

NTP TECHNICAL REPORT ON THE TOXICOLOGY AND CARCINOGENESIS STUDIES OF 2,3,7,8-TETRACHLORODIBENZO-*P*-DIOXIN (TCDD) (CAS No. 1746-01-6) IN FEMALE HARLAN

SPRAGUE-DAWLEY RATS (GAVAGE STUDIES)

NTP TR 521

APRIL 2006

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# **TOXICOLOGY AND CARCINOGENESIS**

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(CAS NO. 1746-01-6)

# **IN FEMALE HARLAN SPRAGUE-DAWLEY RATS**

(GAVAGE STUDIES)



NATIONAL TOXICOLOGY PROGRAM P.O. Box 12233 Research Triangle Park, NC 27709

April 2006

# NTP TR 521

NIH Publication No. 06-4468

National Institutes of Health Public Health Service U.S. DEPARTMENT OF HEALTH AND HUMAN SERVICES

#### FOREWORD

The National Toxicology Program (NTP) is an interagency program within the Public Health Service (PHS) of the Department of Health and Human Services (HHS) and is headquartered at the National Institute of Environmental Health Sciences of the National Institutes of Health (NIEHS/NIH). Three agencies contribute resources to the program: NIEHS/NIH, the National Institute for Occupational Safety and Health of the Centers for Disease Control and Prevention (NIOSH/CDC), and the National Center for Toxicological Research of the Food and Drug Administration (NCTR/FDA) Established in 1978, the NTP is charged with coordinating toxicological testing activities, strengthening the science base in toxicology, developing and validating improved testing methods, and providing information about potentially toxic substances to health regulatory and research agencies, scientific and medical communities, and the public.

The Technical Report series began in 1976 with carcinogenesis studies conducted by the National Cancer Institute. In 1981, this bioassay program was transferred to the NTP. The studies described in the Technical Report series are designed and conducted to characterize and evaluate the toxicologic potential, including carcinogenic activity, of selected substances in laboratory animals (usually two species, rats and mice). Substances selected for NTP toxicity and carcinogenicity studies are chosen primarily on the basis of human exposure, level of production, and chemical structure. The interpretive conclusions presented in NTP Technical Reports are based only on the results of these NTP studies. Extrapolation of these results to other species including characterization of hazards and risks to humans requires analyses beyond the intent of these reports. Selection *per se* is not an indicator of a substance's carcinogenic potential.

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# CONTRIBUTORS

#### **National Toxicology Program**

Evaluated and interpreted results and reported findings

N.J. Walker, Ph.D., Study Scientist
A. Nyska, D.V.M., Study Pathologist
D.W. Bristol, Ph.D.
J.R. Bucher, Ph.D.
L.T. Burka, Ph.D.
J.R. Hailey, D.V.M.
J.K. Haseman, Ph.D.
R.A. Herbert, D.V.M., Ph.D.
R.R. Maronpot, D.V.M.
D.P. Orzech, M.S.
C.S. Smith, Ph.D.
G.S. Travlos, D.V.M.
K.L. Witt, M.S.
M.E. Wyde, Ph.D.

#### **Battelle Columbus Operations**

Conducted study and evaluated pathology findings

M.R. Hejtmancik, Ph.D., Principal Investigator D.M. Sells, D.V.M., Ph.D.

#### **Experimental Pathology Laboratories, Inc.**

Provided pathology review

M.H. Hamlin, II, D.V.M., Principal Investigator A.E. Brix, D.V.M., Ph.D. K.J. Cimon, D.V.M., M.S.

#### **Dynamac Corporation**

Prepared quality assurance audits

S. Brecher, Ph.D., Principal Investigator

### Constella Group, Inc.

Provided statistical analyses

P.W. Crockett, Ph.D., Principal Investigator L.J. Betz, M.S. M.E. Easterling, Ph.D. K.P. McGowan, M.B.A. J.T. Scott, M.S.

### **NTP Pathology Working Group**

*Evaluated slides and prepared pathology report* (June 18, 2002)

M.P. Jokinen, D.V.M., Chairperson Pathology Associates International A.E. Brix, D.V.M., Ph.D. Experimental Pathology Laboratories, Inc. K.J. Cimon, D.V.M., M.S. Experimental Pathology Laboratories, Inc. G.P. Flake, M.D. National Toxicology Program J.R. Hailey, D.V.M. National Toxicology Program B.F. Hamilton, D.V.M., Ph.D. GlaxoSmithKline R.A. Herbert, D.V.M., Ph.D. National Toxicology Program R.R. Maronpot, D.V.M. National Toxicology Program J. Nold, D.V.M. GlaxoSmithKline A. Nyska, D.V.M. National Toxicology Program D.M. Sells, D.V.M., Ph.D. Battelle Columbus Operations

#### **Biotechnical Services, Inc.**

Prepared Technical Report

S.R. Gunnels, M.A., Principal Investigator C.M. Bailey, B.S. Pharm. K.K. Coker, Ph.D. P.A. Gideon, B.A. L.M. Harper, B.S. M.C. Joheim, M.S. E.S. Rathman, M.S. D.C. Serbus, Ph.D.

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#### **SUMMARY**

#### Background

2,3,7,8-Tetrachlorobenzo-*p*-dioxin (TCDD) is one of a large family of hydrocarbons containing chlorine known as dioxins. Some dioxins or dioxin-like compounds are highly toxic and cause cancer, and usually contaminated sites contain many different varieties of these dioxin-like compounds. The National Toxicology Program conducted a series of studies to try to gauge the relative toxicity of some of the more prevalent of these compounds both alone and in mixtures. This study evaluated the effects of TCDD on female rats for comparison with the potency of other chemicals in that family.

#### Methods

We exposed groups of 53 or 54 female rats by depositing solutions of TCDD dissolved in corn oil through tubes directly into their stomachs five days a week for two years. Daily doses of TCDD were 3, 10, 22, 46, or 100 nanograms (ng) per kilogram of body weight. Animals receiving corn oil alone served as the control group. Tissues from more than 40 sites were examined for every animal.

#### Results

Exposure to TCDD caused a variety of diseases in several organs. Cancers of the liver, lung, mouth, and uterus were seen in female rats exposed to TCDD. A variety of other toxic lesions observed in exposed animals included hypertrophy, hyperplasia, fibrosis, and necrosis of the liver, hyperplasia of the oral mucosa, metaplasia of the lung, inflammation and atrophy of the pancreas, kidney nephropathy, cardiomyopathy of the heart, atrophy and hyperplasia of the adrenal gland, atrophy of the thymus, hyperplasia of the forestomach, hypertrophy of the thyroid gland, and inflammation of the mesentery.

#### Conclusions

We conclude that TCDD caused cancer and other toxic effects at several sites in female rats.

# ABSTRACT

# **DIOXIN TOXIC EQUIVALENCY FACTOR EVALUATION OVERVIEW**

Polyhalogenated aromatic hydrocarbons such as 2,3,7,8tetrachlorodibenzo-*p*-dioxin (TCDD) have the ability to bind to and activate the ligand-activated transcription factor, the aryl hydrocarbon receptor (AhR). Structurally related compounds that bind to the AhR and exhibit biological actions similar to TCDD are commonly referred to as "dioxin-like compounds" (DLCs). Ambient human exposure to DLCs occurs through the ingestion of foods containing residues of DLCs that bioconcentrate through the food chain. Due to their lipophilicity and persistence, once internalized, they accumulate in body tissue, mainly adipose, resulting in chronic lifetime human exposure.

Since human exposure to DLCs always involves a complex mixture, the toxic equivalency factor (TEF) methodology has been developed as a mathematical tool to assess the health risk posed by complex mixtures of these compounds. The TEF methodology is a relative potency scheme that ranks the dioxin-like activity of a compound relative to TCDD, which is the most potent congener. This allows for the estimation of the potential dioxin-like activity of a mixture of chemicals based on a common mechanism of action involving an initial binding of DLCs to the AhR.

The toxic equivalency of DLCs was nominated for evaluation because of the widespread human exposure to DLCs and the lack of data on the adequacy of the TEF methodology for predicting relative potency for cancer risk. To address this, the National Toxicology Program conducted a series of 2-year bioassays in female Harlan Sprague-Dawley rats to evaluate the chronic toxicity and carcinogenicity of DLCs and structurally related polychlorinated biphenyls (PCBs) and mixtures of these compounds.



#### 2,3,7,8-Tetrachlorodibenzo-*p*-dioxin TCDD

CAS No. 1746-01-6

Chemical Formula:  $C_{12}H_4Cl_4O_2$  Molecular Weight: 321.98

Synonyms: Dioxin; dioxine; TCDBD; 2,3,7,8-TCDD; 2,3,7,8-tetrachlorodibenzo(b,e)(1,4)dioxin; 2,3,7,8-tetrachlorodibenzo-*p*-dioxin; 2,3,6,7-tetrachlorodibenzo-*p*-dioxin; 2,3,7,8-tetrachlorodibenzo-1,4-dioxin; tetrachlorodibenzodioxin; 2,3,6,7-tetrachlorodibenzo-*p*-dioxin; 2,3,7,8-tetrachlorodibenzo-1,4-dioxin; tetrachlorodibenzodioxin; 2,3,6,7-tetrachlorodibenzo-1,4-dioxin; tetrachlorodibenzodioxin; 2,3,6,7-tetrachlorodibenzo-1,4-dioxin; tetrachlorodibenzo-1,4-dioxin; tetrachlorodibenzo-1,4

TCDD is not manufactured commercially other than for The main sources of scientific research purposes. TCDD releases into the environment are from combustion and incineration; metal smelting, refining, and processing; chemical manufacturing and processing; biological and photochemical processes; and existing reservoir sources that reflect past releases. TCDD (dioxin) was selected for study by the National Toxicology Program as a part of the dioxin TEF evaluation to assess the cancer risk posed by complex mixtures of polychlorinated dibenzodioxins (PCDDs), polychlorinated dibenzofurans (PCDFs), and PCBs. The dioxin TEF evaluation includes conducting multiple 2-year rat bioassays to evaluate the relative chronic toxicity and carcinogenicity of DLCs, structurally related PCBs, and mixtures of these compounds. While one of the aims of the dioxin TEF evaluation was a comparative analysis across studies, in this Technical Report, only the TCDD results are presented and discussed. TCDD was included because it is the reference compound for the dioxin TEF methodology. Female Harlan Sprague-Dawley rats were administered TCDD (at least 98% pure) in corn oil:acetone (99:1) by gavage for 14, 31, or 53 weeks or 2 years. Genetic toxicology studies were conducted in Salmonella typhimurium, L5178Y mouse lymphoma cells, cultured Chinese hamster ovary cells, Drosophila melanogaster, and mouse bone marrow cells.

## **2-YEAR STUDY**

Groups of 81 or 82 female rats were administered 3, 10, 22, 46, or 100 ng TCDD/kg body weight in corn oil:acetone (99:1) by gavage, 5 days per week, for up to 105 weeks; a group of 81 vehicle control female rats received the corn oil/acetone vehicle alone. Up to 10 rats per group were evaluated at 14, 31, or 53 weeks. A stop-exposure group of 50 female rats was administered 100 ng/kg TCDD in corn oil:acetone (99:1) by gavage for 30 weeks and then just the vehicle for the remainder of the study.

Survival of dosed groups was similar to that of the vehicle control group. Mean body weights of 100 ng/kg core study and stop-exposure groups were less than those of the vehicle control group after week 13 of the study. Mean body weights of 46 ng/kg rats were less than those of the vehicle controls during year 2 of the study, and those of 22 ng/kg rats were less than those of the vehicle controls the last 10 weeks of the study.

