. The average division rate constants were based on labeling index data 5 reported by Monticello et al. (1991, 1996). A time-weighted unit length labeling index 6 was calculated for the entire 78 weeks of exposure. The calculated rate constants were 7 plotted against formaldehyde concentrations and resulted in a J-shaped exposure-8 response curve. The probability of mutation per cell generation (a function of the tissue 9 crosslink concentration and the rate of cell division) was used in the clonal growth model 10 to predict tumor yield. A sensitivity analysis indicated that the directly mutagenic 11 pathway had little influence and that the tumor outcome was due primarily to 12 regenerative cellular proliferation.

13 Conolly et al. (2004) extended the approach used by Conolly et al. (2003) to humans. 14 The primary objective was to maximize the use of relevant mechanistic data in predicting 15 human cancer response to inhaled formaldehyde. The only structural difference between 16 the rat and human tumor-response models was that the human model included the entire 17 respiratory tract to provide the capability for predicting tumor risk associated with 18 oronasal breathing at higher exertion levels. The human clonal growth model used three 19 sets of baseline parameters for nonsmokers, smokers, and a mixed population of 20 nonsmokers and smokers in order to estimate human respiratory tract tumor incidences 21 not explicitly related to formaldehyde exposure. Cancer risk predictions were based on J-22 shaped and hockey stick-shaped dose-response curves and included 18 exposure 23 scenarios involving continuous (80-year environmental exposure), and light or heavy 24 working occupational scenarios. Predicted risks for smokers were about an order of 25 magnitude higher than for nonsmokers. Their data indicated that excess risk for 26 continuous environmental exposure to formaldehyde at concentrations below 1 ppm (J-27 shaped dose-response model) or 0.2 ppm (hockey-stick dose-response model) were de 28 *minimis* ( $< 10^{-6}$ ). Breathing rate changes based on various activity levels did not result in 29 large changes to the calculated risk.

1 Results from Conolly et al. (2003, 2004) were later challenged by Subramaniam et al. 2 (2008, 2007) and Crump et al. (2008). These authors identified sources of uncertainty in 3 the CFD models and modified selected features to examine the sensitivity of the 4 predicted dose response to select assumptions. They found that the dose-response 5 predictions below the range of exposures where tumors were observed were highly 6 sensitive to the choice of control data. In contrast to the results reported by Conolly et al. 7 (2003), their reanalysis indicated that up to 74% of the added tumor probability could be 8 attributed to formaldehyde's mutagenic action. Furthermore, slight numerical 9 perturbations in the assumptions regarding the effects of formaldehyde on the division 10 rates and death rates of initiated cells resulted in risk estimates that were up to 10,000 11 times those reported by Conolly et al. (2004).

12 5.7.5.2 Cytotoxicity and cellular proliferation in experimental animals

At high concentrations formaldehyde is highly irritating and cytotoxic, causing loss of cilia and cell death in the nasal cavity (Conaway *et al.* 1996). IARC (2006) provided a comprehensive review of formaldehyde-induced cytotoxicity and cell-proliferation studies. Increased cell proliferation is believed to contribute to carcinogenesis by providing additional cell divisions, thus increasing the probability of spontaneous or chemically induced mutations (Monticello and Morgan 1997).

19 Studies in rats and mice show species differences in the cytotoxicity of inhaled

20 formaldehyde to the respiratory epithelium (Chang et al. 1983, Monticello et al. 1991,

21 Monticello et al. 1996). The sequence of effects, which are more severe in the rat, include

22 rhinitis, epithelial dysplasia, squamous metaplasia and hyperplasia, and squamous-cell

23 carcinoma. Mice were able to compensate for increased concentrations of formaldehyde

24 by reducing minute ventilation, thus reducing deposition and subsequent tissue damage.

25 Eighteen hours after a single 6-hour exposure to 15 ppm, cell proliferation increased 13-

26 fold in rats and 8-fold in mice compared with controls. Cell proliferation was not evident

- 27 until exposure concentrations exceeded 6 ppm following acute, subchronic, or chronic
- 28 exposures; however, histopathological effects and a sustained increase in cell

29 proliferation did not occur at concentrations less than 2 ppm, regardless of the exposure

30 duration.

1	A sustained increase in cellular proliferation subsequent to epithelial-cell toxicity is
2	believed to be an important determinant of neoplastic progression associated with
3	formaldehyde exposure (Liteplo and Meek 2003). Monticello et al. (1996) examined the
4	proliferative response in various regions of the rat nose following exposures to
5	formaldehyde concentrations of 0, 0.7, 2, 6, 10, or 15 ppm for up to 24 months (6
6	hours/day, 5 days/week). Animals were sacrificed at 3, 6, 12, 18, and 24 months. The
7	incidence of regional formaldehyde-induced nasal tumors was correlated with the
8	population-weighted unit length labeling index (i.e., the product of the S-phase nuclei per
9	millimeter of basement membrane and the total number of cells per site) at 3 months.
10	Thus the weighted labeling index incorporates both the cell replication rate and the
11	number of cells at the specific site. A sustained increase in the labeling index was
12	observed only at exposure concentrations that yielded significant numbers of nasal
13	tumors (10 and 15 ppm) (Table 5-29). The authors concluded that target-cell population
14	size, cell proliferation, and local dosimetry play a significant role in the concentration-

15 response curve for formaldehyde-induced nasal cancer in rats.

Exposure	Cell proliferation (population-weighted S-phase nuclei/mm basement membrane × 10 <sup>6</sup> ) <sup>a</sup>			Tumor incidence (%)		
(ppm)	ALM	PLM	AMS	ALM	PLM	AMS
0	9.9	3.9	1.2	0/90	0/90	0/90
0.7	10.3	4.0	1.5	0/90	0/90	0/90
2	9.6	5.7	2.3	0/90	0/90	0/90
6	15.4	4.9	0.8	1/90 (1)	0/90	0/90
10	74.9	7.8	7.2	12/90 (13)	2/90 (2)	0/90
15	91.0	30.2	13.9	17/147 (12)	9/147 (6)	8/147 (5)

 Table 5-29. Formaldehyde exposure, cell proliferation, and nasal tumor incidence

Adapted from Monticello et al. 1996.

ALM = anterior lateral meatus; PLM = posterior lateral meatus; AMS = anterior mid-septum.

<sup>a</sup> Calculated as the product of the unit length labeling index and the total number of nasal epithelial cells at each site. [These data were presented in Figure 8 of Monticello *et al.* (1996); however, the paper incorrectly reported the value as  $10^7$ . The correct value is  $10^6$ .]

- 16 Woutersen et al. (1989) studied the role of cell proliferation in formaldehyde
- 17 carcinogenesis (see Section 4.1.2.2). These authors reported that compound-related
- 18 degenerative, inflammatory, and hyperplastic changes of the nasal respiratory and
- 19 olfactory mucosa were observed when rats with undamaged noses were exposed to 10-
- 20 ppm formaldehyde for 3 months but not when exposed to 0.1 or 1 ppm. These effects

were increased in similarly exposed rats that had severe injury to the nasal mucosa from electrocoagulation. Furthermore, nasal tumors were observed in rats with damaged noses exposed to 10 ppm for 28 months but not in rats with undamaged noses. The authors suggested that tissue damage followed by epithelial regeneration may contribute to formaldehyde-induced carcinogenesis.

6 McGregor *et al.* (2006) reviewed the carcinogenicity and toxicity data of formaldehyde 7 and glutaraldehyde. Although inhalation of these compounds caused similar effects in the 8 nasal epithelium of rats and mice, only formaldehyde induced a dose-related increase in 9 nasal tumors. The postulated mode of action for the carcinogenicity of formaldehyde is 10 that prolonged exposure above a critical concentration induces sustained cytotoxicity and 11 cell proliferation. Genetic changes, occurring secondary to the cytotoxicity, metaplasia, 12 and hyperplasia, result in neoplasia. This mode of action is supported by observations of 13 a consistent, nonlinear dose-response relationship for three key events (sustained cell 14 proliferation, DNA-protein crosslink formation, and tumors) and concordance of these 15 effects across regions of the nasal passages. The nonlinearity of the response may be 16 explained by saturation of glutathione-mediated detoxification at concentrations above 4 17 ppm. However, key events postulated in the mode of action for formaldehyde 18 (cytotoxicity, cell proliferation, and DNA-protein crosslink formation) have been 19 demonstrated with glutaraldehyde exposure without causing nasal tumors in rats and 20 mice. A possible explanation for this discrepancy is that the dialdehyde function of 21 glutaraldehyde may inhibit the macromolecules from further reaction. If these 22 macromolecules are proteins involved in maintenance of survival, then their inhibition 23 may be more likely to lead to cell death instead of a change in the differentiation state. If 24 glutaraldehyde reacts with DNA, then repair of these lesions may be more difficult. This 25 is consistent with the conclusions of Hester et al. (2005) (see Section 5.6.5) based on a 26 comparison of gene-expression profiles, DNA repair, and apoptosis following exposures 27 to formaldehyde or glutaraldehyde, which found that glutaraldehyde had increased 28 apoptosis, greater mitochondrial damage and decreased DNA repair compared to 29 formaldehyde.

## 1 5.7.6 Other tumors

2 Other potential tissue target sites include lymphohematopoietic tumors in humans (acute 3 myelogenous leukemia and other lymphohematopoietic tumors, see Section 3) and 4 experimental animals (hemolymphoreticular tumors, see Section 4), and malignant 5 mammary gland tumors, testicular interstitial-cell adenoma, and gastrointestinal 6 leiomyosarcoma in experimental animals (see Section 4.2). No studies were identified 7 evaluating potential mechanisms for mammary gland, gastrointestinal, or testicular 8 tumors although toxic effects on the testes have been reported in experimental animals 9 (see Section 5.4.3.5). In contrast, numerous mechanistic studies were identified 10 discussing the association between lymphohematopoietic cancers and formaldehyde 11 exposure. This section briefly reviews lymphohematopoietic cancer, and arguments 12 supporting and against the biological plausibility of formaldehyde-induced leukemia. 13 In humans, the bone marrow is the source of all blood cells in the circulation by the time 14 of birth. The blood cells arise from a common pluripotent progenitor cell (stem cell). In 15 the bone marrow, this stem cell forms two multipotent progenitor cells, the common 16 myeloid stem cell and the common lymphoid stem cell. These cells in turn form 17 committed stem cell lines that form fully differentiated blood cells. The myeloid series 18 forms eosinophils, monocytes, polymorphonuclear leukocytes, platelets, erythrocytes, 19 and basophils, whereas the lymphoid series forms plasma cells (B cells), natural killer 20 (NK) cells, and T cells. Hematopoietic progenitor cells have been identified outside of 21 the bone marrow in the peripheral circulation (Fritschi and Siemiatycki 1996), lymph,

22 and in lymphoid tissue and can circulate back to the bone marrow.

23 Malignant blood diseases (leukemia, lymphomas, and myeloma) are a heterogenous 24 group of neoplasms that arise from stem cells at different hierarchical levels of 25 hematopoietic and lymphoid cell development (Greaves 2004). The hierarchical cell 26 population structure includes different stages of stem cells, which are associated with 27 different types of malignancies. Mutations can occur at any stem cell level, and stem cells 28 at any one level undergoing mutations and clonal expansion can produce a variety of 29 different types of neoplasms. The type of neoplasm depends on the target cell undergoing 30 transformation and the phenotype produced as a result of the different genetic

1	abnormalities (Greaves 2004). Examples of lymphoid neoplasms are chronic lymphocytic
2	leukemia, multiple myeloma, Hodgkin's lymphoma, and non-Hodgkin's lymphoma. The
3	terms lymphocytic leukemia and lymphoma are used to describe the usual tissue
4	distribution of the disease (bone marrow and peripheral blood vs. discrete mass in
5	lymphoid tissue) at the time of clinical presentation, but both types of neoplasms can be
6	present in bone marrow, circulating blood, and lymphoid tissues. Acute myelogenous
7	leukemia (AML) is a heterogeneous group of neoplasms that primarily involve the bone
8	marrow. Some lymphatic tumors, especially non-Hodgkin's lymphoma, appear to
9	originate outside the bone marrow (Pyatt et al. 2008).
10	Chromosomal translocations (two-way or reciprocal) are present in the majority of white
11	cell neoplasms, and gene deletion and mutations are also common. Chromosomal
12	translocations in blood neoplasm may arise from disruption of the normal processors of
13	DNA double-strand breakage repair or rearrangements (Greaves 2004).
14	Two groups of researchers have proposed potential mechanisms for formaldehyde-
15	induced leukemia: (1) Zhang et al. (2009a) and (2) the Environmental Protection Agency
16	(EPA) [Note the EPA did not publish their proposed mechanism in the peer-reviewed
17	literature, but the major points are discussed in a criticism published by Pyatt et al. 2008.]
18	The basic concepts of these proposed mechanisms are similar.
19	Zhang et al. (2009a,b) identified three potential mechanisms for formaldehyde-induced
20	leukemia: (1) direct damage to stem cells in bone marrow, (2) damage to circulating
21	hematopoietic stem/progenitor cells in the blood, or (3) damage to pluripotent stem cells
22	present within the nasal turbinates and/or olfactory mucosa. Although the biological
23	plausibility of the first model has been questioned (discussed below), these authors
24	suggested that absorbed formaldehyde would dissolve in the blood and be converted to its
25	hydrated form (methanediol) and could be transported to bone marrow in this form.
26	However, if formaldehyde is not able to reach bone marrow in sufficient quantities to
27	damage stem cells, the two alternate mechanisms involving damage to circulating
28	stem/progenitor cells that travel to bone marrow and become initiated leukemic cells are
29	plausible. Thus, the critical DNA or macromolecular binding occurs in the blood, and

1 when the affected cells proliferate, unrepaired lesions could lead to mutations and cellular 2 toxicity. The initiated stem cell could be re-incorporated into the bone marrow, and 3 eventually lead to leukemia. The authors cited the detection of DNA-protein crosslinks 4 and cytogenetic damage in circulating lymphocytes of exposed workers as supporting 5 evidence. The same type of damage would be expected to occur in circulating 6 hematopoietic stem cells. The third mechanism is similar to the second but involves pre-7 mutagenic or mutagenic damage to primitive pluripotent stem cells that reside in the oral 8 or nasal passages. Damaged stem cells could be released from the nasal passages, perhaps 9 enhanced by formaldehyde-induced cytotoxicity, circulate through the blood, and 10 eventually be incorporated into the bone marrow. Supporting evidence for this 11 mechanism includes toxicity and DNA-protein crosslinks in the nasal passages of 12 laboratory animals exposed to formaldehyde, reports of increased micronuclei in the 13 nasal and oral mucosa of formaldehyde-exposed humans, and a study (Murrell et al. 14 2005) that showed that olfactory epithelial cells obtained from rat nasal passages 15 contained hematopoietic stem/progenitor cells. These cells were shown to re-populate the 16 hematopoietic tissues of irradiated rats and to form hematopoietic stem/progenitor cells 17 of multiple lineages in vivo.

Tang *et al.* reviewed eight studies conducted in China on hematological parameters among formaldehyde-exposed humans. The authors concluded that most of the studies showed that long-term exposure can decrease the number of white blood cells, and possibly lower platelet and hemoglobin (see Section 5.4.2.4). One case report was identified of a previously healthy woman diagnosed with pancytopenia (decreased levels of all formed elements in the blood) shortly after moving into a newly remodeled apartment.

According to Pyatt *et al.* (2008), the EPA-proposed mode of action relies on the
following assumptions: (1) many lymphoid malignancies arise outside of the bone
marrow, (2) lymphoid tissue present at the portal of entry represents a target cell in nasalassociated lymph tissue, (3) circulating stem cells or hematopoietic progenitor cells can
be exposed to formaldehyde in the lungs or nasal passages, (4) formaldehyde has been

1 reported to cause leukemia or lymphomas in rats and mice exposed by inhalation<sup>3</sup> and 2 oral routes, (5) formaldehyde is genotoxic, and (6) some epidemiological studies suggest 3 an association between formaldehyde exposure and lymphohematopoietic malignancies. 4 Several authors have questioned the biological plausibility of an association of 5 formaldehyde and systemic tumors (primarily leukemia) because of formaldehyde 6 reactivity and lack of evidence for bone marrow toxicity (Cole and Axten 2004, Golden 7 et al. 2006, Goldstein 2009, Heck and Casanova 2004, Pyatt et al. 2008). Evidence that 8 suggests that formaldehyde would not be a leukemogen includes the following: (1) 9 normal metabolic processes prevent formaldehyde from entering the systemic circulation 10 as formaldehyde is rapidly metabolized by circulating erythrocytes, and blood 11 concentrations of formaldehyde did not increase in humans exposed to 1.9 ppm for 40 12 minutes, in rats exposed to 14.4 ppm for 2 hours, or in rhesus monkeys exposed to 6 ppm 13 for 4 weeks, (reviewed by Golden et al. 2006); (2) formaldehyde does not cause overt 14 bone marrow toxicity or pancytopenia at high doses, a common feature of known 15 leukemogens; (3) there is no credible evidence that formaldehyde induces leukemia in 16 experimental animals; and (4) epidemiological studies provide limited evidence that 17 occupational exposure to formaldehyde is associated with leukemia. Pyatt et al. (2008) 18 concluded that all known leukemogenic chemicals cause dose-related hematotoxicity, 19 induce bone marrow hypoplasia and dysplastic morphological changes in the bone 20 marrow, and produce hematopoietic neoplasias in rodents. 21 Both EPA (as reviewed by Pyatt et al. 2008) and Zhang et al. (2009a,b) stated that their 22 proposed mechanisms are supported by human studies demonstrating increased 23 micronuclei in nasal and buccal epithelial cells; by the presence of DNA crosslinks, 24 micronuclei, chromosomal aberrations, and SCE in lymphocytes of formaldehyde-

- 25 exposed workers or students; and by animal studies showing increased micronuclei and
- 26 SCE in pulmonary lavage cells of formaldehyde-exposed rats. Pyatt et al. (2008) argued

<sup>&</sup>lt;sup>3</sup> Pyatt *et al.* (2008) stated that the EPA proposal cited the unpublished Batelle data (which is the inhalation study reported by Kerns *et al.* [1983]) as showing a significant increase (and dose-response) in lymphomas in female mice and leukemia in female rats but that the author's review of the data does not support the EPA conclusion.

that the human studies lack consistency, genotoxic effects in animals are limited to local effects, and an *in vitro* study by Schmid and Speit (2007a) found that DNA crosslinks are repaired before lymphocytes begin to replicate. Further, non-Hodgkin's lymphoma is not associated with formaldehyde exposure in human studies, which would argue against nasal tissue as a target of formaldehyde mutagenic effects.

6 Goldstein (2009) noted that although the hypothesis of formaldehyde inducing leukemias 7 through interaction with lymphoid cells in the nose could not be ruled out, it was not 8 supported by the rarity of chloromas (myeloid tumor cells) in the nasal cavity and the fact 9 that other nasal carcinogens such as nickel are not leukemogens. Chloromas, also called 10 granulocytic sarcomas or myeloid sarcomas, are rare tumors that can occur almost 11 anywhere in the body, including the head and neck (Prades et al. 2002). Occurrence of 12 these tumors in the nasal passages has been reported in a few instances (Prades et al. 13 2002, Sanford and Becker 1967, Scully et al. 1990).

#### 14 **5.8 Summary**

15 5.8.1 Adsorption, distribution, metabolism, and excretion

16 Formaldehyde is a metabolic intermediate that is essential for the biosynthesis of purines, 17 thymidine, and some amino acids. The metabolism of formaldehyde is similar in all 18 mammalian species studied. Differences in distribution following inhalation exposure can 19 be related to anatomical differences. For example, rats are obligate nose breathers while 20 monkeys and humans are oronasal breathers. Thus, in humans, some inhaled 21 formaldehyde will bypass the nasal passages and deposit directly into the lower 22 respiratory tract. The endogenous concentrations in the blood of humans, rats and 23 monkeys are about 2 to 3  $\mu$ g/g and do not increase after ingestion or inhalation of 24 formaldehyde from exogenous sources. Although formaldehyde is rapidly and almost 25 completely absorbed from the respiratory or gastrointestinal tracts, it is poorly absorbed 26 from intact skin. When absorbed after inhalation or ingestion, very little formaldehyde 27 reaches the systemic circulation because it is rapidly metabolized at the site of absorption 28 to formate, which is excreted in the urine or oxidized to carbon dioxide and exhaled. 29 Although the metabolic pathways are the same in all tissues, the data indicate that route 30 of absorption does affect the route of elimination. When inhaled, exhalation is the

primary route of elimination; however, when ingested, urinary excretion as formate is more important. Unmetabolized formaldehyde reacts non-enzymatically with sulfhydryl groups or urea, binds to tetrahydrofolate and enters the single-carbon intermediary metabolic pool, or reacts with macromolecules to form crosslinks (primarily between protein and single-stranded DNA).

6 5.8.2 Toxic effects

Formaldehyde is a highly reactive chemical that causes tissue irritation and damage on
contact. Because of its reactivity and rapid metabolism, toxicity is generally limited to
local effects. *In vitro* studies have demonstrated that formaldehyde is cytotoxic and
affects cell viability, cell differentiation and growth, cell proliferation, gene expression,
membrane integrity, mucociliary action, apoptosis, and thiol and ion homeostasis.
Furthermore, cells depleted of glutathione are more susceptible to formaldehyde toxicity.

13 Formaldehyde concentrations that have been associated with various toxic effects in 14 humans show wide interindividual variation and are route dependent. Symptoms are rare 15 at concentrations below 0.5 ppm; however, upper airway and eye irritation, changes in 16 odor threshold, and neurophysiological effects (e.g., insomnia, memory loss, mood 17 alterations, nausea, fatigue) have been reported at concentrations  $\leq 0.1$  ppm. The most 18 commonly reported effects include eye, nose, throat and skin irritation. Other effects 19 include allergic contact dermatitis, histopathological abnormalities (e.g., hyperplasia, 20 squamous metaplasia, and mild dysplasia) of the nasal mucosa, occupational asthma, 21 reduced lung function, and altered immune response. Some studies suggest that long-term 22 exposure to formaldehyde can decrease the number of white blood cells, and possibly 23 lower platelet and hemoglobin, and other studies have shown that formaldehyde exposure 24 affects changes in the percentage of lymphocyte subsets. Higher rates of spontaneous 25 abortion and low birth weights have been reported among women occupationally exposed 26 to formaldehyde. Oral exposure is rare, but there have been several suicides and 27 attempted suicides where individuals drank formaldehyde. These data indicate that the 28 lethal dose is 60 to 90 mL. Formaldehyde ingestion results in severe corrosive damage to 29 the gastrointestinal tract followed by CNS depression, myocardial depression, circulatory 30 collapse, metabolic acidosis, and multiple organ failure.

1 The toxic effects of formaldehyde in experimental animals include irritation, cytotoxicity,

- 2 and cell proliferation in the upper respiratory tract, ocular irritation, pulmonary
- 3 hyperactivity, bronchoconstriction, gastrointestinal irritation, and skin sensitization.
- 4 Histopathological lesions of the upper respiratory tract and cell proliferation have not
- 5 been reported at concentrations less that 2 ppm. Other reported effects include oxidative
- 6 stress, neurotoxicity, immunotoxicity, testicular toxicity, and decreased liver, thyroid
- 7 gland, and testis weights.

## 8 5.8.3 Carcinogenicity of metabolites and analogues

9 Formic acid (formate  $+ H^+$ ), the major metabolite of formaldehyde, has not been tested 10 for carcinogenic effects. Acetaldehyde, an analogue of formaldehyde, is listed as 11 reasonably anticipated to be a human carcinogen by the NTP. Acetaldehyde induced 12 respiratory tract tumors in rats (adenocarcinoma and squamous-cell carcinoma of the 13 nasal mucosa) and laryngeal carcinoma in hamsters. In addition, epidemiological data 14 provide some evidence that acetaldehyde may be associated with oral, esophageal, 15 pharyngeal, laryngeal, and bronchial tumors in humans. Glutaraldehyde and 16 benzaldehyde have also been tested for carcinogenicity in 2-year bioassays by the NTP. 17 Glutaraldehyde was not considered to be carcinogenic in rats and mice, and benzaldehyde 18 was not considered to be carcinogenic in rats. The NTP concluded that there was some 19 evidence of carcinogenicity for benzaldehyde in mice based on an increased incidence of 20 squamous-cell papillomas and hyperplasias in the forestomach of male and female mice.

21 5.8.4 Genetic and related effects

22 Formaldehyde is a direct-acting genotoxic compound that affects multiple gene

- 23 expression pathways, including those involved in DNA synthesis and repair and
- 24 regulation of cell proliferation. Most studies in bacteria were positive for forward or
- 25 reverse mutations without metabolic activation and for microsatellite induction. Studies
- 26 in non-mammalian eukaryotes and plants also were positive for forward and reverse
- 27 mutations, dominant lethal and sex-linked recessive lethal mutations, and DNA single-
- 28 strand breaks. In vitro studies with mammalian and human cells were positive for DNA
- 29 adducts, DNA-protein crosslinks, unscheduled DNA synthesis, single-strand breaks,
- 30 mutations, and cytogeneic effects (chromosomal aberrations, sister chromatid exchange,

1 and micronuclei induction). In *in vivo* studies, formaldehyde caused DNA-protein cross 2 links (in the nasal mucosa and fetal liver but not bone marrow), DNA strand breaks 3 (lymphocytes and liver), dominant lethal mutations, chromosomal aberrations 4 (pulmonary lavage cells and bone marrow in one of two studies), and micronuclei 5 induction in the gastrointestinal tract; however it did not induce sister chromatid 6 exchange or chromosomal aberrations in lymphocytes. P53 mutations were detected in 7 nasal squamous-cell carcinomas from rats. Inhalation exposure of formaldehyde also 8 induced DNA-protein cross links in the nasal turbinates, nasopharynx, trachea, and 9 bronchi of rhesus monkeys. In mice, formaldehyde exposure did not cause dominant 10 lethal mutations, micronuclei induction, or chromosomal aberrations when exposed by 11 intraperitoneal injection, but did induced heritable mutations when exposed by inhalation. 12 In studies of lymphocytes humans exposed to formaldehyde, increased frequencies of 13 chromosomal aberrations were observed in seven of twelve reviewed studies, sister

14 chromatid aberrations in six of thirteen studies, and micronuclei induction in fifteen of

15 sixteen studies reviewed. Increased frequencies of micronuclei were also observed in the

16 buccal or oral epithelium, nasal epithelium in all but one of the available studies. DNA-

17 protein cross links and DNA strand breaks have also been observed in lymphocytes from

- 18 medical personnel exposed to formaldehyde.
- 19 5.8.5 Mechanistic considerations

Although the biological mechanisms associated with formaldehyde-induced cancer are
not completely understood, it is important to recognize that chemicals can act through
multiple toxicity pathways and mechanisms to induce cancer or other health effects.
Potential carcinogenic modes of actions for formaldehyde include DNA reactivity
(covalent binding), gene mutation, chromosomal breakage, aneuploidy, and epigenetic
effects.

26 Studies evaluating nasal tumors in rats have shown that, regional dosimetry, genotoxicity,

27 and cytotoxicity are believed to be important factors. Computational fluid dynamics

28 models have been developed to predict and compare local flux values in the nasal

29 passages of rats, monkeys, and humans. Regions of the nasal passages with the highest

30 flux values are the regions most likely affected by formaldehyde exposure. Similar flux

1 values were predicted for rats and monkeys for regions of the nasal passages with 2 elevated cell proliferation rates, thus providing support for the hypothesis that 3 formaldehyde flux is a key factor for determining toxic response. Furthermore, DNA-4 protein crosslinks and cell-proliferation rates are correlated with the site specificity of 5 tumors. Cell proliferation is stimulated by the cytotoxic effects of formaldehyde. 6 Increased cell proliferation may contribute to carcinogenesis by increasing the probability 7 of spontaneous or chemically induced mutations. The dose-response curves for DNA-8 protein crosslinks, cell proliferation, and tumor formation show similar patterns with 9 sharp increases in slope at concentrations greater than 6 ppm. The observed sequence of 10 nasal lesions is as follows: rhinitis, epithelial dysplasia, squamous metaplasia and 11 hyperplasia, and squamous-cell carcinoma.

12 Biological mechanisms have been proposed for the possible association between 13 lymphohematopoietic cancers and formaldehyde exposure. Proposed mechanisms for 14 formaldehyde-induced leukemia are: (1) direct damage to stem cells in the bone marrow, 15 (2) damage to circulating stem cells, (3) damage to pluripotent stem cells present in the 16 nasal turbinate or olfactory mucosa. Evidence in support of the potential for DNA 17 damage to circulating hematopoietic stem cells is that DNA-protein crosslinks have been 18 identified in the nasal passages of laboratory animals exposed to formaldehyde and 19 increased micronuclei have been identified in the nasal and oral mucosa of formaldehyde-20 exposed humans. In addition, olfactory epithelial cells obtained from rat nasal passages 21 contain hematopoietic stem cells, which have been shown to re-populate the 22 heamtopoietic tissue of irradiated rats. However, some authors have questioned the 23 biologically plausibility of an association between formaldehyde exposure and leukemia, 24 because formaldehyde is rapidly metabolized and would not enter the systemic 25 circulation. They state that formaldehyde does not cause bone marrow toxicity or 26 pancytopenia, which are common features of known leukemogen, and that the genotoxic 27 and carcinogenic effects in animals and humans are limited to local effects.

# 6 References

- 1. Abdel Hameed AA, Khoder MI, Farag SA. 2000. Organic dust and gaseous contaminants at wood working shops. *J Environ Monit* 2(1): 73-6 (as cited in IARC 2006).
- 2. Acheson ED, Barnes HR, Gardner MJ, Osmond C, Pannett B, Taylor CP. 1984. Formaldehyde in the British chemical industry. An occupational cohort study. *Lancet* 1(8377): 611-616. (Supported by the Chemical Industries Association Formaldehyde Health Impact Study Team. Authors affiliated with Department of Health and Social Security, UK; Southampton General Hospital, UK.)
- 3. Ahman M, Alexandersson R, Ekholm U, Bergstrom B, Dahlqvist M, Ulfvarson U. 1991. Impeded lung function in moulders and coremakers handling furan resin sand. *Int Arch Occup Environ Health* 63(3): 175-80 (as cited in IARC 2006).
- 4. Ahmed S, Tsukahara S, Tin Tin Win S, Yamamoto S, Kunugita N, Arashidani K, Fujimaki H. 2007. Effects of low-level formaldehyde exposure on synaptic plasticity-related gene expression in the hippocampus of immunized mice. *J Neuroimmunol* 186(1-2): 104-11. (Supported by the Ministry of Education, Culture, Sports, Science and Technology, Japan. Authors affiliated with National Institute for Environmental Studies, Japan; University of Occupational and Environmental Health, Japan.)
- 5. Akbar-Khanzadeh F, Vaquerano MU, Akbar-Khanzadeh M, Bisesi MS. 1994. Formaldehyde exposure, acute pulmonary response, and exposure control options in a gross anatomy laboratory. *Am J Ind Med* 26(1): 61-75 (as cited in IARC 2006).
- 6. Akbar-Khanzadeh F, Mlynek JS. 1997. Changes in respiratory function after one and three hours of exposure to formaldehyde in non-smoking subjects. *Occup Environ Med* 54(5): 296-300 (as cited in IARC 2006).
- 7. Akron. 2009. *Formaldehyde, Anhydrous*. Department of Chemistry, University of Akron. <u>http://ull.chemistry.uakron.edu/erd/Chemicals/8000/6514.html</u>.
- Albert RE, Sellakumar AR, Laskin S, Kuschner M, Nelson N, Snyder CA. 1982. Gaseous formaldehyde and hydrogen chloride induction of nasal cancer in the rat. *J Natl Cancer Inst* 68(4): 597-603. (Supported by the National Cancer Institute and NIEHS. Authors affiliated with New York University Medical Center, NY.)
- 9. Alexandersson R, Hedenstierna G. 1988. Respiratory hazards associated with exposure to formaldehyde and solvents in acid-curing paints. *Arch Environ Health* 43(3): 222-7 (as cited in IARC 2006).

- 10. Altschuller AP, McPherson SP. 1963. Spectrophotometric analysis of aldehydes in the Los Angeles atomosphere. *J Air Pollut Control Assoc* 13(3): 109-111 (as cited in WHO 1989).
- 11. Andersen ME, Clewell HJ, 3rd, Bermudez E, Willson GA, Thomas RS. 2008. Genomic signatures and dose-dependent transitions in nasal epithelial responses to inhaled formaldehyde in the rat. *Toxicol Sci* 105(2): 368-83. (Supported by the Formaldehyde Council, Inc. Authors affiliated with The Hamner Institutes for Health Sciences, NC.)
- 12. Andersen SK, Jensen OM, Oliva D. 1982. [Exposure to formaldehyde and lung cancer in Danish physicians]. *Ugeskr Laeger* 144(21): 1571-1573. (Support and affiliations not identified due to foreign language.)
- 13. Anderson LG, Lanning JA, Barrell R, Miyagishima J, Jones RH, Wolfe P. 1996. Sources and sinks of formaldehyde and acetaldehyde: An analysis of Denvier's ambient concentration data. *Atmos Environ* 30: 2113-2123 (as cited in IARC 2006).
- Andersson M, Agurell E, Vaghef H, Bolcsfoldi G, Hellman B. 2003. Extendedterm cultures of human T-lymphocytes and the comet assay: a useful combination when testing for genotoxicity in vitro? *Mutat Res* 540(1): 43-55. (Supported by CFN, Astra AB, and AstraZeneca R&D, Sweden. Authors affiliated with Uppsala University, Sweden; AstraZeneca R&D, Sweden; University Hospital, Sweden.)
- Andjelkovich DA, Mathew RM, Richardson RB, Levine RJ. 1990. Mortality of iron foundry workers: I. Overall findings. *J Occup Med* 32(6): 529-540. (Supported by the Chemical Industry Institute of Toxicology. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)
- 16. Andjelkovich DA, Shy CM, Brown MH, Janszen DB, Levine RJ, Richardson RB. 1994. Mortality of iron foundry workers. III. Lung cancer case-control study. *J Occup Med* 36(12): 1301-1309. (Supported by the Chemical Industry Institute of Toxicology, NC. Authors affiliated with Chemical Industry Institute of Toxicology, NC; University of North Carolina, NC.)
- Andjelkovich DA, Janszen DB, Brown MH, Richardson RB, Miller FJ. 1995. Mortality of iron foundry workers: IV. Analysis of a subcohort exposed to formaldehyde. *J Occup Environ Med* 37(7): 826-837. (Supported by the Chemical Industry Institute of Toxicology, NC. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)
- Anwar MI, Khan MZ, Muhammad G, Bachaya A, Babar AM. 2001. Effects of dietary formalin on the health and testicular pathology of male Japanese quails (Coturnix coturnix Japonica). *Vet Hum Toxicol* 43(6): 330-333. (Support not reported. Authors affiliated with University of Agriculture, Pakistan.)

- 19. Appelman LM, Woutersen RA, Zwart A, Falke HE, Feron VJ. 1988. One-year inhalation toxicity study of formaldehyde in male rats with a damaged or undamaged nasal mucosa. *J Appl Toxicol* 8(2): 85-90. (Supported by the 'Koningin Wilhelmina Fonds' (Netherlands Cancer Foundation), Amsterdam, The Netherlands. Authors affiliated with TNO-CIVO Toxicology and Nutrition Institute, Netherlands; Minsitry of Social Affairs and Employment, Netherlands.)
- 20. Armstrong RW, Imrey PB, Lye MS, Armstrong MJ, Yu MC, Sani S. 2000. Nasopharyngeal carcinoma in Malaysian Chinese: occupational exposures to particles, formaldehyde and heat. *Int J Epidemiol* 29(6): 991-998. (Supported by NCI. Authors affiliated with University of Illinois, IL; Institute for Medical Research, Malaysia; University of Southern California School of Medicine, CA; Universiti Kebangsaan Malaysia, Malaysia.)
- Arts JH, Rennen MA, de Heer C. 2006. Inhaled formaldehyde: evaluation of sensory irritation in relation to carcinogenicity. *Regul Toxicol Pharmacol* 44(2): 144-60. (Supported by the Dutch Chemical Industry Association. Authors affiliated with TNO Quality of Life, Netherlands.)
- 22. Aslan H, Songur A, Tunc AT, Ozen OA, Bas O, Yagmurca M, Turgut M, Sarsilmaz M, Kaplan S. 2006. Effects of formaldehyde exposure on granule cell number and volume of dentate gyrus: a histopathological and stereological study. *Brain Res* 1122(1): 191-200. (Support not reported. Authors affiliated with Gaziosmanpasa University School of Medicine, Turkey; Afyon Kocatepe University School of Medicine, Turkey; Adnan Menderes University School of Medicine, Turkey; Firat University School of Medicine, Turkey; Ondokuz Mayis University School of Medicine, Turkey.)
- 23. ATSDR. 1999. *Toxicological Profile for Formaldehyde*. U.S. Department of Health and Human Services, Agency for Toxic Substances and Disease Registry. <u>http://www.atsdr.cdc.gov/toxprofiles/tp111.pdf</u>.
- 24. ATSDR. 2007a. *Formaldehyde Sampling at FEMA Temporary Housing Units, Baton Rouge, LA*. Atlanta, GA: Agency for Toxic Substances and Disease Registry. 14 pp.
- 25. ATSDR. 2007b. *HazDat Database*. Agency for Toxic Substances and Disease Registry. <u>http://www.atsdr.cdc.gov/hazdat.html</u>. Accessed on 8/08/07.
- Ballarin C, Sarto F, Giacomelli L, Bartolucci GB, Clonfero E. 1992. Micronucleated cells in nasal mucosa of formaldehyde-exposed workers. *Mutat Res* 280(1): 1-7. (Support not reported. Authors affiliated with University of Padua, Italy; Occupational Health Inspectorate, Italy; Cytodiagnostic Service, ULSS, Italy.)

