

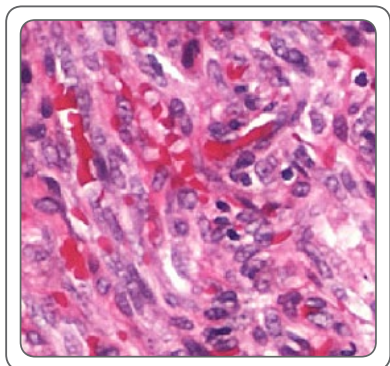


National Toxicology Program  
U.S. Department of Health and Human Services

# Report on Carcinogens

## Monograph on Kaposi Sarcoma-Associated Herpesvirus

August 2016





**National Toxicology Program**

U.S. Department of Health and Human Services

**Report on Carcinogens Monograph on  
Kaposi Sarcoma-Associated Herpesvirus**

August 1, 2016

Office of the Report on Carcinogens  
Division of the National Toxicology Program  
National Institute of Environmental Health Sciences  
U.S. Department of Health and Human Services

**This Page Intentionally Left Blank**

## Foreword

The National Toxicology Program (NTP) is an interagency program within the Public Health Service (PHS) of the Department of Health and Human Services (HHS) and is headquartered at the National Institute of Environmental Health Sciences of the National Institutes of Health (NIEHS/NIH). Three agencies contribute resources to the program: NIEHS/NIH, the National Institute for Occupational Safety and Health of the Centers for Disease Control and Prevention (NIOSH/CDC), and the National Center for Toxicological Research of the Food and Drug Administration (NCTR/FDA). Established in 1978, the NTP is charged with coordinating toxicological testing activities, strengthening the science base in toxicology, developing and validating improved testing methods, and providing information about potentially toxic substances to health regulatory and research agencies, scientific and medical communities, and the public.

The Report on Carcinogens (RoC) is prepared in response to Section 301 of the Public Health Service Act as amended. The RoC contains a list of identified substances (i) that either are *known to be human carcinogens* or are *reasonably anticipated to be human carcinogens* and (ii) to which a significant number of persons residing in the United States are exposed. The NTP, with assistance from other Federal health and regulatory agencies and nongovernmental institutions, prepares the report for the Secretary, Department of HHS. The most recent RoC, the 13th Edition (2014), is available at <http://ntp.niehs.nih.gov/go/roc>.

Nominations for (1) listing a new substance, (2) reclassifying the listing status for a substance already listed, or (3) removing a substance already listed in the RoC are evaluated in a scientific review process (<http://ntp.niehs.nih.gov/go/rocprocess>) with multiple opportunities for scientific and public input and using established listing criteria (<http://ntp.niehs.nih.gov/go/15209>). A list of candidate substances under consideration for listing in (or delisting from) the RoC can be obtained by accessing <http://ntp.niehs.nih.gov/go/37893>.

## Overview and Introduction

This one of a collection of five monographs on selected viruses that provide cancer hazard evaluations for the following human viruses: Epstein-Barr virus, Kaposi sarcoma-associated herpesvirus, human immunodeficiency virus type 1, human T-cell lymphotropic virus type 1, and Merkel cell polyomavirus for potential listing in the Report on Carcinogens (RoC). Viruses currently listed in the RoC include human papillomaviruses: some genital-mucosal types (HPV), hepatitis B virus (HBV), and hepatitis C virus (HCV). Each virus was selected for review for the RoC based on a large database of scientific information (including authoritative reviews), public health concerns for adverse health outcomes, and evidence that a significant number of people are infected with each virus both in the United States and worldwide.

This section provides background information on the preparation of the monographs as well as a discussion of overarching issues related to evaluating the evidence for cancer from human epidemiological studies and evaluating the causation by viruses.

### **Background**

The RoC monograph for each virus consists of the following components: (Part 1) the cancer hazard evaluation component that reviews the relevant scientific information and assesses its quality, applies the RoC listing criteria to the scientific information, and recommends an RoC listing status, and (Part 2) the substance profile containing the NTP's listing recommendation, a summary of the scientific evidence considered key to reaching that recommendation, and information on properties, exposure, and federal regulations and guidelines. Information reviewed in the monographs, with the exception of information on properties and exposure, comes from publicly available and peer-reviewed sources. All sections of the monographs underwent scientific and quality assurance review by independent reviewers.

The cancer hazard evaluation component provides the following information relevant to a RoC listing recommendation: Properties and Detection (Section 1), Human Exposure (Section 2), Human Cancer Studies (Section 3), Mechanistic and Other Relevant Data (Section 4), and Overall Cancer Hazard Evaluation and Listing Recommendation (Section 5). Because these viruses are primarily species-specific for humans, we are not conducting an evaluation of the level of evidence for carcinogenicity from studies in experimental animals and are including studies in animals that inform the mechanisms of carcinogenicity in the Mechanistic and Other Relevant Data section of the monographs, which is similar to the approach used by IARC. Also, specific details about the strains of the viruses are given only if needed to provide context, such as in the viral Properties and Detection section. The monographs relied on the information and data provided in previous IARC monographs on these five viruses in addition to newer key studies or reviews published since the IARC monographs; it is a peer-review assessment of available data through August 17, 2015. Additional publications published after that date were added to the monograph based on recommendations from the peer-review panel that reviewed this document on December 17, 2015. Literature search strategies to obtain information relevant to the cancer evaluation are in Appendix A of each virus monograph; search terms were developed in collaboration with a reference librarian.

***Issues related to evaluating the evidence from human epidemiological studies***

The available studies of specific types of cancer for these human viruses present several challenges with respect to the evaluation of methodological strengths and limitations of the body of evidence. Large prospective cohort studies, particularly those that follow individuals for whom infection status is documented prior to follow-up or cancer diagnosis, have several potential methodological strengths, including evidence that infection precedes cancer diagnosis, adequate statistical power, and, in some studies, have the ability to analyze dose-response relationships. However, there is the potential for misclassification of exposure in studies with a long follow-up period that measure the virus once and have a long follow-up period as new infections might not be identified. For most types of cancer, only cross-sectional or retrospective cohort studies or hospital- or clinic-based case-control studies are available, all of which lack direct evidence of temporality and may lack power or adequate exposure data, e.g., on viral load. However, molecular evidence from human studies and mechanistic data can be used in the evaluation of temporality, distinguishing latent infections caused by the tumor virus and causality. For some (typically rare) outcomes (e.g., cutaneous T-cell lymphoma and human T-cell lymphotropic virus type 1, or lymphoepithelial carcinoma of the salivary gland and Epstein-Barr virus), only case-comparison studies, in which selection of comparison groups may be biased, unmatched, or inadequately described, or case series are available.

For several rare types of cancer, e.g., adult T-cell leukemia/lymphoma and human T-cell lymphotropic virus type 1, or primary effusion lymphoma and Kaposi sarcoma-associated herpesvirus, the presence of the virus in the tumor cells is used as a diagnostic criterion to define the cancer, and thus, evidence of causality relies on cases defined by this criterion and molecular evidence from human studies rather than on epidemiological population-based studies of the association of the virus with a level of cancer risk.

In addition, methodologically adequate studies should include measurement of cofactors and consider potentially confounding factors; however, relatively few studies have measured a panel of other viruses or taken into account other cofactors. Further, while studies comparing cancer risk in treated vs. untreated populations may provide indirect evidence of the role of human immunodeficiency virus type 1, these studies, in particular calendar-period analyses, may not adequately account for changes in risk attributable to improved survival rates or changes in other risk factors.

***Issues related to evaluating causality of viruses***

Approximately 12% of all human cancers have been attributed to viral infections. Although the known oncogenic viruses belong to different virus families, they often share several common traits, such as, viral cancers appear in the context of persistent infections, occur many years to decades after acute infection, and the immune system can play a deleterious or a protective role (Mesri *et al.* 2014). Many viruses generally increase cancer risk in the context of immunosuppression or chronic inflammation (Mesri *et al.* 2014). Similar to other carcinogenic agents, only a small percentage of infected or exposed individuals develop cancer, often decades after the initial infection, reflecting the complex nature of oncogenesis. Some cofactors produced by other organisms or agents in conjunction with risk modifiers such as virus-host cell interactions, host genetic factors, immune dysfunction, or chronic inflammation often can contribute to malignant transformation. In addition, severe immunosuppression, as seen with

congenital immunodeficiency syndromes, chronic human immunodeficiency virus type 1 infection, or as a result of tissue anti-rejection medication, can severely compromise the immune surveillance capabilities of the patient. There are also other challenges that are somewhat unique to the evaluation of the epidemiological studies of viruses and cancer (discussed below) and thus molecular evidence from human tissues is often considered in the evaluation of causality.

In light of these issues, IARC monographs and several other publications have recommended paths to evaluate causality, which are discussed below and incorporated into the NTP approach for evaluating causality of the viruses. What is important for public health in determination of causation of a health effect, such as risk for cancer, is whether the health effect is eliminated or mitigated by removal of the substance (Rothman and Greenland 2005).

A number of attempts have been made to develop criteria or considerations that address causal associations. However, all of them have limitations, especially when applied to infectious agents (Moore and Chang 2010). The following sections identify factors to consider for evaluating causality, some limitations arising from a strict application of the criteria in the context of virally induced cancers, some alternative approaches, and finally, the NTP's approach for evaluating the role of select viral agents in human cancer.

#### *Hill's characteristics of an association for evaluation of causality in epidemiological studies*

Hill proposed nine characteristics to consider when evaluating causality, primarily for epidemiological studies, although they have been expanded for evaluating mechanistic and other types of data (Table 1). Several considerations—strength of the association, consistency across studies, evidence of an exposure-response gradient, and temporality of exposure (Hill 1965)—are used to help guide the RoC evaluations of the human epidemiological data (see RoC Handbook, NTP 2015). However, it should be noted that these are not criteria; and, with the exception of temporality, each and every element is not required in order to demonstrate causality (Rothman and Greenland 2005). Hill (1965) avoided discussing the meaning of “causation,” noting that the “cause” of an illness could be immediate and direct or remote and indirect. The primary question addressed by Hill was “whether the frequency of the undesirable event B will be influenced by a change in the environmental feature A.”

**Table 1. Hill's epidemiological characteristics for evaluating causality**

| Characteristic             | Description  |
|----------------------------|--|
| 1. Strength of association | A strong association between a virus and a cancer increases the confidence for causality unless confounded by some other exposure. However, a weak association does not give evidence against causality. |
| 2. Consistency             | Consistent findings observed among different groups of people, in different places, circumstances, and times.  |
| 3. Specificity             | A viral exposure is limited only to specific types of cancer; this is considered a weak factor because there are well-established examples in which a virus might cause several types of cancer.         |
| 4. Temporality             | Exposure to the virus must occur prior to the onset of the cancer, in contrast to a “passenger infection.”   |
| 5. Biological gradient     | The virus is more likely to be found at the tumor site than at non-tumor sites.  |
| 6. Plausibility            | This characteristic should be applied with caution because it is limited by  |

| Characteristic | Description  |
|----------------|--|
|                | current medical knowledge (e.g., a currently implausible mechanism may gain acceptance with increased understanding of the underlying biology).                            |
| 7. Coherence   | A virus-cancer association should not seriously conflict with known facts on the cancer's natural history and biology.   |
| 8. Experiment  | Changing either exposure or continued infection in a randomized clinical trial should change the measure of clinical outcome (e.g., vaccination programs for HPV and HBV). |
| 9. Analogy     | Are related viruses clearly established to cause cancers in animals or humans?   |

Source: Moore and Chang 2014.

### *Consideration of mechanistic data from studies in humans*

In their evaluation of the evidence for Epstein-Barr virus, the IARC working group noted that the large majority of people are latently infected with Epstein-Barr virus, thus, epidemiological studies may be limited in determining whether the presence of Epstein-Barr virus in tumor tissue is a cause of the cancer or an effect of the tumor. Therefore, in addition to the Hill characteristics, IARC (1997) considered the following factors in their evaluation of Epstein-Barr virus, which are also applicable to other viruses:

- The proportion of Epstein-Barr virus-positive cases in a given tumor entity.
- The proportion of tumor cells that carry the virus.
- The monoclonality of Epstein-Barr virus in the tumor.
- The expression of Epstein-Barr virus proteins.

zur Hausen (2001, 1994) proposed consideration of the following types of mechanistic or epidemiological evidence for evaluating causality of viruses and cancer:

- The presence and persistence of viral DNA in tumor biopsies and cell lines derived from the same tumor type.
- The growth-promoting activity of specific viral genes or of virus-modified host-cell genes in tissue culture systems or in suitable animal systems.
- The continuous expression of viral oncogenes or the modification of host-cell genes containing viral sequences which maintains the malignant phenotype.
- The epidemiological evidence that the virus infection is a major risk factor.

It is difficult to prove that a virus causes cancer, and such determinations almost always generate considerable controversy and debate (Moore and Chang 2010). Viral cancers employ various mechanisms that involve both direct and indirect modes of interaction (Table 2) (zur Hausen and de Villiers 2014). Understanding and managing viral-induced cancers in humans has been hampered by a lack of suitable animal models, the disparate nature of tumor types, a long latency period between primary infection and cancer development, the different types of oncogenic viruses, and the complex nature of the virus-host cell interactions leading to cancer (Mesri *et al.* 2014, zur Hausen and de Villiers 2014).



**Table 2. Direct and indirect modes of interaction of viral infections and cancers**

| Type                    | Description  |
|-------------------------|--|
| Direct carcinogenesis   | Continued presence and expression of viral oncogenes usually after viral genome integration into host cell DNA.<br>Insertional gene activation or suppression.<br>Continued episomal presence of viral nucleic acid and suppression or activation of cellular genes (e.g., by viral microRNA). |
| Indirect carcinogenesis | Induction of immunomodulation, activation of latent tumor virus genomes.<br>Induction of oxygen and nitrogen radicals.<br>Amplification of latent tumor virus DNA.<br>Induction of mutations and/or translocations.<br>Prevention of apoptosis.  |

Source: zur Hausen and de Villiers 2014.

### *Multicausality issues*

Although thousands of viruses are known to cause infection, only a few have been shown to cause cancer in humans (Moore and Chang 2010). An important consideration regarding causality (not limited to viruses) is “multicausality,” that is, the concept that many determinants act together to cause a disease. Rothman and colleagues (Rothman and Greenland 2005) defined a sufficient cause as “complete causal mechanism” – not a signal factor but a set of minimal factors (i.e., component causes)—that if present in an individual will cause disease. Most causes are neither necessary nor sufficient in the absence of other factors to produce the disease; however, a cause does not have to be either necessary or sufficient for its removal to result in disease prevention (Rothman and Greenland 2005, zur Hausen and de Villiers 2014).

### *Application of causality criteria and alternative approaches*

Moore and Chang (2010) investigated the difficulties associated with strict application of the Hill characteristics for two of the most recently discovered oncogenic viruses: Kaposi sarcoma-associated herpesvirus and Merkel cell polyomavirus. Kaposi sarcoma-associated herpesvirus was shown to fulfill Hill’s characteristics for causality of Kaposi sarcoma; however, the application of the characteristics was problematic in the case of Merkel cell polyomavirus and Merkel cell carcinoma (see the monographs for Kaposi sarcoma-associated herpesvirus and Merkel cell polyomavirus). These two examples illustrate the diversity in the patterns of tumor virus epidemiology. Some of the reasons Hill’s characteristics worked for Kaposi sarcoma-associated herpesvirus but not Merkel cell polyomavirus is that all clinical forms of Kaposi sarcoma require infection by Kaposi sarcoma-associated herpesvirus while most studies indicate that not all forms of Merkel cell carcinoma require the presence of Merkel cell polyomavirus. In the case of Merkel cell polyomavirus, additional considerations, as suggested by IARC (1997) and zur Hausen (2001, 1994), provide molecular evidence of the association between Merkel cell polyomavirus and Merkel cell carcinoma, such as mutation and monoclonal integration of the tumor-causing form of the virus into the cellular genome and requirement of tumor cells for the presence of viral oncoproteins for cell survival and proliferation.

While causal criteria can be helpful, there are flaws and practical limitations that restrict their use in cancer biology (Moore and Chang 2010). Therefore, a more probabilistic approach may be more useful for determining whether or not certain viruses cause human cancers. For example, instead of trying to determine if virus A causes cancer B, the probabilistic approach examines if cancer B is more probable in the presence of virus A. Although a correlation does not imply causation, it can be argued that correlations that are strong, reproducible, and predictive have a similar value as a causative conclusion. In a similar fashion, zur Hausen and de Villiers (2014) also expressed concern over all attempts to summarize criteria for “causality” of infectious agents in cancer development and proposed replacing “causal factor” with “risk factor” and grading them according to their contribution to an individual’s cancer risk. This will require a greater understanding of the complexity of factors involved and their mechanistic contribution to individual cancers.

### ***NTP’s approach***

For each virus, the NTP applied the RoC listing criteria (see text box) to the body of literature to reach the listing recommendation. The level of evidence conclusion from studies in humans considers the evidence from epidemiological studies as well as clinical and molecular studies of tissues from exposed (i.e., infected) individuals. In evaluating the mechanistic data and determining the recommendations for its level of evidence conclusion and overall listing recommendation, the NTP considered the principles outlined by Hill, IARC, zur Hausen, and Rothman in its assessment of causality for the five viruses reviewed. However, these factors were not used as a strict checklist to either prove or disprove a causal association but rather as guidance to assess the level of epidemiological or molecular evidence that a virus contributes to a carcinogenic effect.

#### **RoC Listing Criteria**

##### ***Known To Be Human Carcinogen:***

There is sufficient evidence of carcinogenicity from studies in humans\*, which indicates a causal relationship between exposure to the agent, substance, or mixture, and human cancer.

##### ***Reasonably Anticipated To Be Human Carcinogen:***

There is limited evidence of carcinogenicity from studies in humans\*, which indicates that causal interpretation is credible, but that alternative explanations, such as chance, bias, or confounding factors, could not adequately be excluded, OR

there is sufficient evidence of carcinogenicity from studies in experimental animals, which indicates there is an increased incidence of malignant and/or a combination of malignant and benign tumors (1) in multiple species or at multiple tissue sites, or (2) by multiple routes of exposure, or (3) to an unusual degree with regard to incidence, site, or type of tumor, or age at onset, OR

there is less than sufficient evidence of carcinogenicity in humans or laboratory animals; however, the agent, substance, or mixture belongs to a well-defined, structurally related class of substances whose members are listed in a previous Report on Carcinogens as either known to be a human carcinogen or reasonably anticipated to be a human carcinogen, or there is convincing relevant information that the agent acts through mechanisms indicating it would likely cause cancer in humans.

Conclusions regarding carcinogenicity in humans or experimental animals are based on scientific judgment, with consideration given to all relevant information. Relevant information includes, but is not limited to, dose response, route of exposure, chemical structure, metabolism, pharmacokinetics, sensitive sub-populations, genetic effects, or other data relating to mechanism of action or factors that may be unique to a given substance. For example, there may be substances for which there is evidence of carcinogenicity in laboratory animals, but there are compelling data indicating that the agent acts through mechanisms which do not operate in humans and would therefore not reasonably be anticipated to cause cancer in humans.

\*This evidence can include traditional cancer epidemiology studies, data from clinical studies, and/or data derived from the study of tissues or cells from humans exposed to the substance in question that can be useful for evaluating whether a relevant cancer mechanism is operating in people.

## CONTRIBUTORS

### **Office of the Report on Carcinogens (ORoC), Division of the National Toxicology Program (NTP)**

*Conducted technical review and evaluation and proposed the listing recommendation*

Ruth Lunn, DrPH  
(Director, ORoC)

Gloria D. Jahnke, DVM, DABT  
(project lead)

### **Integrated Laboratory Systems, Inc. (Support provided through NIEHS Contract Number HHSN273201100004C)**

*Conducted technical review and evaluation*

Sanford Garner, PhD  
(Principal Investigator)

Andrew Ewens, PhD, DABT

Stanley Atwood, MS, DABT

Jennifer Ratcliffe, PhD, MSc

Jessica Geter, MSLS

Alton Peters, MS

Provided administrative support

Ella Darden, BS

Tracy Saunders, BS

### **Social Scientific Systems, Inc. (Support provided through NIEHS Contract Number HHSN2732015000041)**

*Conducted technical review and evaluation*

Whitney Arroyave, PhD

### **Technical Advisors**

Jim Goedert, M.D.  
Senior Investigator  
Infections and Immunoepidemiology Branch  
Division of Cancer Epidemiology & Genetics  
National Cancer Institute

Elizabeth 'Betsy' Read-Connole, Ph.D.  
Head, Cancer Etiology Section  
Cancer Immunology and Hematology Etiology  
Branch  
Division of Cancer Biology  
National Cancer Institute

Denise Whitby, PhD  
Viral Oncology Section  
Leidos Biomedical Research, Inc.  
National Cancer Institute  
Frederick, MD

## Peer Review

Peer review of the Draft RoC Monograph on Kaposi sarcoma-associated herpesvirus (KSHV) was conducted by an *ad hoc* expert panel at a public meeting held December 17, 2015, in the Rodbell Auditorium at the National Institute of Environmental Health Sciences, David P. Rall Building, Research Triangle Park, NC (see <http://ntp.niehs.nih.gov/go/38854> for materials, minutes, and panel recommendations from the peer-review meeting). The selection of panel members and conduct of the peer review were performed in accordance with the Federal Advisory Committee Act and federal policies and regulations. The panel members served as independent scientists, not as representatives of any institution, company, or governmental agency.

The charge to the Peer-Review Panel was as follows:

1. To comment on the draft cancer evaluation component for KSHV, specifically, whether it was technically correct and clearly stated, whether the NTP has objectively presented and assessed the scientific evidence, and whether the scientific evidence is adequate for applying the RoC listing criteria,
2. To comment on the draft substance profile for KSHV, specifically, whether the scientific justification presented in the substance profile supports the NTP's preliminary policy decision on the RoC listing status of the substance.

The Panel was asked to vote on the following questions:

1. Whether the scientific evidence supports the NTP's preliminary conclusion on the level of evidence for carcinogenicity for the specific types of cancer from cancer studies in humans.
2. Whether the scientific evidence supports the NTP's preliminary listing decision for KSHV in the RoC.

This RoC monograph on KSHV has been revised based on NTP's review of the Panel's peer-review comments. The Peer-Review Panel Report, which captures the Panel recommendations for listing status of KSHV in the RoC and their scientific comments, and the NTP Response to the Peer-Review Report are available on the Peer-Review Meeting webpage for Kaposi sarcoma-associated herpesvirus (<http://ntp.niehs.nih.gov/go/38854>).

**Peer-Review Panel Members****Andrew F. Olshan, PhD (Chair)**

Distinguished Professor and  
Chair  
University of North Carolina  
Chapel Hill, NC

**Edward L. Murphy, Jr., MD, MPH**

Professor, Departments of Laboratory  
Medicine and  
Epidemiology/Biostatistics  
University of California, San Francisco  
San Francisco, CA

**Blossom Damania, PhD**

Assistant Dean of Research, School of  
Medicine, Professor  
University of North Carolina  
Chapel Hill, NC

**Charles S. Rabkin, MD, MSc**

Senior Investigator, Infections and  
Immunoepidemiology Branch, DCEG  
National Cancer Institute  
Bethesda, MD

**Paul F. Lambert, PhD**

Professor and Chair of Oncology, Director,  
McArdle Laboratory for Cancer  
Research  
University of Wisconsin School of  
Medicine and Public Health  
Wisconsin Institutes for Medical Research  
Madison, WI

**Rosemary Rochford, PhD**

Professor, Immunology and Microbiology  
Environmental and Occupational Health  
University of Colorado

**Margaret Madeleine, MPH, PhD**

Associate Member  
Program in Epidemiology  
Fred Hutchinson Cancer Research Center  
Seattle, WA

## Table of Contents

### Part 1: Cancer Hazard Evaluation

|       |   |    |
|-------|---|----|
| 1     | Properties and Detection .....                                    | 1  |
| 1.1   | Biological properties .....                                       | 1  |
| 1.1.1 | Family and type.....  | 1  |
| 1.1.2 | Genome .....  | 2  |
| 1.1.3 | Infection and replication .....                                   | 2  |
| 1.2   | Detection methods .....   | 3  |
| 1.2.1 | Detection in fluids.....  | 4  |
| 1.3   | Summary .....   | 5  |
| 2     | Human Exposure.....   | 7  |
| 2.1   | Prevalence and transmission .....                                 | 7  |
| 2.2   | Diseases, prevention, and treatment .....                         | 8  |
| 2.3   | Summary .....   | 9  |
| 3     | Human Cancer Studies.....   | 11 |
| 3.1   | Introduction .....  | 11 |
| 3.2   | Selection of the literature .....                                 | 11 |
| 3.3   | Cancer evaluation: Kaposi sarcoma .....                           | 12 |
| 3.3.1 | Background information .....                                      | 12 |
| 3.3.2 | Descriptive epidemiological studies .....                         | 13 |
| 3.3.3 | Cohort and nested case-control studies.....                       | 13 |
| 3.3.4 | Case-control studies .....  | 16 |
| 3.3.5 | Cofactors for Kaposi sarcoma.....                                 | 18 |
| 3.3.6 | Integration across studies of Kaposi sarcoma.....                 | 20 |
| 3.4   | Lymphohematopoietic cancers.....                                  | 21 |
| 3.4.1 | Primary effusion lymphoma .....                                   | 21 |
| 3.4.2 | Multicentric Castleman disease .....                              | 23 |
| 3.4.3 | Multiple myeloma.....   | 25 |
| 3.4.4 | Other lymphohematopoietic cancers.....                            | 26 |
| 3.4.5 | Integration across studies .....                                  | 28 |
| 3.5   | Other tumor sites .....   | 28 |
| 3.6   | Synthesis across cancer endpoints.....                            | 29 |
| 4     | Mechanistic and Other Relevant Data .....                         | 31 |
| 4.1   | Characteristics and risk factors.....                             | 31 |
| 4.1.1 | Human immunodeficiency virus type 1 and immunosuppression.....    | 32 |
| 4.1.2 | KSHV transcripts .....  | 33 |
| 4.1.3 | Other possible cofactors.....                                     | 39 |
| 4.2   | Mode of action and evidence for cancer causation .....            | 39 |
| 4.2.1 | Kaposi sarcoma.....   | 40 |
| 4.2.2 | Primary effusion lymphoma .....                                   | 40 |
| 4.2.3 | Multicentric Castleman disease .....                              | 41 |
| 4.3   | Synthesis.....  | 42 |
| 5     | Overall Cancer Hazard Evaluation and Listing Recommendation ..... | 43 |

|   |     |
|---|-----|
| References.....                             | 45  |
| Glossary .....                              | 65  |
| Abbreviations.....                          | 67  |
| Appendix A: Literature Search Strategy..... | A-1 |
| General approach.....                       | A-1 |
| Search strings for KSHV Searches.....       | A-2 |

## Part 1: Cancer Hazard Profile

|                            |     |
|----------------------------|-----|
| Cancer Hazard Profile..... | P-1 |
|----------------------------|-----|

### List of Tables

|  |     |
|--|-----|
| Table 1. Hill's epidemiological characteristics for evaluating causality .....   | iv  |
| Table 2. Direct and indirect modes of interaction of viral infections and cancers .....  | vi  |
| Table 1-1. Detection methods.....  | 5   |
| Table 3-1. Summary of cohort and nested case-control studies of risk of Kaposi sarcoma among<br>KSHV-seropositive vs. KSHV-negative participants ..... | 14  |
| Table 3-2. Summary of case-control studies of Kaposi sarcoma and Kaposi sarcoma-associated<br>herpesvirus (KSHV).....                                  | 17  |
| Table 3-3. Case-control study of co-factors for the development of classic Kaposi sarcoma<br>among a KSHV-positive population in Sicily.....           | 20  |
| Table 3-4. Summary of case-series and case-control/comparison studies of KSHV and<br>multicentric Castleman disease .....                              | 24  |
| Table 3-5. Summary of five case-control/comparison and nested case-control studies of KSHV<br>and multiple myeloma <sup>a</sup> .....                  | 27  |
| Table 3-6. Summary of KSHV cancer endpoints and strength of the epidemiological evidence .....   | 29  |
| Table 4-1. Characteristics of KSHV-associated neoplasms .....  | 32  |
| Table 4-2. Biochemical and biological properties of latent KSHV proteins .....   | 35  |
| Table 4-3. Lytic KSHV proteins and their role in carcinogenesis.....   | 37  |
| Table 5-1. Summary of the evidence for KSHV and Kaposi sarcoma from human studies .....  | 43  |
| Table 5-2. Summary of the evidence for KSHV and primary effusion lymphoma from human<br>studies.....   | 44  |
| Table 5-3. Summary of the evidence for KSHV and multicentric Castleman disease from human<br>studies.....  | 44  |
| Table A-1. Major topics searched.....  | A-1 |

### List of Figures

|  |     |
|--|-----|
| Figure 1-1. KSHV particle.....                         | 1   |
| Figure 1-2. Genome schematic .....                     | 2   |
| Figure 1-3. KSHV infection and replication cycle ..... | 3   |
| Figure A-1. Literature processing flow.....            | A-2 |

---

## **Part 1**

---

# **Cancer Hazard Evaluation**

**Properties and Detection**

**Human Exposure**

**Human Cancer Studies**

**Mechanisms and Other Relevant Data**

**Overall Cancer Hazard Evaluation and Listing  
Recommendation**

---



**This Page Intentionally Left Blank**

---

# 1 Properties and Detection

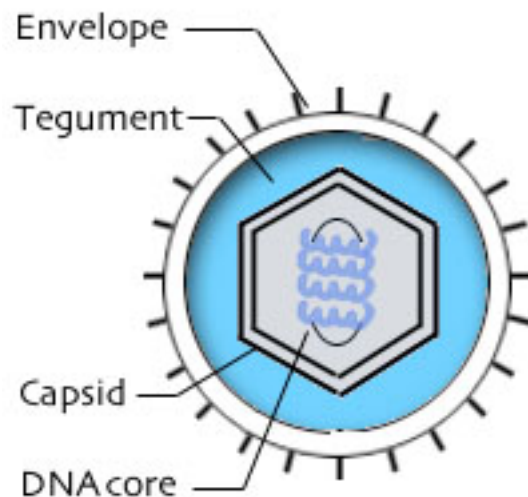
This section reviews the biological properties (Section 1.1) and detection methods (Section 1.2) of the Kaposi sarcoma-associated herpesvirus (KSHV). The information in this section is summarized in Section 1.3.

## 1.1 Biological properties

### 1.1.1 Family and type

KSHV, also designated as human herpesvirus 8 (HHV8), was the first member of the gamma-2 herpesviruses identified in humans in 1994 in association with acquired immunodeficiency syndrome (AIDS) (Chang *et al.* 1994, Fukumoto *et al.* 2011, IARC 1997). Rhadinoviruses also include herpesviruses that infect New World monkeys, macaque Old World monkeys, cattle, and rodents (IARC 1997).

KSHV consists of linear double-stranded DNA (dsDNA) wrapped around a core protein within a protein capsid, surrounded by a membrane envelope containing glycoproteins (Edelman 2005, Giffin and Damania 2014, IARC 2012, Fukumoto *et al.* 2011, IARC 1997) (see Figure 1-1). The capsid is made up of five major capsid proteins forming hexamer and pentamer proteins and a minor capsid protein that binds to the edges of the hexamer and pentamer junctions (Giffin and Damania 2014, IARC 2012). Between the capsid and the membrane envelope are six tegument proteins and 11 viral RNA strands (Giffin and Damania 2014, Fukumoto *et al.* 2011).



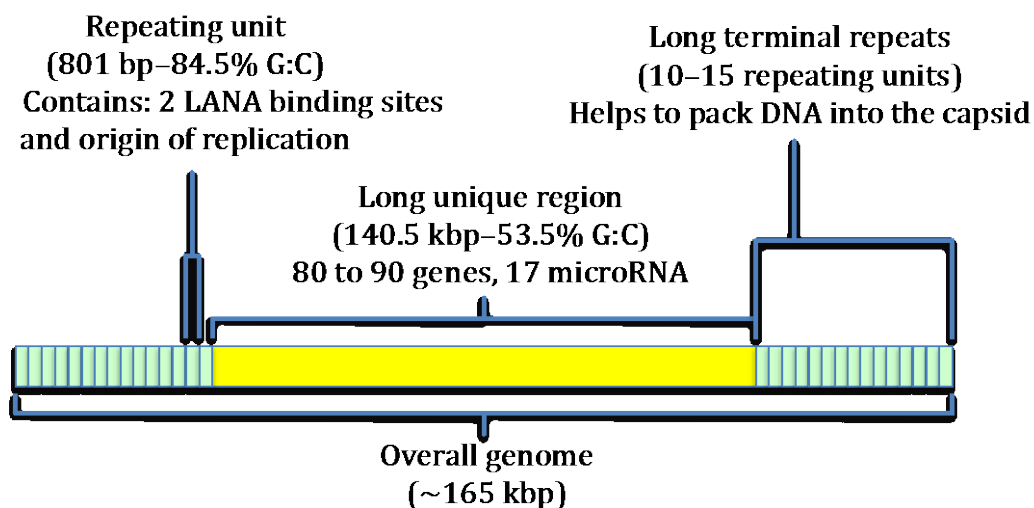
**Figure 1-1. Kaposi sarcoma-associated herpes virus particle**

Source: Los Alamos National Laboratory.

### 1.1.2 Genome

The KSHV genome, with an overall size of 165 kbp, consists of two main sections, (1) the long unique region, which contains over 80 open reading frames (ORF) for protein and microRNA (miRNA) genes, is 140 kbp long and (2) two flanking terminal repeat regions composed of a variable number of repeating units (see Figure 1-2) (IARC 2012, Fukumoto *et al.* 2011, IARC 1997). The terminal repeats do not contain open reading frames, but function by providing the origin of replication, location of genome circularization, and binding sites to tether the viral genome to host chromosomes.

