

Developing Reverse Toxicokinetic Models to Correlate *In Vitro* and *In Vivo* Activity

John Wambaugh
Office of Research and Development
National Center for Computational Toxicology
wambaugh.john@epa.gov

The views expressed in this
presentation are those of the
author and do not necessarily
reflect the views or policies of the
U.S. EPA

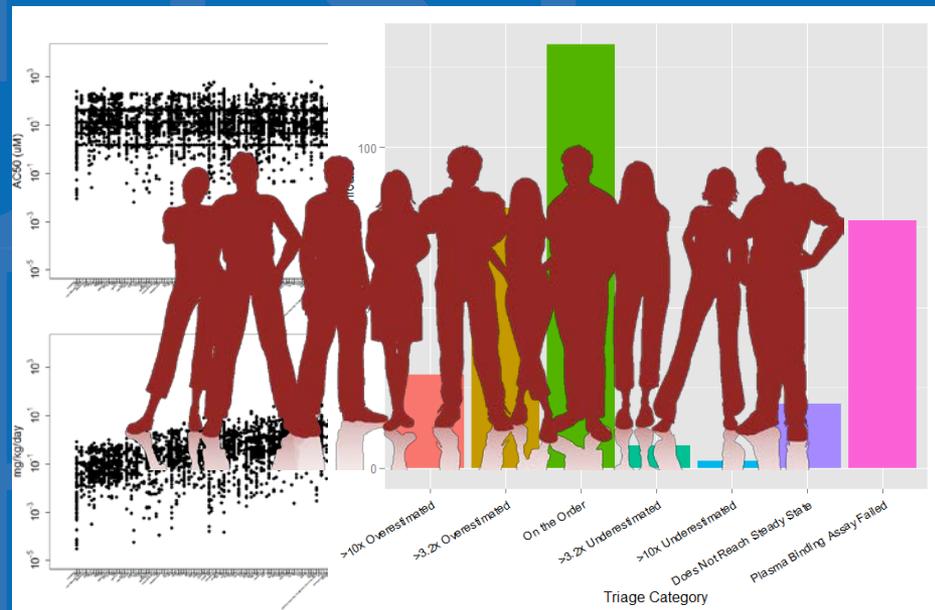


Figure includes image from Thinkstock

Introduction

- High throughput screening (HTS) methods exist for identifying chemical concentrations that may induce changes to human biology (*e.g.*, Tox21, ToxCast)
- High throughput exposure (HTE) methods exist for predicting potential human exposure for arbitrary chemicals (*e.g.*, ExpoCast – Wambaugh *et al.*, 2014)
- Toxicokinetics (TK) provides a bridge between HTS and HTE by predicting tissue concentrations due to exposure

Introduction

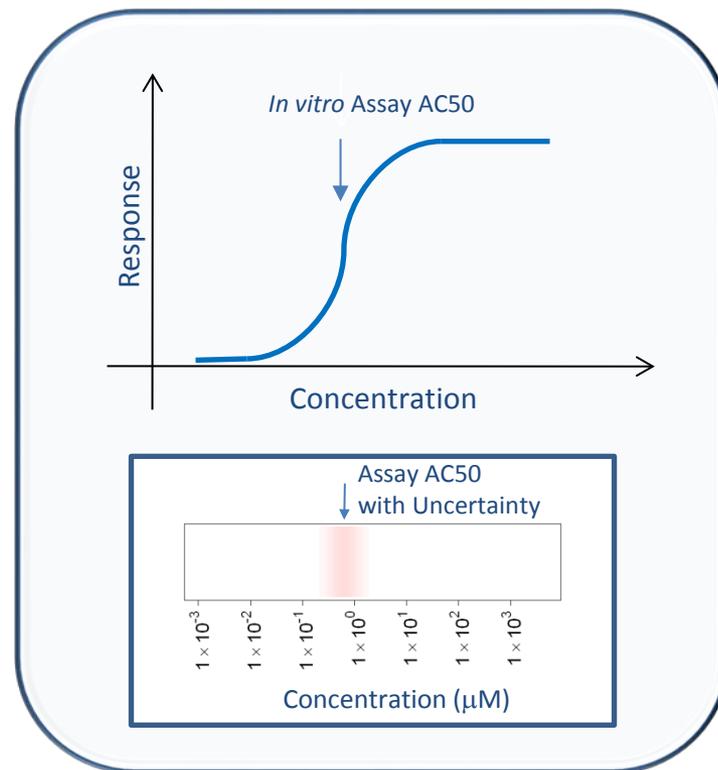
- Toxicokinetics (TK) provides a bridge between HTS and HTE by predicting tissue concentrations due to exposure
 - Traditional TK methods are resource intensive

- Relatively high throughput TK (HTTK) methods have been used by the pharmaceutical industry to determine range of efficacious doses and to prospectively evaluate success of planned clinical trials (Jamei, *et al.*, 2009; Wang, 2010)
 - A key application of HTTK has been “reverse dosimetry” (also called Reverse TK or RTK)

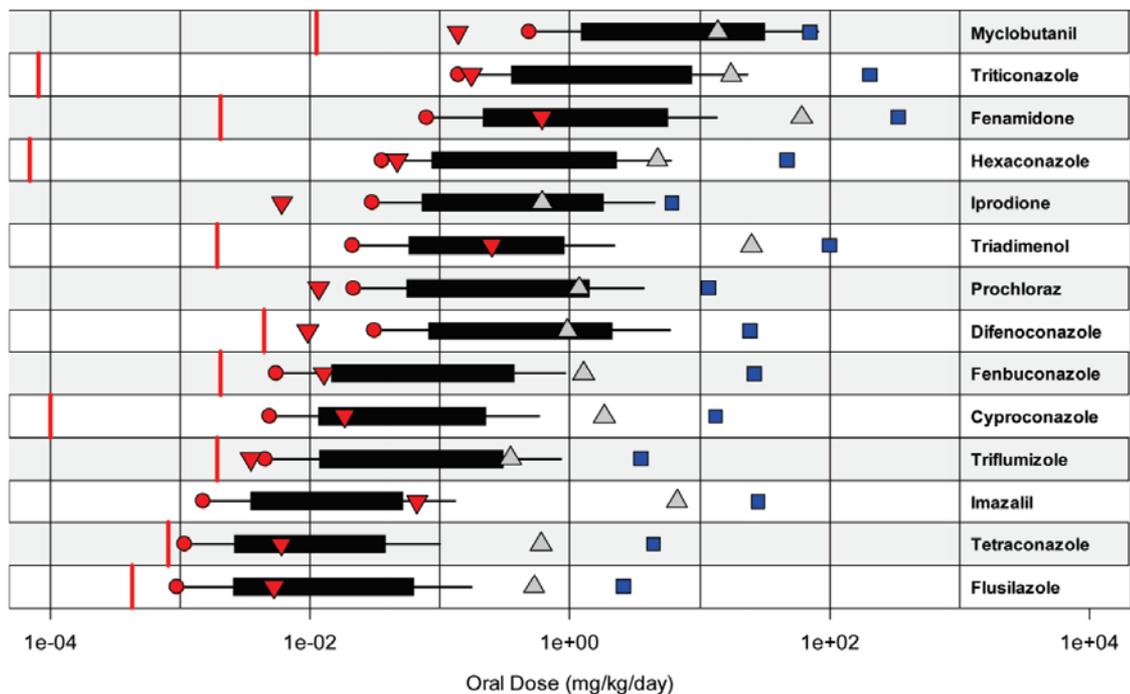
 - RTK can approximately convert *in vitro* HTS results to daily doses needed to produce similar levels in a human for comparison to exposure data (Wetmore, *et al.*, 2012)

High-Throughput Bioactivity

- **Tox21:** Examining >10,000 chemicals using ~50 assays intended to identify interactions with biological pathways (Schmidt, 2009)
- **ToxCast:** For a subset (>1000) of Tox21 chemicals ran >500 additional assays (Judson et al., 2010)
- Most assays conducted in dose-response format (identify 50% activity concentration – AC50 – and efficacy if data described by a Hill function)
- All data is public: <http://actor.epa.gov/>



In vitro Bioactivity, RTK, and *in Vivo* Toxic Doses



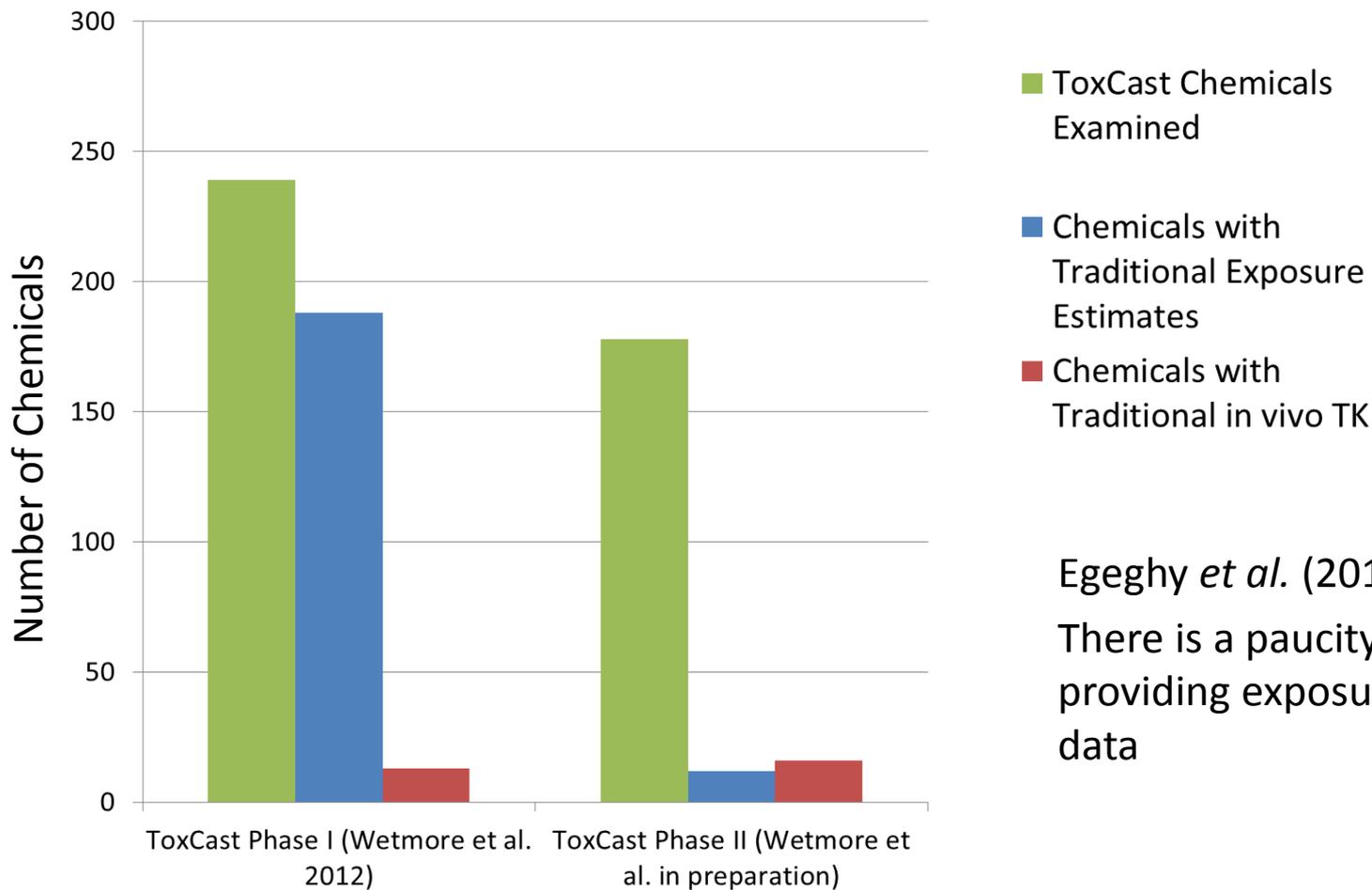
Judson *et al.* (2011)

Comparison of HTTK predicted oral equivalent doses (box and whisker plots in mg/kg/day) with doses for no effect and low effect groups in animal studies

- Lowest Observed Effect Level
- △ No Observed Effect Level (NEL)
- ▼ NEL/100

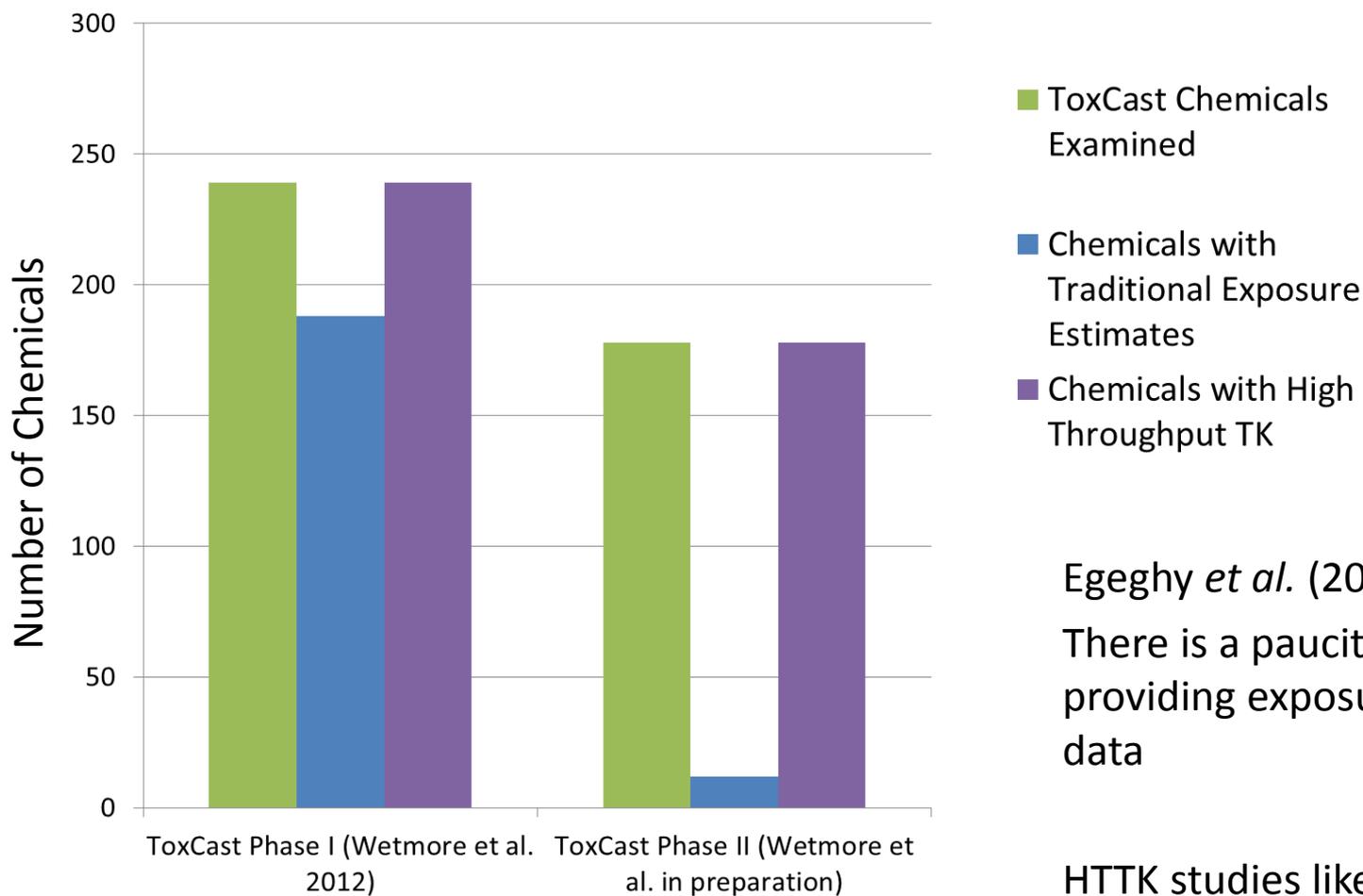
Estimated chronic exposure levels from food residues are indicated by vertical red lines. All values are in mg/kg/day.