- 27. Ballenger JJ. 1984. Some effects of formaldehyde on the upper respiratory tract. *Laryngoscope* 94(11 Pt 1): 1411-1413. (Support not reported. Authors affiliated with Evanston Hospital, IL.)
- 28. Barnes L, Eveson JW, Reichart P, Sidransky D. 2005. World Health Organization classification of tumours. Pathology and genetics of head and neck tumours, Lyon, France: IARC Press.
- 29. Barry JL, Tomé D. 1991. Formaldehyde content of milk in goats fed formaldehyde-treated soybean oil-meal. *Food Addit Contam* 8(5): 633-640. (Support not reported. Authors affiliated with Institut National de la Recherche Agronomique, France.)
- Bartnik FG, Gloxhuber C, Zimmermann V. 1985. Percutaneous absorption of formaldehyde in rats. *Toxicol Lett* 25(2): 167-172. (Support not reported. Authors affiliated with Toxicological Laboratories, Germany.)
- 31. Bartone NF, Grieco RV, Herr BS, Jr. 1968. Corrosive gastritis due to ingestion of formaldehyde: without esophageal impairment. *Jama* 203(1): 50-1. (Support not reported. Authors affiliated with Methodist Hospital, NY.)
- 32. BASF. 2006. *Trioxane*. Cinetic Internet Systemhaus GmbH. http://www2.basf.de/basf2/html/e/produkte/gebiete/trioxan/html/03\_application. htm. Last accessed: 7/28/06.
- Basler A, v d Hude W, Scheutwinkel-Reich M. 1985. Formaldehyde-induced sister chromatid exchanges in vitro and the influence of the exogenous metabolizing systems S9 mix and primary rat hepatocytes. *Arch Toxicol* 58(1): 10-13. (Supported by the Umweltbundesamt. Authors affiliated with Max von Pettenkofer Institut, Germany.)
- 34. Bauchinger M, Schmid E. 1985. Cytogenetic effects in lymphocytes of formaldehyde workers of a paper factory. *Mutat Res* 158(3): 195-9. (Support not reported. Authors affiliated with Abteilung fur Strahlenbiologie, Germany.)
- 35. Beall JR, Ulsamer AG. 1984. Formaldehyde and hepatotoxicity: a review. *J Toxicol Environ Health* 14(1): 1-21. (Support not reported. Authors affiliated with U.S. Department of Energy; U.S. Consumer Product Safety Commission, MD.)
- 36. Beane Freeman LE, Blair A, Lubin JH, Stewart PA, Hayes RB, Hoover RN, Hauptmann M. 2009. Mortality from lymphohematopoietic malignancies among workers in formaldehyde industries: the National Cancer Institute Cohort. *J Natl Cancer Inst* 101(10): 751-61. (Supported my NIH. Authors affiliated with NIH, MD; Stewart Exposure Assessments, LLC, VA; Netherlands Cancer Institute, Netherlands.)

- 37. Beland FA, Fullerton NF, Heflich RH. 1984. Rapid isolation, hydrolysis and chromatography of formaldehyde-modified DNA. *J Chromatogr* 308: 121-131. (Supported by the U.S. Consumer Product Safety Commission and the Veteran's Administration Hospital, AR. Authors affiliated with the National Center for Toxicological Research, AR.)
- 38. Bender J. 2002. The use of noncancer endpoints as a basis for establishing a reference concentration for formaldehyde. *Regul Toxicol Pharmacol* 35(1): 23-31 (as cited in IARC 2006).
- 39. Berk JV, Hollowell CD, Pepper JH, Young RA. 1980. Impact of reduced ventilation on indoor air quality in residential buildings. In *73rd Annual Meeting of the Air Pollution Control Association, Montreal, Quebec, 22-27 June, 1980* (as cited in WHO 1989).
- 40. Berke JH. 1987. Cytologic examination of the nasal mucosa in formaldehydeexposed workers. *J Occup Med* 29(8): 681-684. (Support not reported. Authors affiliated with W.R. Grace & Company, MA.)
- 41. Bermudez E, Chen Z, Gross EA, Walker C, Recio L, Pluta L, Morgan K. 1994. Characterization of cell lines derived from formaldehyde-induced nasal tumors in rats. *Mol Carcinogenesis* 9: 193-199. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC; Cancer Research Institute, China; University of Texas, TX.)
- 42. Berrino F, Richiardi L, Boffetta P, Estève J, Belletti I, Raymond L, Troschel L, Pisani P, Zubiri L, Ascunce N, Gubéran E, Tuyns A, Terracini B, Merletti F. 2003. Occupation and larynx and hypopharynx cancer: a job-exposure matrix approach in an international case-control study in France, Italy, Spain and Switzerland. *Cancer Causes Control* 14(3): 213-223. (Supported by the National Institute for Research on Alcohol Abuse and Alcoholism, the City of Turin: Progetto Finalizzato 'Oncologia', the Associazione Italiana per la Ricerca sul Cancro, CPO-Piemonte, MURST, Special Project 'Oncology," Compagnia di Sa Paolo/FIRMS, the Province of Varese: CNR 'Progetto Finalizzato Oncologia,' the Ministy of Health, Geneva: Fonds national de la recherche scientifique and the Ligue Suisse contre le Cancer. Authors affiliated with National Cancer Institute, Italy; University of Turin, Italy; Karolinska Institutet, Sweden; IARC; Lyon University, France; Geneva Cancer Registry, Switzerland; Zaragoza Cancer Registry, Spain; Institute of Public Health, Spain; Medical Inspectorate of Factories, Switzerland; Milan Job Exposure Matrix Working Group, Italy.)
- 43. Bertazzi PA, Pesatori AC, Radice L, Zocchetti C, Vai T. 1986. Exposure to formaldehyde and cancer mortality in a cohort of workers producing resins. *Scand J Work Environ Health* 12(5): 461-8. (Support not reported. Authors affiliated with University of Milan, Italy; Universita degli Studi, Italy.)

- 44. Bertazzi PA, Pesatori A, Guercilena S, Consonni D, Zocchetti C. 1989. [Carcinogenic risk for resin producers exposed to formaldehyde: extension of follow-up]. *Med Lav* 80(2): 111-122. (Support not identified due to foreign language. Authors affiliated with Universita degli Studi di Milano, Italy; Clinica del Lavoro, Italy.)
- 45. Bizzari SN. 2007. *CEH Marketing Research Report: Formaldehyde*. 658.5000 A. SRI International. 106 pp. (Support and affiliations not reported.)
- 46. Blade LM. 1983. Occupational Exposure to Formaldehyde: Recent NIOSH Involvement. In *Formaldehyde*. Clary JL, Gibson JE, Waritz RS, eds. New York: Marcel Dekker. p. 1-30 (as cited in WHO 1989).
- 47. Blair A, Stewart P, O'Berg M, Gaffey W, Walrath J, Ward J, Bales R, Kaplan S, Cubit D. 1986. Mortality among industrial workers exposed to formaldehyde. *J Natl Cancer Inst* 76(6): 1071-1084. (Support not reported. Authors affiliated with NIH; E.I. du Pont de Nemours & Company, Inc., DE; Health Research Committe of the Formaldehyde Institute; Monsanto Co., MO; WESTAT, Inc., MD; Dynamac Corp., MD.)
- 48. Blair A, Stewart PA. 1989. Comments on the reanalysis of the National Cancer Institute study of workers exposed to formaldehyde. *J Occup Med* 31(11): 881-884. (Support not reported. Authors affiliated with NCI.)
- 49. Blair A, Stewart PA, Hoover RN. 1990b. Mortality from lung cancer among workers employed in formaldehyde industries. *Am J Ind Med* 17(6): 683-99. (Support not reported. Authors affiliated with NCI.)
- 50. Blair A, Zheng T, Linos A, Stewart PA, Zhang YW, Cantor KP. 2001. Occupation and leukemia: a population-based case-control study in Iowa and Minnesota. *Am J Ind Med* 40(1): 3-14. (Support not reported. Authors affiliated with NCI; Yale University School of Public Health, CT; University of Athens, Greece.)
- Blair A, Purdue MP, Weisenburger DD, Baris D. 2007. Chemical exposures and risk of chronic lymphocytic leukaemia. *Br J Haematol* 139(5): 753-761. (Supported by NIH. Authors affiliated with NCI; University of Nebraska Medical Center, NE.)
- 52. Blasiak J, Trzeciak A, Malecka-Panas E, Drzewoski J, Wojewódzka M. 2000. *In vitro* genotoxicity of ethanol and acetaldehyde in human lymphocytes and the gastrointestinal tract mucosa cells. *Toxicol In Vitro* 14(4): 287-295. (Supported by the Committee of Scientific Research. Authors affiliated with University of Lodz, Poland; Medical University of Lodz, Poland; Institute of Nuclear Chemistry and Technology, Poland.)
- 53. Boffetta P, Stellman SD, Garfinkel L. 1989. A case-control study of multiple myeloma nested in the American Cancer Society prospective study. *Int J Cancer*

43(4): 554-9. (Support not reported. Authors affiliated with American Cancer Society, NY; University of Torino, Italy.)

- 54. Boillot C, Bazin C, Tissot-Guerraz F, Droguet J, Perraud M, Cetre JC, Trepo D, Perrodin Y. 2008. Daily physicochemical, microbiological and ecotoxicological fluctuations of a hospital effluent according to technical and care activities. *Sci Total Environ* 403(1-3): 113-29. (Support not reported. Authors affiliated with Universite de Lyon, France; Hospices Civils de Lyon, France.)
- 55. Bolstad-Johnson DM, Burgess JL, Crutchfield CD, Storment S, Gerkin R, Wilson JR. 2000. Characterization of firefighter exposures during fire overhaul. *Aihaj* 61(5): 636-41 (as cited in IARC 2006).
- 56. Bolt HM. 1987. Experimental toxicology of formaldehyde. *J Cancer Res Clin Oncol* 113(4): 305-309. (Support not reported. Authors affiliated with Universitat Dormund, Germany.)
- 57. Bond GG, Flores GH, Shellenberger RJ, Cartmill JB, Fishbeck WA, Cook RR. 1986. Nested case-control study of lung cancer among chemical workers. *Am J Epidemiol* 124(1): 53-66. (Support not reported. Authors affiliated with Dow Chemical, MI; Dow Chemical, TX; University of Michigan, MI. )
- 58. Bono R, Vincenti M, Schiliro T, Scursatone E, Pignata C, Gilli G. 2006. N-Methylenvaline in a group of subjects occupationally exposed to formaldehyde. *Toxicol Lett* 161(1): 10-7. (Supported by the Ministero dell'Universita, della Ricerca Scientifica e Technologica. Authors affiliated with University of Torino, Italy.)
- 59. Bosetti C, McLaughlin JK, Tarone RE, Pira E, La Vecchia C. 2008. Formaldehyde and cancer risk: a quantitative review of cohort studies through 2006. Ann Oncol 19(1): 29-43. (Supported by the Italian Association for Cancer Research and the Italian League Against Cancer. Authors affiliated with Istituto di Ricerche Farmacologiche Mario Negri, Italy; International Epidemiology Institute, MD; Vanderbilt University Medical Center, TN; Universita di Torino, Turin; Universita degli Studi di Milano, Italy.)
- 60. Bostrom CE, Almen J, Steen B, Westerholm R. 1994. Human exposure to urban air pollution. *Environ Health Perspect* 102 Suppl 4: 39-47. (Support not reported. Authors affiliated with Swedish Environmental Protection Agency; Swedish Motor Vehicle Inspection Company; Swedish Environment Research Institute; Stockholm University, Sweden.)
- Boysen M, Zadig E, Digernes V, Abeler V, Reith A. 1990. Nasal mucosa in workers exposed to formaldehyde: a pilot study. *Br J Ind Med* 47(2): 116-21. (Supported by the Norwegian Cancer Society. Authors affiliated with University of Oslo, Norway; Dyno Industries Ltd., Lillestöm; Norwegian Radium Hospital, Norway.)

- 62. Brandt-Rauf PW, Fallon LF, Jr., Tarantini T, Idema C, Andrews L. 1988. Health hazards of fire fighters: exposure assessment. *Br J Ind Med* 45(9): 606-12 (as cited in IARC 2006).
- 63. Bray F, Haugen M, Moger TA, Tretli S, Aalen OO, Grotmol T. 2008. Ageincidence curves of nasopharyngeal carcinoma worldwide: bimodality in lowrisk populations and aetiologic implications. *Cancer Epidemiol Biomarkers Prev* 17(9): 2356-2365. (Supported by Statistics for Innovation. Authors affiliated with Cancer Registry of Norway, Norway; University of Oslo, Norway.)
- 64. Breysse PA. 1984. Formaldehyde levels and accompanying symptoms associated with individuals residing in over 1000 conventional and mobile homes in the state of Washington. *Indoor Air* 3: 403-408 (as cited in WHO 1989).
- 65. Brickus LSR, Cardoso JN, Aquino Neto FR. 1998. Distributions of indoor and outdoor air pollutants in Rio de Janeiro, Brazil; Implications to indoor air quality in bayside offices. *Environ Sci Technol* 32: 3485-3490 (as cited in IARC 2006).
- 66. Brinton LA, Blot WJ, Becker JA, Winn DM, Browder JP, Farmer JC, Jr., Fraumeni JF, Jr. 1984. A case-control study of cancers of the nasal cavity and paranasal sinuses. *Am J Epidemiol* 119(6): 896-906. (Support not reported. Authors affiliated with NCI; University of North Carolina Medical Center, NC; Duke University Medical Center, NC.)
- 67. Broder I, Corey P, Cole P, Lipa M, Mintz S, Nethercott JR. 1988. Comparison of health of occupants and characteristics of houses among control homes and homes insulated with urea formaldehyde foam. II. Initial health and house variables and exposure-response relationships. *Environ Res* 45(2): 156-78. (Supported by Health and Welfare Canada. Authors affiliated with University of Toronto, Canada. )
- 68. Broder I, Corey P, Brasher P, Lipa M, Cole P. 1991. Formaldehyde exposure and health status in households. *Environ Health Perspect* 95: 101-4. (Supported by Health and Welfare Canada. Authors affiliated with University of Toronto, Canada.)
- Brownson RC, Alavanja MC, Chang JC. 1993. Occupational risk factors for lung cancer among nonsmoking women: a case-control study in Missouri (United States). *Cancer Causes Control* 4(5): 449-454. (Supported by NCI. Authors affiliated with Missouri Department of Health; NCI.)
- 70. Buckley KE, Fisher LJ, MacKay VG. 1988. Levels of formaldehyde in milk, blood, and tisues of dairy cows and calves consuming formalin-treated whey. J Agric Food Chem 36: 1146-1150. (Supported by Agriculture Canada under the auspices of the Feed from Waste Program. Authors affiliated with Agriculture Canada.)

- Burgaz S, Cakmak G, Erdem O, Yilmaz M, Karakaya AE. 2001. Micronuclei frequencies in exfoliated nasal mucosa cells from pathology and anatomy laboratory workers exposed to formaldehyde. *Neoplasma* 48(2): 144-147. (Support not reported. Authors affiliated with Gazi University, Turkey.)
- 72. Burgaz S, Erdem O, Cakmak G, Erdem N, Karakaya A, Karakaya AE. 2002. Cytogenetic analysis of buccal cells from shoe-workers and pathology and anatomy laboratory workers exposed to n-hexane, toluene, methyl ethyl ketone and formaldehyde. *Biomarkers* 7(2): 151-61. (Support not reported. Authors affiliated with Gazi University, Turkey; National Institute of Occupational Safety and Health, Turkey; Ankara University, Turkey.)
- 73. Burge PS, Harries MG, Lam WK, O'Brien IM, Patchett PA. 1985. Occupational asthma due to formaldehyde. *Thorax* 40(4): 255-60. (Support not reported. Authors affiliated with East Birmingham Hospital, UK; Cardiothoracic Institute, UK.)
- 74. Burkhart KK, Kulig KW, McMartin KE. 1990. Formate levels following a formalin ingestion. *Vet Hum Toxicol* 32(2): 135-7. (Support not reported. Authors affiliated with University of Colorado, CO; Louisiana State University Medical Center, LA.)
- 75. Callas PW, Pastides H, Hosmer DW, Jr. 1996. Lung cancer mortality among workers in formaldehyde industries. *J Occup Environ Med* 38(8): 747-8; discussion 749-51. (Support not reported. Authors affiliated with University of Massachusetts, MA.)
- 76. Cantor KP, Stewart PA, Brinton LA, Dosemeci M. 1995. Occupational exposures and female breast cancer mortality in the United States. *J Occup Environ Med* 37(3): 336-348. (Support not reported. Authors affiliated with NCI.)
- Carlson RM, Smith MC, Nedorost ST. 2004. Diagnosis and treatment of dermatitis due to formaldehyde resins in clothing. *Dermatitis* 15(4): 169-75. (Support not reported. Authors affiliated with University Hospitals of Cleveland/Case Western Reserve University, OH.)
- Carraro E, Gasparini S, Gilli G. 1999. Identification of a chemical marker of environmental exposure to formaldehyde. *Environ Res* 80(2 Pt 1): 132-7. (Support not reported. Authors affiliated with University of Turin, Italy.)
- 79. Casanova-Schmitz M, Starr TB, Heck HD. 1984a. Differentiation between metabolic incorporation and covalent binding in the labeling of macromolecules in the rat nasal mucosa and bone marrow by inhaled [<sup>14</sup>C]- and [<sup>3</sup>H]formaldehyde. *Toxicol Appl Pharmacol* 76(1): 26-44. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)
- 80. Casanova-Schmitz M, David RM, Heck HD. 1984b. Oxidation of formaldehyde and acetaldehyde by NAD<sup>+</sup>-dependent dehydrogenases in rat nasal mucosal homogenates. *Biochem Pharmacol* 33(7): 1137-1142. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC. )
- Casanova M, Heck Hd A. 1987. Further studies of the metabolic incorporation and covalent binding of inhaled [<sup>3</sup>H]- and [<sup>14</sup>C]formaldehyde in Fischer-344 rats: effects of glutathione depletion. *Toxicol Appl Pharmacol* 89(1): 105-121. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)
- 82. Casanova M, Heck HD, Everitt JI, Harrington WW, Jr., Popp JA. 1988. Formaldehyde concentrations in the blood of rhesus monkeys after inhalation exposure. *Food Chem Toxicol* 26(8): 715-716. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)
- 83. Casanova M, Deyo DF, Heck HD. 1989. Covalent binding of inhaled formaldehyde to DNA in the nasal mucosa of Fischer 344 rats: analysis of formaldehyde and DNA by high-performance liquid chromatography and provisional pharmacokinetic interpretation. *Fundam Appl Toxicol* 12(3): 397-417. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)
- 84. Casanova M, Morgan KT, Steinhagen WH, Everitt JI, Popp JA, Heck HD. 1991. Covalent binding of inhaled formaldehyde to DNA in the respiratory tract of rhesus monkeys: pharmacokinetics, rat-to-monkey interspecies scaling, and extrapolation to man. *Fundam Appl Toxicol* 17(2): 409-428. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)
- 85. Casanova M, Morgan KT, Gross EA, Moss OR, Heck HA. 1994. DNA-protein cross-links and cell replication at specific sites in the nose of F344 rats exposed subchronically to formaldehyde. *Fundam Appl Toxicol* 23(4): 525-536. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)
- 86. Casanova M, Bell DA, Heck HD. 1997. Dichloromethane metabolism to formaldehyde and reaction of formaldehyde with nucleic acids in hepatocytes of rodents and humans with and without glutathione S-transferase T1 and M1 genes. Fundam Appl Toxicol 37(2): 168-180. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC; NIEHS.)
- 87. Casanova M, Heck HA. 1997. Lack of evidence for the involvement of formaldehyde in the hepatocarcinogenicity of methyl *tertiary*-butyl ether in CD-1 mice. *Chem Biol Interact* 105(2): 131-143. (Supported by the Oxygenated Fuels Association. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)

- 88. CDC. 2008. *Final Report on Formaldehyde Levels in FEMA-Supplied Travel Trailers, Park Models and Mobile Homes*. Center for Disease Control and Prevention. 61 pp.
- Chang JC, Gross EA, Swenberg JA, Barrow CS. 1983. Nasal cavity deposition, histopathology, and cell proliferation after single or repeated formaldehyde exposures in B6C3F1 mice and F-344 rats. *Toxicol Appl Pharmacol* 68(2): 161-176. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology. NC.)
- Chaw YF, Crane LE, Lange P, Shapiro R. 1980. Isolation and identification of cross-links from formaldehyde-treated nucleic acids. *Biochemistry* 19(24): 5525-5531. (Supported by NIH. Authors affiliated with New York University, NY; Academica Sinica, China.)
- 91. Chebotarev AN, Titenko NV, Selezneva TG, etal. 1986. Comparison of chromosome aberrations, sister chromatid exchanges, and unscheduled DNA synthesis when evaluating the mutagenicity of environmental factors. *Cytol Genet* 20(2): 21-26. (Support and affiliations not identified due to foreign language.)
- 92. ChemIDPlus. 2009a. *Formaldehyde*. National Library of Medicine. <u>http://chem.sis.nlm.nih.gov/chemidplus/</u> and search "formaldehyde". Accessed on 5/19/09.
- 93. ChemIDPlus. 2009b. *1,3,5-Trioxane*. National Library of Medicine. <u>http://chem.sis.nlm.nih.gov/chemidplus/</u> and search "1,3,5-Trioxane". Accessed on 5/19/09.
- 94. ChemSources. 2009a. *Formaldehyde*. Chemical Sources International. <u>http://www.chemsources.com/</u>. Accessed on 4/30/09.
- 95. ChemSources. 2009b. *Paraformaldehyde*. Chemical Sources International. <u>http://www.chemsources.com/</u>. Accessed on 4/30/09.
- 96. ChemSources. 2009c. *Trioxane*. Chemical Sources International. <u>http://www.chemsources.com/</u>. Accessed on 4/30/09.
- 97. Chen J, So S, Lee H, Fraser MP, Curl RF, Harman T, Tittel FK. 2004. Atmospheric formaldehyde monitoring in the Greater Houston area in 2002. *Appl Spectrosc* 58(2): 243-247. (Supported by the Dreyfus Foundation, the Welch Foundation, the National Science Foundation, and the Gulf Coast Hazardous Substance Research Center. Authors affiliated with Rice University, TX; University of Houston-Clear Lake, TX.)
- Chen SC, Wong RH, Shiu LJ, Chiou MC, Lee H. 2008. Exposure to mosquito coil smoke may be a risk factor for lung cancer in Taiwan. *J Epidemiol* 18(1): 19-25. (Support not reported. Authors affiliated with Chung Shan Medical

University, Taiwan; Central Taiwan University of Science and Technology, Taiwan.)

- 99. Cheng G, Wang M, Upadhyaya P, Villalta PW, Hecht SS. 2008. Formation of formaldehyde adducts in the reactions of DNA and deoxyribonucleosides with alpha-acetates of 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone (NNK), 4- (methylnitrosamino)-1-(3-pyridyl)-1-butanol (NNAL), and N- nitrosodimethylamine (NDMA). *Chem Res Toxicol* 21(3): 746-51. (Supported by the National Cancer Institute. Authors affiliated with University of Minnesota, MN.)
- 100. Cheng Y, Ying C, Yan W, Zhang J, Zhang M. 1995. Nasal mucosa epithelial cell and lymphoblast micronuclei of formaldehyde exposed population. *Sanitation and Antiepidemic Station of Jingsha City* 14(5): 293-295 (as cited in Tang *et al.* 2009).
- 101. Cheng Z, Li Y, Liang B, Wang C. 2004. Investigation of formaldehyde level and health of personnel in clinical pathology. *J Bengbu Med Coll* 29(3): 266-267 (as cited in Tang *et al.* 2009).
- 102. Chiazze L, Jr., Watkins DK, Fryar C, Kozono J. 1993. A case-control study of malignant and non-malignant respiratory disease among employees of a fibreglass manufacturing facility. II. Exposure assessment. *Br J Ind Med* 50(8): 717-725. (Supported by Owens-Corning Fiberglas Corporation. Authors affiliated with Georgetown University School of Medicine, Washington, DC.)
- 103. Chiazze L, Jr., Watkins DK, Fryar C. 1997. Historical cohort mortality study of a continuous filament fiberglass manufacturing plant. I. White men. J Occup Environ Med 39(5): 432-441. (Supported by Owens Corning. Authors affiliated with Georgetown University Medical Center, Washington, DC.)
- 104. Chowdhury AR, Gautam AK, Patel KG, Trivedi HS. 1992. Steroidogenic inhibition in testicular tissue of formaldehyde exposed rats. *Indian J Physiol Pharmacol* 36(3): 162-168. (Supported by the Indian Council of Medical Research. Authors affiliated with National Institute of Occupational Health, India.)
- 105. Chung KY, Cuthbert RJ, Revell GS, Wassel SG, Summers N. 2000. A study on dust emission, particle size distribution and formaldehyde concentration during machining of medium density fibreboard. *Ann Occup Hyg* 44(6): 455-66. (as cited in IARC 2006).
- 106. Cleveland WS, Graedel TE, Kleiner B. 1977. Urban formaldehyde: Observed correlation with source emissions and photochemistry. *Atmos Environ* 11: 357-360 (as cited in WHO 1989).
- 107. Coggon D, Pannett B, Acheson ED. 1984. Use of job-exposure matrix in an occupational analysis of lung and bladder cancers on the basis of death

certificates. *J Natl Cancer Inst* 72(1): 61-65. (Support not reported. Authors affiliated with University of Southampton, UK.)

- 108. Coggon D, Harris EC, Poole J, Palmer KT. 2003. Extended follow-up of a cohort of British chemical workers exposed to formaldehyde. *J Natl Cancer Inst* 95(21): 1608-1615. (Supported by the Colt Foundation. Authors affiliated with University of Southampton, UK.)
- 109. Cohen Hubal EA, Schlosser PM, Conolly RB, Kimbell JS. 1997. Comparison of inhaled formaldehyde dosimetry predictions with DNA-protein cross-link measurements in the rat nasal passages. *Toxicol Appl Pharmacol* 143(1): 47-55. (Supported by the U.S. EPA. Authors affiliated with Chemical Industry Institute of Toxicology, NC; North Carolina State University, NC.)
- 110. Coldiron VR, Ward JB, Jr., Trieff NM, Janssen HE, Jr., Smith JH. 1983. Occupational exposure to formaldehyde in a medical center autopsy service. J Occup Med 25(7): 544-8 (as cited in IARC 2006).
- 111. Cole P, Axten C. 2004. Formaldehyde and leukemia: an improbable causal relationship. *Regul Toxicol Pharmacol* 40(2): 107-112. (Supported by the Formaldehyde Council. Authors affiliated with University of Alabama, AL; Health Risk Solutions, LLC, VA.)
- 112. Collins JJ, Acquavella JF, Esmen NA. 1997. An updated meta-analysis of formaldehyde exposure and upper respiratory tract cancers. *J Occup Environ Med* 39(7): 639-651. (Supported by the Society for Plastic Industries. Authors affiliated with Monsanto Company, MO; University of Oklahoma City, OK.)
- 113. Collins JJ, Esmen NA, Hall TA. 2001a. A review and meta-analysis of formaldehyde exposure and pancreatic cancer. *Am J Ind Med* 39(3): 336-345. (Support not reported. Authors affiliated with Solutia Inc., MO; University of Oklahoma, OK.)
- 114. Collins JJ, Ness R, Tyl RW, Krivanek N, Esmen NA, Hall TA. 2001b. A review of adverse pregnancy outcomes and formaldehyde exposure in human and animal studies. *Regul Toxicol Pharmacol* 34(1): 17-34. (Support not reported. Authors affiliated with Solutia Inc., PA; University of Pittsburgh, PA; Research Triangle Institute, NC; Dupont; University of Oklahoma, OK.)
- 115. Collins JJ, Lineker GA. 2004. A review and meta-analysis of formaldehyde exposure and leukemia. *Regul Toxicol Pharmacol* 40(2): 81-91. (Support not reported. Authors affiliated with Dow Chemical, MI; Peccavis Consulting.)
- 116. Conaway CC, Whysner J, Verna LK, Williams GM. 1996. Formaldehyde mechanistic data and risk assessment: endogenous protection from DNA adduct formation. *Pharmacol Ther* 71(1-2): 29-55. (Supported by NCI. Authors affiliated with American Health Foundation, NY.)

- 117. Conner AH. 2001. Wood: Adhesives. In *Encyclopedia of Materials: Science and Technology*. Elsevier Science, Ltd. p. 9583 9599. (Support and author affiliations not reported.)
- 118. Connor TH, Ward JB, Jr., Legator MS. 1985a. Absence of mutagenicity in the urine of autopsy service workers exposed to formaldehyde: factors influencing mutagenicity testing of urine. *Int Arch Occup Environ Health* 56(3): 225-237. (Supported by the U.S. EPA and NIH. Authors affiliated with University of Texas, TX.)
- 119. Conolly RB, Lilly PD, Kimbell JS. 2000. Simulation modeling of the tissue disposition of formaldehyde to predict nasal DNA-protein cross-links in Fischer 344 rats, rhesus monkeys, and humans. *Environ Health Perspect* 5: 919-924. (Supported by the member companies of the Chemical Industry Institute of Toxicology. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)
- 120. Conolly RB, Kimbell JS, Janszen D, Schlosser PM, Kalisak D, Preston J, Miller FJ. 2003. Biologically motivated computational modeling of formaldehyde carcinogenicity in the F344 rat. *Toxicol Sci* 75(2): 432-447. (Support not reported. Authors affiliated with CIIT Centers for Health Research, NC; Wyeth-Ayerst Research, PA; U.S. EPA.)
- 121. Conolly RB, Kimbell JS, Janszen D, Schlosser PM, Kalisak D, Preston J, Miller FJ. 2004. Human respiratory tract cancer risks of inhaled formaldehyde: dose-response predictions derived from biologically-motivated computational modeling of a combined rodent and human dataset. *Toxicol Sci* 82(1): 279-296. (Support not reported. Authors affiliated with CIIT Centers for Health Research, NC; U.S. EPA; Wyeth-Ayerst Research, PA.)
- 122. ConsensusWorkshop. 1984. Report on the Consensus Workshop on Formaldehyde. *Environ Health Perspect* 58: 323-81.
- 123. Cosma GN, Marchok AC. 1988. Benzo[*a*]pyrene- and formaldehyde-induced DNA damage and repair in rat tracheal epithelial cells. *Toxicology* 51(2-3): 309-320. (Supported by the National Cancer Institute, U.S. Dept. of Energy, Martin Marietta Energy Systems, Inc. and NIH. Authors affiiated with University of Tennessee, TN; Oak Ridge National Laboratory, TN.)
- 124. Cosma GN, Jamasbi R, Marchok AC. 1988a. Growth inhibition and DNA damage induced by benzo[a]pyrene and formaldehyde in primary cultures of rat tracheal epithelial cells. *Mutat Res* 201(1): 161-8. (Supported by the National Cancer Institute, the U.S. Dept. of Energy and NIH. Authors affiliated with Oak Ridge National Laboratory, TN; Bowling Green State University, OH.)
- 125. Cosma GN, Wilhite AS, Marchok AC. 1988b. The detection of DNA-protein cross-links in rat tracheal implants exposed in vivo to benzo[*a*]pyrene and formaldehyde. *Cancer Lett* 42(1-2): 13-21. (Supported by the National Cancer

Institute, NIH and the U.S. Department of Energy. Authors affiliated with Oak Ridge National Laboratory, TN; New York University Medical Center, NY.)

- 126. Costa M, Zhitkovich A, Harris M, Paustenbach D, Gargas M. 1997. DNAprotein cross-links produced by various chemicals in cultured human lymphoma cells. *J Toxicol Environ Health* 50(5): 433-449. (Support not reported. Authors affiliated with New York University Medical Center, NY; Maxus Energy Corporation, NJ; ChemRisk Division of McLaren/Hart, OH.)
- 127. Costa S, Coelho P, Costa C, Silva S, Mayan O, Santos LS, Gaspar J, Teixeira JP. 2008. Genotoxic damage in pathology anatomy laboratory workers exposed to formaldehyde. *Toxicology* 252(1-3): 40-8. (Supported by Fundacao Calouste Gulbenkian. Authors affiliated with National Institute of Health, Portugal; Escola Superior de Biotecnologia da Universidade Catolica Portuguesa, Portugal; Faculty of Medical Sciences UNL, Portugal; Universidade Catolica Portuguesa, Portugal.)
- 128. Covino SJ. 1979. *Evaluation and control of formaldehyde exposure in a hospital autopsy room*. Baltimore, MD: Johns Hopkins University (thesis) (as cited in WHO 1989).
- 129. Craft TR, Bermudez E, Skopek TR. 1987. Formaldehyde mutagenesis and formation of DNA-protein crosslinks in human lymphoblasts in vitro. *Mutat Res* 176(1): 147-55. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)
- 130. Crosby RM, Richardson KK, Craft TR, Benforado KB, Liber HL, Skopek TR. 1988. Molecular analysis of formaldehyde-induced mutations in human lymphoblasts and *E. coli. Environ Mol Mutagen* 12(2): 155-166. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC; Harvard School of Public Health, MA; Eli Lily and Company, IL.)
- 131. Crump KS, Chen C, Fox JF, Van Landingham C, Subramaniam R. 2008. Sensitivity analysis of biologically motivated model for formaldehyde-induced respiratory cancer in humans. *Ann Occup Hyg* 52(6): 481-95. (Supported by the U.S. EPA. Authors affiliated with ENVIRON International Corporation, LA; U.S. EPA.)
- 132. Dai D, Bao Z. 1999. Investigative report on formaldehyde occupational workers. *Ind Health Occup Dis*(1): 43 (as cited in Tang *et al.* 2009).
- 133. Dalbey WE. 1982. Formaldehyde and tumors in hamster respiratory tract. *Toxicology* 24(1): 9-14. (Supported by NCI, U.S. EPA, U.S. Department of Energy, and the Union Carbide Corporation. Authors affiliated with Oak Ridge National Laboratory, TN. )
- 134. Dallas CE, Scott MJ, Ward JB, Jr., Theiss JC. 1992. Cytogenetic analysis of pulmonary lavage and bone marrow cells of rats after repeated formaldehyde

inhalation. *J Appl Toxicol* 12(3): 199-203. (Support not reported. Authors affiliated with University of Georgia College of Pharmacy, GA; University of Texas Medical Branch, TX; Warner-Lambert, MI.)