Most of the genes are conserved among herpesviruses and are named after the genes of the closest related virus, *Herpesvirus saimiri*, which infects non-human primates (IARC 2012, 1997). These genes are designated by the number identifier of their open reading frame, e.g., ORF 16, and are numbered consecutively from left to right across the genome (Giffin and Damania 2014). Other genes originated from host cellular genes during viral evolution and are designated by their cellular homologue, preceded by “v-” denoting a viral origin, e.g., v-Bcl-2 (IARC 2012, 1997). Some genes are unique to KSHV and are designated by the letter “K” in their open reading frame, e.g., ORF K12. MiRNAs are designated by “miR” and a number identifier, e.g., miR-K5.



#### Gene categories

Homologues of Herpes virus Saimiri

Homologues of human genes

Unique to KSHV (designated by “K” before the ORF number, ex. ORF K2)

**Figure 1-2. Genome schematic**

LANA = latency-associated nuclear antigen.

### 1.1.3 Infection and replication

Like other herpesviruses, the KSHV replication cycle includes a latent phase and a lytic phase, which allow the virus to transition between a quiescent infection and active replication (see

Figure 1-3). KSHV has been detected in endothelial cells, epithelial cells, B lymphocytes, and monocytes *in vivo*, while a wider variety of cells can be infected *in vitro* (Giffin and Damania 2014). CD19<sup>+</sup> B lymphocytes are a long-term latency reservoir for the virus. KSHV glycoproteins on the capsid surface mediate fusion with several target cell receptors, and the replication cycle starts as KSHV enters a cell by a receptor-mediated endocytosis or macropinocytosis process (Campbell *et al.* 2014a, Giffin and Damania 2014). The viral capsid then enters the nucleus and releases dsDNA. Inside the virus particle, the genome is linear, but within the host cell nucleus it circularizes, joining the terminal repeat ends together and attaching to host cell chromosomes (Campbell *et al.* 2014a, Fukumoto *et al.* 2011, IARC 1997). In the latent phase, the viral episome persists and is replicated by host machinery to daughter cells with each cell division. No infectious virus particles are produced during latency and only a small number of genes located in the latency locus are expressed. Only a low copy number of the viral genome is present in a latently infected cell, while a high copy number is produced during a lytic infection.

Most of the time, KSHV will remain in the latent phase but cell stressors, such as phorbol esters, sodium butyrate, some signaling cytokines, cell differentiation, reactive oxygen species, and innate immune signaling by Toll-like receptors, can initiate the lytic phase (Giffin and Damania 2014). At the start of the lytic phase the viral lytic transactivator protein, RTA, causes the expression of many viral genes, beginning with those involved in controlling viral gene expression. The next viral genes expressed are those involved in DNA replication and immunomodulation. Viral DNA replication occurs at multiple origins within the terminal repeat region of the circular genome to produce linear copies of newly formed genomes. The viral capsid is then enveloped by the lipid bilayer membrane as it buds off of the nuclear membrane into the cytoplasm and progeny virions are released from the cell. See Section 4.1.2 for a detailed description of latent and lytic genes.

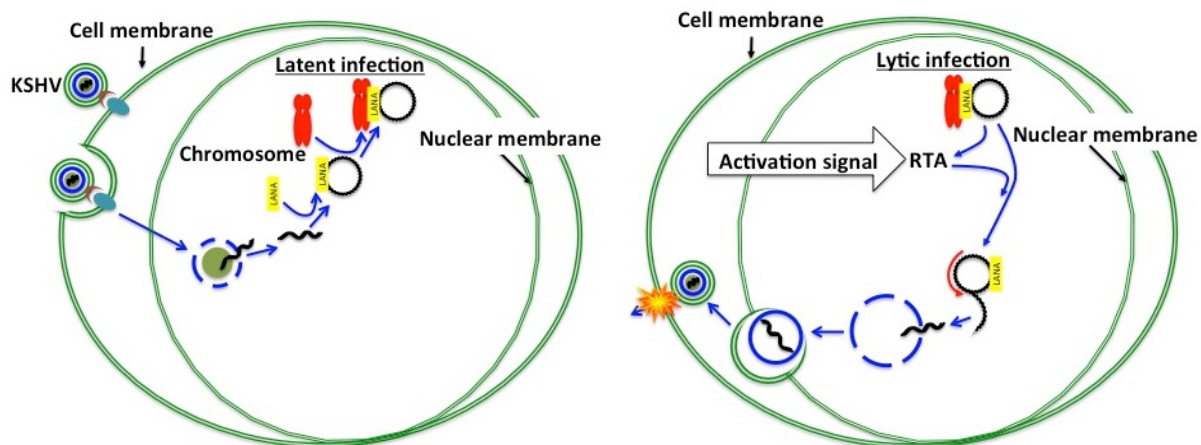


Figure 1-3. KSHV infection and replication cycle

## 1.2 Detection methods

KSHV infections can be detected by measuring viral protein antigens or viral DNA in tissues and antibodies against the virus in blood. The level of KSHV antibodies in the blood only suggests that a viral infection has occurred, as both the specificity and sensitivity of the test methods are low (Xu *et al.* 2015, Fukumoto *et al.* 2011, IARC 2012). KSHV DNA or antibodies against

KSHV-specific antigens in the blood can be used to measure viral load. The nature of the antigens recognized by the antibodies, i.e., either latent or lytic, can help to determine the predominant nature of the infection, but this test alone is not conclusive because of variations in development of antibody responses among individuals (IARC 2012). Both latent and lytic viral infections are associated with carcinogenesis (IARC 2012). In cases of Kaposi sarcoma or primary effusion lymphoma, KSHV latently infects almost all tumor cells while lytic infection is frequently seen in the plasmablastic variant of multicentric Castleman disease (see Mechanisms and Other Relevant Data, Section 4) (Dupin 2000, Damania 2010). Antibodies against two main viral proteins have been measured in humans to detect KSHV, though other protein-specific antibodies can be detected as well (see Mechanisms and Other Relevant Data, Section 4) (IARC 2012, 1997, Fukumoto *et al.* 2011). These include the latent phase protein, latency-associated nuclear antigen (LANA), (ORF 73), and the lytic phase protein, K8.1.

### 1.2.1 Detection in fluids

#### Antibodies

The level of antibody response varies depending on the phase of the infection (IARC 2012). Antibody responses are low in latently infected individuals and higher in patients with lytic infections or in patients with Kaposi sarcoma (IARC 2012). Latently infected cell lines can be used to immunohistochemically detect anti-KSHV antibodies, which stain in the nucleus only. Infected cell lines can be stimulated with phorbol esters or sodium butyrate to increase the number of cells in the lytic phase and to detect antibodies against lytic antigens, which stain the cytoplasm as well as the nucleus (Fukumoto *et al.* 2011, IARC 1997). Recombinant viral proteins and peptides have also been used in ELISA tests for KSHV antibodies (IARC 2012). Polymerase chain reaction (PCR) techniques can detect viral DNA in saliva and to a lesser extent in vaginal secretions and semen (Campbell *et al.* 2014a); however, these techniques are not used clinically.

#### Detection in cells

##### Antigens

KSHV can infect many different cell types including B cells, dendritic cells, monocytes, prostate cells, keratinocytes, fibroblasts, endothelial cells, and epithelial cells, as detected by immunohistochemical staining for KSHV antigens (Campbell *et al.* 2014a, Giffin and Damania 2014, IARC 2012, 1997, Fukumoto *et al.* 2011). KSHV encodes a viral homolog of interleukin-6 (vIL-6), which can be detected in the affected cells and blood of patients with multicentric Castleman disease, primary effusion lymphoma, and Kaposi sarcoma (Bhutani *et al.* 2015, Fukumoto *et al.* 2011, Parravicini *et al.* 1997b, 2000, Mesri *et al.* 2010, Aoki *et al.* 2001). vIL-6 is thought to play a role in multicentric Castleman disease pathogenesis by stimulating the proliferation of plasma cells.

##### DNA

Viral DNA detected in tissues by PCR can indicate KSHV infections in those tissues (IARC 2012, Fukumoto *et al.* 2011, IARC 1997). KSHV DNA can be detected in peripheral blood mononuclear cells, but not all KSHV-infected patients will give positive results (Campbell *et al.* 2014a, Fukumoto *et al.* 2011, IARC 1997). KSHV infection can occur without Kaposi sarcoma

or multicentric Castleman disease, so detecting KSHV DNA in peripheral blood mononuclear cells does not necessarily indicate the presence of these diseases. However, those with Kaposi sarcoma or multicentric Castleman disease tend to have higher viral loads in peripheral blood mononuclear cells.

**Table 1-1. Detection methods**

| Analyte       | Method  | Lytic marker                                  | Latent marker           |
|---------------|---|---|-------------------------|
| <b>Fluids</b> |   |   |                         |
| Antibodies    | Immunohistochemistry – infected primary effusion lymphoma cells | Nuclear and cytoplasmic staining <sup>a</sup> | Only nuclear staining   |
|               | ELISA   | High levels – Suggestive                      | Low levels – Suggestive |
| DNA           | PCR   | Genome detection - Not phase specific         |                         |
| <b>Cells</b>  |   |   |                         |
| Antigen       | Immunohistochemistry – tissue                                   | Nuclear and cytoplasm staining                | Only nuclear staining   |
| DNA           | PCR   | Genome detection - Not phase specific         |                         |

<sup>a</sup>Phorbol esters are used to induce the lytic phase in primary effusion lymphoma cells.

### 1.3 Summary

Kaposi sarcoma-associated herpesvirus (KSHV) is an enveloped DNA virus found in Kaposi sarcomas, primary effusion lymphoma, and some cases of multicentric Castleman disease. The genome circularizes and attaches to the host chromosome but does not integrate into the host genome. There are over 80 genes and 17 mRNAs, which include some genes that originated from host cell genes during the evolution of KSHV. The virus can reproduce lytically or can remain latent, tethered to the host chromosome. The lytic phase can be initiated by innate immune signaling, cytokine signaling, or cell differentiation or by exposure to phorbol esters, sodium butyrate, or reactive oxygen. KSHV can be detected by anti-KSHV antibodies, intracellular viral antigens, and DNA.

**This Page Intentionally Left Blank**

## 2 Human Exposure

This section describes prevalence and transmission (Section 2.1), and non-cancer diseases, prevention, and treatment of Kaposi sarcoma-associated herpesvirus (KSHV) (Section 2.2). The material presented in Sections 2.1 and 2.2 is summarized in Section 2.3.

### 2.1 Prevalence and transmission

Serological tests for detecting the multiple antigens encoded by KSHV vary in sensitivity and specificity and are poorly standardized, resulting in uncertainty in prevalence estimates and complicating comparison of KSHV in different populations (IARC 2012). Considering this caveat, KSHV prevalence rates appear to vary widely in different populations; high-level endemic areas (adult general public seroprevalences between 30% and 70%) are found in many parts of Africa (Martin 2007); intermediate-level endemic areas (general public seroprevalences between 10% and 25%) are found primarily in the Mediterranean area (Whitby *et al.* 1998); and non-endemic areas (general public seroprevalences less than 10%) include North America, Central America, South America, northern Europe, and Asia. However, in non-endemic areas, certain population groups exhibit seroprevalences comparable to those in high- and intermediate-level endemic areas (e.g., between 30% and 60% of human immunodeficiency virus type 1-infected men who have sex with men and between 20% and 30% of human immunodeficiency virus type 1-uninfected men who have sex with men are infected with KSHV (Martin *et al.* 1998, O'Brien *et al.* 1999).

The first systematic evaluation of KSHV epidemiology in the general public in the United States (based on enzyme immunoassays conducted on serum samples from National Health and Examination Survey [NHANES] III data collection) reported that overall KSHV seroprevalence is approximately 7% and is similar among men and women (Engels *et al.* 2007a). These seroprevalence data indicate that a significant number of people in the United States are exposed to KSHV. A previous study of 1,000 U.S. blood donors (collected in 1994 and 1995) tested by six independent laboratories, using various serological tests ranging from single immunofluorescence assays to decision trees based on results of several individual assays, reported KSHV seropositivity estimates ranging from 0.5% to 5% (IARC 2012, Pellett *et al.* 2003). KSHV seropositivity has been found to be lower among human immunodeficiency virus type 1-infected women than in human immunodeficiency virus type 1-infected men who have sex with men, including bisexual men (Kedes *et al.* 1997). Global KSHV seroprevalence has been shown to mirror the geographical distribution of classic or endemic (i.e., non-human immunodeficiency virus type 1-associated) forms of classic Kaposi sarcoma, e.g., high (15% to 60%) in African and southern Mediterranean regions, and low (1% to 5%) in the United States and northern Europe (Minhas and Wood 2014, Ganem 2010).

Transmission of KSHV appears to be primarily via saliva (IARC 2012). The presence of KSHV in peripheral blood suggests that transmission via blood is also possible, and transmission among some intravenous drug users, transfusion recipients, and from transplant donor to transplant recipient has been reported (IARC 2012, Barozzi *et al.* 2003). In populations with high endemic KSHV prevalence, both horizontal and vertical transmission appear to occur, primarily in



childhood between the ages of 6 and 10 years (particularly if the mother is infected) and infection rates increase with age.

Risk factors for infection may include contact with, e.g., infected family members, sources of water, and, in particular, human immunodeficiency virus type 1 infection (IARC 2012), so factors that increase the risk of human immunodeficiency virus type 1 infection (e.g., number of sexual partners) also increase the risk of infection with KSHV. Further, higher KSHV infection rates in human immunodeficiency virus type 1-positive cohorts other than those acquiring human immunodeficiency virus type 1 via parenteral infection were found to reflect sexual activity, for which human immunodeficiency virus type 1 seropositivity serves as a marker (Kedes *et al.* 1996). Men who have sex with men may have a higher risk of KSHV transmission than heterosexual sex partners even in the absence of human immunodeficiency virus type 1 infection (Giuliani *et al.* 2007), and orogenital sex has been shown to be significantly correlated with KSHV seroconversion in men who have sex with men (Dukers *et al.* 2000). The possibility of insect bites facilitating the transmission of KSHV via human saliva (i.e., application of virus-carrying saliva at the bite site to relieve itching and reduce scratching (Amodio *et al.* 2011b, Coluzzi *et al.* 2003) has also been reported.

KSHV is generally not associated with sexual risk factors in heterosexuals (IARC 2012, Engels *et al.* 2007a, Smith *et al.* 1999). However, there is some evidence for spouse-to-spouse transmission among heterosexual couples, which appears to be more efficient for female-to-male than for male-to-female transmission (Dupuy *et al.* 2009). This is based on a study that found higher KSHV seroprevalence among heterosexual spouses of Kaposi sarcoma patients than among spouses of controls without cancer. Overall, KSHV prevalence was higher among male spouses than female spouses.

## **2.2 Diseases, prevention, and treatment**

Most individuals who are otherwise healthy and infected with KSHV are asymptomatic (ACS 2014, NCI 2014, DHHS 2013). There are very few reports of primary infection with KSHV, but reported symptoms associated with initial KSHV infection include a febrile maculopapular skin rash (as observed in immunocompetent children in Egypt) (Andreoni *et al.* 2002), as well as diarrhea, fatigue, localized rash, and lymphadenopathy (in men who have sex with men without human immunodeficiency virus type 1 infection) (Wang *et al.* 2001). Other symptoms include fever, splenomegaly, cytopenia, and bone marrow failure with plasmacytosis (in renal transplant recipients) (Luppi *et al.* 2000a). Active KSHV infection may be associated with fever, cutaneous rash, and hepatitis in autologous peripheral blood stem cell infusion recipients (Luppi *et al.* 2000b).

A limited number of studies have suggested associations between KSHV infection and some non-cancer endpoints, including proliferative and nonproliferative skin diseases such as pemphigus vulgaris and pemphigus foliaceus, and actinic keratosis, but other studies have failed to confirm these associations (Ablashi *et al.* 2002). In addition, KSHV has been found in inflammatory cells in isolated cases of interstitial pneumonitis, in sarcoid tissue, and in histiocytic necrotic lymphadenitis, indicating lymph node infiltration, but an etiologic role of KSHV in these diseases has not been established (Ablashi *et al.* 2002).

Because KSHV transmission is associated with KSHV shedding in saliva and occasional shedding in genital secretions, avoiding salivary exposure and following safe sex practices should theoretically prevent transmission (DHHS 2014, 2013, Chang-Moore Laboratory 2009).

Some drugs have been reported to reduce or inhibit KSHV shedding (e.g., see Cattamanchi *et al.* 2011, Casper *et al.* 2008); however, currently there are no FDA-approved drugs for treatment of KSHV infection. Currently, there is no vaccine against KSHV; however, limited efforts to develop a vaccine are ongoing (ACS 2014, Wu *et al.* 2012).

### **2.3 Summary**

U.S. seroprevalence study data indicate that a significant number of people living in the United States are exposed to KSHV. Most individuals who are otherwise healthy and infected with KSHV are asymptomatic. Transmission appears to be primarily via saliva. Transmission via blood is also possible, and transmission among some intravenous drug users, transfusion recipients and from transplant donor to transplant recipient has been reported. Both horizontal and vertical transmission appears to occur, primarily in childhood (between the ages of 6 and 10 years, with infection rates increasing with age) in populations with high endemic KSHV prevalence, particularly if the mother is infected. Risk factors for infection may include contact with infected family members, sources of water, and particularly human immunodeficiency virus type 1 infection, so factors that increase the risk of human immunodeficiency virus type 1 infection also increase the risk of infection with KSHV. Men who have sex with men may have a higher risk of KSHV transmission than heterosexual sex partners even in the absence of human immunodeficiency virus type 1 infection. A limited number of studies have suggested associations between infection with KSHV and some non-cancer endpoints (e.g., pemphigus vulgaris, pemphigus foliaceus, and actinic keratosis), but other studies have failed to confirm these associations. There is currently no vaccine against KSHV, although limited vaccine development efforts are ongoing.

**This Page Intentionally Left Blank**

## 3 Human Cancer Studies

### 3.1 Introduction

The majority of human epidemiological studies for KSHV have focused on Kaposi sarcoma, which was first described by Dr. Mori Kaposi in 1872 (IARC 1997), prior to the discovery of the virus in 1994 (Chang *et al.* 1994). Since 1994, a number of other cancer endpoints have been evaluated, the major ones are certain B-cell non-Hodgkin lymphomas, in particular rare forms of primary effusion lymphoma, multicentric Castleman disease, multiple myeloma, and other lymphohematopoietic cancers. The evidence for these endpoints, which were identified by IARC (IARC 2012, 1997), is assessed in this section. A limited number of studies of solid tumor endpoints (including prostate, bladder, head and neck, angiosarcoma, skin, hepatocellular, and vulvar cancers, and childhood inflammatory myofibroblastoma) have also been conducted, but only in single or small studies for most endpoints, and thus are only reviewed briefly.

One of the key issues in the cancer hazard evaluation is the extent to which observed associations between KSHV and specific cancer endpoints can be explained by cofactors, including host genetic factors, co-infection with other viruses, or other environmental and/or other risk factors, as distinct from confounding factors. However, while a substantial number of studies have investigated KSHV cancer risk in association with human immunodeficiency virus type 1 infection, relatively few studies have clearly identified other cofactors, in particular the presence of other oncoviruses (e.g., Epstein-Barr virus) and non-viral cofactors.

In the majority of studies, KSHV exposure is detected based on DNA in the peripheral blood mononuclear cells, tumor, normal tissue, semen, saliva, and other biological samples or via serological measures usually measuring antibodies to lytic or latent viral antigens. Earlier studies included in this evaluation primarily used serological measurements, while later studies generally used more sensitive and specific seroprevalence measures or detected KSHV DNA.

This evaluation of the human cancer hazard associated with KSHV is divided into three major parts. First, a summary of the approach for selection of the studies is provided (Section 3.2). Next, the cancer evaluation for each endpoint is presented (Sections 3.3 to 3.5), and lastly, a summary of the evaluation across cancer endpoints is provided (Section 3.6). The level of evidence from cancer studies in humans also considers molecular studies of tissues from humans in addition to epidemiological studies and is provided in Section 5.

The NTP used the body of knowledge published in the IARC (2012, 1997) monographs on KSHV as the resource for studies conducted up to and including 2008. The key human studies identified in these monographs, together with new human studies identified from 2008 to 2015, are considered in the RoC monograph and used to evaluate the scientific evidence for specific cancer endpoints independently of IARC's conclusions. Where available, IARC data tables of the effect estimates have informed the cancer hazard assessment.

### 3.2 Selection of the literature

A literature search of major databases, citations, and other authoritative sources for literature from 2008 to August 2015 was conducted. The literature search strategy (including the databases and search terms, and other sources for identifying literature), procedures for selecting the

literature (systematic screening procedures and inclusion/exclusion criteria), and the results of the literature search are described in Appendix A. Cross-sectional or prospective cohort studies, and case-control studies, which may range from broadly defined, non-matched hospital or population case-comparison designs to formal age-, sex-, and race-matched case-control designs, as well as summaries of multiple case reports or case-series studies are included in the review.

### 3.3 Cancer evaluation: Kaposi sarcoma

#### 3.3.1 Background information

Studies of Kaposi sarcoma incidence or prevalence have been conducted among four main epidemiological types. Although Kaposi sarcoma is histologically indistinguishable in these four types, the age at diagnosis, clinical features, and progression of the disease vary considerably by type.

- “Epidemic” or “human immunodeficiency virus type 1/AIDS-related” Kaposi sarcoma occurs among human immunodeficiency virus type 1-positive, typically immunocompromised individuals;
- “Iatrogenic,” “transplant,” or “immunosuppressive” Kaposi sarcoma occurs primarily among organ recipients, mostly renal and liver transplant patients, generally in the presence of immune suppression, and also among other patients with immune suppression (e.g., hemodialysis patients [Jalilvand *et al.* 2011]);
- “Classic,” “sporadic,” or “indolent” Kaposi sarcoma is rare, less aggressive in its course than human immunodeficiency virus type 1-related Kaposi sarcoma (Hiatt *et al.* 2008), predominantly affects the lower extremities, and occurs mostly among older males 50 to 70 years of age in specific populations, e.g., in some Mediterranean countries such as southern Italy, or among East Europeans of Jewish descent with higher (e.g., 10% to 20%) KSHV prevalence rates (IARC 2012, 1997);
- “Endemic” Kaposi sarcoma also occurs mostly among men, but among younger groups than classic Kaposi sarcoma, including children, in regions of sub-Saharan Africa with relatively high KSHV seroprevalence rates, e.g., of 25% to over 50% (IARC 2012). The course of Kaposi sarcoma in children tends to be much more aggressive than that observed in adults (Senba *et al.* 2011). Since the advent of the human immunodeficiency virus type 1 epidemic, the ratio of Kaposi sarcoma in men to women has decreased from 7:1 to 2:1 in South Africa (Sitas and Newton 2001), despite little evidence that the male to female ratio of KSHV infection differs or has changed over the past two decades (Dedicoat and Newton 2003).

Kaposi sarcoma appears to be rare in immunocompetent individuals among populations with the epidemic type of the cancer, and the expression and course of the cancer may also be affected by compromised immune status in populations with classic or endemic types of Kaposi sarcoma. In the United States, Kaposi sarcoma occurred in less than 1 per 100,000 individuals prior to the human immunodeficiency virus type 1/AIDS epidemic starting in the early 1980s, peaking at approximately 5 to 6 per 100,000 in the U.S. population by the early 1990s, prior to the advent of highly active antiretroviral therapy (HAART) (NCI 2014).

Approximately 12,000 patients were diagnosed with Kaposi sarcoma between 1975 and 2005, according to SEER registry statistics (Armstrong *et al.* 2013). In Europe, the Surveillance of Rare Cancers in Europe (RARECARE) project (Stiller *et al.* 2014) reported 2,667 incident cases of Kaposi sarcoma between 1995 and 2002, an age-standardized rate of 0.3/100,000 across the entire European Union. In the subgroup of southern European countries, where predominantly classic cases have been identified, the average incidence was 0.8/100,000; the incidence rates for other areas in Europe ranged from 0.02 to 0.25. The Kaposi sarcoma incidence rate among U.S. men (approximately 3.3/100,000) (Rouhani *et al.* 2008) is higher than that among European men (0.5/10,000) but comparable among women in the United States and Europe (approximately 0.1/100,000; see Stiller *et al.* [2014]).

### 3.3.2 Descriptive epidemiological studies

A number of case series, descriptive epidemiological, or surveillance studies of Kaposi sarcoma from multiple countries, conducted from 2008 to 2015, were identified (Koski *et al.* 2014 [Tanzania], Mousavi *et al.* 2014 [immigrants to Sweden], Stiller *et al.* 2014 [European cancer surveillance registry], Jalilvand *et al.* 2012 [Iran], Tiussi *et al.* 2012 [Brazil], Jalilvand *et al.* 2011 [Iran], Ogoina *et al.* 2011 [Nigeria], Senba *et al.* 2011 [Kenya], Kanno *et al.* 2010 [Japan], Magri *et al.* 2009 [Brazil], Hiatt *et al.* 2008 [United States], Lanternier *et al.* 2008, Mwakigonja *et al.* 2008 [Tanzania], Nsubuga *et al.* 2008 [Uganda]). As supported by studies reviewed by IARC (2012), descriptive studies that report KSHV seroprevalence (typically LANA) or KSHV-DNA among cases of Kaposi sarcoma indicate ranges from approximately 88% to 98%, irrespective of whether these are human immunodeficiency virus type 1 positive, human immunodeficiency virus type 1 negative, endemic, or classic Kaposi sarcoma cases (IARC 2012, Ogoina *et al.* 2011, Kanno *et al.* 2010, Magri *et al.* 2009, Hiatt *et al.* 2008, Lanternier *et al.* 2008, Mwakigonja *et al.* 2008, Nsubuga *et al.* 2008). However, relatively few case studies of KSHV seroprevalence have been reported among human immunodeficiency virus type 1-negative populations in regions with endemic or classic Kaposi sarcoma.

### 3.3.3 Cohort and nested case-control studies

Table 3-1 summarizes findings from 13 cohort or nested case-control studies conducted prior to 2009 and reviewed by IARC (2012) and from two prospective cohort and one nested case-control studies identified since the IARC review (Labo *et al.* 2015, Wakeham *et al.* 2015, Nawar *et al.* 2008) that reported risk estimates for KSHV seroprevalence or DNA and Kaposi sarcoma. Most of the cohort studies were of immunosuppressed populations either human immunodeficiency virus type 1-positive or transplant recipients and were conducted among U.S., Canadian, or European-based populations; two cohorts were of southern Mediterranean populations who have a high prevalence of classic Kaposi sarcoma (Rezza *et al.* 1999, Parravicini *et al.* 1997a) and the nested case-control study was in Uganda, a country with a high prevalence of endemic Kaposi sarcoma (Labo *et al.* 2015).

Among the studies reviewed by IARC (2012), all of the relative risks were statistically significant, ranging from 2.5 to 75; elevated risks were observed in both human immunodeficiency virus type 1-positive and organ transplant populations, which provide support for KSHV causing Kaposi sarcoma independently of human immunodeficiency virus type 1 infection. Moreover, Newton *et al.* (2006) found that risk of Kaposi sarcoma increased with increasing anti-KSHV titers. The two prospective cohort studies conducted in the United States

(Labo *et al.* 2015, Nawar *et al.* 2008), and the small nested case-control study conducted in Uganda (Wakeham *et al.* 2015), published after the IARC review, are consistent with these findings and reported risk estimates ranging between 1.8 to 9.5. One cohort study, the multi-city U.S. cohort study of 1,354 men who have sex with men (Nawar *et al.* 2008), found that acquiring human immunodeficiency virus type 1 infection prior to KSHV infection was associated with an increase in Kaposi sarcoma risk vs. co-prevalent infection (hazard ratio [HR] = 1.81, 95% CI = 1.32 to 2.48), whereas acquiring KSHV infection prior to human immunodeficiency virus type 1 did not increase risk in comparison with co-prevalent infection (HR = 1.05, 95% CI = 0.59 to 1.87). The nested case-control study calculated risks for both lytic and latent antigens; relative risks were somewhat higher for antibody titers to K8.1 lytic antigen (RR = 4.14, 95% CI = 3.34 to 4.94) than latent antigen (LANA) (RR = 2.2, 95% CI = 1.16 to 3.25) (Wakeham *et al.* 2015). It is not possible to draw conclusions as to whether risk varies among different countries due to the limited number of studies (and few studies reported in detail on the racial or ethnic composition of their cohorts).

IARC also reported on seven cohort studies of human immunodeficiency virus type 1-positive populations or organ transplant recipients (Garcia-Astudillo and Leyva-Cobian 2006, Marcelin *et al.* 2004, Frances *et al.* 1999, 2000, Chatlynne *et al.* 1998, Melbye *et al.* 1998, Regamey *et al.* 1998) that did not calculate risk estimates, primarily due to small numbers; three of these publications reported that KSHV infection was significantly higher in Kaposi sarcoma cases than controls with *P*-values ranging from < 0.001 (Garcia-Astudillo and Leyva-Cobian 2006, Frances *et al.* 2000) to < 0.0001 (Frances *et al.* 1999).

**Table 3-1. Summary of cohort and nested case-control studies of risk of Kaposi sarcoma among KSHV-seropositive vs. KSHV-negative participants**

| Reference                                      | Detection methods         | Cohort (comparison group) and cases and controls                                     | RR       |
|--|---------------------------|--|----------|
| <b>HIV-1-positive/AIDS populations</b>         |                           |  |          |
| <i>Cohort studies</i>                          |                           |  |          |
| Grulich <i>et al.</i> 1999 (USA)               | Latent and lytic antigens | AIDS+ MSM  | 4.4*     |
| O'Brien <i>et al.</i> 1999 (USA)               | Latent nuclear antigen    | HIV-1+ MSM   | 3.6*     |
| Jacobson <i>et al.</i> 2000 (USA) <sup>a</sup> | Lytic antigens            | HIV-1+ MSM (seroconversion to KSHV after HIV-1 infection vs. before HIV-1 infection) | 2.6*     |
| Rezza <i>et al.</i> 1999 (Italy)               | Latent nuclear antigen    | HIV-1+ (various exposures)   | 29.6*    |
| Nawar <i>et al.</i> 2008 (USA)                 | Lytic antigens            | KSHV and HIV-1 concurrent infections; MSM  | 1.8*     |
|  |                           | KSHV infection prior to HIV-1 infection; MSM   | 1.1 (NS) |
| Labo <i>et al.</i> 2015 (USA, Puerto Rico)     | Latent and lytic antigens | HIV-1+ men and women   | 9.5*     |

| Reference  | Detection methods                  | Cohort (comparison group) and cases and controls                            | RR              |
|--|------------------------------------|---|-----------------|
| <b>Range</b>   |                                    |   | <b>1.8–29.6</b> |
| <b><i>Nested case-control studies</i></b>                  |                                    |   |                 |
| Moore <i>et al.</i> 1996<br>Parry and Moore 1997<br>(USA)  | PCR                                | KS cases: AIDS+ MSM<br>Controls: HIV-1+ MSM                                 | 11.6*           |
| Gao <i>et al.</i> 1996<br>(USA)                            | Latent nuclear antigen             | KS cases: AIDS+ MSM<br>Controls: HIV-1+ MSM                                 | [16.3]*         |
| Engels <i>et al.</i> 2003<br>(USA)                         | Lytic antigens<br>PBMC DNA         | KS cases: HIV-1+ MSM<br>Controls: HIV-1+ MSM                                | 11.7*           |
| Quinlivan <i>et al.</i> 2001 <sup>b</sup><br>(Switzerland) | Latent nuclear antigen             | KS cases: HIV-1+ M or F<br>Controls: HIV-1+ M or F                          | 3.0             |
| Newton <i>et al.</i> 2006<br>(UK)                          | Latent and lytic antigens          | KS cases: HIV-1+ M or F<br>Controls: HIV-1+ M or F                          | 4.4*            |
|  |                                    | Titer (latent antigens) -   |                 |
|  |                                    | Min   | 3.5*            |
|  |                                    | Med   | 4.1*            |
|  |                                    | Max   | 8.7*            |
| <i>P<sub>trend</sub></i>                                   | < 0.001                            |   |                 |
| Whitby <i>et al.</i> 1995<br>(UK)                          | PCR                                | KS cases: AIDS+ M or F<br>Controls: HIV-1+ M or F                           | 12.7*           |
| Wakeham <i>et al.</i> 2015<br>(Uganda)                     | Latent and lytic antigens          | KS cases: HIV-1+ M or F<br>Controls: HIV-1+ M or F                          |                 |
|  |                                    | K8.1  | 4.1*            |
|  |                                    | LANA  | 2.2*            |
| <b>Range</b>   |                                    |   | <b>2.2–16.3</b> |
| <b>Organ transplant recipients</b>                         |                                    |   |                 |
| <b><i>Cohort study</i></b>                                 |                                    |   |                 |
| Cattani <i>et al.</i> 2001<br>(Italy)                      | Latent and lytic antigens          | Transplant recipients tested for KSHV before transplant, follow-up 10 years | 34.4*           |
| <b><i>Nested case-control studies</i></b>                  |                                    |   |                 |
| Parravicini <i>et al.</i> 1997 <sup>a</sup><br>(Italy)     | Latent and lytic antigens          | Transplant recipients tested for KSHV before transplant                     | 75.0*           |
| Rabkin <i>et al.</i> 1999<br>(Canada)                      | Latent antigens, whole virus assay | Transplant recipients tested for KSHV before transplant                     | 2.5 (NS)        |

Source: IARC 2012 (with reported or calculated risk estimate), [Table 2.2](#), Nawar *et al.* (2008), Lado *et al.* (2015), Wakeham *et al.* (2015). IARC also described seven other nested case-control or cohort studies that reported the number of KSHV-positive or -negative Kaposi sarcoma, but did not report or provide enough information to calculate a risk estimate [ ] RR or OR calculated by NTP.