In Vitro Bioactivity, In Vivo Toxicokinetics, and Human Exposure



Egeghy *et al.* (2012):
There is a paucity of data for
providing exposure context to HTS
data

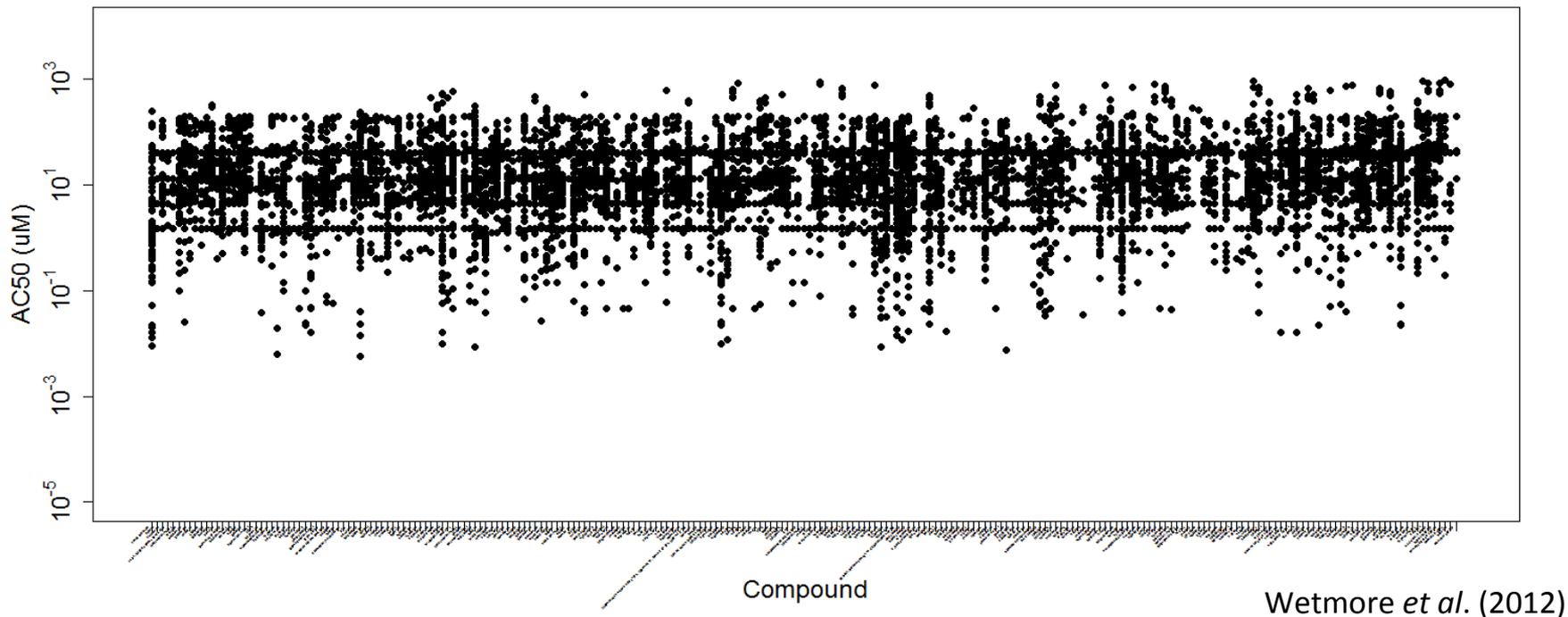
In Vitro Bioactivity, In Vitro Toxicokinetics, and Human Exposure



Egeghy *et al.* (2012):
 There is a paucity of data for providing exposure context to HTS data

HTTK studies like Wetmore *et al.* (2012), can address the need for toxicokinetic data

ToxCast *in vitro* Bioactive Concentrations



- One point for each chemical-*in vitro* assay combination with a systematic (Hill function) concentration response curve

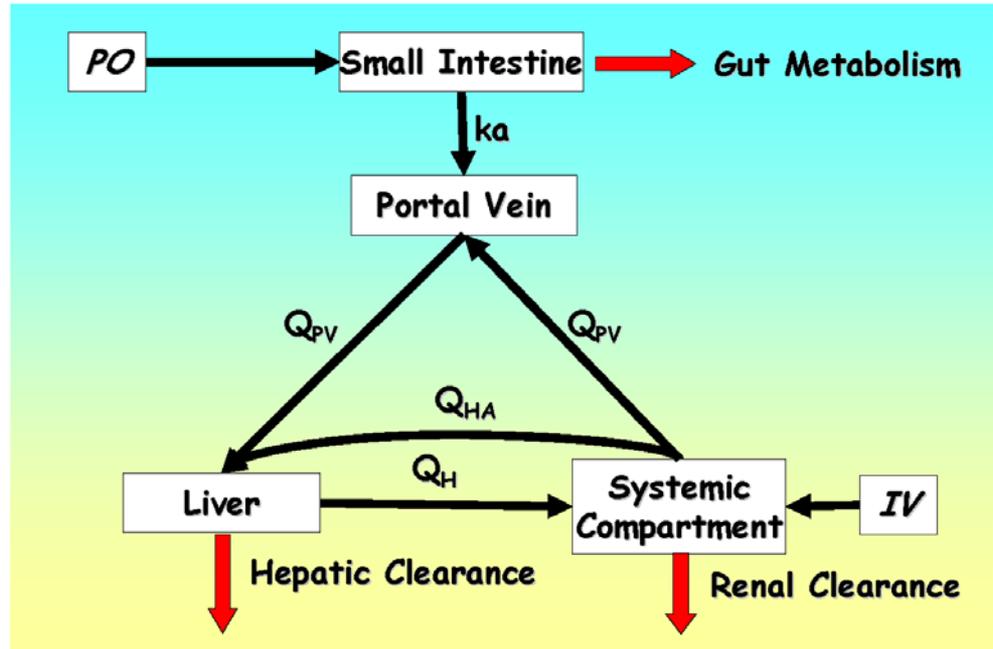
Predicting Steady-State Plasma Concentration

Lam et al (2009)

simCYP
© 2001-2009 Simcp Limited

Minimal Model: Lumped Single Distribution Volume

- *In vitro* plasma protein binding and metabolic clearance assays allow approximate hepatic and renal clearances to be calculated
- At steady state this allows conversion from concentration to administered dose
- No oral absorption/bioavailability included



$$C_{ss} = \frac{\text{oral dose rate}}{(GFR * F_{ub}) + \left(Q_l * F_{ub} * \frac{Cl_{int}}{Q_l + F_{ub} * Cl_{int}} \right)}$$

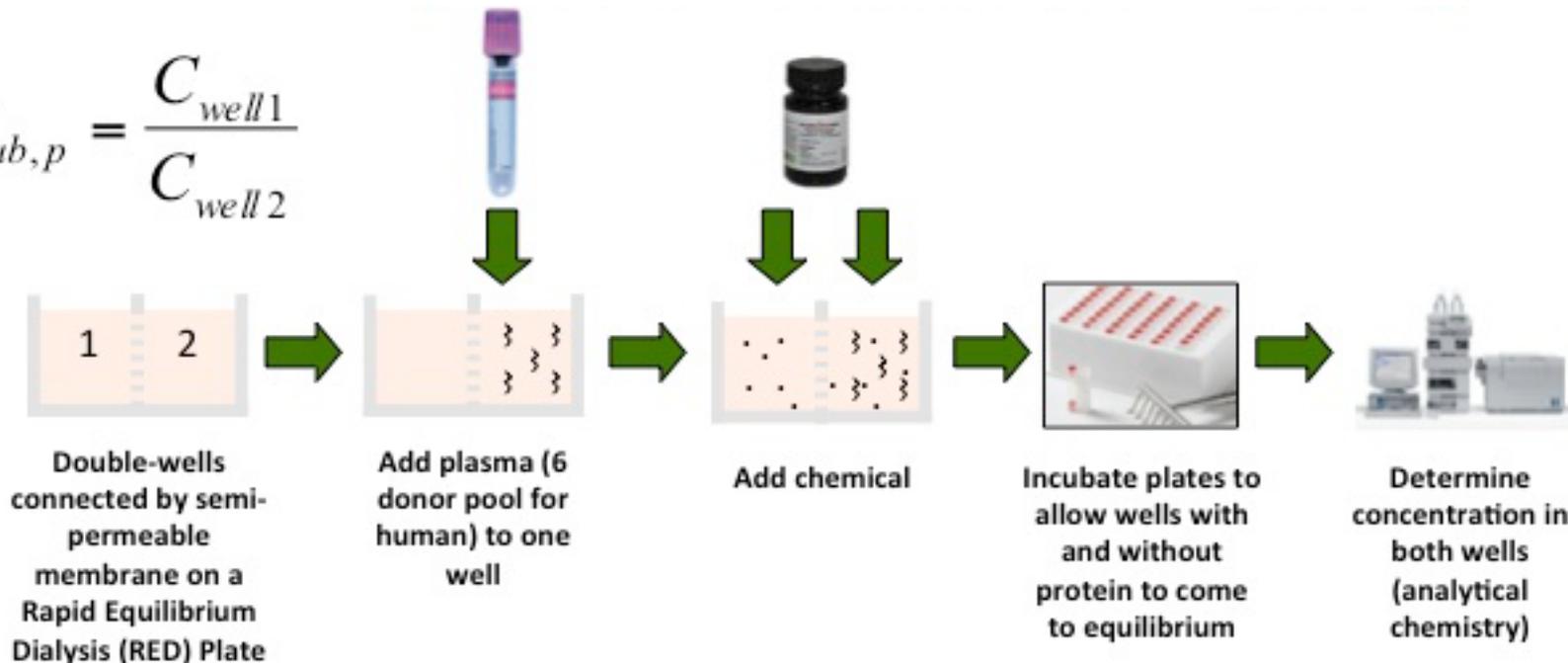
Oral dose in
(mg/kg/day)



Sum of hepatic
and renal
clearance
(mg/kg/day)

Plasma Protein Binding (Fraction Unbound in Plasma)

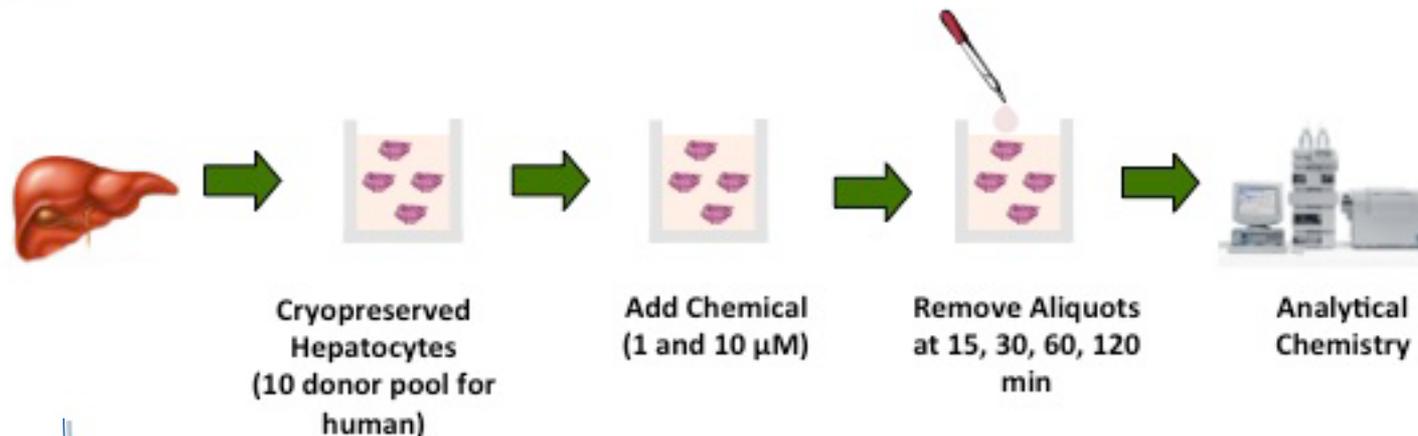
$$F_{ub,p} = \frac{C_{well1}}{C_{well2}}$$



- Data on ToxCast chemicals initially collected at Hamner Institutes
- Published:
 - Rotroff et al. (2010) - Pilot study using 38 Phase I ToxCast Chemicals
 - Wetmore et al. (2012) - Remainder of easily analyzed Phase I chemicals
 - Wetmore et al. (2013) - Rat TK for 50 ToxCast/ToxRefDB compounds
 - Wetmore et al. (2014) – Assessed variability in metabolism for a dozen ToxCast compounds

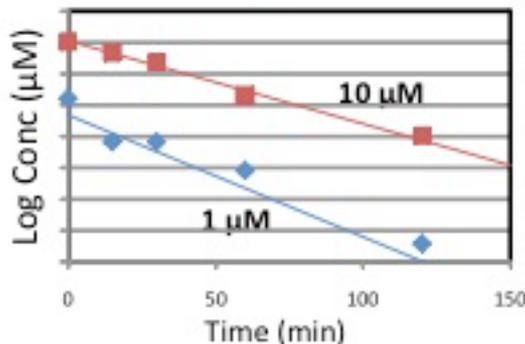
RED Method:
Waters *et al.* (2008)

Intrinsic Hepatic Clearance



The rate of disappearance of parent compound (slope of line) is the **hepatic clearance** ($\mu\text{L}/\text{min}/10^6$ hepatocytes)

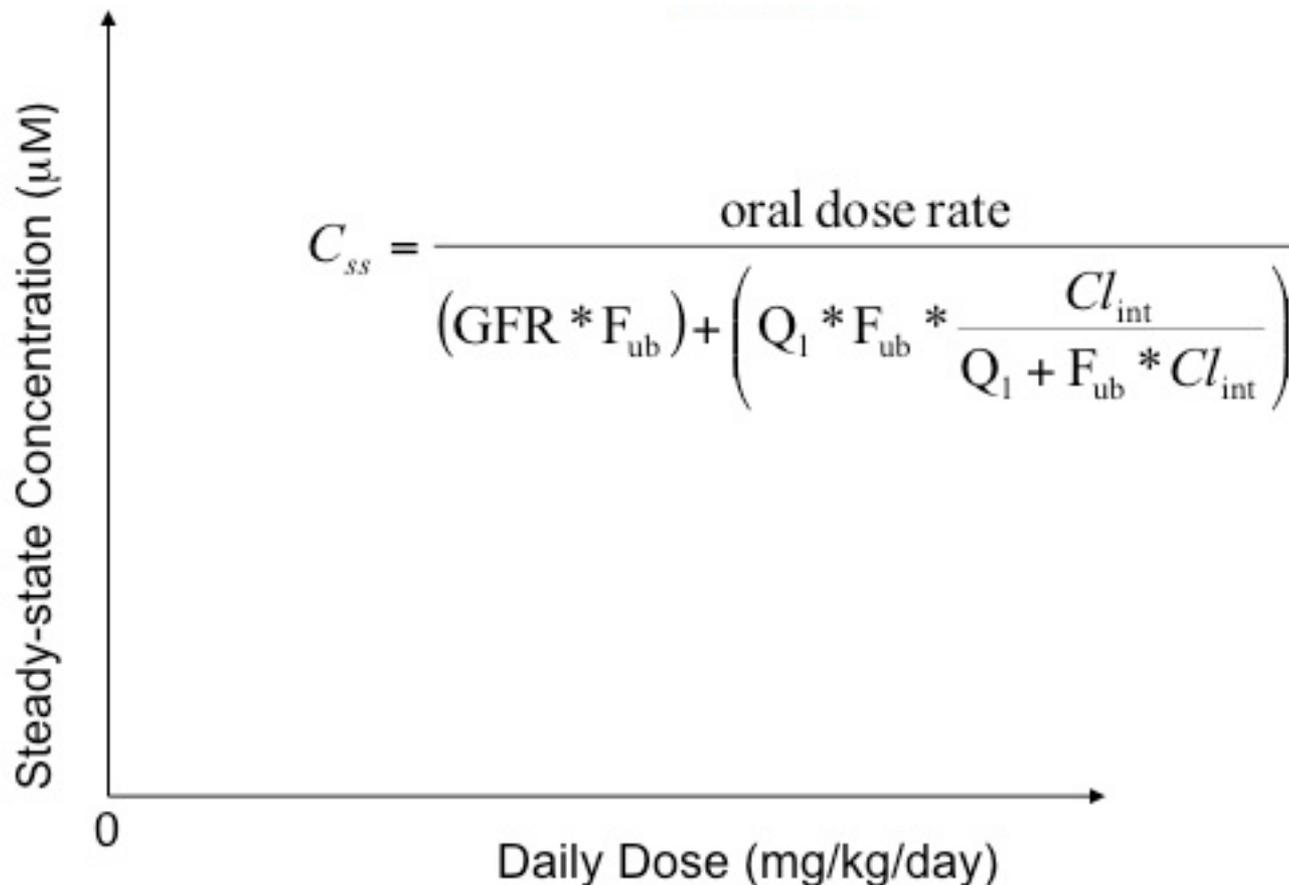
We perform the assay at
Cryopreserved hepatocyte
Method: Shibata *et al.* (2002)



1 and 10 μM to check for saturation of metabolizing enzymes.