- 135. Dally KA, Hanrahan LP, Woodbury MA, Kanarek MS. 1981. Formaldehyde exposure in nonoccupational environments. *Arch Environ Health* 36(6): 277-284. (Supported by the EPA. Authors affiliated with Wisconsin Division of Health, WI; University of Wisconsin, WI.)
- 136. Dascalaki EG, Lagoudi A, Balaras CA, Gaglia AG. 2008. Air quality in hospital operating rooms. *Building And Environment* 43(11): 1945-1952. (Supported by the Hellenic Ministry of Labor and Social Affairs, Centre for Occupational Health and Safety in the framework of the European Project "survey of occupational problems and risks related to indoor conditions in hospital operating rooms" of the European Commission, Directorate General V, Employment, Industrial Relations, and Social Affairs, Public Health and Safety at Work Directorate V/F. Authors affiliated with National Observatory of Athens, Greece; Ioannou Metaxa & Vas. Pavlou, Greece; Terra Nova, Ltd., Greece.)
- 137. Davis ME, Blicharz AP, Hart JE, Laden F, Garshick E, Smith TJ. 2007. Occupational exposure to volatile organic compounds and aldehydes in the U.S. trucking industry. *Environ Sci Technol* 41(20): 7152-8. (Supported by NIH, NCI and HEI. Authors affiliated with Harvard University, MA; VA Boston Healthcare System, MA.)
- 138. Day JH, Lees RE, Clark RH, Pattee PL. 1984. Respiratory response to formaldehyde and off-gas of urea formaldehyde foam insulation. *Can Med Assoc J* 131(9): 1061-5 (as cited in IARC 2006).
- 139. de Groot AC, van Joost T, Bos JD, van der Meeren HL, Weyland JW. 1988. Patch test reactivity to DMDM hydantoin. Relationship to formaldehyde allergy. *Contact Dermatitis* 18(4): 197-201. (Support not reported. Authors affiliated with Carolus & Willem-Alexander Hospital, Netherlands; Academic Hospital Dijkzigt, Netherlands; Academisch Medisch Centrum, Netherlands; Diakonessenhuis, Netherlands; Food Inspection Service, Netherland. )
- 140. de Serres FJ, Brockman HE. 1999. Comparison of the spectra of genetic damage in formaldehyde-induced *ad-3* mutations between DNA repair-proficient and deficient heterokaryons of *Neurospora crassa*. *Mutat Res* 437(2): 151-163. (Supported by NIEHS. Authors affiliated with NIEHS; Illinois State University, IL.)
- 141. Delfino RJ, Gong H, Jr., Linn WS, Pellizzari ED, Hu Y. 2003. Asthma symptoms in Hispanic children and daily ambient exposures to toxic and criteria air pollutants. *Environ Health Perspect* 111(4): 647-56 (as cited in IARC 2006).

- 142. Dell L, Teta MJ. 1995. Mortality among workers at a plastics manufacturing and research and development facility: 1946-1988. *Am J Ind Med* 28(3): 373-384. (Support not reported. Authors affiliated with University of Massachusetts, MA; Union Carbide Corporation, CT.)
- 143. Demkowicz-Dobrzanski K, Castonguay A. 1992. Modulation by glutathione of DNA strand breaks induced by 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone and its aldehyde metabolites in rat hepatocytes. *Carcinogenesis* 13(8): 1447-1454. (Support not reported. Authors affiliated with Laval University, Canada; Medical Academy, Poland.)
- 144. Dingle P, Tapsell P, Hu S. 2000. Reducing formaldehyde exposure in office environments using plants. *Bull Environ Contam Toxicol* 64(2): 302-8 (as cited in IARC 2006).
- 145. Dodson RE, Houseman EA, Levy JI, Spengler JD, Shine JP, Bennett DH. 2007. Measured and modeled personal exposures to and risks from volatile organic compounds. *Environ Sci Technol* 41(24): 8498-505. (Supported by the American Chemistry Council and the Harvard NIEHS Center for Environmental Health. Authors affiliated with Harvard School of Public Health, MA; University of Massachusetts, MA; University of California, CA.)
- 146. Donovan J, Skotnicki-Grant S. 2007. Allergic contact dermatitis from formaldehyde textile resins in surgical uniforms and nonwoven textile masks. *Dermatitis* 18(1): 40-4. (Supported by the Workplace Safety and Insurance Board. Authors affiliated with St. Michael's Hospital, Canada; Bay Dernatology Centre Toronto, Canada.)
- 147. Doolittle DJ, Furlong JW, Butterworth BE. 1985. Assessment of chemically induced DNA repair in primary cultures of human bronchial epithelial cells. *Toxicol Appl Pharmacol* 79(1): 28-38. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC; Duke University Medical Center, NC; Rohm and Haas Company, PA.)
- 148. Dresp J, Bauchinger M. 1988. Direct analysis of the clastogenic effect of formaldehyde in unstimulated human lymphocytes by means of the premature chromosome condensation technique. *Mutat Res* 204(2): 349-352. (Support not reported. Authors affiliated with Institut fur Strahlenbiologie, Germany.)
- 149. Dufresne A, Infante-Rivard C, Malo JL, Gautrin D. 2002. Exposure to formaldehyde among animal health students. *AIHA J (Fairfax, Va)* 63(5): 647-650. (Support not reported. Authors affiliated with McGill University, Canada; Hopital du Sacre-Coeur, Canada.)
- 150. Duhayon S, Hoet P, Van Maele-Fabry G, Lison D. 2008. Carcinogenic potential of formaldehyde in occupational settings: a critical assessment and possible impact on occupational exposure levels. *Int Arch Occup Environ Health* 81(6):

695-710. (Support not reported. Authors affiliated with Universite catholique de Louvain, Belgium.)

- 151. Dumas S, Parent ME, Siemiatycki J, Brisson J. 2000. Rectal cancer and occupational risk factors: a hypothesis-generating, exposure-based case-control study. *Int J Cancer* 87(6): 874-879. (Supported by Health Canada, National Cancer Institute of Canada, Institut de recherche en sante et securite au travail du Quebec, Fonds de la recherche en sante du Quebec and the Medical Research Council of Canada. Authors affiliated with INTR-Institut Armand-Frappier, Canada; Universite Laval, Canada; McGill University, Canada.)
- 152. DuPont. 2009. "Delerin" II Acetal Resin All In Synonym List. DuPont Engineering Polymers. <u>http://eiecomprod.shopforplastics.com/Ensinger/msds/delrin.html</u>. Accessed on 5/20/09.
- 153. Echt A, Burr GA. 1997. Exposure to formaldehyde during garment manufacturing *Appl Occup Environ Hyg* 12: 451-455 (as cited in IARC 2006).
- 154. Edling C, Hellquist H, Ödkvist L. 1987a. Occupational formaldehyde exposure and the nasal mucosa. *Rhinology* 25(3): 181-187. (Supported by the Swedish Work Environment Fund. Authors affiliated with University Hospital, Sweden.)
- 155. Edling C, Jarvholm B, Andersson L, Axelson O. 1987b. Mortality and cancer incidence among workers in an abrasive manufacturing industry. *Br J Ind Med* 44(1): 57-59. (Support not reported. Authors affiliated with University Hospital, Sweden; Sahlgrenska sjukhuset, Sweden.)
- 156. Edling C, Hellquist H, Ödkvist L. 1988. Occupational exposure to formaldehyde and histopathological changes in the nasal mucosa. *Br J Ind Med* 45(11): 761-765. (Supported by the Swedish Work Environment Fund. Authors affiliated with University Hospital, Sweden; College of Medicine, Sweden; King Saud University, Saudi Arabia.)
- 157. Eells JT, McMartin KE, Black K, Virayotha V, Tisdell RH, Tephly TR. 1981.
  Formaldehyde poisoning. Rapid metabolism to formic acid. *Jama* 246(11): 1237-8. (Support not reported. Authors affiliated with University of Iowa, IA; University of Texas Medical Branch, TX.)
- 158. Elci OC, Akpinar-Elci M, Blair A, Dosemeci M. 2003. Risk of laryngeal cancer by occupational chemical exposure in Turkey. *J Occup Environ Med* 45(10): 1100-1106. (Support not reported. Authors affiliated with NCI; Izmir Chest Diseases and Surgery Training Hospital, Turkey; NIOSH.)
- 159. Elliott LJ, Stayner LT, Blade LM, Halperin W, Keenlyside R. 1987. Formaldehyde Exposure Characterization in Garment Manufacturing Plants: A Composite Summary of Three In-Depth Industrial Hygiene Surveys. Cincinnati, OH: U.S. Department of Health and Human Services (as cited in IARC 2006).

- 160. Emri G, Schaefer D, Held B, Herbst C, Zieger W, Horkay I, Bayerl C. 2004. Low concentrations of formaldehyde induce DNA damage and delay DNA repair after UV irradiation in human skin cells. *Exp Dermatol* 13(5): 305-315. (Supported by DAAD. Authors affiliated with University of Debrecen, Hungary; University Medical Center, Manheim, Germany.)
- 161. Enterline PE, Marsh GM, Esmen NA. 1983. Respiratory disease among workers exposed to man-made mineral fibers. *Am Rev Respir Dis* 128: 1-7. (Supported by the Thermal Insulation Manufacturer's Association. Authors affiliated with University of Pittsburgh, PA. )
- 162. Enterline PE, Marsh GM. 1984. The health of workers in the MMMF industry. In *The Biological Effects of Man-Made Mineral Fibers*, vol. 1. Copenhagen: WHO Regional Office for Europe. p. 311 - 339. (Supported by the Thermal Insulation Manufacturer's Association. Author affiliations not reported.)
- 163. Enterline PE, Marsh GM, Henderson V, Callahan C. 1987. Mortality update of a cohort of U.S. man-made mineral fibre workers. *Ann Occup Hyg* 31(4B): 625-656. (Supported by the U.S. Thermal Insulation Manufacturers' Association. Authors affiliated with University of Pittsburgh, PA.)
- 164. EPA. 1981. *Technical document. Formaldehyde*. Washington, D.C.: U.S. Environmental Protection Agency (as cited in WHO 1989).
- 165. EPA. 2007. Anthrax spore decontamination using paraformaldehyde. U.S. Environmental Protection Agency. <u>http://www.epa.gov/pesticides/factsheets/chemicals/paraformaldehyde\_factsheet\_htm</u>.
- 166. Epstein SS, Shafner H. 1968. Chemical mutagens in the human environment. *Nature* 219(5152): 385-387. (Supported by the U.S. Public Health Service and the Department of Health, Education and Welfare, Division of Air Pollution Control. Authors affiliated with Harvard Medical School, MA.)
- 167. Epstein SS, Arnold E, Andrea J, Bass W, Bishop Y. 1972. Detection of chemical mutagens by the dominant lethal assay in the mouse. *Toxicol Appl Pharmacol* 23(2): 288-325. (Supported by NIH and an Environmental Protection Administrator contract. Authors affiliated with Case Western Reserve University Medical School, OH; Children's Cacer Research Foundation, Inc., MA; Harvard School of Public Health, MA.)
- 168. Erdei E, Bobvos J, Brozik M, Paldy A, Farkas I, Vaskovi E, Rudnai P. 2003. Indoor air pollutants and immune biomarkers among Hungarian asthmatic children. Arch Environ Health 58(6): 337-47. (Support not reported. Authors affiliated with National Institute of Environmental Health, Hungary; Metropolitan Institute of State Public Health and Health Officer Service, Hungary; Central Immunology Laboratorium, Hungary.)

- 169. Fan W, Zhou Y, Wang F, Wang X, Gui L, Jin X. 2004. Olfactory function in subjects exposed to formaldehyde. *J Environ Occup Med* 21(3): 202-204 (as cited in Tang *et al.* 2009).
- 170. Fan W, Zhou Y, Jin F, Du L, Jin X. 2006. The health effects of pathologists exposed to formaldehyde. *J Environ Occup Med* 23(6): 466-468 (as cited in Tang *et al.* 2009).
- 171. Fayerweather WE, Pell S, Bender JR. 1983. Chapter 3. Case-Control Study of Cancer Deaths in Du Pont Workers with Potential Exposure to Formaldehyde. In *Formaldehyde: Toxicology, Epidemiology, and Mechanisms*. Clary JC, Gibson JE, Waritz RS, eds. New York, NY: Marcel Dekker. Inc. p. 47-121. (Support not reported. Authors affiliated with I.E. du Pont de Nemours & Company, Inc., DE.)
- 172. FDA. 2006. *FDA Orange Book*. U.S. Food and Drug Administration. http://www.fda.gov/cder/ob/. Accessed on 4/47/09.
- 173. Feick P, Haas SR, Singer MV, Böcker U. 2006. Low-dose exposure of intestinal epithelial cells to formaldehyde results in MAP kinase activation and molecular alteration of the focal adhesion protein paxillin. *Toxicology* 219(1-3): 60-72. (Supported by the Medical Faculty at Mannheim, University Hospital Mannheim, University of Heidelberg, and the Deutsche Forschungsgemeinschaft. Authors affiliated with University of Heidelberg, Germany.)
- 174. Feng Y, Wang W, Jiang Z, Hu G, Zhong H, Zhang H. 1996. Health status of wood workers exposed to formaldehyde. *Anhui J Prev Med* 2(2): 99-100 (as cited in Tang *et al.* 2009).
- 175. Feron VJ, Bruyntjes JP, Woutersen RA, Immel HR, Appelman LM. 1988. Nasal tumours in rats after short-term exposure to a cytotoxic concentration of formaldehyde. *Cancer Lett* 39(1): 101-111. (Supported by the Stichting Koningin Wilhelmina Fonds. Authors affiliated with TNO-CIVO Toxicology and Nutrition Institute, Netherlands; Directorate General of Labor, Netherlands.)
- 176. FIOH. 1994. *Measurements of Formaldehyde, Industrial Hygiene Database*. Helsinki: Finnish Institute of Occupational Health (as cited in IARC 2006).
- 177. Fishbein L. 1992. Exposure from occupational versus other sources. *Scan J Work Environ Health* 18: 5-16. (Support not reported. Author affiliated with International Life Sciences Institute, Washington, D.C.; U.S. FDA.)
- 178. Fleig I, Petri N, Stocker WG, Thiess AM. 1982. Cytogenetic analyses of blood lymphocytes of workers exposed to formaldehyde in formaldehyde manufacturing and processing. *J Occup Med* 24(12): 1009-1012. (Support not reported. Authors affiliated with BASF Aktiengesellschaft, Germany.)

- 179. Flyvholm MA, Hall BM, Agner T, Tiedemann E, Greenhill P, Vanderveken W, Freeberg FE, Menne T. 1997. Threshold for occluded formaldehyde patch test in formaldehyde-sensitive patients. Relationship to repeated open application test with a product containing formaldehyde releaser. *Contact Dermatitis* 36(1): 26-33. (Support not reported. Authors affiliated with National Institute of Occupational Health, Denmark; Proctor & Gamble Company, UK; Proctor & Gamble Company, OH; Gentofte Hospital, Denmark; Proctor & Gamble Company, Belgium.)
- 180. Fondelli MC, Costantini AS, Ercolanelli M, Pizzo AM, Maltoni SA, Quinn MM. 2007. Exposure to carcinogens and mortality in a cohort of restoration workers of water-damaged library materials following the River Arno flooding in Florence, 4 November 1966. *Med Lav* 98(5): 422-431. (Support not reported. Authors affiliated with Scientific Institute of Tuscany Region, Italy; University of Massachusetts Lowell, MA.)
- 181. Fontignie-Houbrechts N. 1981. Genetic effects of formaldehyde in the mouse. *Mutat Res* 88(1): 109-14. (Supported by the Ministere de la Sante Publique de Belgique. Authors affiliated with Universite de Liege, Belgium.)
- 182. Fornace AJ, Jr., Lechner JF, Grafstrom RC, Harris CC. 1982. DNA repair in human bronchial epithelial cells. *Carcinogenesis* 3(12): 1373-1377. (Support not reported. Authors affiliated with National Cancer Institute, MD.)
- 183. Fowler JF. 2003. Formaldehyde as a textile allergen. *Curr Probl Dermatol* 31: 156-65. (Support not reported. Authors affiliated with University of Louisville School of Medicine, KY.)
- 184. Fox CH, Johnson FB, Whiting J, Roller PP. 1985. Formaldehyde fixation. J Histochem Cytochem 33(8): 845-853. (Support not reported. Authors affiliated with NCI; Armed Forces Institute of Pathology, Washington, D.C.)
- 185. Franks SJ. 2005. A mathematical model for the absorption and metabolism of formaldehyde vapour by humans. *Toxicol Appl Pharmacol* 206(3): 309-320. (Supported by the Health and Safety Executive. Authors affiliated with Health and Safety Laboratory, UK.)
- 186. Fransman W, McLean D, Douwes J, Demers PA, Leung V, Pearce N. 2003. Respiratory symptoms and occupational exposures in New Zealand plywood mill workers. *Ann Occup Hyg* 47(4): 287-95 (as cited in IARC 2006).
- 187. Fritschi L, Siemiatycki J. 1996. Lymphoma, myeloma and occupation: results of a case-control study. *Int J Cancer* 67(4): 498-503. (Supported by the Institut de recherche en sante et en securite du travail du Quebec, the Fonds de recherche en sante du Quebec, the National Health Research and Development Program and the National Cancer Institute of Canada. Authors affiliated with Institut Armand-Frappier, Canada.)

- 188. Fujimaki H, Kurokawa Y, Kunugita N, Kikuchi M, Sato F, Arashidani K. 2004. Differential immunogenic and neurogenic inflammatory responses in an allergic mouse model exposed to low levels of formaldehyde. *Toxicology* 197(1): 1-13. (Supported by the Ministry of the Environment. Authors affiliated with National Institute for Environmental Studies, Japan; University of Occupational and Environmental Health, Japan.)
- 189. Gail MH, Santner TJ, Brown CC. 1980. An analysis of comparative carcinogenesis experiments based on multiple times to tumor. *Biometrics* 36(2): 255-266. (Support not reported. Authors affiliated with NCI; Cornell University, NY.)
- 190. Galli CL, Ragusa C, Resmini P, Marinovich M. 1983. Toxicological evaluation in rats and mice of the ingestion of a cheese made from milk with added formaldehyde. *Food Chem Toxicol* 21(3): 313-317. (Supported by "Consorzio per la tutela del formaggio Grana Padano." Authors affiliated with University of Milan, Italy.)
- 191. Galloway SM, Bloom AD, Resnick M, Margolin BH, Nakamura F, Archer P, Zeiger E. 1985. Development of a standard protocol for in vitro cytogenetic testing with Chinese hamster ovary cells: comparison of results for 22 compounds in two laboratories. *Environ Mutagen* 7(1): 1-51. (Supported by NIEHS. Authors affiliated with Litton Bionetics, Inc., MD; Columbia University College of Physicians and Surgeons, NY; NIEHS, NC; University of Colorado Medical School, CO.)
- 192. Gao X, Zhang P, Qian R, Yao N. 1988. The relationship between clinical symptoms and formaldehyde exposure concentrations. *J Labour Med*(1): 14-17 (as cited in Tang *et al.* 2009).
- 193. Garcia CL, Mechilli M, De Santis LP, Schinoppi A, Kobos K, Palitti F. 2009. Relationship between DNA lesions, DNA repair and chromosomal damage induced by acetaldehyde. *Mutat Res* 662(1-2): 3-9. (Supported by AIRC and University of Tuscia. Authors affiliated with University of Tuscia, Italy.)
- 194. Gardner MJ, Pannett B, Winter PD, Cruddas AM. 1993. A cohort study of workers exposed to formaldehyde in the British chemical industry: an update. *Br J Ind Med* 50(9): 827-834. (Support not reported. Authors affiliated with University of Southampton, UK.)
- 195. Garrigós MC, Reche F, Jiménez A. 2001. Potentially toxic colorant precursors and preservatives used in finger-paints. *Bull Environ Contam Toxicol* 66(5): 557-562. (Supported by CICYT. Authors affiliated with University of Alicante, Spain.)
- 196. Geier J, Lessmann H, Hellriegel S, Fuchs T. 2008. Positive patch test reactions to formaldehyde releasers indicating contact allergy to formaldehyde. *Contact*

*Dermatitis* 58(3): 175-7. (Support not reported. Authors affiliated with Georg August University, Germany.)

- 197. Geng Y, Meng X, Li X, Lu G. 2004. Occupational damage in densified wood board producing field and its effect on workers' health. *Occup Health* 20(8): 21-22 (as cited in Tang *et al.* 2009).
- 198. Gérin M, Siemiatycki J, Nadon L, Dewar R, Krewski D. 1989. Cancer risks due to occupational exposure to formaldehyde: results of a multi-site case-control study in Montreal. *Int J Cancer* 44(1): 53-58. (Supported by the Institut de recherche en sante et securite du Travail du Quebec, the National Health Research and Development Program and the National Cancer Institute of Canada. Authors affiliated with Universite de Montreal, Canada; Institut Armand-Frappier, Canada; McGill University, Canada; Health and Welfare Canada; Carlton University, Canada.)
- 199. GI. 2006. *Economic Primer on Formaldehyde*. Lexington, MA: Global Insights. 9 pp. (Support and author affiliations not reported.)
- 200. Gocke E, King MT, Eckhardt K, Wild D. 1981. Mutagenicity of cosmetics ingredients licensed by the European Communities. *Mutat Res* 90(2): 91-109. (Support not reported. Authors affiliated with Zentrallabor fur Mutagenitatsprufung der Deutschen Forschungsgemeinschaft, Germany.)
- 201. Godderis L, Deschuyffeleer T, Roelandt H, Veulemans H, Moens G. 2008. Exposure to metalworking fluids and respiratory and dermatological complaints in a secondary aluminium plant. *Int Arch Occup Environ Health* 81(7): 845-53. (Supported by the company where this study took place. [Company not named in article.] Authors affiliated with IDEWE, Belgium; Katholieke Universiteit Leuven, Belgium.)
- 202. Golalipour MJ, Azarhoush R, Ghafari S, Gharravi AM, Fazeli SA, Davarian A. 2007. Formaldehyde exposure induces histopathological and morphometric changes in the rat testis. *Folia Morphol (Warsz)* 66(3): 167-71. (Supported by Gorgan University. Authors affiliated with Gorgan University, Iran.)
- 203. Golalipour MJ, Kord H, Ghafari S, Gharravi AM, Davarian A, Fazeli SA, Azarhoush R. 2008. Morphometric alterations of the rat spleen following formaldehyde exposure. *Folia Morphol (Warsz)* 67(1): 19-23. (Supported by Gorgan University. Authors affiliated with Gorgan University of Medical Sciences, Iran.)
- 204. Gold KW, Naugle DF, Berry MA. 1993. Indoor Concentrations of Environmental Carcinogens. In *Environmental Carcinogens Methods of Analysis and Exposure Measurement*, vol. 12, IARC Scientific Publications No. 109. Seifert B, van de Wiel HJ, Dodet B, O'Neill IK, eds. Lyon, France: International Agency for Research on Cancer. p. 41-71. (Supported by the U.S. EPA. Authors affiliated with Research Triangle Institute, NC; U.S. EPA.)

- 205. Golden R, Pyatt D, Shields PG. 2006. Formaldehyde as a potential human leukemogen: an assessment of biological plausibility. *Crit Rev Toxicol* 36(2): 135-153. (Supported by the Formaldehyde Council, Inc. Authors affiliated with ToxLogic, MD; Summit Toxicology, CO; University of Colorado, CO; Georgetown University Medical Center, Washington, D.C.)
- 206. Goldmacher VS, Thilly WG. 1983. Formaldehyde is mutagenic for cultured human cells. *Mutat Res* 116(3-4): 417-422. (Supported by the U.S. Department of Energy. Authors affiliated with Massachusetts Institute of Technology, MA.)
- 207. Goldoft M, Weiss N, Vaughan T, Lee J. 1993. Nasal melanoma. *Br J Ind Med* 50(8): 767-768. (Support not reported. Authors affiliated with University of Washington, WA.)
- 208. Goldstein BD. 2009. Clinical and hematotoxicological evaluation of current evidence does not support formaldehyde as a cause of human leukemia. *Hum Exp Toxicol*(Pre-publication). (No grant support was received. Author affiliated with University of Pittsburgh Graduate School of Public Health, PA.)
- 209. Grafström RC, Fornace AJ, Jr., Autrup H, Lechner JF, Harris CC. 1983. Formaldehyde damage to DNA and inhibition of DNA repair in human bronchial cells. *Science* 220(4593): 216-218. (Support not reported. Authors affiliated with NCI, MD.)
- 210. Grafström RC, Fornace A, Jr., Harris CC. 1984. Repair of DNA damage caused by formaldehyde in human cells. *Cancer Res* 44(10): 4323-4327. (Supported by the Swedish Cancer Society and the National Board for Laboratory Animals. Authors affiliated with Karolinska Institutet, Sweden; NCI, MD.)
- 211. Grafström RC, Curren RD, Yang LL, Harris CC. 1985. Genotoxicity of formaldehyde in cultured human bronchial fibroblasts. *Science* 228(4695): 89-91. (Supported by the Swedish Cancer Society. Authors affiliated with Karolinska Institutet, Sweden; NCI, MD; Microbiological Associates, MD.)
- 212. Grafström RC, Willey JC, Sundqvist K, Harris CC. 1986. Pathobiological effects of tobacco smoke-related aldehydes in cultured human bronchial epithelial cells. In *Mechanisms in Tobacco Carcinogenesis*, Banbury Report 23. Hoffman D, Harris CC, eds. Cold Spring Harbor, NY: CSH Press. pp. 273-285. (Supported by the Swedish Cancer Society, the Swedish Medical Research Council, the National Board of Laboratory Animals, the Swedish work Environment Health Fund, and the Swedish Tobacco Company. Authors affiliated with Karolinska Institute, Sweden; NCI. )
- 213. Grafström RC. 1990. In vitro studies of aldehyde effects related to human respiratory carcinogenesis. *Mutat Res* 238(3): 175-184. (Supported by the Swedish Work Environment Fund, Swedish National Board of Laboratory Animals, Swedish Cancer Society, Swedish National Science Research Council,

Swedish Tobacco Company, and the Health Effects Institute. Authors affiliated with Karolinska Institutet, Sweden.)

- 214. Grafström RC, Hsu IC, Harris CC. 1993. Mutagenicity of formaldehyde in Chinese hamster lung fibroblasts: synergy with ionizing radiation and *N*-nitroso-*N*-methylurea. *Chem Biol Interact* 86(1): 41-49. (Supported by the Swedish Work Environment Fund, Swedish National Board of Laboratory Animals, Swedish Cancer Society, Swedish National Science Research Council, Swedish Fund for Research Without Animal Experiments, Swedish Tobacco Company and the U.S. EPA. Authors affiliated with Karolinska Institutet, Sweden; University of Maryland, MD; NCI, MD.)
- 215. Grafström RC, Jernelov MI, Dypbukt JM, Sundqvist K, Atzori L, Zheng X. 1996. Aldehyde toxicity and thiol redox state in cells cultures from human aerodigestive tract. In *Correlations Between In Vitro and In Vivo Investigations in Inhalation Toxicology* Mohr U, Adler KB, Dungworth DI, etal., eds. Washington, D.C.: ILSI Press. p. 319 - 336. (Supported by the Transport Research Board, Volvo Research Foundation, the Council for Forestry and Agricultural Research, the National Board of Laboratory Animals, the Cancer Society, the Fund for Research Without Animal Experiments, HEI and the Tobacco Company in Sweden. Authors affiliated with Karolinksa Institute, Sweden.)
- 216. Greaves MF. 2004. Biological models for leukaemia and lymphoma. *IARC Sci Publ*(157): 351-372. (Supported by the Leukaemia Research Fund, the Kay Kendall Leukaemia Fund and the Institute of Cancer Research. Author affiliated with Chester Beatty Laboratories, UK.)
- 217. Green DJ, Sauder LR, Kulle TJ, Bascom R. 1987. Acute response to 3.0 ppm formaldehyde in exercising healthy nonsmokers and asthmatics. *Am Rev Respir Dis* 135(6): 1261-6 (as cited in IARC 2006).
- 218. Grimsrud DT, Lipschute RD, Girman JR. 1983. *Indoor Air Quality in Energy Efficient Residences* LBL-14795. Berkely, CA: Energy and Environment Division, Lawrence Berkely Laboratory, University of California (as cited by WHO 1989).
- 219. Grosjean D. 1982. Formaldehyde and other carbonyls in Los Angeles ambient air. *Environ Sci Technol* 16: 254-262 (as cited in ATSDR 1999).
- 220. Grosjean D, Swanson RD. 1983. Carbonyls in Los Angeles air: contribution of direct emissions and photochemistry. *Sci Total Environ* 29: 65-85 (as cited in WHO 1989).
- 221. Grosjean E, Williams ELI, Grosjean D. 1993. Ambient levels of formaldehyde and acetaldehyde in Atlanta, Georgia. *J Air Waste Manage Assoc* 43: 469-474 (as cited in IARC 2006).

- 222. Grosjean E, Grosjean D, Fraser MP, Cass GR. 1996. Air quality model evaluation data for organics. 2. C<sub>1</sub>-C<sub>14</sub> carbonyls in Los Angeles air. *Environ Sci Technol* 30: 2687-2703. (Supported by the Electric Power Research Institute and DGA, Inc. Authors affiliated with DGA, Inc., CA; California Institute of Technology, CA. )
- 223. Gulec M, Songur A, Sahin S, Ozen OA, Sarsilmaz M, Akyol O. 2006. Antioxidant enzyme activities and lipid peroxidation products in heart tissue of subacute and subchronic formaldehyde-exposed rats: a preliminary study. *Toxicol Ind Health* 22(3): 117-24. (Support not reported. Authors affiliated with Ankara Numune Education and Research Hospital, Turkey; Afyon Kocatepe University Medical School, Turkey; Gaziomanpasa University Medical School, Turkey; Firat University Medical Faculty, Turkey; Hacettepe University Medical School, Turkey.)
- 224. Gurel A, Coskun O, Armutcu F, Kanter M, Ozen OA. 2005. Vitamin E against oxidative damage caused by formaldehyde in frontal cortex and hippocampus: biochemical and histological studies. *J Chem Neuroanat* 29(3): 173-8. (Support not reported. Authors affiliated with Zonguldak Karaelmas University, Turkey.)
- 225. Gustavsson P, Jakobsson R, Johansson H, Lewin F, Norell S, Rutkvist LE. 1998. Occupational exposures and squamous cell carcinoma of the oral cavity, pharynx, larynx, and oesophagus: a case-control study in Sweden. Occup Environ Med 55(6): 393-400. (Supported by the Cancer Society of Stockholm, the Swedish Cancer Fund and the research fund of the Swedish tobacco company. Authors affiliated with Karolinska Hospital, Sweden; Kungsklippen, Sweden.)
- 226. Guyton KZ, Kyle AD, Aubrecht J, Cogliano VJ, Eastmond DA, Jackson M, Keshava N, Sandy MS, Sonawane B, Zhang L, Waters MD, Smith MT. 2009. Improving prediction of chemical carcinogenicity by considering multiple mechanisms and applying toxicogenomic approaches. *Mutat Res* 681(2-3): 230-240. (Supported by the U.S. EPA, Integrated Laboratory Systems, Inc., Pfizer, Inc. and NIEHS. Authors affiliated with U.S. EPA; University of California Berkeley, CA; Pfizer Global Research and Development, CT; IARC; University of California Riverside, CA; Integrated Laboratory Systems, NC; California Environmental Protection Agency.)
- 227. Haas EM, Bailey HR, Farragher I. 2007. Application of 10 percent formalin for the treatment of radiation-induced hemorrhagic proctitis. *Diseases of the Colon and Rectum* 50(2): 213. (Support not reported. Authors affiliated with Methodist Hospital, TX; UT Health Science Center, TX; Western Hospital, Australia.)
- 228. Hagberg M, Kolmodin-Hedman B, Lindahl R, Nilsson CA, Norstrom A. 1985. Irritative complaints, carboxyhemoglobin increase and minor ventilatory function changes due to exposure to chain-saw exhaust. *Eur J Respir Dis* 66(4): 240-7 (as cited in IARC 2006).

- 229. Hagiwara M, Watanabe E, Barrett JC, Tsutsui T. 2006. Assessment of genotoxicity of 14 chemical agents used in dental practice: ability to induce chromosome aberrations in Syrian hamster embryo cells. *Mutat Res* 603(2): 111-120. (Support not reported. Authors affiliated with The Nippon Dental University, Japan; Novartis Institutes for BioMedical Research Inc., MA; NIH, MD.)
- 230. Hall A, Harrington JM, Aw TC. 1991. Mortality study of British pathologists. *Am J Ind Med* 20(1): 83-89. (Support not reported. Authors affiliated with University of Birmingham, UK.)
- 231. Hamaguchi F, Tsutsui T. 2000. Assessment of genotoxicity of dental antiseptics: ability of phenol, guaiacol, p-phenolsulfonic acid, sodium hypochlorite, pchlorophenol, m-cresol or formaldehyde to induce unscheduled DNA synthesis in cultured Syrian hamster embryo cells. *Jpn J Pharmacol* 83(3): 273-6. (Support not reported. Authors affiliated with Nippon Dental University, Japan.)
- 232. Hansen J, Olsen JH. 1995. Formaldehyde and cancer morbidity among male employees in Denmark. *Cancer Causes Control* 6(4): 354-360. (Support not reoprted. Authors affiliated with Danish Cancer Society, Denmark; National Institute of Occupational Health, Denmark.)
- 233. Hansen J, Olsen JH. 1996. [Occupational exposure to formaldehyde and risk of cancer]. *Ugeskr Laeger* 158(29): 4191-4194. (Support and affiliations not identified due to foreign language.)
- 234. Harrington JM, Shannon HS. 1975. Mortality study of pathologists and medical laboratory technicians. *Br Med J* 4(5992): 329-332. (Supported by the Department of Health and Social Security and the Scottish Home and Health Department. Authors affiliated with London School of Hygiene and Tropical Medicine, UK.)
- 235. Harrington JM, Oakes D. 1984. Mortality study of British pathologists 1974-80. Br J Ind Med 41(2): 188-191. (Supported by the DHSS. Authors affiliated with University of Birmingham, UK; London School of Hygiene and Tropical Medicine, UK.)
- 236. Harris JC, Rumack BH, Aldrich FD. 1981. Toxicology of urea formaldehyde and polyurethane foam insulation. *Jama* 245(3): 243-3. (Support not reported. Authors affiliated with University of Colorado, CO; Denver General Hospital, CO; B.F. Stolinsky Laboratories, CO; IBM Corporation, CO.)
- 237. Hauptmann M, Lubin JH, Stewart PA, Hayes RB, Blair A. 2003. Mortality from lymphohematopoietic malignancies among workers in formaldehyde industries. *J Natl Cancer Inst* 95(21): 1615-1623. (Supported by the National Cancer Institute, National Institutes of Health, Department of Health and Human Services. Authors affiliated with NCI.)

- 238. Hauptmann M, Lubin JH, Stewart PA, Hayes RB, Blair A. 2004. Mortality from solid cancers among workers in formaldehyde industries. *Am J Epidemiol* 159(12): 1117-1130. (Support not reported. Authors affiliated with NCI.)
- 239. Hawthorne AR, Gammage RB, Dudney CS, Wormack DR, Morris SA, Wesley RR. 1983. Preliminary results of a forty home indoor air pollutant monitoring study. In *Proceedings of the Conference on Mesaurements and Monitoring of Non-Criteria (Toxic) Contaminants in Air*. Pittsburgh, PA: Air Pollution Control Association. 514 pp (as cited in WHO 1989).
- 240. Hayes RB, Raatgever JW, de Bruyn A, Gerin M. 1986. Cancer of the nasal cavity and paranasal sinuses, and formaldehyde exposure. *Int J Cancer* 37(4): 487-92. (Supported by the Ministry of Health and Environmental Hygiene, Netherlands. Authors affiliated with Erasmus University Rotterdam, Netherlands; University of Montreal, Canada.)
- 241. Hayes RB, Blair A, Stewart PA, Herrick RF, Mahar H. 1990. Mortality of U.S. embalmers and funeral directors. *Am J Ind Med* 18(6): 641-652. (Support not reported. Authors affiliated with NCI; NIOSH; NIH.)
- 242. Hayes RB, Klein S, Suruda A, Schulte P, Boeniger M, Stewart P, Livingston GK, Oesch F. 1997. O<sup>6</sup>-alkylguanine DNA alkyltransferase activity in student embalmers. *Am J Ind Med* 31(3): 361-365. (Supported by the Bundesministerium fur Forschung und Technologie. Authors affiliated with NCI, MD; University of Mainz, Germany; NIOSH, OH; University of Cincinnati School of Medicine, OH.)
- 243. He JL, Jin LF, Jin HY. 1998. Detection of cytogenetic effects in peripheral lymphocytes of students exposed to formaldehyde with cytokinesis-blocked micronucleus assay. *Biomed Environ Sci* 11(1): 87-92. (Support not reoprted. Authors affiliated with Zhejiang Medical University, China.)
- 244. Heck H, Casanova M. 2004. The implausibility of leukemia induction by formaldehyde: a critical review of the biological evidence on distant-site toxicity. *Regul Toxicol Pharmacol* 40(2): 92-106. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)
- 245. Heck HD, White EL, Casanova-Schmitz M. 1982. Determination of formaldehyde in biological tissues by gas chromatography/mass spectrometry. *Biomed Mass Spectrom* 9(8): 347-353. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)
- 246. Heck HD, Casanova-Schmitz M, Dodd PB, Schachter EN, Witek TJ, Tosun T. 1985. Formaldehyde (CH<sub>2</sub>O) concentrations in the blood of humans and Fischer-344 rats exposed to CH<sub>2</sub>O under controlled conditions. *Am Ind Hyg Assoc J* 46(1): 1-3. (Supported by the Formaldehyde Institute. Authors affiliated with Chemical Industry Institute of Toxicology, NC; Yale University School of Medicine, CT.)