AIDS = acquired immunodeficiency syndrome; HIV-1 = human immunodeficiency virus type 1; KS = Kaposi sarcoma; KSHV = Kaposi sarcoma-associated herpesvirus; LANA = latent antinuclear antigen; MSM = men who have sex with men; PBMC =



peripheral blood mononuclear cells; PCR = polymerase chain reaction; RR = relative risk; NS = lower 95% confidence interval (CI) < 1.0.

\*Lower 95% confidence interval (CI) > 1.0.

<sup>a</sup>KSHV- and HIV-1-positive MSM with detectable KSHV viral load 1 year prior to diagnosis vs. KSHV- and HIV-1-positive MSM with no detectable viral load.

<sup>b</sup>Note that membrane antigen RR = 1.7 (95% CI = 0.5 to 6.7) and lytic antigen RR = 0.9 (95% CI = 0.2 to 6.8).

### 3.3.4 Case-control studies

IARC (2012) reviewed 80 case-control studies of Kaposi sarcoma; 68 studies included AIDS cases, 7 included human immunodeficiency virus type 1-positive or “non-AIDS” cases, and 10 studies included human immunodeficiency virus type 1-negative cases (plus 4 studies in which human immunodeficiency virus type 1 status was unclear or not specified); 27 included classic cases, 14 included endemic cases, and 9 included transplant cases (some studies had more than one group). Table 3-2 summarizes data from 39 case-control studies in which risk estimates for KSHV were reported. One new case-control study, of classic cases in a Sicilian population (Amodio *et al.* [2011b], based on an earlier study by Goedert *et al.* [2002]), was identified after the IARC (2012) review. (Further studies among KSHV-positive patients in this population were conducted by Pelsler *et al.* [2010] and Anderson *et al.* [2008]) and are discussed under cofactors in Section 3.2.5, below.) Cases that fall into more than one category of Kaposi sarcoma were sometimes combined by study authors for analysis. Comparison groups may be human immunodeficiency virus type 1-positive or AIDS patients, or blood donors, oncology patients, or patients with other skin diseases or other non-cancer conditions, or combinations of such comparison groups from which a single risk estimate was calculated by authors or IARC working groups.

Among the studies of patients with human immunodeficiency virus type 1 (some of which had AIDS), odds ratios (ORs), almost all statistically significant, ranged from 1.0 to 1,683 (Table 3-2). The two studies of human immunodeficiency virus type 1-negative cases and controls, both conducted in African populations with, as noted above, high endemic KSHV prevalence rates, reported ORs of 1.5 to 12.0, depending on the level of KSHV titer (Newton *et al.* 2003b, Sitas *et al.* 1999). The studies designated as grouped populations consist of combinations of human immunodeficiency virus type 1-positive, human immunodeficiency virus type 1-negative or undetermined, endemic, classic, or transplant-related Kaposi sarcoma patients in comparison with similar Kaposi sarcoma-negative groups, as noted in Table 3-2, and for which only single risk estimates were reported, ORs ranging from 3.8 to 924 were observed. ORs could also be calculated for some constituent groups within these studies, although in some instances, only a single combined control group was used and/or no formal matching of controls was conducted, so that such risk estimates must be interpreted with caution. Nevertheless, for both classic and endemic Kaposi sarcoma, KSHV was observed in > 80% to 100% of cases (Table 3-2). Among the few studies that included transplant patients with Kaposi sarcoma, KSHV was observed in approximately 66% to 100% of cases.

Three studies reviewed by IARC (2012) reported on dose-response relationships between titers of KSHV exposure and risk for Kaposi sarcoma. Sitas *et al.* (1999) and Newton *et al.* (2003b) observed statistically significant trends ( $P < 0.001$ ) among both human immunodeficiency virus type 1-positive and human immunodeficiency virus type 1-negative groups with a higher prevalence of endemic Kaposi sarcoma in Africa; Albrecht *et al.* (2004) reported a similar trend among human immunodeficiency virus type 1-positive AIDS patients in Germany.

**Table 3-2. Summary of case-control studies of Kaposi sarcoma and Kaposi sarcoma-associated herpesvirus (KSHV)**

| Reference   | Detection method   | OR <sup>a</sup>              | Comments                                    |
|---|--------------------|------------------------------|---|
| <b>HIV-1-positive/AIDS populations</b>                          |                    |                              |   |
| Chang <i>et al.</i> 1994 (USA)                                  | PCR                | 68.8                         |   |
| Whitby <i>et al.</i> 1995 (UK)                                  | PCR                | 29.0                         |   |
| Monini <i>et al.</i> 1996b, Monini <i>et al.</i> 1996a (Italy)  | PCR                | 1.7 <sup>a</sup>             |   |
| Parry and Moore 1997 (USA)                                      | PCR                | 7.3 MSM<br>21.8 hemophiliacs |   |
| Humphrey <i>et al.</i> 1996 (USA)                               | PCR                | 2.3                          |   |
| Decker <i>et al.</i> 1996 (USA)                                 | PCR                | 1.1 <sup>b</sup>             |   |
| Gao <i>et al.</i> 1996 (USA)                                    | LANA, WB           | 18.9                         |   |
| Lefrere <i>et al.</i> 1996 (France)                             | PCR                | 285.0                        |   |
| Miller <i>et al.</i> 1996 (USA)                                 | WB, IFA            | 13.4 (WB)/12.2 (IFA)         |   |
| Noel 1995 (Belgium)   | PCR                | 42.2                         |   |
| Howard <i>et al.</i> 1997 (UK)                                  | PCR                | 10.1                         |   |
| Sitas <i>et al.</i> 1999 (South Africa)                         | LANA, IFA          | 10.8–1683                    | Sig. ↑ inc. KSHV titer                      |
| Min and Katzenstein 1999 (USA)                                  | PCR                | 16.3                         |   |
| Boivin <i>et al.</i> 1999 (Canada)                              | PCR                | 6.7                          |   |
| Greenblatt <i>et al.</i> 2001 (USA)                             | PCR                | 148.0                        |   |
| Keller <i>et al.</i> 2001 (Brazil)                              | PCR                | 7.4–17.0                     |   |
| Cannon <i>et al.</i> 2003 (USA)                                 | PCR                | 8.6                          |   |
| Newton <i>et al.</i> 2003b (Uganda)                             | LANA, IFA          | 4.1–0.8                      | Sig. ↑ inc. KSHV titer                      |
| Alagiozoglou <i>et al.</i> 2003 (South Africa)                  | LANA, IFA, PCR     | 15.4–25.1                    |   |
| Albrecht <i>et al.</i> 2004 (Germany)                           | IFA                | 1.04–23.2                    | Sig. ↑ inc. KSHV titer                      |
| Szalai <i>et al.</i> 2005 (Hungary)                             | LANA, IFA, ELISA   | 2.3 <sup>b</sup> –23.1       |   |
| van der Kuyl <i>et al.</i> 2005 (Netherlands)                   | PCR                | 6.3                          |   |
| Martro <i>et al.</i> 2007 (Spain)                               | IFA                | 4.8                          |   |
| Widmer <i>et al.</i> 2006 (Switzerland)                         | PCR                | 1.2 <sup>b</sup>             |   |
| Laney <i>et al.</i> 2007 (USA)                                  | PCR                | 1.6 <sup>b</sup> –3.4        |   |
| Zago <i>et al.</i> 2000 (Brazil)                                | PCR                | 23.4–104.8                   |   |
| <b>HIV-1-negative populations</b>                               |                    |                              |   |
| Sitas <i>et al.</i> 1999 (South Africa)                         | LANA, IFA          | 1.5–12.0                     | Endemic—sign. trend with ↑ KSHV titer       |
| Newton <i>et al.</i> 2003b, Newton <i>et al.</i> 2003a (Uganda) | LANA, IFA          | 2.3–7.1                      | Endemic—sign. trend with ↑ KSHV titer       |
| Cattani <i>et al.</i> 2001 (Italy)                              | IFA                | 34.4                         | Transplant patients                         |
| Amodio <i>et al.</i> 2011a (Sicily)                             | LANA, lytic (K8.1) | 5.13 <sup>b</sup>            | Classic                                     |
| <b>Grouped populations</b>                                      |                    |                              | <b>Case groups</b>                          |
| Moore and Chang 1995 (USA)                                      | PCR                | 400 (19–17,300)              | Total (HIV-1+ men, HIV-1-neg. MSM, Classic) |
|   |                    | [210 (12–3,712)]             | HIV-1+                                      |
|   |                    | [∞]                          | HIV-1-neg. MSM                              |
|   |                    | [∞]                          | Classic                                     |

| Reference                                | Detection method | OR <sup>a</sup>  | Comments   |
|--|------------------|--|--|
| Chang <i>et al.</i> 1996 (USA)           | PCR              | 49.4 (9.0–328.0)<br>[66 (5.1–857.7)]<br>[36 (5.3–253.6)]                               | HIV-1+, endemic<br>HIV-1 +<br>Endemic  |
| Luppi <i>et al.</i> 1996 (Italy)         | PCR              | 41.6 (7.0–327.1)<br>[∞]<br>[∞]   | HIV-1+, classic<br>HIV-1+<br>Classic   |
| Albini <i>et al.</i> 1996 (Italy)        | PCR              | 924.3 (125–10,194)<br><br>[∞]<br>[∞]<br>[∞]<br>[198 (10.8–3,617)]                      | Total: HIV-1+,<br>endemic, classic,<br>transplant<br>AIDS vs. HIV-1+<br>Endemic vs. HIV-1+<br>Classic vs. HIV-1+<br>Transplant vs. HIV-1<br>neg. |
| Herman <i>et al.</i> 1998 (USA)          | PCR              | 336 (14.9–47,561)  | HIV-1+, classic (vs.<br>HIV-1+ and HIV-1<br>neg. controls<br>combined)   |
| Cattani <i>et al.</i> 1998 (Italy)       | PCR              | 7.4 (2.4–23.8)<br>[5.9 (1.8–19.2)]<br>[20.0 (1.7–238.6)]                               | Classic, transplant<br>Classic<br>Transplant   |
| Kazakov <i>et al.</i> 2002 (Switzerland) | PCR              | 543 (52–14,041)<br>[600 (34.2–10,541.2)]<br>[150 (11.1–2,025)]<br>1,275 (75.9–21,420)] | Total: HIV-1+,<br>transplant, classic<br>HIV-1+<br>Transplant<br>Classic   |
| Lager <i>et al.</i> 2003 (South Africa)  | PCR              | 572 (26–79,693)  | HIV-1 status unknown   |
| Massambu <i>et al.</i> 2003 (Tanzania)   | PCR              | 3.8 (0.3–60.9)   | HIV-1+, endemic  |

Source: IARC 2012, [Table 2.1](#), Amodio *et al.* 2011a.

[ ] = OR calculated by NTP.

CI = 95% confidence interval; ELISA = enzyme-linked immunosorbent assay; HIV-1 = human immunodeficiency virus type 1; IFA = immunofluorescence assay; KS = Kaposi sarcoma; KSHV = Kaposi sarcoma-associated herpesvirus; LANA = latent antinuclear antigen; MSM = men who have sex with men; OR = odds ratio; PCR = polymerase chain reaction; WB = Western blot assay.

<sup>a</sup>Lower 95% CI = 0.0.

<sup>b</sup>Lower 95% CI < 1.0.

### 3.3.5 Cofactors for Kaposi sarcoma

KSHV has been identified in over 90% (including all four types) of Kaposi sarcoma lesions (IARC 2012, Mesri *et al.* 2010, Dedicoat and Newton 2003, Moore and Chang 1998). The principal cofactor identified for Kaposi sarcoma in the presence of KSHV infection is human immunodeficiency virus type 1 infection via immune suppression, among populations in both the United States and Europe (“epidemic” Kaposi sarcoma) and those with an increased prevalence of “classic” or “endemic” Kaposi sarcoma (Tables 3-1 and 3-2). Prospective studies, largely

among men who have sex with men and infrequently among human immunodeficiency virus type 1-positive intravenous drug users or women, suggest that human immunodeficiency virus type 1 infection significantly increases the risk of Kaposi sarcoma development in the presence of KSHV in a range of geographical populations, particularly among AIDS patients. There is additional evidence that infection with human immunodeficiency virus type 1 prior to infection with KSHV may increase the risk of Kaposi sarcoma by up to 2.6-fold compared with those infected with KSHV before human immunodeficiency virus type 1, and increasing duration of human immunodeficiency virus type 1 infection, and decreasing CD4 counts are associated with a more rapid course of development of the cancer (Nawar *et al.* 2008, Jacobson *et al.* 2000, Renwick *et al.* 1998). (See also accompanying human immunodeficiency virus type 1 monograph.)

Immune suppression among organ recipients, primarily renal and liver transplant patients, has also been identified as a major cofactor among patients who were KSHV positive prior to transplantation or who acquired KSHV during organ transplantation (see Table 3-1). In a review of 38 case-series studies by Hosseini-Moghaddam *et al.* (2012), approximately 13% of previously KSHV-seropositive patients reportedly developed Kaposi sarcoma after renal transplantation; approximately 5% of the patients developed the cancer as a result of primary KSHV (donor KSHV positive, recipient KSHV negative) infection.

Few studies have investigated the role of recreational drugs with immunomodulatory properties. In one prospective study, prior use of marijuana, amyl nitrate “poppers,” cocaine, or amphetamines was weakly associated with Kaposi sarcoma risk among co-KSHV and human immunodeficiency virus type 1-infected men who have sex with men, although a clear dose-response relationship was lacking (Chao *et al.* 2009).

The role of co-factors is less clear for the endemic and classic forms of Kaposi sarcoma. Never smoking, diabetes, and the use of oral corticosteroids have all been identified as potential risk factors for classic Kaposi sarcoma among KSHV-positive individuals in the absence of human immunodeficiency virus type 1 infection or iatrogenic or other causes of immune suppression in a well-conducted case-control study of a population in Sicily (Amodio *et al.* 2011a, Anderson *et al.* 2008, Goedert *et al.* 2002). While this study provides limited evidence that these factors increase the risk of Kaposi sarcoma among KSHV-positive individuals, few other data in other populations include examination of these risk factors and no firm conclusions can be drawn about their role, or that of other risk factors.

With respect to co-infection with other viruses, the role of Epstein-Barr virus is uncertain. It appears that the prevalence of Epstein-Barr virus infection is similar in Kaposi sarcoma and non-Kaposi sarcoma patients and does not appear to play a direct role in the development of the sarcoma, at least according to one well-conducted case-control study among classic cases of Kaposi sarcoma (Table 3-3) (Pelser *et al.* 2010). Other viruses, such as herpes simplex type 2, human cytomegalovirus, human herpesvirus-6 and 7, and human papillomavirus have also been implicated in the activating lytic replication of KSHV and thus in Kaposi sarcoma pathogenesis (reviewed by Purushothaman *et al.* 2015). However, no cohort or case-control studies specifically addressing the risk of Kaposi sarcoma in the presence of these viruses as cofactors among KSHV-positive populations have been identified to date.

Among populations with high rates of endemic Kaposi sarcoma, particularly prior to the global human immunodeficiency virus type 1 epidemic, other potential cofactors, including malaria (see e.g., Nalwoga *et al.* 2015), other parasitic infections, soil types causing immunosuppression, or oncogenic plants have been suggested as increasing the risk of KSHV infection and/or reactivation (IARC 2012, Dedicoat and Newton 2003). Tuberculosis has been reported as exacerbating the severity of endemic cases (e.g., in Tanzania, Koski *et al.* [2014]) but it is not clear if it is a risk factor for the initial development of the sarcoma. Finally, prior to the human immunodeficiency virus type 1 epidemic, Kaposi sarcoma was more prevalent among men than women in, e.g., populations with endemic cases, despite similar KSHV seroprevalence in both sexes, suggesting that cofactors not yet clearly established might play a role in the development of the cancer (Dedicoat and Newton 2003).

**Table 3-3. Case-control study of co-factors for the development of classic Kaposi sarcoma among a KSHV-positive population in Sicily**

| Reference                   | Cofactor                | # KS Cases//KSHV+controls   | OR (95% CI) for KS            |
|-----------------------------|-------------------------|---|-------------------------------|
| Anderson <i>et al.</i> 2008 | Diabetes                | Cases (45/142)<br>Controls (15/123)                               | 4.02 (1.73–9.37)              |
|                             | Never smoking           | Cases (23/142)<br>Controls (12/123)                               | 2.66 (1.06–6.64)              |
|                             | Oral corticosteroid use | Cases (53/142)<br>Controls (26/123)                               | 2.25 (1.16–4.38)              |
| Pelser <i>et al.</i> 2010   | EBV                     | Cases (low EBNA-1) (43/119)<br>Controls (low EBNA-1) (34/105)     | 0.97 (0.52–1.82) <sup>a</sup> |
|                             |                         | Cases (high EBV VCA) (45/119)<br>Controls (high EBV VCA) (45/105) | 0.87 (0.45–1.69) <sup>a</sup> |

CI = confidence interval; EBNA = Epstein-Barr virus nuclear antigen; EBV = Epstein-Barr virus; EBV VCA = EBV viral capsid antigen; KS = Kaposi sarcoma; KSHV = Kaposi sarcoma-associated herpesvirus; OR = odds ratio.

<sup>a</sup>Adjusted for age, sex, smoking, diabetes, and oral corticosteroid use.

### 3.3.6 Integration across studies of Kaposi sarcoma

At least 25 cohort or nested case-control studies and 81 case-control studies conducted in populations differing by race or ethnicity and in various geographical locations have demonstrated that KSHV infection causes Kaposi sarcoma. KSHV has been identified in over 80% of cases by serology and in virtually all Kaposi sarcoma tumors, and is considered to be the diagnostic prerequisite for a diagnosis of Kaposi sarcoma. The histopathology of all four epidemiological types of Kaposi sarcoma (epidemic, iatrogenic, classic, and endemic) is identical. Case-control studies conducted among populations exhibiting the different types showed statistically significant increased risks for Kaposi sarcoma generally exceeding 10 and up to 300 or more (using serology and/or DNA in the tumor to identify KSHV infection). Most cohort studies report risks from approximately 2 to 16 although a few studies, primarily among transplant patients, have reported risks up to 75. In addition, statistically significant dose-response relationships for measures of KSHV infection and Kaposi sarcoma risk among both human immunodeficiency virus type 1-positive and/or human immunodeficiency virus type 1-

negative populations have been reported (Newton *et al.* 2006, Albrecht *et al.* 2004, Newton *et al.* 2003b, Newton *et al.* 2003a, Sitas *et al.* 1999).

The heterogeneity in risk observed particularly in case-control studies may be at least partly due to methodological limitations. A majority of the case-control or case-comparison studies have small numbers of cases, and lack formal methods of selection or matching of controls, or have inadequate reporting. In addition, temporality (evidence that KSHV infection preceded the development of Kaposi sarcoma) cannot generally be clearly established in these studies. The majority of cohort or nested case-control studies have somewhat limited statistical power, and the sensitivity and specificity of KSHV detection methods in earlier studies is generally more limited than in later studies. However, several prospective cohort studies and nested case-control studies (e.g., Whitby *et al.* 1995, Gao *et al.* 1996, Moore *et al.* 1996, Nawar *et al.* 2008, Labo *et al.* 2015, Wakeham *et al.* 2015) provide evidence of temporality. Overall, confounding by other factors can be ruled out with confidence because of the findings of highly statistically significant increased risks across studies with different designs and conducted in different populations, and evidence of dose-response relationships.

With respect to cofactors, immune suppression—primarily resulting from human immunodeficiency virus type 1 infection or iatrogenically in, for example, transplant cases—is the principal cofactor. human immunodeficiency virus type 1 infection prior to KSHV infection may increase the risk of epidemic Kaposi sarcoma between 50% and 100% compared with human immunodeficiency virus type 1 infection during or after KSHV infection. While a recent, well-conducted case-control study in Sicily (Anderson *et al.* 2008) provides limited evidence that diabetes, corticosteroid use, and never smoking increase the risk of Kaposi sarcoma among KSHV-positive individuals, few other data in other populations have included examination of these risk factors and no firm conclusions can be drawn. Clear evidence for a modulatory role of other specific cofactors, such as genetic factors, malaria, hypoxic stress, or environmental and dietary factors, particularly among endemic and classic cases, is lacking. Co-infection with other viruses, such as Epstein-Barr virus, cannot be completely ruled out as cofactors in the development of Kaposi sarcoma in KSHV-infected individuals; although one case-control study (Pelser *et al.* 2010) suggests no association with co-infection with Epstein-Barr virus, there are few other data to evaluate the role of this or other viruses.

### **3.4 Lymphohematopoietic cancers**

#### **3.4.1 Primary effusion lymphoma**

##### **Background information**

Primary effusion lymphoma, a monoclonal lymphoproliferative disorder also called body cavity-based B-cell lymphoma, is a rare type of B-cell non-Hodgkin lymphoma that presents primarily as pleural, peritoneal, or pericardial lymphomatous effusions and makes up approximately 2% to 4% of human immunodeficiency virus type 1-related non-Hodgkin lymphomas (Sullivan *et al.* 2008, Simonelli *et al.* 2003). In addition to lymphomatous effusions, diagnosis of primary effusion lymphoma includes KSHV infection; large-cell immunoblastic or anaplastic large-cell lymphomas; expression of CD45, clonal immunoglobulin gene rearrangements; and lack of *c-myc*, *bcl-2*, *ras*, and *p53* gene alterations (Ablashi *et al.* 2002, Nador *et al.* 1996). Primary effusion lymphomas are thought to arise from post-germinal center B cells because they have

mutations in their immunoglobulin genes. Because a large number of cases of primary effusion lymphoma identified in association with KSHV have distinctive morphological, immunophenotypical, and molecular genetic characteristics, they have been grouped as a single distinct pathological entity (IARC 2012, 1997). Such entities should be distinguished from other large body cavity-based B-cell lymphomas or lymphoproliferative disorders observed in KSHV-negative patients and may also have a different prognosis (see e.g., Xiao *et al.* 2013, Chen *et al.* 2009, Kishimoto *et al.* 2009).

### **Epidemiological studies**

Epidemiological evidence of an association between primary effusion lymphoma and KSHV relies primarily on a large number of case reports, case-series, and case-comparison studies rather than formal case-control or cohort studies, due to the rarity of the condition, and the fact that the presence of KSHV is a diagnostic criterion for the specific pathological entity that constitutes primary effusion lymphoma. KSHV-positive cases of primary effusion lymphoma are occur among human immunodeficiency virus type 1 positive and human immunodeficiency virus type 1 negative individuals approximately half of the cases have Kaposi sarcoma. Primary effusion lymphoma among human immunodeficiency virus type 1-positive patients exhibits a rapid progression with short survival times, whereas in human immunodeficiency virus type 1-negative and immunocompetent patients, progression appears to be much slower (IARC 1997). The large majority of primary effusion lymphoma cases occur among men, irrespective of human immunodeficiency virus type 1 or Kaposi sarcoma status, presumably reflecting a higher prevalence of KSHV infection among men (IARC 2012).

IARC (2012, [Table 2.5](#)) reviewed 97 individual cases reported in the literature to be primary effusion lymphoma; 44 cases were reported as both KSHV and human immunodeficiency virus type 1 positive, 31 cases were KSHV positive but human immunodeficiency virus type 1 negative, and three case were uncertain as to KSHV (N = 2)) or human immunodeficiency virus type 1 status (N = 1). The remaining 19 cases were KSHV-negative and human immunodeficiency virus type 1-negative, and associated with genetic or other pre-existing conditions and/or observed among patients 67 years of age or older; however, IARC noted that the KSHV-negative cases probably represented another form of non-Hodgkin lymphoma because they do not meet the diagnostic criteria for primary effusion lymphoma. Several small case-series studies (Oksenhendler *et al.* 2002, Aoki *et al.* 2001, Judde *et al.* 2000, Parravicini *et al.* 2000) were identified that reported a total of 12 additional (i.e., not included in the 97 cases reviewed by IARC) KSHV-positive cases of primary effusion lymphoma, 10 of which were from human immunodeficiency virus type 1-positive/AIDS patients.

KSHV-positive, human immunodeficiency virus type 1-negative cases (11 of the 31 cases reported by IARC and one additional case) have been observed among organ transplant patients (IARC 2012, Testa *et al.* 2010, Boulanger *et al.* 2008, Dotti *et al.* 1999) and among populations (including ~14 reviewed by IARC and 8 additional cases) that are known to have a high risk of endemic or classic epidemiological Kaposi sarcoma, leading to the proposition that a “classic” form of primary effusion lymphoma can be identified (Yiakoumis *et al.* 2010, reviewed by Ascoli *et al.* 2002).

Among 47 KSHV-positive cases of primary effusion lymphoma with known human immunodeficiency virus type 1 status (reported by IARC), approximately half of the cases in

both human immunodeficiency virus type 1-positive (14/30) and human immunodeficiency virus type 1-negative patients (8/17) also had Kaposi sarcoma; some also presented with multicentric Castleman disease (reviewed below).

In a large case-comparison study of 191 patients with a range of lymphoproliferative disorders (Asou *et al.* 2000), 21 patients were identified with primary effusion lymphoma, all of whom were KSHV DNA positive, in contrast to 1 of 170 patients with AIDS-related non-Hodgkin lymphoma or with non-AIDS-related lymphomas (Castleman disease, reactive lymphadenopathy, mucosa-associated lymphoid tissue lymphoma, and non-AIDS non-Hodgkin lymphoma). (In this study, the human immunodeficiency virus type 1 status of primary effusion lymphoma cases is unclear.) No additional cohort or case-control studies have been identified (see review by Zhang *et al.* 2010).

### **Cofactors for primary effusion lymphoma**

Immune suppression resulting from human immunodeficiency virus type 1 infection is a cofactor for primary effusion lymphoma development among KSHV-positive cases. Due in part to its rarity, there are few data on primary effusion lymphoma cases among other immunosuppressed groups although one study reviewed by IARC (2012), Kapelushnik *et al.* (2001), reported a case in a KSHV-positive renal transplant patient. However, no other case studies were identified for the current review. A substantial proportion (50%) of KSHV-related primary effusion lymphoma cases also have evidence of Epstein-Barr virus infection (IARC 2012, 1997), which has also been postulated as a cofactor (Gloghini *et al.* 2013, Boulanger *et al.* 2005, Bryant-Greenwood *et al.* 2003).

### **3.4.2 Multicentric Castleman disease**

#### **Background and epidemiological evidence**

Multicentric Castleman disease is a subset (20%) (Talat and Schulte 2011) of Castleman disease, first identified by Castleman (1956) and characterized inflammatory symptoms, plasmacytic lymphadenopathy, splenomegaly, and cytopenia with polyclonal hyperimmunoglobulinemia and high serum levels of IL-6 (Oksenhendler *et al.* 2013, Fukumoto *et al.* 2011), and has an aggressive course and poor prognosis (IARC 1997).

Evidence of a positive association between multicentric Castleman disease and KSHV has been reported in several early case reports and case series (Barozzi *et al.* 1996, Corbellino *et al.* 1996, Gessain *et al.* 1996, and Soulier *et al.* 1995); and in four case-comparison studies (63 cases of multicentric Castleman disease and 213 controls) (Bélec *et al.* 1999, Parravicini *et al.* 1997a, Soulier *et al.* 1995, and Asou *et al.* 2000) reviewed by IARC (2012). These studies reported a higher frequency of KSHV, primarily in human immunodeficiency virus type 1-positive but infrequently in human immunodeficiency virus type 1-negative groups, in comparison with controls (Table 3-4). Several histological variants of multicentric Castleman disease exist (hyaline vascular, plasma cell, mixed, and plasmablastic) but KSHV in individuals with and without human immunodeficiency virus type 1 infection, is almost always associated with the plasmablastic variant and rarely in the hyaline vascular form of the disease (Fukumoto *et al.* 2011, Dupin *et al.* 1999, 2000, Parravicini *et al.* 2000, Ascoli *et al.* 1999, Damania 2010). Dupin *et al.* 2000 reported that KSHV was specifically associated with a plasmablastic variant of



multicentric Castleman disease in which KSHV-positive plasmablasts showing  $\gamma$  light-chain restriction localized in the mantle zone of B-cell follicles (Dupin *et al.* 2000). In some cases, these plasmablasts coalesce to form foci of microlymphoma. These plasmablasts are not present in KSHV-negative multicentric Castleman disease (see Section 3.5 for a discussion of the two different forms of multicentric Castleman disease). Findings described in Table 3-4 illustrate the preponderance of plasmablastic types.

While rare, the incidence and prevalence of KSHV-multicentric Castleman disease across populations are uncertain, as it is not tracked in cancer registries and is likely to be under-diagnosed due to its irregular course. Among 32 cases of MCD followed at the NIH Clinical Center in Bethesda, MD, five were in recent African immigrants (Uldrick *et al.* 2012); Gopal *et al.* (2014) reported a single KSHV-multicentric Castleman disease case in Malawi.

**Table 3-4. Summary of case-series and case-control/comparison studies of KSHV and multicentric Castleman disease**

| Reference                                | KSHV detection method | HIV-1 status of MCD cases/controls        | KSHV-exposed cases/total cases or OR (95% CI) <sup>a</sup> exposed cases vs. exposed controls/ |
|--|-----------------------|---|--|
| <b>Case reports and case series</b>      |                       |   |  |
| Corbellino <i>et al.</i> 1996            | PCR, Southern blot    | HIV-1 negative                            | 4/6 (4/4 with plasma cell type)  |
| Barozzi <i>et al.</i> 1996               | PCR                   | HIV-1 negative                            | 1/16 (1/1 is hyaline vascular type)  |
| Gessain <i>et al.</i> 1996               | PCR                   | HIV-1 positive<br>HIV-1 negative          | 3/4 (mixed cell type) <sup>c</sup><br>1/6 (plasma cell type)                                   |
| Soulier <i>et al.</i> 1995               | PCR, Southern blot    | HIV-1 positive<br>HIV-1 negative          | 14/14 (6/14 plasma type)<br>7/17 (3/7 plasma type)   |
| Ascoli <i>et al.</i> 1999                | PCR                   | HIV-1 positive<br>HIV-1 negative          | 1/1 (type not mentioned)<br>1/1 (type not mentioned)   |
| Gopal <i>et al.</i> 2014                 | LANA/viral load       | HIV-1 positive                            | 1/1 (plasmablastic variant)  |
| Parravicini <i>et al.</i> 2000           | Immuno-histochemistry | HIV-1 positive<br>HIV-1 negative          | 4/4 (plasma cell type)<br>6/6 (plasma cell type)   |
| Dupin <i>et al.</i> 1999                 | Immuno-histochemistry | HIV-1 positive<br>HIV-1 negative          | 5/5 (plasmablastic)<br>0/1 <sup>b</sup>  |
| Dupin <i>et al.</i> 2000                 | PCR                   | HIV-1 positive<br>HIV-1 negative          | 8/8 (plasmablastic)<br>2/2 (plasmablastic)   |
| <b>Case-comparison studies</b>           |                       |   |  |
| Belec <i>et al.</i> 1999<br>France       | IFA antibodies        | 8 cases, 15 controls                      | $[\infty (6/8 \text{ vs. } 0/15)]^b$   |
| Parravicini <i>et al.</i> 1997b<br>Italy | PCR                   | All HIV-1 negative                        | $[\infty (6/14 \text{ vs. } 0/25)]^c$  |
| Soulier <i>et al.</i> 1995<br>France     | PCR                   | HIV-1 positive<br>HIV-1 negative<br>Total | 14/14 vs. 1/34<br>7/17<br>$[30.2.(3.8-237.9)][23.0 (2.9 - 181.5)]$                             |

| Reference                        | KSHV detection method | HIV-1 status of MCD cases/controls | KSHV-exposed cases/total cases or OR (95% CI) <sup>a</sup> exposed cases vs. exposed controls/ (21/31 vs. 1/34) <sup>c, d</sup> |
|----------------------------------|-----------------------|------------------------------------|---|
| Asou <i>et al.</i> 2000<br>Japan | PCR                   | Unclear                            | [∞ (1/10 vs. 0/139)] <sup>b</sup>   |

Source: IARC 2012, [Table 2.6](#) and [Table 2.7](#).

CI = confidence interval; IFA = immunofluorescence assay; HIV-1 = human immunodeficiency virus type 1; OR = odds ratio; PCR = polymerase chain reaction.

<sup>a</sup>[ ] ORs calculated by NTP, <sup>b</sup>cell type not specified; <sup>c</sup>plasma cell or mixed type; <sup>d</sup>RoC calculation based on 2x2 table

### Cofactors for multicentric Castleman disease

Human immunodeficiency virus type 1 co-infection substantially increases the risk of multicentric Castleman disease among KSHV-positive patients (Table 3-5; also see Section 4), and KSHV appears to be found in almost all cases of human immunodeficiency virus type 1-related multicentric Castleman disease and approximately half of human immunodeficiency virus type 1-negative cases (Oksenhendler *et al.* 2013, Oksenhendler *et al.* 2002, Parravicini *et al.* 1997b).