#### **Thyroid Hormone Concentrations**

Alterations in serum thyroid hormone levels were evaluated at the 14-, 31- and 53-week interim evaluations. At 14 weeks, there were significant decreases in serum total and free thyroxine ( $T_4$ ) levels and increases in serum total triiodothyronine ( $T_3$ ) and thyroid stimulating hormone (TSH). At 31 weeks, there were significant decreases in serum total and free  $T_4$  levels and increases in serum total  $T_3$  but no significant effect on TSH. At 53 weeks, there were significant decreases in serum total  $T_4$  levels and increases in serum total  $T_3$ . There were no significant effects on total  $T_4$  or TSH levels.

#### Hepatic Cell Proliferation Data

To evaluate hepatocyte replication, analysis of labeling of replicating hepatocytes with 5-bromo-2'-deoxyuridine was conducted at the 14-, 31-, and 53-week interim evaluations. The hepatocellular labeling index was significantly higher in the 22 ng/kg group compared to vehicle controls at 14 weeks. At the 31-week interim evaluation, the labeling indices in hepatocytes were significantly higher in all dosed groups than in the vehicle controls. At 53 weeks, labeling indices were significantly higher in the 46 and 100 ng/kg groups than in the vehicle controls.

#### Cytochrome P450 Enzyme Activities

To evaluate the expression of known dioxin-responsive genes, CYP1A1-associated 7-ethoxyresorufin-*O*deethylase (EROD) activity and CYP1A2-associated acetanilide-4-hydroxylase (A4H) activity were evaluated at 14, 31, and 53 weeks. In addition, pentoxyresorufin-*O*-deethylase (PROD) activity was analyzed. Hepatic EROD, PROD, and A4H activities were significantly higher in all dosed groups relative to vehicle controls at the 14-, 31-, and 53-week interim evaluations. Pulmonary EROD was also significantly higher in all dosed groups compared to vehicle controls at 14, 31, and 53 weeks.

# Determinations of TCDD Concentrations in Tissues

The tissue disposition of TCDD was analyzed in the liver, lung, fat, and blood of all animals in each group at the 14-, 31-, and 53-week interim evaluations and in 10 animals per group at the end of the 2-year study (105 weeks). The highest concentrations of TCDD were observed in the liver, followed by fat tissue. Liver and fat tissue concentrations of TCDD increased with increasing doses of TCDD. No measurable concentrations of TCDD were observed in blood from vehicle control or treated rats at any of the interim evaluations. Mean levels of TCDD in the liver and fat in the 100 ng/kg group at the end of the 2-year study were 9.3 and 3.2 ng/g, respectively. In liver tissue from the stopexposure group, TCDD concentrations were slightly higher than those observed in the 3 ng/kg group. In the stop-exposure group, TCDD concentrations in fat were below the limits of quantitation.

Absolute and/or relative liver weights were increased at 14, 31, and 53 weeks, with more severe effects occurring in the higher dosed groups. Increased liver weights correlated with increased incidences of hepatocyte hypertrophy at 14, 31, and 53 weeks.

Exposure led to a treatment-related increase in hepatic toxicity with a broad spectrum of lesions. Incidences and severities of lesions increased at higher doses and longer durations of exposure. The earliest effects were increased incidences and severities of hepatocyte hypertrophy at 14 weeks. At 2 years, there was a significant increase in toxic hepatopathy characterized by increased incidences of numerous nonneoplastic liver lesions including hepatocyte hypertrophy, multinucleated hepatocytes, altered hepatocellular foci, inflammation, pigmentation, diffuse fatty change, necrosis, portal fibrosis, oval cell hyperplasia, bile duct hyperplasia, bile duct cysts, cholangiofibrosis, and nodular hyperplasia

At 2 years, the incidence of hepatocellular adenoma was significantly increased in the 100 ng/kg core study group. Dose-related increased incidences of cholangiocarcinoma were seen in core study rats administered 22 ng/kg or greater. The highest incidence of cholangiocarcinoma was seen in the 100 ng/kg core study group and included a significant number of animals with multiple cholangiocarcinomas. Two cholangiocarcinomas and two hepatocellular adenomas were seen in the 100 ng/kg stop-exposure group. Two hepatocholangiomas were seen in the 100 ng/kg core study group, and one cholangioma was seen in the 100 ng/kg stop-exposure group.

In the lung, the incidence of cystic keratinizing epithelioma of the lung was significantly increased at 2 years in the 100 ng/kg core study group. Nonneoplastic effects in the lung included increased incidences of bronchiolar metaplasia.

The incidence of gingival squamous cell carcinoma of the oral mucosa was significantly increased in the 100 ng/kg core study group at 2 years and was accompanied by an increased incidence of gingival squamous hyperplasia.

At 2 years, the incidence of squamous cell carcinoma of the uterus in the 46 ng/kg group was significantly increased, and there were two squamous cell carcinomas in the 100 ng/kg stop-exposure group.

At 2 years, one acinar adenoma and two acinar cell carcinomas of the pancreas were seen in the 100 ng/kg core study group; one acinar carcinoma was seen in the 100 ng/kg stop-exposure group. The incidences of acinar cell adenoma or carcinoma (combined) exceeded the historical vehicle control range. Nonneoplastic effects in the lung included acinar cytoplasmic vacuolization, chronic active inflammation, acinar atrophy, and arterial chronic active inflammation.

Numerous nonneoplastic effects were seen in other organs including thymic atrophy, adrenal cortex atrophy, adrenal cortex hyperplasia, cardiomyopathy, mesenteric artery inflammation, clitoral gland cysts, nephropathy, squamous hyperplasia of the forestomach, and thyroid gland follicular cell hypertrophy. A decrease in the incidence of ovarian atrophy was also observed.

# **GENETIC TOXICOLOGY**

TCDD was not mutagenic in any of several *in vitro* and *in vivo* short-term tests. No induction of gene mutations was seen in *S. typhimurium* strains TA98, TA100, TA1535, or TA1537 exposed to TCDD with or without S9 activation enzymes. No induction of trifluorothymidine resistance (gene mutations) was observed in L5178Y tk<sup>+/-</sup> mouse lymphoma cells tested with or without S9 activation. In cytogenetic tests with cultured Chinese hamster ovary cells, no consistently reproducible induction of sister chromatid exchanges or chromosomal aberrations were noted after culturing with

TCDD with or without S9. *In vivo* tests in male *D. melanogaster* showed no induction of sex-linked recessive lethal mutations in germ cells following treatment of adult flies by injection with TCDD. In male  $B6C3F_1$  mice, no increases in the frequency of chromosomally aberrant cells were seen in bone marrow samples taken at two different posttreatment sampling times.

#### CONCLUSIONS

Under the conditions of this 2-year gavage study, there was *clear evidence of carcinogenic activity*\* of TCDD in female Harlan Sprague-Dawley rats based on increased incidences of cholangiocarcinoma and hepatocellular adenoma of the liver, cystic keratinizing epithelioma of the lung, and gingival squamous cell carcinoma of the oral mucosa. The increased incidence of squamous cell carcinoma of the uterus was also considered to be related to TCDD administration. The marginally increased incidences of pancreatic acinar neoplasms and occurrences of hepatocholangioma and cholangioma of the liver may have been related to TCDD administration.

TCDD administration caused increased incidences of nonneoplastic lesions of the liver, lung, oral mucosa, pancreas, thymus, adrenal cortex, heart, clitoral gland, kidney, forestomach, mesentery, and thyroid gland in female rats.

<sup>\*</sup> Explanation of Levels of Evidence of Carcinogenic Activity is on page 11. A summary of the Technical Report Review Subcommittee comments and public discussion on this Technical Report appears on page 13.

# Summary of the 2-Year Carcinogenesis and Genetic Toxicology Studies of TCDD in Female Sprague-Dawley Rats

#### Doses in corn oil/acetone by gavage

0, 3, 10, 22, 46, or 100 ng/kg, and 100 ng/kg (stop-exposure)

#### **Body weights**

22, 46, and 100 ng/kg core study and 100 ng/kg stop-exposure groups less than the vehicle control group

#### Survival rates

25/53, 21/54, 23/53, 19/53, 22/53, 21/53, 21/50

#### Nonneoplastic effects

#### Liver:

hepatocyte hypertrophy (0/53, 19/54, 19/53, 42/53, 41/53, 52/53, 22/50); multinucleated hepatocyte (0/53, 0/54, 16/53, 26/53, 36/53, 51/53, 32/50); eosinophilic focus (11/53, 14/54, 21/53, 27/53, 27/53, 44/53, 27/50); inflammation (33/53, 46/54, 47/53, 50/53, 52/53, 49/53, 43/50); pigmentation (4/53, 9/54, 34/53, 48/53, 52/53, 53/53, 45/50); diffuse fatty change (0/53, 2/54, 12/53, 17/53, 30/53, 48/53, 10/50); necrosis (1/53, 4/54, 4/53, 8/53, 10/53, 17/53, 8/50); oval cell hyperplasia (0/53, 4/54, 3/53, 20/53, 38/53, 53/53, 1/50); bile duct hyperplasia (5/53, 4/54, 7/53, 22/53, 40/53, 53/53, 7/50); bile duct cyst (3/53, 1/54, 2/53, 2/53, 0/53, 21/53, 6/50); nodular hyperplasia (0/53, 0/54, 0/53, 3/53, 7/53, 36/53, 0/50); portal fibrosis (0/53, 0/54, 0/53, 0/53, 5/53, 27/53, 1/50); toxic hepatopathy (0/53, 2/54, 8/53, 30/53, 45/53, 53/53, 16/50); cholangiofibrosis (1/53, 1/54, 2/53, 1/53, 11/53, 31/53, 1/50) Lung: alveolar epithelium, metaplasia, bronchiolar (2/53, 19/54, 33/53, 35/52, 45/53, 46/52, 31/50) Oral Mucosa: gingival squamous hyperplasia (1/53, 7/54, 14/53, 13/53, 15/53, 16/53, 8/50) Pancreas: acinar cytoplasmic vacuolization (1/51, 0/54, 0/52, 1/53, 15/52, 42/51, 0/49); chronic active inflammation (0/51, 0/54, 2/52, 2/53, 3/52, 6/51, 4/49); acinar atrophy (1/51, 2/54, 4/52, 4/53, 4/52, 9/51, 4/49); arterial chronic active inflammation (0/51, 1/54, 1/52, 2/53, 2/52, 7/51, 2/49) Thymus: atrophy (36/51, 41/52, 44/52, 41/49, 44/46, 42/42, 45/49); severity of atrophy (2.6, 2.7, 3.0, 3,1, 3.6, 3.9, 3.3) Adrenal Cortex: atrophy (2/53, 0/54, 4/53, 5/53, 5/53, 27/53, 4/50); hyperplasia (16/53, 16/54, 18/53, 25/53, 29/53, 30/53, 20/50) Heart: cardiomyopathy (10/53, 12/54, 22/53, 25/52, 32/53, 36/52, 22/50) Clitoral Gland: cystic duct (34/50, 37/52, 41/53, 42/52, 41/51, 48/53, 35/49) Kidney: nephropathy (34/53, 26/54, 32/53, 36/53, 39/53, 52/53, 41/50); severity of nephropathy (1.2, 1.1, 1.3, 1.4, 1.4, 2.2, 1.4) Forestomach: squamous hyperplasia (3/53, 4/54, 4/53, 2/53, 7/53, 11/53, 5/50) Mesentery chronic active artery inflammation (0/53, 1/54, 0/53, 0/53, 4/53, 7/53, 1/50) Thyroid Gland: follicular cell hypertrophy (3/52, 4/54, 4/53, 7/51, 10/53, 17/52, 6/49)