- 247. Heck HD, Casanova M, Starr TB. 1990. Formaldehyde toxicity--new understanding. *Crit Rev Toxicol* 20(6): 397-426. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)
- 248. Heck Hd A, Chin TY, Schmitz MC. 1983. Distribution of [<sup>14</sup>C] Formaldehyde in Rats after Inhalation Exposure. In *Formaldehyde Toxicity*. Gibson JE, ed. Washington, D.C.: Hemisphere Publishing Corporation. pp. 26-36. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)
- 249. Heck HdA, Casanova M, C-W. L, Swenberg JA. 1986. The formation of DNAprotein crosslinks by aldehydes present in tobacco smoke. In *Mechanisms in Tobacco Carcinogenesis*, Banbury Report 23. Hoffman D, Harris CC, eds. Cold Spring Harbor, NY: CSH Press. pp. 215-230. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)
- 250. Heck HdA, Casanova M, Steinhagen WH, Everitt JI, Morgan KT, Popp JA. 1989. Formaldehyde toxicity: DNA-protein crosslinking studies in rats and nonhuman primates. In *Nasal Carcinogenesis in Rodents: Relevance to Human Risk*. Feron VJ, Bosland MC, eds. Wageningen: Pudoc. pp. 159-164. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)
- 251. Heikkila P, Priha E, Savela A. 1991. *Formaldehyde: Exposure and Work #14*. Helsinki: Finnish Institute of Occupational Health and the Finnish Work Environment Fund (as cited in IARC 2006).
- 252. Heineman EF, Olsen JH, Pottern LM, Gomez M, Raffn E, Blair A. 1992. Occupational risk factors for multiple myeloma among Danish men. *Cancer Causes Control* 3(6): 555-68. (Support not reported. Authors affiliated with NCI, MD; Danish Cancer Registry, Denmark; Arbedjdsmedicinsk Klinsk, Denmark.)
- 253. Hemminki K, Mutanen P, Saloniemi I, Niemi ML, Vainio H. 1982. Spontaneous abortions in hospital staff engaged in sterilising instruments with chemical agents. *Br Med J (Clin Res Ed)* 285(6353): 1461-3. (Support not reported. Authors affiliated with Institute of Occupational Health, Finland.)
- 254. Hemminki K, Kyyronen P, Lindbohm ML. 1985. Spontaneous abortions and malformations in the offspring of nurses exposed to anaesthetic gases, cytostatic drugs, and other potential hazards in hospitals, based on registered information of outcome. *J Epidemiol Community Health* 39(2): 141-7. (Supported by the Medical Research Council, Finland. Authors affiliated with Institute of Occupational Health, Finland.)
- 255. Hendrick DJ, Lane DJ. 1975. Formalin asthma in hospital staff. *Br Med J* 1(5958): 607-8 (as cited in IARC 2006).

- 256. Hendrick DJ, Lane DJ. 1977. Occupational formalin asthma. *Br J Ind Med* 34(1): 11-8 (as cited in IARC 2006).
- 257. Hendrick DJ, Rando RJ, Lane DJ, Morris MJ. 1982. Formaldehyde asthma: challenge exposure levels and fate after five years. *J Occup Med* 24(11): 893-7 (as cited in IARC 2006).
- 258. Herbert FA, Hessel PA, Melenka LS, Yoshida K, Nakaza M. 1995. Pulmonary effects of simultaneous exposures to MDI formaldehyde and wood dust on workers in an oriented strand board plant. *J Occup Environ Med* 37(4): 461-5 (as cited in IARC 2006).
- 259. Hernberg S, Westerholm P, Schultz-Larsen K, Degerth R, Kuosma E, Englund A, Engzell U, Hansen HS, Mutanen P. 1983a. Nasal and sinonasal cancer. Connection with occupational exposures in Denmark, Finland and Sweden. *Scand J Work Environ Health* 9(4): 315-326. (Supported by the Nordic Council of Ministers and the Swedish Work Environment Fund. Authors affiliated with Institute of Occupational Health, Finland; Swedish Confederation of Trade Unions, Sweden; Copenhagen County Hospital, Denmark; Foundation for Industrial Safety and Health in the Construction Industry, Sweden; Huddinge Hospital, Sweden; Finsen Institute, Denmark.)
- 260. Hernberg S, Collan Y, Degerth R, Englund A, Engzell U, Kuosma E, Mutanen P, Nordlinder H, Hansen HS, Schultz-Larsen K, Søgaard H, Westerholm P. 1983b. Nasal cancer and occupational exposures. Preliminary report of a joint Nordic case-referent study. *Scand J Work Environ Health* 9(2): 208-213. (Supported by the Nordc Council of Ministers and the Swedish Work Environment Fund. Authors affiliated with Institute of Occupational Health, Finland; University of Kuopio, Finland; Foundation for Industrial Safety and Health in the Construction Industry, Sweden; Huddinge Hospital, Sweden; Uppsala University, Sweden; Finsen Institute, Denmark; City Hospital, Denmark; Swedish Workers Confederation, Sweden.)
- 261. Herrick HF, Alacrese AA, Reisdorf RP, Rumsey DL. 1983. Industrial Hygiene Characterization of Urea-Formaldehyde and Polyurethane Foam Insulation. Technical Report no. 83-108. Washington, D.C.: National Institute of Occupational Safety and Health (as cited in WHO 1989).
- 262. Hester SD, Benavides GB, Yoon L, Morgan KT, Zou F, Barry W, Wolf DC. 2003. Formaldehyde-induced gene expression in F344 rat nasal respiratory epithelium. *Toxicology* 187(1): 13-24. (Support not reported. Authors affiliated with U.S. EPA; GlaxoSmithKline Inc., NC; University of North Carolina, NC.)
- 263. Hester SD, Barry WT, Zou F, Wolf DC. 2005. Transcriptomic analysis of F344 rat nasal epithelium suggests that the lack of carcinogenic response to glutaraldehyde is due to its greater toxicity compared to formaldehyde. *Toxicol*

*Pathol* 33(4): 415-424. (Support not reported. Authors affiliated with University of North Carolina, NC; U.S. EPA.)

- 264. Hikiba H, Watanabe E, Barrett JC, Tsutsui T. 2005. Ability of fourteen chemical agents used in dental practice to induce chromosome aberrations in Syrian hamster embryo cells. *J Pharmacol Sci* 97(1): 146-52. (Support not reported. Authors affiliated with Nippon Dental University, Japan; NIH, MD.)
- 265. Hilbert G, Gruson D, Bedry R, Cardinaud JP. 1997. Circulatory shock in the course of fatal poisoning by ingestion of formalin. *Intensive Care Med* 23(6): 708. (Support not reported. Authors affiliated with Hopital Pellegrin, France.)
- 266. Hildesheim A, Dosemeci M, Chan CC, Chen CJ, Cheng YJ, Hsu MM, Chen IH, Mittl BF, Sun B, Levine PH, Chen JY, Brinton LA, Yang CS. 2001. Occupational exposure to wood, formaldehyde, and solvents and risk of nasopharyngeal carcinoma. *Cancer Epidemiol Biomarkers Prev* 10(11): 1145-1153. (Support not reported. Authors affiliated with NCI; Chang-Gung Memorial Hospital, Taiwan; George Washington University, Washington, D.C; National Health Research Institute, Taiwan.)
- 267. Hilton J, Dearman RJ, Basketter DA, Scholes EW, Kimber I. 1996. Experimental assessment of the sensitizing properties of formaldehyde. *Food Chem Toxicol* 34(6): 571-8. (Support not reported. Authors affiliated with Zeneca Central Toxicology Laboratory, UK; Unilever Environmental Safety Laboratory, UK.)
- 268. Hodgson AT, Rudd AF, Beal D, Chandra S. 2000. Volatile organic compound concentrations and emission rates in new manufactured and site-built houses. *Indoor Air* 10(3): 178-92 (as cited in IARC 2006).
- 269. Hodgson AT, Beal D, McIlvaine JE. 2002. Sources of formaldehyde, other aldehydes and terpenes in a new manufactured house. *Indoor Air* 12(4): 235-42 (as cited in IARC 2006).
- 270. Holly EA, Aston DA, Ahn DK, Smith AH. 1996. Intraocular melanoma linked to occupations and chemical exposures. *Epidemiology* 7(1): 55-61. (Supported by NCI. Authors affiliated with University of California San Francisco, CA; Stanford University, CA; University of California Berkeley, CA.)
- 271. Holmström M, Wilhelmsson B, Hellquist H. 1989a. Histological changes in the nasal mucosa in rats after long-term exposure to formaldehyde and wood dust. *Acta Otolaryngol* 108(3-4): 274-283. (Support not reported. Authors affiliated with Karolinska Institutet, Sweden; King Saud University, Saudi Arabia.)
- 272. Holmström M, Wilhelmsson B, Hellquist H, Rosén G. 1989b. Histological changes in the nasal mucosa in persons occupationally exposed to formaldehyde alone and in combination with wood dust. *Acta Otolaryngol* 107(1-2): 120-129. (Supported by the Swedish Work Environment Fund. Authors affiliated with

Karolinska Institutet, Sweden; King Saud University, Saudia Arabia; National Board of Occupational Safety and Health, Sweden.)

- 273. Holness DL, Nethercott JR. 1989. Health status of funeral service workers exposed to formaldehyde. *Arch Environ Health* 44: 222-228. (Supported by the Ontario Ministry of Labour. Authors affiliated with University of Toronto, Canada.)
- 274. Homma Y, Nowels K, Oyasu R. 1986. Effects of formalin-induced injuries on urinary bladder carcinogenesis. *Cancer Lett* 32(2): 117-123. (Supported by NCI. Authors affiliated with Northwestern University Medical School, IL.)
- 275. Horton AW, Tye R, Stemmer KL. 1963. Experimental carcinogenesis of the lung. Inhalation of gaseous formaldehyde or an aerosol of coal tar by C3H mice. *J Natl Cancer Inst* 30: 31-43. (Support not reported. Authors affiliated with University of Cincinnati, OH.)
- 276. Horvath EP, Jr., Anderson H, Jr., Pierce WE, Hanrahan L, Wendlick JD. 1988. Effects of formaldehyde on the mucous membranes and lungs. A study of an industrial population. *Jama* 259(5): 701-7 (as cited in ATSDR 1999).
- 277. Howard PH. 1989. *Handbook of Environmental Fate and Exposure Data for Organic Chemicals* Chelsea, MI: Lewis Publishers, Inc. p. 342-350.
- 278. HPD. 2009. *Household Products Database: Formaldehyde*. U.S. Department of Health and Human Services. Updated on 9/26/08. <u>http://householdproducts.nlm.nih.gov/</u>. Accessed on 5/15/09.
- 279. HSDB. 2007. *Hazardous Substances Data Bank*. National Library of Medicine. <u>http://toxnet.nlm.nih.gov/cgi-bin/sis/htmlgen?HSDB</u> and search " formaldehyde". Accessed on 6/28/07.
- 280. HSDB. 2009a. *Hazardous Substances Data Bank. Formaldehyde*. National Library of Medicine. <u>http://toxnet.nlm.nih.gov/cgi-bin/sis/htmlgen?HSDB</u> and search " formaldehyde". Accessed on 5/18/09.
- 281. HSDB. 2009b. *Hazardous Substances Data Bank. Paraformaldehyde*. National Library of Medicine. <u>http://toxnet.nlm.nih.gov/cgi-bin/sis/htmlgen?HSDB</u> and search " paraformaldehyde". Accessed on 5/18/09.
- 282. HSDB. 2009c. *Hazardous Substances Data Bank. 1,3,5-Trioxane*. National Library of Medicine. <u>http://toxnet.nlm.nih.gov/cgi-bin/sis/htmlgen?HSDB</u> and search " 1,3,5-Trioxane". Accessed on 5/18/09.
- 283. Huan P, Fan W, Jin F. 2001. The investigation of combined effect of formaldehyde and noise on blood pressure. *Occup Health and Emerg Rescue* 19(1): 6-7 (as cited in Tang *et al.* 2009).

- 284. Humphries P, Pretorius E, Naude H. 2008. Direct and indirect cellular effects of aspartame on the brain. *Eur J Clin Nutr* 62(4): 451-62. (Support not reported. Authors affiliated with University of Pretoria, South Africa; University of Limpopo, South Africa.)
- 285. IARC. 1982. *IARC Monographs on the Evaluation of Carcinogenic Risks to Humans. Some Industrial Chemicals and Dyestuffs*, vol. 29, Lyon, France: International Agency for Research on Cancer. 416 pp.
- 286. IARC. 1995. *IARC Monographs on the Evaluation of Carcinogenic Risks to Humans. Wood Dust and Formaldehyde*, vol. 62, Lyon, France: International Agency for Research on Cancer. p. 217-362.
- IARC. 2006. IARC Monographs on the Evaluation of Carcinogenic Risks to Humans. Formaldehyde, 2-Butoxyethanol and 1-tert-Butoxypropan-2-ol, vol. 88, Lyon, France: International Agenc for Research on Cancer. p. 39-325.
- 288. Iarmarcovai G, Bonassi S, Sari-Minodier I, Baciuchka-Palmaro M, Botta A, Orsiere T. 2007. Exposure to genotoxic agents, host factors, and lifestyle influence the number of centromeric signals in micronuclei: a pooled reanalysis. *Mutat Res* 615(1-2): 18-27. (Supported by Direction Regionale du Travail, de l'Emploi et de la Formation Professionnelle of the Provence-Alpes-Cote d'Azur region, the Fondation Philippe Daher, the Associazione Italiana per la Ricerca sul Cancro and the Agenzia Spaziale Italiana. Authors affiliated with Universite de la Mediterranee, France; National Cancer Research Institute, Italy.)
- 289. ICIS. 2007. Chemical Profile: Formaldehyde. *ICIS Chemical Business Americas* April 30 May 6: 34. (Support and author affiliations not reported.)
- 290. Im H, Oh E, Mun J, Khim JY, Lee E, Kang HS, Kim E, Kim H, Won NH, Kim YH, Jung WW, Sul D. 2006. Evaluation of toxicological monitoring markers using proteomic analysis in rats exposed to formaldehyde. *J Proteome Res* 5(6): 1354-66. (Supported by the Medical Research Center for Environmental Toxico-Genomics and Proteomics of Korea University and by the Ministry of Environment as "The Eco-Technopia 21 project." Authors affiliated with Korea University, Korea; Chonnam University, Korea; MyGene Bioscience Institute, Korea.)
- 291. Inventro. 2009. *Nylon 6*. Inventro Polymers. <u>http://www.inventro.net/NYLON.HTM</u>. Accessed on 5/28/09.
- 292. IRSST. 2006. *Prevention Guide: Formaldehye in the Workplace*. Montreal, Canada: Institut de recherche Robert-Sauve en sante et en securite du travail 55 pp. (Supported by the Commission de la sante et de la securite du travail. Authors affiliated with IRSST; University of Montreal, Canada.)

- 293. Ishidate Jr M, Sofuni T, Yoshikawa K. 1981. Chromosomal aberration tests *in vitro* as a primary screening tool for environmental mutagens and/or carcinogens. *GANN Monogr Cancer Res* 27: 95-108. (Supported by the Ministry of Education, Science and Culture and the Ministry of Health and Welfare of Japan. Authors affiliated with National Institute of Hygienic Sciences, Japan.)
- 294. Iversen OH. 1986. Formaldehyde and skin carcinogenesis. *Environ Int* 12: 541-544. (Support not reported. Authors affiliated with University of Oslo, Norway; National Hospital, Norway.)
- 295. Jankovic J, Jones W, Burkhart J, Noonan G. 1991. Environmental study of firefighters. *Ann Occup Hyg* 35: 581-602. (Supported by the Federal Emergency Management Agency, U.S. Fire Administration. Authors affiliated with NIOSH.)
- 296. Jeffcoat AR, Chasalow F, Feldman DB, Marr H. 1983. Disposition of [<sup>14</sup>C] Formaldehyde after Topical Exposure to Rats, Guinea Pigs and Monkeys. In *Formaldehyde Toxicity*. Gibson JE, ed. Washington, D.C.: Hemisphere Publishing Corporation. pp. 38-50. (Supported by the American Textile Manufacturer's Institute and the Formaldehyde Institute. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)
- 297. Jensen OM, Andersen SK. 1982. Lung cancer risk from formaldehyde. *Lancet* 1(8277): 913. (Support not reported. Authors affiliated with Danish Cancer Registry, Denmark.)
- 298. Jiang SF, Yu LQ, Leng SG, Zhang YS, Cheng J, Dai YF, Niu Y, He FS, Zheng YX. 2006. [Association between XRCC1 gene polymorphisms and DNA damage of workers exposed to formaldehyde]. *Wei Sheng Yan Jiu* 35(6): 675-677 (as cited in Tang *et al.* 2009).
- 299. Jin F, Zhu R. 1992. Cytogenetic effects on peripheral blood lymphocytes of workers occupationally exposed to formaldehyde. *Chin J Ind Hyg Occup Dis* 10(5): 277-278, 281 (as cited in Tang *et al.* 2009).
- 300. Kagi N, Fujii S, Tamura H, Namiki N. 2009. Secondary VOC emissions from flooring material surfaces exposed to ozone or UV irradiation. *Building And Environment* 44(6): 1199-1205. (Support not reported. Authors affiliated with National Institute of Public Health, Japan; Tokyo Institute of Technology, Japan; Techno Ryowa Ltd., Japan; Kogakuin University, Japan.)
- 301. Kalinic N, Sega K, SiSovic A. 1985. Formaldehyde Levels in Selected Indoor Environments. In *Proceedings of the Third International Conference on Indoor Air Quality and Climate, Stockholm, 20-24 August*, vol. 3. Berglund B, Lindvall T, Sundell J, eds. Stockholm: Swedish Council for Building Research. p. 145-148 (as cited in WHO 1989).

- 302. Kamata E, Nakadate M, Uchida O, Ogawa Y, Suzuki S, Kaneko T, Saito M, Kurokawa Y. 1997. Results of a 28-month chronic inhalation toxicity study of formaldehyde in male Fisher-344 rats. *J Toxicol Sci* 22(3): 239-54. (Support not reported. Authors affiliated with National Institute of Health Sciences, Japan.)
- 303. Katakura Y, Kishi R, Okui T, Ikeda T, Miyake H. 1993. Distribution of radioactivity from <sup>14</sup>C-formaldehyde in pregnant mice and their fetuses. *British J Ind Med* 50: 176-182. (Support not reported. Authors affiliated with Hokkaido Institute of Public Health, Japan.)
- 304. Katakura Y, Kishi R, Okui T, Ikeda T, Miyake H. 1993. Distribution of radioactivity from 14C-formaldehyde in pregnant mice and their fetuses. *Br J Ind Med* 50(2): 176-82. (Support not reported. Authors affiliated with Hokkaido Institute of Public Health, Japan.)
- 305. Kauppinen T. 1986. Occupational exposure to chemical agents in the plywood industry. *Ann Occup Hyg* 30(1): 19-29 (as cited in IARC 2006).
- 306. Kauppinen T, Partanen T. 1988. Use of plant- and period-specific job-exposure matrices in studies on occupational cancer. *Scand J Work Environ Health* 14(3): 161-167. (Support not reported. Authors affiliated with Institute of Occupational Health, Finland.)
- 307. Kauppinen TP, Niemela RI. 1985. Occupational exposure to chemical agents in the particleboard industry. *Scand J Work Environ Health* 11(5): 357-63 (as cited in IARC 2006).
- 308. Keefer LK, Streeter AJ, Leung LY, Perry WC, Hu HS, Baillie TA. 1987. Pharmacokinetic and deuterium isotope effect studies on the metabolism of formaldehyde and formate to carbon dioxide in rats in vivo. *Drug Metab Dispos* 15(3): 300-304. (Support not reported. Authors affiliated with National Cancer Institute; University of Washington; Pathfinder Laboratories.)
- 309. Keil CB, Akbar-Khanzadeh F, Konecny KA. 2001. Characterizing formaldehyde emission rates in a gross anatomy laboratory. *Appl Occup Environ Hyg* 16(10): 967-72 (as cited in IARC 2006).
- 310. Keller DA, Heck HD, Randall HW, Morgan KT. 1990. Histochemical localization of formaldehyde dehydrogenase in the rat. *Toxicol Appl Pharmacol* 106(2): 311-26. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC; E. I. Du Pont de Nemours & Co., DE.)
- 311. Kelly TJ, Mukund R, Spicer CW, Pollack AJ. 1994. Concentrations and transformations of hazardous air pollutants. *Environ Sci Technol* 28: 378A-387A. (Supported by the U.S. EPA. Authors affiliated with Battelle, OH.)

- 312. KEMI. 1993. *Formaldehyde*. Swedish Chemical Agency. <u>http://apps.kemi.se/flodessok/floden/kemamne\_Eng/formaldehyd\_eng.htm</u>. Accessed on 5/20/09.
- 313. Kennedy ER, Gagnon YT, Teass AW, Seitz T. 1992. Development and evaluation of a method to estimate potential formaldehyde dose from inhalable dust fibers. *Appl Occup Environ Hyg* 7: 231-240 (as cited in IARC 2006).
- 314. Kepler GM, Richardson RB, Morgan KT, Kimbell JS. 1998. Computer simulation of inspiratory nasal airflow and inhaled gas uptake in a rhesus monkey. *Toxicol Appl Pharmacol* 150(1): 1-11. (Supported by U.S. EPA. Authors affiliated with Chemical Industry Institute of Toxicology, NC; North Carolina State University, NC.)
- 315. Kerfoot EJ, Mooney TF. 1975. Formaldehyde and paraformaldehyde study in funeral homes. *Am Ind Hyg Assoc J* 36(7): 533-7 (as cited in IARC 2006).
- 316. Kernan GJ, Ji BT, Dosemeci M, Silverman DT, Balbus J, Zahm SH. 1999. Occupational risk factors for pancreatic cancer: a case-control study based on death certificates from 24 U.S. states. *Am J Ind Med* 36(2): 260-270. (Support not reported. Authors affiliated with WS Atkins-Polska Ltd., Poland; NCI.)
- 317. Kerns WD, Pavkov KL, Donofrio DJ, Gralla EJ, Swenberg JA. 1983. Carcinogenicity of formaldehyde in rats and mice after long-term inhalation exposure. *Cancer Res* 43(9): 4382-92. (Supported by the Chemical Industry Institute of Toxicology. Authors affiliated with Battelle, OH; Chemical Industry Institute of Toxicology, NC; Smith, Kline & French Laboratories, PA.)
- 318. Khan MZ, Ali Z, Muhammad G, Khan A, Mahmood F. 2003. Pathological effects of formalin (37% formaldehyde) mixed in feed or administered into the crops of White Leghorn cockerels. *J Vet Med A Physiol Pathol Clin Med* 50(7): 354-358. (Support not reported. Authors affiliated with University of Agriculture, Pakistan.)
- 319. Khwaja HA. 1995. Atmospheric concentrations of carboxylic acids and related compounds at a semiurban site. *Atmos Environ* 29: 127-139. (as cited in IARC 2006).
- 320. Kiec-Swierczynska M. 1996. Occupational allergic contact dermatitis in Lodz: 1990-1994. Occup Med (Lond) 46(3): 205-8. (Supported by the National Committee for Scientific Research. Authors affiliated with Nofer Institute of Occupational Medicine, Poland.)
- 321. Kilburn KH, Seidman BC, Warshaw R. 1985a. Neurobehavioral and respiratory symptoms of formaldehyde and xylene exposure in histology technicians. *Arch Environ Health* 40(4): 229-33. (Support not reported. Authors affiliated with University of Southern California, CA.)

- 322. Kilburn KH, Warshaw R, Thornton JC. 1987. Formaldehyde impairs memory, equilibrium, and dexterity in histology technicians: effects which persist for days after exposure. *Arch Environ Health* 42(2): 117-20. (Support not reported. Authors affiliated with University of Southern California, CA; Mount Sinai School of Medicine, NY.)
- 323. Kilburn KH, Warsaw RH. 1992. Neurobehavioral effects of formaldehyde and solvents on histology technicians: repeated testing across time. *Environ Res* 58(2): 134-146. (Support not reported. Authors affiliated with University of Southern California, CA; Workers Disease Detection Services, Inc., CA.)
- 324. Kilburn KH. 2000. Indoor air effects after building renovation and in manufactured homes. Am J Med Sci 320(4): 249-54. (Support not reported. Authors affiliated with University of Southern California, CA.)
- 325. Kilburn KH. 2001. Function testing for chemical brain damage: a review. *Arch Environ Health* 56(2): 132-7. (Support not reported. Authors affiliated with University of Southern California, CA.)
- 326. Kim CW, Song JS, Ahn YS, Park SH, Park JW, Noh JH, Hong CS. 2001. Occupational asthma due to formaldehyde. *Yonsei Med J* 42(4): 440-5 (as cited in IARC 2006).
- 327. Kim H, Kim YD, Cho SH. 1999. Formaldehyde exposure levels and serum antibodies to formaldehyde-human serum albumin of Korean medical students. *Arch Environ Health* 54(2): 115-8 (as cited in IARC 2006).
- 328. Kim WJ, Terada N, Nomura T, Takahashi R, Lee SD, Park JH, Konno A. 2002. Effect of formaldehyde on the expression of adhesion molecules in nasal microvascular endothelial cells: the role of formaldehyde in the pathogenesis of sick building syndrome. *Clin Exp Allergy* 32(2): 287-95. (Support not reported. Authors affiliated with Chiba University School of Medicine, China; Hana Nose Institute, Korea.)
- 329. Kimbell JS, Gross EA, Joyner DR, Godo MN, Morgan KT. 1993. Application of computational fluid dynamics to regional dosimetry of inhaled chemicals in the upper respiratory tract of the rat. *Toxicol Appl Pharmacol* 121(2): 253-263. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)
- 330. Kimbell JS, Gross EA, Richardson RB, Conolly RB, Morgan KT. 1997. Correlation of regional formaldehyde flux predictions with the distribution of formaldehyde-induced squamous metaplasia in F344 rat nasal passages. *Mutat Res* 380(1-2): 143-154. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)
- 331. Kimbell JS, Subramaniam RP. 2001. Use of computational fluid dynamics models for dosimetry of inhaled gases in the nasal passages. *Inhal Toxicol* 13(5):

325-34. (Support not reported. Authors affiliated with CIIT Centers for Health Research, NC; U.S. EPA.)

- 332. Kimbell JS, Subramaniam RP, Gross EA, Schlosser PM, Morgan KT. 2001a. Dosimetry modeling of inhaled formaldehyde: comparisons of local flux predictions in the rat, monkey, and human nasal passages. *Toxicol Sci* 64(1): 100-110. (Supported by CIIT member companies and the American Chemistry Council. Authors affiliated with CIIT, NC; U.S. EPA; GlaxoSmithKline, NC.)
- 333. Kimbell JS, Overton JH, Subramaniam RP, Schlosser PM, Morgan KT, Conolly RB, Miller FJ. 2001b. Dosimetry modeling of inhaled formaldehyde: binning nasal flux predictions for quantitative risk assessment. *Toxicol Sci* 64(1): 111-121. (Supported by CIIT member companies and the American Chemistry Council. Authors affiliated with CIIT, NC; U.S. EPA; GlaxoSmithKline, NC.)
- 334. Kinney PL, Chillrud SN, Ramstrom S, Ross J, Spengler JD. 2002. Exposures to multiple air toxics in New York City. *Environ Health Perspect* 110 Suppl 4: 539-46 (as cited in IARC 2006).
- 335. Kitaeva LV, Kitaev EM, Pimenova MN. 1990. [The cytopathic and cytogenetic sequelae of chronic inhalational exposure to formaldehyde on female germ cells and bone marrow cells in rats]. *Tsitologiia* 32(12): 1212-1216. (Support not identified due to foreign language. Authors affiliated with Acedemy of the Medical Sciences of the USSR, Russia.)
- 336. Kitaeva LV, Mikheeva EA, Shelomova LF, Shvartsman PY. 1996. Genotoxic effects of formaldehyde in somatic human cells *in vivo*. *Genetika* 32: 1287-1290. (Support not identified due to foreign language. Authors affiliated with Mechnikov State Medical Academy, Russia; Gertsen State Pedagogical University, Russia.)
- 337. Kligerman AD, Phelps MC, Erexson GL. 1984. Cytogenetic analysis of lymphocytes from rats following formaldehyde inhalation. *Toxicol Lett* 21(3): 241-246. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)
- 338. Konopinski VJ. 1983. Formaldehyde in office and commercial environments. *Am Ind Hyg Assoc J* 44(3): 205-8 (as cited in ATSDR 1999).
- 339. Koppel C, Baudisch H, Schneider V, Ibe K. 1990. Suicidal ingestion of formalin with fatal complications. *Intensive Care Med* 16(3): 212-4. (Support not reported. Authors affiliated with Freie Universitat Berlin, Germany; Landesuntersuchungsinstitut fur Lebensmittel, Germany.)
- 340. Korczynski RE. 1994. Formaldehyde exposure in the funeral industry. *Appl Occup Environ Hyg* 9: 575-579 (as cited in IARC 2006).

- 341. Korczynski RE. 1996. Effectiveness of downdraft ventilation in morgues. *Appl Occup Environ Hyg* 11: 5-8 (as cited in IARC 2006).
- 342. Korhonen K, Liukkonen T, Ahrens W, Astrakianakis G, Boffetta P, Burdorf A, Heederik D, Kauppinen T, Kogevinas M, Osvoll P, Rix BA, Saalo A, Sunyer J, Szadkowska-Stanczyk I, Teschke K, Westberg H, Widerkiewicz K. 2004. Occupational exposure to chemical agents in the paper industry. *Int Arch Occup Environ Health* 77(7): 451-460. (Supported by the European Commission. Authors affiliated with Lappeenranta Regional Institute of Occupational Health, Finland; Bremen Institute for Prevention Research and Social Medicine, Germany; BC Cancer Agency, Canada; IARC; Erasmus University, Netherlands; University of Utrecht, Netherlands; Institute of Occupational Health, Finland; Institut Municipal d'Investigacio Medica, Spain; National Institute of Occupational Health, Norway; Danish Cancer Society, Denmark; Nofer Institute of Occupational Medicine, Poland; University of British Columbia, Canada; Orebro Medical Center Hospital, Sweden; Regional Sanitary Epidemiological Station, Poland.)
- 343. Korky JK, Schwarz SR, Lustigman BK. 1987. Formaldehyde concentrations in biology department teaching facilities. *Bull Environ Contam Toxicol* 38(5): 907-10 (as cited in IARC 2006).
- 344. Krakowiak A, Gorski P, Pazdrak K, Ruta U. 1998. Airway response to formaldehyde inhalation in asthmatic subjects with suspected respiratory formaldehyde sensitization. *Am J Ind Med* 33(3): 274-81 (as cited in IARC 2006).
- 345. Krasner SW, McGuire MJ, Jacangelo JG, Patania NL, Reagan KM, Aieta EM. 1989. The occurrence of disinfection by-products in United States drinking water. *J Amer Water Works Assoc* 81(8): 41-53. (Supported by USEPA, AMWA, CDHS, and CPHF. Authors affiliated with Metropolitan Water District of Southern California, CA; James M. Montgomery Consulting Engineers, Inc., CA.)
- 346. Kreiger RA, Garry VF. 1983. Formaldehyde-induced cytotoxicity and sisterchromatid exchanges in human lymphocyte cultures. *Mutat Res* 120(1): 51-55. (Supported by the University of Minnesota and the Minnesota Medical Foundation. Authors affiliated with University of Minnesota, MN.)
- 347. Kriebel D, Sama SR, Cocanour B. 1993. Reversible pulmonary responses to formaldehyde. A study of clinical anatomy students. *Am Rev Respir Dis* 148(6 Pt 1): 1509-15 (as cited in IARC 2006).
- 348. Krones CJ, Conze J, Hoelzl F, Stumpf M, Klinge U, Moller M, Dott W, Schumpelick V, Hollender J. 2007. Chemical composition of surgical smoke produced by electrocautery, harmonic scalpel and argon beaming - a short study.

*European Surgery-Acta Chirurgica Austriaca* 39(2): 118-121. (Support not reported. Authors affiliated with Technical University of Aachen, Germany.)

- 349. Krzyzanowski M, Quackenboss JJ, Lebowitz MD. 1990. Chronic respiratory effects of indoor formaldehyde exposure. *Environ Res* 52: 117-125. (Supported by the EPA, EPRI and NIH. Authors affiliated with University of Arizona, AZ; National Institute of Hygiene, Poland.)
- 350. Ku RH, Billings RE. 1984. Relationships between formaldehyde metabolism and toxicity and glutathione concentrations in isolated rat hepatocytes. *Chem Biol Interact* 51(1): 25-36. (Supported by NIEHS. Authors affiliated with University of Texas Health Science Center, TX.)
- 351. Kuljac J. 1983. Formaldehyde in residences and workrooms. *J Yugosl Union Air Pollut Prev Assoc* 11: 11-14 (as cited in WHO 1989).
- 352. Kulle TJ, Sauder LR, Hebel JR, Green DJ, Chatham MD. 1987. Formaldehyde dose-response in healthy nonsmokers. *J Air Pollut Control Assoc* 37: 919-924 (as cited in IARC 2006).
- 353. Kulle TJ. 1993. Acute odor and irritation response in healthy nonsmokers with formaldehyde exposure. *Inhal Toxicol* 5: 323-332 (as cited in IARC 2006).
- 354. Kum C, Kiral F, Sekkin S, Seyrek K, Boyacioglu M. 2007a. Effects of xylene and formaldehyde inhalations on oxidative stress in adult and developing rats livers. *Exp Anim* 56(1): 35-42. (Support not reported. Authors affiliated with Adnan Menderes University, Turkey.)
- 355. Kuo H, Jian G, Chen C, Liu C, Lai J. 1997. White blood cell count as an indicator of formaldehyde exposure. *Bull Environ Contam Toxicol* 59(2): 261-7. (Support not reported. Authors affiliated with China Medical College, China.)
- 356. Laforest L, Luce D, Goldberg P, Bégin D, Gérin M, Demers PA, Brugère J, Leclerc A. 2000. Laryngeal and hypopharyngeal cancers and occupational exposure to formaldehyde and various dusts: a case-control study in France. Occup Environ Med 57(11): 767-773. (Support not reported. Authors affiliated with Institut National de la Sante et de la Recherche Medicale, France; Universite de Montreal, Canada; University of British Columbia, Canada; Institut Curie, France.)
- 357. Lam CW, Casanova M, Heck HD. 1985. Depletion of nasal mucosal glutathione by acrolein and enhancement of formaldehyde-induced DNA-protein crosslinking by simultaneous exposure to acrolein. *Arch Toxicol* 58(2): 67-71. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)

- 358. Lamb B, Westberg H, Bryant P. 1985. Air filtration rates in pre- and postweathered houses. *J Air Pollut Control Assoc* 35: 541-551 (as cited in ATSDR 1999).
- 359. Lamont Moore L, Ogrodnik EC. 1986. Occupational exposure to formaldehyde in mortuaries. *J Environ Health* 49: 32-35 (as cited in IARC 2006).
- 360. Lang I, Bruckner T, Triebig G. 2008. Formaldehyde and chemosensory irritation in humans: a controlled human exposure study. *Regul Toxicol Pharmacol* 50(1): 23-36. (Supported by FormaCare sector group of CE-FIC. Authors affiliated with University of Heidelberg, Germany.)
- 361. Lavoue J, Begin D, Beaudry C, Gerin M. 2007. Monte Carlo simulation to reconstruct formaldehyde exposure levels from summary parameters reported in the literature. *Annals Of Occupational Hygiene* 51(2): 161-172. (Support not reported. Authors affiliated with Universite de Montreal, Canada.)
- 362. Lavoue J, Vincent R, Gerin M. 2008. Formaldehyde exposure in U.S. industries from OSHA air sampling data. *J Occup Environ Hyg* 5(9): 575-87. (Supported by the Quebec Association for Occupational Hygiene, Health and Safety and the IRSST. Authors affiliated with Universite de Montreal, Canada; INRS, France.)
- 363. Lazutka JR, Lekevicius R, Dedonyte V, Maciuleviciute-Gervers L, Mierauskiene J, Rudaitiene S, Slapsyte G. 1999. Chromosomal aberrations and sister-chromatid exchanges in Lithuanian populations: effects of occupational and environmental exposures. *Mutat Res* 445(2): 225-239. (Supported by Lithuanian Ministry of Health Care, the Municipality of the Kedainiai district, the Open Society Fund-Lithuania, and the National Sciences Program 'Atomic Energetics and the Environment'. Authors affiliated with Vilnius University, Lithuania.)
- 364. Lee S, Radtke T. 1998. Exposure to formaldehyde among fish hatchery workers. *Appl Occup Environ Hyg* 13: 3-6 (as cited in IARC 2006).
- 365. Leiper K, Morris AI. 2007. Treatment of radiation proctitis. *Clin Oncol (R Coll Radiol)* 19(9): 724-9. (Support not reported. Authors affiliated with Royal Liverpool University Hospital, UK.)
- 366. Lemus R, Abdelghani AA, Akers TG, Horner WE. 1998. Potential health risks from exposure to indoor formaldehyde. *Rev Environ Health* 13(1-2): 91-8 (as cited in IARC 2006).
- 367. Levine RJ, Andjelkovich DA, Shaw LK. 1984. The mortality of Ontario undertakers and a review of formaldehyde-related mortality studies. *J Occup Med* 26(10): 740-746. (Supported by the Chemical Industry Institute of Toxicology. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)

- 368. Levy S, Nocentini S, Billardon C. 1983. Induction of cytogenetic effects in human fibroblast cultures after exposure to formaldehyde or X-rays. *Mutat Res* 119(3): 309-317. (Support not reported. Authors affiliated with Institut Curie, France.)
- 369. Li H, Wang J, Konig R, Ansari GA, Khan MF. 2007a. Formaldehyde-protein conjugate-specific antibodies in rats exposed to formaldehyde. *J Toxicol Environ Health A* 70(13): 1071-5. (Supported by the Texas Advanced Technology Program, Texas Higher Education Coordinating Board. Authors affiliated with University of Texas Medical Branch, TX.)
- 370. Li JR, Zhu JL, Ye LF. 2007b. Determination of formaldehyde in squid by highperformance liquid chromatography. *Asia Pacific Journal Of Clinical Nutrition* 16: 127-130. (Support not reported. Authors affiliated with Zhejiang Gongshang University, China.)
- 371. Li M, Cheng J, Zhao Y, Zhu Y, Pan Y. 1988. The study on sister chromatid exchange in lymphocytes of workers exposed to formaldehyde. *Shanghai Med* 11(10): 581-583 (as cited in Tang *et al* 2009).
- 372. Li Z, Lian S, Cai H, Zhang F, Pei H. 1999. Hazard on worker occupationally exposed to formaldehyde. *Chin Occup Med* 26(4): 57-58 (as cited in Tang *et al.* 2009).
- 373. Li ZG, Chen BC. 2002. Effect of low concentration formaldehyde on the health of workers. *Chin J Ind Med* 15(5): 302-303 (as cited in Tang *et al* 2009).
- 374. Liber HL, Benforado K, Crosby RM, Simpson D, Skopek TR. 1989. Formaldehyde-induced and spontaneous alterations in human *hprt* DNA sequence and mRNA expression. *Mutat Res* 226(1): 31-37. (Support not reported. Authors affiliated with Harvard School of Public Health, MA; Chemical Industry Institute of Toxicology, NC.)
- 375. Liebling T, Rosenman KD, Pastides H, Griffith RG, Lemeshow S. 1984. Cancer mortality among workers exposed to formaldehyde. *Am J Ind Med* 5(6): 423-428. (Support not reported. Authors affiliated with University of Massachusetts, MA; State of New Jersey Department of Health.)
- 376. Lillienberg L, Burdorf A, Mathiasson L, Thorneby L. 2008. Exposure to metalworking fluid aerosols and determinants of exposure. Ann Occup Hyg 52(7): 597-605. (Supported by the Swedish Council for Working Life and Social Research. Authors affiliated with Sahlgrenska University Hospital, Sweden; Erasmus University, Netherlands; Lund University, Sweden.)
- 377. Lindstrom AB, Proffitt D, Fortune CR. 1995. Effects of modified residential construction on indoor air quality. *Indoor Air* 5: 258-269 (as cited in IARC 2006).