Based on 24 cases of human immunodeficiency virus type 1-associated MCD, Powles *et al.* (2009) measured the incidence of MCD as 4.3/10 000 patient-years (95% CI 2.7 to 6.4). The incidence increased significantly from 2.3 (95% CI = 0.02 to 4.2) in the pre-HAART era to 8.3 (95% CI = 4.6 to 12.6) in the post-HAART era. A nadir CD4 count > 200/mm<sup>3</sup> increased age, no previous HAART exposure and non-Caucasian ethnicity were all associated with an increased risk of multicentric Castleman disease. In a U.K. study of 41 human immunodeficiency virus type 1-positive patients with KSHV-related Kaposi sarcoma or multicentric Castleman disease, Westrop *et al.* (2012) reported that KSHV-infected human immunodeficiency virus type 1-positive people of African ancestry were significantly more likely to develop multicentric Castleman disease than their European counterparts (relative risk [RR] = 2.42, 95% CI = 1.28 to 4.55). An analysis of the frequency of A299G, a toll-like receptor 4 (TLR4) single nucleotide polymorphism associated with increased multicentric Castleman disease incidence, showed the polymorphism to be three-fold higher in patients of native African descent compared to those of European descent. Finally, the role of Epstein-Barr virus in KSHV-associated multicentric Castleman disease has not been extensively investigated, but the mantle zone plasmablastic cells predominantly infected with KSHV do not appear to be co-infected with Epstein-Barr virus (Oksenhendler *et al.* 2002, Dupin *et al.* 2000).

#### 3.4.3 Multiple myeloma

Multiple myeloma is a comparatively rare type of lymphohematopoietic cancer, making up less than 2% of incident cancers in the United States. The annual (2008 to 2011) U.S. incidence and death rate has been stable at approximately 6.3/100,000 in men and 3.3/100,000 in women with an approximately 47% 5-year survival rate (SEER 2015).

Twenty hospital-based case-control/comparison studies and two nested case-control studies of the association between multiple myeloma and KSHV were reviewed by IARC (2012). One large case-control study, including 254 cases of multiple myeloma, has been identified since

2008 (Benavente *et al.* 2011), together with two case series of multiple myeloma patients that reported a higher frequency of KSHV in bone marrow tissue (Ismail *et al.* 2011, Sadeghian *et al.* 2008) than among healthy controls. Excluding five studies using a case-control or case-comparison design (Benavente *et al.* 2011, Sitas *et al.* 1999, Schonrich *et al.* 1998) or nested case-control design (Tedeschi *et al.* 2005, Tedeschi *et al.* 2001) with age-matched controls (see Section 3.7 and Table 3-6), the majority of studies were clinic based and did not report methods for the selection of cases and controls or comparison groups and few appear to have controlled for age, sex, or other variables. Among these lower quality studies, a total of 118 cases of multiple myeloma were associated with KSHV (22%) compared with 233 of 1,524 patients without multiple myeloma (23%) in 17 of 18 studies where rates among controls were reported. The five case-control or nested case-control studies also do not support a positive association between KSHV infection and multiple myeloma risk (Table 3-5). In the nested case-control studies (Tedeschi *et al.* 2005, Tedeschi *et al.* 2001), increased risks were observed for KSHV detected by latent antibodies; however, IARC (2012) reported that after stratifying by detection method, no statistically significant differences in KSHV positivity between cases and controls were observed by the Working Group.

#### 3.4.4 Other lymphohematopoietic cancers

Several case-series and case-control studies have been conducted of other lymphohematopoietic cancers, including other lymphomas. An increased incidence of non-Hodgkin lymphoma has been reported in association with KSHV-related multicentric Castleman disease or primary effusion lymphoma in some patients (Oksenhendler *et al.* 2013, 2002). Pan *et al.* (2012) recently described a rare KSHV-associated large B-cell lymphoma in lymph nodes or extranodal sites that do not exhibit lymphomatous effusions and appear to have a different immunological profile but which may form a continuum with classic primary effusion lymphoma. Deloose *et al.* (2005) also reported that 38% of human immunodeficiency virus type 1-positive cases of solid immunoblastic/non-Hodgkin lymphoma were KSHV positive among 99 cases of non-Hodgkin lymphoma without evidence of primary effusion lymphoma or multicentric Castleman disease. Burbelo *et al.* (2012) also reported a higher frequency of KSHV among a small series of human immunodeficiency virus type 1-positive non-Hodgkin lymphoma cases than among healthy controls, but no differences in KSHV frequency were reported in a series of human immunodeficiency virus type 1-positive non-Hodgkin lymphoma cases compared according to duration of human immunodeficiency virus type 1 immunosuppression (Gerard *et al.* 2009).

**Table 3-5. Summary of five case-control/comparison and nested case-control studies of KSHV and multiple myeloma<sup>a</sup>**

| Reference Study type   | KSHV detection method   | HIV-1 status   | OR or RR (95% CI) (KSHV-exposed cases vs. exposed controls)   |
|--|---|----------------|---|
| Schonrich <i>et al.</i> 1998<br>Germany<br>Case-comparison                           | IFA lytic KS-1  | HIV-1 negative | [1.39 (0.15–13.3)]<br>(4/99 cases vs. 1/34 healthy controls <sup>b</sup> )  |
| Sitas <i>et al.</i> 1999<br>S Africa<br>Combined 2 case-control studies              | IFA latent<br>Titer<br>1:100<br>1:> 20,000                        | Not specified  | Adjusted prevalence rate <sup>c</sup> (108 cases and 108 controls)<br>24% cases vs. 32% blood-donor controls<br>1.5% cases vs. 3% blood-donor controls                |
| Tedeschi <i>et al.</i> 2001<br>Finland<br>Nested case-control study                  | IFA lytic<br>Latent (LANA)<br>Western blot (ORF 65, 73, or K8.1A) | Not specified  | 2.02 (0.94–4.33)<br>(13/47 cases vs. 36/224 controls)<br>10.0 (0.91–110.3)<br>(2/47 cases vs. 1/224 controls)<br>0.89 (0.25–3.25)<br>(3/47 cases vs. 16/224 controls) |
| Tedeschi <i>et al.</i> 2005<br>Scandinavia<br>5 cohorts, nested case-control studies | IFA lytic<br>IFA latent   | Not specified  | 1.08 (0.5–1.1)<br>(39/329 cases vs. 238/1,393 controls <sup>d</sup> )<br>0.6 (0.1–2.7)<br>(2/329 cases vs. 16/1,615 controls <sup>d</sup> )                           |
| Benavente <i>et al.</i> 2011<br>Europe<br>EpiLymph<br>multisite case-control study   | Latent (ORF 73 antigen)<br>Lytic (K8.1)                           | Not specified  | 0.31 (0.11–0.85)<br>(4/254 cases; NR controls <sup>e</sup> )<br>0.43 (0.15–1.2)<br>(4 cases; NR controls <sup>e</sup> )   |

Source: IARC 2012, [Table 2.8](#) and [Table 2.9](#) except for Benavente *et al.* (2011); [ ] = OR calculated by NTP.

CI = confidence interval, IFA = immunofluorescence assay; LANA = latent nuclear antigen; KS = Kaposi sarcoma; KSHV = Kaposi sarcoma-associated herpesvirus; MM = multiple myeloma, NR = not reported, OR = odds ratio, RR = relative risk.

<sup>a</sup>Studies with unclear selection of cases or controls or no adjustment for age or other variables were excluded.

<sup>b</sup>Age- and sex-matched controls.

<sup>c</sup>Adjusted for age, sex, and education and sex partner.

<sup>d</sup>Age-, sex-, and area-matched controls; OR adjusted for age.

<sup>e</sup>Age-, sex-, sampling date-, length of follow-up- and area-matched controls.

A nested case-control study of 155 incident cases of non-Hodgkin lymphoma among human immunodeficiency virus type 1-positive men who have sex with men vs. 155 non-cancer controls (Beachler *et al.* 2011) did not detect a difference in serum KSHV after adjustment for age and CD4 status. The multisite EpiLymph study in Europe (Benavente *et al.* 2011), comprising 2,083 cases of lymphoma and 2,048 age-, sex-, and country-matched controls with KSHV serotyping, also reported no associations between KSHV seropositivity and B-cell lymphoma, T-cell lymphoma or Hodgkin lymphoma. Positive associations for splenic marginal zone lymphoma (OR = 4.1, 95% CI = 1.57 to 10.83; 6 cases, using the lytic K8.1 antigen) and mantle-cell lymphoma (OR = 2.63, 95% CI = 0.88 to 7.86; 4 cases, using lytic K8.1 antigen). Although this was a large study, exposure prevalence was low resulting in few exposed cases for some subtypes of B-cell lymphomas, T-cell lymphoma, and Hodgkin lymphoma. KSHV-related non-Hodgkin lymphoma is uncommon in sub-Saharan African populations despite a high prevalence of endemic KSHV (Engels *et al.* 2007b).

### 3.4.5 Integration across studies

Over 115 KSHV-positive cases of primary effusion lymphoma have been reported in the literature, primarily in case reports or case series, and have demonstrated a form of KSHV-related primary effusion lymphoma with distinguishing clinical, morphological, and immunophenotypic features that are uniquely associated with KSHV infection; cases occur in both human immunodeficiency virus type 1-positive and human immunodeficiency virus type 1 negative patients (IARC 2012). KSHV has also been associated with the plasmablastic variant of multicentric Castleman disease (see also Section 4). A majority of KSHV-related cases occur in human immunodeficiency virus type 1-positive patients (IARC 2012). To date, it is unclear if other viruses, including Epstein-Barr virus, or other cofactors, play a role in the development of these conditions, although their rarity has precluded large-scale studies of these factors. No association of multiple myeloma with KSHV infection was found in several studies. Data are insufficient to evaluate other lymphomas.

### 3.5 Other tumor sites

Few studies have been conducted on other tumor sites, with the partial exception of prostate cancer. Case-control or nested case-control studies of prostate cancer identified since 2008 have reported no or small increases in risk in association with KSHV (Sutcliffe *et al.* 2015, McDonald *et al.* 2011, Huang *et al.* 2008). In the most recent nested case-control study within a prospective clinical cancer prevention trial, Sutcliffe *et al.* reported no difference in KSHV-positive serostatus (11.6% in cases, 11.1% in controls,  $P = 0.81$ ). These findings are consistent with those reported in earlier studies and reviewed by IARC (2012). IARC (1997) reviewed six case-series studies of angiosarcoma, four of which were reported as showing no association with KSHV; five case studies of skin cancer were reviewed, four of which showed no association with KSHV (IARC 1997). In addition, IARC (1997) reviewed approximately 28 case-series and small case-control studies comparing KSHV DNA in skin and other tissue samples from patients with Kaposi sarcoma lesions with patients with other skin tumors or tissue lesions resembling Kaposi sarcoma and found, overall, only 3% of non-Kaposi sarcoma tissues contained KSHV DNA.

Since 2008, single studies have been identified for KSHV and hepatocellular (Su *et al.* 2014), vulvar (Simbiri *et al.* 2014), bladder cancers (Panagiotakis *et al.* 2013), laryngeal cancer (Mohamadian Roshan *et al.* 2014, Guvenc *et al.* 2008), head and neck squamous-cell carcinoma

(McLemore *et al.* 2010), and childhood inflammatory myofibroblastoma (Alaggio *et al.* 2010). The number of studies on specific solid tumors is insufficient to evaluate these endpoints, with the possible exception of prostate cancer, for which no associations with KSHV serostatus have been reported to date.

### 3.6 Synthesis across cancer endpoints

A summary of the evidence for KSHV infection and the different cancer endpoints from epidemiological studies is provided in Table 3-6. The level of evidence from cancer studies in human also considers studies of tissues from humans in addition to epidemiological studies and is provided in Section 5.

**Table 3-6. Summary of KSHV cancer endpoints and strength of the epidemiological evidence**

| Cancer endpoint   | Strength of evidence   |
|---|--|
| Kaposi sarcoma  | <ul style="list-style-type: none"> <li>Consistent evidence of highly increased, statistically significant risk in many prospective cohort and case-control studies.</li> <li>KSHV found in &gt; 90% of Kaposi sarcoma tumors in biopsies from Kaposi sarcoma case series.</li> <li>Dose-response relationships observed in several studies.</li> </ul> |
| Primary effusion lymphoma                               | <ul style="list-style-type: none"> <li>Consistent evidence of KSHV seropositivity or KSHV DNA in over 115 cases (individual case reports, case-series, case-comparison studies; morphologically and immunophenotypically distinct primary effusion lymphoma)</li> </ul>  |
| Multicentric Castleman disease                          | <ul style="list-style-type: none"> <li>Consistent evidence of KSHV seropositivity in MCD. KSHV is found in more than 99% of the plasmablastic variant of MCD. Molecular studies support the association with this specific form.</li> <li>Most cases observed in HIV-1-positive individuals.</li> </ul>  |
| Multiple myeloma  | <ul style="list-style-type: none"> <li>Inconsistent evidence, the majority with no or non-significant modest positive associations, from several case-control or case-comparison studies.</li> </ul>   |
| Other lymphohematopoietic cancers and solid tumor sites | <ul style="list-style-type: none"> <li>Inconsistent evidence for prostate cancer risk based on few studies.</li> <li>Inadequate number of epidemiological studies to evaluate other endpoints.</li> </ul>  |

KSHV = Kaposi sarcoma-associated herpesvirus; MCD = multicentric Castleman disease.

**This Page Intentionally Left Blank**

## 4 Mechanistic and Other Relevant Data

KSHV-associated oncogenesis is a complex process that involves interactions among various viral, host, and environmental cofactors (Mesri *et al.* 2014, Fukumoto *et al.* 2011). Although many of the mechanisms by which KSHV is able to establish and maintain infection are known, and many KSHV-encoded proteins and their functions have been characterized *in vitro*, some aspects of infection and transformation are still not well understood (Giffin and Damania 2014, Fukumoto *et al.* 2011). This section reviews the characteristics of KSHV-associated neoplasms and risk factors (Section 4.1), evaluates the mode of action and evidence that KSHV is an oncogenic virus (Section 4.2), and provides a synthesis of the information (Section 4.3).

As discussed in Section 3, three neoplasms have been associated with KSHV in humans, although the level of evidence for each endpoint varies: (1) Kaposi sarcoma, (2) primary effusion lymphoma, and (3) a plasmablastic variant of multicentric Castleman disease. These cancers occur most frequently in the context of immunosuppression and are the focus of this section (Giffin and Damania 2014, Mesri *et al.* 2014). Although other lymphohematopoietic neoplasms and several solid tumor sites (see Section 3) have been investigated, the available data were insufficient to establish a causal relationship with KSHV and are not discussed in this section.

### 4.1 Characteristics and risk factors

Some basic characteristics of the neoplasms associated with KSHV are shown in Table 4-1. The epidemiological data reviewed in Section 3 indicated that human immunodeficiency virus type 1 infection and immunosuppression are important host-related cofactors that increase the risk of developing KSHV-associated neoplasms. However, it is also clear that not all host and environmental cofactors have been identified. Further, KSHV latent and lytic transcripts include genes and non-coding RNAs that promote acquisition of various cancer hallmarks (e.g., insensitivity to antigrowth signals, avoiding cell-cycle arrest, immune evasion, genetic instability, cell proliferation, resisting apoptosis, and angiogenesis) (Mesri *et al.* 2014). This section reviews the role of immunosuppression, viral proteins, and non-coding RNAs, and other possible cofactors in KSHV-induced neoplasms.



**Table 4-1. Characteristics of KSHV-associated neoplasms**

| Neoplasm | Presentation  | Lineage and primary tumor cell   | Clonality                 | KSHV genomes  |
|----------|---|--|---------------------------|---|
| KS       | Highly angiogenic lesions found on skin, visceral organs, or mucosal surfaces; leaky vasculature                | Endothelial cell origin; tumor cells are spindle cells with mixed blood and lymphatic endothelial cell markers | Oligoclonal or monoclonal | > 99% of tumor cells contain single copy of KSHV genome |
| PEL      | Non-Hodgkin lymphoma; B-cell expansion (usually serous effusion without detectable tumor masses) in body cavity | B cells; CD20–; markers resemble partially differentiated plasma cells   | Monoclonal                | Tumor cells contain 50 to 100 copies of KSHV genome     |
| MCD      | Plasmablastic variant; disseminated lymphadenopathy   | B cell; IgM $\gamma$ -restricted plasmablasts  | Typically polyclonal      | Unknown, detected in most HIV-1+ MCD cases              |

Source: Adapted from (Giffin and Damania 2014).

KS = Kaposi sarcoma; MCD = multicentric Castleman disease; PEL = primary effusion lymphoma.

#### 4.1.1 Human immunodeficiency virus type 1 and immunosuppression

Although KSHV encodes oncogenic genes that could potentially induce all Kaposi sarcoma phenotypes, the risk of malignancy is greatly enhanced in the presence of cofactors, particularly immunosuppression by human immunodeficiency virus type 1 or iatrogenically during organ transplantation (Mesri *et al.* 2010). The importance of human immunodeficiency virus type 1 infection as a potent cofactor for KSHV oncogenesis is demonstrated by the dramatic increase in Kaposi sarcoma incidence in human immunodeficiency virus type 1-infected individuals (see Section 3). The risk for developing Kaposi sarcoma is strongly associated with defects in both cellular and humoral immunity. Decreasing CD4 cell counts are associated with an increasing risk of Kaposi sarcoma in both AIDS-associated and classic Kaposi sarcoma cases; however, human immunodeficiency virus type 1-infected individuals with higher CD4 counts are also developing Kaposi sarcoma as they age (Krown *et al.* 2008). Decreased CD19 cell counts are also associated with an increased risk of Kaposi sarcoma in human immunodeficiency virus type 1-positive and human immunodeficiency virus type 1-negative individuals (Uldrick and Whitby 2011). Iatrogenic immunosuppression also is associated with an increased risk (primarily in kidney allograft patients), but it is not nearly as great as that seen with human immunodeficiency virus type 1 infection (Mesri *et al.* 2010). This also may reflect differences in KSHV infection rates rather than human immunodeficiency virus type 1-specific causes, or differences in immune dysfunction. Further, human immunodeficiency virus type 1 infection causes changes in cytokine profiles and produces several viral proteins (human immunodeficiency virus type 1 transactivating protein [tat], human immunodeficiency virus type 1 negative factor protein [Nef]) that may be potent cofactors that can activate KSHV lytic replication and influence pathogenesis (Purushothaman *et al.* 2015, Mesri *et al.* 2014, Zhu *et al.* 2014). The biological basis for the role of immunosuppression in KSHV oncogenesis also involves the interactions of latent and lytic infection and is discussed in the following sections.

#### 4.1.2 KSHV transcripts

The KSHV genome contains 87 open reading frames including many genes found in other herpesviruses, several unique open reading frames (designated K1 to K15) not found in other herpesviruses, and at least 14 viral genes that are homologues of human cellular genes (Mesri *et al.* 2010, Wen and Damania 2010). Viral transcripts dysregulate several cellular signaling pathways including those involved in cell-cycle progression (e.g., vCyclin, LANA), angiogenesis (e.g., LANA, K1, vCCL, vGPCR, vIL6), apoptosis (e.g., LANA, K1, vFLIP, vBCL-2), immune modulation/evasion (e.g., vFLIP, K3, K5, vIRFs), signal transduction (e.g., K1, K15), and antiviral responses (IARC 2012, Mesri *et al.* 2010, Wen and Damania 2010). While latent infection is dominant in KSHV-infected cells *in vivo* and *in vitro*, both latent and lytic genes contribute to the malignant phenotype (Fukumoto *et al.* 2011, Mesri *et al.* 2010). Viral transcription profiling identified significant differences in the degree of KSHV transcription in human immunodeficiency virus type 1-infected Kaposi sarcoma patients that had not received antiviral treatment and/or chemotherapy (Hosseiniipour *et al.* 2014). Two primary subtypes of Kaposi sarcoma transcriptional signatures were identified: (1) lesions that displayed only a latent transcription pattern (i.e., LANA, vCyclin, vFLIP, miRNAs), and (2) lesions that exhibited extended but incomplete viral RNA transcription (e.g., vIRFs, K1, K15, Orf21/thymidine kinase, Orf36/protein kinase, and others). Only one sample exhibited a complete lytic transcription profile. The data demonstrate multiple subtypes of Kaposi sarcoma lesions and is consistent with other studies reporting similar “extended” KSHV transcripts under conditions of incomplete or abortive replication (discussed below).

A large portion of the KSHV genome is devoted toward evading the innate immune response of the host (e.g., interferon production, interferon regulatory factor activation, NK cell activity, complement activation, inflammasome activation, and chemokine activity), thus facilitating life-long infection (Giffin and Damania 2014, Fukumoto *et al.* 2011). In addition, KSHV evades adaptive immune responses by repressing viral antigen presentation, T-cell activation, BCR-mediated B-cell activation and B-cell differentiation (Giffin and Damania 2014). Professional antigen-presenting cells, including B cells, dendritic cells, and macrophages, are primary targets for KSHV infection (Campbell *et al.* 2014a, Knowlton *et al.* 2013). Cytokine profiles, surface expression of major histocompatibility complex (MHC) molecules, and T-cell activation are altered in infected antigen-presenting cells. This likely explains why T-cell responses to KSHV antigens are not very robust. Both complete and abortive virus replication cycles in antigen-presenting cells (APCs) can affect viral pathogenesis and progression to Kaposi sarcoma and B-cell cancers. Host cell transformation and oncogenesis are thought to result inadvertently from the survival mechanisms used by KSHV and other oncogenic viruses (Wen and Damania 2010).

In an immune-compromised host, lytic infected cells escape immunosurveillance and are able to express the full oncogenic repertoire, including genes that induce angiogenesis and inflammation (Cavallin *et al.* 2014, Mesri *et al.* 2014). In addition, some latently infected cells also express some of the early lytic genes but do not complete the lytic cycle. These cells may become progressively transformed and acquire oncogenic alterations that allow them to switch back to a less immunogenic latent form. Latent proteins and miRNAs play an important role in KSHV-associated neoplasms through inhibition of apoptosis and maintenance of latency while KSHV lytic proteins mimic or disrupt host cytokine signals, resulting in microenvironments that promote tumor growth through mechanisms called “paracrine neoplasia” (Cavallin *et al.* 2014,

Fukumoto *et al.* 2011, Mesri *et al.* 2010). This section briefly reviews the role of latent proteins, lytic proteins, non-coding RNAs, and their interactions in KSHV pathogenesis.

### **Latent proteins**

Latent KSHV infection is characterized by a circularized, extra-chromosomal viral genome (episome) that is tethered to the host chromosomes by latency-associated nuclear antigen (LANA) (Giffin and Damania 2014). All KSHV-infected cells express LANA, and antibody-based detection methods are the most powerful tool for diagnosis of pathological samples of KSHV infection (Fukumoto *et al.* 2011). Only a small portion of the viral genome is actively transcribed during latency and no functional or infectious viral particles are produced (Giffin and Damania 2014). LANA is the most consistently detected viral protein in KSHV-associated tumor cells and is necessary for replicating the episomal viral DNA (IARC 2012). In addition to LANA, latent transcripts include vCyclin, viral FLICE-inhibitory protein (vFLIP), kaposin A and B, viral interferon regulatory factor 3 (vIRF-3, also known as LANA-2) (expressed in primary effusion lymphoma and multicentric Castleman disease cells but not in Kaposi sarcoma) and viral miRNAs. Although the primary function of LANA is to maintain the viral episome, it also interferes with several anti-tumorigenic pathways, including p53 and anti-proliferative transforming growth factor- $\beta$  (TGF $\beta$ ), and might contribute to angiogenesis by stabilizing hypoxia-inducible factor-1 $\alpha$  (HIF-1 $\alpha$ ) and by targeting von Hippel Lindau for degradation (Mesri *et al.* 2010). Recent evidence also suggests that caspase cleavage sites in LANA have an important role in delaying apoptosis and blunting the caspase-1-mediated inflammasome, thus inhibiting key cellular defense mechanisms (Davis *et al.* 2015). vCyclin is a constitutive activator of cyclin-dependent kinase 6 (CDK6), a gene that is overexpressed in lymphomas and leukemias, induces cytokinesis defects and polyploidy, and promotes viral replication (Kollmann *et al.* 2013, Mesri *et al.* 2010). vFLIP and kaposin A and B contribute to the inflammatory microenvironment. Thus, these transcripts are associated with growth and proliferative signals, evasion of apoptosis, angiogenic and inflammatory signals, and a limitless replicative potential that support tumor growth and progression (Mesri *et al.* 2010). Transgenic mice expressing some or all of the KSHV latency genes develop phenotypes characteristic of KSHV malignancies (e.g., B-cell hyperplasia and lymphoma) (Sin and Dittmer 2013, Fakhari *et al.* 2006). Some biochemical and biological properties, including their role in carcinogenesis, of KSHV latent proteins are shown in Table 4-2.

**Table 4-2. Biochemical and biological properties of latent KSHV proteins**

| KSHV gene/protein                                   | Function in viral life cycle   | Homology to human protein             | Biochemical properties  | Role in carcinogenesis   |
|---|--|---------------------------------------|---|--|
| ORF 73/LANA   | Replication and maintenance of latent viral episome; partition of episomes to daughter cells         | None                                  | Interacts with histones, pRB, BET proteins, GSK-3 $\beta$ and others; inactivates p53-dependent transcriptional activation; induces S-phase entry and activates hTERT transcription; recruits origin-binding proteins | Anti-apoptotic activity, cell survival, deregulation of cell cycle, enhances ras-mediated transforming properties <i>in vitro</i> , tumorigenic in transgenic mice |
| ORF 71, K13/vFLIP                                   | Viral persistence; spindle cell formation and lymphomagenesis; inhibition of lytic viral replication | FLICE (caspase-8)-inhibitory proteins | Activates NF- $\kappa$ B; inhibits CD95/Fas-induced apoptosis, anoikis, superoxide-induced cell death; modulates MHC-1 expression   | Anti-apoptotic activity, cell survival, immune evasion   |
| ORF 72/vCYC   | Viral cyclin; strongly activates CDK6 protein kinase activity  | D-type cyclin                         | Phosphorylates H1, pRB, BCL-2, p27 <sup>KIP1</sup> in tandem with CDK6  | Deregulation of cell cycle   |
| K12/Kaposin A <sup>a</sup> , Kaposin B <sup>a</sup> | Modulation of cytokine mRNAs regulated by the p38 pathway  | None                                  | Interacts with cytohesin-1<br>Modulates p38/MK2   | Transforming properties in cultured cells  |
| K10.5/vIRF3, LANA-2 <sup>b</sup>                    | Modulates viral interferon response  | Interferon regulatory factor          | Modulates p53 function; activates IRF-3, IRF-7, c-myc; inhibits IRF-5   | Anti-apoptotic activity, immune evasion, cell survival   |
| K15/K15 protein <sup>b</sup>                        | Recruits endothelial cells to infected cells   | None                                  | Activates NF- $\kappa$ B, MEK/Erk; induces inflammatory cytokines; interacts with proteins involved in signal transduction, with members of src family of PTK, and with an apoptotic regulatory protein HAX-1         | Possible anti-apoptotic and angiogenic activity  |

Source: (IARC 2012).

BET = bromodomain and extra-terminal; CDK6 = cyclin-dependent kinase 6; FLICE = FADD-like interleukin-1 beta-converting enzyme; HAX-1 = HS1-associated protein X1; hTERT = human telomerase reverse transcriptase; IRF = interferon regulatory factor; LANA = latency associated nuclear antigen; MHC = major histocompatibility complex; NF- $\kappa$ B = nuclear factor kappa-light-chain-enhancer of activated B cells; pRB = retinoblastoma protein; PTK = protein tyrosine kinase; vCYC = viral cyclin; vIRF3 = viral interferon regulatory factor-3.

<sup>a</sup>Expression increases after activation of the lytic cycle.

<sup>b</sup>Lytic in endothelial cells; latent in B cells.

### **Lytic KSHV proteins**

The lytic cycle is characterized by expression of all viral transcripts in an orchestrated temporal order, replication of the viral genome, viral assembly, and viral egress (Wen and Damania 2010). Lytic reactivation requires expression of the replication and transcription activator (RTA) protein and is under epigenetic control (Giffin and Damania 2014, Li *et al.* 2014, Pantry and Medveczky 2009). RTA plays an important role as both an initiator and controller of KSHV lytic DNA replication (Purushothaman *et al.* 2015). Many of the lytic viral proteins, including K1, viral interferon response factors (vIRFs), vIL-6, viral-encoded chemokines (vCCLs), viral G protein-coupled receptor (vGPCR) and K15 contribute to the angiogenic and inflammatory phenotype of oncogenic lesions (Purushothaman *et al.* 2015, Sodhi *et al.* 2000). However, lytic infection generally results in cellular lysis and is unlikely to have a direct role in endothelial cell growth, transformation, or immortalization (Mesri *et al.* 2010). Two hypotheses (paracrine oncogenesis and abortive lytic replication) have been proposed to explain how the interactions between latent and lytic KSHV-infected cells contribute to a malignant phenotype (Cavallin *et al.* 2014, Mesri *et al.* 2010, Bais *et al.* 2003).

The paracrine hypothesis recognizes that host and viral growth factors can enhance latent genes and cytokines supplied by a minority of lytically infected cells or latently infected cells expressing early lytic genes (Cavallin *et al.* 2014, Mesri *et al.* 2010). Lytic genes induce a number of paracrine-acting factors such as vascular endothelial growth factor (VEGF), platelet-derived growth factor (PDGF), IL-6, angiopoietin 2 (ANGP2), and others that could drive latently infected cell proliferation, angiogenesis, and inflammation and support immune escape and survival. Experimental evidence supporting the role of lytic infection in oncogenesis include the following: (1) lytic viral proteins are expressed and virions are present in a minority of cells within Kaposi sarcoma lesions, (2) immunosuppression increases KSHV re-activation and lytic replication, (3) interruption of lytic replication by immune reconstitution or anti-lytic herpes anti-virals inhibits or prevents tumor development, (4) lytic infection is necessary to support viral episomal maintenance by recruitment of new cells to latency, and (5) endothelial cells expressing latent genes were tumorigenic in nude mice when co-injected with a few vGPCR-expressing cells (at a ratio that approximated the proportion of vGPCR-expressing and latent gene-expressing tumor cells found in human Kaposi sarcoma) but lost their tumorigenic potential in the absence of paracrine secretions from vGPCR-expressing cells (Mesri *et al.* 2010, Montaner *et al.* 2006). In addition, controlling KSHV lytic infection decreases the risk of developing Kaposi sarcoma in patients with AIDS (Uldrick and Whitby 2011) and transgenic mice expressing vGPCR develop angiogenic lesions that resemble Kaposi sarcoma (Guo *et al.* 2003, Yang *et al.* 2000).

The abortive lytic hypothesis recognizes that not all cells undergoing lytic reactivation complete the lytic cycle (i.e., abortive lytic replication). Such cells express only the oncogenic early lytic genes and can be transformed by genetic or epigenetic alterations, including vGPCR-mediated immortalization, and switch back to a less immunogenic latent status (Cavallin *et al.* 2014, Bais *et al.* 2003). Lytically infected cells can stimulate these cells through paracrine mechanisms.

Biochemical and biological properties of some KSHV lytic proteins are shown in Table 4-3.

**Table 4-3. Lytic KSHV proteins and their role in carcinogenesis**

| <b>KSHV gene/protein</b> | <b>Function in viral lifecycle</b>              | <b>Homology to human protein</b>  | <b>Biochemical properties</b>   | <b>Role in carcinogenesis</b>   |
|--------------------------|---|---|---|---|
| K1/vIP                   | May increase/decrease viral reactivation        | None  | Activates intracellular signaling cascades; induces angiogenic cytokines; blocks intracellular transport of BCR-complexes to cell surface | Transforming properties, angiogenesis, anti-apoptotic activity                                  |
| ORF 74/vGPCR             | Stimulates cellular proliferation               | G-protein-coupled receptor  | Activates Akt, MEK/Erk, JNK, p38; induces secretion of proinflammatory cytokines and angiogenic growth factors                            | Transforming properties in cells, tumorigenicity in mice, angiogenesis, anti-apoptotic activity |
| K2/vIL6                  | Viral IL6                                       | Interleukin-6   | Induces proliferation of PEL cell lines; induces VEGF and STAT3 phosphorylation   | Tumor cell survival, angiogenesis, hematopoiesis  |
| K3/miR-1, ZMP-B          | Down modulates HLA, ICAM-1, B7-2                | None  | E3 ubiquitin ligase   | Immune evasion  |
| K5/miR-2                 | Down modulates HLA-A, HLA-B, ICAM-1, CD86, CD1d | Part of a family of membrane-bound-E3-ubiquitin ligases                         | E3 ubiquitin ligase   | Immune evasion  |
| K6/vCCL-1, vMIP-I        | Viral chemokine                                 | Chemokine homologue related to MIP-1 $\alpha$                                   | CCR8 agonist; induces monocyte chemotaxis (VEGF production)   | Angiogenic properties, anti-apoptotic activity  |
| K4/vCCL-2, vMIP-II       | Viral chemokine                                 | Chemokine homologue related to macrophage inflammatory protein (MIP)-1 $\alpha$ | CCR3, CCR5, CCR8 agonist; induces monocyte chemotaxis   | Angiogenic properties   |
| K4.1/vCCL-3, vMIP-III    | Viral chemokine                                 | Some homology to chemokines TARC and eotaxin                                    | CCR4, XCR1 agonist  | Angiogenic properties   |
| K7/vIAP                  | Apoptosis inhibitor                             | Structurally related to a splice variant of survivin                            | Binds to and inhibits several proteins involved in apoptosis; induces degradation of I $\kappa$ B, p53, vGCR                              | Anti-apoptotic activity   |
| ORF 16/vBCL-2            | Viral Bcl-2                                     | Bcl-2   | Heterodimerizes with human Bcl-2  | Anti-apoptotic activity   |

| <b>KSHV gene/protein</b>         | <b>Function in viral lifecycle</b>   | <b>Homology to human protein</b> | <b>Biochemical properties</b>  | <b>Role in carcinogenesis</b>   |
|----------------------------------|--|----------------------------------|--|---|
| ORF 45/ORF 45                    | Virion protein important for lytic replication; inhibits virus-mediated induction of type 1 interferon | None                             | Binds to and inhibits phosphorylation of IRF-7   | Virion infectivity, immune evasion  |
| ORF 50/K-RTA                     | Immediate-early transactivator; ubiquitin E3 ligase  | None                             | Binds to and activates several lytic viral promoters directly or by interacting with RBPJk; promotes ubiquitination and degradation of IRF-7; represses p53  | Reactivation of lytic viral replication from latency, immune evasion, anti-apoptotic activity |
| K8/K-bZIP                        | Modulates cell cycle and lytic reactivation  | None                             | EBV Zta homologue; binds to lytic replication origin; binds to, antagonizes, and recruits p53 to ND10/PML bodies; inhibits G1/S transition; co-regulator of K-RTA  | Deregulation of cell cycle  |
| K9/vIRF-1                        | Modulates viral interferon responses   | Interferon regulatory factor     | Prevents IRF-3-mediated transcription; inhibits p53-transcriptional activity and prevents p53-dependent apoptosis  | Transformation of rodent fibroblasts, immune evasion, anti-apoptotic activity                 |
| K11/vIRF-2                       | Modulates viral interferon responses   | Interferon regulatory factor     | Inhibits interferon induction; inhibits induction of CD95L   | Immune evasion, anti-apoptotic activity   |
| K10.5/vIRF3, LANA-2 <sup>a</sup> | Modulates viral interferon responses   | Interferon regulatory factor     | Modulates p53 function; activates IRF-3, IRF-7, c-myc; inhibits IRF-5  | Anti-apoptotic activity, immune evasion, cell survival  |
| K15/K15 protein <sup>a</sup>     | Recruits endothelial cells to infected cells   | None                             | Activates NF- $\kappa$ B, MEK/Erk; induces inflammatory cytokines; interacts with proteins involved in signal transduction, with members of src family of protein tyrosine kinases, and with an apoptotic regulatory protein HAX-1 | Possible anti-apoptotic and angiogenic activity   |

Sources: Purushothaman *et al.* 2015, IARC 2012.