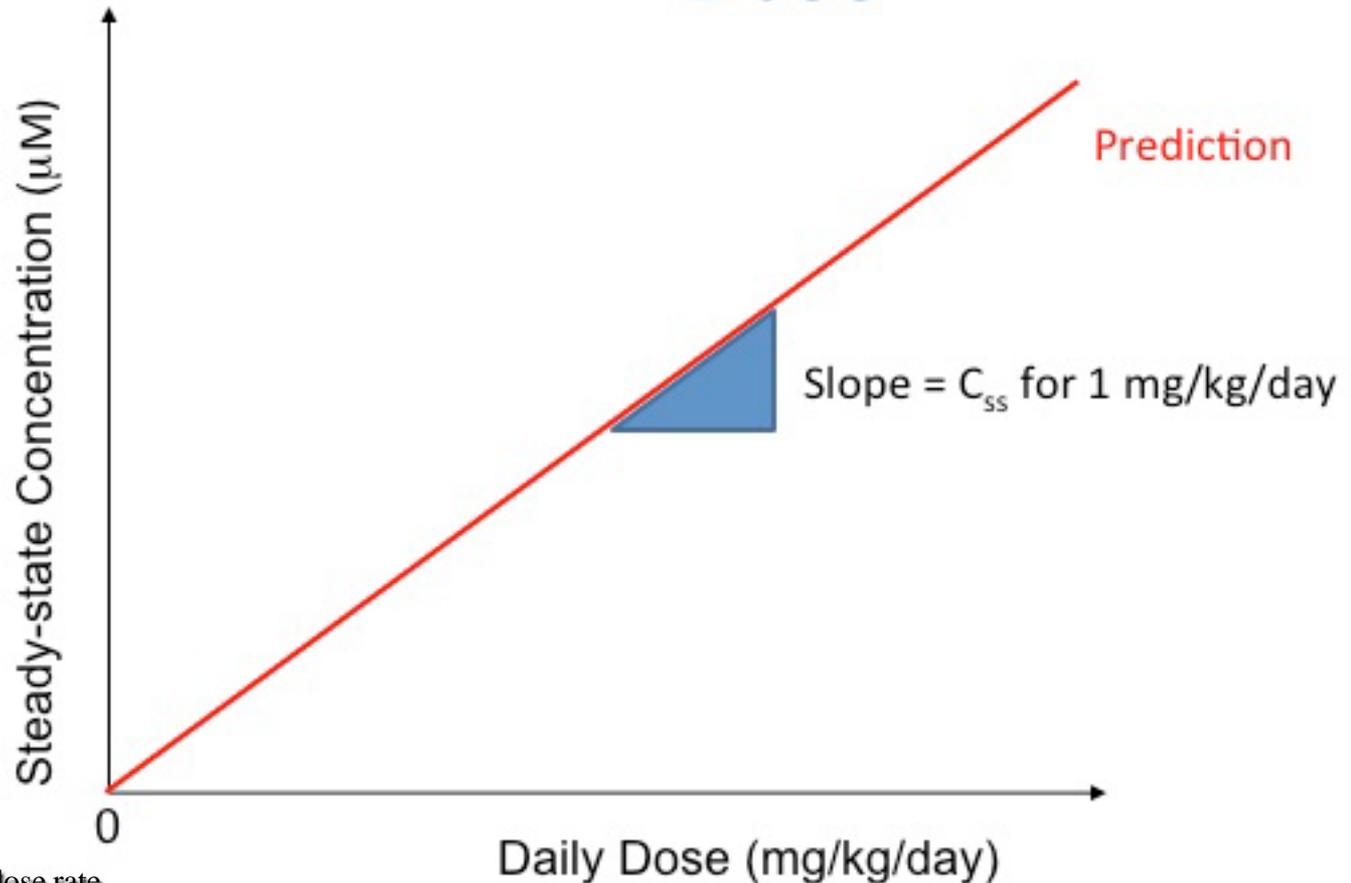
- Data on ToxCast chemicals initially collected at Hamner Institutes

Steady-State is Linear with Dose



- Can calculate predicted steady-state concentration (C_{ss}) for a 1 mg/kg/day dose and multiply to get concentrations for other doses

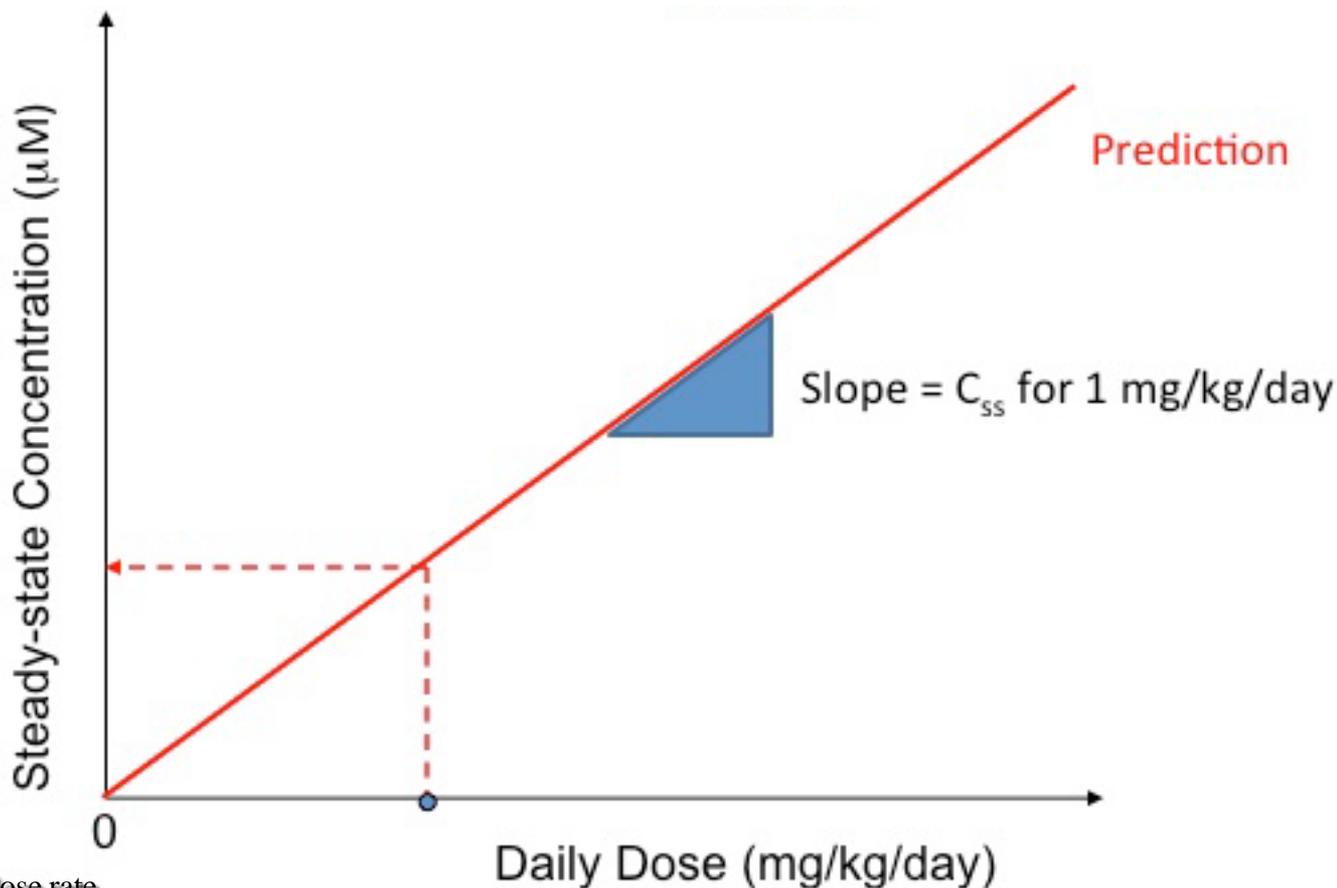
Steady-State is Linear with Dose



$$C_{ss} = \frac{\text{oral dose rate}}{\left(\text{GFR} * F_{ub} \right) + \left(Q_l * F_{ub} * \frac{Cl_{int}}{Q_l + F_{ub} * Cl_{int}} \right)}$$

- Can calculate predicted steady-state concentration (C_{ss}) for a 1 mg/kg/day dose and multiply to get concentrations for other doses

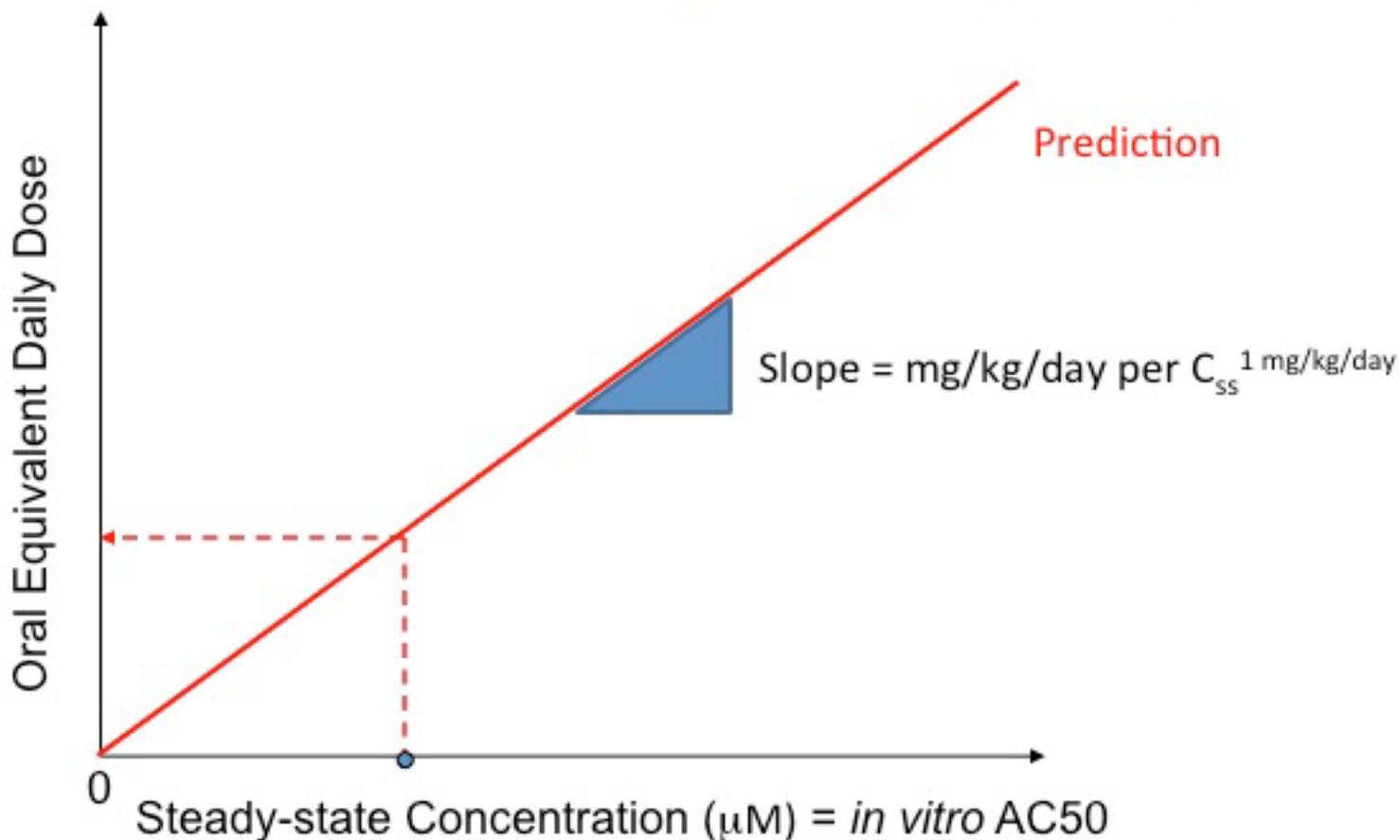
Steady-State is Linear with Dose



$$C_{ss} = \frac{\text{oral dose rate}}{\left(\text{GFR} * F_{ub} \right) + \left(Q_l * F_{ub} * \frac{Cl_{int}}{Q_l + F_{ub} * Cl_{int}} \right)}$$

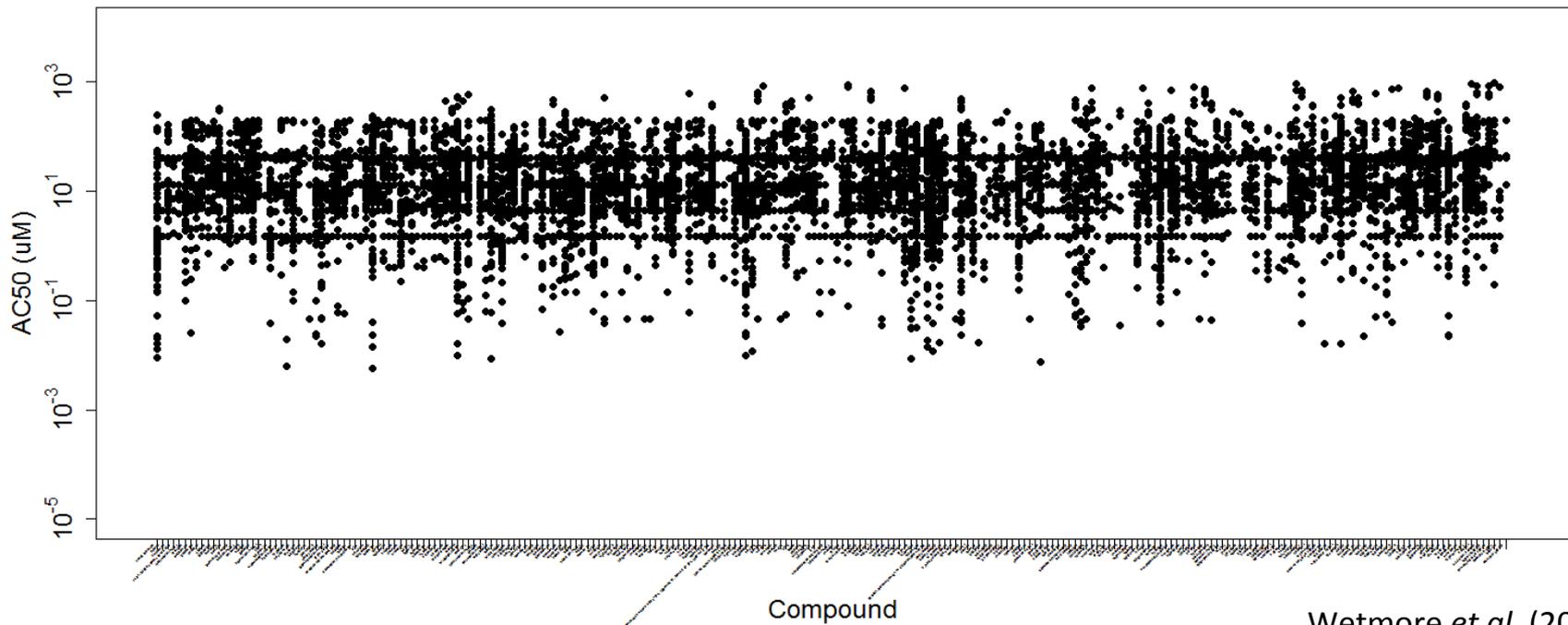
- Can calculate predicted steady-state concentration (C_{ss}) for a 1 mg/kg/day dose and multiply to get concentrations for other doses