- 378. Lino dos Santos Franco A, Damazo AS, Beraldo de Souza HR, Domingos HV, Oliveira-Filho RM, Oliani SM, Costa SK, Tavares de Lima W. 2006. Pulmonary neutrophil recruitment and bronchial reactivity in formaldehyde-exposed rats are modulated by mast cells and differentially by neuropeptides and nitric oxide. *Toxicol Appl Pharmacol* 214(1): 35-42. (Supported by Fundacao de Amparo a Pesquisa do Estado de Sao Paulo and Conselho Nacional de Pesquisa. Authors affiliated with University of Sao Paulo, Brazil; University of Sao Paulo State, Brazil; UNIFESP, Brazil. )
- 379. Lino dos Santos Franco A, Domingos HV, Damazo AS, Breithaupt-Faloppa AC, de Oliveira AP, Costa SK, Oliani SM, Oliveira-Filho RM, Vargaftig BB, Tavares-de-Lima W. 2009. Reduced allergic lung inflammation in rats following formaldehyde exposure: long-term effects on multiple effector systems. *Toxicology* 256(3): 157-63. (Supported by Fundacao de Amparo a Pesquisa do Estado de Sao Paulo and Conselho Nacional de Pequisa. Authors affiliated with University of Sao Paulo, Brazil; Sao Paulo State University, Brazil.)
- 380. Linos A, Blair A, Cantor KP, Burmeister L, VanLier S, Gibson RW, Schuman L, Everett G. 1990. Leukemia and non-Hodgkin's lymphoma among embalmers and funeral directors. *J Natl Cancer Inst* 82(1): 66. (Support not reported. Authors affiliated with Athens Medical School, Greece; NCI; University of Iowa, IO; University of Minnesota, MN; Orlando Regional Medical Center, FL.)
- 381. Liteplo RG, Meek ME. 2003. Inhaled formaldehyde: exposure estimation, hazard characterization, and exposure-response analysis. *J Toxicol Environ Health B Crit Rev* 6(1): 85-114. (Supported by the Formaldehyde Epidemiology, Toxicology and Environmental Group, Inc. Authors affiliated with Health Canada.)
- 382. Liu Y, Li CM, Lu Z, Ding S, Yang X, Mo J. 2006. Studies on formation and repair of formaldehyde-damaged DNA by detection of DNA-protein crosslinks and DNA breaks. *Front Biosci* 11: 991-997. (Supported by the China National Key Technologies R&D Program for the 10th 5-Year Plan from the Chinese Ministry of Science and Technology and the Nanyang Technological University of Singapore. Authors affiliated with Central China Normal University, China; Nanyang Technological University, Singapore; Kumetrix, Inc., CA.)
- 383. Liu YR, Zhou Y, Qiu W, Zeng JY, Shen LL, Li AP, Zhou JW. 2009b. Exposure to formaldehyde induces heritable DNA mutations in mice. *J Toxicol Environ Health A* 72(11): 767-73. (Supported by the National Natural Science Foundation of China. Authors affiliated with Nanjing Medical University, China.)
- 384. Liu YY, Lin TC, Wang YJ, Ho WL. 2009a. Carbonyl Compounds and Toxicity Assessments of Emissions from a Diesel Engine Running on Biodiesels. *Journal Of The Air & Waste Management Association* 59(2): 163-171. (Supported by the

Taiwan National Science Council. Authors affiliated with National Cheng Kung University, China.)

- 385. Lodén M. 1986b. The *in vitro* permeability of human skin to benzene, ethylene glycol, formaldehyde, and n-hexane. *Acta Pharmacol Toxicol (Copenh)* 58(5): 382-389. (Supported by the Swedish Work Environment Fund. Authors affiliated with National Defense Research Institute, Sweden.)
- 386. Logue JN, Barrick MK, Jessup GL, Jr. 1986. Mortality of radiologists and pathologists in the Radiation Registry of Physicians. *J Occup Med* 28(2): 91-99. (Supported by the American College of Radiology and the College of American Pathologists. Authors affiliated with U.S. FDA; Pennsylvania Department of Health.)
- 387. Loomis TA. 1979. Formaldehyde toxicity. Arch Pathol Lab Med 103(7): 321-4.(Support not reported. Authors affiliated with University of Washington, WA.)
- 388. Lovschall H, Eiskjaer M, Arenholt-Bindslev D. 2002. Formaldehyde cytotoxicity in three human cell types assessed in three different assays. *Toxicol In Vitro* 16(1): 63-69. (Supported by Calcin-fondin. Authors affiliated with University of Aarhus, Denmark.)
- 389. Lu K, Boysen G, Gao L, Collins LB, Swenberg JA. 2008a. Formaldehydeinduced histone modifications in vitro. *Chem Res Toxicol* 21(8): 1586-93. (Supported by NIH and the Formaldehyde Council, Inc. Authors affiliated with University of North Carolina, NC.)
- 390. Lu K, Ye W, Gold A, Ball LM, Swenberg JA. 2009. Formation of S-[1-(N(2)-Deoxyguanosinyl)methyl]glutathione between Glutathione and DNA Induced by Formaldehyde. *J Am Chem Soc*. (Supported by NIH and the Formaldehyde Council, Inc. Authors affiliated with University of North Carolina, NC.)
- 391. Lu Y, Chen XJ, Yang XY, Xue ZQ. 2007. A survey of the effection to teachers' health from formaldehyde contact. *J Xinjiang Med Univ* 30(3): 234 (as cited in Tang *et al.* 2009).
- 392. Lu Z, Li CM, Qiao Y, Yan Y, Yang X. 2008b. Effect of inhaled formaldehyde on learning and memory of mice. *Indoor Air* 18(2): 77-83. (Supported by the China National Science Foundation, the Chinese Ministry of Science and the Nanyang Technological University. Authors affiliated with Central China Normal University, China; Nanyang Technological University, China.)
- 393. Luce D, Gérin M, Leclerc A, Morcet JF, Brugère J, Goldberg M. 1993a. Sinonasal cancer and occupational exposure to formaldehyde and other substances. *Int J Cancer* 53(2): 224-231. (Support not reported. Authors affiliated with INSERM, France; Universite de Montreal, Canada; Institut Curie, France.)

- 394. Luce D, Leclerc A, Bégin D, Demers PA, Gérin M, Orlowski E, Kogevinas M, Belli S, Bugel I, Bolm-Audorff U, Brinton LA, Comba P, Hardell L, Hayes RB, Magnani C, Merler E, Preston-Martin S, Vaughan TL, Zheng W, Boffetta P. 2002. Sinonasal cancer and occupational exposures: a pooled analysis of 12 case-control studies. *Cancer Causes Control* 13(2): 147-157. (Supported by the Commission of the European Union, Directorate General for Employment, Industrial Relations, and Social Affairs and the Biomedical and Health Research Programme. Authors affiliated with INSERM, France; Universite de Montreal, Canada; University of British Columbia, Canada; Institut Municipal d'Investigacio Medica, Spain; Instituto Superiore di Sanita, Italy; Hessiches Ministerium fur Frauen, Arbeit und Sozialordnung, Germany; NCI; Orebro Medical Center, Sweden; University of Turin, Italy; Centro per lo Studio et la Prevenzione Oncologica, Italy; University of Southern California, CA; Fred Hutchinson Cancer Research Center, WA; Vanderbilt University, TN; IARC.)
- 395. Luker MA, Van Houten RW. 1990. Control of formaldehyde in a garment sewing plant. Am Ind Hyg Assoc J 51: 541-544. (Support not reported. Authors affiliated with Liberty Mutual Insurance Company, TN and MA; Metcalf & Eddy, Inc., GA.)
- 396. Lutz WK. 1986. Endogenous formaldehyde does not produce detectable DNAprotein crosslinks in rat liver. *Toxicol Pathol* 14(4): 462-465. (Supported by the Swiss National Science Foundation. Authors affiliated with University of Zurich, Switzerland.)
- 397. Lyapina M, Zhelezova G, Petrova E, Boev M. 2004. Flow cytometric determination of neutrophil respiratory burst activity in workers exposed to formaldehyde. *Int Arch Occup Environ Health* 77(5): 335-40. (Supported by the Faculty of Medicine at the Medical University, Sofia. Authors affiliated with Medical University, Bulgaria; University Hospital of St. Ivan Rilski, Bulgaria.)
- 398. Ma TH, Harris MM. 1988. Review of the genotoxicity of formaldehyde. *Mutat Res* 196(1): 37-59. (Support not reported. Authors affiliated with Western Illinois University, IL.)
- 399. Mackerer CR, Angelosanto FA, Blackburn GR, Schreiner CA. 1996. Identification of formaldehyde as the metabolite responsible for the mutagenicity of methyl *tertiary*-butyl ether in the activated mouse lymphoma assay. *Proc Soc Exp Biol Med* 212(4): 338-341. (Support not reported. Authors affiliated with Stonybrook Laboratories, Inc., NJ, a subsidiary of Mobil Oil.)
- 400. Madison RE, Broughton A, Thrasher JD. 1991. Immunologic biomarkers associated with an acute exposure to exothermic byproducts of a ureaformaldehyde spill. *Environ Health Perspect* 94: 219-23. (Support not reported. Authors affiliated with California State University, CA; Antibody Assay Laboratories, CA; Thrasher and Associates, CA.)
- 401. Magaña-Schwencke N, Ekert B, Moustacchi E. 1978. Biochemical analysis of damage induced in yeast by formaldehyde. I. Induction of single-strand breaks in DNA and their repair. *Mutat Res* 50(2): 181-193. (Supported by Electricite de France, the SEITA, and Euratom. Authors affiliated with Fondation Curie Institut du Radium, France. )
- 402. Maibach HI. 1983. Formaldehyde: Effects on Animal and Human Skin. In *Formaldehyde Toxicity*. Gibson JE, ed. Washington, D.C.: Hemisphere. p. 166-174. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)
- 403. Main DM, Hogan TJ. 1983. Health effects of low-level exposure to formaldehyde. *J Occup Med* 25(12): 896-900. (Support not reported. Authors affiliated with University of Illinois.)
- 404. Maitre A, Soulat JM, Masclet P, Stoklov M, Marques M, de Gaudemaris R. 2002. Exposure to carcinogenic air pollutants among policemen working close to traffic in an urban area. *Scand J Work Environ Health* 28(6): 402-10 (as cited in IARC 2006).
- 405. Majumder PK, Kumar VL. 1995. Inhibitory effects of formaldehyde on the reproductive system of male rats. *Indian J Physiol Pharmacol* 39(1): 80-82. (Support not reported. Authors affiliated with All India Institute of Medical Sciences, India)
- 406. Makar AB, McMartin KE, Palese M, Tephly TR. 1975. Formate assay in body fluids: application in methanol poisoning. *Biochem Med* 13(2): 117-26 (as cited in WHO 1989).
- 407. Makinen M, Kalliokoski P, Kangas J. 1999. Assessment of total exposure to phenol-formaldehyde resin glue in plywood manufacturing. *Int Arch Occup Environ Health* 72(5): 309-14 (as cited in IARC 2006).
- 408. Malaka T, Kodama AM. 1990. Respiratory health of plywood workers occupationally exposed to formaldehyde. *Arch Environ Health* 45: 288-294. (Support not reported. Authors affiliated with University of Hawaii, Hawaii.)
- 409. Malek FA, Moritz KU, Fanghanel J. 2003. Formaldehyde inhalation & open field behaviour in rats. *Indian J Med Res* 118: 90-6. (Support not reported. Authors affiliated with Ernst-Moritz-Arndt-University, Germany.)
- 410. Malek FA, Moritz KU, Fanghanel J. 2004. Effects of a single inhalative exposure to formaldehyde on the open field behavior of mice. *Int J Hyg Environ Health* 207(2): 151-158. (Support not reported. Authors affiliated with Ernst-Moritz-Arndt-University, Germany.)

- 411. Mallinckrodt. 2009. *Material Safety Data Sheet: Paraformaldehyde*. Mallinckrodt Baker, Inc. <u>http://www.jtbaker.com/msds/englishhtml/p0154.htm</u>. Accessed on 5/27/09.
- 412. Marsh GM. 1982. Proportional mortality patterns among chemical plant workers exposed to formaldehyde. *Br J Ind Med* 39(4): 313-322. (Supported by the Monsanto Company. Authors affiliated with the University of Pittsburgh, PA.)
- 413. Marsh GM, Stone RA, Henderson VL. 1992a. A reanalysis of the National Cancer Institute study on lung cancer mortality among industrial workers exposed to formaldehyde. *J Occup Med* 34(1): 42-44. (Supported by the Formaldehyde Institute. Authors affiliated with University of Pittsburgh, PA.)
- 414. Marsh GM, Stone RA, Henderson VL. 1992b. Lung cancer mortality among industrial workers exposed to formaldehyde: a Poisson regression analysis of the National Cancer Institute Study. *Am Ind Hyg Assoc J* 53(11): 681-691. (Supported by the Formaldehyde Institute. Authors affiliated with University of Pittsburgh, PA.)
- 415. Marsh GM, Stone RA, Esmen NA, Henderson VL. 1994a. Mortality patterns among chemical plant workers exposed to formaldehyde and other substances. *J Natl Cancer Inst* 86(5): 384-386. (Supported by the American Cyanamid Company. Authors affiliated with University of Pittsburgh, PA; Esmen Research and Engineering, PA.)
- 416. Marsh GM, Stone RA, Henderson VL, Esmen NA. 1994b. Misclassification of nasopharyngeal cancer reply. *J Natl Cancer Inst* 86: 1557. (Support not reported. Authors affiliated with University of Pittsburgh, PA; Esmen Research and Engineering, PA.)
- 417. Marsh GM, Stone RA, Esmen NA, Henderson VL, Lee KY. 1996. Mortality among chemical workers in a factory where formaldehyde was used. *Occup Environ Med* 53(9): 613-627. (Supported by the American Cyanamid Company. Authors affiliated with University of Pittsburgh, PA; University of Oklahoma, OK; Department of Veteran Affairs, Washington, D.C.)
- 418. Marsh GM, Youk AO, Stone RA, Buchanich JM, Gula MJ, Smith TJ, Quinn MM. 2001. Historical cohort study of US man-made vitreous fiber production workers: I. 1992 fiberglass cohort follow-up: initial findings. *J Occup Environ Med* 43(9): 741-756. (Supported by the North American Insulation Manufacturers Association. Authors affiliated with University of Pittsburgh, PA; Harvard School of Public Health, MA; University of Massachusetts Lowell, MA.)
- 419. Marsh GM, Youk AO, Buchanich JM, Cassidy LD, Lucas LJ, Esmen NA, Gathuru IM. 2002. Pharyngeal cancer mortality among chemical plant workers exposed to formaldehyde. *Toxicol Ind Health* 18(6): 257-268. (Supported by

Cytec Industries, Inc. Authors affiliated with University of Pittsburgh, PA; University of Oklahoma, OK.)

- 420. Marsh GM, Youk AO. 2004. Reevaluation of mortality risks from leukemia in the formaldehyde cohort study of the National Cancer Institute. *Regul Toxicol Pharmacol* 40(2): 113-124. (Supported by the Formaldehyde Council, Inc. Authors affiliated with University of Pittsburgh, PA.)
- 421. Marsh GM, Youk AO. 2005. Reevaluation of mortality risks from nasopharyngeal cancer in the formaldehyde cohort study of the National Cancer Institute. *Regul Toxicol Pharmacol* 42(3): 275-283. (Supported by the Formaldehyde Council, Inc. Authors affiliated with University of Pittsburgh, PA.)
- 422. Marsh GM, Youk AO, Buchanich JM, Erdal S, Esmen NA. 2007a. Work in the metal industry and nasopharyngeal cancer mortality among formaldehyde-exposed workers. *Regul Toxicol Pharmacol* 48(3): 308-319. (Supported by Cytec Industries Inc. (Cytec). Authors affiliated with University of Pittsburgh, PA; University of Illinois at Chicago, IL.)
- 423. Marsh GM, Youk AO, Morfeld P. 2007b. Mis-specified and non-robust mortality risk models for nasopharyngeal cancer in the National Cancer Institute formaldehyde worker cohort study. *Regul Toxicol Pharmacol* 47(1): 59-67. (Supported by the European Chemical Industry Council. Authors affiliated with University of Pittsburgh, PA; Cologne University Medical School, Germany; Institute for Occupational Sciences of RAG Aktiengesellschaft, Germany.)
- 424. Martin CN, McDermid AC, Garner RC. 1978. Testing of known carcinogens and noncarcinogens for their ability to induce unscheduled DNA synthesis in HeLa cells. *Cancer Res* 38(8): 2621-2627. (Supported by the Yorkshire Cancer Research Campaign and the Medical Research Council. Authors affiliated with University of York, UK.)
- 425. Materna BL, Jones JR, Sutton PM, Rothman N, Harrison RJ. 1992.
  Occupational exposures in California wildland fire fighting. *Am Ind Hyg Assoc J* 53(1): 69-76 (as cited in IARC 2006).
- 426. McDuffie HH, Pahwa P, McLaughlin JR, Spinelli JJ, Fincham S, Dosman JA, Robson D, Skinnider LF, Choi NW. 2001. Non-Hodgkin's lymphoma and specific pesticide exposures in men: cross-Canada study of pesticides and health. *Cancer Epidemiol Biomarkers Prev* 10(11): 1155-63. (Supported by Health Canada, the British Columbia Health Research Foundation, and the University of Saskatchewan. Authors affiliated with University of Saskatchewan, Canada; University of Toronto, Canada; St. Paul's Hospital, Canada; Alberta Cancer Board, Canada; Saskatchewan Cancer Agency, Canada; Manitoba Cancer Treatment and Research Foundation, Canada.)

- 427. McGregor D, Bolt H, Cogliano V, Richter-Reichhelm HB. 2006. Formaldehyde and glutaraldehyde and nasal cytotoxicity: case study within the context of the 2006 IPCS Human Framework for the Analysis of a cancer mode of action for humans. *Crit Rev Toxicol* 36(10): 821-35. (Support not reported. Authors affiliated with Toxicity Evaluation Consultants, UK; Institut fur Arbeitsphysiologie, Germany; IARC; Federal Institute for Risk Assessment, Germany.)
- 428. McGuire MT, Casserly DM, Greff RM. 1992. formaldehyde concentrations in fabric stores. *Appl Occup Environ Hyg* 7: 112-119 (as cited in IARC 2006).
- 429. McMartin KE, Martin-Amat G, Noker PE, Tephly TR. 1979. Lack of a role for formaldehyde in methanol poisoning in the monkey. *Biochem Pharmacol* 28(5): 645-9. (Supported by NIH. Authors affiliated with University of Iowa, IA.)
- 430. McNary JE, Jackson EM. 2007. Inhalation exposure to formaldehyde and toluene in the same occupational and consumer setting. *Inhal Toxicol* 19(6-7): 573-6. (Support not reported. Authors affiliated with Clayton Environmental Consultants, CA; Jackson Research Associates, Inc., WA.)
- 431. MedScape. 2006. *Monograph Methenamine, Methenamine Hippurate, Methenamine Mandelate* MedScape. <u>http://www.medscape.com/druginfo/monograph?cid=med&drugid=5658&drugn</u> <u>ame=Mandelamine+Oral&monotype=monograph&print=1</u>.
- 432. Merk O, Speit G. 1998. Significance of formaldehyde-induced DNA-protein crosslinks for mutagenesis. *Environ Mol Mutagen* 32(3): 260-268. (Supported by the Department for Environment Baden-Wurttemberg. Authors affiliated with Universitatklinikum Ulm, Germany.)
- 433. Merk O, Speit G. 1999. Detection of crosslinks with the comet assay in relationship to genotoxicity and cytotoxicity. *Environ Mol Mutagen* 33(2): 167-172. (Supported by the Department for Environment Baden-Wurttemberg. Authors affiliated with Universitatklinikum Ulm, Germany.)
- 434. Merletti F, Boffetta P, Ferro G, Pisani P, Terracini B. 1991. Occupation and cancer of the oral cavity or oropharynx in Turin, Italy. *Scand J Work Environ Health* 17(4): 248-254. (Supported by the Consiglio Nazionale della Ricerche, Rome, the Associazione Italiana per le Ricerca sul Cancro, the Ministry of Public Education and the Consorzio per il Sistema Informativo. Authors affiliated with University of Turin, Italy; IARC; National Cancer Institute, Italy.)
- 435. Mery S, Gross EA, Joyner DR, Godo M, Morgan KT. 1994. Nasal diagrams: a tool for recording the distribution of nasal lesions in rats and mice. *Toxicol Pathol* 22(4): 353-72. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)

- 436. Migliore L, Ventura L, Barale R, Loprieno N, Castellino S, Pulci R. 1989. Micronuclei and nuclear anomalies induced in the gastro-intestinal epithelium of rats treated with formaldehyde. *Mutagenesis* 4(5): 327-334. (Supported by the National Research Council and Progetto Strategico Mutagenesai. Authors affiliated with University of Pisa, Italy; University of Ferrara, Italy; Farmitalia Carlo Erba, Italy.)
- 437. Miguel A, H., De Aquino Neto FR, Cardoso JN, Vasconcellos PC, Pereira AS, Marquez KSG. 1995. Characterization of indoor air quality in the cities of Sao Paulo and Rio de Janeiro, Brazil. *Environ Sci Technol* 29: 338-345 (as cited in IARC 2006).
- 438. Milton DK, Walters MD, Hammond K, Evans JS. 1996. Worker exposure to endotoxin, phenolic compounds, and formaldehyde in a fiberglass insulation manufacturing plant. *Am Ind Hyg Assoc J* 57: 889-896. (Supported by Owens-Corning Corporation, NIEHS and NIOSH. Authors affiliated with Harvard School of Public Health, MA; University of Massachusetts, MA; Polaroid Corp; University of California - Berkeley, CA.)
- 439. Miretskaya, Shvartsman PY. 1982. Studies of chromosome aberrations in human lymphocytes under the influence of formaldehyde. 1. Formaldehyde treatment of lymphocytes in vitro. *Tsitologiia* 24: 1056-1060 (as cited in IARC).
- 440. Mohammed MF, Kang D, Aneja VP. 2002. Volatile organic compounds in some urban locations in the United States. *Chemosphere* 47: 863-882. (Supported by the Commission for Education & Culture Exchange. Authors affiliated with North Carolina State University, NC; Alexandria University, Egypt; U.S. EPA.)
- 441. Monticello TM, Morgan KT, Everitt JI, Popp JA. 1989. Effects of formaldehyde gas on the respiratory tract of rhesus monkeys. Pathology and cell proliferation. *Am J Pathol* 134(3): 515-27. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)
- 442. Monticello TM, Miller FJ, Morgan KT. 1991. Regional increases in rat nasal epithelial cell proliferation following acute and subchronic inhalation of formaldehyde. *Toxicol Appl Pharmacol* 111(3): 409-421. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC; Pathology Associates, Inc., NC.)
- 443. Monticello TM, Swenberg JA, Gross EA, Leininger JR, Kimbell JS, Seilkop S, Starr TB, Gibson JE, Morgan KT. 1996. Correlation of regional and nonlinear formaldehyde-induced nasal cancer with proliferating populations of cells. *Cancer Res* 56(5): 1012-1022. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC; Analytical Sciences, Inc., NC; Pharmaceutical Research Institute, NJ; University of North Carolina, NC; Environ Corp, NC; DowElanco, IN.)

- 444. Monticello TM, Morgan KT. 1997. Chemically-induced nasal carcinogenesis and epithelial cell proliferation: a brief review. *Mutat Res* 380(1-2): 33-41. (Support not reported. Authors affiliated with Bristol Myers-Squibb Pharmaceutical Research Institute, NJ; Glaxo Wellcome, NC.)
- 445. Morgan KT, Jiang XZ, Starr TB, Kerns WD. 1986b. More precise localization of nasal tumors associated with chronic exposure of F-344 rats to formaldehyde gas. *Toxicol Appl Pharmacol* 82(2): 264-271. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC; Shanghai First Medical College, China; Smith Kline & French Laboratories, PA.)
- 446. Morgan KT, Monticello TM. 1990. Airflow, gas deposition, and lesion distribution in the nasal passages. *Environ Health Perspect* 85: 209-218. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)
- 447. Morgan KT. 1997. A brief review of formaldehyde carcinogenesis in relation to rat nasal pathology and human health risk assessment. *Toxicol Pathol* 25(3): 291-307. (Support not reported. Author affiliated with CIIT, NC; GlaxoWellcome, Inc., NC.)
- 448. Murrell W, Feron F, Wetzig A, Cameron N, Splatt K, Bellette B, Bianco J, Perry C, Lee G, Mackay-Sim A. 2005. Multipotent stem cells from adult olfactory mucosa. *Dev Dyn* 233(2): 496-515. (Supported by Queensland Health, the Garnett Passe and Rodney Williams Foundation and the Stanley Medical Research Institute. Authors affiliated with Griffith University, Australia; Princess Alexandra Hospital, Australia; Royal Adelaide Hospital, Australia; Universite de Marseille, France.)
- 449. Mutsuga M, Kawamura Y, Sugita-Konishi Y, Hara-Kudo Y, Takatori K, Tanamoto K. 2006. Migration of formaldehyde and acetaldehyde into mineral water in polyethylene terephthalate (PET) bottles. *Food Addit Contam* 23(2): 212-8. (Support not reported. Authors affiliated with National Institute of Health Sciences, Japan.)
- 450. Myers JA, Mall J, Doolas A, Jakate SM, Saclarides TJ. 1997. Absorption kinetics of rectal formalin instillation. *World J Surg* 21(8): 886-889. (Support not reported. Authors affiliated with Rush-Presbyterian-St. Luke's Medical Center, IL.)
- 451. Na K, Cocker DR. 2008. Fine organic particle, formaldehyde, acetaldehyde concentrations under and after the influence of fire activity in the atmosphere of Riverside, California. *Environ Res* 108(1): 7-14. (Support not reported. Authors affiliated with University of California Riverside, CA.)
- 452. Nacher V, Llombart C, Carretero A, Navarro M, Ysern P, Calero S, Figols E, Ruberte J. 2007. A new system to reduce formaldehyde levels improves safety conditions during gross veterinary anatomy learning. *J Vet Med Educ* 34(2):

168-71. (Support not reported. Authors affiliated with Autonomous University of Barcelona, Spain; Tecnologia y Distribucion Medico y Científica, Spain.)

- 453. Natarajan AT, Darroudi F, Bussman CJ, van Kesteren-van Leeuwen AC. 1983. Evaluation of the mutagenicity of formaldehyde in mammalian cytogenetic assays in vivo and vitro. *Mutat Res* 122(3-4): 355-360. (Supported by the Koningin Wilhelmina Fonds and the EEC Chemical Mutagen Program. Authors affiliated with University of Leiden, Netherlands; J.A. Cohen Institute of Radiopathology and Protection, Netherlands.)
- 454. Neri M, Bonassi S, Knudsen LE, Sram RJ, Holland N, Ugolini D, Merlo DF. 2006. Children's exposure to environmental pollutants and biomarkers of genetic damage. I. Overview and critical issues. *Mutat Res* 612(1): 1-13. (Supported by the European Union and the Associazione Italiana per la Ricerca sul Cancro. Authors affiliated with National Cancer Research Institute, Italy; University of Copenhagen, Denmark; Institute of Experimental Medicine, Czech Republic; Health Institute of Central Bohemia, Czech Republic; University of California Berkeley, CA; University of Genoa, Italy.)
- 455. Neuss S, Speit G. 2008. Further characterization of the genotoxicity of formaldehyde in vitro by the sister chromatid exchange test and co-cultivation experiments. *Mutagenesis* 23(5): 355-7. (Supported by the European Chemical Industry Council. Authors affiliated with Universitat Ulm, Germany.)
- 456. Newell GW. 1983. Chapter 1. Overview of Formaldehyde. In *Formaldehyde Toxicity*. Gibson JE, ed. Washington, D.C.: Hemispehere Publishing Corporation. p. 3 12. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)
- 457. Nilsson JA, Zheng X, Sundqvist K, Liu Y, Atzori L, Elfwing A, Arvidson K, Grafstrom RC. 1998. Toxicity of formaldehyde to human oral fibroblasts and epithelial cells: influences of culture conditions and role of thiol status. *J Dent Res* 77(11): 1896-903. (Supported by the Swedish Council for Forestry and Agricultural Research, Swedish Transport Research Board, Swedish National Board of Laboratory Animals, Swedish Cancer Society, Swedish Tobacco Company and the Health Effects Institute. Authors affiliated with Karolinska Institutet, Sweden.)
- 458. NIOSH. 1976a. *NIOSH Criteria for a Recommended Standard Occupational Exposure to Formaldehyde*. DHEW (NIOSH) Publication No. 77-126. Cincinnati, OH: National Institute for Occupational Safety and Health. 165 pp. (as cited in WHO 1989).
- 459. NIOSH. 1976b. *Health Hazard Evaluation Determination Report, Unitog Col, Warrensburg, Missouri*. Report 75-143-333. Washington, D.C.: National Institute for Occupational Safety and Health (as cited in WHO 1989).

- 460. NIOSH. 1976c. *Health Hazard Evaluation Report, Formica Corporation*. Report 75-145-327. Cincinnati, OH: National Institute for Occupational Safety and Health (as cited in WHO 1989).
- 461. NIOSH. 1979a. *Health Hazard Evaluation Determination, Commonwealth Trading Company*. Report HE78-116-557. Washington, D.C.: National Institute of Occupational Safety and Health (as cited in WHO 1989).
- 462. NIOSH. 1979b. *Industry Selection for Determination of Extent of Exposure*. Cincinnati, OH: National Institute for Occupational Safety and Health (as cited in WHO 1989).
- 463. NIOSH. 1980a. *Walk-Through Survey Report of Georgia-Pacific Chemical and Resin Division*. Cincinnati, OH: National Institute for Occupational Safety and Health (as cited in WHO 1989).
- 464. NIOSH. 1980b. An Industrial Hygiene Survey of Urea-Formaldehyde Foam Insulation Manufacturing at Rapco Foam, Inc., Enviro Control, Inc., Rockville, Maryland. NIOSH Contract No. 210-78-0081. Cincinnati, OH: National Institute for Occupational Safety and Health (as cited in WHO 1989).
- 465. NIOSH. 1980c. *Health Hazard Evaluation Determination, St. Regis Paper Company, Buckport, Maine*. HE 80-126-777. Washington, D.C.: National Institute for Occupational Safety and Health (as cited in WHO 1989).
- 466. NIOSH. 1981. Health Hazard Evaluation Determination, Rock Hill Printing and Finishing Company, Rock Hill, North Carolina. Report 80-126-777. Cincinnati, OH: National Institute for Occupational Safety and Health (as cited in WHO 1989).
- 467. NIOSH. 2001. *Metal Working Fluids: Recommendation for Chronic Inhalation Studies*. Cincinnati, OH: National Institute for Occupational Safety and Health. 90 pp.
- 468. Nishi K, Yamada M, Wakasugi C. 1988. Formaldehyde poisoning: report of an autopsy case. *Nihon Hoigaku Zasshi* 42(1): 85-9. (Support not reported. Authors affiliated with Wakayama Medical College, Japan; Nara Medical University, Japan.)
- 469. Nisse C, Haguenoer JM, Grandbastien B, Preudhomme C, Fontaine B, Brillet JM, Lejeune R, Fenaux P. 2001. Occupational and environmental risk factors of the myelodysplastic syndromes in the North of France. *Br J Haematol* 112(4): 927-935. (Supported by the Centre de Recherche en Sante-Travail-Ergonomie, the Ministere du travail, the Ligue contra le Cancer and the Caisse Nationale de l'Assurance Maladie. Authors affiliated with Universite de Lille; CHU; Hopital Calmette; Comite pour le developpement de la Medecine du Travail; Direction Regionale du Travail et de l'Emploi; Caisse Regionale d'Assurance Maladie.)

- 470. Norbäck D, Björnsson E, Janson C, Widström J, Boman G. 1995. Asthmatic symptoms and volatile organic compounds, formaldehyde, and carbon dioxide in dwellings. *Occup Environ Med* 52: 388-395. (Supported by the Swedish Association Against Asthma and Allergy, Swedish Medical Research Council, Swedish Society of Medicine, Swedish Heart and Lung Foundation, Bror Hjerpstedts Foundation, Pharmacia and the County Council of Uppsala. Authors affiliated with Uppsala University, Sweden.)
- 471. Nordman H, Keskinen H, Tuppurainen M. 1985. Formaldehyde asthma--rare or overlooked? *J Allergy Clin Immunol* 75(1 Pt 1): 91-99. (Support not reported. Authors affiliated with Institute of Occupational Health, Finland.)
- 472. Norsted SW, Kozinetz CA, Annegers JF. 1985. Formaldehyde complaint investigations in mobile homes by the Texas Department of Health. *Environ Res* 37: 93-100 (as cited in ATSDR 1999).
- 473. Nousiainen P, Lindqvist J. 1979. *Chemical hazards in the textile industry. Air contaminents. (Tiendonanto 16), Tampere, Valtion teknillinen tutkimsukeskus* (as cited in IARC 2006).
- 474. NRC. 2009. *National Response Center*. United States Coast Guard. <u>http://www.nrc.uscg.mil</u> and <u>http://www.nrc.uscg.mil/nrcback.html</u>.
- 475. NTP. 1981. Report on Carcinogens, 2nd Edition, Research Triangle Park, NC: U.S. Department of Health and Human Services, Public Health Service, National Toxicology Program.
- 476. NTP. 1990. Toxicology and Carcinogenesis Studies of Benzaldehyde (Cas No. 100-52-7) in F344/N Rats and B6C3F<sub>1</sub> Mice (Gavage Studies). Technical Report Series No. 378. Research Triangle Park, NC: National Toxicology Program. 195 pp.
- 477. NTP. 1999. Toxicology and Carcinogenesis Studies of Glutaraldehyde (Cas No. 111-30-8) in F344/N Rats And B6C3F<sub>1</sub> Mice (Inhalation Studies). NTP TR 490, NIH Publication No. 99-3980. Research Triangle Park, NC: National Toxicology Program. 236 pp.
- 478. NTP. 2005a. *Report on Carcinogens, 11th Edition*, Research Triangle Park, NC: U.S. Department of Health and Human Services, Public Health Service, National Toxicology Program.
- 479. O'Connor PM, Fox BW. 1987. Comparative studies of DNA cross-linking reactions following methylene dimethanesulphonate and its hydrolytic product, formaldehyde. *Cancer Chemother Pharmacol* 19(1): 11-15. (Supported by the Cancer Research Campaign. Authors affiliated with Paterson Laboratories, UK.)
- 480. O'Neil MJ, Heckelman PE, Koch CB, Roman KJ, Kenny CM, D'Arecca MR, eds. 2006. *The Merck Index: An Encyclopedia of Chemicals, Drugs and*

*Biologicals*. 14th ed. Whitehouse Station, NJ: Merck & Co., Inc. p. 726, 1211, 1672.