<sup>a</sup>Lytic in endothelial cells; latent in B cells.

## Non-coding RNAs

KSHV-encoded miRNAs are expressed in latently infected cells and are involved with suppressing lytic reactivation by activating NF- $\kappa$ B, endothelial cell differentiation, and angiogenesis (Moody *et al.* 2013, Mesri *et al.* 2010). KSHV infection also induces cellular miRNAs, particularly miR-132. miR-132 inhibits anti-viral innate immune responses and induces abnormal endothelial cell proliferation, thus, linking viral immune escape with angiogenesis, two important features of Kaposi sarcoma. Moody *et al.* (2013) demonstrated that KSHV miRNAs mediated cellular transformation and tumorigenesis using a KSHV-induced cellular transformation system of primary rat mesenchymal precursor cells. A mutant virus with a cluster of precursor miRNAs deleted failed to transform primary cells and caused cell-cycle arrest and apoptosis. The oncogenicity of the mutant virus was restored by complementation with the miRNA cluster or several individual precursor miRNAs. Qin *et al.* (2010) also demonstrated that KSHV miRNAs induce IL-6 and IL-10 secretion in murine macrophages and human myelomonocytic cells and support a role in programming macrophage cytokine responses that favor KSHV-associated tumor progression. Transgenic mice expressing KSHV miRNAs as well as multiple viral latent proteins (e.g., LANA, vFLIP, vCYC) recapitulated the precancerous latent state of KSHV infection (i.e., chronic activation of mature B cells, increased plasma cell frequency, marginal zone B-cell hyperplasia, and hyperglobulinemia) (Sin and Ditmer 2013). A subsequent study that crossed these KSHV latency locus transgenic mice with Myc transgenic mice demonstrated that the KSHV latency locus cooperates with the deregulated Myc pathways to induce lymphoma at a higher rate than in single latency or Myc transgenic mice (Sin *et al.* 2015).

In addition to the miRNAs, KSHV also encodes a long, non-coding RNA known as polyadenylated nuclear (PAN) RNA during the early lytic stage (Campbell *et al.* 2014b). PAN RNA has been implicated in KSHV gene expression, replication, and immune modulation and is required for optimal expression of all KSHV lytic genes. It also facilitates LANA-episomal dissociation during lytic replication.

### 4.1.3 Other possible cofactors

It is likely that other cofactors are involved in KSHV-associated neoplasms as evidenced by the geographic variation in incidence before the human immunodeficiency virus type 1 epidemic (see Section 3.2.1). Co-infection with other human herpesviruses (e.g., Epstein-Barr virus; see monograph on Epstein-Barr virus), or human cytomegalovirus can activate KSHV lytic replication and influence its pathogenesis (Purushothaman *et al.* 2015). While other cofactors have been proposed (see Section 3), their mode of action and role as risk factors for specific cancers have not been established.

## 4.2 Mode of action and evidence for cancer causation

As discussed in the Overview and Introduction Section, it is often difficult to apply stringent criteria, such as Koch's postulates or Hill's considerations, for determining that a human tumor virus is oncogenic (Moore and Chang 2010, zur Hausen 2001). Therefore, in addition to the usual criteria used by epidemiologists to determine causality, other factors should be considered as proposed by IARC (1997) and zur Hausen (2001, 1994). Unlike other oncogenic viruses, Hill's epidemiological considerations for causality are met for KSHV and Kaposi sarcoma



(Moore and Chang 2010). Thus, application of various criteria applicable to oncogenic viruses and mechanistic data provide strong evidence that KSHV is oncogenic in humans and are briefly reviewed below by tumor type.

#### 4.2.1 Kaposi sarcoma

Chang *et al.* (1994) was the first to report KSHV in Kaposi sarcoma tissues obtained from AIDS patients. There is now substantial evidence that KSHV is the etiological agent of Kaposi sarcoma. KSHV is present in the vast majority of Kaposi sarcoma tumor cells, thus meeting Hill's consideration for strength and consistency (each cell contains one copy of the KSHV genome and its seroprevalence was correlated with the global incidence of the disease) (Fukumoto *et al.* 2011, Mesri *et al.* 2010). In addition, the most difficult of the Hill considerations, experimental epidemiological evidence in a randomized clinical trial, was fulfilled when AIDS patients who were treated with an antih herpesvirus drug were protected from new occurrences of Kaposi sarcoma but not against established tumors (Moore and Chang 2010). Other lines of evidence linking KSHV and Kaposi sarcoma include the following (Cavallin *et al.* 2014):

- KSHV is associated with all four clinical forms of the disease,
- KSHV is found in spindle cells,
- KSHV infection precedes the onset of Kaposi sarcoma,
- KSHV seroprevalence is highest in areas of high Kaposi sarcoma incidence,
- KSHV encodes many viral oncogenes, and
- KSHV transforms and induces tumorigenesis in endothelial cells.
- Monoclonal expansion of KSHV tumors supports an etiologic role for the virus (Duprez *et al.* 2007, Judde *et al.* 2000).

In addition, *in vitro* models and animal models have been developed that accurately reproduce many features observed in KSHV-associated neoplasms (Dittmer *et al.* 2015, Ashlock *et al.* 2014, Mesri and Cesarman 2011, Mutlu *et al.* 2007, An *et al.* 2006).

#### 4.2.2 Primary effusion lymphoma

Primary effusion lymphoma is rare. It occurs most frequently in human immunodeficiency virus type 1-infected homosexual males; however, it has been described in association with post-transplant immunosuppression, and in human immunodeficiency virus type 1-negative elderly patients of Eastern European/Mediterranean and Jewish descent (two populations with high seropositivity for KSHV and elevated incidence of classic Kaposi sarcoma) (IARC 2012, Fukumoto *et al.* 2011, Dourmishev *et al.* 2003). Because of inadequate pathology and public health infrastructures, the rates of primary effusion lymphoma in the areas of highest KSHV prevalence (i.e., sub-Saharan Africa) are currently unknown.

KSHV DNA sequences were first identified in eight body-cavity-based lymphomas (later identified as primary effusion lymphoma) taken from patients with AIDS, but they were not detected in 185 other lymphomas examined from patients with or without AIDS (Cesarman *et al.* 1995). Asou *et al.* (2000) reported similar findings—KSHV was present in 21 of 21 primary effusion lymphoma patients but was not detected in 139 patients with other AIDS- and non-

AIDS-related lymphomas. KSHV infection in primary effusion lymphoma cells is predominantly latent with only a small subpopulation of cells that stain for markers of lytic reactivation (Fukumoto *et al.* 2011). vIL-6 is the most frequently detected lytic protein and induces VEGF expression and cytokine signals in a broad range of cell types (Fukumoto *et al.* 2011, Aoki *et al.* 2001). Further evidence that supports KSHV infection as a causal agent in primary effusion lymphoma is as follows:

- Primary effusion lymphoma neoplasms always carry KSHV and are commonly co-infected with Epstein-Barr virus (Fukumoto *et al.* 2011, Cesarman *et al.* 1995);
- KSHV has been incorporated as a diagnostic criterion for primary effusion lymphoma (IARC 2012);
- About half of primary effusion lymphoma patients also have Kaposi sarcoma, and a previous Kaposi sarcoma diagnosis confers an increased risk of primary effusion lymphoma relative to all other AIDS-associated non-Hodgkin lymphomas (Fukumoto *et al.* 2011, Dourmishev *et al.* 2003);
- Primary effusion lymphoma cells contain 50 to 100 copies per cell of KSHV DNA, and the expression pattern of KSHV-encoded proteins is almost identical in primary effusion lymphoma and Kaposi sarcoma (Giffin and Damania 2014, IARC 2012, Fukumoto *et al.* 2011);
- Several KSHV-infected cell lines have been established from primary effusion lymphoma cells that retain a stable latent viral genome including a KSHV+/EBV- cell line established from KSHV+ and Epstein-Barr virus+ primary effusion lymphoma cases, suggesting that KSHV plays an essential role in primary effusion lymphoma pathogenesis (Fukumoto *et al.* 2011, Katano *et al.* 1999); and
- Expression of several KSHV viral genes was required for survival of primary effusion lymphoma cells in culture (IARC 2012).

#### 4.2.3 Multicentric Castleman disease

Castleman disease is an uncommon lymphoproliferative disorder with several histological variants (hyaline vascular, plasma cell, mixed, and plasmablastic types) and two clinical forms; unicentric (or localized) and multicentric (Fajgenbaum *et al.* 2014, Al-Maghrabi 2011, Bonekamp *et al.* 2011, El-Osta and Kurzrock 2011). The plasmablastic form of multicentric Castleman disease has been specifically associated with KSHV in both human immunodeficiency virus type 1-positive and human immunodeficiency virus type 1-negative cases (Dupin *et al.* 2000). Thus, KSHV-associated multicentric Castleman disease is now recognized as a distinct entity from other forms of multicentric Castleman disease and is classified by WHO as “large B-cell lymphoma arising in HHV8-associated multicentric Castleman disease” (Swerdlow *et al.* 2008). This plasmablastic variant of diffuse large B cell lymphoma has KSHV expression of lytic antigens, cytoplasmic IgM with light chain restriction, and viral interleukin 6 expression and is characterized by severe inflammatory symptoms (attributed primarily to elevated levels of IL-6), an aggressive course, and a poor prognosis (Fajgenbaum *et al.* 2014, Venkataraman *et al.* 2013, Polizzotto *et al.* 2012, Bonekamp *et al.* 2011). Dysregulated IL-6 levels are thought to contribute to multicentric Castleman disease development (Cai *et al.* 2010). Fewer studies have investigated the association of multicentric

Castleman disease and KSHV compared to Kaposi sarcoma or primary effusion lymphoma; however, additional supporting evidence is as follows:

- Several case-control, case-comparison or case series studies have reported an association between KSHV and multicentric Castleman disease (see Section 3 and Table 3-4);
- Cases of Kaposi sarcoma and multicentric Castleman disease frequently occur together in the human immunodeficiency virus type 1-positive population (see Section 3, Reddy and Mitsuyasu 2011, Cronin and Warnke 2009);
- The level of KSHV in peripheral blood monocytes or plasma corresponds with symptoms during multicentric Castleman disease flares in human immunodeficiency virus type 1-positive individuals (Reddy and Mitsuyasu 2011);
- KSHV-positive multicentric Castleman disease cases are associated with KSHV lytic infection, and high levels of vIL-6 have been detected in infected B cells and sera and likely are associated with B-cell proliferation seen in multicentric Castleman disease (IARC 2012, Fukumoto *et al.* 2011, Aoki *et al.* 2001);
- vIL-6, a frequently detected lytic protein, enhances cytokine signaling, increases human IL-6 and induces VEGF expression, and likely exacerbates inflammation and disease progression and may be a mechanism leading to multicentric Castleman disease (Giffin and Damania 2014, Al-Maghrabi 2011, Fukumoto *et al.* 2011); and
- Treatment of multicentric Castleman disease cases with an inhibitor of KSHV infection has shown some success (Casper *et al.* 2004).

### 4.3 Synthesis

Human viral oncogenesis is a complex process that involves interactions among many viral, host, and environmental factors. Immunosuppression is an important cofactor based on the dramatic increase in Kaposi sarcoma incidence among human immunodeficiency virus type 1-infected individuals. KSHV infection has been clearly linked with all clinical forms of Kaposi sarcoma, as well as primary effusion lymphoma and a plasmablastic variant of multicentric Castleman disease, and biologically plausible modes-of-action have been described. One or more KSHV transcripts are expressed in all KSHV-associated tumors and are required for survival of tumor cells in culture. Latent transcripts drive viral persistence and replication, promote host cell proliferation, and prevent apoptosis while lytic transcripts dysregulate cell signaling pathways that contribute to the angiogenic and inflammatory phenotype of oncogenic lesions via paracrine mechanisms.

## 5 Overall Cancer Hazard Evaluation and Listing Recommendation

Kaposi sarcoma-associated virus (KSHV) is known to be a human carcinogen based on sufficient evidence from studies in humans. This conclusion is based on epidemiological and molecular studies showing that it causes Kaposi sarcoma (Table 5-1) primary effusion lymphoma (Table 5-2), and a plasmablastic variant of multicentric Castleman disease (Table 5-3) in humans, together with supporting evidence from mechanistic studies demonstrating the biological plausibility of its carcinogenicity in humans.

Data are inadequate to evaluate the association between KSHV and multiple myeloma, which has inconsistent evidence from epidemiological studies and no available evidence from mechanistic studies.

The following tables provide the level of evidence recommendations for the carcinogenicity of KSHV for each endpoint from studies in humans, including the key data from both epidemiological and molecular studies in humans.

**Table 5-1. Summary of the evidence for KSHV and Kaposi sarcoma from human studies**

| Types of studies                     | Kaposi sarcoma  |
|--------------------------------------|---|
| <b>Epidemiological</b>               |   |
| Positive associations                | 39/39 <sup>a</sup> case-control studies (all ORs > 1.0) and all 16 <sup>a</sup> cohort/nested case-control studies; ORs/RRs for all studies are mostly statistically significant and some very high (10-→ 100-fold)<br><br>Elevated RR in both HIV-1-negative and HIV-1-positive populations and in all types of Kaposi sarcoma<br><br>Dose response in several studies |
| <b>Molecular (human tissue)</b>      |   |
| Clonality                            | Mono- or oligoclonal  |
| % KSHV-infected tumors               | > 99% (~1 copy/cell)  |
| KSHV protein expression <sup>b</sup> |   |
| Latent                               | LANA, vCyclin, vFLIP, kaposin A and B   |
| Lytic                                | RTA, K1, vIRFs, vIL-6, vGPCR, vCCLs, K15  |
| <b>Level of evidence</b>             | <b>Sufficient</b>   |

HIV-1 = human immunodeficiency virus type 1; K1 = unique KSHV protein 1; K15 = unique KSHV protein 15; KSHV = Kaposi sarcoma-associated herpes virus; LANA = latency-associated nuclear antigen; OR = odds ratio; RR = risk ratio; RTA = replication and transactivation protein; vCCLs = viral-encoded chemokines; vFLIP = viral FLICE-inhibitory protein; vGPCR = viral G protein-coupled receptor; vIL-6 = viral homolog of interleukin-6; vIRF = viral interferon regulatory factors.

<sup>a</sup>The number of studies is based on those reporting risk estimates.

<sup>b</sup>See Section 4.1.2 for more details.

**Table 5-2. Summary of the evidence for KSHV and primary effusion lymphoma from human studies**

| Types of studies                | Primary effusion lymphoma (PEL)   |
|---------------------------------|---|
| <b>Epidemiological</b>          |   |
| Positive associations           | <p>Found in over 115 cases; at least 95 individual case reports and case-series studies and 21 cases in one comparison study</p> <p>PEL is a single distinct pathological entity</p> <p>Found in both HIV-1-negative and -positive cases and in transplant patients</p> <p>Approximately 50% cases also have Kaposi sarcoma; some PEL cases found in geographical regions associated with endemic or classic Kaposi sarcoma</p> |
| <b>Molecular (human tissue)</b> |   |
| Clonality                       | Monoclonal  |
| % KSHV-infected tumors          | 100% (high copy number); cell lines established from PEL tumors retain a stable latent viral genome   |
| KSHV protein expression         | Similar to Kaposi sarcoma   |
| Other                           | <p>KSHV is part of diagnostic criteria</p> <p>Expression of KSHV viral genes was required for survival of PEL cells in culture</p>  |
| <b>Level of evidence</b>        | <b>Sufficient</b>   |

HIV-1 = human immunodeficiency virus type 1; KSHV = Kaposi sarcoma-associated herpesvirus; PEL = primary effusion lymphoma.

**Table 5-3. Summary of the evidence for KSHV and multicentric Castleman disease from human studies**

| Types of studies                     | Multicentric Castleman disease (MCD)   |
|--------------------------------------|--|
| <b>Epidemiological</b>               |  |
| Positive associations                | <p>Found in most cases of MCD (plasmablastic variant) in several case series studies;</p> <p>4/4 case comparison studies; very high ORs or no cases of KSHV among controls</p> |
| <b>Molecular (human tissue)</b>      |  |
| Clonality                            | Typically polyclonal <sup>b</sup>  |
| % KSHV-infected tumors               | 100% of cancerous lymph nodes KSHV+ with HIV-1+ and 50% KSHV+ with HIV-1-  |
| KSHV protein expression <sup>a</sup> | LANA, vIL-6  |
| <b>Level of evidence</b>             | <b>Sufficient (plasmablastic variant)</b>  |

HIV-1 = human immunodeficiency virus type 1; KSHV = Kaposi sarcoma-associated herpesvirus; LANA = latency-associated nuclear antigen; OR = odds ratio; vIL-6 = viral homolog of interleukin-6.

<sup>a</sup>Multicentric Castleman disease is associated with lytic infection.

<sup>b</sup>Monoclonal B-cell expansion have been reported.

## References

1. Ablashi DV, Chatlynne LG, Whitman JE, Jr., Cesarman E. 2002. Spectrum of Kaposi's sarcoma-associated herpesvirus, or human herpesvirus 8, diseases. *Clin Microbiol Rev* 15(3): 439-464.
2. ACS. 2014. *Kaposi Sarcoma*. American Cancer Society. 34 pp. <http://www.cancer.org/>.
3. Al-Maghrabi JA. 2011. Castleman's disease. Update on pathogenesis. *Saudi Med J* 32(5): 451-458.
4. Alaggio R, Cecchetto G, Bisogno G, Gambini C, Calabro ML, Inserra A, Boldrini R, De Salvo GL, ES GdA, Dall'igna P. 2010. Inflammatory myofibroblastic tumors in childhood: a report from the Italian Cooperative Group studies. *Cancer* 116(1): 216-226.
5. Alagiozoglou L, Morris L, Bredell H, Martin DJ, Sitas F. 2003. Human herpesvirus-8 antibodies and DNA in HIV-1 infected patients in South Africa. *Epidemiol Infect* 131(3): 1125-1129.
6. Albini A, Aluigi M, Benelli R, Berti E, Biberfeld P, Blasig C, Calabro M, Calvo F, Chiecobianchi L, Corbellino M, Delmistro A, Ekman M, Favero A, Hofschneider P, Kaaya E, Lebbe C, Morel P, Neipel F, Noonan D, Parravicini C, Repetto L, Schalling M, Sturzl M, Tschachler E. 1996. Oncogenesis in HIV-infection. *Int J Oncol* 9(1): 5-8.
7. Albrecht D, Meyer T, Lorenzen T, Stoehr A, Arndt R, Plettenberg A. 2004. Epidemiology of HHV-8 infection in HIV-positive patients with and without Kaposi sarcoma: diagnostic relevance of serology and PCR. *J Clin Virol* 30(2): 145-149. (as cited in IARC 2012)
8. Amodio E, Goedert JJ, Barozzi P, Riva G, Firenze A, Bonura F, Viviano E, Romano N, Luppi M. 2011a. Differences in Kaposi sarcoma-associated herpesvirus-specific and herpesvirus-non-specific immune responses in classic Kaposi sarcoma cases and matched controls in Sicily. *Cancer Sci* 102(10): 1769-1773.
9. Amodio E, Valentini M, Gori-Savellini G, Valenti RM, Romano N, Goedert JJ, Cusi MG. 2011b. Prevalence of Toscana and Sicilian phlebovirus antibodies in classic Kaposi sarcoma case patients and control subjects in Sicily. *J Infect Dis* 204(9): 1423-1426.
10. An FQ, Folarin HM, Compitello N, Roth J, Gerson SL, McCrae KR, Fakhari FD, Dittmer DP, Renne R. 2006. Long-term-infected telomerase-immortalized endothelial cells: a model for Kaposi's sarcoma-associated herpesvirus latency in vitro and in vivo. *J Virol* 80(10): 4833-4846.
11. Anderson LA, Lauria C, Romano N, Brown EE, Whitby D, Graubard BI, Li Y, Messina A, Gafa L, Vitale F, Goedert JJ. 2008. Risk factors for classical Kaposi sarcoma in a population-based case-control study in Sicily. *Cancer Epidemiol Biomarkers Prev* 17(12): 3435-3443.

12. Andreoni M, Sarmati L, Nicastrì E, El Sawaf G, El Zalabani M, Uccella I, Bugarini R, Parisi SG, Rezza G. 2002. Primary human herpesvirus 8 infection in immunocompetent children. *JAMA* 287(10): 1295-1300.
13. Aoki Y, Yarchoan R, Wyvill K, Okamoto S, Little RF, Tosato G. 2001. Detection of viral interleukin-6 in Kaposi sarcoma-associated herpesvirus-linked disorders. *Blood* 97(7): 2173-2176.
14. Armstrong AW, Lam KH, Chase EP. 2013. Epidemiology of classic and AIDS-related Kaposi's sarcoma in the USA: incidence, survival, and geographical distribution from 1975 to 2005. *Epidemiol Infect* 141(1): 200-206.
15. Ascoli V, Sirianni MC, Mezzaroma I, Mastroianni CM, Vullo V, Andreoni M, Narciso P, Scalzo CC, Nardi F, Pistilli A, Lo Coco F. 1999. Human herpesvirus-8 in lymphomatous and nonlymphomatous body cavity effusions developing in Kaposi's sarcoma and multicentric Castleman's disease. *Ann Diagn Pathol* 3(6): 357-363.
16. Ascoli V, Lo Coco F, Torelli G, Vallisa D, Cavanna L, Bergonzi C, Luppi M. 2002. Human herpesvirus 8-associated primary effusion lymphoma in human immunodeficiency virus-negative patients: a clinico-epidemiologic variant resembling classic Kaposi's sarcoma. *Haematologica* 87(4): 339-343.
17. Ashlock BM, Ma Q, Issac B, Mesri EA. 2014. Productively infected murine Kaposi's sarcoma-like tumors define new animal models for studying and targeting KSHV oncogenesis and replication. *PLoS One* 9(1): e87324.
18. Asou H, Tasaka T, Said JW, Daibata M, Kamada N, Koeffler HP. 2000. Co-infection of HHV-6 and HHV-8 is rare in primary effusion lymphoma. *Leuk Res* 24(1): 59-61.
19. Bais C, Van Geelen A, Eroles P, Mutlu A, Chiozzini C, Dias S, Silverstein RL, Rafii S, Mesri EA. 2003. Kaposi's sarcoma associated herpesvirus G protein-coupled receptor immortalizes human endothelial cells by activation of the VEGF receptor-2/ KDR. *Cancer Cell* 3(2): 131-143.
20. Barozzi P, Luppi M, Masini L, Marasca R, Savarino M, Morselli M, Ferrari MG, Bevini M, Bonacorsi G, Torelli G. 1996. Lymphotropic herpes virus (EBV, HHV-6, HHV-8) DNA sequences in HIV negative Castleman's disease. *Clin Mol Pathol* 49(4): M232-235.
21. Barozzi P, Luppi M, Facchetti F, Mecucci C, Alu M, Sarid R, Rasini V, Ravazzini L, Rossi E, Festa S, Crescenzi B, Wolf DG, Schulz TF, Torelli G. 2003. Post-transplant Kaposi sarcoma originates from the seeding of donor-derived progenitors. *Nat Med* 9(5): 554-561.
22. Beachler DC, Gellert LL, Jacobson LP, Ambinder RF, Breen EC, Martinez-Maza O, Rabkin CS, Kaslow RA, D'Souza G. 2011. Kaposi sarcoma-associated herpesvirus serum DNA and antibodies not associated with subsequent non-Hodgkin lymphoma risk. *J Acquir Immune Defic Syndr* 56(2): 188-192.
23. Benavente Y, Mbisa G, Labo N, Casabonne D, Becker N, Maynadie M, Foretova L, Cocco PL, Nieters A, Staines A, Bofetta P, Brennan P, Whitby D, de Sanjose S. 2011. Antibodies

- against lytic and latent Kaposi's sarcoma-associated herpes virus antigens and lymphoma in the European EpiLymph case-control study. *Br J Cancer* 105(11): 1768-1771.
24. Bhutani M, Polizzotto MN, Uldrick TS, Yarchoan R. 2015. Kaposi sarcoma-associated herpesvirus-associated malignancies: epidemiology, pathogenesis, and advances in treatment. *Semin Oncol* 42(2): 223-246.
  25. Boivin G, Gaudreau A, Toma E, Lalonde R, Routy JP, Murray G, Handfield J, Bergeron MG. 1999. Human herpesvirus 8 DNA load in leukocytes of human immunodeficiency virus-infected subjects: correlation with the presence of Kaposi's sarcoma and response to anticytomegalovirus therapy. *Antimicrob Agents Chemother* 43(2): 377-380.
  26. Bonekamp D, Horton KM, Hruban RH, Fishman EK. 2011. Castleman disease: the great mimic. *Radiographics* 31(6): 1793-1807.
  27. Boulanger E, Duprez R, Delabesse E, Gabarre J, Macintyre E, Gessain A. 2005. Mono/oligoclonal pattern of Kaposi sarcoma-associated herpesvirus (KSHV/HHV-8) episomes in primary effusion lymphoma cells. *Int J Cancer* 115(4): 511-518.
  28. Boulanger E, Afonso PV, Yahiaoui Y, Adle-Biassette H, Gabarre J, Agbalika F. 2008. Human herpesvirus-8 (HHV-8)-associated primary effusion lymphoma in two renal transplant recipients receiving rapamycin. *Am J Transplant* 8(3): 707-710.
  29. Bryant-Greenwood P, Sorbara L, Filie AC, Little R, Yarchoan R, Wilson W, Raffeld M, Abati A. 2003. Infection of mesothelial cells with human herpes virus 8 in human immunodeficiency virus-infected patients with Kaposi's sarcoma, Castleman's disease, and recurrent pleural effusions. *Mod Pathol* 16(2): 145-153.
  30. Burbelo PD, Kovacs JA, Wagner J, Bayat A, Rhodes CS, De Souza Y, Greenspan JS, Iadarola MJ. 2012. The cancer-associated virus landscape in HIV patients with oral hairy leukoplakia, Kaposi's sarcoma, and non-Hodgkin lymphoma. *AIDS Res Treat* 2012: 634523.
  31. Cai Q, Verma SC, Lu J, Robertson ES. 2010. Molecular biology of Kaposi's sarcoma-associated herpesvirus and related oncogenesis. *Adv Virus Res* 78: 87-142.
  32. Campbell DM, Rappocciolo G, Jenkins FJ, Rinaldo CR. 2014a. Dendritic cells: key players in human herpesvirus 8 infection and pathogenesis. *Front Microbiol* 5: Article 452.
  33. Campbell M, Kung HJ, Izumiya Y. 2014b. Long non-coding RNA and epigenetic gene regulation of KSHV. *Viruses* 6(11): 4165-4177.
  34. Cannon MJ, Dollard SC, Black JB, Edlin BR, Hannah C, Hogan SE, Patel MM, Jaffe HW, Offermann MK, Spira TJ, Pellett PE, Gunthel CJ. 2003. Risk factors for Kaposi's sarcoma in men seropositive for both human herpesvirus 8 and human immunodeficiency virus. *AIDS* 17(2): 215-222.
  35. Casper C, Nichols WG, Huang ML, Corey L, Wald A. 2004. Remission of HHV-8 and HIV-associated multicentric Castleman disease with ganciclovir treatment. *Blood* 103(5): 1632-1634.



36. Casper C, Krantz EM, Corey L, Kuntz SR, Wang J, Selke S, Hamilton S, Huang ML, Wald A. 2008. Valganciclovir for suppression of human herpesvirus-8 replication: a randomized, double-blind, placebo-controlled, crossover trial. *J Infect Dis* 198(1): 23-30.
37. Castleman B, Iverson L, Menendez VP. 1956. Localized mediastinal lymphnode hyperplasia resembling thymoma. *Cancer* 9(4): 822-830. (as cited in IARC 1997)
38. Cattamanchi A, Saracino M, Selke S, Huang ML, Magaret A, Celum C, Corey L, Wald A, Casper C. 2011. Treatment with valacyclovir, famciclovir, or antiretrovirals reduces human herpesvirus-8 replication in HIV-1 seropositive men. *J Med Virol* 83(10): 1696-1703.
39. Cattani P, Capuano M, Lesnoni La Parola I, Guido R, Santangelo R, Cerimele F, Masini C, Nanni G, Fadda G, Cerimele D. 1998. Human herpesvirus 8 in Italian HIV-seronegative patients with Kaposi sarcoma. *Arch Dermatol* 134(6): 695-699.
40. Cattani P, Capuano M, Graffeo R, Ricci R, Cerimele F, Cerimele D, Nanni G, Fadda G. 2001. Kaposi's sarcoma associated with previous human herpesvirus 8 infection in kidney transplant recipients. *J Clin Microbiol* 39(2): 506-508.
41. Cavallin LE, Goldschmidt-Clermont P, Mesri EA. 2014. Molecular and cellular mechanisms of KSHV oncogenesis of Kaposi's sarcoma associated with HIV/AIDS. *PLoS Pathog* 10(7): e1004154.
42. Cesarman E, Chang Y, Moore PS, Said JW, Knowles DM. 1995. Kaposi's sarcoma-associated herpesvirus-like DNA sequences in AIDS-related body-cavity-based lymphomas. *N Engl J Med* 332(18): 1186-1191.
43. Chang Y, Cesarman E, Pessin MS, Lee F, Culpepper J, Knowles DM, Moore PS. 1994. Identification of herpesvirus-like DNA sequences in AIDS-associated Kaposi's sarcoma. *Science* 266(5192): 1865-1869.
44. Chang Y, Moore PS, Talbot SJ, Boshoff CH, Zarkowska T, Godden K, Paterson H, Weiss RA, Mitnacht S. 1996. Cyclin encoded by KS herpesvirus. *Nature* 382(6590): 410.
45. Chang-Moore Laboratory. 2009. *Kaposi's Sarcoma-Associated Herpesvirus (KSHV)*. Chang-Moore Laboratory. <http://www.tumorvirology.pitt.edu/kshvrsch.html>. Accessed on 1/26/15.
46. Chao C, Jacobson LP, Jenkins FJ, Tashkin D, Martinez-Maza O, Roth MD, Ng L, Margolick JB, Chmiel JS, Zhang ZF, Detels R. 2009. Recreational drug use and risk of Kaposi's sarcoma in HIV- and HHV-8-coinfected homosexual men. *AIDS Res Hum Retroviruses* 25(2): 149-156.
47. Chatlynne LG, Lapps W, Handy M, Huang YQ, Masood R, Hamilton AS, Said JW, Koeffler HP, Kaplan MH, Friedman-Kien A, Gill PS, Whitman JE, Ablashi DV. 1998. Detection and titration of human herpesvirus-8-specific antibodies in sera from blood donors, acquired immunodeficiency syndrome patients, and Kaposi's sarcoma patients using a whole virus enzyme-linked immunosorbent assay. *Blood* 92(1): 53-58.