HTTK Allows Steady-State *In Vitro*- *In Vivo* Extrapolation (IVIVE)



- Swap the axes (this is the “reverse” part of reverse dosimetry)
- Can divide bioactive concentration by C_{ss} for for a 1 mg/kg/day dose to get oral equivalent dose

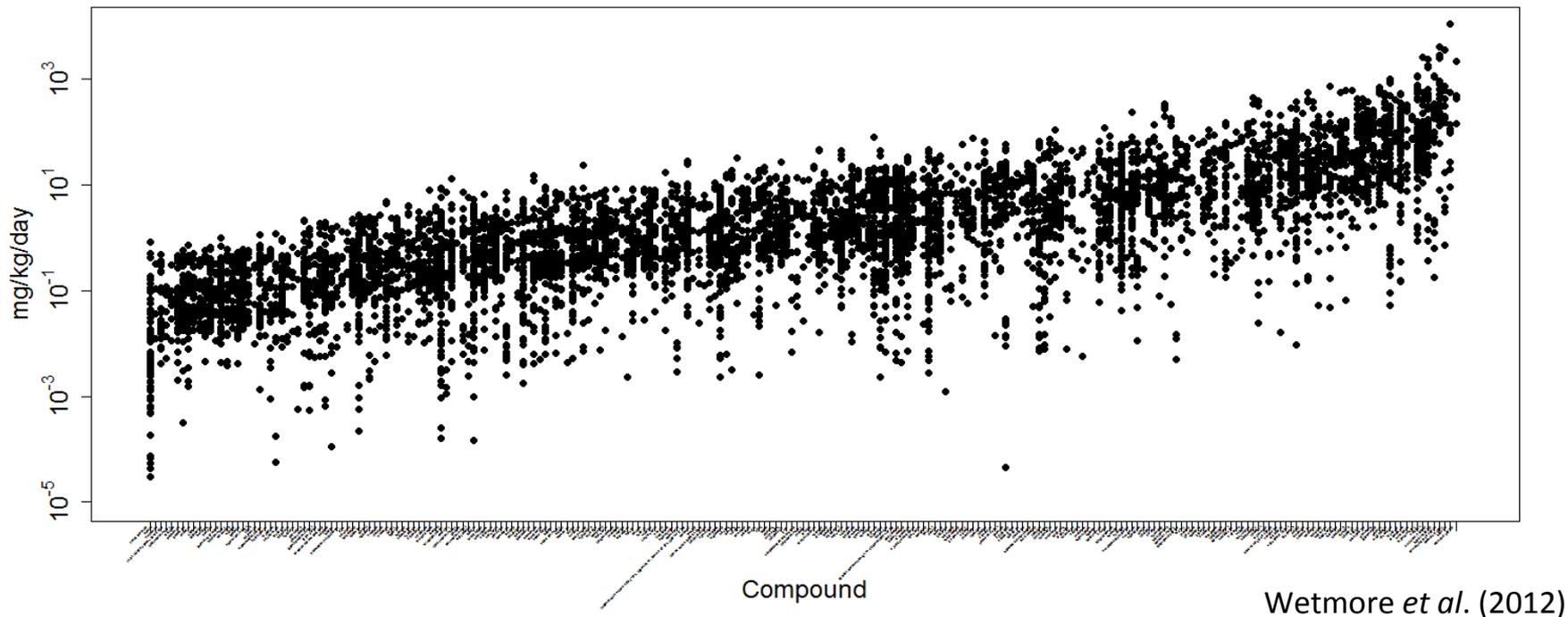
ToxCast *in vitro* Bioactive Concentrations



Wetmore *et al.* (2012)

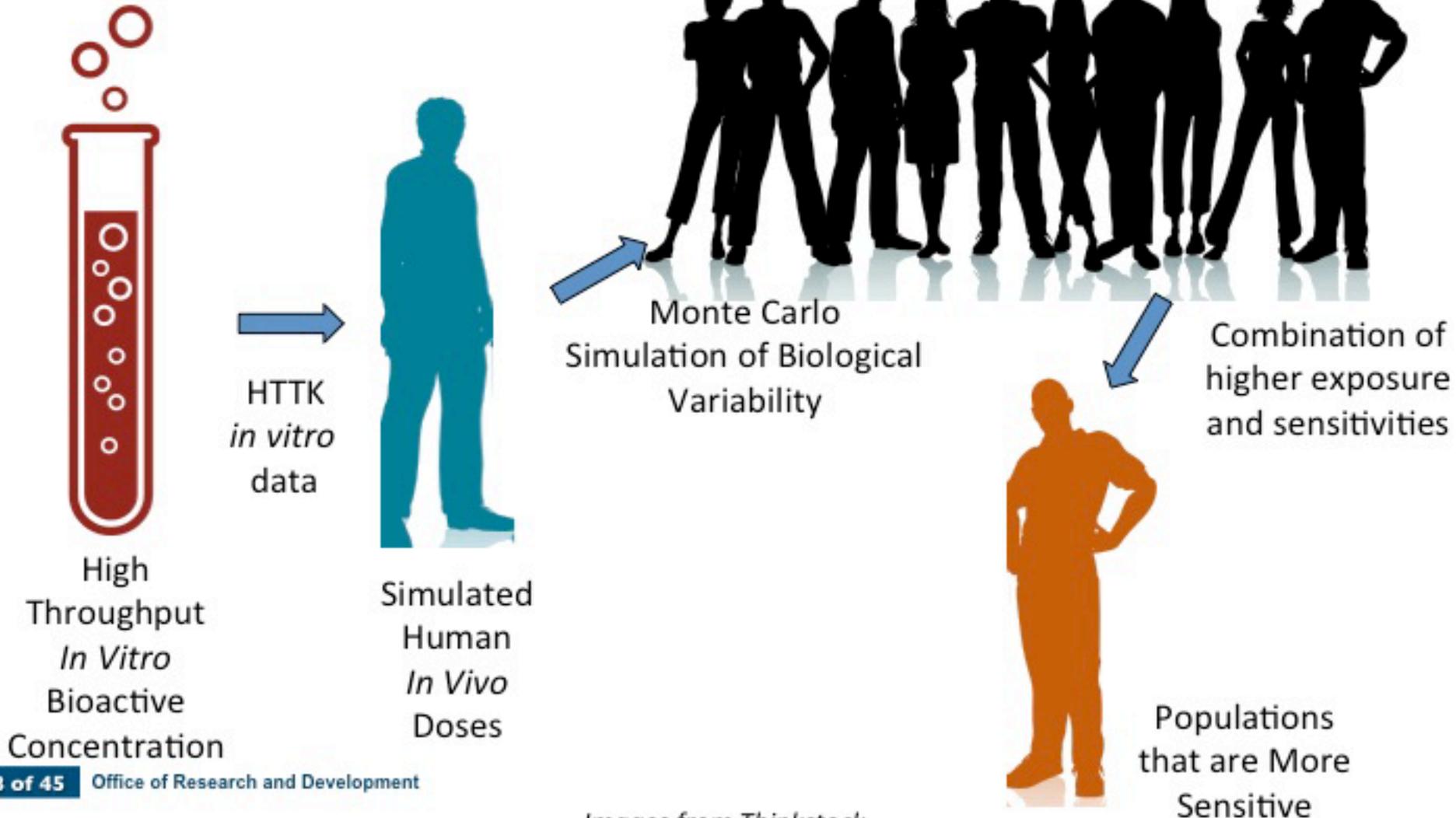
- It appears harder to prioritize on bioactive *in vitro* concentration without *in vivo* context

RTK Oral Equivalents



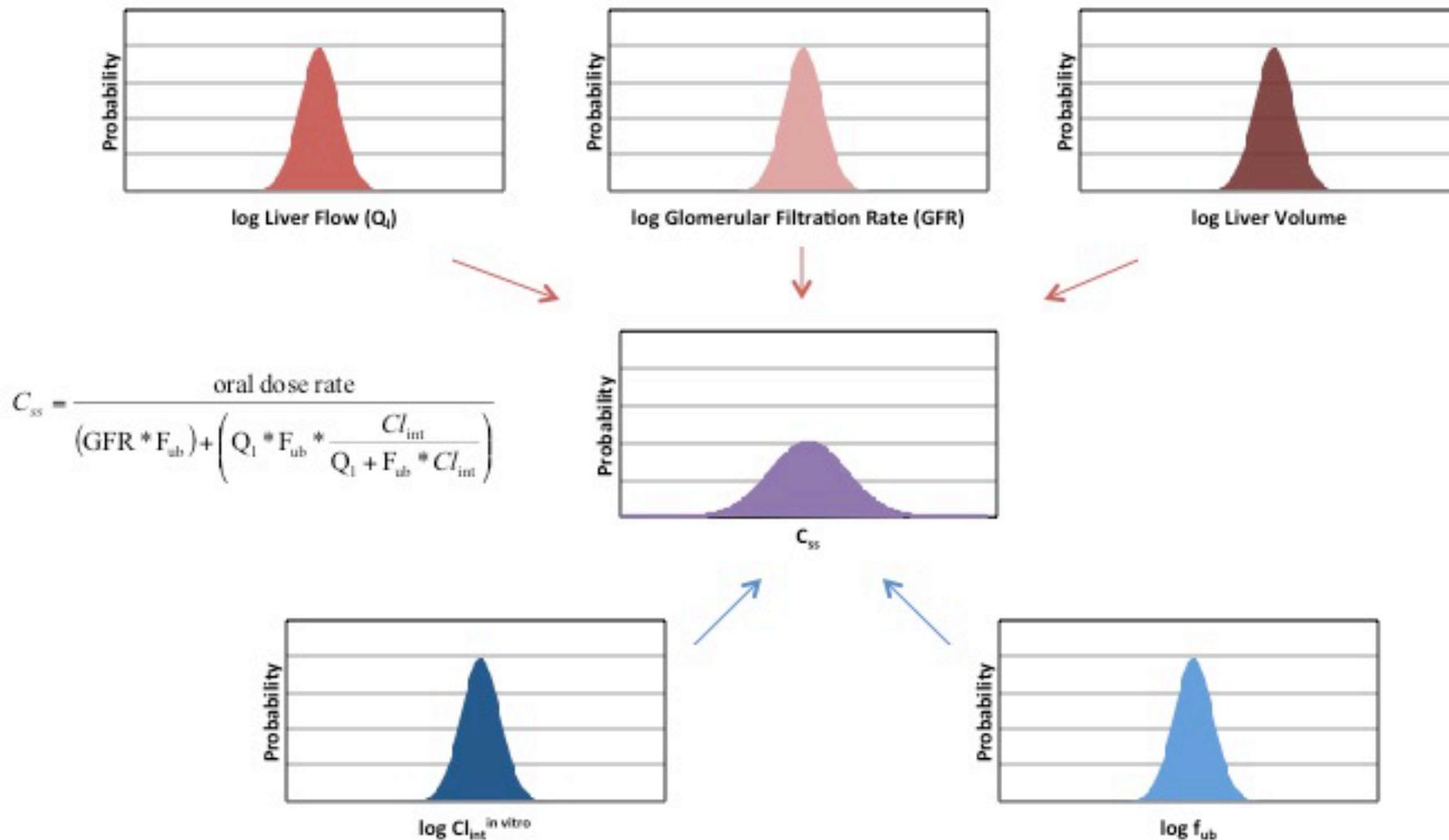
- Translation from *in vitro* to steady-state oral equivalent doses allow greater discrimination between effective chemical potencies