- 481. O'Quinn SE, Kennedy CB. 1965. Contact dermatitis due to formaldehyde in clothing textiles. *Jama* 194(6): 593-6. (Support not reported. Authors affiliated with Louisiana University State School of Medicine, LA.)
- 482. Obe G, Beek B. 1979. Mutagenic activity of aldehydes. *Drug Alcohol Depend* 4(1-2): 91-94. (Support not reported. Authors affiliated with Freie Universitat Berlin, Germany.)
- 483. Odeigah PG. 1997. Sperm head abnormalities and dominant lethal effects of formaldehyde in albino rats. *Mutat Res* 389(2-3): 141-148. (Supported by the University of Lagos. Author affiliated with University of Lagos, Nigeria.)
- 484. Offerman FJ, Hollowell CD, Znazaroff WW, Roseme GD, Rizzuto JR. 1982. Low infiltration housing in Rochester, New York. A atudy of air-exchange rates and indoor air quality. *Environ Int* 8: 435-445 (as cited in WHO 1989).
- 485. Ohmichi K, Komiyama M, Matsuno Y, Sawabe Y, Miyaso H, Fukata H, Ohmichi M, Kadota T, Nomura F, Mori C. 2006b. Relationship between exposure to formaldehyde and immunoglobulin E (IgE) production during the gross anatomy laboratory (vol 52, pg 642, 2006). *Journal Of Health Science* 52(5): 642-647. (Support not reported. Authors affiliated with Chiba University Hospital, Japan; Chiba City Social Welfare Administrative Office, Japan.)
- 486. Ohmichi K, Matsuno Y, Miyaso H, Yamamoto H, Toriuchi M, Shimane M, Mori C. 2007. Pilot study of a dissection table for gross anatomy laboratory equipped with a photocatalytic device that decomposes formaldehyde. *J Occup Health* 49(6): 499-503. (Support not reported. Authors affiliated with Chiba University, Japan; Natsume Seisakusho Co. Ltd., Japan; Nisshin EM Co. Ltd., Japan.)
- 487. Ojajärvi IA, Partanen TJ, Ahlbom A, Boffetta P, Hakulinen T, Jourenkova N, Kauppinen TP, Kogevinas M, Porta M, Vainio HU, Weiderpass E, Wesseling CH. 2000. Occupational exposures and pancreatic cancer: a meta-analysis. *Occup Environ Med* 57(5): 316-324. (Supported by the Finnish Work Environment Fund. Authors affiliated with Finnish Institute of Occupational Health, Finland; Karolinska Institutet, Sweden; IARC, France; Finnish Cancer Registry, Finland; Institut National de la Santé et de la Reserche Médicale, France; Institut Municipal d'Investigacio Medica, Spain; Universidad Nacional, Costa Rica.)
- 488. Olin KL, Cherr GN, Rifkin E, Keen CL. 1996. The effects of some redox-active metals and reactive aldehydes on DNA-protein cross-links in vitro. *Toxicology* 110(1-3): 1-8. (Support not reported. Authors affiliated with University of California - Davis, CA.)

- 489. Oliva-Teles MT, Paiga P, Delerue-Matos CM, Alvim-Ferraz MCM. 2009. Evaluation of Formaldehyde in Foundry Waste Sands Using Liquid Chromatography. *Analytical Letters* 42(3): 492-504. (Support not reported. Authors affiliated with REQUIMTE, Portugal; LEPAE, Portugal.)
- 490. Olsen JH, Jensen SP, Hink M, Faurbo K, Breum NO, Jensen OM. 1984. Occupational formaldehyde exposure and increased nasal cancer risk in man. *Int J Cancer* 34(5): 639-644. (Support not reported. Authors affiliated with Danish Cancer Registry, Denmark; Labour Inspection Service, Denmark; Danish National Institute of Occupational Health.)
- 491. Olsen JH, Asnaes S. 1986. Formaldehyde and the risk of squamous cell carcinoma of the sinonasal cavities. *Br J Ind Med* 43(11): 769-774. (Support not reported. Authors affiliated with Danish Cancer Registry, Denmark; Institute of Forensic Medicine, Denmark.)
- 492. Orsiere T, Sari-Minodier I, Iarmarcovai G, Botta A. 2006. Genotoxic risk assessment of pathology and anatomy laboratory workers exposed to formaldehyde by use of personal air sampling and analysis of DNA damage in peripheral lymphocytes. *Mutat Res* 605(1-2): 30-41. (Supported by the Direction Regionale du Travail, de l'Emploi et de la Formation Professionnelle of the Provence-Alpes-Cote d'Azur region and the Fondation Philippe Daher. Authors affiliated with Universite de la Mediterranee, France; Hopitaux de Marseille, France.)
- 493. OSHA. 1990. Enforcement Procedure for Occupational Exposure to Formaldehyde U.S. Department of Labor. <u>http://www.osha.gov/pls/oshaweb/owadisp.show\_document?p\_table=DIRECTI\_VES&p\_id=1566</u>. Accessed on 5/29/07.
- 494. Ott MG, Teta MJ, Greenberg HL. 1989. Lymphatic and hematopoietic tissue cancer in a chemical manufacturing environment. *Am J Ind Med* 16(6): 631-643. (Support not reported. Authors affiliated with Arthur D. Little, Inc., MA; Union Carbide Corporation, CT; ARCO, CA.)
- 495. Overton JH, Kimbell JS, Miller FJ. 2001. Dosimetry modeling of inhaled formaldehyde: the human respiratory tract. *Toxicol Sci* 64(1): 122-134. (Support not reported. Authors affiliated with U.S. EPA; Chemical Industry Institute of Toxicology, NC.)
- 496. Ovrebo S, Haugen A, Skaug V. 2002. Biotransformation of formaldehyde in cultured human bronchus. *Environ Res* 89(1): 38-42. (Supported by the National Institute of Occupational Health, Norway. Authors affiliated with National Institute of Occupational Health, Norway.)
- 497. Owen BA, Dudney CS, Tan EL, Easterly CE. 1990. Formaldehyde in drinking water: comparative hazard evaluation and an approach to regulation. *Regul*

*Toxicol Pharmacol* 11(3): 220-236. (Supported by the U.S. Navy. Authors affiliated with Oak Ridge National Laboratory, TN.)

- 498. Ozen OA, Yaman M, Sarsilmaz M, Songur A, Kus I. 2002. Testicular zinc, copper and iron concentrations in male rats exposed to subacute and subchronic formaldehyde gas inhalation. *J Trace Elem Med Biol* 16(2): 119-122. (Support not reported. Authors affiliated with Firat University, Turkey.)
- 499. Ozen OA, Akpolat N, Songur A, Kus I, Zararsiz I, Ozacmak VH, Sarsilmaz M. 2005. Effect of formaldehyde inhalation on Hsp70 in seminiferous tubules of rat testes: an immunohistochemical study. *Toxicol Ind Health* 21(10): 249-254. (Support not reported. Authors affiliated with Afyon Kocatepe University, Turkey; Firat University, Turkey; Z. Karaelmas University, Turkey.)
- 500. Ozen OA, Kus MA, Kus I, Alkoc OA, Songur A. 2008. Protective effects of melatonin against formaldehyde-induced oxidative damage and apoptosis in rat testes: an immunohistochemical and biochemical study. *Syst Biol Reprod Med* 54(4-5): 169-176. (Support not reported. Authors affiliated with Afyon Kocatepe University, Turkey; Firat University, Turkey; Namik Kemal University, Turkey.)
- 501. Pala M, Ugolini D, Ceppi M, Rizzo F, Maiorana L, Bolognesi C, Schiliro T, Gilli G, Bigatti P, Bono R, Vecchio D. 2008. Occupational exposure to formaldehyde and biological monitoring of Research Institute workers. *Cancer Detect Prev* 32(2): 121-6. (Support not reported. Authors affiliated with National Cancer Research Institute, Italy; University of Turin, Italy; University of Genoa, Italy.)
- 502. Pan S, Lin S, Luo SC. 2000. The combined effects of formaldehyde and noise on neurobehavioral function. *J Labour Med* 17(3): 159-161 (as cited in Tang *et al.* 2009).
- 503. Pan S, Guo W, Li Y, Wang X, Cheng H, Li W. 2001. Formaldehyde's effect on superoxide dimutase and neurobehavorial functions. *Chin Occup Med* 2001(28): 1 (as cited in Tang *et al.* 2009).
- 504. Pang XB, Mu YJ. 2007. Characteristics of carbonyl compounds in public vehicles of Beijing city: Concentrations, sources, and personal exposures. *Atmospheric Environment* 41(9): 1819-1824. (Supported by the Chinese National Natural Science Foundation and the National Basic Research Program of China. Authors affiliated with Chinese Academy of Sciences, China.)
- 505. Parfett CL. 2003. Combined effects of tumor promoters and serum on proliferin mRNA induction: a biomarker sensitive to saccharin, 2,3,7,8-TCDD, and other compounds at minimal concentrations promoting C3H/10T1/2 cell transformation. *J Toxicol Environ Health A* 66(20): 1943-66. (Supported by the Safe Environments Programme of the Healthy Environments and Consumer Safety Branch of Health Canada. Authors affiiated with Health Canada. )

- 506. Park JS, Ikeda K. 2006. Variations of formaldehyde and VOC levels during 3 years in new and older homes. *Indoor Air* 16(2): 129-35. (Supported by the ERC program of MOST and Hanyang University. Authors affiliated with Hanyang University, Korea; National Institute of Public Health, Japan.)
- 507. Partanen T, Kauppinen T, Nurminen M, Nickels J, Hernberg S, Hakulinen T, Pukkala E, Savonen E. 1985. Formaldehyde exposure and respiratory and related cancers. A case-referent study among Finnish woodworkers. *Scand J Work Environ Health* 11(6): 409-415. (Supported by the Finnish Work Environment Fund. Authors affiliated with Institute of Occupational Health, Finland; Finnish Cancer Registry, Finland.)
- 508. Partanen T, Kauppinen T, Hernberg S, Nickels J, Luukkonen R, Hakulinen T, Pukkala E. 1990. Formaldehyde exposure and respiratory cancer among woodworkers--an update. *Scand J Work Environ Health* 16(6): 394-400. (Supported by the Academy of Finland. Authors affiliated with Institute of Occupational Health, Finland; Finnish Cancer Registry, Finland.)
- 509. Partanen T, Kauppinen T, Luukkonen R, Hakulinen T, Pukkala E. 1993. Malignant lymphomas and leukemias, and exposures in the wood industry: an industry-based case-referent study. *Int Arch Occup Environ Health* 64(8): 593-596. (Supported by the Academy of Finland and the Finnish Work Environment Fund. Authors affiliated with Institute of Occupational Health, Finland; Finnish Cancer Registry, Finland.)
- 510. Patel KG, Bhatt HV, Choudhury AR. 2003. Alteration in thyroid after formaldehyde (HCHO) treatment in rats. *Ind Health* 41(3): 295-7. (Support not reported. Authors affiliated with National Institute of Occupational Health, India; Regional Occupational Health Centre, India.)
- 511. Paustenbach D, Alarie Y, Kulle T, Schachter N, Smith R, Swenberg J, Witschi H, Horowitz SB. 1997. A recommended occupational exposure limit for formaldehyde based on irritation. *J Toxicol Environ Health* 50(3): 217-263. (Support not reported. Authors affiliated with McLaren/Hart, CA; University of Pittsburgh, PA; Environmental Health Sciences, MD; Mount Sinai Medical Center, NY; University of Michigan, MI; University of North Carolina, NC; University of California at Davis, CA; ChemRisk, CA.)
- 512. Pazdrak K, Gorski P, Krakowiak A, Ruta U. 1993. Changes in nasal lavage fluid due to formaldehyde inhalation. *Int Arch Occup Environ Health* 64(7): 515-9 (as cited in IARC 2006).
- 513. Peng JS, Sun F, Chen WG, Wei F, Chang W. 2003. Study of the formaldehyde pollution in the human anatomy laboratory of a medical college. *J Environ Health* 20(5): 294-295 (as cited in Tang *et al.* 2009).
- 514. Pesch B, Pierl CB, Gebel M, Gross I, Becker D, Johnen G, Rihs HP, Donhuijsen K, Lepentsiotis V, Meier M, Schulze J, Bruning T. 2008. Occupational risks for

adenocarcinoma of the nasal cavity and paranasal sinuses in the German wood industry. *Occup Environ Med* 65(3): 191-196. (Support not reported. Authors affiliated with BGFA-Forschungsinstitut fur Arbeitsmedizin der Deutschen Gesetzlichen Unfallversicherung, Germany; Stadtisches Klinikum Braunschweig, Germany; Holz-Berufsgenossenschaft, Germany.)

- 515. Pickrell JA, Mokler B, Griffis LC, Hobbs C. 1983. Formaldehyde release rate coefficients from selected consumer products. *Environ Sci Technol* 17: 753-757. (Support not reported. Authors affiliated with Lovelace Biomedical and Environmental Research Institute, NM; Chevron Environmental Health Center, Inc., CA; Small Particle Technology, NM; U.S. Consumer Product Safety Commission, Washington, D.C.)
- 516. Pickrell JA, Griffis LC, Mokler B, Kanapilly GM, Hobbs CH. 1984. Formaldehyde release from selected consumer products: influence of chamber loading, multiple products, relative humidity, and temperature. *Environ Sci Technol* 18: 682-686. (Support not reported. Authors affiliated with Lovelace Biomedical and Environmental Research Institute, NM; Chevron Environmental Health Center, Inc., CA; Small Particle Technology, NM.)
- 517. Pilidis GA, Karakitsios SP, Kassomenos PA, Kazos EA, Stalikas CD. 2009. Measurements of benzene and formaldehyde in a medium sized urban environment. Indoor/outdoor health risk implications on special population groups. *Environ Monit Assess* 150(1-4): 285-94. (Supported by the European Union. Authors affiliated with University of Ioannina, Greece.)
- 518. Pinkerton LE, Hein MJ, Stayner LT. 2004. Mortality among a cohort of garment workers exposed to formaldehyde: an update. *Occup Environ Med* 61(3): 193-200. (Support not reported. Authors affiliated with NIOSH.)
- 519. Pitten FA, Kramer A, Herrmann K, Bremer J, Koch S. 2000. Formaldehyde neurotoxicity in animal experiments. *Pathol Res Pract* 196(3): 193-8. (Support not reported. Authors affiliated with Ernst-Moritz-Arndt-University, Germany; HUMAINE Klinikum of Bad Saarow, Germany.)
- 520. PolymerProcessing. 2009. *Poly(oxymethylene)*. PolymerProcessing.com. http://www.polymerprocessing.com/polymers/POM.html. Accessed on 5/28/09.
- 521. Popa V, Teculescu D, Stanescu D, Gavrilescu N. 1969. Bronchial asthma and asthmatic bronchitis determined by simple chemicals. *Dis Chest* 56(5): 395-402 (as cited in IARC 2006).
- 522. Porter JA. 1975. Letter: Acute respiratory distress following formalin inhalation. *Lancet* 2(7935): 603-4. (Support not reported. Authors affiliated with Medical College of Wisconsin, WI.)
- 523. Pottern LM, Heineman EF, Olsen JH, Raffn E, Blair A. 1992. Multiple myeloma among Danish women: employment history and workplace exposures. *Cancer*

*Causes Control* 3(5): 427-32. (Support not reported. Authors affiliated with NCI, MD; Danish Cancer Registry, Denmark; Arbedjdsmedicinsk Klinsk, Denmark.)

- 524. Prades JM, Alaani A, Mosnier JF, Dumollard JM, Martin C. 2002. Granulocytic sarcoma of the nasal cavity. *Rhinology* 40(3): 159-161. (Support not reported. Authors affiliated with Saint-Etienne University Hospital Centre, France.)
- 525. Pratt GC, Palmer K, Wu CY, Oliaei F, Hollerbach C, Fenske MJ. 2000. An assessment of air toxics in Minnesota. *Environ Health Perspect* 108(9): 815-25 (as cited in IARC 2006).
- 526. Prescher KE. 1984. Indoor formaldehyde determination with passive collectors. *Schriftenr Verh Wabolu* 59(33-40) (as cited in WHO 1989).
- 527. Priha E, Riipinen H, Korhonen K. 1986. Exposure to formaldehyde and solvents in Finnish furniture factories in 1975-1984. *Ann Occup Hyg* 30(3): 289-94 (as cited in IARC 2006).
- 528. Priha E, Vuorinen R, Schimberg R. 1988. *Textile Finishing Agents (Tyoolot 65)*. Helsinki: Finnish Institute of Occupational Health (as cited in IARC 2006).
- 529. Pyatt D, Natelson E, Golden R. 2008. Is inhalation exposure to formaldehyde a biologically plausible cause of lymphohematopoietic malignancies? *Regul Toxicol Pharmacol* 51(1): 119-33. (Supported by the Formaldehyde Council, Inc. Authors affiliated with Summit Toxicology, L.L.P., CO; ToxLogic, L.L.C., MD; University of Colorado, CO; Cornell University, TX.)
- 530. Qian RJ, Zhang PH, Duang TL, Yao NL. 1988. Investigation on occupational hazards of formaldehyde exposure. *Ind Hyg Occup Dis* 14(2): 101 (as cited in Tang *et al* 2009).
- 531. Quievryn G, Zhitkovich A. 2000. Loss of DNA-protein crosslinks from formaldehyde-exposed cells occurs through spontaneous hydrolysis and an active repair process linked to proteosome function. *Carcinogenesis* 21(8): 1573-1580. (Supported by Brown University. Authors affiliated with Brown University, RI.)
- 532. Quinn MM, Smith TJ, Youk AO, Marsh GM, Stone RA, Buchanich JM, Gula MJ. 2001. Historical cohort study of US man-made vitreous fiber production workers: VIII. Exposure-specific job analysis. *J Occup Environ Med* 43(9): 824-834. (Supported by the North American Insulation Manufacturers Association and University of Massachusetts Lowell. Authors affiliated with University of Massachusetts Lowell, MA; Harvard School of Public Health, MA; University of Pittsburgh, PA.)

- 533. Ragan DL, Boreiko CJ. 1981. Initiation of C3H/10T1/2 cell transformation by formaldehyde. *Cancer Lett* 13(4): 325-331. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)
- 534. Ravis SM, Shaffer MP, Shaffer CL, Dehkhaghani S, Belsito DV. 2003. Glutaraldehyde-induced and formaldehyde-induced allergic contact dermatitis among dental hygienists and assistants. *J Am Dent Assoc* 134(8): 1072-8. (Supported by the American Dental Association Health Foundation. Authors affiliated with University of Miami, FL; University of Iowa, IA; University of Kansas, MO.)
- 535. Recio L, Sisk S, Pluta L, Bermudez E, Gross EA, Chen Z, Morgan K, Walker C. 1992. *p53* mutations in formaldehyde-induced nasal squamous cell carcinomas in rats. *Cancer Res* 52(21): 6113-6116. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)
- 536. Recio L. 1997. Oncogene and tumor suppressor gene alterations in nasal tumors. *Mutat Res* 380(1-2): 27-31. (Support not reported. Author affiliated with Chemical Industry Institute of Toxicology, NC.)
- 537. Reh CM, Letts D, Deitchman S. 1994. National Park Service, Yosemite National Park, CA. Health Hazard Evaluation Report, HETA 90-0365-2415. Cincinnati, OH: National Institute for Occupational Safety and Health (as cited in IARC 2006).
- 538. Reiss R, Ryan PB, Tibbetts SJ, Koutrakis P. 1995. Measurement of organic acids, aldehydes, and ketones in residential environments and their relation to ozone. *J Air Waste Manage Assoc* 45: 811-822. (Supported by the Center for Indoor Air Research and NIH. Authors affiliated with Harvard University, MA.)
- 539. Rephaeli A, Waks-Yona S, Nudelman A, Tarasenko I, Tarasenko N, Phillips DR, Cutts SM, Kessler-Icekson G. 2007. Anticancer prodrugs of butyric acid and formaldehyde protect against doxorubicin-induced cardiotoxicity. *Br J Cancer* 96(11): 1667-74. (Supported by the Israel Ministry of Science, Art and Sports, Israel Science Foundation; Israel Cancer Research Fund, Israel Cancer Association, Ministry of Absorption in Science, and the Australian Research Council. Authors affiliated with Tel-Aviv University, Israel; Bar-Ilan University, Israel; La Trobe University, Australia.)
- 540. Reynolds SJ, Black DW, Borin SS, Breuer G, Burmeister LF, Fuortes LJ, Smith TF, Stein MA, Subramanian P, Thorne PS, Whitten P. 2001. Indoor environmental quality in six commercial office buildings in the midwest United States. *Appl Occup Environ Hyg* 16(11): 1065-1077. (Supported by the U.S. EPA, NIH and the University of Iowa. Authors affiliated with College of Public Health University of Iowa, IA.)
- 541. Riala RE, Riihimaki HA. 1991. Solvent and formaldehyde exposure in parquet and carpet work. *Appl Occup Environ Hyg* 6: 301-308 (as cited in IARC 2006).

- 542. Ridpath JR, Nakamura A, Tano K, Luke AM, Sonoda E, Arakawa H, Buerstedde JM, Gillespie DA, Sale JE, Yamazoe M, Bishop DK, Takata M, Takeda S, Watanabe M, Swenberg JA, Nakamura J. 2007. Cells deficient in the FANC/BRCA pathway are hypersensitive to plasma levels of formaldehyde. *Cancer Res* 67(23): 11117-11122. (Supported by NIEHS and University of North Carolina. Authors affiliated with University of North Carolina at Chapel Hill, NC; University of Virginia, VA; Kyoto University, Japan; Department of Radiation Genetics Graduate School of Medicine, Japan; GSF-National Research Center for Environment and Health, Germany; Beatson Institute for Cancer Research, UK; Medical Research Council Laboratory of Molecular Biology, UK; University of Chicago, IL; Hiroshima University, Japan.)
- 543. Rietbrock N. 1965. Formaldehyde oxidation in the rat. *Naunyn-Schmiedeberg's Arch Exp Pathol Pharmakol* 251: 189-190 (as cited in IARC 2006).
- 544. Ritchie IM, Lehnen RG. 1987. Formaldehyde-related health complaints of residents living in mobile and conventional homes. *Am J Public Health* 77(3): 323-8. (Support not reported. Authors affiliated with Indiana University, IN.)
- 545. Robins JM, Pambrun M, Chute C, Blevins D. 1988. Estimating the effect of formaldehyde exposure on lung cancer and non-malignant respiratory disease (NMRD) mortality using a new method to control for the healthy worker survivor effect. In *Progress in Occupational Epidemiology*. Hogstedt C, Reuterwall C, eds. Amsterdam: Elsevier Science. p. 75-78. (Support not reported. Authors affiliated with Harvard School of Public Health, MA.)
- 546. Rosen G, Bergstrom B, Ekholm U. 1984. Occupational exposure to formaldehyde in Sweden. *Arbete Halsa* 50: 16-21 (as cited in IARC 2006).
- 547. Ross WE, Shipley N. 1980. Relationship between DNA damage and survival in formaldehyde-treated mouse cells. *Mutat Res* 79(3): 277-283. (Support not reported. Authors affiliated with University of Florida College of Medicine, FL.)
- 548. Ross WE, McMillan DR, Ross CF. 1981. Comparison of DNA damage by methylmelamines and formaldehyde. *J Natl Cancer Inst* 67(1): 217-221. (Supported by the National Cancer Institute. Authors affiliated with University of Florida, FL.)
- 549. Roush GC, Walrath J, Stayner LT, Kaplan SA, Flannery JT, Blair A. 1987. Nasopharyngeal cancer, sinonasal cancer, and occupations related to formaldehyde: a case-control study. *J Natl Cancer Inst* 79(6): 1221-1224. (Supported by NCI. Authors affiliated with Yale University School of Medicine, CT; E.I. du Pont de Nemours & Company, Inc., DE; NIOSH; NCI; Connecticut Tumor Registry, CT.)
- 550. Rusch GM, Clary JJ, Rinehart WE, Bolte HF. 1983. A 26-week inhalation toxicity study with formaldehyde in the monkey, rat, and hamster. *Toxicol Appl*

*Pharmacol* 68(3): 329-343. (Supported by the Formaldehyde Institute. Authors affiliated with Bio/dynamics Inc., NJ; Allied Corp., NJ.)

- 551. Saito Y, Nishio K, Yoshida Y, Niki E. 2005. Cytotoxic effect of formaldehyde with free radicals via increment of cellular reactive oxygen species. *Toxicology* 210(2-3): 235-245. (Support not reported. Authors affiliated with National Institute of Advanced Industrial Science and Technology, Japan.)
- 552. Saladino AJ, Willey JC, Lechner JF, Grafstrom RC, LaVeck M, Harris CC. 1985. Effects of formaldehyde, acetaldehyde, benzoyl peroxide, and hydrogen peroxide on cultured normal human bronchial epithelial cells. *Cancer Res* 45(6): 2522-2526. (Support not reported. Authors affiliated with NCI, MD; Baltimore Veterans Administration Medical Center, MD; Heart, Lung and Blood Institute, MD; CDC, GA; Karolinska Institutet, Sweden.)
- 553. Salas LJ, Singh HB. 1986. Measurements of formaldehyde and acetaldehyde in teh urban ambient air. *Atmos Environ* 20: 1301-1304 (as cited in ATSDR 1999).
- 554. Salonen H, Pasanen AL, Lappalainen S, Riuttala H, Tuomi T, Pasanen P, Back B, Reijula K. 2009. Volatile organic compounds and formaldehyde as explaining factors for sensory irritation in office environments. *J Occup Environ Hyg* 6(4): 239-47. (Support not reported. Authors affiliated with Finnish Institute of Occupational Health, Finland; University of Kuopio, Finland.)
- 555. Sanford DM, Becker GD. 1967. Acute leukemia presenting as nasal obstruction. *Arch Otolaryngol* 85(1): 102-104. (Support not reported. Authors affiliated with Gorgas Hospital; Womack Army Hospital, NC.)
- 556. Sarsilmaz M, Kaplan S, Songur A, Colakoglu S, Aslan H, Tunc AT, Ozen OA, Turgut M, Bas O. 2007. Effects of postnatal formaldehyde exposure on pyramidal cell number, volume of cell layer in hippocampus and hemisphere in the rat: a stereological study. *Brain Res* 1145: 157-67. (Support not reported. Authors affiliated with Firat University School of Medicine, Turkey; Ondokuz Mayıs University School of Medicine, Turkey; Afyon Kocatepe University School of Medicine, Turkey; Gaziosmanpasa University School of Medicine, Turkey; Adnan Menderes University School of Medicine, Turkey.)
- 557. Sasaki Y, Ohtani T, Ito Y, Mizuashi M, Nakagawa S, Furukawa T, Horii A, Aiba S. 2009. Molecular events in human T cells treated with diesel exhaust particles or formaldehyde that underlie their diminished interferon-gamma and interleukin-10 production. *Int Arch Allergy Immunol* 148(3): 239-250. (Supported by Tohoku University, Japan Society for the Promotion of Science, and the New Energy and Industrial Technology Development Organization. Authors affiliated with Tohoku University Graduate School of Medicine, Japan.)
- 558. Sass-Kortsak AM, Holness DL, Pilger CW, Nethercott JR. 1986. Wood dust and formaldehyde exposures in the cabinet-making industry. *Am Ind Hyg Assoc J* 47(12): 747-53 (as cited in IARC 2006).

- 559. Sauder LR, Chatham MD, Green DJ, Kulle TJ. 1986. Acute pulmonary response to formaldehyde exposure in healthy nonsmokers. *J Occup Med* 28(6): 420-4 (as cited in IARC 2006).
- 560. Sauder LR, Green DJ, Chatham MD, Kulle TJ. 1987. Acute pulmonary response of asthmatics to 3.0 ppm formaldehyde. *Toxicol Ind Health* 3(4): 569-78 (as cited in IARC 2006).
- 561. Saurel-Cubizolles MJ, Hays M, Estryn-Behar M. 1994. Work in operating rooms and pregnancy outcome among nurses. *Int Arch Occup Environ Health* 66(4): 235-241. (Supported by the Direction regionale du travail et de l'emploi d'lle-de-France. Authors affiliated with INSERM, France; Hopital Henri Mondor, France.)
- 562. Sawant AA, Shah SD, Zhu XN, Miller JW, Cocker DR. 2007. Real-world emissions of carbonyl compounds from in-use heavy-duty diesel trucks and diesel Back-Up Generators (BUGS). *Atmospheric Environment* 41(21): 4535-4547. (Supported by the Strategic Environmental Research Development Program, Cummins, and the California Energy Commission. Authors affiliated with University of California, CA; Johnson Matthey, Inc., PA; Ford Motor Company, MI.)
- 563. Sax SN, Bennett DH, Chillrud SN, Kinney PL, Spengler JD. 2004. Differences in source emission rates of volatile organic compounds in inner-city residences of New York City and Los Angeles. *J Expo Anal Environ Epidemiol* 14(Suppl 1): S95-109. (Supported by the Mickey Leland National Urban Air Toxics Research Center, NIEHS, the Center for Environmental Health in Northern Manhattan, the Columbia Center for Children's Environmental Health and the Akira Yamaguchi endowment fund. Authors affiliated with University of Harvard School of Public Health, MA; Columbia University, NY.)
- 564. Schachter EN, Witek TJ, Jr., Brody DJ, Tosun T, Beck GJ, Leaderer BP. 1987. A study of respiratory effects from exposure to 2.0 ppm formaldehyde in occupationally exposed workers. *Environ Res* 44(2): 188-205 (as cited in IARC 2006).
- 565. Schafer D, Brommer C, Riechelmann H, Mann JW. 1999. In vivo and in vitro effect of ozone and formaldehyde on human nasal mucociliary transport system. *Rhinology* 37(2): 56-60.
- 566. Schlink K, Janßen K, Nitzsche S, Gebhard S, Hengstler JG, Klein S, Oesch F. 1999. Activity of O<sup>6</sup>-methylguanine DNA methyltransferase in mononuclear blood cells of formaldehyde-exposed medical students. *Arch Toxicol* 73(1): 15-21. (Supported by the Bundesministerium fur Forschung und Technologie. Authors affiliated with University of Mainz, Germany.)
- 567. Schmid E, Göggelmann W, Bauchinger M. 1986. Formaldehyde-induced cytotoxic, genotoxic and mutagenic response in human lymphocytes and

*Salmonella typhimurium. Mutagenesis* 1(6): 427-431. (Support not reported. Authors affiliated with Institut fur Strahlenbiologie, Germany; Institut fur Toxikologie, Germany.)

- 568. Schmid O, Speit G. 2007. Genotoxic effects induced by formaldehyde in human blood and implications for the interpretation of biomonitoring studies. *Mutagenesis* 22(1): 69-74. (Supported by the European Chemical Industry Council. Authors affiliated with Universitat Ulm, Germany.)
- 569. Schulam P, Newbold R, Hull LA. 1985. Urban and rural ambient air aldehyde levels in Schenectady, New York and on Whiteface Mountain, New York. *Atmos Environ* 19: 623-626 (as cited in ATSDR 1999).
- 570. Schupp T, Bolt HM, Hengstler JG. 2005. Maximum exposure levels for xylene, formaldehyde and acetaldehyde in cars. *Toxicology* 206(3): 461-470. (Support not reported. Authors affiliated with Elastogran, Germany; Institut fur Arbeitsphysiologie an der Universitat Dortmund, Germany; Institute of Legal Medicine and Rudolf-Broehm Institute of Pharmacology and Toxicology, Germany; Universitat Mainz, Germany.)
- 571. ScienceLab. 2009a. *Material Safety Data Sheet: s-Trioxane*. ScienceLab.com. http://www.sciencelab.com.
- 572. ScienceLab. 2009b. *Material Safety Data Sheet: Paraformaldehyde*. ScienceLab.com. <u>http://www.sciencelab.com</u>.
- 573. Scully RE, Mark EJ, McNeely WF, McNeely BU. 1990. Case records of the Massachusetts General Hospital. *New Eng J Med* 323(24): 1689-1691. (Support and author affiliations not reported.)
- 574. Seiber JN. 1996. Toxic air contaminants in urban atmospheres: experience in California. *Atmos Environ* 30: 751-756. (Support not reported. Authors affiliated with University of Nevada, NV.)
- 575. Sellakumar AR, Snyder CA, Solomon JJ, Albert RE. 1985. Carcinogenicity of formaldehyde and hydrogen chloride in rats. *Toxicol Appl Pharmacol* 81(3 Pt 1): 401-406. (Supported by NCI. Authors affiliated with New York University Medical Center, NY.)
- 576. Sexton K, Liu KS, Petreas MX. 1986. Formaldehyde concentrations inside private residences: A mail-out approach to indoor air monitoring. *J Air Pollut Control Assoc* 36: 698-704 (as cited in IARC 2006).
- 577. Sexton K, Petreas MX, Liu KS. 1989. Formaldehyde exposures inside mobile homes. *Environ Sci Technol* 23: 985-988 (as cited in ATSDR 1999).

- 578. Shah JJ, Singh HB. 1988. Distribution of volatile organic chemicals in outdoor and indoor air. A national VOCs data base. *Environ Sci Technol* 22: 1381-1388 (as cited in IARC 2006).
- 579. Shaham J, Bomstein Y, Meltzer A, Kaufman Z, Palma E, Ribak J. 1996a. DNA--protein crosslinks, a biomarker of exposure to formaldehyde--*in vitro* and *in vivo* studies. *Carcinogenesis* 17(1): 121-125. (Supported by the Committe for Preventive Action and Research in Occupational Health, Ministry of Labor and Social Welfare. Authors affiliated with Occupational Health and Rehabilitation Institute, Israel; Tel-Aviv University, Israel; Occupational Health Clinic, Israel.)
- 580. Shaham J, Bomstein Y, Meltzer A, Ribak J. 1996b. Response. *Carcinogenesis* 17: 2098-2101 (as cited in IARC 2006).
- 581. Shaham J, Bomstein Y, Melzer A, Ribak J. 1997. DNA-Protein Crosslinks and Sister Chromatid Exchanges as Biomarkers of Exposure to Formaldehyde. Int J Occup Environ Health 3(2): 95-104. (Supported by the Committee for Preventive Action and Research in Occupational Health, Ministry of Labor and Social Welfare, Israel. Authors affiliated with Occupational Health and Rehabilitation Institute, Israel; Tel-Aviv University, Israel.)
- 582. Shaham J, Gurvich R, Kaufman Z. 2002. Sister chromatid exchange in pathology staff occupationally exposed to formaldehyde. *Mutat Res* 514(1-2): 115-123. (Supported by the Committee for Preventive Action and Research in Occupational Health, Ministry of Labor and Social Welfare. Authors affiliated with Tel-Aviv University, Israel.)
- 583. Shaham J, Bomstein Y, Gurvich R, Rashkovsky M, Kaufman Z. 2003. DNAprotein crosslinks and p53 protein expression in relation to occupational exposure to formaldehyde. *Occup Environ Med* 60(6): 403-409. (Supported by the Preventive Activities Program of the Ministry of Labor and Social Welfare, Israel. Authors affiliated with Occupational Health and Rehabilitation Institute, Israel; Tel-Aviv University, Israel.)
- 584. Shellow H, Altman AT. 1966. Dermatitis from formaldehyde resin textiles. *Arch Dermatol* 94(6): 799-801. (Support and affiliations not reported.)
- 585. Shi J, Zhu SX, Tong ZM, Sun DX, Yang H, Jiang RM, Etal. 2006. Epidemiological study of health effect for occupational exposure staffs to formaldehyde. *Chin Occup Med* 33(3): 237-239 (as cited in Tang *et al.* 2009).
- 586. Siemiatycki J, Dewar R, Nadon L, Gérin M. 1994. Occupational risk factors for bladder cancer: results from a case-control study in Montreal, Quebec, Canada. *Am J Epidemiol* 140(12): 1061-1080. (Supported by the Institut de recherche en sante et en securite du Travail du Quebec, the Fonds de recherche en sante du Quebec, the National Health and Research Development Program, and the National Cancer Institute of Canada. Authors affiliated with Institut Armand-Frappier, Canada; McGill University, Canada; Universite de Montreal, Canada.)