48. Chen W, Huang Q, Zuppan CW, Rowsell EH, Cao JD, Weiss LM, Wang J. 2009. Complete absence of KSHV/HHV-8 in posttransplant lymphoproliferative disorders: an immunohistochemical and molecular study of 52 cases. *Am J Clin Pathol* 131(5): 632-639.
49. Coluzzi M, Calabro ML, Manno D, Chieco-Bianchi L, Schulz TF, Ascoli V. 2003. Reduced seroprevalence of Kaposi's sarcoma-associated herpesvirus (KSHV), human herpesvirus 8 (HHV8), related to suppression of Anopheles density in Italy. *Med Vet Entomol* 17(4): 461-464.
50. Corbellino M, Poirel L, Aubin JT, Paulli M, Magrini U, Bestetti G, Galli M, Parravicini C. 1996. The role of human herpesvirus 8 and Epstein-Barr virus in the pathogenesis of giant lymph node hyperplasia (Castleman's disease). *Clin Infect Dis* 22(6): 1120-1121.
51. Cronin DMP, Warnke RA. 2009. Castleman disease: an update on classification and the spectrum of associated lesions. *Advances in Anatomic Pathology* 16(4): 236-246.
52. Damania B. Targeting the PI3K/Akt/mTOR Pathway in AIDS-Associated Malignancies in Molecular Basis for Therapy of AIDS-Defining cancers. 2010. Editors: Dirk Dittmer and Susan Krown. Springer Science+Business Media, LLC, 256 pp.
53. Davis DA, Naiman NE, Wang V, Shrestha P, Haque M, Hu D, Anagho HA, Carey RF, Davidoff KS, Yarchoan R. 2015. Identification of caspase cleavage sites in KSHV latency-associated nuclear antigen and their effects on caspase-related host defense responses. *PLoS Pathog* 11(7): e1005064.
54. Decker LL, Shankar P, Khan G, Freeman RB, Dezube BJ, Lieberman J, Thorley-Lawson DA. 1996. The Kaposi sarcoma-associated herpesvirus (KSHV) is present as an intact latent genome in KS tissue but replicates in the peripheral blood mononuclear cells of KS patients. *J Exp Med* 184(1): 283-288.
55. Dedicoat M, Newton R. 2003. Review of the distribution of Kaposi's sarcoma-associated herpesvirus (KSHV) in Africa in relation to the incidence of Kaposi's sarcoma. *Br J Cancer* 88(1): 1-3.
56. Deloose ST, Smit LA, Pals FT, Kersten MJ, van Noesel CJ, Pals ST. 2005. High incidence of Kaposi sarcoma-associated herpesvirus infection in HIV-related solid immunoblastic/plasmablastic diffuse large B-cell lymphoma. *Leukemia* 19(5): 851-855.
57. DHHS. 2013. *Guidelines for the Prevention and Treatment of Opportunistic Infections Among HIV-Exposed and HIV-Infected Children*. Bethesda, MD: National Institutes of Health. 384 pp.  
[https://aidsinfo.nih.gov/contentfiles/lvguidelines/oi\\_guidelines\\_pediatrics.pdf](https://aidsinfo.nih.gov/contentfiles/lvguidelines/oi_guidelines_pediatrics.pdf).
58. DHHS. 2014. *Guidelines for Prevention and Treatment of Opportunistic Infections in HIV-Infected Adults and Adolescents*. Bethesda, MD: National Institutes of Health. 407 pp.  
[https://aidsinfo.nih.gov/contentfiles/lvguidelines/adult\\_oi.pdf](https://aidsinfo.nih.gov/contentfiles/lvguidelines/adult_oi.pdf).
59. Dittmer DP, Damania B, Sin SH. 2015. Animal models of tumorigenic herpesviruses-an update. *Curr Opin Virol* 14: 145-150.

60. Dotti G, Fiocchi R, Motta T, Facchinetti B, Chiadini B, Borleri GM, Gavazzeni G, Barbui T, Rambaldi A. 1999. Primary effusion lymphoma after heart transplantation: a new entity associated with human herpesvirus-8. *Leukemia* 13(5): 664-670.
61. Dourmishev LA, Dourmishev AL, Palmeri D, Schwartz RA, Lukac DM. 2003. Molecular genetics of Kaposi's sarcoma-associated herpesvirus (human herpesvirus-8) epidemiology and pathogenesis. *Microbiol Mol Biol Rev* 67(2): 175-212.
62. Dukers NH, Renwick N, Prins M, Geskus RB, Schulz TF, Weverling GJ, Coutinho RA, Goudsmit J. 2000. Risk factors for human herpesvirus 8 seropositivity and seroconversion in a cohort of homosexual men. *Am J Epidemiol* 151(3): 213-224.
63. Dupin N, Fisher C, Kellam P, Ariad S, Tulliez M, Franck N, van Marck E, Salmon D, Gorin I, Escande JP, Weiss RA, Alitalo K, Boshoff C. 1999. Distribution of human herpesvirus-8 latently infected cells in Kaposi's sarcoma, multicentric Castleman's disease, and primary effusion lymphoma. *Proc Natl Acad Sci USA* 96(8): 4546-4551.
64. Dupin N, Diss TL, Kellam P, Tulliez M, Du MQ, Sicard D, Weiss RA, Isaacson PG, Boshoff C. 2000. HHV-8 is associated with a plasmablastic variant of Castleman disease that is linked to HHV-8-positive plasmablastic lymphoma. *Blood* 95(4): 1406-1412.
65. Duprez R, Lacoste V, Briere J, Couppie P, Frances C, Sainte-Marie D, Kassa-Kelembho E, Lando MJ, Essame Oyono JL, Nkegoum B, Hbid O, Mahe A, Lebbe C, Tortevoye P, Huerre M, Gessain A. 2007. Evidence for a multiclonal origin of multicentric advanced lesions of Kaposi sarcoma. *J Natl Cancer Inst* 99(14): 1086-1094.
66. Dupuy A, Schulz T, Chevret S, Agbalika F, Pellet C, Janier M, Dupin N, Verola O, Calvo F, Lebbe C. 2009. Asymmetrical transmission of human herpesvirus 8 among spouses of patients with Kaposi sarcoma. *Br J Dermatol* 160(3): 540-545.
67. Edelman DC. 2005. Human herpesvirus 8—a novel human pathogen. *Viol J* 2: 78, 32 pp.
68. El-Osta HE, Kurzrock R. 2011. Castleman's disease: from basic mechanisms to molecular therapeutics. *Oncologist* 16(4): 497-511.
69. Engels EA, Biggar RJ, Marshall VA, Walters MA, Gamache CJ, Whitby D, Goedert JJ. 2003. Detection and quantification of Kaposi's sarcoma-associated herpesvirus to predict AIDS-associated Kaposi's sarcoma. *AIDS* 17(12): 1847-1851.
70. Engels EA, Atkinson JO, Graubard BI, McQuillan GM, Gamache C, Mbisa G, Cohn S, Whitby D, Goedert JJ. 2007a. Risk factors for human herpesvirus 8 infection among adults in the United States and evidence for sexual transmission. *J Infect Dis* 196(2): 199-207.
71. Engels EA, Mbulaiteye SM, Othieno E, Gomez M, Mathew S, Cesarman E, Knowles DM, Chadburn A. 2007b. Kaposi sarcoma-associated herpesvirus in non-Hodgkin lymphoma and reactive lymphadenopathy in Uganda. *Hum Pathol* 38(2): 308-314.
72. Fajgenbaum DC, van Rhee F, Nabel CS. 2014. HHV-8-negative, idiopathic multicentric Castleman disease: novel insights into biology, pathogenesis, and therapy. *Blood* 123(19): 2924-2933.

73. Fakhari FD, Jeong JH, Kanan Y, Dittmer DP. 2006. The latency-associated nuclear antigen of Kaposi sarcoma-associated herpesvirus induces B cell hyperplasia and lymphoma. *J Clin Invest* 116(3): 735-742.
74. Frances C, Mouquet C, Calvez V. 1999. Human herpesvirus 8 and renal transplantation. *N Engl J Med* 340(13): 1045; author's reply p. 1046.
75. Frances C, Mouquet C, Marcelin AG, Barete S, Agher R, Charron D, Benalia H, Dupin N, Piette JC, Bitker MO, Calvez V. 2000. Outcome of kidney transplant recipients with previous human herpesvirus-8 infection. *Transplantation* 69(9): 1776-1779.
76. Fukumoto H, Kanno T, Hasegawa H, Katano H. 2011. Pathology of Kaposi's sarcoma-associated herpesvirus infection. *Front Microbiol* 2: 175.
77. Ganem D. 2010. KSHV and the pathogenesis of Kaposi sarcoma: listening to human biology and medicine. *J Clin Invest* 120(4): 939-949.
78. Gao SJ, Kingsley L, Li M, Zheng W, Parravicini C, Ziegler J, Newton R, Rinaldo CR, Saah A, Phair J, Detels R, Chang Y, Moore PS. 1996. KSHV antibodies among Americans, Italians and Ugandans with and without Kaposi's sarcoma. *Nat Med* 2(8): 925-928.
79. Garcia-Astudillo LA, Leyva-Cobian F. 2006. Human herpesvirus-8 infection and Kaposi's sarcoma after liver and kidney transplantation in different geographical areas of Spain. *Transpl Immunol* 17(1): 65-69.
80. Gerard L, Meignin V, Galicier L, Fieschi C, Leturque N, Piketty C, Fonquernie L, Agbalika F, Oksenhendler E. 2009. Characteristics of non-Hodgkin lymphoma arising in HIV-infected patients with suppressed HIV replication. *AIDS* 23(17): 2301-2308.
81. Gessain A, Sudaka A, Briere J, Fouchard N, Nicola MA, Rio B, Arborio M, Troussard X, Audouin J, Diebold J, de The G. 1996. Kaposi sarcoma-associated herpes-like virus (human herpesvirus type 8) DNA sequences in multicentric Castleman's disease: is there any relevant association in non-human immunodeficiency virus-infected patients? *Blood* 87(1): 414-416.
82. Giffin L, Damania B. 2014. KSHV: pathways to tumorigenesis and persistent infection. *Adv Virus Res* 88: 111-159.
83. Giuliani M, Cordiali-Fei P, Castilletti C, Di Carlo A, Palamara G, Boros S, Rezza G. 2007. Incidence of human herpesvirus 8 (HHV-8) infection among HIV-uninfected individuals at high risk for sexually transmitted infections. *BMC Infect Dis* 7: 143.
84. Gloghini A, Dolcetti R, Carbone A. 2013. Lymphomas occurring specifically in HIV-infected patients: from pathogenesis to pathology. *Semin Cancer Biol* 23(6): 457-467.
85. Goedert JJ, Vitale F, Lauria C, Serraino D, Tamburini M, Montella M, Messina A, Brown EE, Rezza G, Gafa L, Romano N, the Classical Kaposi's Sarcoma Working Group. 2002. Risk factors for classical Kaposi's sarcoma. *J Natl Cancer Inst* 94(22): 1712-1718.

86. Gopal S, Fedoriw Y, Montgomery ND, Kampani C, Krysiak R, Sanders MK, Dittmer DP, Liomba NG. 2014. Multicentric Castleman's disease in Malawi. *Lancet* 384(9948): 1158.
87. Greenblatt RM, Jacobson LP, Levine AM, Melnick S, Anastos K, Cohen M, DeHovitz J, Young MA, Burns D, Miotti P, Koelle DM. 2001. Human herpesvirus 8 infection and Kaposi's sarcoma among human immunodeficiency virus-infected and -uninfected women. *J Infect Dis* 183(7): 1130-1134.
88. Grulich AE, Olsen SJ, Luo K, Hendry O, Cunningham P, Cooper DA, Gao SJ, Chang Y, Moore PS, Kaldor JM. 1999. Kaposi's sarcoma-associated herpesvirus: a sexually transmissible infection? *J Acquir Immune Defic Syndr Hum Retrovirol* 20(4): 387-393.
89. Guo HG, Sadowska M, Reid W, Tschachler E, Hayward G, Reitz M. 2003. Kaposi's sarcoma-like tumors in a human herpesvirus 8 ORF74 transgenic mouse. *J Virol* 77(4): 2631-2639.
90. Guvenc MG, Midilli K, Ozdogan A, Inci E, Tahamiler R, Enver O, Sirin G, Ergin S, Kuskucu M, Divanoglu EO, Yilmaz G, Altas K. 2008. Detection of HHV-8 and HPV in laryngeal carcinoma. *Auris Nasus Larynx* 35(3): 357-362.
91. Herman PS, Shogreen MR, White WL. 1998. The evaluation of human herpesvirus 8 (Kaposi's sarcoma-associated herpesvirus) in cutaneous lesions of Kaposi's sarcoma: a study of formalin-fixed paraffin-embedded tissue. *Am J Dermatopathol* 20(1): 7-11.
92. Hiatt KM, Nelson AM, Lichy JH, Fanburg-Smith JC. 2008. Classic Kaposi Sarcoma in the United States over the last two decades: a clinicopathologic and molecular study of 438 non-HIV-related Kaposi Sarcoma patients with comparison to HIV-related Kaposi Sarcoma. *Mod Pathol* 21(5): 572-582.
93. Hill AB. 1965. The environment and disease: Association or causation? *Proc R Soc Med* 58: 295-300.
94. Hosseini-Moghaddam SM, Soleimanirahbar A, Mazzulli T, Rotstein C, Husain S. 2012. Post renal transplantation Kaposi's sarcoma: a review of its epidemiology, pathogenesis, diagnosis, clinical aspects, and therapy. *Transpl Infect Dis* 14(4): 338-345.
95. Hosseinipour MC, Sweet KM, Xiong J, Namarika D, Mwafongo A, Nyirenda M, Chiwoko L, Kamwendo D, Hoffman I, Lee J, Phiri S, Vahrson W, Damania B, Dittmer DP. 2014. Viral profiling identifies multiple subtypes of Kaposi's sarcoma. *MBio* 5(5): e01633-01614.
96. Howard MR, Whitby D, Bahadur G, Suggett F, Boshoff C, Tenant-Flowers M, Schulz TF, Kirk S, Matthews S, Weller IV, Tedder RS, Weiss RA. 1997. Detection of human herpesvirus 8 DNA in semen from HIV-infected individuals but not healthy semen donors. *AIDS* 11(2): F15-19.
97. Huang WY, Hayes R, Pfeiffer R, Viscidi RP, Lee FK, Wang YF, Reding D, Whitby D, Papp JR, Rabkin CS. 2008. Sexually transmissible infections and prostate cancer risk. *Cancer Epidemiol Biomarkers Prev* 17(9): 2374-2381.

98. Humphrey RW, O'Brien TR, Newcomb FM, Nishihara H, Wyvill KM, Ramos GA, Saville MW, Goedert JJ, Straus SE, Yarchoan R. 1996. Kaposi's sarcoma (KS)-associated herpesvirus-like DNA sequences in peripheral blood mononuclear cells: association with KS and persistence in patients receiving anti-herpesvirus drugs. *Blood* 88(1): 297-301.
99. IARC. 1997. Kaposi sarcoma herpesvirus/human herpesvirus 8. In *Epstein-Barr Virus and Kaposi's Sarcoma Herpesvirus/Human Herpesvirus 8*. IARC Monographs on the Evaluation of Carcinogenic Risks to Humans, vol. 70. Lyon, France: International Agency for Research on Cancer. pp. 375-492.
100. IARC. 2012. Kaposi sarcoma herpesvirus. In *Biological Agents*. IARC Monographs on the Evaluation of Carcinogenic Risks to Humans, vol. 100B. Lyon, France: International Agency for Research on Cancer. pp. 169-214.
101. Ismail SI, Mahmoud IS, Salman MA, Sughayer MA, Mahafzah AM. 2011. Frequent detection of human herpes virus-8 in bone marrow of Jordanian patients of multiple myeloma. *Cancer Epidemiol* 35(5): 471-474.
102. Jacobson LP, Jenkins FJ, Springer G, Munoz A, Shah KV, Phair J, Zhang Z, Armenian H. 2000. Interaction of human immunodeficiency virus type 1 and human herpesvirus type 8 infections on the incidence of Kaposi's sarcoma. *J Infect Dis* 181(6): 1940-1949.
103. Jalilvand S, Shoja Z, Mokhtari-Azad T, Nategh R, Gharehbaghian A. 2011. Seroprevalence of human herpesvirus 8 (HHV-8) and incidence of Kaposi's sarcoma in Iran. *Infect Agent Cancer* 6: 5.
104. Jalilvand S, Tornesello ML, Buonaguro FM, Buonaguro L, Naraghi ZS, Shoja Z, Ziaee AA, Hamkar R, Shahmahmoodi S, Nategh R, Mokhtari-Azad T. 2012. Molecular epidemiology of human herpesvirus 8 variants in Kaposi's sarcoma from Iranian patients. *Virus Res* 163(2): 644-649.
105. Judde JG, Lacoste V, Briere J, Kassa-Kelembho E, Clyti E, Couppie P, Buchrieser C, Tulliez M, Morvan J, Gessain A. 2000. Monoclonality or oligoclonality of human herpesvirus 8 terminal repeat sequences in Kaposi's sarcoma and other diseases. *J Natl Cancer Inst* 92(9): 729-736.
106. Kanno T, Sato Y, Nakamura T, Sakamoto K, Sata T, Katano H. 2010. Genotypic and clinicopathological characterization of Kaposi's sarcoma-associated herpesvirus infection in Japan. *J Med Virol* 82(3): 400-406.
107. Kapelushnik J, Ariad S, Benharroch D, Landau D, Moser A, Delsol G, Brousset P. 2001. Post renal transplantation human herpesvirus 8-associated lymphoproliferative disorder and Kaposi's sarcoma. *Br J Haematol* 113(2): 425-428. (as cited in IARC 2012)
108. Katano H, Hoshino Y, Morishita Y, Nakamura T, Satoh H, Iwamoto A, Herndier B, Mori S. 1999. Establishing and characterizing a CD30-positive cell line harboring HHV-8 from a primary effusion lymphoma. *J Med Virol* 58(4): 394-401.
109. Kazakov DV, Prinz BM, Michaelis S, Schmid M, Muller B, Adams V, Burg G, Kempf W. 2002. Study of HHV-8 DNA sequences in archival biopsies from lesional skin of Kaposi's

- sarcoma, various mesenchymal tumors and related reactive conditions. *J Cutan Pathol* 29(5): 279-281.
110. Kedes DH, Operskalski E, Busch M, Kohn R, Flood J, Ganem D. 1996. The seroepidemiology of human herpesvirus 8 (Kaposi's sarcoma-associated herpesvirus): distribution of infection in KS risk groups and evidence for sexual transmission. *Nat Med* 2(8): 918-924.
  111. Kedes DH, Ganem D, Ameli N, Bacchetti P, Greenblatt R. 1997. The prevalence of serum antibody to human herpesvirus 8 (Kaposi sarcoma-associated herpesvirus) among HIV-seropositive and high-risk HIV-seronegative women. *JAMA* 277(6): 478-481.
  112. Keller R, Zago A, Viana MC, Bourboulia D, Desgranges C, Casseb J, Moura WV, Dietze R, Collandre H. 2001. HHV-8 infection in patients with AIDS-related Kaposi's sarcoma in Brazil. *Braz J Med Biol Res* 34(7): 879-886.
  113. Kishimoto K, Kitamura T, Hirayama Y, Tate G, Mitsuya T. 2009. Cytologic and immunocytochemical features of EBV negative primary effusion lymphoma: report on seven Japanese cases. *Diagn Cytopathol* 37(4): 293-298.
  114. Knowlton ER, Lepone LM, Li J, Rappocciolo G, Jenkins FJ, Rinaldo CR. 2013. Professional antigen presenting cells in human herpesvirus 8 infection. *Front Immunol* 3: 427.
  115. Kollmann K, Heller G, Schneckenleithner C, Warsch W, Scheicher R, Ott RG, Schafer M, Fajmann S, Schleder M, Schiefer AI, Reichart U, Mayerhofer M, Hoeller C, Zochbauer-Muller S, Kerjaschki D, Bock C, Kenner L, Hoefler G, Freissmuth M, Green AR, Moriggl R, Busslinger M, Malumbres M, Sexl V. 2013. A kinase-independent function of CDK6 links the cell cycle to tumor angiogenesis. *Cancer Cell* 24(2): 167-181.
  116. Koski L, Ngoma T, Mwaiselage J, Le L, Soliman AS. 2014. Changes in the pattern of Kaposi's sarcoma at Ocean Road Cancer Institute in Tanzania (2006-2011). *Int J STD AIDS*, doi:10.1177/0956462414544724.
  117. Krown SE, Lee JY, Dittmer DP, Consortium AM. 2008. More on HIV-associated Kaposi's sarcoma. *N Engl J Med* 358(5): 535-536; author reply 536.
  118. Labo N, Miley W, Benson CA, Campbell TB, Whitby D. 2015. Epidemiology of Kaposi's sarcoma-associated herpesvirus in HIV-1-infected US persons in the era of combination antiretroviral therapy. *AIDS* 29(10): 1217-1225.
  119. Lager I, Altini M, Coleman H, Ali H. 2003. Oral Kaposi's sarcoma: a clinicopathologic study from South Africa. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 96(6): 701-710.
  120. Laney AS, Cannon MJ, Jaffe HW, Offermann MK, Ou CY, Radford KW, Patel MM, Spira TJ, Gunthel CJ, Pellett PE, Dollard SC. 2007. Human herpesvirus 8 presence and viral load are associated with the progression of AIDS-associated Kaposi's sarcoma. *AIDS* 21(12): 1541-1545. (as cited in IARC 2012)

121. Lanternier F, Lebbe C, Schartz N, Farhi D, Marcelin AG, Kerob D, Agbalika F, Verola O, Gorin I, Janier M, Avril MF, Dupin N. 2008. Kaposi's sarcoma in HIV-negative men having sex with men. *AIDS* 22(10): 1163-1168.
122. Lefrere JJ, Meyohas MC, Mariotti M, Meynard JL, Thauvin M, Frottier J. 1996. Detection of human herpesvirus 8 DNA sequences before the appearance of Kaposi's sarcoma in human immunodeficiency virus (HIV)-positive subjects with a known date of HIV seroconversion. *J Infect Dis* 174(2): 283-287.
123. Li Q, He M, Zhou F, Ye F, Gao SJ. 2014. Activation of Kaposi's sarcoma-associated herpesvirus (KSHV) by inhibitors of class III histone deacetylases: identification of sirtuin 1 as a regulator of the KSHV life cycle. *J Virol* 88(11): 6355-6367.
124. Luppi M, Barozzi P, Maiorana A, Collina G, Ferrari MG, Marasca R, Morselli M, Rossi E, Ceccherini-Nelli L, Torelli G. 1996. Frequency and distribution of herpesvirus-like DNA sequences (KSHV) in different stages of classic Kaposi's sarcoma and in normal tissues from an Italian population. *Int J Cancer* 66(4): 427-431.
125. Luppi M, Barozzi P, Schulz TF, Setti G, Staskus K, Trovato R, Narni F, Donelli A, Maiorana A, Marasca R, Sandrini S, Torelli G. 2000a. Bone marrow failure associated with human herpesvirus 8 infection after transplantation. *N Engl J Med* 343(19): 1378-1385.
126. Luppi M, Barozzi P, Schulz TF, Trovato R, Donelli A, Narni F, Sheldon J, Marasca R, Torelli G. 2000b. Nonmalignant disease associated with human herpesvirus 8 reactivation in patients who have undergone autologous peripheral blood stem cell transplantation. *Blood* 96(7): 2355-2357.
127. Magri MC, Carbone PH, de los Santos-Fortuna E, Caterino-de-Araujo A. 2009. A comparative study of the frequency of antibody and titers against human herpesvirus 8 latent and lytic antigens in "at-risk" individuals and among patients with Kaposi's sarcoma. *J Med Virol* 81(7): 1292-1297.
128. Marcelin AG, Roque-Afonso AM, Hurtova M, Dupin N, Tulliez M, Sebah M, Arkoub ZA, Guettier C, Samuel D, Calvez V, Dussaix E. 2004. Fatal disseminated Kaposi's sarcoma following human herpesvirus 8 primary infections in liver-transplant recipients. *Liver Transpl* 10(2): 295-300.
129. Martin JN, Ganem DE, Osmond DH, Page-Shafer KA, Macrae D, Kedes DH. 1998. Sexual transmission and the natural history of human herpesvirus 8 infection. *N Engl J Med* 338(14): 948-954.
130. Martin JN. 2007. The epidemiology of KSHV and its association with malignant disease. In *Human Herpesviruses: Biology, Therapy, and Immunoprophylaxis*. Arvin A, Campadelli-Fiume G, Mocarski E *et al.*, eds. Cambridge: Cambridge University Press.
131. Martro E, Esteve A, Schulz TF, Sheldon J, Gambus G, Munoz R, Whitby D, Casabona J, Euro-Shaks study g. 2007. Risk factors for human herpesvirus 8 infection and AIDS-associated Kaposi's sarcoma among men who have sex with men in a European multicentre study. *Int J Cancer* 120(5): 1129-1135. (as cited in IARC 2012)



132. Massambu C, Pyakurel P, Kaaya E, Enbom M, Urassa W, Demirhan I, Loewer J, Linde A, Chandra A, Heiden T, Doerr HW, Chandra P, Biberfeld P. 2003. Serum HHV8 DNA and Tat antibodies in Kaposi's sarcoma patients with and without HIV-1 infection. *Anticancer Res* 23(3B): 2389-2395.
133. McDonald AC, Jenkins FJ, Bunker CH, Wilson JW, Patrick AL, Weissfeld JL. 2011. A case-cohort study of human herpesvirus 8 seropositivity and incident prostate cancer in Tobago. *Infect Agent Cancer* 6: 25.
134. McLemore MS, Haigentz M, Jr., Smith RV, Nuovo GJ, Alos L, Cardesa A, Brandwein-Gensler M. 2010. Head and neck squamous cell carcinomas in HIV-positive patients: a preliminary investigation of viral associations. *Head Neck Pathol* 4(2): 97-105.
135. Melbye M, Cook PM, Hjalgrim H, Begtrup K, Simpson GR, Biggar RJ, Ebbesen P, Schulz TF. 1998. Risk factors for Kaposi's-sarcoma-associated herpesvirus (KSHV/HHV-8) seropositivity in a cohort of homosexual men, 1981-1996. *Int J Cancer* 77(4): 543-548.
136. Mesri EA, Cesarman E, Boshoff C. 2010. Kaposi's sarcoma and its associated herpesvirus. *Nat Rev Cancer* 10(10): 707-719.
137. Mesri EA, Cesarman E. 2011. Kaposi's sarcoma herpesvirus oncogenesis is a notch better in 3D. *Cell Host Microbe* 10(6): 529-531.
138. Mesri EA, Feitelson MA, Munger K. 2014. Human viral oncogenesis: a cancer hallmarks analysis. *Cell Host Microbe* 15(3): 266-282.
139. Miller G, Rigsby MO, Heston L, Grogan E, Sun R, Metroka C, Levy JA, Gao SJ, Chang Y, Moore P. 1996. Antibodies to butyrate-inducible antigens of Kaposi's sarcoma-associated herpesvirus in patients with HIV-1 infection. *N Engl J Med* 334(20): 1292-1297.
140. Min J, Katzenstein DA. 1999. Detection of Kaposi's sarcoma-associated herpesvirus in peripheral blood cells in human immunodeficiency virus infection: association with Kaposi's sarcoma, CD4 cell count, and HIV RNA levels. *AIDS Res Hum Retroviruses* 15(1): 51-55.
141. Minhas V, Wood C. 2014. Epidemiology and transmission of Kaposi's sarcoma-associated herpesvirus. *Viruses* 6(11): 4178-4194.
142. Mohamadian Roshan N, Jafarian A, Ayatollahi H, Ghazvini K, Abbas Tabatabaee S. 2014. Correlation of laryngeal squamous cell carcinoma and infections with either HHV-8 or HPV-16/18. *Pathol Res Prac* 210: 205-209.
143. Monini P, de Lellis L, Fabris M, Rigolin F, Cassai E. 1996a. Kaposi's sarcoma-associated herpesvirus DNA sequences in prostate tissue and human semen. *N Engl J Med* 334(18): 1168-1172.
144. Monini P, de Lellis L, Cassai E. 1996b. Absence of HHV-8 in prostate and semen: author's reply. *N Engl J Med* 335(6): 1238-1239.

145. Montaner S, Sodhi A, Ramsdell AK, Martin D, Hu J, Sawai ET, Gutkind JS. 2006. The Kaposi's sarcoma-associated herpesvirus G protein-coupled receptor as a therapeutic target for the treatment of Kaposi's sarcoma. *Cancer Res* 66(1): 168-174.
146. Moody R, Zhu Y, Huang Y, Cui X, Jones T, Bedolla R, Lei X, Bai Z, Gao SJ. 2013. KSHV microRNAs mediate cellular transformation and tumorigenesis by redundantly targeting cell growth and survival pathways. *PLoS Pathog* 9(12): e1003857.
147. Moore PS, Chang Y. 1995. Detection of herpesvirus-like DNA sequences in Kaposi's sarcoma in patients with and without HIV infection. *N Engl J Med* 332(18): 1181-1185.
148. Moore PS, Chang Y. 1998. Kaposi's sarcoma (KS), KS-associated herpesvirus, and the criteria for causality in the age of molecular biology. *Am J Epidemiol* 147(3): 217-221.
149. Moore PS, Chang Y. 2014. The conundrum of causality in tumor virology: the cases of KSHV and MCV. *Semin Cancer Biol* 26: 4-12.
150. Moore PS, Kingsley LA, Holmberg SD, Spira T, Gupta P, Hoover DR, Parry JP, Conley LJ, Jaffe HW, Chang Y. 1996. Kaposi's sarcoma-associated herpesvirus infection prior to onset of Kaposi's sarcoma. *AIDS* 10(2): 175-180.
151. Moore PS, Chang YA. 2010. Why do viruses cause cancer? Highlights of the first century of human tumour virology. *Nature Reviews Cancer* 10(12): 878-889.
152. Mousavi SM, Sundquist J, Hemminki K. 2014. Risk of Kaposi sarcoma among immigrants to Sweden. *Acta Derm Venereol* 94(4): 476-477.
153. Mutlu AD, Cavallin LE, Vincent L, Chiozzini C, Eroles P, Duran EM, Asgari Z, Hooper AT, La Perle KM, Hilsher C, Gao SJ, Dittmer DP, Rafii S, Mesri EA. 2007. *In vivo* growth-restricted and reversible malignancy induced by human herpesvirus-8 KSHV: a cell and animal model of virally induced Kaposi's sarcoma. *Cancer Cell* 11(3): 245-258.
154. Mwakigonja AR, Pyakurel P, Kokhaei P, Pak F, Lema LK, Kaaya EE, Biberfeld P. 2008. Human herpesvirus-8 (HHV-8) sero-detection and HIV association in Kaposi's sarcoma (KS), non-KS tumors and non-neoplastic conditions. *Infect Agent Cancer* 3: 10.
155. Nador RG, Cesarman E, Chadburn A, Dawson DB, Ansari MQ, Sald J, Knowles DM. 1996. Primary effusion lymphoma: a distinct clinicopathologic entity associated with the Kaposi's sarcoma-associated herpes virus. *Blood* 88(2): 645-656.
156. Nalwoga A, Cose S, Wakeham K, Miley W, Ndibazza J, Drakeley C, Elliott A, Whitby D, Newton R. 2015. Association between malaria exposure and Kaposi's sarcoma-associated herpes virus seropositivity in Uganda. *Trop Med Int Health* 20(5): 665-672.
157. Nawar EW, Cole SR, Farzadegan H, Witt MD, Jenkins FJ, Margolick JB, Phair JP, Jacobson LP. 2008. Sexual activity and Kaposi's sarcoma among human immunodeficiency virus type 1 and human herpesvirus type 8-coinfected men. *Ann Epidemiol* 18(7): 517-521.

158. NCI. 2014. *A Snapshot of Kaposi Sarcoma*. National Cancer Institute. <http://www.cancer.gov/researchandfunding/snapshots/kaposisarcoma>. Accessed on 2/20/15.
159. Newton R, Ziegler J, Bourboulia D, Casabonne D, Beral V, Mbidde E, Carpenter L, Reeves G, Parkin DM, Wabinga H, Mbulaiteye S, Jaffe H, Weiss R, Boshoff C, Uganda Kaposi's Sarcoma Study Group. 2003a. The sero-epidemiology of Kaposi's sarcoma-associated herpesvirus (KSHV/HHV-8) in adults with cancer in Uganda. *Int J Cancer* 103(2): 226-232.
160. Newton R, Ziegler J, Bourboulia D, Casabonne D, Beral V, Mbidde E, Carpenter L, Parkin DM, Wabinga H, Mbulaiteye S, Jaffe H, Weiss R, Boshoff C. 2003b. Infection with Kaposi's sarcoma-associated herpesvirus (KSHV) and human immunodeficiency virus (HIV) in relation to the risk and clinical presentation of Kaposi's sarcoma in Uganda. *Br J Cancer* 89(3): 502-504.
161. Newton R, Carpenter L, Casabonne D, Beral V, Babiker A, Darbyshire J, Weller I, Weiss R, Kwan A, Bourboulia D, Munoz F, Lagos D, Boshoff C. 2006. A prospective study of Kaposi's sarcoma-associated herpesvirus and Epstein-Barr virus in adults with human immunodeficiency virus-1. *Br J Cancer* 94(10): 1504-1509.
162. Noel JC. 1995. Kaposi's sarcoma and KSHV. *Lancet* 346(8986): 1359.
163. Nsubuga MM, Biggar RJ, Combs S, Marshall V, Mbisa G, Kambugu F, Mehta M, Biryahwaho B, Rabkin CS, Whitby D, Mbulaiteye SM. 2008. Human herpesvirus 8 load and progression of AIDS-related Kaposi sarcoma lesions. *Cancer Lett* 263(2): 182-188.
164. NTP. 2015. *Handbook for Preparing the Report on Carcinogens Monographs* Research Triangle Park, NC: National Toxicology Program. 89 pp. [http://ntp.niehs.nih.gov/ntp/roc/handbook/roc\\_handbook\\_508.pdf](http://ntp.niehs.nih.gov/ntp/roc/handbook/roc_handbook_508.pdf).
165. O'Brien TR, Kedes D, Ganem D, Macrae DR, Rosenberg PS, Molden J, Goedert JJ. 1999. Evidence for concurrent epidemics of human herpesvirus 8 and human immunodeficiency virus type 1 in US homosexual men: rates, risk factors, and relationship to Kaposi's sarcoma. *J Infect Dis* 180(4): 1010-1017.
166. Ogoina D, Onyemelukwe GC, Musa BO, Babadoko A. 2011. Human herpesvirus 8 infections and AIDS-associated Kaposi sarcoma in Zaria, Northern Nigeria. *J Int Assoc Physicians AIDS Care (Chic)* 10(1): 43-48.
167. Oksenhendler E, Boulanger E, Galicier L, Du MQ, Dupin N, Diss TC, Hamoudi R, Daniel MT, Agbalika F, Boshoff C, Clauvel JP, Isaacson PG, Meignin V. 2002. High incidence of Kaposi sarcoma-associated herpesvirus-related non-Hodgkin lymphoma in patients with HIV infection and multicentric Castleman disease. *Blood* 99(7): 2331-2336.
168. Oksenhendler E, Boutboul D, Beldjord K, Meignin V, de Labarthe A, Fieschi C, Dossier A, Agbalika F, Parravicini C, Tosato G, Gerard L, Galicier L. 2013. Human herpesvirus 8+ polyclonal IgM $\lambda$  B-cell lymphocytosis mimicking plasmablastic leukemia/lymphoma in HIV-infected patients. *Eur J Haematol* 91(6): 497-503.