# Summary of the 2-Year Carcinogenesis and Genetic Toxicology Studies of TCDD in Female Sprague-Dawley Rats

#### Neoplastic effects

Liver:				
hepatocellular adenoma (0/53, 0/54, 0/53, 0/53, 1/5	53, 13/53, 2/50);			
cholangiocarcinoma (0/53, 0/54, 0/53, 1/53, 4/53, 2	25/53, 2/50)			
Lung:				
cystic keratinizing epithelioma (0/53, 0/54, 0/53, 0/	/52, 0/53, 9/52, 0/50)			
Oral Mucosa:				
gingival squamous cell carcinoma (1/53, 2/54, 1/53	8, 0/53, 4/53, 10/53, 5/50)			
Uterus:				
squamous cell carcinoma (0/53, 0/54, 0/53, 0/53, 5	/53, 0/53, 2/50)			
1				
Equivocal findings				
Pancreas:				
acinar adenoma or carcinoma (0/51, 0/54, 0/52, 0/5	53, 0/52, 3/51, 1/49)			
Liver:				
hepatocholangioma (0/53, 0/54, 0/53, 0/53, 0/53, 2	/53, 0/50);			
cholangioma (0/53, 0/54, 0/53, 0/53, 0/53, 0/53, 1/2	50)			
	,			
Level of evidence of carcinogenic activity				
Clear evidence				
Genetic toxicology				
Salmonella typhimurium gene mutations:	Negative in strains TA98, TA100, TA1535, and TA1537, with and without S9			
Mouse lymphoma gene mutations:	Negative with and without S9			
Sister chromatid exchanges	-			
Cultured Chinese hamster ovary cells in vitro: Negative with and without S9				
Chromosomal aberrations				
Cultured Chinese hamster ovary cells in vitro:	Negative with and without S9			
Sex-linked recessive lethal mutations				
Drosophila melanogaster:	No induction of sex-linked recessive lethal mutations			
Chromosomal aberrations				

Negative

Mouse bone marrow *in vivo*:

#### EXPLANATION OF LEVELS OF EVIDENCE OF CARCINOGENIC ACTIVITY

The National Toxicology Program describes the results of individual experiments on a chemical agent and notes the strength of the evidence for conclusions regarding each study. Negative results, in which the study animals do not have a greater incidence of neoplasia than control animals, do not necessarily mean that a chemical is not a carcinogen, inasmuch as the experiments are conducted under a limited set of conditions. Positive results demonstrate that a chemical is carcinogenic for laboratory animals under the conditions of the study and indicate that exposure to the chemical has the potential for hazard to humans. Other organizations, such as the International Agency for Research on Cancer, assign a strength of evidence for conclusions based on an examination of all available evidence, including animal studies such as those conducted by the NTP, epidemiologic studies, and estimates of exposure. Thus, the actual determination of risk to humans from chemicals found to be carcinogenic in laboratory animals requires a wider analysis that extends beyond the purview of these studies.

Five categories of evidence of carcinogenic activity are used in the Technical Report series to summarize the strength of the evidence observed in each experiment: two categories for positive results (clear evidence and some evidence); one category for uncertain findings (equivocal evidence); one category for no observable effects (no evidence); and one category for experiments that cannot be evaluated because of major flaws (inadequate study). These categories of interpretative conclusions were first adopted in June 1983 and then revised in March 1986 for use in the Technical Report series to incorporate more specifically the concept of actual weight of evidence of carcinogenic activity. For each separate experiment (male rats, female rats, male mice, female mice), one of the following five categories is selected to describe the findings. These categories refer to the strength of the experimental evidence and not to potency or mechanism.

- Clear evidence of carcinogenic activity is demonstrated by studies that are interpreted as showing a dose-related (i) increase of malignant neoplasms, (ii) increase of a combination of malignant and benign neoplasms, or (iii) marked increase of benign neoplasms if there is an indication from this or other studies of the ability of such tumors to progress to malignancy.
- Some evidence of carcinogenic activity is demonstrated by studies that are interpreted as showing a chemical-related increased incidence of neoplasms (malignant, benign, or combined) in which the strength of the response is less than that required for clear evidence.
- Equivocal evidence of carcinogenic activity is demonstrated by studies that are interpreted as showing a marginal increase of neoplasms that may be chemical related.
- No evidence of carcinogenic activity is demonstrated by studies that are interpreted as showing no chemical-related increases in malignant or benign neoplasms.
- **Inadequate study** of carcinogenic activity is demonstrated by studies that, because of major qualitative or quantitative limitations, cannot be interpreted as valid for showing either the presence or absence of carcinogenic activity.

For studies showing multiple chemical-related neoplastic effects that if considered individually would be assigned to different levels of evidence categories, the following convention has been adopted to convey completely the study results. In a study with clear evidence of carcinogenic activity at some tissue sites, other responses that alone might be deemed some evidence are indicated as "were also related" to chemical exposure. In studies with clear or some evidence of carcinogenic activity, other responses that alone might be termed equivocal evidence are indicated as "may have been" related to chemical exposure.

When a conclusion statement for a particular experiment is selected, consideration must be given to key factors that would extend the actual boundary of an individual category of evidence. Such consideration should allow for incorporation of scientific experience and current understanding of long-term carcinogenesis studies in laboratory animals, especially for those evaluations that may be on the borderline between two adjacent levels. These considerations should include:

- adequacy of the experimental design and conduct;
- · occurrence of common versus uncommon neoplasia;
- progression (or lack thereof) from benign to malignant neoplasia as well as from preneoplastic to neoplastic lesions;
- some benign neoplasms have the capacity to regress but others (of the same morphologic type) progress. At present, it is impossible to
  identify the difference. Therefore, where progression is known to be a possibility, the most prudent course is to assume that benign
  neoplasms of those types have the potential to become malignant;
- · combining benign and malignant tumor incidence known or thought to represent stages of progression in the same organ or tissue;
- · latency in tumor induction;
- multiplicity in site-specific neoplasia;
- · metastases;
- supporting information from proliferative lesions (hyperplasia) in the same site of neoplasia or in other experiments (same lesion in another sex or species);
- presence or absence of dose relationships;
- · statistical significance of the observed tumor increase;
- · concurrent control tumor incidence as well as the historical control rate and variability for a specific neoplasm;
- · survival-adjusted analyses and false positive or false negative concerns;
- · structure-activity correlations; and
- in some cases, genetic toxicology.

### NATIONAL TOXICOLOGY PROGRAM BOARD OF SCIENTIFIC COUNSELORS TECHNICAL REPORTS REVIEW SUBCOMMITTEE

The members of the Technical Reports Review Subcommittee who evaluated the draft NTP Technical Report on TCDD on February 17, 2004, are listed below. Subcommittee members serve as independent scientists, not as representatives of any institution, company, or governmental agency. In this capacity, subcommittee members have five major responsibilities in reviewing the NTP studies:

- · to ascertain that all relevant literature data have been adequately cited and interpreted,
- to determine if the design and conditions of the NTP studies were appropriate,
- · to ensure that the Technical Report presents the experimental results and conclusions fully and clearly,
- · to judge the significance of the experimental results by scientific criteria, and
- to assess the evaluation of the evidence of carcinogenic activity and other observed toxic responses.

Mary Anna Thrall, D.V.M., Chairperson Department of Pathology Colorado State University Fort Collins, CO

Larry S. Andrews, Ph.D. Toxicology Department Rohm and Haas Company Spring House, PA

Diane F. Birt, Ph.D. Department of Food Science & Human Nutrition Iowa State University Ames, IA

Kim Boekelheide, M.D., Ph.D., Principal Reviewer Division of Biology and Medicine Department of Pathology and Laboratory Medicine Brown University Providence, RI

Michael R. Elwell, D.V.M., Ph.D., Principal Reviewer Pathology, Drug Safety Evaluation Pfizer Global Research and Development Groton, CT

Thomas A. Gasiewicz, Ph.D., Principal Reviewer Department of Environmental Medicine Environmental Health Sciences Center University of Rochester School of Medicine Rochester, NY

Shuk-Mei Ho, Ph.D. Department of Surgery, Division of Urology University of Massachusetts Medical School Worcester, MA James E. Klaunig, Ph.D. Indiana University School of Medicine Indianapolis, IN

Charlene A. McQueen, Ph.D. Department of Pharmacology and Toxicology College of Pharmacy University of Arizona Tucson, AZ

Walter W. Piegorsch, Ph.D. Director of Undergraduate Studies Department of Statistics University of South Carolina Columbia, SC

Stephen M. Roberts, Ph.D. Center for Environmental & Human Toxicology University of Florida Gainesville, FL

Richard D. Storer, M.P.H., Ph.D. Department of Genetic and Cellular Toxicology Merck Research Laboratories West Point. PA

Mary Vore, Ph.D. Graduate Center for Toxicology University of Kentucky Lexington, KY

#### SUMMARY OF TECHNICAL REPORTS REVIEW SUBCOMMITTEE COMMENTS

On February 17, 2004, the draft Technical Report on the toxicology and carcinogenesis studies of 2,3,7,8-tetrachlorobenzo-*p*-dioxin (TCDD) received public review by the National Toxicology Program's Board of Scientific Counselors' Technical Reports Review Subcommittee. The review meeting was held at the National Institute of Environmental Health Sciences, Research Triangle Park, NC.

Dr. N.J. Walker, NIEHS, introduced the toxicology and carcinogenesis studies of TCDD by noting that the chemical is the benchmark base of reference for the toxic equivalency factor (TEF) methodology. He described the study design and the spectrum of hormonal and histopathologic alterations seen in the liver, lung, oral mucosa, and pancreas as well as nonneoplastic lesions in a variety of other tissues. The proposed conclusion was *clear evidence of carcinogenic activity* of TCDD in female Harlan Sprague-Dawley rats.

Dr. Elwell, the first principal reviewer, said the study was well designed and included a number of useful mechanistic studies. He suggested the inclusion of inflammation of the mesenteric artery as another nonneoplastic effect in the conclusions.

Dr. Gasiewicz, the second principal reviewer, suggested that references to increases or decreases in lesion incidences implied statistical significance, and differences that were not statistically significant should be so specified. He inquired about variations in control values for thyroid hormones and the BrdU labeling index at different time points and whether more quantitative criteria could be assigned for severity grades for nonneoplastic lesions. He also questioned inclusion of squamous cell carcinoma of the uterus in one dose group as treatment related in the conclusions. Dr. Boekelheide, the third principal reviewer, also suggested attempts to quantify the diagnostic criteria for histopathologic diagnoses.

Dr. Walker explained that the measurements for clinical chemistry parameters were done sequentially at the time of analysis with emphasis on comparison between dose groups, so seeming differences between different time points might be artifactual. Differences in water consumption could also have been a factor.

Dr. Walker said that carcinomas of the uterus were also observed in the stop-exposure study, which lent support to their being chemical related. Dr. Elwell added that the occurrence of five such tumors in one dose group seemed sufficiently significant, particularly since another database of industry studies reported only two such tumors in 900 historical control female Sprague-Dawley rats. Dr. A. Nyska, NIEHS, said that fuller diagnostic criteria for nonneoplastic lesions would be included in the final document.

Dr. W.T. Allaben, NCTR, inquired about the choice of corn oil as the gavage vehicle. Dr. Walker replied that corn oil was used for more ready comparison with other studies in the literature, which used the same route.

Dr. Thrall suggested including bone marrow smears along with histopathology routinely in studies of chemicals associated with lymphoproliferative or myeloproliferative.

Dr. Elwell moved that the conclusions be accepted as written, with the inclusion of inflammation of the mesenteric artery to the conclusions and mention of ovarian atrophy in the text of the Abstract. Dr. Boekelheide seconded the motion, which was approved unanimously with 12 votes.

# **OVERVIEW**

# **DIOXIN TOXIC EQUIVALENCY FACTOR**

#### **EVALUATION**

## Polyhalogenated Aromatic Hydrocarbons and Human Exposure

Polyhalogenated aromatic hydrocarbons (PHAHs) comprise a large class of compounds including polychlorinated dibenzodioxins (PCDDs), polychlorinated dibenzofurans (PCDFs), polychlorinated biphenyls (PCBs), polychlorinated naphthalenes (PCNs), and polybrominated diphenyl ethers (PBDEs).