Reverse Dosimetry with HTTK

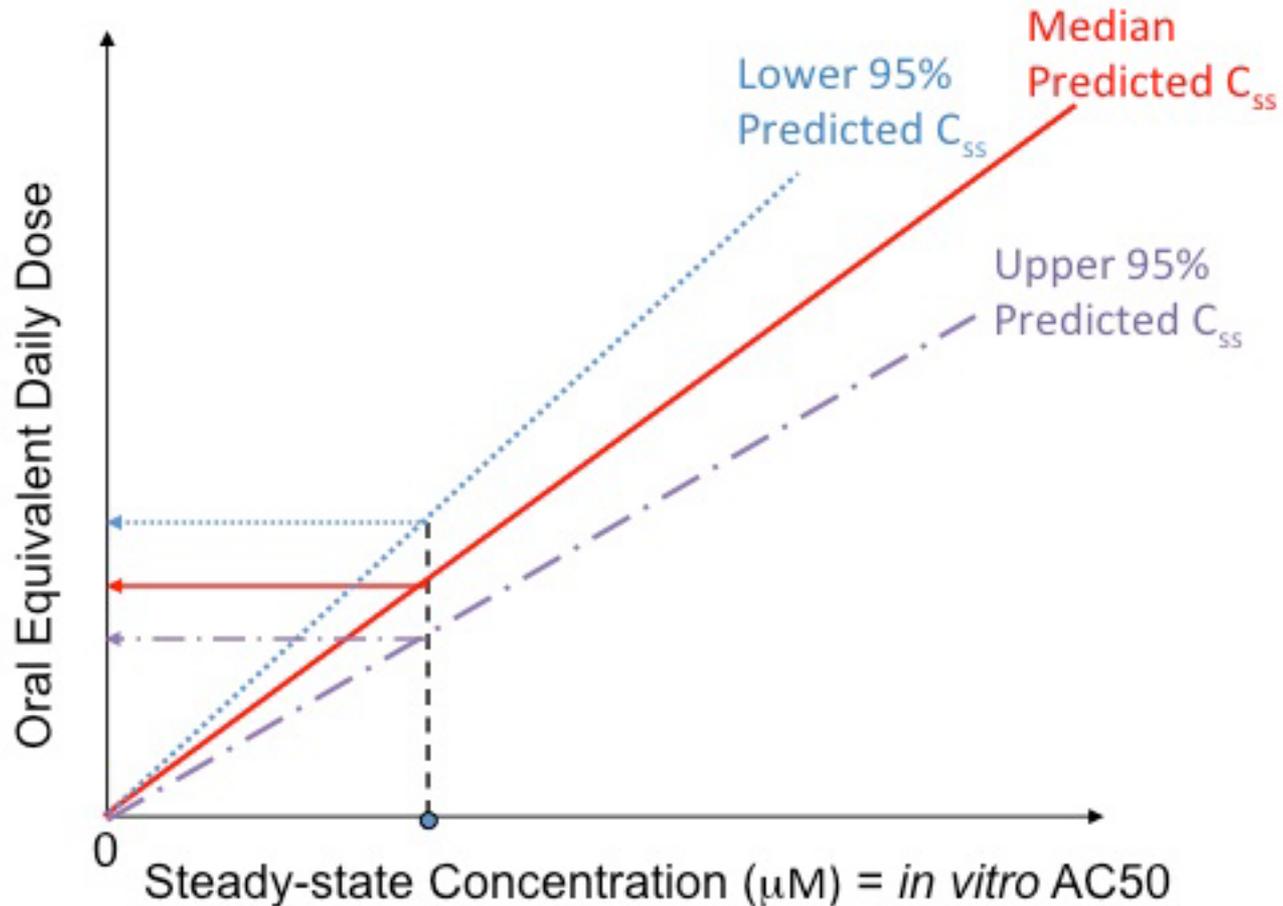


Images from Thinkstock

Monte Carlo (MC) Approach to Variability: SimCYP (Pharma) Approach



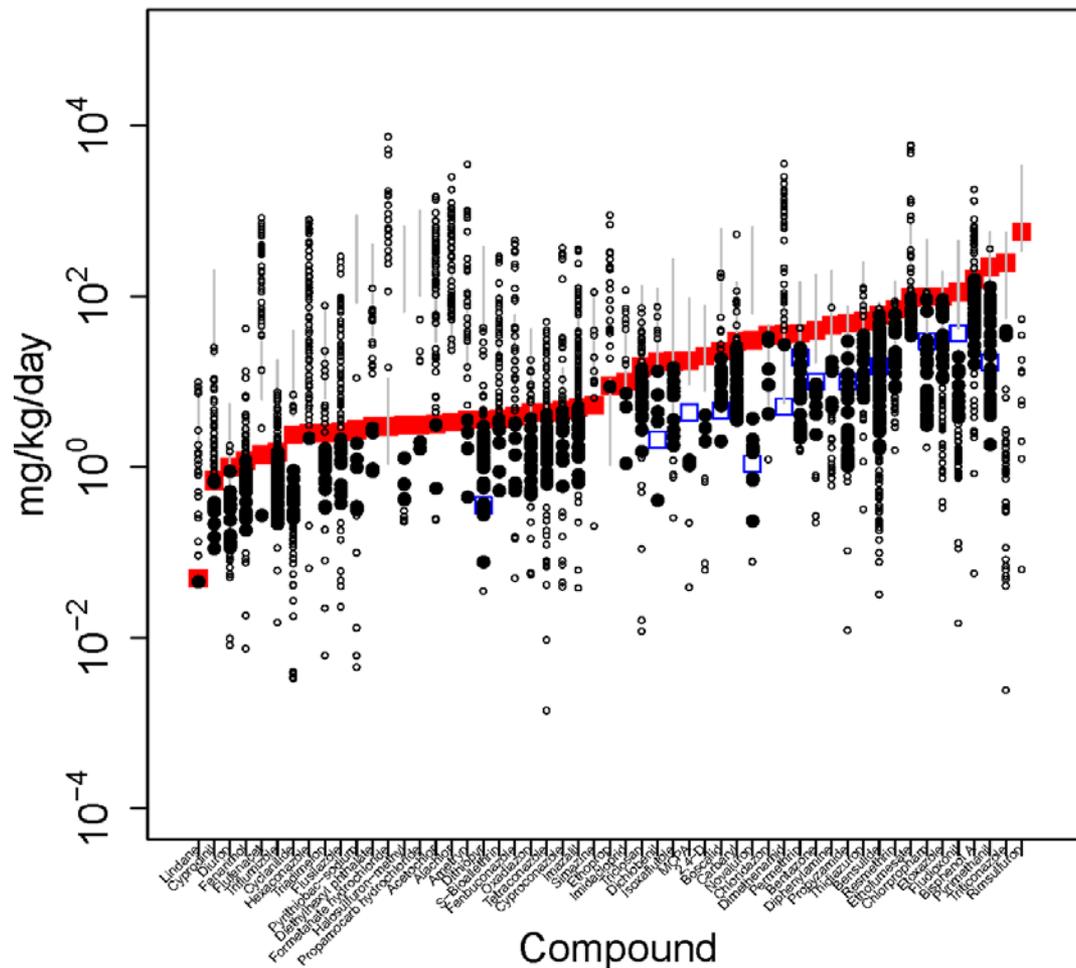
Steady-State In Vitro-In Vivo Extrapolation (IVIVE)



- The higher the predicted C_{ss} , the lower the oral equivalent dose, so the upper 95% predicted C_{ss} from the MC has a lower oral equivalent dose

Human HTS With Rat HTTK

- Concordance of steady state oral equivalent doses and *in vivo* No Observed (□) and Lowest Observed (■) dose levels provides an additional method for evaluation
- Wetmore *et al.* (2013) chose 53 chemicals with good coverage of *in vivo* endpoints in the Toxicity Reference Database (ToxRefDB – Martin *et al.*, 2009)

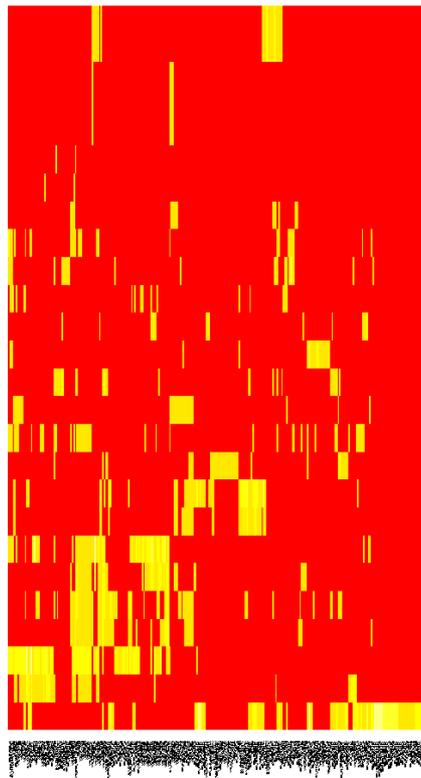
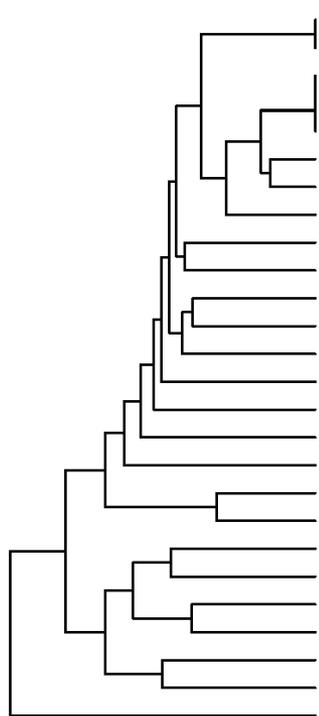
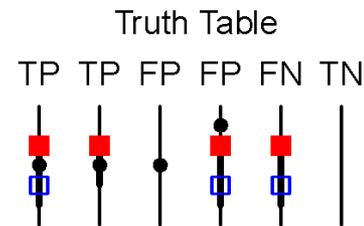
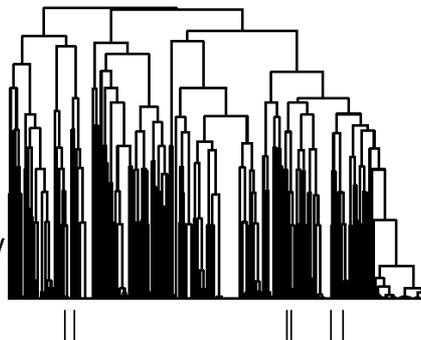
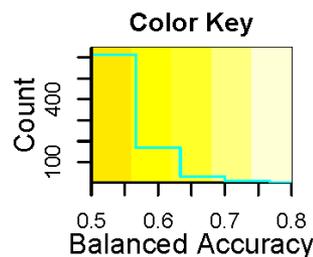


<http://actor.epa.gov/toxrefdb/>

Correlating Human *in vitro* and Rat *in vivo*



- Can find statistical associations with individual ToxCast *in vitro* assays at ToxRefDB endpoints
- Correlations are weak – the *n* for any given endpoint-assay combination is too low
 - You still need to know biology, *i.e.*, must have biological hypothesis
- Only looking at steady-state doses



- D-Maternal-PregnancyRelated-MaternalPregLoss (14)
- D-Maternal-PregnancyRelated (14)
- D-Maternal-GeneralMaternal-Systemic (42)
- D-Maternal-GeneralMaternal (42)
- D-Maternal (43)
- M-Liver (27)
- C-Tumorigen (27)
- D-Developmental-GeneralFetal-FetalWeightReduction (18)
- C-LiverTumors (12)
- C-SpleenPathology (10)
- C-LiverHypertrophy (26)
- D-Prenatal-Loss (14)
- C-KidneyNephropathy (10)
- M-FemaleReproductiveTract (11)
- M-LactationPND21 (10)
- C-LiverProliferativeLesions (25)
- M-Kidney (16)
- M-ViabilityPND4 (19)
- M-OffspringSurvival (22)
- M-GestationalInterval (10)
- M-ReproductivePerformance (18)
- M-LitterSize (14)
- M-ReproductiveOutcome (16)
- C-ThyroidHyperplasia (6)
- C-ThyroidTumors (12)
- C-LiverNecrosis (10)

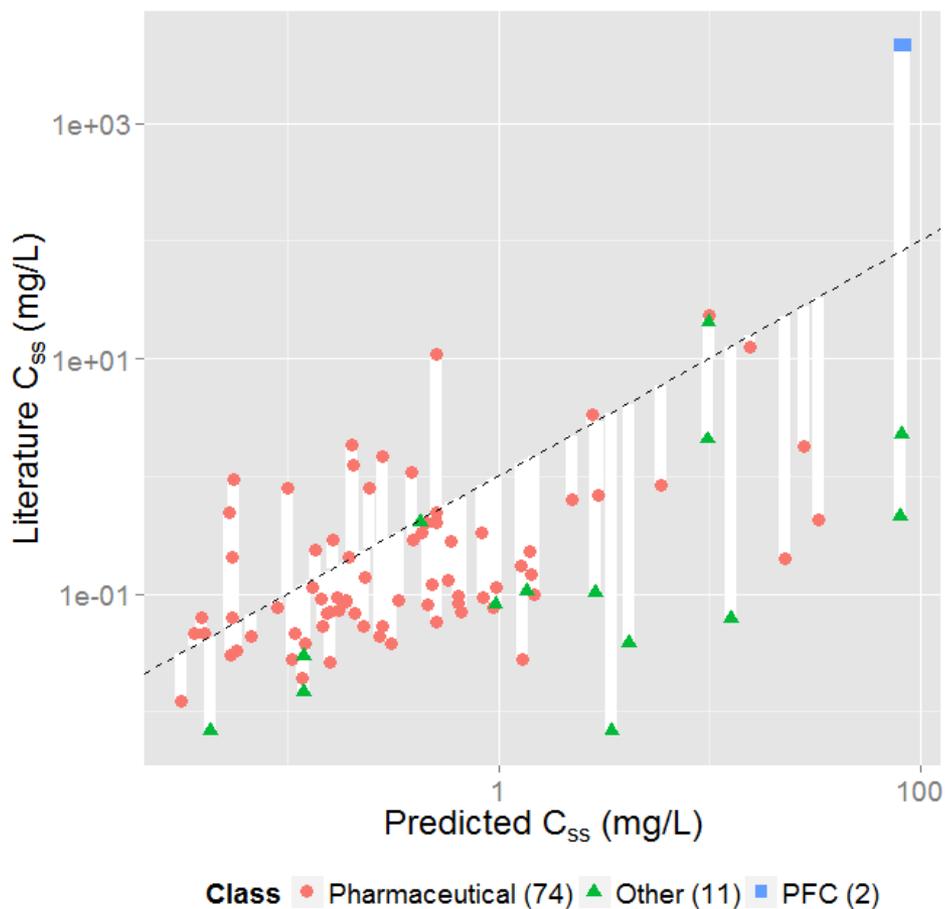
ToxCast Assays

Wetmore, *et al.* (2013)

In vivo Predictive Ability and Domain of Applicability

- In drug development, HTK methods estimate therapeutic doses for clinical studies – predicted concentrations are typically on the order of values measured in clinical trials (Wang, 2010)
- For environmental compounds, there will be no clinical trials
- Uncertainty must be well characterized ideally with rigorous statistical methodology
 - We will use direct comparison to *in vivo* data in order to get an empirical estimate of our uncertainty
 - Any approximations, omissions, or mistakes should work to increase the estimated uncertainty when evaluated systematically across chemicals

Using *in vivo* Data to Evaluate RTK



- When we compare the C_{ss} predicted from *in vitro* HTKK with *in vivo* C_{ss} values determined from the literature we find limited correlation ($R^2 \sim 0.34$)
- The dashed line indicates the identity (perfect predictor) line:
 - Over-predict for 65
 - Under-predict for 22
- The white lines indicate the discrepancy between measured and predicted values (the residual)

Predicting When RTK Will Work

- We can use computer algorithms to analyze chemical descriptors to try to predict when the residual will be small
- Factors included are:
 - Physico-chemical properties
 - Log(Kow), molecular weight, acid/base association constants (pKa), general pharmaceutical or perfluorinated compound classification
 - *In vitro* HTTK data
 - Plasma protein binding (F_{ub}) and hepatic clearance
 - Active chemical transport
 - Use quantitative structure activity relationships (QSARs) to predict likelihood each compound is a substrate for 17 different transporters (e.g, Sedykh et al, 2013)

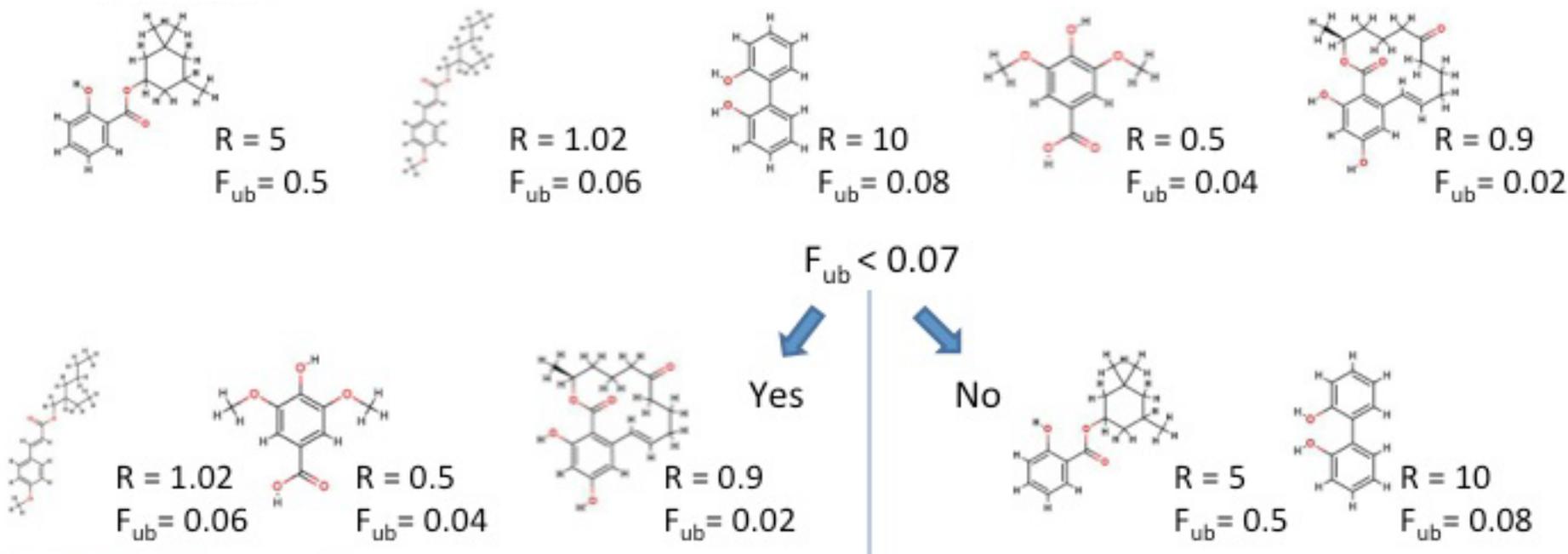
Transporter QSAR Predictions

From Alexander Sedykh and Alex Tropsha (UNC) and Sieto Bosgra (TNO)