169. Pan ZG, Zhang QY, Lu ZB, Quinto T, Rozenvald IB, Liu LT, Wilson D, Reddy V, Huang Q, Wang HY, Ren YS. 2012. Extracavitary KSHV-associated large B-Cell lymphoma: a distinct entity or a subtype of primary effusion lymphoma? Study of 9 cases and review of an additional 43 cases. *Am J Surg Pathol* 36(8): 1129-1140.
170. Panagiotakis GI, Papadogianni D, Chatziioannou MN, Lasithiotaki I, Delakas D, Spandidos DA. 2013. Association of human herpes, papilloma and polyoma virus families with bladder cancer. *Tumour Biol* 34(1): 71-79.
171. Pantry SN, Medveczky PG. 2009. Epigenetic regulation of Kaposi's sarcoma-associated herpesvirus replication. *Semin Cancer Biol* 19(3): 153-157.
172. Parravicini C, Olsen SJ, Capra M, Poli F, Sirchia G, Gao SJ, Berti E, Nocera A, Rossi E, Bestetti G, Pizzuto M, Galli M, Moroni M, Moore PS, Corbellino M. 1997a. Risk of Kaposi's sarcoma-associated herpes virus transmission from donor allografts among Italian posttransplant Kaposi's sarcoma patients. *Blood* 90(7): 2826-2829. (as cited in IARC 2012)
173. Parravicini C, Corbellino M, Paulli M, Magrini U, Lazzarino M, Moore PS, Chang Y. 1997b. Expression of a virus-derived cytokine, KSHV vIL-6, in HIV-seronegative Castleman's disease. *Am J Pathol* 151(6): 1517-1522.
174. Parravicini C, Chandran B, Corbellino M, Berti E, Paulli M, Moore PS, Chang Y. 2000. Differential viral protein expression in Kaposi's sarcoma-associated herpesvirus-infected diseases: Kaposi's sarcoma, primary effusion lymphoma, and multicentric Castleman's disease. *Am J Pathol* 156(3): 743-749.
175. Parry JP, Moore PS. 1997. Corrected prevalence of Kaposi's sarcoma (KS)-associated herpesvirus infection prior to onset of KS. *AIDS* 11(1): 127-128.
176. Pellett PE, Wright DJ, Engels EA, Ablashi DV, Dollard SC, Forghani B, Glynn SA, Goedert JJ, Jenkins FJ, Lee TH, Neipel F, Todd DS, Whitby D, Nemo GJ, Busch MP, for the Retrovirus Epidemiology Donor Study. 2003. Multicenter comparison of serologic assays and estimation of human herpesvirus 8 seroprevalence among US blood donors. *Transfusion* 43(9): 1260-1268.
177. Pelsler C, Middeldorp J, Mbulaiteye SM, Lauria C, Messina A, Viviano E, Romano N, Vitale F, Goedert JJ. 2010. Risk of classical Kaposi sarcoma by plasma levels of Epstein-Barr virus antibodies, sCD26, sCD23 and sCD30. *Infect Agent Cancer* 5: 18.
178. Polizzotto MN, Uldrick TS, Hu D, Yarchoan R. 2012. Clinical manifestations of Kaposi sarcoma herpesvirus lytic activation: multicentric Castleman disease (KSHV-MCD) and the KSHV inflammatory cytokine syndrome. *Front Microbiol* 3: 73.
179. Powles T, Stebbing J, Bazeos A, Hatzimichael E, Mandalia S, Nelson M, Gazzard B, Bower M. 2009. The role of immune suppression and HHV-8 in the increasing incidence of HIV-associated multicentric Castleman's disease. *Ann Oncol* 20(4): 775-779.
180. Purushothaman P, Uppal T, Verma SC. 2015. Molecular biology of KSHV lytic reactivation. *Viruses* 7(1): 116-153.

181. Qin Z, Kearney P, Plaisance K, Parsons CH. 2010. Pivotal advance: Kaposi's sarcoma-associated herpesvirus (KSHV)-encoded microRNA specifically induce IL-6 and IL-10 secretion by macrophages and monocytes. *J Leukoc Biol* 87(1): 25-34.
182. Quinlivan EB, Wang RX, Stewart PW, Kolmoltri C, Regamey N, Erb P, Vernazza PL, Swiss HIVCS. 2001. Longitudinal sero-reactivity to human herpesvirus 8 (KSHV) in the Swiss HIV Cohort 4.7 years before KS. *J Med Virol* 64(2): 157-166.
183. Rabkin CS, Shepherd FA, Wade JA. 1999. Human herpesvirus 8 and renal transplantation. *N Engl J Med* 340(13): 1045-1046. (as cited in IARC 2012)
184. Reddy D, Mitsuyasu R. 2011. HIV-associated multicentric Castleman disease. *Curr Opin Oncol* 23(5): 475-481.
185. Regamey N, Cathomas G, Schwager M, Wernli M, Harr T, Erb P. 1998. High human herpesvirus 8 seroprevalence in the homosexual population in Switzerland. *J Clin Microbiol* 36(6): 1784-1786.
186. Renwick N, Halaby T, Weverling GJ, Dukers NH, Simpson GR, Coutinho RA, Lange JM, Schulz TF, Goudsmit J. 1998. Seroconversion for human herpesvirus 8 during HIV infection is highly predictive of Kaposi's sarcoma. *AIDS* 12(18): 2481-2488.
187. Rezza G, Andreoni M, Dorrucchi M, Pezzotti P, Monini P, Zerboni R, Salassa B, Colangeli V, Sarmati L, Nicastrì E, Barbanera M, Pristera R, Aiuti F, Ortona L, Ensoli B. 1999. Human herpesvirus 8 seropositivity and risk of Kaposi's sarcoma and other acquired immunodeficiency syndrome-related diseases. *J Natl Cancer Inst* 91(17): 1468-1474.
188. Rothman KJ, Greenland S. 2005. Causation and causal inference in epidemiology. *Am J Public Health* 95 Suppl 1: S144-150.
189. Rouhani P, Fletcher CD, Devesa SS, Toro JR. 2008. Cutaneous soft tissue sarcoma incidence patterns in the U.S.: An analysis of 12,114 cases. *Cancer* 113(3): 616-627.
190. Sadeghian MH, Katebi M, Ayatollahi H, Keramati MR. 2008. Immunohistochemical study association between human herpesvirus 8 and multiple myeloma. *Int J Hematol* 88(3): 283-286.
191. Schonrich G, Raftery M, Schnitzler P, Rohr U, Goldschmidt H. 1998. Absence of a correlation between Kaposi's sarcoma-associated herpesvirus (KSHV/HHV-8) and multiple myeloma. *Blood* 92(9): 3474-3475.
192. SEER. 2015. *SEER Stat Fact Sheets: Myeloma*. National Cancer Institute. <http://seer.cancer.gov/statfacts/html/mulmy.html>. Accessed on 9/13/15.
193. Senba M, Buziba N, Mori N, Morimoto K, Nakamura T. 2011. Increased prevalence of Kaposi's sarcoma-associated herpesvirus in the Kaposi's sarcoma-endemic area of western Kenya in 1981-2000. *Acta Virol* 55(2): 161-164.
194. Simbiri KO, Jha HC, Kayembe MK, Kovarik C, Robertson ES. 2014. Oncogenic viruses associated with vulva cancer in HIV-1 patients in Botswana. *Infect Agent Cancer* 9: 28.

195. Simonelli C, Spina M, Cinelli R, Talamini R, Tedeschi R, Gloghini A, Vaccher E, Carbone A, Tirelli U. 2003. Clinical features and outcome of primary effusion lymphoma in HIV-infected patients: a single-institution study. *J Clin Oncol* 21(21): 3948-3954.
196. Sin SH, Dittmer DP. 2013. Viral latency locus augments B-cell response in vivo to induce chronic marginal zone enlargement, plasma cell hyperplasia, and lymphoma. *Blood* 121(15): 2952-2963.
197. Sin SH, Kim Y, Eason A, Dittmer DP. 2015. KSHV Latency Locus Cooperates with Myc to Drive Lymphoma in Mice. *PLoS Pathog* 11(9): e1005135.
198. Sitas F, Carrara H, Beral V, Newton R, Reeves G, Bull D, Jentsch U, Pacella-Norman R, Bourboulia D, Whitby D, Boshoff C, Weiss R. 1999. Antibodies against human herpesvirus 8 in black South African patients with cancer. *N Engl J Med* 340(24): 1863-1871.
199. Sitas F, Newton R. 2001. Kaposi's sarcoma in South Africa. *J Natl Cancer Inst Monogr*(28): 1-4.
200. Smith NA, Sabin CA, Gopal R, Bourboulia D, Labbet W, Boshoff C, Barlow D, Band B, Peters BS, de Ruiter A, Brown DW, Weiss RA, Best JM, Whitby D. 1999. Serologic evidence of human herpesvirus 8 transmission by homosexual but not heterosexual sex. *J Infect Dis* 180(3): 600-606.
201. Sodhi A, Montaner S, Patel V, Zohar M, Bais C, Mesri EA, Gutkind JS. 2000. The Kaposi's sarcoma-associated herpes virus G protein-coupled receptor up-regulates vascular endothelial growth factor expression and secretion through mitogen-activated protein kinase and p38 pathways acting on hypoxia-inducible factor 1alpha. *Cancer Res* 60(17): 4873-4880.
202. Soulier J, Grollet L, Oksenhendler E, Cacoub P, Cazals-Hatem D, Babinet P, d'Agay MF, Clauvel JP, Raphael M, Degos L, et al. 1995. Kaposi's sarcoma-associated herpesvirus-like DNA sequences in multicentric Castleman's disease. *Blood* 86(4): 1276-1280.
203. Stiller CA, Trama A, Brewster DH, Verne J, Bouchardy C, Navarro C, Chirlaque MD, Marcos-Gragera R, Visser O, Serraino D, Weiderpass E, Dei Tos AP, Ascoli V, Group RW. 2014. Descriptive epidemiology of Kaposi sarcoma in Europe. Report from the RARECARE project. *Cancer Epidemiol* 38(6): 670-678.
204. Su CC, Tseng KC, Lin MN, Hsieh TC, Chu TY. 2014. High seroprevalence of human herpesvirus type 8 in patients with hepatocellular carcinoma. *Eur J Clin Microbiol Infect Dis*.
205. Sullivan RJ, Pantanowitz L, Casper C, Stebbing J, Dezube BJ. 2008. HIV/AIDS: epidemiology, pathophysiology, and treatment of Kaposi sarcoma-associated herpesvirus disease: Kaposi sarcoma, primary effusion lymphoma, and multicentric Castleman disease. *Clin Infect Dis* 47(9): 1209-1215.
206. Sutcliffe S, Till C, Jenkins FJ, Gaydos CA, Goodman PJ, Hoque AM, Hsing AW, Thompson IM, Nelson WG, De Marzo AM, Platz EA. 2015. Prospective study of human

- herpesvirus type 8 serostatus and prostate cancer risk in the placebo arm of the Prostate Cancer Prevention Trial. *Cancer Causes Control* 26(1): 35-44.
207. Swerdlow SH, Campo E, Harris NL, Jaffe ES, Pieleri SA, Stein H, Thiele J, Vardiman JW. 2008. *WHO Classification of Tumours* 4th ed., vol. 2, Geneva, Switzerland: World Health Organization.
  208. Szalai E, Gerlei Z, Szlavik J, Szladek G, Patel R, Hunyadi J, Gergely L, Juhasz A. 2005. Prevalence of human herpesvirus-8 infection in HIV-positive patients with and without Kaposi's sarcoma in Hungary. *FEMS Immunol Med Microbiol* 43(2): 265-268.
  209. Talat N, Schulte KM. 2011. Castleman's disease: systematic analysis of 416 patients from the literature. *Oncologist* 16(9): 1316-1324.
  210. Tedeschi R, Kvarnung M, Knekt P, Schulz TF, Szekely L, De Paoli PD, Aromaa A, Teppo L, Dillner J. 2001. A prospective seroepidemiological study of human herpesvirus-8 infection and the risk of multiple myeloma. *Br J Cancer* 84(1): 122-125.
  211. Tedeschi R, Luostarinen T, De Paoli P, Gislefoss RE, Tenkanen L, Virtamo J, Koskela P, Hallmans G, Lehtinen M, Dillner J. 2005. Joint Nordic prospective study on human herpesvirus 8 and multiple myeloma risk. *Br J Cancer* 93(7): 834-837.
  212. Testa A, Baiocchini A, Comandini UV, Falasca L, Nardacci R, Maritti M, Loiacono L, Bibbolino C, Rizzi EB, Cristofaro M, Ettorre GM, Vennarecci G, Antonucci G, Del Nonno F. 2010. Fatal sclerosing peritonitis associated with primary effusion lymphoma after liver transplantation: a case report. *Transplant Proc* 42(9): 3849-3853.
  213. Tiussi RM, Caus AL, Diniz LM, Lucas EA. 2012. Kaposi's sarcoma: clinical and pathological aspects in patients seen at the Hospital Universitario Cassiano Antonio Moraes - Vitoria - Espirito Santo - Brazil. *An Bras Dermatol* 87(2): 220-227.
  214. Uldrick TS, Whitby D. 2011. Update on KSHV epidemiology, Kaposi sarcoma pathogenesis, and treatment of Kaposi sarcoma. *Cancer Lett* 305(2): 150-162.
  215. Uldrick T, Polizzotto MN, Wyvill K, Aleman K, Marshall V, Little R, Filie A, Raffeld M, Steinberg SM, Pittaluga S, Whitby D, Yarchoan R. 2012. Kaposi sarcoma herpesvirus (KSHV)-associated lymphomas are associated with markedly elevated serum IL-10, elevated IL-6, IL-17 and circulating KSHV. *Infect Agent Cancer* 7(Suppl 1): 39.
  216. van der Kuyl AC, Polstra AM, van den Burg R, Jan Weverling G, Goudsmit J, Cornelissen M. 2005. Cytomegalovirus and human herpesvirus 8 DNA detection in peripheral blood monocyctic cells of AIDS patients: correlations with the presence of Kaposi's sarcoma and CMV disease. *J Med Virol* 76(4): 541-546.
  217. Venkataraman G, Uldrick TS, Aleman K, O'Mahony D, Karcher DS, Steinberg SM, Raffeld MA, Marshall V, Whitby D, Little RF, Yarchoan R, Pittaluga S, Maric I. 2013. Bone marrow findings in HIV-positive patients with Kaposi sarcoma herpesvirus-associated multicentric Castleman disease. *Am J Clin Pathol* 139(5): 651-661.

218. Wakeham K, Johnston WT, Nalwoga A, Webb EL, Mayanja BN, Miley W, Elliott AM, Whitby D, Newton R. 2015. Trends in Kaposi's sarcoma-associated Herpesvirus antibodies prior to the development of HIV-associated Kaposi's sarcoma: a nested case-control study. *Int J Cancer* 136(12): 2822-2830.
219. Wang QJ, Jenkins FJ, Jacobson LP, Kingsley LA, Day RD, Zhang ZW, Meng YX, Pellett PE, Kousoulas KG, Baghian A, Rinaldo CR, Jr. 2001. Primary human herpesvirus 8 infection generates a broadly specific CD8(+) T-cell response to viral lytic cycle proteins. *Blood* 97(8): 2366-2373.
220. Wen KW, Damania B. 2010. Kaposi sarcoma-associated herpesvirus (KSHV): molecular biology and oncogenesis. *Cancer Lett* 289(2): 140-150.
221. Westrop SJ, Lagos D, Boshoff C, Bower M, Imami N. 2012. African ancestry and innate immunity contribute to the incidence of multicentric Castleman's disease in HIV-1/Kaposi's sarcoma herpesvirus-coinfected individuals. *Future Virol* 7(7): 729-734.
222. Whitby D, Luppi M, Barozzi P, Boshoff C, Weiss RA, Torelli G. 1998. Human herpesvirus 8 seroprevalence in blood donors and lymphoma patients from different regions of Italy. *J Natl Cancer Inst* 90(5): 395-397.
223. Whitby D, Howard MR, Tenant-Flowers M, Brink NS, Copas A, Boshoff C, Hatzioannou T, Suggett FE, Aldam DM, Denton AS, *et al.* 1995. Detection of Kaposi sarcoma associated herpesvirus in peripheral blood of HIV-infected individuals and progression to Kaposi's sarcoma. *Lancet* 346(8978): 799-802.
224. Widmer IC, Erb P, Grob H, Itin P, Baumann M, Stalder A, Weber R, Cathomas G. 2006. Human herpesvirus 8 oral shedding in HIV-infected men with and without Kaposi sarcoma. *J Acquir Immune Defic Syndr* 42(4): 420-425.
225. Wu TT, Qian J, Ang J, Sun R. 2012. Vaccine prospect of Kaposi sarcoma-associated herpesvirus. *Curr Opin Virol* 2(4): 482-488.
226. Xiao J, Selvaggi SM, Leith CP, Fitzgerald SA, Stewart J, 3rd. 2013. Kaposi sarcoma herpesvirus/human herpesvirus-8-negative effusion-based lymphoma: report of 3 cases and review of the literature. *Cancer Cytopathol* 121(11): 661-669.
227. Xu GJ, Kula T, Xu Q, Li MZ, Vernon SD, Ndung'u T, Ruxrungtham K, Sanchez J, Brander C, Chung RT, O'Connor KC, Walker B, Larman HB, Elledge SJ. 2015. Viral immunology. Comprehensive serological profiling of human populations using a synthetic human virome. *Science* 348(6239): aaa0698.
228. Yang TY, Chen SC, Leach MW, Manfra D, Homey B, Wiekowski M, Sullivan L, Jenh CH, Narula SK, Chensue SW, Lira SA. 2000. Transgenic expression of the chemokine receptor encoded by human herpesvirus 8 induces an angioproliferative disease resembling Kaposi's sarcoma. *J Exp Med* 191(3): 445-454.
229. Yiakoumis X, Pangalis GA, Kyrtsonis MC, Vassilakopoulos TP, Kontopidou FN, Kalpadakis C, Korkolopoulou P, Levidou G, Androulaki A, Siakantaris MP, Sachanas S,



- Andreopoulos A. 2010. Primary effusion lymphoma in two HIV-negative patients successfully treated with pleurodesis as first-line therapy. *Anticancer Res* 30(1): 271-276.
230. Zago A, Bourboulia D, Viana MC, Collandre H, Dietze R, Boshoff C, Keller R. 2000. Seroprevalence of human herpesvirus 8 and its association with Kaposi sarcoma in Brazil. *Sex Transm Dis* 27(8): 468-472.
231. Zhang H, Yang XY, Hong T, Feldman T, Bhattacharyya PK. 2010. Kaposi sarcoma-associated herpesvirus (human herpesvirus type 8)-associated extracavitary lymphoma: Report of a case in an HIV-positive patient with simultaneous Kaposi sarcoma and a review of the literature. *Acta Haematol* 123(4): 237-241.
232. Zhu X, Guo Y, Yao S, Yan Q, Xue M, Hao T, Zhou F, Zhu J, Qin D, Lu C. 2014. Synergy between Kaposi's sarcoma-associated herpesvirus (KSHV) vIL-6 and HIV-1 Nef protein in promotion of angiogenesis and oncogenesis: role of the AKT signaling pathway. *Oncogene* 33(15): 1986-1996.
233. zur Hausen H. 1994. Molecular pathogenesis of cancer of the cervix and its causation by specific human papillomavirus types. *Curr Top Microbiol Immunol* 186: 131-156.
234. zur Hausen H. 2001. Oncogenic DNA viruses. *Oncogene* 20(54): 7820-7823.
235. zur Hausen H, de Villiers EM. 2014. Cancer "causation" by infections--individual contributions and synergistic networks. *Semin Oncol* 41(6): 860-875.

## Glossary

**Case report:** Detailed descriptions of a few patients or clinical cases (frequently, just one sick person) with an unusual disease or complication, uncommon combinations of diseases, an unusual or misleading semiology, cause, or outcome (maybe a surprising recovery). They often are preliminary observations that are later refuted. They cannot estimate disease frequency or risk (e.g., for lack of a valid denominator).

**Case series:** A collection of subjects (usually, patients) with common characteristics used to describe some clinical, pathophysiological, or operational aspect of a disease, treatment, exposure, or diagnostic procedure. A case series does not include a comparison group and is often based on prevalent cases and on a sample of convenience. Common selection biases and confounding severely limit their power to make causal inferences.

**Case-comparison study (case-control study, case referent study):** The observational epidemiological study of persons with the disease (or another outcome variable) of interest and a suitable control group of persons without the disease (comparison group, reference group). The potential relationship of a suspected risk factor or an attribute to the disease is examined by comparing the diseased and non-diseased subjects with regard to how frequently the factor or attribute is present (or, if quantitative, the levels of the attribute) in each of the groups (diseased and non-diseased).

**Cellular immunity:** Immunity independent of antibody but dependent on the recognition of antigen by T cells and their subsequent destruction of cells bearing the antigen or on the secretion by T cells of lymphokines that enhance the ability of phagocytes to eliminate the antigen.

**Cofactor:** A factor that activates or enhances the action of another entity such as a disease-causing agent. Cofactors may influence the progression of a disease or the likelihood of becoming ill.

**Diagnostic criteria:** The specific combination of signs, symptoms, and test results that a clinician uses to identify a person as representing a case of a particular disease or condition.

**Enzyme immunoassay:** An assay that uses an enzyme-bound antibody to detect antigen. The enzyme catalyzes a color reaction when exposed to substrate.

**Highly active antiretroviral therapy:** Treatment regimens that stop or slow the HIV virus from reproducing and keep HIV disease from progressing. The usual HAART regimen combines 3 or more HIV drugs from at least 2 different classes.

**Horizontal transmission:** The spread of an infectious agent from one individual to another, usually through contact with bodily excretions or fluids, such as sputum or blood, which contains the agent.

**Humoral immunity:** The component of the immune system involving antibodies that are secreted by B-cells and circulate as soluble proteins in blood.

**Immunohistochemical staining:** A method to detect specific antigens in cells based on an antigen-antibody reaction, which can be recognized at the light microscopic level.

**Latent phase:** A phase of the virus life cycle during which the virus is not replicating.

**Lytic phase:** A phase of the virus life cycle during which the virus replicates within the host cell, releasing a new generation of viruses when the infected cell lyses.

**Lytic reactivation:** The biological events beginning with emergence of a virus from latency and ending with lysis of the host cell and release of progeny virions.

**microRNA:** Small, non-coding RNA molecules approximately 22 nucleotides in length that act post translationally in a regulatory role to target messenger RNAs for cleavage or translational expression.

**Monoclonal:** Pertaining to or designating a group of identical cells or organisms derived from a single cell or organism.

**Non-coding RNAs:** Functional RNA molecules that are transcribed from DNA but are not translated into proteins.

**Oligoclonal:** A few different clones, or the product of a few different clones.

**Paracrine neoplasia:** A process whereby KSHV-infected cells induce neighboring uninfected cells to produce cytokines and growth factors that are necessary as autocrine and paracrine factors for driving tumor production.

**Parenteral:** By some other means than through the gastrointestinal tract; the parenteral route of infection involves breaks in the skin such as cuts and scrapes, puncture wounds, bites and burns.

**Peripheral blood monocytes:** Circulating precursors of macrophages and dendritic cells that migrate from the blood stream across vascular endothelium for immunological surveillance, as well as respond to inflammation.

**Polyclonal:** Pertaining to or designating a group of cells or organisms derived from several cells.

**Polymerase chain reaction:** A laboratory technique used to produce large amounts of specific DNA fragments. Polymerase chain reaction is used for genetic testing and to diagnose disease.

**Professional antigen-presenting cells (APCs):** Specialized white blood cells that express MHC class II and are able to activate a helper T-cell that has never encountered its antigen before. The three main types of professional APCs are macrophages, dendritic cells, and B cells.

**Titer:** A laboratory measurement of the concentration of a substance in a solution (e.g., an antibody titer measures the presence and amount of antibodies in the blood).

**Toll-like receptors:** A family of pattern recognition receptors involved in the detection of structures associated with pathogens or damaged host tissues.

**Vertical transmission:** The transmission of infection from one generation to the next (e.g., from mother to infant prenatally, during delivery, or in the postnatal period via breast milk).

## Abbreviations

|                  |  |
|------------------|--|
| AIDS:            | Acquired Immune Deficiency Syndrome            |
| ANGP2:           | angiopoietin 2                                 |
| APC:             | antigen-presenting cell                        |
| BET:             | bromodomain and extra-terminal                 |
| CDC:             | Centers for Disease Control and Prevention     |
| CDK6:            | cyclin-dependent kinase 6                      |
| CI:              | confidence interval                            |
| DNA:             | deoxyribonucleic acid                          |
| DOT:             | Department of Transportation                   |
| dsDNA:           | double-stranded DNA                            |
| EBNA:            | Epstein-Barr virus nuclear antigen             |
| EBV:             | Epstein-Barr virus                             |
| EBV VCA:         | EBV viral capsid antigen                       |
| ELISA:           | enzyme-linked immunosorbent assays             |
| F:               | female   |
| FDA:             | Food and Drug Administration                   |
| FLICE:           | FADD-like interleukin-1 beta-converting enzyme |
| HAART:           | highly active antiretroviral therapy           |
| HAX-1:           | HS1-associated protein X1                      |
| HHV8:            | human herpesvirus 8                            |
| HIF-1 $\alpha$ : | hypoxia-inducible factor-1 $\alpha$            |
| HIV:             | human immunodeficiency virus                   |
| HR:              | hazard ratio                                   |
| hTERT:           | human telomerase reverse transcriptase         |
| IARC:            | International Agency for Research on Cancer    |
| IFA:             | immunofluorescence assay                       |
| HHV8:            | human herpesvirus 8                            |
| IFA:             | immunofluorescence assay                       |
| K1:              | unique KSHV protein 1                          |
| K15:             | unique KSHV protein 15                         |
| KS:              | Kaposi sarcoma                                 |

|                 |  |
|-----------------|--|
| KSHV:           | Kaposi sarcoma-associated herpesvirus                          |
| LANA:           | latency-associated nuclear antigen                             |
| MHC:            | major histocompatibility complex                               |
| miRNA:          | microRNA   |
| M:              | male   |
| Max:            | maximum  |
| MCD:            | multicentric Castleman disease                                 |
| Med:            | median   |
| Min:            | minimum  |
| MM:             | multiple myeloma   |
| MSM:            | men who have sex with men                                      |
| Nef:            | negative factor protein  |
| NF- $\kappa$ B: | nuclear factor kappa-light-chain enhancer of activated B cells |
| NHANES:         | National Health and Examination Survey                         |
| NR:             | not reported   |
| NS:             | not significant  |
| NTP:            | National Toxicology Program                                    |
| OR:             | odds ratio   |
| ORF:            | open reading frames  |
| OSHA:           | Occupational Safety and Health Administration                  |
| PAN:            | polyadenylated nuclear   |
| PBMC:           | peripheral blood mononuclear cells                             |
| PCR:            | polymerase chain reaction                                      |
| PDGF:           | platelet-derived growth factor                                 |
| PEL:            | primary effusion lymphoma                                      |
| pRB:            | retinoblastoma protein   |
| PTK:            | protein tyrosine kinase  |
| RARECARE:       | Surveillance of Rare Cancers in Europe                         |
| RNA:            | ribonucleic acid   |
| RR:             | relative risk  |
| RTA:            | replication and transcription activator                        |
| SEER:           | Surveillance, Epidemiology, and End Results Program            |

---

|          |                                     |
|----------|-------------------------------------|
| TGFBeta: | transforming growth factor- $\beta$ |
| TLR4     | toll-like receptor 4                |
| U.K. :   | United Kingdom                      |
| U.S. :   | United States                       |
| U.S.A.:  | United States of America            |
| vCCL:    | viral-encoded chemokines            |
| VEGF:    | vascular endothelial growth factor  |
| vFLIP:   | viral FLICE-inhibitory protein      |
| vGPCR:   | viral G protein-coupled receptor    |
| vIL-6:   | interleukin-6                       |
| vIRF:    | viral interferon regulatory factors |
| WB:      | western blot assay                  |

**This Page Intentionally Left Blank**

## Appendix A: Literature Search Strategy

The objective of the literature search approach is to identify published literature that is relevant for evaluating the potential carcinogenicity of the Kaposi sarcoma-associated herpes virus (KSHV). As discussed in the Viruses Concept Document ([https://ntp.niehs.nih.gov/ntp/roc/concept\\_docs/2014/virusesconcept\\_508.pdf](https://ntp.niehs.nih.gov/ntp/roc/concept_docs/2014/virusesconcept_508.pdf)), the monograph relies on the IARC monograph and studies published since the monograph (new studies). The literature search strategy was used to identify new human cancer studies and recent reviews of mechanistic data.

### General approach

Database searching encompasses selecting databases and search terms and conducting the searches. Searches of several citation databases are generally conducted using search terms for the individual viruses of interest, combined with search terms for cancer and/or specific topics, including epidemiological and mechanistic studies. A critical step in the process involves consultation with an information specialist to develop relevant search terms. These terms are used to search bibliographic databases. IARC used literature found by searching PubMed for KSHV through 12/2008, so PubMed, Web of Science and Scopus were searched for new information about HTLV from > 2008 to August 2015. Table 1 highlights the general concepts searched with selected example terms. To review all the terms used, please refer the to full search strings below.

**Table A-1. Major topics searched**

| Topics                                   | Example terms  |
|--|--|
| Kaposi's sarcoma-associated herpes virus | Herpesvirus 8, human(Mesh), Kaposi's sarcoma-associated herpesvirus, human herpesvirus 8, KSHV, HHV8 |

The literature for KSHV was searched without using narrowing terms within the bibliographic databases. The results were then processed in EndNote to remove duplicates before being transferred to DistillerSR for screening.

The bibliographic database search results (3314) were processed in Endnote then imported into DistillerSR for first and second tier screening. Relevant studies found through the citations of review articles and other secondary searched were also included. Tagging in DistillerSR categorized the useful articles into Human Epidemiologic literature (212) or Mechanistic literature (704).



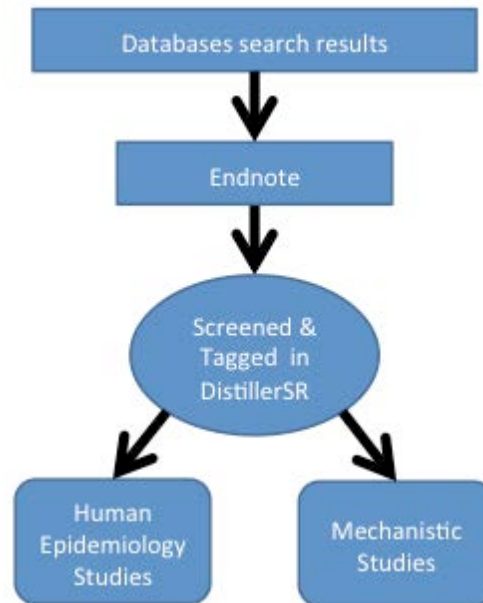


Figure A-1. Literature processing flow

#### Search strings for KSHV Searches

Pubmed: 2008-2015

"Herpesvirus 8, human"[mh] OR "Kaposi's sarcoma-associated herpes virus"[tiab] OR "Kaposi's sarcoma-associated herpesvirus"[tiab] OR "Kaposi sarcoma-associated herpes virus"[tiab] OR "Kaposi sarcoma-associated herpesvirus"[tiab] OR KSHV[tiab] OR "human herpesvirus 8"[tiab] OR "human herpes virus 8"[tiab] OR HHV8[tiab] OR "HHV-8"[tiab]

Web of Science and Scopus: 2008-2015

"Kaposi's sarcoma-associated herpes virus" OR "Kaposi's sarcoma-associated herpesvirus" OR KSHV OR "human herpesvirus 8" OR "human herpes virus 8" OR HHV8

---

**Part 2**

---

**Cancer Hazard Profile**

---

**This Page Intentionally Left Blank**

---

## Kaposi Sarcoma-Associated Herpesvirus

CAS No.: none assigned

Known to be a human carcinogen<sup>1</sup>

Also known as KSHV or human herpesvirus 8 (HHV-8)

### Carcinogenicity

Kaposi sarcoma-associated herpesvirus (KSHV) is *known to be a human carcinogen* based on sufficient evidence from studies in humans. This conclusion is based on evidence from epidemiological and molecular studies, which show that KSHV causes Kaposi sarcoma, primary effusion lymphoma, and a plasmablastic variant of multicentric Castleman disease, and on supporting mechanistic data. KSHV causes cancer, primarily but not exclusively in people with suppressed immune systems, by coding for protein and RNA products that work together to transform host cells into cancer cells and promote their survival and growth. These viral products are made (through the use of host-cell machinery) both when KSHV is in the lytic phase (destroying the infected cell during replication) and when it is latent (maintaining its DNA in the infected cell without destroying the cell) (Mesri *et al.* 2010, Fukumoto *et al.* 2011). KSHV virus is latent in most infected cells.