PCDDs and PCDFs were not manufactured for commercial purposes. They are unwanted by-products of many anthropogenic activities, including combustion processes, such as forest and backyard trash fires, and manufacturing processes for herbicides and paper. PCB mixtures were commercially produced and used in the electric power industry as dielectric insulating fluids in transformers and capacitors and were used in hydraulic fluids, plastics, and paints. PCNs were produced and used as dielectric fluids in capacitors, transformers, and cables. PBDEs are flame retardants, used in the manufacture of items including paints, foams, textiles, furniture, and household plastics (USEPA, 2000a).

Because these compounds are resistant to degradation and persistent in the environment, they have the ability to bioaccumulate and become more concentrated. Ambient human exposure to PHAHs occurs through the ingestion of foods containing PHAH residues. Due to their persistence and lipophilicity, once internalized, they accumulate in adipose tissue, resulting in chronic lifetime human exposure (Schecter *et al.*, 1994a).

#### **Dioxin-like** Compounds

Depending on the location and type of the halogenation, some PHAHs, most notably certain PCDDs, PCDFs, and PCBs, have the ability to bind to a cytosolic receptor known as the aryl hydrocarbon receptor (AhR) (Safe, 1990; Whitlock, 1990). 2,3,7,8-Tetrachlorodibenzo-*p*dioxin (TCDD), commonly referred to as "dioxin," is the most well characterized member of these structurally related compounds and exhibits the highest potency of binding to the AhR. Depending upon the number and

position of the substitutions, there are potentially 75 PCDDs, 135 PCDFs, and 209 PCBs. Structurally related compounds that bind to the AhR and exhibit biological actions similar to TCDD are commonly referred to as dioxin-like compounds (DLCs). There are seven PCDDs, ten PCDFs, and thirteen PCBs that exhibit such dioxin-like activity (USEPA, 2000b). In addition to the persistent DLCs, there is a wide variety of other compounds that can also bind to the AhR, including polycyclic aromatic hydrocarbons, (e.g., benzo(a)pyrene found in cigarette smoke), dietary indoles (e.g., indole-3-carbinol found in cruciferous vegetables), dietary flavonoids (e.g., quercetin, kaempferol), and heme degradation products (e.g., bilirubin/biliverdin).

The persistent PHAHs and DLCs have been the subject of an extensive amount of research regarding environmental levels, transport, and fate; human exposure; mechanisms of action; and toxicity that is beyond the scope of this report. The extensive body of knowledge on TCDD and related compounds has been fully reviewed by the International Agency for Research on Cancer (1997), the Agency for Toxic Substances and Disease Registry (1998, 2000), and the United States Environmental Protection Agency (2000a,b,c); therefore, it will not be re-reviewed in depth in this Technical Report.

# Mechanism of Action via the Aryl Hydrocarbon Receptor

Based on the extensive body of research on the induction of the cytochrome P450 1A1 (CYP1A1) gene by TCDD, the primary mechanism of action of DLCs involves initial binding to the AhR (Schmidt and Bradfield, 1996). The AhR is a protein found as a multimeric complex in the cytosol of all vertebrate species and acts as a ligandactivated transcription factor. Initial binding of ligand to the receptor disrupts the receptor complex leading to receptor activation and translocation into the nucleus where it heterodimerizes with the AhR nuclear translocator protein (ARNT) (Gu *et al.*, 2000). The AhR-ARNT heterodimer binds to specific cognate DNA sequence elements known as dioxin/xenobiotic response elements (DRE/XRE) present in the regulatory region of specific genes such as CYP1A1. Binding of the AhR-ARNT heterodimer to these elements leads to increased transcription of the specific gene. The characteristic response to TCDD is the transcriptional induction of CYP1A1, which is mediated by binding of the heterodimer to DREs present in the 5' flanking region of the gene. The AhR is expressed in all tissues with a definite tissue specificity in terms of level of expression and diversity of response. TCDD has been shown to modulate numerous growth factor, cytokine, hormone, and metabolic pathways in animals and experimental systems. Many, if not all, are parts of pathways involved in cellular proliferation and differentiation; taken together, they provide a plausible mechanism for toxicity and carcinogenicity. Most of the molecular details for induction of gene expression via the AhR have been characterized for the transcriptional activation of the CYP1A1 gene. The expressions of many genes have been shown to be affected by TCDD (Puga et al., 2000; Frueh et al., 2001; Martinez et al., 2002), yet there is evidence for direct transcriptional activation through the AhR for only a very few of these (Sutter and Greenlee, 1992).

#### Toxicity of Dioxin-like Compounds

High doses of and/or continuous exposure to dioxins leads to a broad spectrum of toxic responses including death, immunosuppression, carcinogenicity, and impaired reproduction and development (Whitlock, 1990; ATSDR, 1998; Grassman *et al.*, 1998; USEPA, 2000c). The type of toxicity is dependent on the magnitude of dose, duration and pattern of exposure, timing of exposure, species, and gender. A generalized mode of action for toxicity induced by dioxins is one that involves initial binding of the compounds to the AhR. Subsequent alterations in expression of specific genes and alterations in biological signal transduction pathways lead to an alteration in growth regulation and differentiation that leads to pathology and toxicity.

The broad spectrum of DLC effects on hormone and growth factor systems, cytokines, and signal transduction pathways indicates powerful growth dysregulators. The effect of DLCs on growth regulation may be manifested through alterations in genes involved in cellular growth and homeostasis. Although the relationship between these effects and carcinogenesis can only be inferred, all of these effects are involved in cellular growth and differentiation; disruption of normal cellular processes could be a risk factor for carcinogenicity. The initial involvement of the AhR in initiating this cascade of events is supported by studies showing the lower potency of structurally related compounds with lower affinity for the AhR, reduction of effects in rodents with lower AhR affinities (Pohjanvirta *et al.*, 1993; Birnbaum, 1994a), and the lack of effects using transgenic mice that lack AhR functionality (Gonzalez *et al.*, 1996; Gonzalez and Fernandez-Salguero, 1998; Gonzalez, 2001; Vorderstrasse *et al.*, 2001). These data indicate that the AhR is necessary, but may not be sufficient, for mediating the toxic action of DLCs.

# Polyhalogenated Aromatic Hydrocarbon Mixtures and Toxic Equivalency Factors

PHAHs always exist in the environment as complex mixtures; therefore, normal background human exposure to PHAHs always occurs as a complex mixture. The toxic equivalency factor (TEF) approach has been developed to assess risks posed by complex mixtures of PCDDs, PCDFs, and PCBs (Ahlborg et al., 1992; Van den Berg et al., 1998; USEPA, 2000c). The TEF methodology is a relative potency scheme to estimate the total exposure and dioxin-like effects of a mixture of chemicals based on a common mechanism of action involving an initial binding of the compound to the AhR. The TEF methodology is currently the most feasible interim approach for assessing and managing the risk posed by these mixtures and has been formally adopted by a number of countries including Canada, Germany, Italy, the Netherlands, Sweden, the United Kingdom, and the United States. The method is also used by the International Programme on Chemical Safety and the World Health Organization (WHO). Criteria for inclusion of a compound in the TEF methodology are structural relationship to PCDD/PCDFs, binding to the AhR, elicitation of AhR-mediated biochemical and toxic responses, and persistence and accumulation in the food chain.

The current WHO TEFs are based on a subjective evaluation of individual studies that examined the relative potency of a given chemical to the reference compound, TCDD, which is assigned a potency of 1. TEF values are an order of magnitude *estimate* of the overall "toxic potency" of a given compound and therefore do not specifically refer to the potency from any single study with a particular endpoint. By comparison, a relative potency factor is determined for a specific chemical in a single study relative to a specific endpoint. Therefore, a single TEF is based on an evaluation of multiple relative potency factors. The TEF determination is a subjective assessment because the relative potency factors are derived from the literature and there is considerable variability in the types of studies, endpoints analyzed, and quality of procedures. Types of procedures for calculation of relative potency factors vary from a comparative dose response assessment (e.g., ratio of  $ED_{50}$  or  $EC_{50}$ ) to a simple administered dose ratio calculation. In evaluating different studies and endpoints, *in vivo* studies are weighted more than *in vitro* studies, chronic studies are weighted more than simple biochemical responses.

An implicit assumption of the TEF methodology is that the combined effects of the different congeners are dose additive, which is supported by *in vivo* studies with mixtures of PCDDs and PCDFs, mixtures of PCDFs, and mixtures of PCBs and TCDD and by *in vitro* studies with mixtures of PCBs and PCDFs (Birnbaum *et al.*, 1987; Schrenk *et al.*, 1991, 1994; Birnbaum and DeVito, 1995; USEPA, 2000c). Therefore, the total toxic equivalents (TEQs) for the AhR-mediated toxic potency of a mixture of PCDDs, PCDFs, and PCBs may be estimated by the summation of the mass of each congener in the mixture after adjustment for its potency. Currently, only PCDDs, PCDFs, and certain PCBs are included in this TEF scheme.

$$\begin{split} \text{TEQ} &= \sum_{ni} \left( \text{PCDD}_i \times \text{TEF}_i \right)_n + \sum_{ni} \left( \text{PCDF}_i \times \text{TEF}_i \right)_n \\ &+ \sum_{ni} (\text{PCB}_i \times \text{TEF}_i)_n \end{split}$$

where i = the individual congener and its respective TEF, and n = all congeners within each class of DLCs

# Uncertainties in the Use of Toxic Equivalency Factors

While TEFs were developed initially as an interim approach to facilitate exposure assessment and hazard identification, there has been an increasing use of this scheme to determine TEQs in human tissues for dose-response assessment of effects in human populations (Flesch-Janys *et al.*, 1998). While the database for development of TEFs for DLCs is extensive, these data are for dioxin-regulated noncancer endpoints that often reflect simply the activation of the AhR. No mammalian studies have formally evaluated relative potency factors for a neoplastic endpoint. The mechanism by which activation of the AhR and subsequent changes in dioxinOne of the implicit assumptions in the use of TEFs is that the TEQ for different compounds is dose additive. While dose additivity is supported for certain mixtures, this may not be true for some biological endpoints in some models. As outlined by Van den Berg *et al.* (1998), the TEF methodology is likely valid for biological responses that are clearly AhR dependent but may not be true for more complex biological responses such as neoplasia.

## The Dioxin Toxic Equivalency Factor Evaluation Studies

To test the validity of the TEF approach for the prediction of cancer risk, the NTP has conducted multiple 2year bioassays in female Sprague-Dawley rats to evaluate the chronic toxicity and carcinogenicity of DLCs, structurally related PCBs, and mixtures of these compounds. Specific hypotheses to be tested by these studies are:

- 1. TEFs for PCDDs, PCDFs, and PCBs can predict the relative carcinogenic potency of single congeners in female Sprague-Dawley rats.
- 2. TEFs for PCDDs, PCDFs, and planar PCBs can predict the relative carcinogenic potency of an environmentally relevant mixture of these chemicals in the female Sprague-Dawley rat.
- 3. The carcinogenicity of a dioxin-like, non-orthosubstituted PCB is not altered by the presence of a mono-ortho- or di-ortho-substituted PCB.
- 4. Relative potencies for DLCs are dose additive.
- 5. The relative potencies for activation of biochemical endpoints, such as CYP1A1 induction, in the 2-year studies are equivalent to the relative potency for induction of carcinogenesis when estimated based on administered dose.
- 6. The relative potencies for activation of biochemical endpoints, such as CYP1A1 induction, in the 2-year studies are equivalent to the relative potency for induction of carcinogenesis when estimated based on target tissue dose.
- 7. The relative potencies for alteration of a given response are the same, regardless of the dose metric used (e.g., administered dose, serum or whole blood concentrations, or tissue dose).