- 587. Singh HB, Salas LJ, Stiles RE. 1982. Distribution of selected gaseous organic mutagens and suspect carcinogens in ambient air. *Environ Sci Technol* 16: 872-880 (as cited in ATSDR 1999).
- 588. Snyder RD, Van Houten B. 1986. Genotoxicity of formaldehyde and an evaluation of its effects on the DNA repair process in human diploid fibroblasts. *Mutat Res* 165(1): 21-30. (Support not reported. Authors affiliated with Stauffer Chemical Company, CT; University of Tennessee, TN.)
- 589. Soffritti M, Maltoni C, Maffei F, Biagi R. 1989. Formaldehyde: an experimental multipotential carcinogen. *Toxicol Ind Health* 5(5): 699-730. (Supported by the Institute of Oncology "F. Addarrii," Italy. Authors affiliated with Institute of Oncology "F. Addarrii," Italy.)
- 590. Soffritti M, Belpoggi F, Lambertin L, Lauriola M, Padovani M, Maltoni C. 2002a. Results of long-term experimental studies on the carcinogenicity of formaldehyde and acetaldehyde in rats. *Ann N Y Acad Sci* 982: 87-105. (Supported by the Regional Agency for Prevention and Environment of the Emilia-Romagna Region, Italy. Authors affiliated with European Ramazzini Foundation for Oncology and Environmental Sciences, Italy.)
- 591. Sogut S, Songur A, Ozen OA, Ozyurt H, Sarsilmaz M. 2004. Does the subacute (4-week) exposure to formaldehyde inhalation lead to oxidant/antioxidant imbalance in rat liver? *Eur J Gen Med* 1(3): 26-32. (Support not reported. Authors affiliated with Mustafa Kemmel University; Afyon Kocatepe University; Gaziosmanpasa University; Firat University.)
- 592. Songur A, Akpolat N, Kus I, Ozen OA, Zararsiz I, Sarsilmaz M. 2003. The effects of inhaled formaldehyde during the early postnatal period in the hippocampus of rats: a morphological and immunohistochemical study. *Neurosci Res Commun* 33: 168-178. (Supported by Firat University. Authors affiliated with Afyon Kocatepe University School of Medicine, Turkey; Firat University School of Medicine, Turkey.)
- 593. Speit G, Schütz P, Merk O. 2000. Induction and repair of formaldehyde-induced DNA-protein crosslinks in repair-deficient human cell lines. *Mutagenesis* 15(1): 85-90. (Supported by the program Environment and Health (PUG) at the Forschungszentrum Karlsruhe with funds from the Department for the Environment Baden-Wurttemburg. Authors affiliated with Universitatsklinikum Ulm, Germany.)
- 594. Speit G, Merk O. 2002. Evaluation of mutagenic effects of formaldehyde *in vitro*: detection of crosslinks and mutations in mouse lymphoma cells. *Mutagenesis* 17(3): 183-187. (Support not reported. Authors affiliated with Universitatsklinikum Ulm, Germany.)

- 595. Speit G. 2006. The implausibility of systemic genotoxic effects measured by the comet assay in rats exposed to formaldehyde. *J Proteome Res* 5(10): 2523-4. (Support not reported. Authors affiliated with Universitat Ulm, Germany.)
- 596. Speit G, Schutz P, Hogel J, Schmid O. 2007a. Characterization of the genotoxic potential of formaldehyde in V79 cells. *Mutagenesis* 22(6): 387-94. (Supported by the European Chemical Industry Council. Authors affiliated with Universitat Ulm, Germany.)
- 597. Speit G, Schmid O, Frohler-Keller M, Lang I, Triebig G. 2007b. Assessment of local genotoxic effects of formaldehyde in humans measured by the micronucleus test with exfoliated buccal mucosa cells. *Mutat Res* 627(2): 129-35. (Supported by the European Chemical Industry Council. Authors affiliated with Universitat Ulm, Germany; Universitatsklinikum Heidelberg, Germany.)
- 598. Speit G, Schmid O, Neuss S, Schutz P. 2008b. Genotoxic effects of formaldehyde in the human lung cell line A549 and in primary human nasal epithelial cells. *Environ Mol Mutagen* 49(4): 300-7. (Supported by the European Chemical Industry Council. Authors affiliated with Universitat Ulm, Germany.)
- 599. Speit G, Zeller J, Schmid O, Elhajouji A, Ma-Hock L, Neuss S. 2009. Inhalation of formaldehyde does not induce systemic genotoxic effects in rats. *Mutat Res* 677(1-2): 76-85. (Supported by the European Chemical Industry Council. Authors affiliated with Universitat Ulm, Germany; Novartis Pharma AG, Switzerland; BASF SE, Germany.)
- 600. Spicer CW, Buxton BE, Holdren MW, Smith DL, Kelly TJ, Rust SW, Pate AD, Sverdrup GM, Chuang JC. 1996. Variability of hazardous air pollutants in an urban area. *Atmos Environ* 30: 3443-3456. (Supported by the U.S. EPA. Authors affiliated with Battelle, OH.)
- 601. SRI. 2009a. *Directory of Chemical Producers. Formaldehyde*. SRI Consulting. <u>http://www.sriconsulting.com/DCP/Public/index.html</u>. Accessed on 4/30/09.
- 602. SRI. 2009b. *Directory of Chemical Producers. Paraformaldehyde*. SRI Consulting. <u>http://www.sriconsulting.com/DCP/Public/index.html</u>. Accessed on 4/30/09.
- 603. SRI. 2009c. *Directory of Chemical Producers. Trioxane*. SRI Consulting. <u>http://www.sriconsulting.com/DCP/Public/index.html</u>. Accessed on 4/30/09.
- 604. Stayner L, Smith AB, Reeve G, Blade L, Elliott L, Keenlyside R, Halperin W. 1985. Proportionate mortality study of workers in the garment industry exposed to formaldehyde. *Am J Ind Med* 7(3): 229-240. (Support not reported. Authors affiliated with NIOSH; Rhode Island Department of Health; City of Houston Health Department, TX.)

- 605. Stayner LT, Elliott L, Blade L, Keenlyside R, Halperin W. 1988. A retrospective cohort mortality study of workers exposed to formaldehyde in the garment industry. *Am J Ind Med* 13(6): 667-681. (Support not reported. Authors affiliated with NIOSH; Caribbean Epidemiologic Research Center.)
- 606. Steinsvag K, Bratveit M, Moen BE. 2007. Exposure to carcinogens for defined job categories in Norway's offshore petroleum industry, 1970 to 2005. *Occupational And Environmental Medicine* 64(4): 250-258. (Supported by the Norwegian Oil Industry Association. Authors affiliated with University of Bergen, Norway.)
- 607. Stellman SD, Demers PA, Colin D, Boffetta P. 1998. Cancer mortality and wood dust exposure among participants in the American Cancer Society Cancer Prevention Study-II (CPS-II). *Am J Ind Med* 34(3): 229-237. (Supported by the American Cancer Society, NCI and the European Commission. Authors affiliated with American Health Foundation, NY; University of British Columbia, Canada; IARC.)
- 608. Sterling TD, Collett CW, Sterling EM. 1987. Environmental tobacco smoke and indoor air quality in modern office work environments. *J Occup Med* 29(1): 57-62 (as cited in ATSDR 1999).
- 609. Sterling TD, Weinkam JJ. 1988. Reanalysis of lung cancer mortality in a National Cancer Institute study on mortality among industrial workers exposed to formaldehyde. *J Occup Med* 30(11): 895-901. (Supported by L'Office de Protection du Consommateur of the Province of Quebec, Canada. Authors affiliated with Simon Fraser University, Canada.)
- 610. Sterling TD, Weinkam JJ. 1989a. Reanalysis of lung cancer mortality in a National Cancer Institute Study on "Mortality among industrial workers exposed to formaldehyde". *Exp Pathol* 37(1-4): 128-132. (Supported by L'Office de Protection du Consommateur of the Province of Quebec, Canada. Authors affiliated with Simon Fraser University, Canada.)
- 611. Sterling TD, Weinkam JJ. 1989b. Reanalysis of lung cancer mortality in a National Cancer Institute Study of "Mortality among industrial workers exposed to formaldehyde": additional discussion. *J Occup Med* 31: 881-884. (Support not reported. Authors affiliated with Simon Fraser University, Canada.)
- 612. Sterling TD, Weinkam JJ. 1994. Mortality from respiratory cancers (including lung cancer) among workers employed in formaldehyde industries. *Am J Ind Med* 25(4): 593-602. (Supported by L'Office de Protection du Consommateur of the Province of Quebec, Canada. Authors affiliated with Simon Fraser University, Canada.)
- 613. Stern DR, Steinhagen RM. 2007. Anorectal cancer following topical formalin application for haemorrhagic radiation proctitis. *Colorectal Dis* 9(3): 275-8.

(Support not reported. Authors affiliated with Mount Sinai School of Medicine, NY.)

- 614. Stern FB, Beaumont JJ, Halperin WE, Murthy LI, Hillis BW, Fajen JM. 1987. Mortality of chrome leather tannery workers and chemical exposures in tanneries. *Scan J Work Environ Health* 13(2): 108-117. (Support not reported. Authors affiliated with NIOSH; University of California Davis, CA.)
- 615. Stewart P, Cubit D, Blair A. 1987a. Formaldehyde levels in seven industries. *Appl Ind Hyg* 2(6): 213-236. (Support not reported. Authors affiliated with NCI.)
- 616. Stewart PA, Schairer C, Blair A. 1989. Comparison of jobs, exposures, and mortality risks for short-term and long-term workers. *J Occup Med* 32(8): 703-708. (Support not reported. Authors affiliated with NCI.)
- 617. Stewart PA, Herrick RF, Feigley CE, Utterback DF, Hornung R, Mahar H, Hayes R, Douthit DE, Blair A. 1992. Study design for assessing exposures of embalmers for a case-control study. Part I. Monitoring results. *Appl Occup Environ Hyg* 7: 532-540.(as cited in IARC 2006).
- 618. Stone R, Tucker B, Depaso D, Shepard E, Villanneal E. 1981. Evaulation of Formaldehyde Problems in Residential Mobile Houses. Report HUJD-0002070. Order No. PB82-144619. Washington, D.C.: U.S. Environmental Protection Agency (as cited in WHO 1989).
- 619. Stone RA, Youk AO, Marsh GM, Buchanich JM, McHenry MB, Smith TJ. 2001. Historical cohort study of US man-made vitreous fiber production workers: IV. Quantitative exposure-response analysis of the nested case-control study of respiratory system cancer. *J Occup Environ Med* 43(9): 779-792. (Supported by the North American Insulation Manufacturers Association. Authors affiliated with University of Pittsburgh, PA; Harvard School of Public Health, MA.)
- 620. Stone RA, Youk AO, Marsh GM, Buchanich JM, Smith TJ. 2004. Historical cohort study of U.S. man-made vitreous fiber production workers IX: summary of 1992 mortality follow up and analysis of respiratory system cancer among female workers. *J Occup Environ Med* 46(1): 55-67. (Supported by the North American Insulation Manufacturer's Association. Authors affiliated with University of Pittsburgh, PA; Harvard School of Public Health, MA.)
- 621. Stroup NE, Blair A, Erikson GE. 1986. Brain cancer and other causes of death in anatomists. *J Natl Cancer Inst* 77(6): 1217-1224. (Supported by the Public Health Service, NIEHS and Brown University. Authors affiliated with Yale University, CT; CDC; NIH; Brown University, RI.)
- 622. Stumpf JM, Blehm KD, Buchan RM, Gunter BJ. 1986. Characterization of particleboard aerosol--size distribution and formaldehyde content. *Am Ind Hyg*

*Assoc J* 47(12): 725-730. (Supported by NIOSH and the American Industrial Hygiene Foundation. Authors affiliated with Hewlett Packard, CA; Colorado State University, CO; NIOSH.)

- 623. Subramaniam RP, Richardson RB, Morgan KT, Kimbell JS, Guilmette RA. 1998. Computational fluid dynamics simulations of inspiratory airflow in the human nose and nasopharynx. *Inhal Toxicol* 10: 91-120. (Supported by the U.S. EPA. Authors affiliated with Chemical Industry Institute of Toxicology, NC; Lovelace Respiratory Institute, NM.)
- 624. Subramaniam RP, Crump KS, Van Landingham C, White P, Chen C, Schlosser PM. 2007. Uncertainties in the CIIT model for formaldehyde-induced carcinogenicity in the rat: a limited sensitivity analysis-I. *Risk Anal* 27(5): 1237-54. (Supported by the U.S. EPA. Authors affiliated with U.S. EPA; ENVIRON International Corporation, LA.)
- 625. Subramaniam RP, Chen C, Crump KS, Devoney D, Fox JF, Portier CJ, Schlosser PM, Thompson CM, White P. 2008. Uncertainties in biologicallybased modeling of formaldehyde-induced respiratory cancer risk: identification of key issues. *Risk Anal* 28(4): 907-23. (Support not reported. Authors affiliated with U.S. EPA; Louisiana Tech University, LA; NIEHS.)
- 626. Sul D, Kim H, Oh E, Phark S, Cho E, Choi S, Kang HS, Kim EM, Hwang KW, Jung WW. 2007. Gene expression profiling in lung tissues from rats exposed to formaldehyde. *Arch Toxicol*. (Supported by the Medical Research Center for Environmental Toxico-Genomics and Proteomics, Korea Science and Engineering Foundations, the Ministry of Science and Technology and the Ministry of Environment. Authors affiliated with Korea University, South Korea; MyGene Bioscience Institute, South Korea; Chonnam University, South Korea; Chung-Ang University, South Korea.)
- 627. Suruda A, Schulte P, Boeniger M, Hayes RB, Livingston GK, Steenland K, Stewart P, Herrick R, Douthit D, Fingerhut MA. 1993. Cytogenetic effects of formaldehyde exposure in students of mortuary science. *Cancer Epidemiol Biomarkers Prev* 2(5): 453-460. (Supported by NIOSH and NCI. Authors affiliated with NIOSH; NCI; University of Cincinnati, OH; Cincinnati College of Mortuary Science, OH.)
- 628. Suskov, II, Sazonova LA. 1982. Cytogenetic effects of epoxy, phenolformaldehyde and polyvinylchloride resins in man. *Mutat Res* 104(1-3): 137-40. (Support not reported. Authors affiliated with Academy of Sciences of the USSR, Russia.)
- 629. Swenberg J, Kerns W, Pavkov K, Mitchell R, Gralla EJ. 1980b. Carcinogenicity of formaldehyde vapor: interim findings in a long-term bioassay of rats and mice. *Dev Toxicol Environ Sci* 8: 283-286. (Support not reported. Authors

affiliated with Chemical Industry Institute of Toxicology, NC; Battelle Columbus Laboratories, OH.)

- 630. Swenberg J, al. E. 1983b. Mechanisms of formaldehyde toxicity. In *Formaldehyde Toxicity*. Gibson JE, ed. New York: Hemisphere. pp. 132-147. (Support and author affiliations not reported.)
- 631. Swenberg JA, Kerns WD, Mitchell RI, Gralla EJ, Pavkov KL. 1980a. Induction of squamous cell carcinomas of the rat nasal cavity by inhalation exposure to formaldehyde vapor. *Cancer Res* 40(9): 3398-3402. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC; Battelle Columbus Laboratories, OH.)
- 632. Swenberg JA, Barrow CS, Boreiko CJ, Heck HD, Levine RJ, Morgan KT, Starr TB. 1983a. Non-linear biological responses to formaldehyde and their implications for carcinogenic risk assessment. *Carcinogenesis* 4(8): 945-952. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)
- 633. Takahashi M, Hasegawa R, Furukawa F, Toyoda K, Sato H, Hayashi Y. 1986. Effects of ethanol, potassium metabisulfite, formaldehyde and hydrogen peroxide on gastric carcinogenesis in rats after initiation with N-methyl-N'-nitro-N-nitrosoguanidine. *Jpn J Cancer Res* 77(2): 118-24. (Supported by the Ministry of Education, Science and Culture and the Ministry of Health and Welfare, Japan. Authors affiliated with National Institute of Hygienic Sciences, Japan.)
- 634. Takahashi S, Tsuji K, Fujii K, Okazaki F, Takigawa T, Ohtsuka A, Iwatsuki K. 2007. Prospective study of clinical symptoms and skin test reactions in medical students exposed to formaldehyde gas. *J Dermatol* 34(5): 283-9. (Supported by the Japan Chemical Industry Association. Authors affiliated with Okayama Graduate School of Medicine, Japan.)
- 635. Tan Y-M, DiBerardinis L, Snmith T. 1999. Exposure assessment of laboratory students. *Appl Occup Environ Hyg* 14: 530-538 (as cited in IARC 2006).
- 636. Tanaka K, Nishiyama K, Yaginuma H, Sasaki A, Maeda T, Kaneko SY, Onami T, Tanaka M. 2003. [Formaldehyde exposure levels and exposure control measures during an anatomy dissecting course]. *Kaibogaku Zasshi* 78(2): 43-51 (as cited in IARC 2006).
- 637. Tang LX, Zhang YS. 2003. Health investigation on workers exposed to formaldehyde. *Occup Health* 19(7): 34-35 (as cited in Tang *et al* 2009).
- 638. Tang X, Bai Y, Duong A, Smith MT, Li L, Zhang L. 2009. Formaldehyde in China: Production, consumption, exposure levels, and health effects. *Environ Int*. (Supported by the Department of Science and Technology of Guangdong Province, China, and the Northern California Center for Occupational and

Environmental Health. Authors affiliated with University of California Berkeley, CA; Guangdong Poison Control Center, China.)

- 639. Tao XM, Liu CH, Zhao XQ. 1990. Investigation on workers occupationally exposed to low concentration of formaldehyde. *Ind Health Occup Dis* 16(3): 152-153 (as cited in Tang *et al.* 2009).
- 640. Tarvainen L, Kyyronen P, Kauppinen T, Pukkala E. 2008. Cancer of the mouth and pharynx, occupation and exposure to chemical agents in Finland [in 1971-95]. *Int J Cancer* 123(3): 653-659. (Supported by the Finnish Dental Society Apollonia and the Finnish Dental Association for Women. Authors affiliated with University of Helsinki, Finland; Finnish Cancer Registry, Finland; Finnish Institute of Occupational Health, Finland.)
- 641. Taskinen H, Kyyronen P, Hemminki K, Hoikkala M, Lajunen K, Lindbohm ML. 1994. Laboratory work and pregnancy outcome. *J Occup Med* 36(3): 311-9 (as cited in IARC 2006).
- 642. Taskinen HK, Kyyronen P, Sallmen M, Virtanen SV, Liukkonen TA, Huida O, Lindbohm ML, Anttila A. 1999. Reduced fertility among female wood workers exposed to formaldehyde. *Am J Ind Med* 36(1): 206-12. (Support not reported. Authors affiliated with Finnish Institute of Occupational Health, Finland; Lappeenranta Regional Institute of Occupational Health, Finland; Finnish Cancer Registry, Finland; University of Tampere, Finland.)
- 643. Tatham L, Tolbert P, Kjeldsberg C. 1997. Occupational risk factors for subgroups of non-Hodgkin's lymphoma. *Epidemiology* 8(5): 551-558.
  (Supported by NCI. Authors affiliated with American Cancer Society, GA; Emory University, GA; University of Utah, UT.)
- 644. Teng S, Beard K, Pourahmad J, Moridani M, Easson E, Poon R, O'Brien PJ. 2001. The formaldehyde metabolic detoxification enzyme systems and molecular cytotoxic mechanism in isolated rat hepatocytes. *Chem Biol Interact* 132(1-3): 285-296. (Support not reported. Authors affiliated with University of Toronto, Canada; Health Canada.)
- 645. Thomson EJ, Shackleton S, Harrington JM. 1984. Chromosome aberrations and sister-chromatid exchange frequencies in pathology staff occupationally exposed to formaldehyde. *Mutat Res* 141(2): 89-93. (Support not reported. Authors affiliated with Western General Hospital, UK; University of Birmingham, UK.)
- 646. Thrasher JD, Kilburn KH. 2001. Embryo toxicity and teratogenicity of formaldehyde. *Arch Environ Health* 56(4): 300-11. (Support provided by the law firm, Gordon, Edlestein, Krepack, Grant, Felton, and Goldstein (Los Angeles, CA), for the cost of translating papers published in Russian and Japanese. Authors affiliated with Sam-1 Trust, NM; University of Southern California, CA.)

- 647. Thun MJ, Lakat MF, Altman R. 1982. Symptom survey of residents of homes insulated with urea--formaldehyde foam. *Environ Res* 29(2): 320-34. (Support not reported. Authors affiliated with New Jersey State Health Department.)
- 648. TIG. 2005. *Urea Formaldehyde Resins*. Morristown, NJ: The Innovation Group. 16 pp. (Support and author affiliations not reported.)
- 649. Tikuisis T, Phibbs MR, Sonnenberg KL. 1995. Quantitation of employee exposure to emission products generated by commercial-scale processing of polyethylene. *Am Ind Hyg Assoc J* 56(8): 809-14 (as cited in IARC 2006).
- 650. Til HP, Woutersen RA, Feron VJ, Hollanders VH, Falke HE, Clary JJ. 1989. Two-year drinking-water study of formaldehyde in rats. *Food Chem Toxicol* 27(2): 77-87. (Support not reported. Authors affiliated with TNO-CIVO Toxicology and Nutrition Institute, Netherlands.)
- 651. Titenko-Holland N, Levine AJ, Smith MT, Quintana PJ, Boeniger M, Hayes R, Suruda A, Schulte P. 1996. Quantification of epithelial cell micronuclei by fluorescence in situ hybridization (FISH) in mortuary science students exposed to formaldehyde. *Mutat Res* 371(3-4): 237-248. (Supported by NIOSH, NIH and NCI. Authors affiliated with University of California at Berkeley, CA; NIOSH, OH; NCI, MD; San Diego State University, CA; University of Utah, UT.)
- 652. Titford ME, Horenstein MG. 2005. Histomorphologic assessment of formalin substitute fixatives for diagnostic surgical pathology. *Arch Pathol Lab Med* 129(4): 502-6. (Support not reported. Authors affiliated with University of South Alabama, AL; The Dermatology Group, NJ.)
- 653. Tobe M, Naito K, Kurokawa Y. 1989. Chronic toxicity study on formaldehyde administered orally to rats. *Toxicology* 56(1): 79-86. (Support not reported. Authors affiliated with National Institute of Hygienic Sciences, Japan.)
- 654. Tomkins BA, McMahon JM, Caldwell WM, Wilson DL. 1989. Liquid chromatographic determination of total formaldehyde in drinking water. *J Assoc Off Anal Chem* 72(5): 835-839. (Support not reported. Authors affiliated with Oak Ridge National Laboratory, TN.)
- 655. Tong ZM, Shi J, Zhao JS, Yang H, Jiang RM, Kong L, Etal. 2006. Analysis on genetic toxicity of formaldehyde on occupational exposure population. *Chin J Public Health* 22(7): 783-784 (as cited in Tang *et al* 2009).
- 656. Tong ZM, Zhu SX, Shi J. 2007. Effect of formaldehyde on blood component and blood biochemistry of exposed workers. *Chin J Ind Med* 20(6): 409-410 (as cited in Tang *et al* 2009).
- 657. TRI. 2009. Trends Report: TRI On-site and Off-site Reported Disposed of or Otherwise Released (in pounds), Trend Report for facilities in All Industries,

*FORMALDEHYDE, U.S., 1988-2007.* U.S. Environmental Protection Agency. <u>http://www.epa.gov/triexplorer/</u>. Accessed on 5/19/09.

- 658. Triebig G, Schaller KH, Beyer B, Muller J, Valentin H. 1989. Formaldehyde exposure at various workplaces. *Sci Total Environ* 79(2): 191-5 (as cited in IARC 2006).
- 659. Tsai CF, Shiau HW, Lee SC, Chou SS. 2003. Determination of low-moleculeweight aldehydes in packed drinking water by high performance liquid chromatography. *J Food Drug Anal* 11(1): 46-52. (Support not reported. Authors affiliated with Department of Health, Taiwan.)
- 660. Tsukahara S, Yamamoto S, Tin Tin Win S, Ahmed S, Kunugita N, Arashidani K, Fujimaki H. 2006. Inhalation of low-level formaldehyde increases the Bcl-2/Bax expression ratio in the hippocampus of immunologically sensitized mice. *Neuroimmunomodulation* 13(2): 63-8. (Supported by the Ministry of the Environment and the Ministry of Education, Culture, Sports, Science and Technology. Authors affiliated with National Institute for Environmental Studies, Japan; University of Occupational and Environmental Health, Japan.)
- 661. Tuazon EC, Graham RA, Winer AM, Easton RR, Pitts Jr JN, Hanst PL. 1978. A kilometer path length fourier-transforma infrared system for the study of trace pollutants in ambient and synthetic atmospheres. *Atmos Environ* 12: 865-875 (as cited in WHO 1989).
- 662. Tyihák E, Bocsi J, Timár F, Rácz G, Szende B. 2001. Formaldehyde promotes and inhibits the proliferation of cultured tumour and endothelial cells. *Cell Prolif* 34(3): 135-141. (Support not reported. Authors affiliated with Hungarian Academy of Sciences, Hungary; Semmelweis University, Hungary.)
- 663. Ulsamer AG, Gupta KC, Cohn MS, Pruss PW. 1982. Formaldehyde in indoor air: Toxicity at risk. In 75th Annual Meeting of the Air Pollution Control Association, New Orleans, Louisiana, 20-25 June, 1982 (as cited in WHO 1989).
- 664. Usanmaz SE, Akarsu ES, Vural N. 2002. Neurotoxic effects of acute and subacute formaldehyde exposures in mice. *Environ Toxicol Pharmacol* 11: 93-100. (Support not reported. Authors affiliated with Ankara University, Turkey.)
- 665. USDA. 1999. *Wood Handbook. Wood as an Engineering Material*. General Technical Report FPL-GTR-113. Madison, WI: United States Department of Agriculture. 486 pp.
- 666. USDHEW. 1966. *Report on a Study of Formaldehyde Exposures: Perfection Garment Manufacturing, Martinsburg, West Virgina*. Washington, D.C.: U.S. Department of Health, Education and Welfare (as cited in WHO 1989).

- 667. USDHEW. 1968. Formaldehyde Emissions in the Permanent Press Fabrics Industry. TR-52. Cincinnati, OH: U.S. Department of Health, Education and Welfare, Consumer Protection and Environmental Health Service (as cited in WHO 1989).
- 668. Vargová M, Janota S, Karelova J, Barancokova M, Sulcova M. 1992. Analysis of the health risk of occupational exposure to formaldehyde using biological markers. *Analusis* 20(8): 451-454. (Support not reported. Authors affiliated with Institute of Preventive and Clinical Medicine, Czechoslovakia; Derer's Hospital, Czechoslovakia; National Institute of Health, Czechoslovakia.)
- 669. Vargová M, Wagnerova J, Liskova A, Jakubovsky J, Gajdova M, Stolcova E, Kubova J, Tulinska J, Stenclova R. 1993. Subacute immunotoxicity study of formaldehyde in male rats. *Drug Chem Toxicol* 16(3): 255-75. (Support not reported. Authors affiliated with Institute of Clinical and Preventive Medicine, Czechoslovakia; Comenius University, Czechoslovakia.)
- 670. Vasudeva N, Anand C. 1996. Cytogenetic evaluation of medical students exposed to formaldehyde vapor in the gross anatomy dissection laboratory. *J Am Coll Health* 44(4): 177-179. (Support not reported. Authors affiliated with All India Institute of Medical Sciences, India; Lady Hardinge Medical College, India.)
- 671. Vaughan TL, Strader C, Davis S, Daling JR. 1986a. Formaldehyde and cancers of the pharynx, sinus and nasal cavity: I. Occupational exposures. *Int J Cancer* 38(5): 677-683. (Supported by the U.S. EPA. Authors affiliated with Fred Hutchinson Cancer Research Center, WA; University of Washington, WA.)
- 672. Vaughan TL, Strader C, Davis S, Daling JR. 1986b. Formaldehyde and cancers of the pharynx, sinus and nasal cavity: II. Residential exposures. *Int J Cancer* 38(5): 685-688. (Supported by the U.S. EPA. Authors affiliated with Fred Hutchinson Cancer Research Center, WA; University of Washington, WA.)
- 673. Vaughan TL, Stewart PA, Teschke K, Lynch CF, Swanson GM, Lyon JL, Berwick M. 2000. Occupational exposure to formaldehyde and wood dust and nasopharyngeal carcinoma. *Occup Environ Med* 57(6): 376-384. (Supported by NCI. Authors affiliated with Fred Hutchinson Cancer Research Center, WA; University of Washington, WA; NCI; University of British Columbia, Canada; University of Iowa, IO; Michigan State University, MI; University of Utah, UT; Memorial Sloan-Kettering Cancer Research Center, NY.)
- 674. Veraldi A, Costantini AS, Bolejack V, Miligi L, Vineis P, van Loveren H. 2006. Immunotoxic effects of chemicals: A matrix for occupational and environmental epidemiological studies. *Am J Ind Med* 49(12): 1046-55. (Supported by the "Fondazione S. Paolo" in Turin. Authors affiliated with Unit of Environmental and Occupational Epidemiology, Italy; University of Turin, Italy; Imperial

College London, UK; Dutch National Institute of Public Health and the Environment, Netherlands; Maastricht University, Netherlands.)

- 675. Vinzents P, Laursen B. 1993. A national cross-sectional study of the working environment in the Danish wood and furniture industry--air pollution and noise. *Ann Occup Hyg* 37(1): 25-34 (as cited in IARC 2006).
- 676. Vock EH, Lutz WK, Ilinskaya O, Vamvakas S. 1999. Discrimination between genotoxicity and cytotoxicity for the induction of DNA double-strand breaks in cells treated with aldehydes and diepoxides. *Mutat Res* 441(1): 85-93. (Support not reported. Authors affiliated with University of Wurzburg, Germany; Kazan State University, Russia.)
- 677. Von Hippel PH, Wong KY. 1971. Dynamic aspects of native DNA structure: kinetics of the formaldehyde reaction with calf thymus DNA. *J Mol Biol* 61(3): 587-613. (Supported by the U.S. Public Health Service. Authors affiliated with Unversity of Oregon, OR.)
- 678. Walrath J, Fraumeni JF, Jr. 1983. Mortality patterns among embalmers. *Int J Cancer* 31(4): 407-411. (Support not reported. Authors affiliated with NIH.)
- 679. Walrath J, Fraumeni JF, Jr. 1984. Cancer and other causes of death among embalmers. *Cancer Res* 44(10): 4638-4641. (Support not reported. Authors affiliated with NCI.)
- 680. Wang B, Liu DD. 2006. [Detection of formaldehyde induced developmental toxicity assessed with single cell gel electrophoresis]. *Fen Zi Xi Bao Sheng Wu Xue Bao* 39(5): 462-466. (Support not identified due to foreign language. Authors affiliated with Central China Normal University, China.)
- 681. Wang L, Hang ZY, Sun ZH, Chen YQ. 1997. The research of micronuclei in peripheral lymphocyte of workers occupationally exposed to formaldehyde. *Carcinog Teratog Mutagen* 9(2): 123-125 (as cited in Tang *et al.* 2009).
- 682. Wang R, Zhang Y, Lan Q, Holford TR, Leaderer B, Zahm SH, Boyle P, Dosemeci M, Rothman N, Zhu Y, Qin Q, Zheng T. 2009. Occupational exposure to solvents and risk of non-Hodgkin lymphoma in Connecticut women. *Am J Epidemiol* 169(2): 176-185. (Supported by NCI and NIH. Authors affiliated with Yale University School of Public Health, CT; NCI; IARC; University of South Maine, ME.)
- 683. Wang W, Xu J, Xu L, Yue B, Zou F. 2007. The instability of (GpT)n and (ApC)n microsatellites induced by formaldehyde in Escherichia coli. *Mutagenesis* 22(5): 353-357. (Support not reported. Authors affiliated with Sichuan University, China; Southwest Jiaotong University, China.)
- 684. Wantke F, Focke M, Hemmer W, Tschabitscher M, Gann M, Tappler P, Gotz M, Jarisch R. 1996b. Formaldehyde and phenol exposure during an anatomy

dissection course: A possible source of IgE-mediated sensitization? *Allergy* 51: 837-841 (as cited in IARC 2006).

- 685. Wantke F, Focke M, Hemmer W, Bracun R, Wolf-Abdolvahab S, Gotz M, Jarisch R, Gotz M, Tschabitscher M, Gann M, Tappler P. 2000. Exposure to formaldehyde and phenol during an anatomy dissecting course: sensitizing potency of formaldehyde in medical students. *Allergy* 55(1): 84-7 (as cited in IARC 2006).
- 686. Ward JB, Jr., Hokanson JA, Smith ER, Chang LW, Pereira MA, Whorton EB, Jr., Legator MS. 1984. Sperm count, morphology and fluorescent body frequency in autopsy service workers exposed to formaldehyde. *Mutat Res* 130(6): 417-424. (Supported by the U.S. EPA. Authors affiliated with University of Texas, TX; U.S. EPA.)
- 687. Warshaw EM, Ahmed RL, Belsito DV, DeLeo VA, Fowler JF, Jr., Maibach HI, Marks JG, Jr., Toby Mathias CG, Pratt MD, Rietschel RL, Sasseville D, Storrs FJ, Taylor JS, Zug KA. 2007. Contact dermatitis of the hands: cross-sectional analyses of North American Contact Dermatitis Group Data, 1994-2004. *J Am Acad Dermatol* 57(2): 301-14. (Supported by NIH. Authors affiliated with Veterans Affairs Medical Center, MN; University of Minnesota, MN; American Dermatology Associates; Columbia University; University of Louisville; University of California San Francisco; Pennsylvania State University; University of Cincinnati; University of Ottawa; University of Arizona; McGill University; Oregon Health Science University; Cleveland Clinic; Dartmouth-Hitchcock Medical Center.)
- 688. West RR, Stafford DA, Farrow A, Jacobs A. 1995. Occupational and environmental exposures and myelodysplasia: a case-control study. *Leuk Res* 19(2): 127-139. (Supported by the Medical Research Council and the Health and Safety Executive. Authors affiliated with University of Wales College of Medicine, UK.)
- 689. West S, Hildesheim A, Dosemeci M. 1993. Non-viral risk factors for nasopharyngeal carcinoma in the Philippines: results from a case-control study. *Int J Cancer* 55(5): 722-727. (Supported by the Phillipine National Research Council. Authors affiliated with Johns Hopkins University School of Medicine, MD; NCI.)
- 690. Wetter DA, Davis MD, Yiannias JA, Cheng JF, Connolly SM, el-Azhary RA, Farmer SA, Fett DD, Johnson JS, Linehan DL, Richardson DM, Schroeter AL. 2005. Patch test results from the Mayo Clinic Contact Dermatitis Group, 1998-2000. *J Am Acad Dermatol* 53(3): 416-21. (Support not reported. Authors affiliated with Mayo Clinic, MN.)
- 691. Whitehead MC, Savoia MC. 2008. Evaluation of methods to reduce formaldehyde levels of cadavers in the dissection laboratory. *Clin Anat* 21(1):

75-81. (Supported by the University of California. Authors affiliated with University of California, CA.)