### Cancer Studies in Humans

The majority of human cancer studies of KSHV have focused on Kaposi sarcoma (a cancer of the cells that line blood or lymph vessels). Kaposi sarcoma has four main subtypes: (1) epidemic, or related to human immunodeficiency virus type 1 (HIV-1) infection, (2) iatrogenic (resulting from medical treatment, such as organ transplants), (3) classic (a slow-growing form found mostly in older men in specific populations, such as in Mediterranean countries or among East Europeans of Jewish descent), and (4) endemic (found in sub-Saharan Africa, mostly among men but also among children).

Other cancer end points, including two rare B-cell non-Hodgkin lymphomas (primary effusion lymphoma and multicentric Castleman disease), also have been linked to KSHV. Primary effusion lymphoma (also called body-cavity-based B-cell lymphoma) arises in a specific type of immune cell (B lymphocytes) and comprises approximately 2% to 4% of HIV-1-related non-Hodgkin lymphomas (Simonelli *et al.* 2003, Sullivan *et al.* 2008). Multicentric Castleman disease also arises in B lymphocytes; it has several histological variants (hyaline vascular, plasma cell, mixed, and plasmablastic), and KSHV is associated with the plasmablastic variant.

### Kaposi Sarcoma

Evidence for an association between KSHV infection and Kaposi sarcoma is based on consistent findings of increased risk in epidemiological studies with different designs and in different populations and on the presence of an exposure-response relationship between the degree of viral infection and the cancer.

Over 90% of Kaposi sarcoma patients are infected with KSHV, and KSHV DNA is found in virtually all Kaposi sarcoma tumors; KSHV is therefore considered to be a prerequisite for diagnosis of this cancer (Chang *et al.* 1994, Mesri *et al.* 2010, Cavallin *et al.* 2014). The

---

<sup>1</sup>NTP listing recommendation for the RoC.

association between KSHV and Kaposi sarcoma has been evaluated in about 80 case-control studies and 25 cohort studies or studies of cases and controls identified within the cohorts (IARC 2012, NTP 2016). In the case-control studies, patients with all four types of Kaposi sarcoma were often 10 times and sometimes over 100 times more likely to be infected with KSHV than were individuals without Kaposi sarcoma. In the cohort studies, KSHV-infected individuals were 2 to 16 times more likely to develop Kaposi sarcoma than were uninfected individuals. In some studies, the risk of Kaposi sarcoma increased with increasing viral load of KSHV (Sitas *et al.* 1999, Newton *et al.* 2003a,b, 2006, Albrecht *et al.* 2004).

Most KSHV-infected patients who develop Kaposi sarcoma have immune systems compromised either by HIV-1 infection or as a result of drug treatments after organ or tissue transplants. The timing of infection with HIV-1 may also play a role in development of Kaposi sarcoma. Acquiring HIV-1 infection prior to KSHV infection may increase the risk of epidemic Kaposi sarcoma by 50% to 100%, compared with acquiring HIV-1 infection at the same time as or after KSHV infection. Nevertheless, patients with the classic or endemic forms of Kaposi sarcoma are not suspected of having suppressed immune systems, suggesting that immunosuppression is not required for development of Kaposi sarcoma.

#### *Primary Effusion Lymphoma*

Evidence for an association between KSHV infection and primary effusion lymphoma is based on consistent findings from over 115 KSHV-infected patients reported in a large number of case reports, several small case series, and a case-comparison study (IARC 2012, see also Section 3, Human Cancer Studies in Part 1: Cancer Hazard Evaluation), together with histological confirmation of the tumors' specific morphological and immunological features. These cases of primary effusion lymphoma were identified in different populations, including both HIV-1-positive and HIV-1-negative patients, organ-transplant recipients, and people from areas where the risk of endemic or classic Kaposi sarcoma is known to be high (Dotti *et al.* 1999, Boulanger *et al.* 2008, Testa *et al.* 2010, IARC 2012). In addition, some patients with primarily effusion lymphoma also had other KSHV-associated cancers, including Kaposi sarcoma and multicentric Castleman disease. The strong association of KSHV with primary effusion lymphoma led to the adoption of the presence of KSHV infection as a diagnostic criterion for this specific pathologic entity. Primary effusion lymphoma in HIV-1-positive patients exhibits rapid progression with short survival times, whereas progression in HIV-1-negative patients and patients with normal immune responses appears to be much slower (IARC 1997), suggesting that immunosuppression is an important cofactor in KSHV carcinogenicity.

Studies of tumor tissue provide evidence for a causal role of KSHV in primary effusion lymphoma. These studies have shown that primary effusion lymphoma cells contain high levels (50 to 100 copies) of KSHV DNA and have patterns of viral gene products (RNA or protein) very similar to those observed in Kaposi sarcoma. Importantly, primary effusion lymphoma lesions arise from a single KSHV-infected B cell (i.e., are monoclonal), suggesting that KSHV infection precedes tumor growth (Katano *et al.* 1999, Judde *et al.* 2000, Fukumoto *et al.* 2011, IARC 2012, Giffin and Damania 2014).

#### *Multicentric Castleman Disease*

Two types of evidence establish a link between KSHV infection and the plasmablastic variant of multicentric Castleman disease. The first consists of nine case-series studies and four case-comparison studies, which provide, although not conclusively, evidence for an association

between KSHV infection and all types of multicentric Castleman disease, and the second consists of molecular analysis of this type of the tumor tissue, which demonstrates that the association between KSHV infection and multicentric Castleman disease is strongest for the plasmablastic variant of this disease (see Section 3, Human Cancer Studies in Part 1: Cancer Hazard Evaluation). (The plasmablastic variant arises in plasmablasts, which are immature precursors of antibody-producing B lymphocytes.)

KSHV has been detected in about half of all multicentric Castleman disease cases (all variants) reported in the literature, but has only rarely been detected in patients without cancer (i.e., in only 1 of almost 200 controls in the case-comparison studies). It is more common among multicentric Castleman disease patients who are also infected with HIV-1 than in HIV-1-negative patients, suggesting that immunosuppression is important for cancer development (Parravicini *et al.* 1997a, Oksenhendler *et al.* 2002, 2013). Moreover, the level of KSHV DNA in circulating white blood cells or blood plasma is related to the occurrence of symptoms during multicentric Castleman disease flares in HIV-1-positive individuals (Reddy and Mitsuyasu 2011).

Studies characterizing the tumor tissue have shown that plasmablasts in KSHV-associated multicentric Castleman disease have a unique molecular profile and produce a distinctive monotypic form of immunoglobulin M; these plasmablasts are not found in KSHV-negative multicentric Castleman disease (Dupin *et al.* 2000). Therefore, KSHV-associated (plasmablastic) multicentric Castleman disease is now recognized as an entity distinct from other forms of multicentric Castleman disease; it is classified by the World Health Organization as “a large B-cell lymphoma arising in HHV8- [KSHV-] associated multicentric Castleman disease” (IARC 2008). In KSHV-associated multicentric Castleman disease cells, it appears that KSHV proteins are produced both when the virus is latent and when it is replicating. These proteins include a virally encoded interleukin 6, which stimulates proliferation of mature B lymphocytes and causes inflammation, and thus may play a role in carcinogenicity (Aoki *et al.* 2001, Burbelo *et al.* 2010, Fukumoto *et al.* 2011, Giffin and Damania 2014).

### **Studies on Mechanisms of Carcinogenesis**

KSHV-associated cancer develops through a complex process that involves interactions among many viral, host, and environmental factors (Fukumoto *et al.* 2011, Mesri *et al.* 2014). Both *in vitro* and animal models have been developed that accurately reproduce many features observed in KSHV-associated cancer (An *et al.* 2006, Mutlu *et al.* 2007, Mesri and Cesarman 2011, Ashlock *et al.* 2014, Dittmer *et al.* 2015); however, not all aspects of viral infection and transformation into cancer cells are well understood (Fukumoto *et al.* 2011, Giffin and Damania 2014).

In an immune-compromised individual, KSHV-infected cells escape recognition and destruction by the immune system and are able to produce cancer-causing viral RNA or proteins (Cavallin *et al.* 2014, Mesri *et al.* 2014). Viral products made during the lytic phase mimic or disrupt host cytokine signals (communication among cells), creating conditions that promote tumor growth, proliferation of latently infected cells, development of blood vessels, inflammation, and evasion or alteration of the immune response and antiviral response (Mesri *et al.* 2010, Wen and Damania 2010, Fukumoto *et al.* 2011, Cavallin *et al.* 2014). In addition, some latently infected cells are less likely to provoke an immune response and may progressively transform into cancer cells through inhibition of programmed cell death (apoptosis) and

maintenance of viral latency (see also Section 3, Mechanisms and Other Relevant Data in Part 1: Cancer Hazard Evaluation).

### **Biological Properties**

KSHV is an enveloped double-stranded DNA gamma-2 herpesvirus (rhadinovirus) that was first identified in humans in 1994 in association with acquired immunodeficiency syndrome (AIDS) (Chang *et al.* 1994, IARC 1997, Fukumoto *et al.* 2011). A lipid membrane envelope surrounds a layer of viral proteins, which encloses a viral capsid (protein shell) and a linear 165-kb genome (IARC 1997, 2012, Fukumoto *et al.* 2011, Giffin and Damania 2014). KSHV infects many types of cells, including both immune cells (B lymphocytes, dendritic cells, and monocytes) and non-immune cells (keratinocytes, fibroblasts, and prostate cells) (IARC 1997, 2012, Fukumoto *et al.* 2011, Campbell *et al.* 2014, Giffin and Damania 2014). Certain immune cells (CD19<sup>+</sup> B lymphocytes) are a long-term reservoir for the latent virus. KSHV glycoproteins bind to several host-cell receptors and initiate viral entry by an inward folding of the host cell membrane to form a small capsule within the cell that contains the virus attached to its receptor (Giffin and Damania 2014). The virus can reproduce through the lytic cycle, which destroys the infected cell, or can remain latent as a viral episome, consisting of circular DNA separate from the infected cell's chromosomes, which can use the cell's machinery to replicate along with the infected cell's own DNA.

### **Detection**

KSHV is detected most commonly by measurement of anti-KSHV antibodies, and also by detection of DNA and viral proteins in tissues (Parravicini *et al.* 1997b, 2000, Fukumoto *et al.* 2011, Bhutani *et al.* 2015, Xu *et al.* 2015). Tests are available for detecting antibodies to proteins produced in the latent and lytic phases. These tests vary in sensitivity and specificity, but have generally improved over time. Different viral proteins are made during different phases of the viral life cycle (latent or lytic), and an individual's antibody response to these proteins varies, which makes it difficult to compare the prevalence of KSHV in different populations (IARC 1997, 2012, Fukumoto *et al.* 2011). Viral DNA can be detected in tumor tissue by polymerase chain reaction (IARC 1997, Fukumoto *et al.* 2011, Campbell *et al.* 2014).

### **Exposure**

Prevalence studies measuring antibodies to KSHV in serum have shown that a significant number of people in the United States are infected with KSHV. In the first systematic evaluation of KSHV epidemiology in the U.S. general population (based on tests for KSHV antibodies in serum samples from the Third National Health and Nutrition Examination Survey, 1988–1994), overall prevalence of KSHV antibodies was approximately 7% and was similar in men and women (Engels *et al.* 2007). A previous study of 1,000 U.S. blood donors (sampled in 1994 and 1995) reported estimated prevalence of KSHV antibodies ranging from 0.5% to 5% (Pellett *et al.* 2003, IARC 2012). KSHV prevalence rates appear to vary widely among different populations, from 2% to 3% in northern Europe to over 50% in some sub-Saharan African populations (IARC 2012). However, even outside of areas where KSHV is endemic, prevalence rates among men having sex with men have been reported in the range of 30% to 60% for HIV-1-positive men and 20% to 30% for HIV-1-negative men (Martin *et al.* 1998, O'Brien *et al.* 1999, Phillips *et al.* 2008).

## **Transmission**

KSHV is thought to be transmitted primarily via saliva (IARC 2012). The presence of KSHV in peripheral blood suggests that it can also be spread via blood, and transmission has been reported in injection drug users, in transfusion recipients, and from organ-transplant donors to recipients (Barozzi *et al.* 2003, IARC 2012). In populations in which KSHV infection is endemic, it can be transmitted from mother to child, especially among children between the ages of 6 and 10 years, and infection rates increase with age. Risk factors for infection may include contact with infected family members, contact with contaminated water, and, in particular, HIV-1 infection (IARC 2012). There is also some evidence for spouse-to-spouse transmission among heterosexual couples, which appears to be more efficient from female to male than male to female (Dupuy *et al.* 2009).

Factors that increase the risk of HIV-1 infection (e.g., number of sexual partners) also increase the risk of infection with KSHV (Smith *et al.* 1999, Engels *et al.* 2007, IARC 2012), and orogenital sex has been shown to be significantly correlated with development of KSHV antibodies in men who have sex with men (Dukers *et al.* 2000). It is unclear whether KSHV is sexually transmitted in heterosexuals. It has also been suggested that application of virus-carrying saliva to the sites of insect bites (to relieve itching) could facilitate the transmission of KSHV (Coluzzi *et al.* 2003, Amodio *et al.* 2011).

## **Diseases (Non-Cancer), Prevention, and Treatment**

Most otherwise healthy individuals who are infected with KSHV show no symptoms (DHHS 2013a, ACS 2014, NCI 2014). There are very few reports of primary infection with KSHV. Symptoms associated with initial KSHV infection include fever, a measles-like skin rash, diarrhea, fatigue, swollen lymph nodes, enlarged spleen, blood-cell deficiencies, and bone-marrow failure with an excess of B lymphocytes (Luppi *et al.* 2000a, Wang *et al.* 2001, Andreoni *et al.* 2002). Active KSHV infection may be associated with fever, skin rash, and hepatitis (Luppi *et al.* 2000b). There is conflicting evidence regarding suggested associations of KSHV infection with actinic keratosis or with the autoimmune skin diseases pemphigus vulgaris and pemphigus foliaceus (Ablashi *et al.* 2002). KSHV has been found in inflammatory cells in isolated cases of interstitial pneumonitis (an autoimmune-related lung disease), in sarcoid tissue (lesions formed in sarcoidosis, an inflammatory disease), and in histocytic necrotizing lymphadenitis (a lymph-node disorder), but a causal role for KSHV in these diseases has not been established.

Because KSHV transmission is associated with KSHV shedding in saliva and occasional shedding in genital secretions, avoiding salivary exposure (e.g., via kissing or sharing food, drink, or toothbrushes) and following safe sexual practices may theoretically prevent transmission (Chang-Moore Laboratory 2009, DHHS 2013a,b). Some drugs have been reported to reduce or inhibit KSHV shedding; however, no FDA-approved drugs currently exist for treatment of KSHV infection. Highly active antiretroviral therapy was associated with an 89% decrease in KSHV shedding frequency (Cattamanchi *et al.* 2011). There is no vaccine against KSHV, but limited vaccine development efforts are ongoing (Wu *et al.* 2012, ACS 2014).



## Regulations

### ***Department of Transportation (DOT)***

Infectious substances are considered hazardous materials, and special requirements have been set for marking, labeling, and transporting these materials.

### ***Occupational Safety and Health Administration (OSHA)***

Comprehensive regulations have been developed for employers to develop and adhere to exposure control plans for bloodborne pathogens.

All work-related needlestick injuries and cuts from sharp objects that are contaminated with another person's blood or other potentially infectious material must be recorded.

First-aid training program trainees must have adequate instruction in the value of universal precautions for preventing infectious diseases.

## Guidelines

### ***American Society of Transplantation (AST)***

The AST has issued guidance for the diagnosis, prevention, and treatment of KSHV infection after solid (vascular) organ transplantation. AST guidelines do not specifically prohibit solid organ transplantation because of KSHV seropositivity in either the donor or the recipient. They do advise that serologic screening be considered for donors and recipients from geographic regions with high rates of KSHV infection.

### ***Food and Drug Administration (FDA, an HHS agency)***

The FDA has issued numerous guidance documents prescribing procedures (e.g., use of standardized labels, abbreviated donor screening questionnaires) for reducing the risk of virus transmission by blood and blood products.

### ***Health Resources and Services Administration (HRSA, an HHS agency)***

KSHV infection is included as part of a list of potential donor-derived disease transmission events (PDDTE) reported through 2010 in the Organ Procurement and Transplantation Network's (OPTN) guidance for reporting PDDTE; however, these guidelines do not specifically prohibit solid organ transplantation because of KSHV seropositivity in the donor.

## References

Ablashi DV, Chatlynne LG, Whitman JE Jr, Cesarman E. 2002. Spectrum of Kaposi's sarcoma-associated herpesvirus, or human herpesvirus 8, diseases. *Clin Microbiol Rev* 15(3): 439-464.

ACS. 2014. *Kaposi Sarcoma*. American Cancer Society. Last updated: 8/19/14. <http://www.cancer.org/acs/groups/cid/documents/webcontent/003106-pdf.pdf>.

Albrecht D, Meyer T, Lorenzen T, Stoehr A, Arndt R, Plettenberg A. 2004. Epidemiology of HHV-8 infection in HIV-positive patients with and without Kaposi sarcoma: Diagnostic relevance of serology and PCR. *J Clin Virol* 30(2): 145-149.

- Amodio E, Valentini M, Gori-Savellini G, Valenti RM, Romano N, Goedert JJ, Cusi MG. 2011. Prevalence of Toscana and Sicilian phlebovirus antibodies in classic Kaposi sarcoma case patients and control subjects in Sicily. *J Infect Dis* 204(9): 1423-1426.
- An FQ, Folarin HM, Compitello N, Roth J, Gerson SL, McCrae KR, Fakhari FD, Dittmer DP, Renne R. 2006. Long-term-infected telomerase-immortalized endothelial cells: A model for Kaposi's sarcoma-associated herpesvirus latency *in vitro* and *in vivo*. *J Virol* 80(10): 4833-4846.
- Andreoni M, Sarmati L, Nicastrì E, El Sawaf G, El Zalabani M, Uccella I, Bugarini R, Parisi SG, Rezza G. 2002. Primary human herpesvirus 8 infection in immunocompetent children. *JAMA* 287(10): 1295-1300.
- Aoki Y, Yarchoan R, Wyvill K, Okamoto S, Little RF, Tosato G. 2001. Detection of viral interleukin-6 in Kaposi sarcoma-associated herpesvirus-linked disorders. *Blood* 97(7): 2173-2176.
- Ashlock BM, Ma Q, Issac B, Mesri EA. 2014. Productively infected murine Kaposi's sarcoma-like tumors define new animal models for studying and targeting KSHV oncogenesis and replication. *PLoS One* 9(1): e87324. 15 pp.
- Barozzi P, Luppi M, Facchetti F, Mecucci C, Alu M, Sarid R, *et al.* 2003. Post-transplant Kaposi sarcoma originates from the seeding of donor-derived progenitors. *Nat Med* 9(5): 554-561.
- Bhutani M, Polizzotto MN, Uldrick TS, Yarchoan R. 2015. Kaposi sarcoma-associated herpesvirus-associated malignancies: Epidemiology, pathogenesis, and advances in treatment. *Semin Oncol* 42(2): 223-246.
- Boulanger E, Afonso PV, Yahiaoui Y, Adle-Biassette H, Gabarre J, Agbalika F. 2008. Human herpesvirus-8 (HHV-8)-associated primary effusion lymphoma in two renal transplant recipients receiving rapamycin. *Am J Transplant* 8(3): 707-710.
- Burbelo PD, Issa AT, Ching KH, Wyvill KM, Little RF, Iadarola MJ, Kovacs JA, Yarchoan R. 2010. Distinct profiles of antibodies to Kaposi sarcoma-associated herpesvirus antigens in patients with Kaposi sarcoma, multicentric Castleman disease, and primary effusion lymphoma. *J Infect Dis* 201(12): 1919-1922.
- Campbell DM, Rappocciolo G, Jenkins FJ, Rinaldo CR. 2014. Dendritic cells: Key players in human herpesvirus 8 infection and pathogenesis. *Front Microbiol* 5: 452.
- Cattamanchi A, Saracino M, Selke S, Huang ML, Magaret A, Celum C, Corey L, Wald A, Casper C. 2011. Treatment with valacyclovir, famciclovir, or antiretrovirals reduces human herpesvirus-8 replication in HIV-1 seropositive men. *J Med Virol* 83(10): 1696-1703.
- Cavallin LE, Goldschmidt-Clermont P, Mesri EA. 2014. Molecular and cellular mechanisms of KSHV oncogenesis of Kaposi's sarcoma associated with HIV/AIDS. *PLoS Pathog* 10(7): e1004154. 4 pp.
- Chang Y, Cesarman E, Pessin MS, Lee F, Culpepper J, Knowles DM, Moore PS. 1994. Identification of herpesvirus-like DNA sequences in AIDS-associated Kaposi's sarcoma. *Science* 266(5192): 1865-1869.
- Chang-Moore Laboratory. 2009. *Kaposi's Sarcoma-Associated Herpesvirus (KSHV)*. Chang-Moore Cancer Virology Laboratory. <http://www.tumorvirology.pitt.edu/kshvrsh.html>.

- Coluzzi M, Calabro ML, Manno D, Chieco-Bianchi L, Schulz TF, Ascoli V. 2003. Reduced seroprevalence of Kaposi's sarcoma-associated herpesvirus (KSHV), human herpesvirus 8 (HHV8), related to suppression of Anopheles density in Italy. *Med Vet Entomol* 17(4): 461-464.
- DHHS. 2013a. Human herpesvirus-8 disease. In *Guidelines for the Prevention and Treatment of Opportunistic Infections Among HIV-Exposed and HIV-Infected Children*. Panel on Opportunistic Infections in HIV-Exposed and HIV-Infected Children, Department of Health and Human Services. Last updated: 11/6/13. [https://aidsinfo.nih.gov/contentfiles/lvguidelines/oi\\_guidelines\\_pediatics.pdf](https://aidsinfo.nih.gov/contentfiles/lvguidelines/oi_guidelines_pediatics.pdf).
- DHHS. 2013b. Human herpesvirus-8 disease. In *Guidelines for Prevention and Treatment of Opportunistic Infections in HIV-Infected Adults and Adolescents*. Panel on Opportunistic Infections in HIV-Infected Adults and Adolescents. Department of Health and Human Services. Last updated: 5/7/13. [https://aidsinfo.nih.gov/contentfiles/lvguidelines/Adult\\_OI.pdf](https://aidsinfo.nih.gov/contentfiles/lvguidelines/Adult_OI.pdf).
- Dittmer DP, Damania B, Sin SH. 2015. Animal models of tumorigenic herpesviruses — an update. *Curr Opin Virol* 14: 145-150.
- Dotti G, Fiocchi R, Motta T, Facchinetti B, Chiodini B, Borleri GM, Gavazzeni G, Barbui T, Rambaldi A. 1999. Primary effusion lymphoma after heart transplantation: A new entity associated with human herpesvirus-8. *Leukemia* 13(5): 664-670.
- Dupin N, Diss TL, Kellam P, Tulliez M, Du MQ, Sicard D, Weiss RA, Isaacson PG, Boshoff C. 2000. HHV-8 is associated with a plasmablastic variant of Castleman disease that is linked to HHV-8-positive plasmablastic lymphoma. *Blood* 95(4): 1406-1412.
- Dupuy A, Schulz T, Chevret S, Agbalika F, Pellet C, Janier M, *et al.* 2009. Asymmetrical transmission of human herpesvirus 8 among spouses of patients with Kaposi sarcoma. *Br J Dermatol* 160(3): 540-545.
- Engels EA, Atkinson JO, Graubard BI, McQuillan GM, Gamache C, Mbisa G, Cohn S, Whitby D, Goedert JJ. 2007. Risk factors for human herpesvirus 8 infection among adults in the United States and evidence for sexual transmission. *J Infect Dis* 196(2): 199-207.
- Fukumoto H, Kanno T, Hasegawa H, Katano H. 2011. Pathology of Kaposi's sarcoma-associated herpesvirus infection. *Front Microbiol* 2: 175.
- Giffin L, Damania B. 2014. KSHV: Pathways to tumorigenesis and persistent infection. *Adv Virus Res* 88: 111-159.
- IARC. 1997. Kaposi sarcoma herpesvirus/human herpesvirus 8. In *Epstein-Barr Virus and Kaposi's Sarcoma Herpesvirus/Human Herpesvirus 8*. IARC Monographs on the Evaluation of Carcinogenic Risks to Humans, vol. 70. Lyon, France: International Agency for Research on Cancer. pp. 375-492.
- IARC. 2008. *WHO Classification of Tumours of Haematopoietic and Lymphoid Tissues*, 4th ed., vol. 2. Swerdlow SH, Campo E, Harris NL, Jaffe ES, Pileri SA, Stein H, Thiele J, Vardiman JW, eds. Lyon, France: International Agency for Research on Cancer. 439 pp.
- IARC. 2012. Kaposi sarcoma herpesvirus. In *Biological Agents*. IARC Monographs on the Evaluation of Carcinogenic Risks to Humans, vol. 100B. Lyon, France: International Agency for Research on Cancer. pp. 169-214.

- Judde JG, Lacoste V, Briere J, Kassa-Kelembho E, Clyti E, Couppie P, *et al.* 2000. Monoclonality or oligoclonality of human herpesvirus 8 terminal repeat sequences in Kaposi's sarcoma and other diseases. *J Natl Cancer Inst* 92(9): 729-736.
- Katano H, Hoshino Y, Morishita Y, Nakamura T, Satoh H, Iwamoto A, Herndier B, Mori S. 1999. Establishing and characterizing a CD30-positive cell line harboring HHV-8 from a primary effusion lymphoma. *J Med Virol* 58(4): 394-401.
- Luppi M, Barozzi P, Schulz TF, Setti G, Staskus K, Trovato R, *et al.* 2000a. Bone marrow failure associated with human herpesvirus 8 infection after transplantation. *N Engl J Med* 343(19): 1378-1385.
- Luppi M, Barozzi P, Schulz TF, Trovato R, Donelli A, Narni F, Sheldon J, Marasca R, Torelli G. 2000b. Nonmalignant disease associated with human herpesvirus 8 reactivation in patients who have undergone autologous peripheral blood stem cell transplantation. *Blood* 96(7): 2355-2357.
- Martin JN, Ganem DE, Osmond DH, Page-Shafer KA, Macrae D, Kedes DH. 1998. Sexual transmission and the natural history of human herpesvirus 8 infection. *N Engl J Med* 338(14): 948-954.
- Mesri EA, Cesarman E, Boshoff C. 2010. Kaposi's sarcoma and its associated herpesvirus. *Nat Rev Cancer* 10(10): 707-719.
- Mesri EA, Cesarman E. 2011. Kaposi's sarcoma herpesvirus oncogenesis is a notch better in 3D. *Cell Host Microbe* 10(6): 529-531.
- Mesri EA, Feitelson MA, Munger K. 2014. Human viral oncogenesis: A cancer hallmarks analysis. *Cell Host Microbe* 15(3): 266-282.
- Mutlu AD, Cavallin LE, Vincent L, Chiozzini C, Eroles P, Duran EM, *et al.* 2007. *In vivo* growth-restricted and reversible malignancy induced by human herpesvirus-8 KSHV: A cell and animal model of virally induced Kaposi's sarcoma. *Cancer Cell* 11(3): 245-258.
- NCI. 2014. *A Snapshot of Kaposi Sarcoma*. National Cancer Institute. <http://www.cancer.gov/researchandfunding/snapshots/kaposisarcoma>.
- Newton R, Ziegler J, Bourbouli D, Casabonne D, Beral V, Mbidde E, *et al.* 2003a. Infection with Kaposi's sarcoma-associated herpesvirus (KSHV) and human immunodeficiency virus (HIV) in relation to the risk and clinical presentation of Kaposi's sarcoma in Uganda. *Br J Cancer* 89(3): 502-504.
- Newton R, Ziegler J, Bourbouli D, Casabonne D, Beral V, Mbidde E, *et al.* 2003b. The sero-epidemiology of Kaposi's sarcoma-associated herpesvirus (KSHV/HHV-8) in adults with cancer in Uganda. *Int J Cancer* 103(2): 226-232.
- Newton R, Carpenter L, Casabonne D, Beral V, Babiker A, Darbyshire J, *et al.* 2006. A prospective study of Kaposi's sarcoma-associated herpesvirus and Epstein-Barr virus in adults with human immunodeficiency virus-1. *Br J Cancer* 94(10): 1504-1509.
- O'Brien TR, Kedes D, Ganem D, Macrae DR, Rosenberg PS, Molden J, Goedert JJ. 1999. Evidence for concurrent epidemics of human herpesvirus 8 and human immunodeficiency virus type 1 in US homosexual men: rates, risk factors, and relationship to Kaposi's sarcoma. *J Infect Dis* 180(4): 1010-1017.

- Oksenhendler E, Boulanger E, Galicier L, Du MQ, Dupin N, Diss TC, *et al.* 2002. High incidence of Kaposi sarcoma-associated herpesvirus-related non-Hodgkin lymphoma in patients with HIV infection and multicentric Castleman disease. *Blood* 99(7): 2331-2336.
- Oksenhendler E, Boutboul D, Beldjord K, Meignin V, de Labarthe A, Fieschi C, *et al.* 2013. Human herpesvirus 8+ polyclonal IgM $\lambda$  B-cell lymphocytosis mimicking plasmablastic leukemia/lymphoma in HIV-infected patients. *Eur J Haematol* 91(6): 497-503.
- Parravicini C, Corbellino M, Paulli M, Magrini U, Lazzarino M, Moore PS, Chang Y. 1997a. Expression of a virus-derived cytokine, KSHV vIL-6, in HIV-seronegative Castleman's disease. *Am J Pathol* 151(6): 1517-1522.
- Parravicini C, Olsen SJ, Capra M, Poli F, Sirchia G, Gao SJ, *et al.* 1997b. Risk of Kaposi's sarcoma-associated herpes virus transmission from donor allografts among Italian posttransplant Kaposi's sarcoma patients. *Blood* 90(7): 2826-2829.
- Parravicini C, Chandran B, Corbellino M, Berti E, Paulli M, Moore PS, Chang Y. 2000. Differential viral protein expression in Kaposi's sarcoma-associated herpesvirus-infected diseases: Kaposi's sarcoma, primary effusion lymphoma, and multicentric Castleman's disease. *Am J Pathol* 156(3): 743-749.
- Pellett PE, Wright DJ, Engels EA, Ablashi DV, Dollard SC, Forghani B, *et al.* 2003. Multicenter comparison of serologic assays and estimation of human herpesvirus 8 seroprevalence among US blood donors. *Transfusion* 43(9): 1260-1268.
- Phillips AM, Jones AG, Osmond DH, Pollack LM, Catania JA, Martin JN. 2008. Awareness of Kaposi's sarcoma-associated herpesvirus among men who have sex with men. *Sex Transm Dis* 35(12): 1011-1014.
- Reddy D, Mitsuyasu R. 2011. HIV-associated multicentric Castleman disease. *Curr Opin Oncol* 23(5): 475-481.
- Simonelli C, Spina M, Cinelli R, Talamini R, Tedeschi R, Gloghini A, Vaccher E, Carbone A, Tirelli U. 2003. Clinical features and outcome of primary effusion lymphoma in HIV-infected patients: A single-institution study. *J Clin Oncol* 21(21): 3948-3954.
- Sitas F, Carrara H, Beral V, Newton R, Reeves G, Bull D, *et al.* 1999. Antibodies against human herpesvirus 8 in black South African patients with cancer. *N Engl J Med* 340(24): 1863-1871.
- Smith NA, Sabin CA, Gopal R, Bourboullia D, Labbet W, Boshoff C, *et al.* 1999. Serologic evidence of human herpesvirus 8 transmission by homosexual but not heterosexual sex. *J Infect Dis* 180(3): 600-606.
- Sullivan RJ, Pantanowitz L, Casper C, Stebbing J, Dezube BJ. 2008. HIV/AIDS: epidemiology, pathophysiology, and treatment of Kaposi sarcoma-associated herpesvirus disease: Kaposi sarcoma, primary effusion lymphoma, and multicentric Castleman disease. *Clin Infect Dis* 47(9): 1209-1215.
- Testa A, Baiocchi A, Comandini UV, Falasca L, Nardacci R, Maritti M, *et al.* 2010. Fatal sclerosing peritonitis associated with primary effusion lymphoma after liver transplantation: A case report. *Transplant Proc* 42(9): 3849-3853.

---

Wang QJ, Jenkins FJ, Jacobson LP, Kingsley LA, Day RD, Zhang ZW, *et al.* 2001. Primary human herpesvirus 8 infection generates a broadly specific CD8(+) T-cell response to viral lytic cycle proteins. *Blood* 97(8): 2366-2373.

Wen KW, Damania B. 2010. Kaposi sarcoma-associated herpesvirus (KSHV): Molecular biology and oncogenesis. *Cancer Lett* 289(2): 140-150.

Wu TT, Qian J, Ang J, Sun R. 2012. Vaccine prospect of Kaposi sarcoma-associated herpesvirus. *Curr Opin Virol* 2(4): 482-488.

Xu GJ, Kula T, Xu Q, Li MZ, Vernon SD, Ndung'u T, *et al.* 2015. Viral immunology. Comprehensive serological profiling of human populations using a synthetic human virome. *Science* 348(6239): aaa0698. 23 pp.

**This Page Intentionally Left Blank**



National Toxicology Program  
National Institute of Environmental Health Sciences  
National Institutes of Health  
P.O. Box 12233, MD K2-14  
Research Triangle Park, NC 27709

ISSN# 2331-267X