# Individual Compounds, Mixtures, and Rationale for Choice



2,3,7,8-Tetrachlorodibenzo-p-dioxin
TCDD

CAS No. 1746-01-6

Chemical Formula:	$C_{12}H_4Cl_4O_2$
Molecular weight:	321.97

TCDD is the most potent DLC and the reference compound to which all DLCs are compared in the TEF methodology. As such, it has a TEF value of 1.0. TCDD is classified as a known human carcinogen by the NTP and the International Agency for Research on Cancer.



2,3,4,7,8-Pentachlorodibenzofuran PeCDF

CAS No. 57117-31-4

Chemical Formula:	$C_{12}H_3Cl_5O$
Molecular weight:	340.42

PeCDF is a dioxin-like PHAH with high bioaccumulation in the food chain and a TEF value of 0.5. This compound represents the most potent PCDF present in human tissues.



CAS No. 57465-28-8

Chemical Formula:	$C_{12}H_5Cl_5$
Molecular weight:	326.42

PCB 126 is a non-ortho-substituted PCB with high bioaccumulation in the food chain and a TEF value of 0.1. PCB 126 is considered the most potent dioxin-like PCB congener present in the environment and accounts for 40% to 90% of the total toxic potency of PCBs having a "dioxin-like" activity.



2,3',4,4',5-Pentachlorobiphenyl PCB 118

CAS No. 31508-00-6

Chemical Formula:	$C_{12}H_5Cl_5$
Molecular weight:	326.43

PCB 118 is a mono-ortho-substituted PCB that has partial dioxin-like activity. A tentative TEF value of 0.0001 has been assigned although there is controversy over whether mono-ortho-substituted PCBs should be included in the TEF methodology.



2,2',4,4',5,5'-Hexachlorobiphenyl PCB 153

CAS No. 35065-27-1

Chemical Formula:	$C_{12}H_4Cl_6$
Molecular weight:	360.88

PCB 153 is a di-ortho-substituted nonplanar PCB and is present at the highest concentrations in human samples on a molar basis. Nonplanar PCBs do not have dioxinlike activity and are not included in the TEF methodology; therefore, PCB 153 has no TEF value. Some studies have shown that nondioxin PCBs such as PCB 153 can antagonize the effects of DLCs.

#### **Mixture Studies**

Several mixture studies were conducted to assess the dose additivity of DLCs and interactions of PCBs.

#### Mixture of TCDD, PCB 126, and PeCDF

This mixture was designed to test for dose-additivity of the highest potency DLCs in each of the three classes of PHAHs covered by the TEF methodology. The mixture was composed of equal TEQ ratios (1:1:1) of TCDD, PCB 126, and PeCDF. Total TEQ dosages ranged from 10 to 100 ng TEQ/kg per day. These compounds were chosen because they are the most potent members of the PCDDs, PCDFs, and coplanar PCBs. Based on average human tissue levels of these compounds, they represent approximately 48% of the human tissue burden of dioxin TEQs.

#### Binary mixture study of PCB 126 and PCB 153

Several studies have indicated an antagonism of the effects of DLCs by di-ortho-substituted PCBs such as PCB 153. This binary mixture study consisted of two parts:

- 1. PCB 126 and PCB 153 at the environmentally relevant ratio of 1:1,000. The dosage levels of PCB 126 were chosen to span the range used in the individual dose-response study of PCB 126.
- Varying ratios of PCB 153 at the mid-dose of PCB 126 (300 ng/kg per day).

#### Binary mixture of PCB 118 and PCB 126

This binary mixture was not designed a priori as part of the dioxin TEF evaluation. While the individual PCB 118 study was at the in-life phase, it was found that the PCB 118 compound being used contained not only PCB 118 but also 0.622% PCB 126 (PCB 118:PCB 126 of 161:1). Given the large TEF difference between PCB 118 (0.0001) and PCB 126 (0.1), this resulted in a TEQ ratio for PCB 126:PCB 118 of 6:1. As such, the effects of the compound would be expected to be due mainly to dioxin-like effects of PCB 126 rather than effects of PCB 118. In human tissues, the ratio of PCB 126:PCB 118, on a TEQ basis, ranges from 0.9:1 in blood, 3.9:1 in breast milk, and 15:1 in adipose tissue (USEPA, 2000b). The mass ratio of PCB 118:PCB 126 is on average 135:1 in beef fat and 190:1 in milk. Consequently, the PCB 118:PCB 126 ratio in this compound (161:1) represented an environmentally relevant mixture of PCBs on both a mass and TEQ basis. Since PCB 126 was already being studied and the PCB 118 study was already in life, the PCB 118 study was continued to test for the effect of a mono-ortho-substituted PCB on a coplanar PCB at an environmentally relevant ratio. The PCB 118 was resynthesized and checked for the absence of high TEQ contributing compounds, and a new study was started.

# STUDY DESIGN, SPECIES, AND DOSE SELECTION RATIONALE

These studies were conducted in female Harlan Sprague-Dawley rats based on the prior observations by Kociba *et al.* (1978) of the carcinogenicity of TCDD in Spartan Sprague-Dawley rats. Female rats were chosen based on the high potency of hepatocarcinogenicity in females in this strain. Male rats were not studied due to the lack of induction of liver and lung neoplasms in the previous studies of Sprague-Dawley rats with TCDD. Animals were dosed by oral gavage because the majority of human exposure is oral.

Dose selection for TCDD of 3 to 100 ng/kg per day was based on the range used in the Kociba *et al.* (1978) study and on the demonstrated induction of liver tumor incidence over this dose range. Dosage levels for other compounds were based on the TCDD dosage range after adjustment for the current TEF values or relative potency values (Table 1). These studies were designed to examine dose additivity rather than response additivity, and dose spacing was weighted in the 10 to 100 ng/kg range to increase dose density in the region where an increase in liver tumors was expected. Doses higher than 100 ng/kg were not used in order to limit the known effects on body weight and liver toxicity seen with TCDD at this dose level. Prior studies of TCDD suggest that this dose is at or near the predicted maximum tolerated dose.

Interim necropsies at 14, 31, and 53 weeks were incorporated into the studies for the examination of mechanistically based biomarkers of AhR- or PCB-mediated effects. These endpoints included alterations in cytochromes P450 1A1, 1A2 and 2B, thyroid hormone levels, and hepatocyte replication. Tissue analyses of the

parent compound in the liver, lung, blood, and adipose were included at each interim necropsy and at terminal necropsy for dose response analysis using administered dose, total body burden, and target tissue dose as the dose metric.

Additional "special study" animals were included at each interim necropsy. Tissues from these animals were provided to specific extramural grantees to facilitate the conduct of additional mechanistic studies. These animals were not evaluated as part of the core study.

 TABLE 1

 Compounds and Associated Doses Used in the Dioxin TEF Evaluation Studies

Compound	TEF <sup>a</sup>	Core Study	Stop-Exposure Study
TCDD	1	3, 10, 22, 46, 100 ng/kg	100 ng/kg
PCB 126	0.1	10 <sup>b</sup> , 30, 100, 175, 300, 550, 1,000 ng/kg	1,000 ng/kg
PeCDF	0.5	6, 20, 44, 92, 200 ng/kg	200 ng/kg
TEF Mixture <sup>c</sup>		10 ng TEQ/kg (3.3 ng/kg TCDD, 6.6 ng/kg PeCDF, 33.3 ng/kg PCB 126) 22 ng TEQ/kg (7.3 ng/kg TCDD, 14.5 ng/kg PeCDF, 73.3 ng/kg PCB 126) 46 ng TEQ/kg (15.2 ng/kg TCDD, 30.4 ng/kg PeCDF, 153 ng/kg PCB 126) 100 ng TEQ/kg (33 ng/kg TCDD, 66 ng/kg PeCDF, 333 ng/kg PCB 126)	None
PCB 153	None	10, 100, 300, 1,000, 3,000 µg/kg	3,000 µg/kg
PCB 126/PCB 153 <sup>d</sup>		10/10, 100/100, 300/100, 300/300, 300/3,000, 1,000/1,000	None
PCB 126/PCB 118 <sup>e</sup>		7 ng TEQ/kg (62 ng/kg PCB 126, 10 μg/kg PCB 118) 22 ng TEQ/kg (187 ng/kg PCB 126, 30 μg/kg PCB 118) 72 ng TEQ/kg (622 ng/kg PCB 126, 100 μg/kg PCB 118) 216 ng TEQ/kg (1,866 ng/kg PCB 126, 300 μg/kg PCB 118) 360 ng TEQ/kg (3,110 ng/kg PCB 126, 500 μg/kg PCB 118)	360 ng TEQ/kg
PCB 118	0.0001	10 <sup>b</sup> , 30 <sup>b</sup> , 100, 220, 460, 1,000, 4,600 µg/kg	4,600 µg/kg

<sup>a</sup> Van den Berg *et al.* (1998)

<sup>b</sup> 14-, 31-, and 53-week scheduled sacrifices only

<sup>c</sup> 10, 22, 46, 100 ng TEQ/kg (TCDD:PeCDF:PCB 126, 1:2:10)

PCB 126 dose units are ng/kg, PCB 153 units are  $\mu$ g/kg.

PCB 126 dose units are ng/kg, PCB 118 units are μg/kg. Doses are based on PCB 126 levels that are 0.622% of the administered PCB 118 bulk.

# **INTRODUCTION**



#### 2,3,7,8-Tetrachlorodibenzo-*p*-dioxin TCDD

CAS No. 1746-01-6

Chemical Formula: C<sub>12</sub>H<sub>4</sub>Cl<sub>4</sub>O<sub>2</sub> Molecular Weight: 321.98

Synonyms: Dioxin; dioxine; TCDBD; 2,3,7,8-TCDD; 2,3,7,8-tetrachlorodibenzo(b,e)(1,4)dioxin; 2,3,7,8-tetrachlorodibenzo-*p*-dioxin; 2,3,6,7-tetrachlorodibenzo-*p*-dioxin; 2,3,7,8-tetrachlorodibenzo-1,4-dioxin; tetrachlorodibenzodioxin; 2,3,6,7-tetrachlorodibenzoidioxin; tetradioxin

## **CHEMICAL AND PHYSICAL PROPERTIES**

TCDD belongs to a family of chemicals designated polyhalogenated aromatic hydrocarbons (PHAHs). These include the polychlorinated dibenzo-*p*-dioxins (PCDDs), polychlorinated dibenzofurans (PCDFs), and polychlorinated biphenyls (PCBs). Depending on the position and number of chlorine substitutions, the structure of these chemicals allows for 75 chlorinated dioxins, 135 chlorinated dibenzofurans, and 209 chlorinated biphenyls (USEPA, 2000b).

TCDD is a colorless to white crystalline powder with a melting point of 305° C. It is insoluble in water, slightly soluble in n-octanol, methanol, and lard oil, and soluble in organic solvents including *o*-dichlorobenzene, chlorobenzene, benzene, and chloroform (IARC, 1997; Lewis, 1997; ATSDR, 1998). TCDD is stable in DMSO, 95% ethanol, or acetone. TCDD can undergo a slow photochemical and bacterial degradation, but it is normally extremely stable. Although TCDD is nonflammable, it degrades when heated in excess of 500° C (Sittig, 1985; Lewis, 1988).

# PRODUCTION, USE, AND HUMAN EXPOSURE

TCDD is not manufactured commercially other than for The main sources of scientific research purposes. TCDD releases into the environment are from combustion and incineration sources, metal smelting, refining, and processing, chemical manufacturing and processing, biological and photochemical processes, and existing reservoir sources that reflect past releases (USEPA, 2000a). Chemical reactions that result in the formation of TCDD are used in the production of phenoxy herbicides (e.g., 2,4,5-trichlorophenoxyacetic acid), hexachlorophene, chlorophenols, chlorodiphenyl ether herbicides [e.g., 1,3,4-trichloro-2-(4-nitrophenoxy)benzene], hexachlorobenzene, pulp bleaching, and the manufacture of dyes and pigments. TCDD is formed as an unwanted combustion by-product during the incineration of municipal waste, sewage sludge, hospital waste, polyvinyl chloride, and wood and from automobile emissions. Photochemical dechlorination of dioxins has been shown to produce TCDD. Biochemical reactions that occur in sewage sludge can also result in the formation of dioxins.