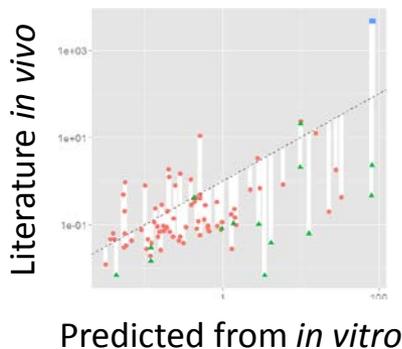
Compound	ASBT_pKm	ASBTx	BCRP	BSEP	MCT1	MDR1	MDR1_pKm	MRP1	MRP2	MRP3x	MRP4x	MRP5x	NTCPx	OATP2B1	OCT1x	PEPT1_pKm	PEPT1x
2,2-Bis(4-hydroxyphenyl)-1,1,1-trichloroethane (HPTe)	NA	0.006	0.482	0.394	0.564	0.163	4.425	0.45	0.301	0.36	0.224	0.464	0.341	0.532	NA	4.142	0.071
2,4-D	NA	0.012	0.405	0.378	0.584	0.082	4.389	0.388	0.233	0.252	0.213	0.387	0.259	0.283	0.748	3.765	0.121
2,5-Hexanedione	NA	0.031	0.288	NA	0.71	0.246	4.5	0.368	0.174	0.126	0.437	0.185	0.197	0.105	0.832	3.253	0.193
2-Phenylphenol	NA	0.007	0.451	0.456	0.744	0.168	4.638	0.097	0.244	0.192	0.2	0.443	0.192	0.283	0.957	3.969	0.116
4-(2,4-Dichlorophenoxy)butyric acid	NA	0.007	0.439	0.394	0.438	0.086	4.454	0.413	0.249	0.23	0.351	0.264	0.278	0.312	0.701	3.551	0.097
6-Desisopropylatrazine	NA	0.024	0.414	0.447	0.603	0.178	4.408	0.34	0.16	0.145	0.589	0.384	0.18	0.207	0.887	3.595	0.141
Abamectin	NA	0.167	0.388	0.45	NA	0.93	5.185	0.916	0.698	0.924	0.29	NA	NA	0.076	NA	NA	NA
Abamectin	NA	0.167	0.388	0.45	NA	0.93	5.185	0.916	0.698	0.924	0.29	NA	NA	0.076	NA	NA	NA
Acephate	NA	0.015	0.266	NA	0.626	0.129	4.444	0.585	0.187	0.216	0.546	0.17	0.203	0.143	0.648	3.23	0.124
Acetaminophen	NA	0.016	0.247	0.479	0.699	0.058	4.477	0.254	0.183	0.188	0.062	0.279	0.18	0.218	0.796	3.352	0.237
Acetamiprid	NA	0.011	0.6	0.34	0.545	0.162	4.326	0.415	0.169	0.143	0.431	0.29	0.159	0.276	0.797	3.572	0.208
Acetochlor	NA	0.013	0.327	0.54	0.403	0.162	4.511	0.456	0.233	0.296	0.538	0.177	0.142	0.31	0.528	3.831	0.153
Acetylsalicylic acid	NA	0.005	0.194	0.596	0.466	0.055	4.524	0.366	0.238	0.235	0.08	0.318	0.182	0.229	0.747	3.553	0.286
Acifluorfen	NA	NA	0.641	0.348	0.364	0.309	4.328	0.338	0.542	0.485	0.208	NA	0.37	0.711	0.169	NA	NA
Acrylamide	NA	NA	0.331	NA	0.913	0.268	4.639	0.298	0.162	0.126	0.22	0.206	NA	0.096	0.885	3.132	0.138
Aflatoxin	NA	0.012	0.537	0.626	0.468	0.468	4.565	0.783	0.386	0.23	0.179	0.509	0.144	0.224	0.49	NA	0.082
Alachlor	NA	0.012	0.326	0.537	0.413	0.19	4.522	0.451	0.211	0.236	0.538	0.179	0.137	0.299	0.574	3.872	0.143
Aldicarb	NA	0.02	0.27	0.51	0.495	0.064	4.463	0.553	0.178	0.239	0.569	0.156	NA	0.173	0.587	3.597	0.157

Recursive Partitioning Tree for Residuals

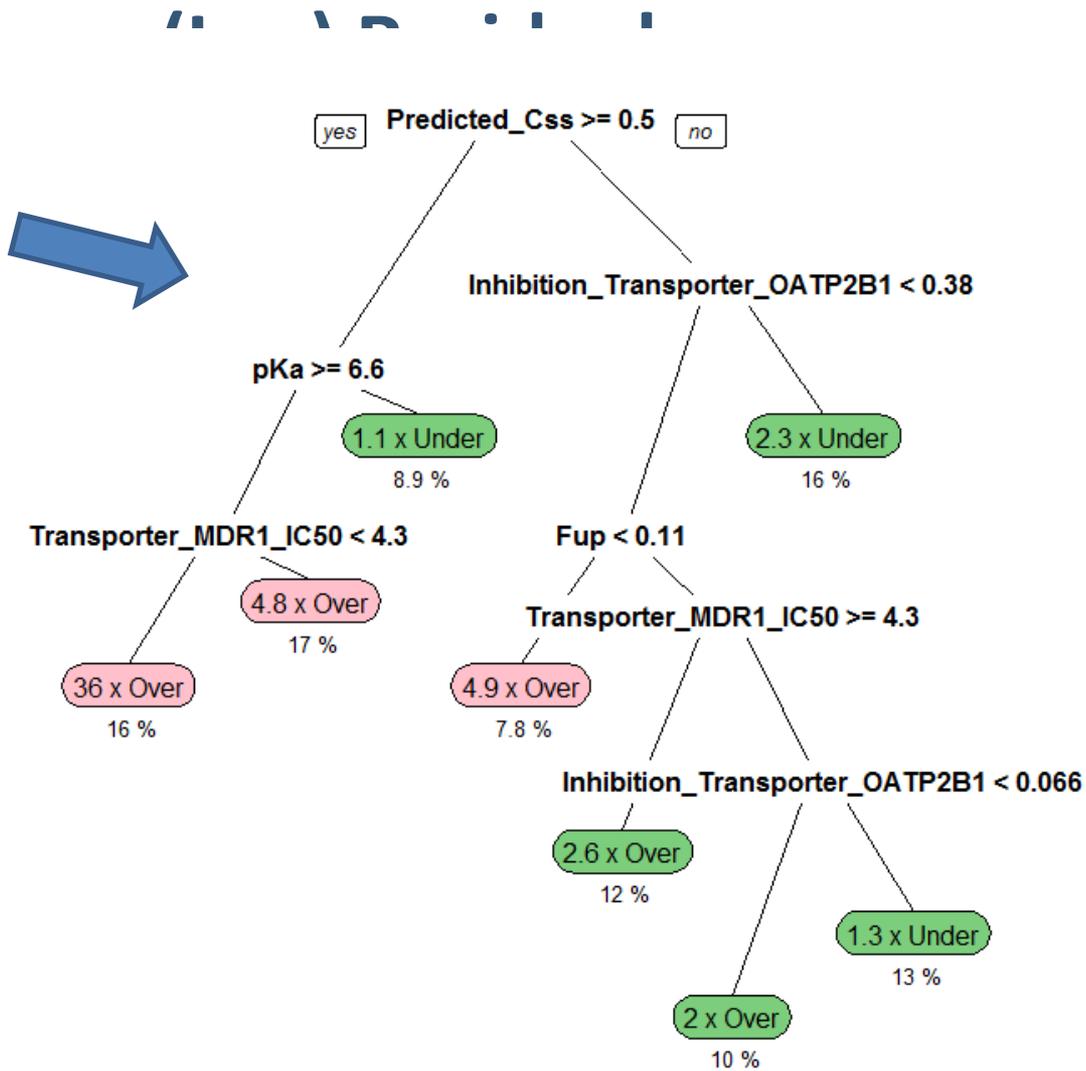
- The higher the C_{ss} , the lower the oral equivalent dose
- Ideally the residuals (difference between the literature value and the prediction) are small or $R \equiv C_{ss}^{lit.}/C_{ss}^{pred.} \approx 1$
- If a residual is large, we would prefer to over-predict C_{ss} to be conservative, *i.e.* $R < 1$



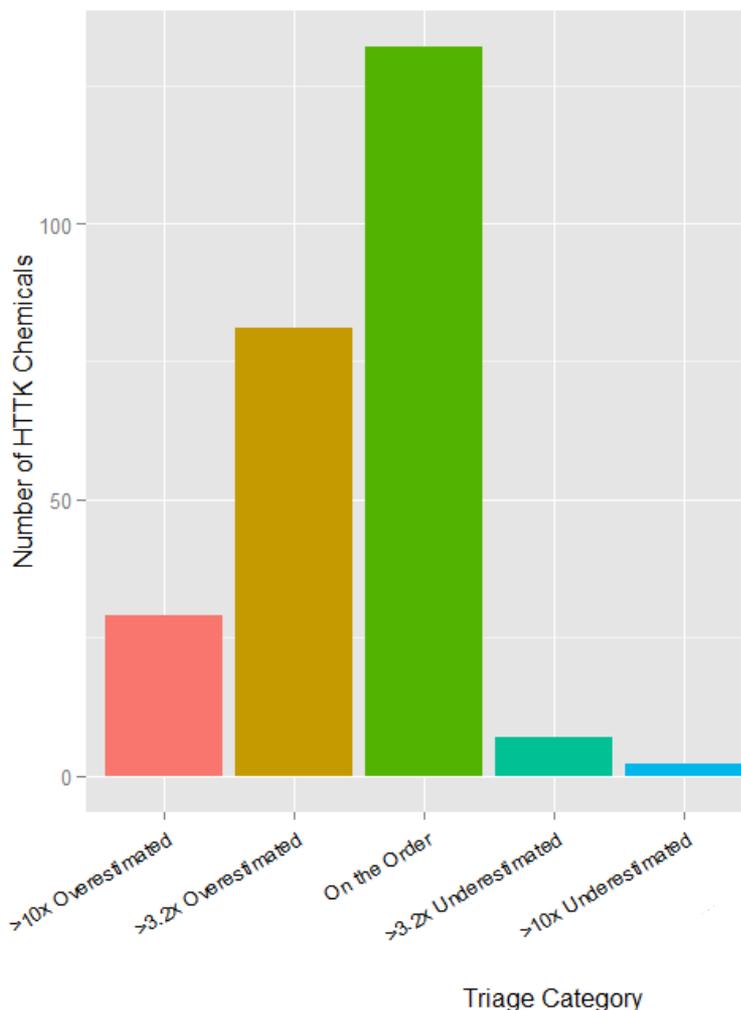
Recursive Partitioning Tree for



- Regression tree predicts expected residual based on physico-chemical properties, transporter QSARs, and *in vitro* HTTK data
- We can use this predicted error as a chemical-specific estimate of the accuracy of HTTK predict
- If the predicted C_{ss} underestimates value, the necessary exposure ρ will be higher



Evaluation of HTTK Performance and Domain of Applicability



- Through comparison to *in vivo* data, a cross-validated predictor (random forest, using 50,000 classifier trees) of success or failure of HTTK has been constructed
- The largest single class of chemicals fall into the category of “On the order” (within approximately a factor of three)
- More likely to overestimate (conservative error) than underestimate plasma concentrations from an exposure

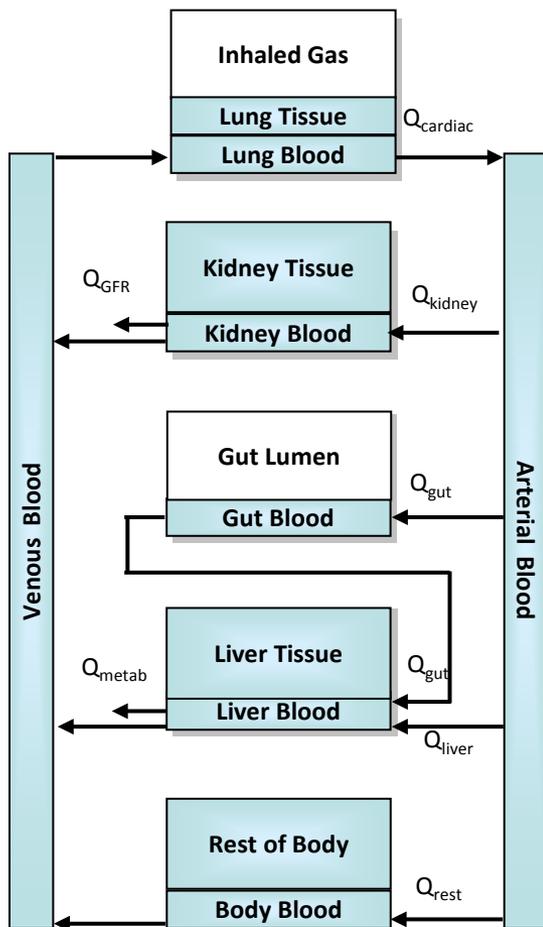
RTK Assumptions

- To date, the TK models used for environmental chemicals have been relatively simple, making three key assumptions:
 - 1) Whole body is at the same concentration (*i.e.*, plasma)
 - 2) Environmental exposure is constant and uniform (*i.e.*, constant infusion)
 - 3) Enough time has passed that the plasma concentration is at steady-state with respect to the environment
- QSARs for tissue-specific properties address the first
- We can test the second two assumptions using dynamic simulation (*e.g.*, more realistic, sporadic dosing) of physiologically-based toxicokinetic (PBTK) models

High Throughput Physiologically-based Toxicokinetic Models (HTPBTK)

- PBTK models typically require more data than simpler models like we used for C_{ss} , but we can build generic, high throughput PBTK (HTPBTK) models parameterized with:
 - the same *in vitro* HTTK data used for RTK, **plus**
 - QSARs for tissue-specific properties
 - Assumptions about unknown dynamic processes, such as absorption
- We use these HTPBTK models perform both simulation experiments and compare model predictions from *in vitro* data with human and rat *in vivo* measurements

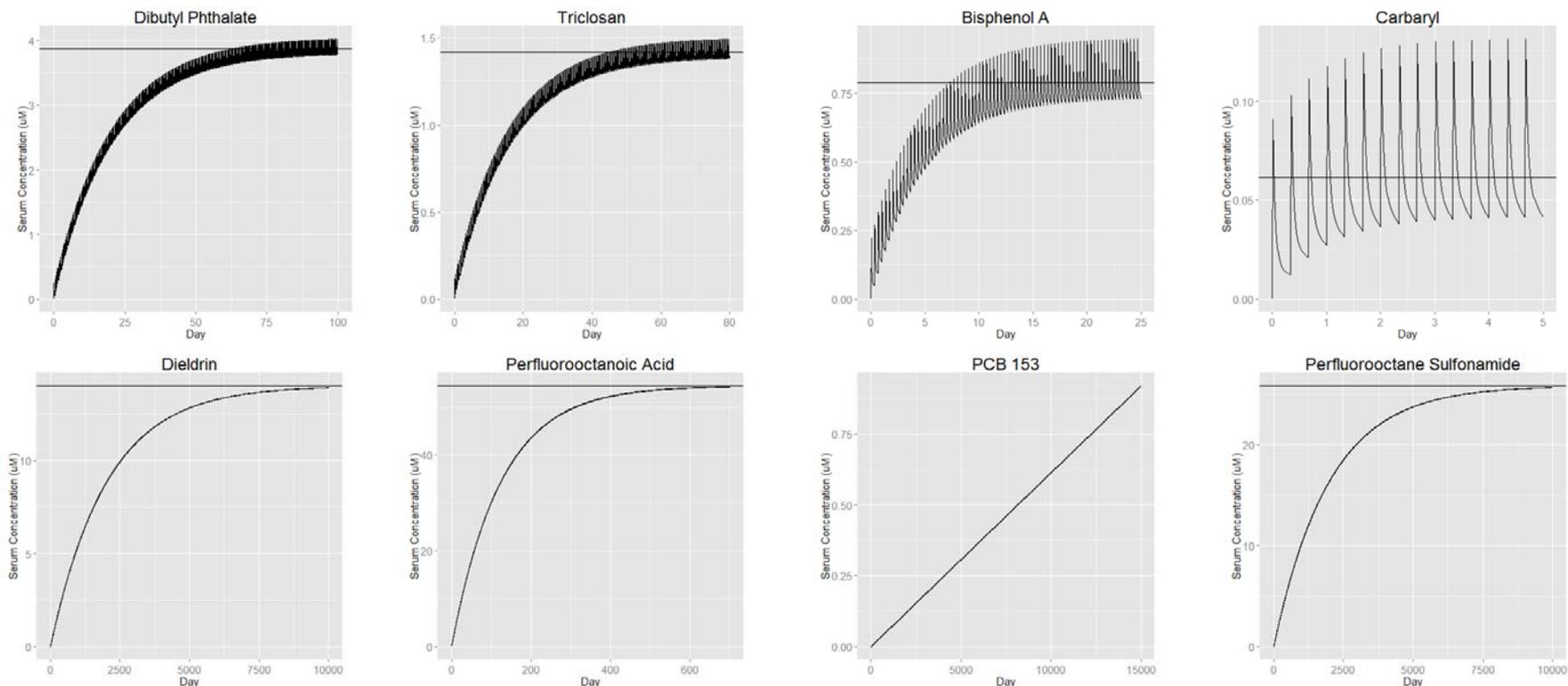
Physiologically-based Toxicokinetic (PBTK) Model