- 692. WHO. 1989. *Environmental Health Criteria 89: Formaldehyde*. International Programme on Chemical Safety. <u>http://www.inchem.org/documents/ehc/ehc/ehc89.htm</u>. Last accessed: 2/16/06.
- 693. WHO. 2000. Air Quality Guidelines for Europe. 2nd Edition. WHO Regional Publications, European Series, No. 91. Copenhagen: World Health Organization.
- 694. WHO. 2002. Formaldehyde: Concise International Chemical Assessment Document 40. World Health Organization. http://www.inchem.org/documents/cicads/cicad40.htm.
- 695. WHO. 2005. Formaldehyde in Drinking-water. Background Document for Development of WHO Guidelines for Drinking-water Quality.
   WHO/SDE/WSH/05.08/48. World Health Organization. 18 pp.
- 696. Wieslander G, Norback D, Walinder R, Erwall C, Venge P. 1999a. Inflammation markers in nasal lavage, and nasal symptoms in relation to relocation to a newly painted building: a longitudinal study. *Int Arch Occup Environ Health* 72(8): 507-15 (as cited in IARC 2006).
- 697. Williams GM, Mori H, McQueen CA. 1989. Structure-activity relationships in the rat hepatocyte DNA-repair test for 300 chemicals. *Mutat Res* 221(3): 263-286. (Support not reported. Authors affiliated with American Health Foundation, NY; Gifu University School of Medicine, Japan.)
- 698. Williams TM, Levine RJ, Blunden PB. 1984. Exposure of embalmers to formaldehyde and other chemicals. *Am Ind Hyg Assoc J* 45(3): 172-6 (as cited in IARC 2006 and WHO 1989).
- 699. Wilmer JW, Woutersen RA, Appelman LM, Leeman WR, Feron VJ. 1987. Subacute (4-week) inhalation toxicity study of formaldehyde in male rats: 8hour intermittent versus 8-hour continuous exposures. *J Appl Toxicol* 7(1): 15-16. (Supported by the Directorate-General of Labour, Ministry of Social Affairs and Employment, Netherlands. Authors affiliated with TNO-CIVO Toxicology and Nutrition Institute, Netherlands.)
- 700. Wilmer JW, Woutersen RA, Appelman LM, Leeman WR, Feron VJ. 1989. Subchronic (13-week) inhalation toxicity study of formaldehyde in male rats: 8-hour intermittent versus 8-hour continuous exposures. *Toxicol Lett* 47(3): 287-293. (Supported by the Directorate General of Labor, Ministry of Social Affairs and Employment, Netherlands. Authors affiliated with TNO-CIVO Toxicology and Nutrition Institute, Netherlands.)

- 701. Wilson RT, Moore LE, Dosemeci M. 2004. Occupational exposures and salivary gland cancer mortality among African American and white workers in the United States. *J Occup Environ Med* 46(3): 287-297. (Support not reported. Authors affiliated with NCI.)
- 702. Witek TJ, Jr., Schachter EN, Tosun T, Beck GJ, Leaderer BP. 1987. An evaluation of respiratory effects following exposure to 2.0 ppm formaldehyde in asthmatics: lung function, symptoms, and airway reactivity. *Arch Environ Health* 42(4): 230-7 (as cited in IARC 2006).
- 703. Wolf DC, Gross EA, Lyght O, Bermudez E, Recio L, Morgan K. 1995. Immunohistochemical localization of p53, PCNA, and TGF-α proteins in formaldehyde-induced rat nasal squamous cell carcinomas. *Toxicol Appl Pharmacol* 132: 27-35. (Support not reported. Authors affiliated with Chemical Industry Institute of Toxicology, NC.)
- 704. Wong EY, Ray R, Gao DL, Wernli KJ, Li W, Fitzgibbons ED, Feng Z, Thomas DB, Checkoway H. 2006. Reproductive history, occupational exposures, and thyroid cancer risk among women textile workers in Shanghai, China. *Int Arch Occup Environ Health* 79(3): 251-258. (Supported by the National Cancer Institute [NCI] and the National Institute of Environmental Health Sciences [NIEHS]. Authors affiliated with University of Washington, WA; Fred Hutchinson Cancer Research Center, WA; Zhong Shan Hospital Cancer Center, China.)
- 705. Wong O. 1983. An epidemiologic mortality study of a cohort of chemical workers potentially exposed to formaldehyde, with a discussion on SMR and PMR. In *Formaldehyde Toxicity*. Gibson JE, ed. New York: Hemisphere. p. 256-272. (Support and affiliations not reported.)
- 706. Wortley P, Vaughan TL, Davis S, Morgan MS, Thomas DB. 1992. A casecontrol study of occupational risk factors for laryngeal cancer. *Br J Ind Med* 49(12): 837-844. (Support not reported. Authors affiliated with University of Washington, WA; Fred Hutchinson Cancer Research Center.)
- 707. Woutersen RA, Appelman LM, Wilmer JW, Falke HE, Feron VJ. 1987. Subchronic (13-week) inhalation toxicity study of formaldehyde in rats. *J Appl Toxicol* 7(1): 43-9. (Supported by the Stichting Koningin Wilhelmina Fonds. Authors affiliated with TNO-CIVO Toxicology and Nutrition Institute, Netherlands.)
- 708. Woutersen RA, van Garderen-Hoetmer A, Bruijntjes JP, Zwart A, Feron VJ. 1989. Nasal tumours in rats after severe injury to the nasal mucosa and prolonged exposure to 10 ppm formaldehyde. *J Appl Toxicol* 9(1): 39-46. (Supported by the Stichting Koningin Wilhelmina Fonds. Authors affiliated with TNO-CIVO Toxicology and Nutrition Institute, Netherlands.)

- 709. WSDE. 1998. Washington State Air Toxic Sources and Emission Estimation Methods. Publication No. 98-207. Olympia, WA: Washington State Department of Ecology. 129 pp.
- 710. Wu PC, Li YY, Lee CC, Chiang CM, Su HJ. 2003. Risk assessment of formaldehyde in typical office buildings in Taiwan. *Indoor Air* 13(4): 359-363. (Supported by the Taiwan National Science Council and the Environmental Protection Agency. Authors affiliated with National Cheng-Kung University, Taiwan.)
- 711. Xu SY, Yi GL, Li SH. 2007b. Hygienic investigation of the effect of formaldehyde on the workers' health. *Occup Health* 23(7): 491-492 (as cited in Tang *et al* 2009).
- 712. Yager JW, Cohn KL, Spear RC, Fisher JM, Morse L. 1986. Sister-chromatid exchanges in lymphocytes of anatomy students exposed to formaldehydeembalming solution. *Mutat Res* 174(2): 135-139. (Support not reported. Authors affiliated with University of California Berkeley, CA; University of California San Francisco, CA.)
- 713. Yanagawa Y, Kaneko N, Hatanaka K, Sakamoto T, Okada Y, Yoshimitu S. 2007. A case of attempted suicide from the ingestion of formalin. *Clin Toxicol* (*Phila*) 45(1): 72-6. (Support not reported. Authors affiliated with National Defense Medical College, Japan.)
- 714. Yang WH. 2007a. Hemogram of workers exposed to low concetration of formaldehyde. *Pract Prev Med* 14(3): 792 (as cited in Tang *et al.* 2009).
- 715. Yanysheva NA, Balenko NV, Chernichenko IA, Litvichenko ON, Sovertkova LS. 1998. Characteristics of modifying effects of formaldehyde on carcinogenesis. *Gig Sanit* 8: 51 54 (ascited in IARC).
- 716. Ye JX, Hong X, Song SY, Li QJ, Zhong LF. 2000. Investigation of formaldehyde concentration in department of anatomy. *J Dalian Med Univ* 22(3): 222-223 (as cited in Tang *et al.* 2009).
- 717. Ye X, Yan W, Xie H, Zhao M, Ying C. 2005. Cytogenetic analysis of nasal mucosa cells and lymphocytes from high-level long-term formaldehyde exposed workers and low-level short-term exposed waiters. *Mutat Res* 588(1): 22-27. (Support not reported. Authors affiliated with WenZhou Medical College, China; Tongji Medical College, China.)
- 718. Ying CJ, Yan WS, Zhao MY, Ye XL, Xie H, Yin SY, Zhu XS. 1997. Micronuclei in nasal mucosa, oral mucosa and lymphocytes in students exposed to formaldehyde vapor in anatomy class. *Biomed Environ Sci* 10(4): 451-5. (Support not reported. Authors affiliated with Tongji Medical University, China; Wengzhou Medical College, China. )
- 719. Ying CJ, Ye XL, Xie H, Yan WS, Zhao MY, Xia T, Yin SY. 1999. Lymphocyte subsets and sister-chromatid exchanges in the students exposed to formaldehyde vapor. *Biomed Environ Sci* 12(2): 88-94. (Support not reported. Authors affiliated with Tongji Medical University, China; Wenzhou Medical College, China.)
- 720. Youk AO, Marsh GM, Stone RA, Buchanich JM, Smith TJ. 2001. Historical cohort study of US man-made vitreous fiber production workers: III. Analysis of exposure-weighted measures of respirable fibers and formaldehyde in the nested case-control study of respiratory system cancer. *J Occup Environ Med* 43(9): 767-778. (Supported by the North American Insulation Manufacturers Association. Authors affiliated with University of Pittsburgh, PA; Harvard School of Public Health, MA.)
- 721. Yu LQ, Jiang SF, Leng SG, He FS, Zheng YX. 2005. [Early genetic effects on workers occupationally exposed to formaldehyde]. *Zhonghua Yu Fang Yi Xue Za Zhi* 39(6): 392-395. (Support not identified due to foreign language. Authors affiliated with North China Coal Medical College, China; Chinese Center for Disease Control and Prevention, China.)
- 722. ZenStoves. 2009. Zen Stoves. http://zenstoves.net/SolidFuelBurner.htm#Trioxane. Accessed on 5/20/09.
- 723. Zhang DZ, Zhang FL, Jin SY, Liu YH. 1999. Investigation on the health of workers occupationally exposed to low levels of formaldehyde. *Chin J Ind Hyg Occup Dis* 17(5): 13-14 (as cited in Tang *et al.* 2009).
- 724. Zhang GS, Li TT, Luo M, Liu JF, Liu ZR, Bai YH. 2008b. Air pollution in the microenvironment of parked new cars. *Building And Environment* 43(3): 315-319. (Support not reported. Authors affiliated with Peking University, China.)
- 725. Zhang J, Wilson WE, Lioy PJ. 1994b. Indoor air chemistry: Formation of organic acids and aldehydes. *Environ Sci Technol* 28: 1975-1982 (as cited in ATSDR 1999).
- 726. Zhang L, Chung FL, Boccia L, Colosimo S, Liu W, Zhang J. 2003. Effects of garage employment and tobacco smoking on breathing-zone concentrations of carbonyl compounds. *AIHA J (Fairfax, Va)* 64(3): 388-93 (as cited in IARC 2006).
- 727. Zhang L, Steinmaus C, Eastmond DA, Xin XK, Smith MT. 2009a. Formaldehyde exposure and leukemia: a new meta-analysis and potential mechanisms. *Mutat Res* 681(2-3): 150-68. (Support not reported. Authors affiliated with University of California Berkeley, CA; California Environmental Protection Agency; University of California Riverside, CA.)
- 728. Zhang L, Beane Freeman L, Nakamura J, Hecht S, Vandenburg J, Smith M, Sonawane B. 2009b. Formaldehyde and leukemia: Epidemiology, potential

mechanisms, and implications for risk assessment. *Environ Mol Mutagen*(in press): 25 pp. (Supported by NCI and the U.S. EPA. Authors affiliated with University of California Berkeley, CA; NCI; University of North Carolina. NC; University of Minnesota, MN; U.S. EPA; )

- 729. Zhang XB, Sun GG, Xiao HW. 2007d. Research on formaldehye pollution in anatomy lab of medical college. *Chin J Clin Anat* 25(3): 38-40 (as cited in Tang *et al.* 2009).
- 730. Zhitkovich A, Costa M. 1992. A simple, sensitive assay to detect DNA-protein crosslinks in intact cells and in vivo. *Carcinogenesis* 13(8): 1485-9. (Supported by the U.S. EPA and NIEHS. Authors affiliated with New York University Medical Center, NY.)
- 731. Zhong W, Hee SQ. 2004a. Quantitation of normal and formaldehyde-modified deoxynucleosides by high-performance liquid chromatography/UV detection. *Biomed Chromatogr* 18(7): 462-469. (Supported by the UCLA Center for Occupational and Environmental Health and NIOSH Southern California Educational Research Center. Authors affiliated with University of California at Los Angeles, CA.)
- 732. Zhong W, Que Hee SS. 2004b. Formaldehyde-induced DNA adducts as biomarkers of in vitro human nasal epithelial cell exposure to formaldehyde. *Mutat Res* 563(1): 13-24. (Supported by the UCLA Center for Occupational and Environmental Health, NIOSH Southern California Educational Research Center, and the University of California Toxic Substances Research and Teaching Program. Authors affiliated with University of California at Los Angeles.)
- 733. Zug KA, Rietschel RL, Warshaw EM, Belsito DV, Taylor JS, Maibach HI, Mathias CG, Fowler JF, Jr., Marks JG, Jr., DeLeo VA, Pratt MD, Sasseville D, Storrs FJ. 2008. The value of patch testing patients with a scattered generalized distribution of dermatitis: retrospective cross-sectional analyses of North American Contact Dermatitis Group data, 2001 to 2004. J Am Acad Dermatol 59(3): 426-31. (Supported by the Dartmouth-Hitchcock Medical Center. Authors affiliated with Dartmouth-Hitchcock Medical Center, NH.)

This Page Intentionally Left Blank

## **Glossary of Terms**

- Acinar: Pertaining to one of the granular masses which constitute a racemose or compound gland such as the pancreas.
- Acute lymphocytic leukemia (also called: Acute lymphoblastic leukemia, Acute lymphoid leukemia, Acute lymphatic leukemia): A group of neoplasms composed of immature precursor B or T lymphocytes (lymphoblasts).
- Acute myeloid leukemias: Leukemias characterized by accumulation of immature myeloid forms in the bone marrow and suppression of normal hematopoiesis.
- Acute: The clinical term is used for a disease having a short and relatively severe course. In rodent testing, usually pertains to administration of an agent in a single dose.
- Adduct: A complex that forms when a chemical binds to a biological molecule such as DNA or a protein.
- Adenocarcinoma: A cancer that develops in the lining or inner surface of an organ.
- Adenoma: An ordinarily benign neoplasm of epithelial tissue in which the neoplastic cells form glands or gland-like structures in the stroma.
- Adipose tissue: Fatty tissue.
- Aleukemia: A condition where the leukemic cells are primarily in the bone marrow and not in the peripheral circulation; white blood cell count is normal or depressed.
- Allele: Any one of a series of two or more different genes that occupy the same position (locus) on a chromosome.
- Alveolar/bronchiolar: Pertaining to the alveoli or bronchi of the lungs.
- Ambient air: Outdoor air to which the general public is exposed.
- Ameloblastoma: A malignant jaw tumor which stems from the ameloblasts, cells which form tooth enamel.
- Anemia: Lower than normal limits of circulating red blood cells.
- Aneuploidy: One or a few chromosomes above or below the normal chromosome number.
- Anthropogenic: Caused by humans.
- Apoptosis: A mechanism of cellular suicide which occurs after sufficient cellular damage, also called programmed cell death.

- Aquifer: Geologic formations containing sufficient saturated porous and permeable material to transmit water.
- Aromatic hydrocarbon: An organic chemical compound formed primarily from carbon and hydrogen atoms with a structure based on benzene rings and resembling benzene in chemical behavior; substituents on the rings(s) may contain atoms other than carbon or hydrogen.
- Ascites: Effusion and accumulation of serous fluid in the abdominal cavity.
- Atypia: an abnormality in cells.
- Autoignition: The temperature at or above which a material will spontaneously ignite (catch fire) without an external spark or flame.
- Bacteriostatic: Inhibiting the growth or multiplication of bacteria.
- **Benign tumor:** An abnormal mass of tissue that does not spread and that is not life-threatening.
- **Betel nut:** The nut of the Areca palm tree and an ingredient of betel nut quid, an addictive mix chewed in some Pacific and Asian cultures. Its use is associated with aggressive oral cancers affecting especially the inner lining of the cheeks and lips; other sites include the tongue, lower lip, tonsil and floor of the mouth.
- Bilirubin: A pigment produced when the liver processes waste products.
- **Bioaccumulation:** The process by which a material in an organism's environment progressively concentrates within the organism.
- **Bioassay:** The determination of the potency or concentration of a compound by its effect upon animals: Isolated tissues: Or microorganisms: As compared with a chemical or physical assay.
- **Bioconcentrate:** Accumulation of a chemical in tissues of a fish or other organism to levels greater than in the surrounding medium.
- **Biodegradation:** Biotransformation; the conversion within an organism of molecules from one form to another: A change often associated with change in pharmacologic activity.
- Bronchiogenic carcinoma: a carcinoma originating in the bronchi of the lung.
- Bronchioloalveolar: Derived from epithelium of terminal bronchioles.
- **Buccal cavity:** The vestibule in the mouth between the teeth and the cheeks.
- **Calendaring:** A process of smoothing or glazing paper or cloth by pressing it between plates or passing it through rollers.

**Cannula:** A tube for insertion into a duct or cavity.

**Carcinoma:** A malignant neoplasm of the epithelium.

- Carina: A projection of the lowest tracheal cartilage.
- **Chelating agent:** A substance used to reduce the concentration of free metal ion in solution by complexing it; often used to remove toxic metals from the body.
- Chromosomal aberrations: Any abnormality of a chromosome's number or structure.
- Chronic lymphocytic leukemia: A lymphoid leukemia arising from B-cells.
- **Chronic myeloid leukemia:** A cancer of the blood-forming tissues associated with an increased production of terminally differentiated myeloid cells.
- **Chronic:** Continuing for a long period time. In rodent testing, pertains to dosing schedules of greater than 3 months.
- **Cicatrical stricture:** A scar formed in the healing of a wound that causes a decrease in the diameter of a canal, duct, or other passage.
- Clastogen: Any substance which causes chromosomal breaks.
- Colitis: Inflammation of the colon.
- **Confounding:** A relationship between the effects of two or more causal factors observed in a set of data such that it is not logically possible to separate the contribution of any single causal factor to the observed effects.
- Copolymers: A polymer of two or more different monomers.
- **Creatinine:** A waste product of protein metabolism that is found in the urine.
- **Critical temperature:** the temperature above which a gas cannot be liquefied, regardless of the pressure applied.
- **Critical temperature:** The temperature of a gas above which it is no longer possible by use of any pressure: However great: To convert it into a liquid.
- Cytogenetic: The cellular constituents concerned in heredity.
- Cytotoxic: An agent that is toxic to cells.
- **Dam:** Female parent.
- **Dehydrogenation:** The removal of one or more hydrogen ions or protons from a molecule.

- **Differentiated squamous-cell types:** Neoplastic squamous cells similar in appearance to normal squamous cells, but are less orderly.
- **Diffusion coefficient:** The rate at which a substance moves from an area of high concentration to an area of low concentration.
- **Dissociation constant (pka):** The equilibrium constant for the breaking apart of a weak acid into its hydrogen and conjugate base in a water solution.
- **Dorsal:** relating to the back or posterior of a structure.
- Dysplasia: Alteration in the size, shape, and organization of adult cells.
- **Dysplasia:** an abnormality of development; in pathology, alteration in size, shape, and organization of adult cells.
- **Ectoparasitic infection:** An infection caused by a parasite that lives on the outside of the body.
- **Effluents:** Waste material such as water from sewage treatment or manufacturing plants discharged into the environment.
- **Electrocoagulation:** use of a high-frequency electric current to bring about the coagulation and destruction of tissue.
- Endogenous: Originating within an organism.
- Endogenously: Derived or produced internally.
- **Eosinophil:** A granular leukocyte with a nucleus that usually has two lobes connected by a slender thread of chromatin and is readily stained by eosin.
- **Epidemiology:** A science concerned with the occurrence and distribution of disease in populations.
- **Epididymis:** The epididymis is a coiled segment of the spermatic ducts that serves to store and transport spermatozoa between the testis and the vas deferens.
- **Epigenetics:** Changes in phenotype (appearance) or gene expression caused by mechanisms other than changes in the underlying DNA sequence.
- **Epithelial:** Relating to or consisting of epithelium.
- **Epithelium:** the cellular covering of internal and external surfaces of the body, including the lining of vessels and other small cavities.
- Erythema: Redness of the skin produced by congestion of the capillaries.

Erythrocytes: Cells that carry oxygen to all parts of the body (red blood cells).

- Esthesioneuroepithelioma: tumor consisting of undifferentiated cells of sensory nerve epithelium.
- **Esthesioneuroma:** (Olfactory neuroma) A nasal cavity tumor of nervous tissue from olfactory epithelium.
- **Eukaryote:** An organism whose cells contain a limiting membrane around the nuclear material and which undergoes mitosis.
- Ever hourly: Workers who had ever worked in an hourly job.
- **Exogenous:** Developed or originating outside the body.
- Extrahepatic: Outside of, or unrelated to, the liver.
- Fibroblasts: Connective tissue cells.
- **Fibrosarcoma:** a type of soft tissue sarcoma that begins in fibrous tissue, which holds bones, muscles, and other organs in place.
- **Flash point:** The lowest temperature at which the vapor of a combustible liquid can be made to ignite momentarily in air.
- Flux: The rate of mass flow across a unit area.
- Follicular lymphoma: The most common form of Non-Hodgkin's lymphoma in the US.
- **Forestomach:** A non-glandular expansion of the alimentary canal between the esophagus and the glandular stomach. Rodents have a forestomach and a glandular stomach, whereas, humans have a glandular stomach.
- **Formalin:** a solution of formaldehyde in water typically containing 37% formaldehyde by mass and 10% to 15% methanol as a stabilizer.
- **Fundus:** in anatomy, it is used for the bottom or base of an organ, or the part of a hollow organ farthest from its mouth.
- Gastrectomy: Surgical removal of the stomach.
- **Gavage:** In animal experiments, the introduction of material through a tube passed through the mouth into the stomach.
- Genotoxicity: The amount of damage caused to a DNA molecule.
- **Glandular stomach:** the muscular sac between the esophagus and the small intestine containing glandular tissue. The glands of the stomach secrete mucous, hydrochloric acid and digestive enzymes.

- **Grana cheese:** a class of hard, mature cheeses from Italy which have a granular texture and are often used for grating (e.g., Parmigiano-Reggiano or parmesan cheese).
- **Half-life:** The time required for a substance to be reduced to one-half its present value through degradation or through elimination from an organism.
- **Healthy-worker effect:** Phenomenon of workers usually exhibiting overall death rates lower than those of the general population due to the fact that the severely ill and disabled are ordinarily excluded from employment.
- Hematocrit: The volume percentage of the erythrocytes in the whole blood.
- Hematopoietic: Pertaining to the formation of blood or blood cells.
- **Hemolymphoreticular:** pertaining to the network of cells and tissues of the blood and lymph nodes found throughout the body.
- **Henry's law:** The relationship that defines the partition of a soluble or partially soluble species between the gas and solution phases.
- **Hepatoblastoma:** A malignant neoplasm occurring in young children, primarily in the liver, composed of tissue resembling embryonal or fetal hepatic epithelium, or mixed epithelial and mesenchymal tissues.
- Hepatocellular: Pertaining to cells of the liver.
- Hepatotoxic: A substance that is toxic to the liver.
- Heterozygotes: An organism that has different alleles at a particular gene locus on homologous chromosomes.
- **Histones:** The chief protein components of chromatin. They act as spools around which DNA winds, and they play a role in gene regulation.
- **Hodgkin's disease:** (Hodgkin's lymphoma) A form of malignant lymphoma characterized by painless progressive enlargement of the lymph nodes, spleen, and general lymphoid tissue.
- **Homozygotes:** An organism that has the same alleles at a particular gene locus on homologous chromosomes.
- **Hydrolysis:** a chemical reaction in which the interaction of a compound with water results in the decomposition of that compound.
- Hydrolysis: The chemical breakdown of a compound due to reaction with water.
- **Hydroxyl radicals:** A particularly reactive, damaging type of free radical that is formed when superoxide radicals react with hydrogen peroxide.

- **Hyperkeratosis:** excessive thickening of the outer layer of the skin, which contains keratin.
- Hyperplasia: an abnormal increase in the number of normal cells in an organ or tissue.
- **Hyperplasia:** The abnormal multiplication or increase in the number of normal cells in normal arrangement in a tissue.
- **Hypertrophy:** increase in volume of a tissue or organ produced entirely by enlargement of existing cells.
- Hypopharynx: The lowermost section of the pharynx.
- Hypopharynx: The lowermost section of the pharynx.
- **Ileitis:** Inflammation of the ileum (distal portion of the small intestine extending from the jejunum to the cecum).
- *In situ*: Latin phrase meaning confined to the site of origin; a cancer that has not metastasized or invaded neighboring tissues
- *In vitro*: Biological process taking place in a test tube: Culture dish: Or elsewhere outside a living organism.
- *In vivo*: Biological processes taking place in a living organism.
- **Intraperitoneal [i.p.] injection:** Injection within the peritoneal cavity, i.e., the area that contains the abdominal organs.
- Intravesical: occurring within the urinary bladder.
- **Isoenzymes:** Any of the chemically distinct forms of an enzyme that perform the same biochemical function.
- **Jejunitis:** Inflammation of the jejunum (a portion of the small intestine extending from the duodenum to the ileum).
- Keratinizing squamous-cell types: Neoplastic squamous cells with keratin in the cytoplasm.
- $\mathbf{K}_{oc}$  (soil organic carbon-water partitioning coefficient): A measure of the tendency for organics to be adsorbed by soil and sediment which is useful in predicting the mobility of organic contaminants in soil.
- Lacrimation: the production, secretion, and shedding of tears.
- Large B-cell lymphomas: Types of lymphomas of the B cell lineage; a common form of non-Hodgkin's lymphoma.

Large-cell diffuse lymphoma: An aggressive B cell non-Hodgkin's lymphoma.

Larynx: Also called the voice box, it is located below the pharynx in the neck.

- Larynx: Also called the voicebox, it is located below the pharynx in the neck.
- **Latency:** The time between the instant of stimulation (exposure to a substance) and the beginning of a response (disease).
- LD50: The dose that kills 50 percent of a group of test animals.
- **Leachate:** The liquid produced in a landfill from the decomposition of waste within the landfill.
- Leiomyosarcoma: a malignant (cancer) tumor of smooth muscle cells that can arise almost anywhere in the body, but is most common in the uterus, abdomen, or pelvis.
- Leukemia: A cancer of the blood-forming tissues that is characterized by a marked increase in the number of abnormal white blood cells (leukocytes).
- Leukemia: A cancer of the blood-forming tissues that is characterized by a marked increase in the number of abnormal white blood cells (leukocytes) in the peripheral blood.
- Leukocyte: White blood cell.
- **Lipid peroxidation:** The oxidative degradation of lipids by free radicals resulting in cell damage.
- **Lipophilicity:** The affinity of a molecule or a moiety for a lipophilic (as fats) environment.
- **Lymphatic:** A small sac or node in which lymph is stored; or pertaining to the lymph, lymph nodes, or vascular channels that transport lymph to the lymph nodes.
- **Lymphocyte:** A mononuclear leukocyte that is primarily a product of lymphoid tissue and participates in humoral and cell-mediated immunity.
- Lymphohaematopoietic: Of, relating to, or involved in the production of lymphocytes and cells of blood, bone marrow, spleen, lymph nodes, and thymus.
- **Lymphohematopoietic:** Of, relating to, or involved in the production of lymphocytes and cells of blood, bone marrow, spleen, lymph nodes, and thymus.
- Lymphoma: A neoplasm of the lymphatic tissue.
- Lymphoma: A neoplasm of the lymphatic tissue.

- Lymphosarcoma: Any of various malignant neoplastic disorders of lymphoid tissue; excluding Hodgkin's disease.
- Lymphosarcoma: Any of various malignant neoplastic disorders of lymphoid tissue; excluding Hodgkin's disease.
- Macroarray: A term for microarrays with larger and fewer spots in the array.
- **Macrophage:** A large cell that is present in blood, lymph, and connective tissues, removing waste products, harmful microorganisms, and foreign material from the bloodstream.
- Malignant: Tending to become progressively worse; life-threatening.
- **Meta-analysis:** The process or technique of synthesizing research results by using various statistical methods to retrieve, select, and combine results from previous separate but related studies.
- **Metabolism:** The whole range of biochemical processes that occur within living organisms, consisting both of anabolism and catabolism (the buildup and breakdown of substances, respectively).
- Metabolite: A substance produced by metabolism.
- **Metaplasia:** a change in morphology of one differentiated cell type to a differentiated cell type that does not normally occur in that tissue.
- Metaplasia: The change in the type of mature cells in a tissue to a form that is not normal for that tissue.
- **Micronuclei:** Nuclei separate from, and additional to, the main nucleus of a cell, produced during the telophase of mitosis or meiosis by lagging chromosomes or chromosome fragments derived from spontaneous or experimentally induced chromosomal structural changes.
- **Microsatellite instability:** A condition manifested by damaged DNA due to defects in the normal DNA repair process. Sections of DNA called microsatellites, which consist of a sequence of repeating units of 1 to 6 base pairs in length, become unstable and can shorten or lengthen.
- Mitogen: A substance that induces mitosis.
- Monocyte: A mononuclear phagocytic leukocyte.
- **Monomer:** A chemical subunit that is joined to other similar subunits so as to produce a polymer.

- **Multiple myeloma:** A malignant neoplasm derived from plasma cells and found at several locations in the body.
- **Multiple myeloma:** A malignant neoplasm derived from plasma cells and found at several locations in the body.
- Myelodysplasia: A description for hemopoietic stem cells that do not mature normally.
- **Myelodysplastic syndromes:** A group of clonal stem cell disorders associated with ineffective hematopoiesis and associated cytopenias.
- **Myeloid leukemias:** A heterogeneous group of neoplasms that originate from hematopoietic progenitor cells of the myeloid series (red blood cells, white blood cells, and platelets).
- Nasal cavity: Air-filled space above and behind the nose.
- **Nasal turbinates:** (nasal conchae, nasoturbinates) Scrolled spongy bones in the posterior part of the nasal cavity.
- Nasopharyngeal: Associated with the nasal (uppermost) part of the pharynx
- **Nasopharynx:** The upper part of the pharynx, posterior to the nasal cavity and above the soft palate.
- **Nasopharynx:** The upper part of the pharynx, which leads from the nasal passages to the trachea.
- **Necropsy:** The examination of the dead body of an animal by dissection so as to detail the effects of the disease.
- **Necrosis:** The pathologic death of one or more cells, or of a portion of tissue or organ, resulting from irreversible damage.
- Neoplasm: An abnormal mass of cells.
- **Neutrophil:** A granular leukocyte having a nucleus with three to five lobes connected by slender threads of chromatin.
- **Non-Hodgkin's lymphoma:** A heterogeneous group of malignant lymphomas; the only common feature being an absence of the giant Reed-Sternberg cells characteristic of Hodgkin's disease.
- **Non-Hodgkin's lymphoma:** A heterogeneous group of malignant lymphomas; the only common feature being an absence of the giant Reed-Sternberg cells characteristic of Hodgkin's disease.
- **Nucleoside:** An organic compound consisting of a purine or pyrimidine base linked to a sugar but lacking the phosphate residues that would make it a nucleotide.

- **Nucleotide:** The molecular subunit of nucleic acids; consists of a purine or pyrimidine base, a sugar, and phosphoric acid.
- Octanol-water partition coefficient ( $K_{ow}$ ): A measure of the equilibrium concentration of a compound between octanol and water.
- **Oral cavity:** The cavity of the mouth, bounded above by the hard and soft palates and below by the tongue and the mucous membrane connecting it with the inner part of the mandible.
- **Oronasal:** Pertaining to the mouth and the nose.
- **Oropharyngeal:** Associated with the part of the pharynx between the soft palate and the epiglottis.
- **Oropharynx:** The part of the pharynx between the soft palate and the epiglottis; located below the nasopharynx.
- **Oropharynx:** The part of the pharynx consisting of the base of the tongue, soft palate, and tonsils; it is located below the nasopharynx.
- **Osteochondroma:** a benign bone tumor consisting of projecting adult bone capped by cartilage.
- **Oxidation:** the addition of oxygen to a compound with a loss of electrons; always occurs accompanied by reduction.
- **Pancytopenia:** Lower than normal circulating red blood cells, white blood cells, and platelets.
- **Pantropic:** Having an affinity for many tissues; capable of attacking derivatives of any of the three embryonic layers.
- **Papilloma:** a benign tumor derived from epithelium that can arise from skin, mucous membranes, or glandular ducts.
- Paraformaldehyde: a polymer of formaldehyde.
- **Paranasal sinuses**: Air-filled cavities surrounding the nasal cavity. There are 4 pairs of paranasal sinuses: maxillary, frontal, ethmoid, and sphenoid.
- **Parenchyma:** The distinguishing or specific cells of a gland or organ, contained in and supported by the connective tissue, framework, or stroma.
- Percutaneous: Effected or performed through the skin.
- **Perirenal:** Of, relating to, occurring in, or being the tissues surrounding the kidney.
- Phagocyte: Any cell that ingest microorganisms or other cells and foreign particles.

**Pharyngitis:** Inflammation of the pharynx.

**Pharynx:** A tube leading from the nose to the esophagus and trachea, which then leads to the lungs.

**Pharynx:** The passageway connecting the oral and nasal cavities to the larynx and esophagus.

Photolysis: The decomposition or separation of molecules by the action of light.

**Polymer:** A chemical formed by the joining together of similar chemical subunits.

**Polymorphism:** A variation in the DNA that is too common to be due merely to new mutation.

**Polypoid:** resembling a polyp; i.e., a growth that protrudes from a mucous membrane.

**Prills:** Granules or pellets that flow freely and do not clump together.

**Proctitis:** Inflammation of the mucous membrane that lines the rectum.

Prokaryote: An organism that does not have a true nucleus (e.g., bacteria).

**Pulmonary:** of or relating to the lungs.

**Pyknosis:** Contraction of nuclear contents to a deep staining irregular mass; a sign of cell death.

Pylorus: a small circular opening between the stomach and the duodenum.

- **Racemic:** Denoting a mixture that is optically inactive, being composed of an equal number of dextro- and levorotary substances which are separable.
- **Rales:** wet, crackly lung noises heard on inspiration which indicate fluid in the air sacs of the lungs; often indicative of pneumonia.
- **Resin:** any of a class of solid or semisolid viscous substances obtained either as exudations from certain plants or prepared by polymerization of simple molecules.
- Rhabdomyosarcomas: a highly malignant tumor of striated muscle.
- **Rhinitis:** a nonspecific term that covers infections, allergies, and other disorders in which the mucous membranes become infected or irritated, producing a discharge, congestion, and swelling of the tissues.

Rhinitis: Inflammation of the mucous membrane of the nose.

Rhinosinusitis: Inflammation of the nose and sinuses.

- Sarcoma: A malignant tumor of connective tissue.
- Seroprevalence: The overall occurrence of a disease within a defined population at one time, as measured by blood tests.
- Sinonasal: Pertaining to the nasal and sinus cavities.
- **Sister chromatid exchange (SCE):** The exchange during mitosis of homologous genetic material between sister chromatids; increased as a result of inordinate chromosomal fragility due to genetic or environmental factors.
- Small-cell diffuse lymphoma: Lymphoma affecting immature B cells.
- **Specific gravity:** the ratio of the density of a substance to the density of a standard substance. For liquids and solids the standard substance is usually water, for gases the standard substance is air.
- Spelt-wheat: hardy wheat of inferior quality, grown mostly in Europe for livestock feed.
- Squamous-cell histotype: Cellular structure that is stratified.
- **Subacute:** Between acute and chronic; denoting the course of a disease of moderate duration or severity. In rodent testing, usually pertains to a dosing schedule of less than one month.
- **Subchronic:** In rodent testing, generally refers to a dosing schedule lasting from one to three months.
- Subcutaneous injection: Injection beneath the skin.
- **Syngenic:** Individuals or tissues that have identical genotypes (i.e., identical twins or animals of the same inbred strain, or their tissues).
- Tachycardia: Abnormally rapid heart rate.
- **Thermosetting resin:** a resin that has the property of becoming permanently hard and rigid when heated or cured.
- Thoracolumbar: pertaining to the thoracic and lumbar vertebrae.
- **Threshold limit value (TLV):** The maximum permissible concentration of a material, generally expressed in parts per million in air for some defined period of time.
- **Time-weighted average (TWA):** The average exposure concentration of a chemical measured over a period of time (not an instantaneous concentration).
- **Trioxane:** a trimer of formaldehyde used as fuel and in plastics manufacture.

Ubiquitous: Present everywhere at once.

- **Upper respiratory tract:** Consists of the nasal and oral cavities, pharynx, larynx, and trachea.
- **Urticaria:** A vascular reaction of the skin marked by the transient appearance of smooth, slightly elevated patches (wheals) and often attended by severe itching (also called hives).
- **Uveal carcinoma (intraocular melanoma):** A malignant tumor arising from melanocytes in the uvea (iris, ciliary body, choroid) of the eye.
- **Vacuolation:** Creation of small cavities containing air or fluid in the tissues of an organism.
- **Vapor density:** The ratio of the weight of a given volume of one gas to the weight of an equal volume of another gas at the same temperature and pressure.
- **Vapor pressure:** The pressure exerted by a vapor in equilibrium with its solid or liquid phase.
- **Vestibulum:** an anatomical cavity, chamber, or channel; vestibule.
- **Volatile:** Quality of a solid or liquid allowing it to pass into the vapor state at a given temperature.
- **Xenobiotic:** A pharmacologically, endocrinologically, or toxicologically active substance not endogenously produced and therefore foreign to an organism.
- **Z-DNA:** a form of DNA in which the double helix twists in a left-hand direction, thus producing a zigzag appearance.

553