Due to high lipophilicity and low solubility of TCDD in aqueous media, TCDD accumulates in the fat tissues of animals. The highest concentrations of TCDD are found in fish, meat, eggs, and dairy products (Schecter *et al.*, 1994b; USEPA, 2000b). This results in widespread exposure of the general population to TCDD and related DLCs (Schecter *et al.*, 1994a). It is estimated that 90% of human exposure to TCDD and related dioxins occurs through ingestion of food contaminated with these compounds. A small fraction of exposure occurs via inhalation or dermal absorption.

Adult daily intake of DLCs, including PCDDs, PCDFs, and dioxin-like PCBs, from all sources is estimated to be approximately 70 pg toxic equivalents (TEQ)/day, where TEQ reflects the potency adjusted mass of all DLCs covered by the World Health Organization TEFs. The intake from all sources of PCDDs and PCDFs is estimated at 45 pg TEQ/day, and intake from dioxin-like PCBs is 25 pg TEQ/day. Approximately 90% of the daily intake is from food sources (40 pg TEQ/day for PCDDs and PCDFs and 22 pg TEQ/day for dioxin-like PCBs). Intake of TCDD, PeCDF, and PCB 126 from food is estimated to be 5 pg TEQ/day, 6.6 pg TEQ/day, and 13 pg TEQ/day, respectively. TCDD, PeCDF, and PCB 126 combined (24.6 pg TEQ/day) represent 40% of the estimated daily intake of DLCs from food sources. TCDD alone represents 8% of the estimated daily intake of DLCs.

This level of exposure, together with the long half-life of DLCs in humans, leads to persistent levels of DLCs in human tissues (USEPA, 2000b). Depending on dietary practices and proximity to specific sources of exposure, some populations may have additional exposure/body burdens above this level. Exposure levels in the United States are similar to those seen in other industrialized countries. In contrast to the general population, several specific populations have been exposed to much higher levels of PeCDF as a result of occupational exposure.

The average level of PCDDs, PCDFs, and dioxin-like PCBs, combined, on a TEQ basis, in human adipose tissue is 68 pg TEQ/g lipid (USEPA, 2000b). The average levels of TCDD, PeCDF, and PCB 126 in human adipose tissues are 5.5 pg TEQ/g, 5.0 pg TEQ/g, and 22.4 pg TEQ/g, respectively. Consequently, TCDD, PeCDF, and PCB 126 combined (32.9 pg TEQ/g lipid) represent 48% of the total level of DLCs present in human adipose tissue. TCDD alone represents 8% of the DLCs present in human adipose tissue.

## **TOXICOKINETICS**

There is an extensive body of literature examining the toxicokinetics of TCDD and related compounds (USEPA, 2000c); for the purpose of brevity, only pertinent information is provided here. Several studies have examined absorption of TCDD from the gastrointestinal tract (Piper et al., 1973; Rose et al., 1976). The absorption of TCDD from the gastrointestinal tract in Sprague-Dawley rats given a single dose of 1 µg TCDD/kg body weight in corn oil:acetone (25:1) is 84% (range 66% to 93%). Similar results have been observed after repeated exposure (0.1 to 1 µg/kg per day) and higher doses. Once absorbed, TCDD is transported primarily through the lymphatic system by chylomicrons and is readily distributed throughout the body. The main sites of distribution in rats within the first few days after exposure are to the liver and adipose tissue; lesser amounts are distributed to skin and the thyroid gland (Pohjanvirta et al., 1990). In blood, TCDD is associated mainly with lipoproteins, serum lipids, and a smaller fraction of albumin and cellular components. The pattern of distribution of TCDD in rats is due to its lipophilicity and binding to cytochrome P450 1A2 (Gillner et al., 1987; Diliberto et al., 1997). Cytochrome P450 1A2 is a known TCDDbinding protein that is also inducible by TCDD. Because CYP1A2 is inducible only in the liver and nasal passages (Goldstein and Linko, 1984), TCDD tends to sequester in the liver at levels that would not be predicted based on its lipophilicity alone. The hepatic sequestration by TCDD is not seen in CYP1A2 knockout mice, demonstrating the critical involvement of CYP1A2 in this process (Diliberto et al., 1999).

Estimates of the half-life of TCDD in rats depend on the organs examined and doses used. Whole-body elimination half-life in rats has been estimated as 12 to 31 days (USEPA, 2000c). In rat adipose tissue, TCDD exhibits first-order kinetics with an elimination half-life of 24.5 days. In Wistar rat liver, TCDD exhibits a biphasic elimination pattern with half-lives of 11.5 days and 16.9 days for the short- and long-term components. The elimination of TCDD from the liver of female Sprague-Dawley rats is also biphasic, and the half-lives decrease when the concentration of TCDD is greater than 1,000 ppt wet weight (Walker et al., 2000). For example, when the tissue concentration is below 1,000 ppt, the half-life is estimated to be 33.1 days, but it drops to just 16.7 days when the tissue concentration is above 1,000 ppt. Similarly, for animals that began TCDD exposure 18 weeks following diethylnitrosamine initiation, when the tissue concentration was below 1,000 ppt, the half-life was estimated to be 45.3 days, but it dropped to just 19.2 days [95% confidence interval (CI) 9.1 to 66.0 days] when the tissue concentration was above 1,000 ppt.

In the lung of female Sprague-Dawley rats, the half-life of TCDD is 39.7 days (95% CI 34.4 to 45.8 days) (Tritscher *et al.*, 2000). In serum, the half-life is estimated to be 44.6 days (95% CI 39.1 to 50.9 days). By comparison, it has been estimated that the whole body half-life of TCDD in humans is approximately 7 to 10 years.

#### **TCDD** Toxic Equivalency Factor

TCDD is the reference compound for the dioxin toxic equivalency factor (TEF) scheme and therefore has a TEF of 1.0 (Van den Berg *et al.*, 1998).

# Τοχιζιτγ

TCDD is one of the most studied environmental toxicants, and a vast body of literature is available on toxicities associated with TCDD exposure in multiple species (Poland and Knutson, 1982; Birnbaum, 1994b; ATSDR, 1998; Grassman *et al.*, 1998; USEPA, 2000c). Consequently, it is beyond the scope of this Technical Report to fully describe all the toxicities in all species that have been observed. Some of the toxic effects observed include developmental and reproductive alterations, immunotoxicity, teratogenicity, carcinogenicity, and lethality. Different animal species vary widely in sensitivity to the lethal toxicity of TCDD; the oral LD<sub>50</sub> of the chemical varies from 1  $\mu$ g/kg (guinea pigs) to 5,000  $\mu$ g/kg (hamsters). In rats, the LD<sub>50</sub> is 26  $\mu$ g/kg (male) or 22  $\mu$ g/kg (female), and in monkeys, the LD<sub>50</sub> is 70 μg/kg (Poland and Knutson, 1982). An acute lethal dose of TCDD has a latency period of 1 to 2 weeks in all species tested, during which time the animals exhibit a wasting syndrome. Characteristic effects associated with TCDD exposure are chloracne, reduced body weight and body weight gain, porphyria, thymic atrophy, gastric hyperplasia/hypoplasia, hepatotoxicity, increased serum concentrations of liver enzymes, hypertriglyceridemia, increased liver weight, hepatic Vitamin A depletion, altered thyroid homeostasis, and increased metabolizing enzymes (Poland and Knutson, 1982).

## CARCINOGENICITY

#### **Experimental** Animals

The carcinogenicity of TCDD has been clearly established in rodents by the dermal, dosed feed, and gavage routes of administration (Table 2) (Kociba et al., 1978; Toth et al., 1979; NTP, 1982a,b; Della Porta et al., 1987; Rao et al., 1988; IARC, 1997; USEPA, 2000c). TCDD administered by gavage induces tumors in male and female Osborne-Mendel rats and B6C3F, mice (NTP, In the NTP (1982a) study, significantly 1982a). increased incidences of thyroid gland follicular cell adenoma were observed in high dose male and female rats and high dose female mice. TCDD gavage administration also produced an increased incidence of neoplastic liver nodules in high dose female mice and an increased incidence of hepatocellular carcinoma in high dose male and female mice. TCDD administered to Swiss-Webster mice by dermal application caused an increased incidence of fibrosarcoma of the integumentary system in high dose females and yielded equivocal evidence in males (NTP, 1982b). Based on the NTP (1982b) studies, there is substantial evidence of carcinogenicity of TCDD in male and female rats and mice (Table 3).

Tissue Sites of Increased Neoplasm Incidences in Rodent Bioassays of TCDD

Species/Strain	Route of Administration	Sex	Tissue Site	Reference	
Rats/Sprague-Dawley	Dosed feed	Male	Tongue Nasal turbinates/hard palate	Kociba et al., 1978	
		Female	Lung Nasal turbinates/hard palate Liver		
Rats/Osborne-Mendel	Gavage	Male Female	Thyroid gland Liver	NTP, 1982a	
Mice/B6C3F <sub>1</sub>	Gavage	Male Female	Liver Liver Thyroid gland	NTP, 1982a	
Mice/Swiss Webster	Dermal	Female	Integumentary fibrosarcoma	NTP, 1982b	
Mice/B6C3 and B6C	Gavage	Male Female	Thymic lymphomas Liver	Della Porta et al., 1987	
Mice/Swiss	Gavage	Male	Liver	Toth et al., 1979	
Hamsters/Syrian Golden	Dermal	Male	Facial skin carcinoma	Rao et al., 1988	

## TABLE 3 Levels of Evidence for Carcinogenicity of TCDD in NTP Rodent Bioassays

Sex/Species	Route of Administration	
	Gavage <sup>a</sup>	Dermal <sup>b</sup>
Male/Rats <sup>c</sup>	Positive	Not Tested
Female/Rats <sup>c</sup>	Positive	Not Tested
Male/Mice <sup>d</sup>	Positive	Equivocal
Female/Mice <sup>u</sup>	Positive	Positive

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<sup>a</sup> NTP, 1982a b NTP, 1982b

c Osborne-Mendel rats d B6C3F<sub>1</sub> mice (gavage); Swiss-Webster mice (dermal)

One of the most highly cited carcinogenicity studies for TCDD is a 2-year study conducted by Dow Chemical Company (Kociba *et al.*, 1978). In that study, increased incidences of tumors were seen at multiple sites in Sprague-Dawley rats administered up to 100 ng TCDD/kg per day in feed for 2 years (Table 4). Increased incidences of hepatocellular hyperplastic nod-

ules (females), hepatocellular carcinoma (females), keratinizing squamous cell carcinoma of the lung (females), adenoma of the adrenal cortex (males), squamous cell carcinoma of the nasal turbinates/hard palate (males and females), and stratified squamous cell carcinoma of the tongue (males) were observed. Significantly decreased tumor incidences were observed for pheochromocytoma of the adrenal gland (males), subcutaneous skin lipoma, fibroma, or fibroadenoma (combined) (males), benign uterine tumors, benign neoplasms of the mammary gland, mammary gland carcinoma, pituitary gland adenoma (females), and acinar adenoma of the pancreas (males). Two evaluations of the pathology of the female liver tumor data confirmed significant increases in the incidences of hepatocellular adenoma and hepatocellular carcinoma (Squire, 1980; Goodman and Sauer, 1992; Table 5).