- Some tissues (*e.g.*, arterial blood) are simple compartments, while others (*e.g.*, kidney) are compound compartments consisting of separate blood and tissue sections.
- Some specific tissues (lung, kidney, gut, and liver) are modeled explicitly, others (*e.g.*, fat, brain, bones) are lumped into the “Rest of Body” compartment.
 - Partition coefficients from Schmitt (2008a and b)
 - Describe a specific species using chemical-independent physiology (Davies and Morris, 1993)
- Chemical enters the body primarily through oral absorption, but we don’t know absorption rate and bioavailability (assume “fast”, *i.e.* 1/h and 100%)
- The only way chemicals “leave” the body are through metabolism (change into a metabolite) in the liver or excretion by glomerular filtration into the proximal tubules of the kidney and out of the body

Evaluating RTK Assumptions

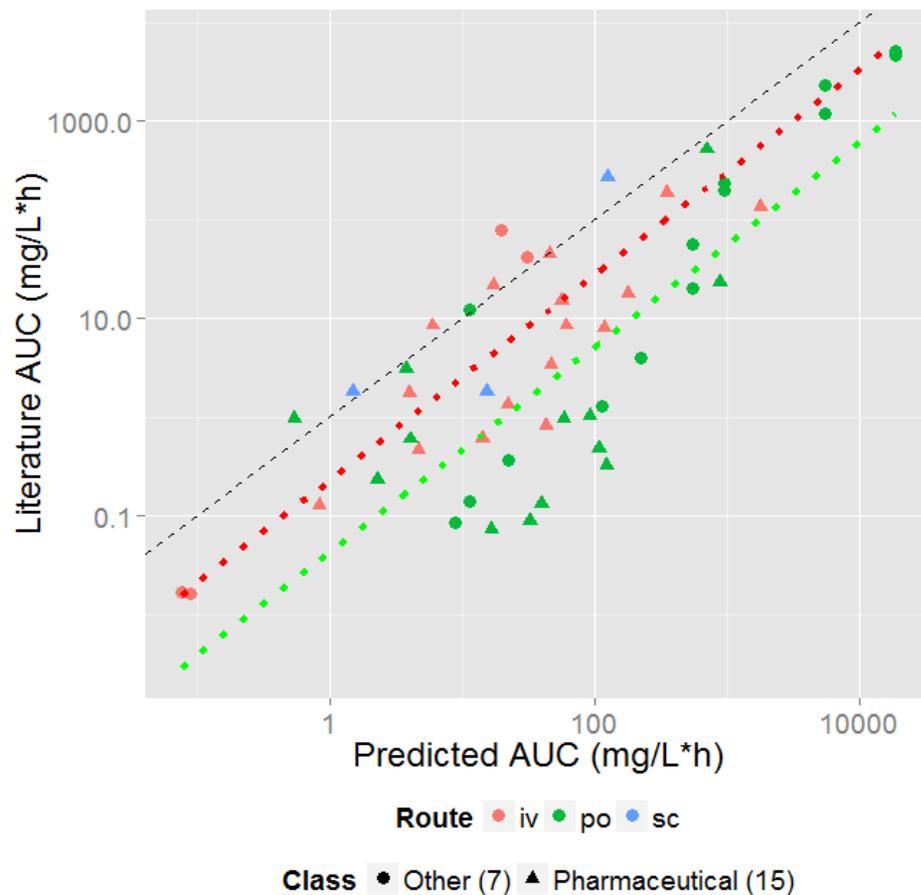
- Can use HTPBTK simulations to evaluate appropriateness of RTK assumptions
- Can use HTPBTK to predict traditional TK metrics, such as peak concentration (C_{max}) and time integrated area under the curve (AUC) for various tissues
- Below we show approach to “steady-state” due to three simulated daily doses



Evaluation Data for HTTK

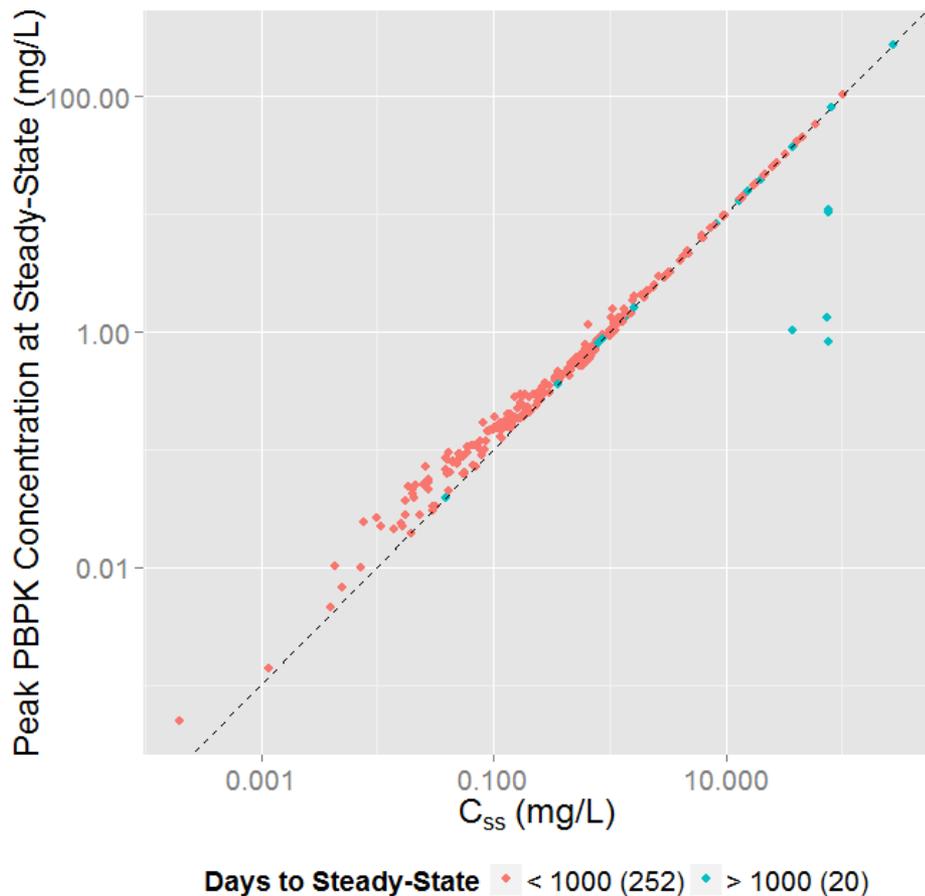
- For most non-therapeutic compounds, it is unlikely that there will ever be controlled human TK data
- Though some *in vivo* TK data exists to allow statistical assessment of HTTK predictions, these data are predominantly for pharmaceutical compounds
- Wetmore *et al.*, (2013): Rat HTTK data for 53 chemicals
- Compile and collect *in vivo* TK data for some or all of the rat HTTK compounds:
 - Allows evaluation of predictions based on *in vitro* and QSAR (*e.g.*, clearance and volume of distribution)
 - Allows measurement of other key processes (*e.g.*, absorption rates, extra-hepatic metabolism).

Evaluating HTPBTK Predictions from *In Vitro* Data



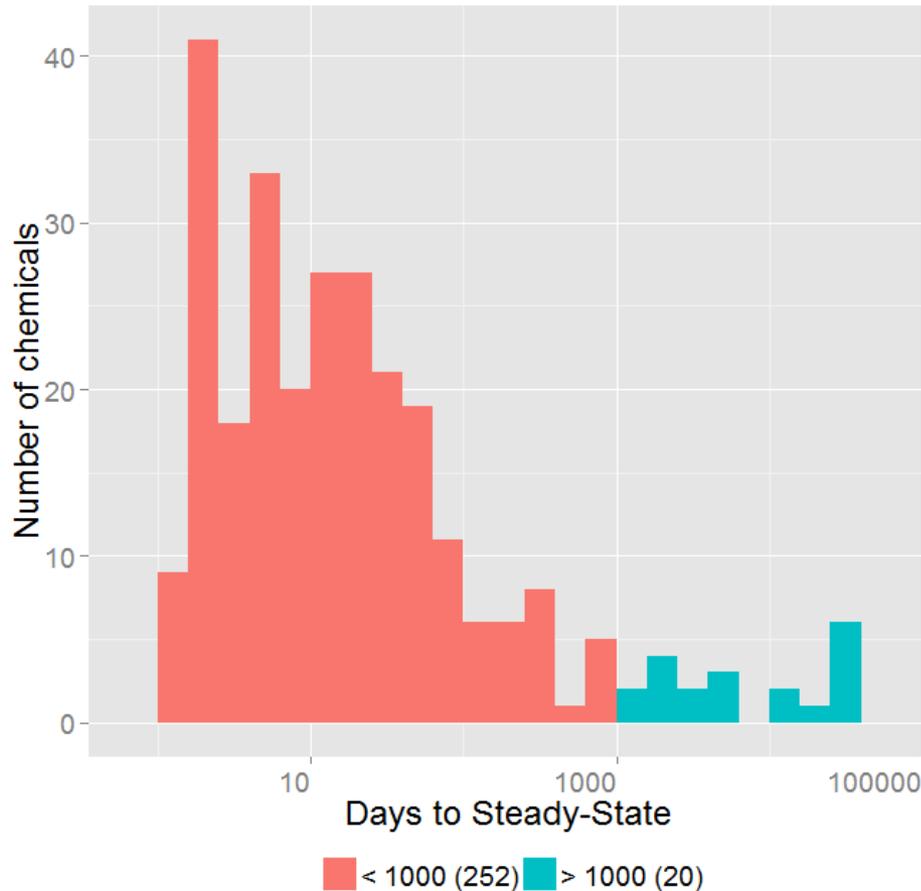
- HTPBTK predictions for the AUC (time integrated plasma concentration or Area Under the Curve)
- *in vivo* measurements from the literature for various treatments (dose and route) of rat ($R^2 \sim 0.69$)
- Predictions are generally conservative – *i.e.*, predicted AUC higher than measured
- Oral dose AUC $\sim 5.4x$ higher than intravenous dose AUC

Peak Concentration vs. C_{ss}



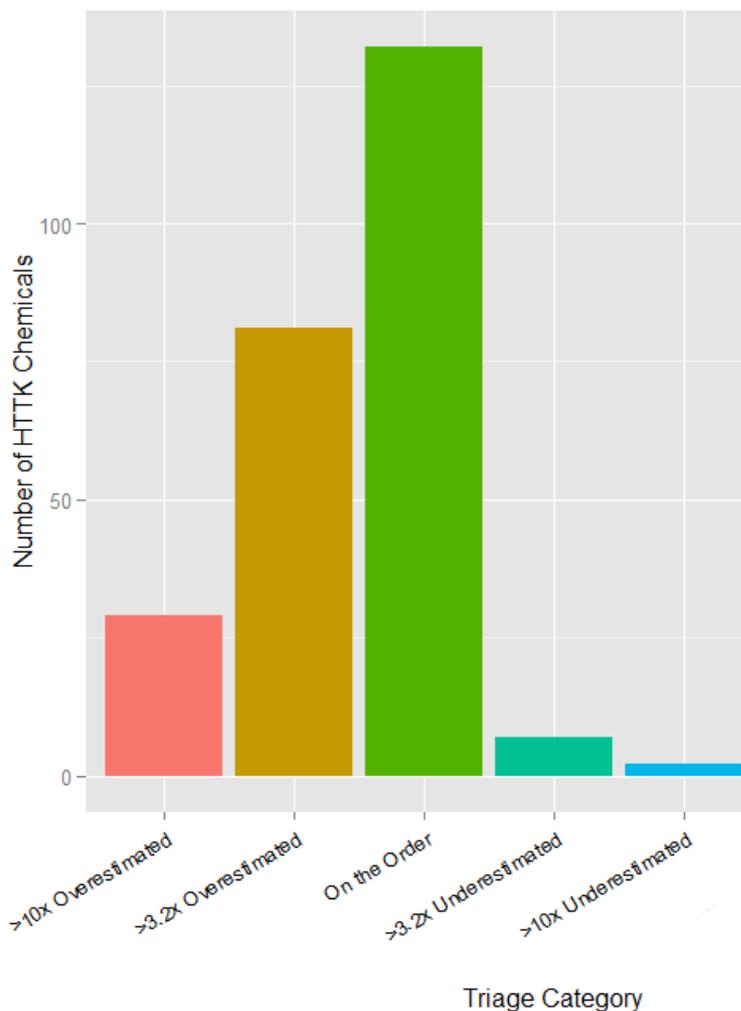
- Peak serum concentrations from the HTPBTK model are compared against the steady-state concentration predicted by the three compartment model for a constant infusion exposure (as in Wetmore *et al.* 2012)
- The dashed, identity (1:1) line indicates that for most compounds the peak concentrations are very similar to C_{ss}

Evaluation of Steady-State Predictions



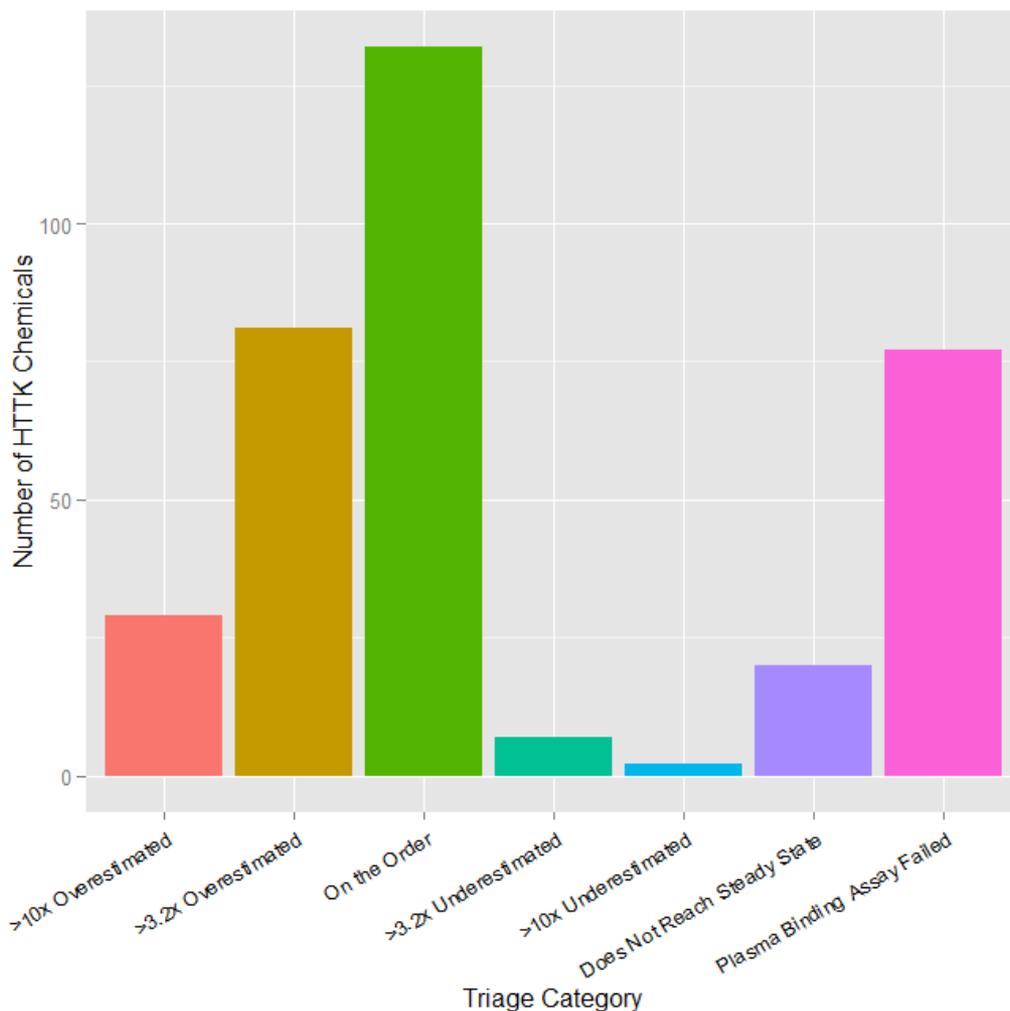
- Using HTPBTK and assuming three daily doses (every eight hours)
- This allows us to evaluate the plausibility of the steady-state dosing assumption.
- We find that the majority of chemicals reach steady state in a few weeks
- A second population of chemicals never reach steady state.