 TABLE 4

 Incidences of Tumors in Sprague-Dawley Rats in a 2-Year Feed Study of TCDD<sup>a</sup>

	Male			Female				
	0 ng/kg	1 ng/kg	10 ng/kg	100 ng/kg	0 ng/kg	1 ng/kg	10 ng/kg	100 ng/kg
- b	85	50	50	50	86	50	50	
Hepatocellular Hyperplastic Nodules	6	0	3	2	8	3	18*	23*
Hepatocellular Carcinoma	2	0	0	1	1	0	2	11*
Lung	85	50	50	50	86	50	50	49
Keratinizing Squamous Cell Carcinoma	a 0	0	0	1	0	0	0	7*
Nose, Hard Palate	85	50	50	50	86	50	50	49
Stratified Squamous Cell Carcinoma	0	0	0	4*	0	0	1	4*
Tongue	85	50	50	50	86	50	50	49
Stratified Squamous Cell Carcinoma	0	1	1	3*	1	0	0	2
Adrenal Gland	85	50	50	50	86	50	50	49
Adrenal Cortex Adenoma	0	0	2	5*	9	6	2	5
Adrenal Medulla, Pheochromocytoma	28	6	10	4*	7	2	1	3
Skin Subautaneous Linoma Fibroma	85	50	50	50	86	50	50	49
or Fibroadenoma	10	1*	5	6	1	1	0	0
Uterus					86	50	50	49
Benign Neoplasms					28	12	11	7*
Mammary Gland					86	50	50	49
Benign Neoplasms					73	35	36	24*
Carcinoma					8	4	4	0*
Pituitary Gland	85	50	50	50	86	50	50	49
Adenoma	26	6	11	13	43	18	13	12*
Pancreas	85	50	50	50	86	50	50	49
Acinar Adenoma	14	7	5	2*	0	1	0	1

\* Significantly different (P≤0.05) from the control group by the Fisher exact test

<sup>a</sup> Data presented by Kociba *et al.*, 1978

 $^{\circ}$  Number of animals with tissue examined microscopically (only sites where significant changes were observed are shown)

Number of animals with lesion

Table	5
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Incidences of Liver Tumors in Female Sprague-Dawley Rats Exposed to TCDD in Feed for 2 Years<sup>a</sup>

Evaluation	Tumor Classification		TCDD (ng/kg per day)			
		0 ng/kg	1 ng/kg	10 ng/kg	100 ng/kg	
Kociba <i>et al.</i> (1978) <sup>b</sup>	Hyperplastic Nodule	8/86▲▲	3/50	18/50**	23/49**	
	Hepatocellular Carcinoma Hyperplastic Nodule or	1/86▲▲	0/50	2/50	11/49**	
	Hepatocellular Carcinoma	9/86▲▲	3/50	18/50**	34/48**	
Squire (1980)	Neoplastic Nodule or Hepatocellular Carcinoma	16/86	8/50	27/50**	33/47**	
Goodman and Sauer (1992)	Hepatocellular Adenoma	2/86▲▲	1/50	9/50**	14/45**	
	Hepatocellular Carcinoma Hepatocellular Adenoma or	0/86▲▲	0/50	0/50	4/45**	
	Carcinoma	2/86▲▲	1/50	9/50**	18/45**	

\*\* Significantly different (P≤0.01) from the control group by the Fisher exact test

▲ Significant (P≤0.01) Mantel-Haenszel trend test (Huff *et al.*, 1991).

<sup>a</sup> Hyperplastic nodule, neoplastic nodule and hepatocellular adenoma are interchangeable descriptions for the same lesion.

<sup>b</sup> Incidence data for hyperplastic nodule or hepatocellular carcinoma (combined) in this study are as described by Huff *et al.*, 1991.

#### Humans

Cancer incidence has been evaluated in several human populations with elevated dioxin exposures (USEPA, 2000c). These are either occupational cohort studies of phenoxy herbicide workers who were exposed to mixtures of DLCs or studies of a population in Seveso, Italy, that was accidentally exposed to TCDD after an explosion at a chemical plant in 1976. The most recent follow-up of the Seveso cohort that was primarily exposed to TCDD indicates that exposure is associated with an increase in all cancers combined and with several specific cancers: rectal cancer, lung cancer, Hodgkins disease, non-Hodgkins lymphoma, and myeloid leukemia (Bertazzi et al., 2001). TCDD has been classified as a known human carcinogen by the IARC and the NTP based on an evaluation of all the human and rodent cancer data with consideration of the role of the aryl hydrocarbon receptor (AhR) in the mechanism of action of TCDD.

## **TUMOR PROMOTION STUDIES**

Numerous studies have examined the promotion of putative preneoplastic liver lesions by TCDD within the framework of a two-stage initiation-promotion protocol (Dragan and Schrenk, 2000). In the liver, clonal expansion of genetically altered cells leads to the formation of putative preneoplastic altered hepatocellular focal lesions identified by alterations in histomorphology or gene expression. These lesions are believed to be precursors in the development of liver tumors (Pitot *et al.*, 1991). These studies demonstrate that TCDD is a potent liver tumor promoter and that this effect is dose-dependent (Pitot *et al.*, 1980; Maronpot *et al.*, 1993; Teeguarden *et al.*, 1999), duration-of-exposure dependent, and reversible (Dragan *et al.*, 1992; Walker *et al.*, 1998, 2000). Studies also show that TCDD promotes more tumors in female rat liver than in male rat liver, and that this is due to the enhancing effect of estrogens on the promotion of preneoplastic lesions (Lucier *et al.*, 1991; Wyde *et al.*, 2001a, 2002). Moreover, this may explain why TCDD is a hepatocarcinogen in female but not male rats (Kociba *et al.*, 1978).

Tests of the tumor initiating and promoting capacity of TCDD have also been conducted in two-stage (initiation-TCDD promotion) models of mouse skin tumorgenesis (IARC, 1997; Dragan and Schrenk, 2000; USEPA, 2000c). Similar studies demonstrate that TCDD is at least two orders of magnitude more potent than the prototypical tumor promoter tetradecanoyl phorbol acetate (Poland et al., 1982) in those skin tumor promotion models. Studies looking at AhR-active congeners indicate that TCDD-mediated skin tumor promotion is likely to be AhR-dependent (Hebert et al., 1990). Overall, there appears to be insufficient evidence that TCDD is a tumor initiator in mouse skin (Dragan and Schrenk, 2000). Transgenic models have also been used to examine the mechanism of carcinogenicity of TCDD in mice (Eastin et al., 1998). These include the Tg.AC transgenic mouse that harbors an activated mouse v-Ha-*ras* oncogene (an intermediate in growth factor signaling). Dermal application of TCDD results in significant increases in the incidences of squamous cell papillomas in both male and female Tg.AC mice, supporting the conclusion that TCDD is a tumor promoter. Subsequent studies by the NTP showed that the induction of papillomas and squamous cell carcinomas by dermal application of TCDD to hemizygous Tg.AC mice was dose-dependent (Van Birgelen *et al.*, 1999; Dunson *et al.*, 2000). In addition, the induction of skin papillomas in this model occurred when TCDD was given by oral administration. These data provide further support for the potent tumor promoting and weak initiating activities of TCDD.

In addition to the liver and skin, TCDD is a tumor promoter in the lung (Anderson et al., 1991; Beebe et al., 1995). An increased trend in the incidence of lung tumors was observed in male mice in the NTP carcinogenesis bioassay of TCDD (NTP, 1982a). In subsequent studies, control or N-nitrosodimethylamine- (NDMA) initiated male Swiss mice were given single (1.6, 16, or  $48 \,\mu g/kg$ ) or multiple (0.05  $\mu g/kg$  per week for 20 weeks) intraperitoneal injections of TCDD and sacrificed at 52 weeks of age (Anderson et al., 1991). A 100% incidence rate of lung tumors in NDMA-initiated mice was observed in the absence of TCDD treatment. Tumor multiplicity was significantly increased in two NDMA/TCDD treatment groups (1.6 and 16 µg/kg) when compared to NDMA-initiated controls. In Sprague-Dawley rats, which have a much lower spontaneous incidence rate of lung tumors, TCDD alone can promote the development of bronchiolar hyperplasia and alveolar-bronchiolar metaplasia (Tritscher et al., 2000). It was demonstrated that the induction of these lesions was reversible; incidences of these lesions returned to control levels following withdrawal of TCDD for 16 or 30 weeks.

Overall, these data demonstrate that TCDD's dominant mode of action for carcinogenesis is likely as a potent tumor promoter.

## **Mechanistic Studies**

TCDD is generally classified as nongenotoxic and nonmutagenic. The common mechanism of action of TCDD and related compounds involves an initial binding to the AhR (Poland and Knutson, 1982; Safe, 1990; Whitlock, 1990; Schmidt and Bradfield, 1996). The broad spectrum of TCDD effects on hormone and growth factor systems, cytokines, and other signal transducer pathways indicates that TCDD is an extremely powerful growth dysregulator (Birnbaum, 1994a; Table 6). Because TCDD is not directly genotoxic (Wassom *et al.*, 1977), it is believed that the pathological responses associated with exposure to TCDD are fundamentally due to binding to and activation of the AhR, subsequent alterations in expression of TCDD-regulated genes, and altered signaling of biological pathways that interact with the AhR signal transduction mechanism (Poland and Knutson, 1982).

Alterations in expression of TCDD-regulated genes occur via a mechanism that involves a high-affinity interaction of TCDD and related polycyclic and polyhalogenated aromatic hydrocarbons with an intracellular receptor protein, the AhR, which functions as a ligandactivated transcription factor (Okey et al., 1994; Schmidt and Bradfield, 1996). Ligand binding initiates a signaling pathway in which the cytosolic AhR translocates to the nucleus (Whitlock, 1993). At some point subsequent to ligand binding, the AhR associates with an aromatic hydrocarbon nuclear translocator protein (ARNT) to form the nuclear DNA-binding and transcriptionally active AhR complex. Both the AhR and ARNT proteins are members of the basic helix-loophelix family of transcription factors (Hoffman et al., 1991; Burbach et al., 1992; Ema et al., 1992). The AhR-ARNT heterodimer binds with high affinity to a specific DNA sequence termed the dioxin response element (DRE). DREs have been identified in the enhancer regions of genes encoding several drug-metabolizing enzymes (Lai et al., 1996). The characteristic response to TCDD is the transcriptional induction of the cytochrome P450 1A1 gene (CYP1A1), which is mediated by binding of the AhR-ARNT complex to DREs present in the 5' flanking region of the gene. The AhR is expressed in all tissues examined (Dolwick et al., 1993), with a definite tissue specificity in terms of level of expression and diversity of response, indicating that TCDD is likely to have some effect in every tissue. However, even with the same receptor and the same ligand, there are both qualitative and quantitative differences in response, and these differences are likely to be involved in the tissue- and species-specificity of the response to TCDD. It is still not known how alterations in gene expression ultimately lead to the development of pathologies and adverse health effects associated with TCDD exposure. However, it is generally accepted that most, if not all, TCDD responses require an initial step of binding to the AhR.

The most studied response to TCDD is induction of the CYP1A class of cytochromes P450 (Whitlock, 1999). CYP1A1 is induced in many tissues including the liver, lung, kidney, nasal passages, and small intestine, with the highest induction in rats occurring in the liver.