Evaluation of HTTK Performance and Domain of Applicability



- Through comparison to *in vivo* data, a cross-validated (random forest) predictor of success or failure of HTTK has been constructed

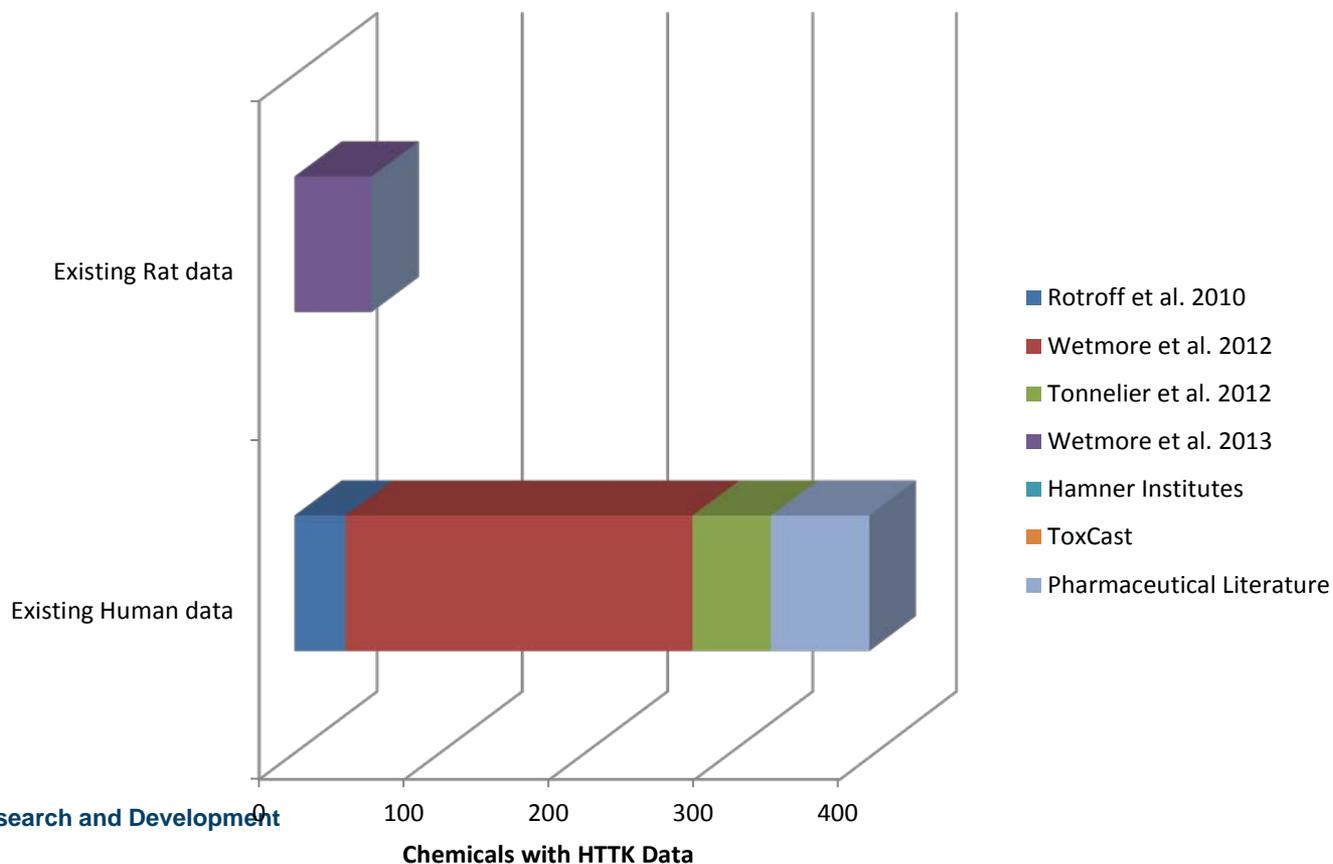
Evaluation of HTTK Performance and Domain of Applicability



- Through comparison to *in vivo* data, a cross-validated (random forest) predictor of success or failure of HTTK has been constructed
- Add categories for chemicals that do not reach steady-state or for which plasma binding assay fails

Chemicals with HTK Data

- *In vitro* assays limited by time needed to develop chemical-specific analytical chemistry method

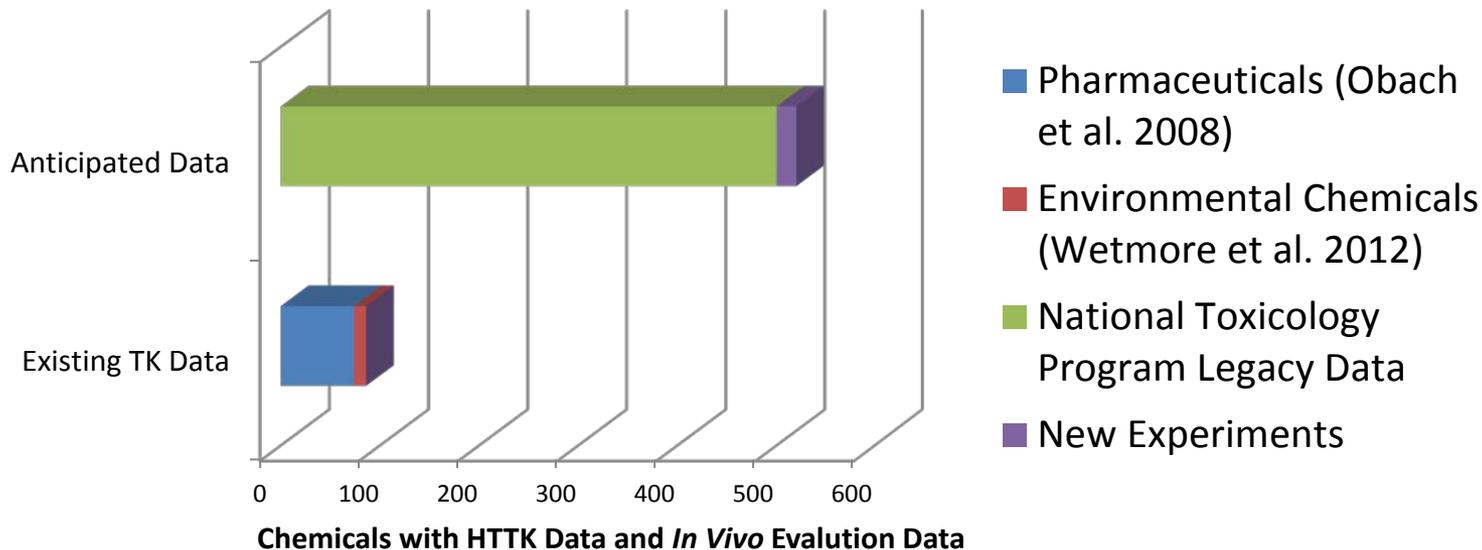


New Data for HHTK

	Intrinsic hepatic clearance and plasma protein binding data	
239 Chemicals	Wetmore <i>et al.</i> , (2012)	
181	To be published in 2015	
88	Collected Summer 2014	

- ToxCast HHTK testing:
 - Measuring metabolism by human hepatocytes
 - Improved assays for measuring binding of chemicals to human plasma protein
 - Obtain data on ToxCast chemicals not investigated by the Hamner Institute studies
 - Reinvestigate chemicals that proved difficult in previous efforts
- This data will eventually allow determination of human oral equivalent doses (mg/kg BW/day) for most ToxCast chemicals.

New Data for Evaluating HTTK Predictions



- Goal of HTTK modeling of pharmaceuticals is to determine efficacious doses for follow-on human clinical trials, the degree of confidence needed is different
- For environmental compounds, their uncertainty must be well characterized ideally with rigorous statistical methodology

HTTK Future Directions

- Working to make all data and models available as R package (“httk”)
- Collecting new HTTK data on chemicals using improved (serial dilution of plasma) methodology for measuring plasma binding
- Current MC simulations focus on adults
 - Expanding populations variability analysis to include sensitive population groups to reflect NHANES and beyond (e.g., children <6yo)
- Current *in vivo* data for evaluation of models is from heterogeneous studies. Ongoing efforts to
 - Collect data from limited *in vivo* studies (EPA/NHEERL and Research Triangle Institute)
 - Organize data from larger, systematic studies (e.g., National Toxicology Program) into computable format
 - Improved evaluation data will allow better assessment of predictive ability and determination of domain of applicability

The views expressed in this presentation are those of the author and do not necessarily reflect the views or policies of the U.S. EPA

Summary

- Toxicokinetics (TK) provides a bridge between HTS and HTE by predicting tissue concentrations due to exposure
- HTTK methods developed for pharmaceuticals have been adapted to environmental testing
- A primary application of HTTK is “Reverse Dosimetry” or RTK
 - Can infer daily doses that produce plasma concentrations equivalent to the bioactive concentrations, **but:**
 - Must consider domain of applicability
 - Chemical-specific analytical chemistry methods make HTTK slower than bioactivity HTS or HTE
- Although we used MC simulation to characterize some aspects of human variability (*e.g.*, body weight of adults), any key determinants of variability that are not included in our simulation have not been assessed
- **We must carefully characterize the uncertainty in our approach**

The views expressed in this presentation are those of the author and do not necessarily reflect the views or policies of the U.S. EPA



Chemical Safety for Sustainability (CSS) Rapid Exposure and Dosimetry (RED) Project

NCCT

Chris Grulke

Richard Judson

Thomas Knudsen

Chantel Nicolas*

Robert Pearce*

James Rabinowitz

Caroline Ring*

Woody Setzer

Imran Shah

Rusty Thomas

NRMRL

Xiaoyu Liu

NHEERL

Jane Ellen Simmons

Marina Evans

Mike Hughes

NERL

Craig Barber

Dalizza Colon

Kathie Dionisio*

Peter Egeghy

Kim Gaetz

Kristin Isaacs

Haluk Ozkaynak

Julia Rager*

Mark Strynar

Jon Sobus

Mike Tornero-Velez

Dan Vallero

High Throughput Toxicokinetics Researchers

***Trainees**

Collaborators

Arnot Research and Consulting

Jon Arnot

Chemical Computing Group

Rocky Goldsmith

Environmental Protection Agency

Alicia Frame

Hamner Institutes

Barbara Wetmore

Cory Strobe

Indiana University

James Sluka

Michigan State University

Jade Mitchell

**National Institute for Environmental Health
Sciences (NIEHS)**

Mike Devito

Nisha Sipes

Kyla Taylor

Kristina Thayer

**Netherlands Organisation for Applied
Scientific Research (TNO)**

Sieto Bosgra

North Carolina State University

Anran Wang

Research Triangle Institute

Timothy Fennell

Silent Spring Institute

Robin Dodson

University of California, Davis

Deborah Bennett

University of Michigan

Olivier Jolliet

University of North Carolina, Chapel Hill

Alexander Sedykh*

Alex Tropsha

References

- Davies, B. and Morris, T. (1993). Physiological Parameters in Laboratory Animals and Humans. *Pharmaceutical Research* **10**(7), 1093-1095, 10.1023/a:1018943613122.
- Egeghy, Peter P., et al. "The exposure data landscape for manufactured chemicals." *Science of the Total Environment* 414: 159-166 (2012)
- Jamei, M., et al. (2009). The Simcyp population-based ADME simulator. *Expert opinion on drug metabolism & toxicology* **5**(2), 211-23, 10.1517/17425250802691074.
- Judson, R. S., et al. (2011). Estimating Toxicity-Related Biological Pathway Altering Doses for High-Throughput Chemical Risk Assessment. *Chemical Research in Toxicology* **24**(4), 451-462, 10.1021/tx100428e.
- Martin, M. T., et al. (2009) Profiling the Reproductive Toxicity of Chemicals from Multigeneration Studies in the Toxicity Reference Database. *Toxicological Sciences* **110**(1), 181-190
- Rotroff, D. M., et al. (2010). Incorporating human dosimetry and exposure into high-throughput in vitro toxicity screening. *Toxicological Sciences* **117**(2), 348-358, 10.1093/toxsci/kfq220.
- Schmitt, W. (2008a). General approach for the calculation of tissue to plasma partition coefficients. *Toxicology in vitro : an international journal published in association with BIBRA* **22**(2), 457-67, 10.1016/j.tiv.2007.09.010.
- Schmitt, W. (2008b). Corrigendum to: "General approach for the calculation of tissue to plasma partition coefficients" [*Toxicology in Vitro* 22 (2008) 457-467]. *Toxicology in Vitro* **22**(6), 1666, <http://dx.doi.org/10.1016/j.tiv.2008.04.020>.
- Sedykh, A., et al. (2013). Human intestinal transporter database: QSAR modeling and virtual profiling of drug uptake, efflux and interactions. *Pharm Res* **30**(4), 996-1007, 10.1007/s11095-012-0935-x.
- Shibata, Y., et al. (2002). Prediction of Hepatic Clearance and Availability by Cryopreserved Human Hepatocytes: An Application of Serum Incubation Method. *Drug Metabolism and Disposition* **30**(8), 892-896, 10.1124/dmd.30.8.892.
- Wambaugh, J. F., et al. (2014). High Throughput Heuristics for Prioritizing Human Exposure to Environmental Chemicals. *Environmental Science & Technology*, 10.1021/es503583j.
- Wang, Y.-H. (2010). Confidence Assessment of the Simcyp Time-Based Approach and a Static Mathematical Model in Predicting Clinical Drug-Drug Interactions for Mechanism-Based CYP3A Inhibitors. *Drug Metabolism and Disposition* **38**(7), 1094-1104, 10.1124/dmd.110.032177.
- Waters, N. J., et al. (2008). Validation of a rapid equilibrium dialysis approach for the measurement of plasma protein binding. *Journal of Pharmaceutical Sciences* **97**(10), 4586-4595, 10.1002/jps.21317.
- Wetmore, B. A., et al. (2012). Integration of dosimetry, exposure, and high-throughput screening data in chemical toxicity assessment. *Toxicological sciences : an official journal of the Society of Toxicology* **125**(1), 157-74, 10.1093/toxsci/kfr254.
- Wetmore, B. A., et al. (2013). Relative Impact of Incorporating Pharmacokinetics on Predicting In Vivo Hazard and Mode of Action from High-Throughput In Vitro Toxicity Assays. *Toxicological Sciences* **132**(2), 327-346, 10.1093/toxsci/kft012.
- Wetmore, B. A., et al. (2014). Incorporating Population Variability and Susceptible Subpopulations into Dosimetry for High-Throughput Toxicity Testing. *Toxicological Sciences*, kfu169.