Enzyme Induction	Modulation of Hormones and Receptors	Modulation of Growth Factors and Receptors		
CYP1A1	Androgens	Vitamin A		
CYP1B1	Estrogens	EGF		
CYP1A2	Estrogen receptor	TGFα		
Quinone reductase	Glucocorticoids	EGF receptor		
UDP-glucuronosyltransferase	Glucocorticoid receptor	TGFβ		
Glutathione-S-transferase	Insulin	ΤΝΓα		
Aldehyde dehydrogenase	Gastrin	IL-1β		
Ornithine decarboxylase	Thyroid hormones	c-Ras		
Tyrosine kinase	Melatonin	c-ErbA		
Terminal deoxynucleotidyltransferase				
Phosphoenolpyruvate carboxykinase				
Plasminogen activator inhibitor-2				

# TABLE 6Biochemical Effects of TCDD<sup>a</sup>

<sup>a</sup> Adapted from Birnbaum, 1994a

CYP1A1 induction is a very sensitive response to TCDD and, as such, serves as a useful marker for exposure to TCDD. TCDD induces CYP1A1 in vivo and in vitro in animal models and humans. Induction of 7-ethoxyresorufin-O-deethylase activity is a marker of CYP1A1 activity. CYP1A2 is constitutively expressed in the liver at low levels and is inducible by TCDD only in the liver and possibly nasal turbinates of rats (Goldstein and Linko, 1984). CYP1A2 is induced by TCDD, and induction of acetanilide-4-hydroxylase activity is a marker of CYP1A2 activity. In addition to the well-characterized induction of CYP1A1 and CYP1A2, TCDD induces another cytochrome P450, CYP1B1, in human cells (Sutter et al., 1994) and rodent tissues (Walker et al., 1995). CYP1B1 is active in the metabolism of numerous polycyclic aromatic hydrocarbons and arylamines and can catalyze the 4-hydroxylation of 17<sup>β</sup>-estradiol (Hayes et al., 1996; Murray et al., 2001).

TCDD is believed to disrupt thyroid hormone homeostasis via the induction of the phase II enzymes UDPglucuronosyltransferases (UGTs). Thyroxine ( $T_4$ ) production and secretion is controlled by thyroid stimulating hormone (TSH), which is under negative and positive regulation from the hypothalamus, pituitary gland, and thyroid gland by thyrotrophin releasing hormone, TSH itself,  $T_4$ , and triiodothyronine. TCDD induces the synthesis of UDP-glucuronosyltransferase-1 mRNA by an AhR-dependent transcriptional mechanism. Consequently, decreased serum  $T_4$  levels via induction of UGT may lead to a decrease in the negative feedback inhibition on the pituitary gland. This would then lead to a rise in secreted TSH, resulting in chronic hyperstimulation of the follicular cells of the thyroid gland.

TCDD has been shown to modulate numerous growth factor, cytokine, hormone and metabolic pathways in animals and experimental systems (Sutter and Greenlee, 1992; Birnbaum, 1994b). Many, if not all, are part of pathways involved in cellular proliferation and differentiation. These include the glucocorticoid receptor tyrosine kinases, interleukin-1-beta, plasminogen activator inhibitor-2, urokinase type plasminogen activator, tumor necrosis factor-alpha, gonadotrophin releasing hormone, testosterone, and prostaglandin endoperoxide H synthase-2. More recently, the application of toxicogenomic analyses has increased our understanding of the number of genes/proteins altered by TCDD both in vitro (Puga et al., 2000; Martinez et al., 2002) and in vivo (Bruno et al., 2002; Kurachi et al., 2002; Zeytun et al., 2002). Most of the molecular details for induction of gene expression via the AhR have been characterized for the transcriptional activation of the CYP1A1 gene (Whitlock, 1999). While TCDD has been shown to affect the expression of many genes, there is detailed characterization of transcriptional activation through the AhR for only a few of these.

## **GENETIC TOXICOLOGY**

In general, polychlorinated biphenyl compounds are not mutagenic in standard short-term mutagenicity tests conducted *in vitro* or *in vivo*. TCDD follows this same activity pattern. TCDD (tested up to the limits of toxicity) was not mutagenic in *Salmonella typhimurium* with or without exogenous liver S9 metabolic activation enzymes (Mortelmans et al., 1984; Blevins, 1991). No induction of mutations was seen at the trifluorothymidine locus in mouse lymphoma L5178Y tk<sup>+/-</sup> cells treated with up to 1.0 µg/mL TCDD with or without S9 (McGregor et al., 1991). In tests for induction of chromosomal damage in cultured Chinese hamster ovary cells, TCDD did not induce sister chromatid exchanges or chromosomal aberrations when tested up to toxic levels (0.8 µg/mL) with or without S9 (Galloway et al., 1987). In in vivo tests, no induction of sex-linked recessive lethal mutations was observed in germ cells of male Drosophila melanogaster following injection of adult flies with 50, 250, or 500 ppm TCDD (Zimmering et al., 1985). TCDD, administered for 6 weeks at a dose of 2 µg/kg to male and female Big Blue transgenic rats, did not alter the frequency or spectrum of mutations (Thornton et al., 2001). There is one report of a positive result with TCDD (and two other polychlorinated aromatic hydrocarbons) in a test that measured induction of chromosomal deletions, resulting from intrachromosomal recombination, in mouse embryos in vivo (Schiestl et al., 1997); in this study, the two other compounds, Aroclor 1221 and Aroclor 1260, also induced deletions in yeast cells. An earlier study of the potential for TCDD-induced cytogenetic damage in laboratory mice showed no increase in the frequencies of sister chromatid exchanges, chromosomal aberrations, or micronuclei in bone marrow cells of either C57Bl/6J or DBA/21 mice, following administration of a single dose of up to

150 μg TCDD/kg (Meyne *et al.*, 1985). In summary, no mutagenic activity was detected with TCDD in a variety of *in vitro* and *in vivo* short-term tests.

### **STUDY RATIONALE**

The female Sprague-Dawley rat was selected as the model for the current study because this sex and species has been used frequently in chronic and subchronic studies of dioxins. In addition, this was the model in which TCDD was demonstrated as a carcinogen in a dosedfeed study conducted by the Dow Chemical Company (Kociba et al., 1978). Moreover, the incidence of liver tumors in female Sprague-Dawley rats has frequently been the primary rodent carcinogenicity dataset used by regulatory agencies worldwide for development of cancer risk guidelines for TCDD exposure. The doses chosen for the current study were based on the 1 to 100 ng/kg per day range used in the Kociba et al. (1978) feed study where increased incidences of liver tumors were seen. These studies were not specifically designed to determine a no-observed-adverse-effect level or lowest-observed-adverse-effect level; rather, doses used in the present study were selected to increase doseresponse data density in the 10 to 100 ng/kg range, where increases in liver and lung tumors were expected, to facilitate derivation of relative potency factors for carcinogenesis. Male rats were not studied due to the lack of induction of liver and lung tumors in the previous studies of Sprague-Dawley rats.

# **MATERIALS AND METHODS**

## PROCUREMENT

## AND CHARACTERIZATION OF TCDD

TCDD was obtained from IIT Research Institute (Chicago, IL) in one lot (CR82-2-2) that was used for the 2-year study. Identity and purity analyses were conducted by the analytical chemistry laboratory, Research Triangle Institute (Research Triangle Park, NC), and the study laboratory, Battelle Columbus Operations (Columbus, OH). Reports on analyses performed in support of the TCDD study are on file at the National Institute of Environmental Health Sciences.

The chemical, a white crystalline powder, was identified as TCDD by infrared spectroscopy, proton nuclear magnetic resonance spectroscopy, direct probe mass spectroscopy (MS), low resolution gas chromotography (GC) coupled with MS, and melting point determination. A precise melting point range was not determined as the chemical appeared to sublime at approximately 260° C. The purity was determined by GC. The purity profiles detected two impurities with a combined relative area of 2.0% and two impurities with a combined relative area of 1.6%. The major impurity in each profile (1.5% of the major peak) was identified using GC/MS as 1,2,4trichlorodibenzo-p-dioxin. A small peak eluting immediately after the main component was believed to be a dimethyl isomer of trichloro-p-dioxin (positional substitution unknown). Also, a trace amount of a higher molecular weight tetrachlorinated dioxin (parent ion = 426) was observed, but due to the relatively weak intensity of the signal, precise identification could not be made. The overall purity was determined to be 98% or greater.

### **PREPARATION OF STOCK SAMPLES**

Lot CR82-2-2 was dissolved in acetone and prealiquotted for use as analytical stock or formulation stock in the study because of the very small amount of chemical that was required to prepare the dose formulations at the intended concentrations. Details concerning the preparation and use of these stock solutions are provided in Appendix D. The test article was stored at room temperature and protected from light in amber glass bottles. Purity was monitored with periodic reanalysis using GC. No degradation was observed during the course of the study.

# **PREPARATION AND ANALYSIS**

# **OF DOSE FORMULATIONS**

The dose formulations were prepared by mixing TCDD with acetone and then diluting the mixture in corn oil (Spectrum Quality Products) such that the final concentration contained 1.0% acetone (Table D2). The dose formulations were stored at room temperature in amber glass bottles with minimal headspace, sealed with Teflon<sup>®</sup>-lined lids, for up to 35 days.

Homogeneity studies of 1.2 and 40 ng/mL dose formulations and simulated animal room stability studies of a 1.2 ng/mL dose formulation were performed by the study laboratory using GC/MS. Homogeneity was confirmed, and stability was confirmed for 3 hours under simulated animal room conditions. An additional stability study of a 0.2 ng/mL dose formulation was performed by the analytical laboratory, and stability was confirmed for at least 35 days for the dose formulation stored in sealed glass containers at 5° C and room temperature. Gavagability was confirmed by the study laboratory for the 40 ng/mL formulation.

Periodic analyses of the dose formulations of TCDD were conducted by the study laboratory using GC/MS. During the 2-year study, the dose formulations were analyzed at least every 3 months (Table D3). Of the dose formulations analyzed, 56 of 58 were within 10% of the target concentrations, and 19 of 21 animal room samples were within 10% of the target concentrations. Periodic analyses of the corn oil vehicle performed by the study laboratory using potentiometric titration demonstrated peroxide concentrations less than 3 mEq/kg.

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# **2-YEAR STUDY**

#### **Study Design**

Groups of 81 or 82 (3 ng/kg) female rats received TCDD in corn oil:acetone (99:1) by gavage at doses of 3, 10, 22, 46, or 100 ng/kg 5 days per week for up to 105 weeks; a group of 81 female rats received the corn oil:acetone (99:1) vehicle alone. Up to 10 rats per group were evaluated at 14, 31, or 53 weeks. For stop-exposure evaluation, a group of 50 female rats was given 100 ng/kg for 30 weeks and then the vehicle for the remainder of the study.

Additional "special study" animals were included at each interim necropsy. Tissues from these animals were provided to specific extramural grantees to facilitate the conduct of additional mechanistic studies. These animals were not evaluated as part of the core study.

#### Source and Specification of Animals

Male and female Sprague-Dawley rats were obtained from Harlan Sprague-Dawley, Inc. (Indianapolis, IN), for use in the 2-year study. Sufficient male rats were included in this study to ensure normal estrous cycling of female rats. Male rats were not administered the test compound. Rats were quarantined for 11 days before and were approximately 8 weeks old at the beginning of the study. Rats were evaluated for parasites and gross observation of disease, and the health of the rats was monitored during the studies according to the protocols of the NTP Sentinel Animal Program (Appendix F). Sentinel rats included five males and five females at 1 month, five males at 6, 12, and 18 months, and five 100 ng/kg females at the end of the study.

#### **Animal Maintenance**

Male rats were housed three per cage, and female rats were housed three or five per cage. Feed and water were available *ad libitum*. Cages were changed twice weekly; racks were changed and rotated every 2 weeks. Further details of animal maintenance are given in Table 7. Information on feed composition and contaminants is provided in Appendix E.

#### **Clinical Examinations and Pathology**

All animals were observed twice daily. Clinical findings were recorded on day 29, monthly thereafter, and at the end of the study. Body weights were recorded on the first day of the study, weekly for 13 weeks, monthly thereafter, and at the end of the study. At 14, 31, and 53 weeks, blood was taken from the retroorbital sinus of up to ten female rats per group (except stop-exposure) and processed into serum for thyroid hormone determinations. Radioimmunoassays were performed for thyroid stimulating hormone, triiodothyronine, and free thyroxine ( $T_4$ ) using a Packard Cobra II gamma counter (Packard Instrument Company, Meriden, CT). The assay for total  $T_4$  was performed on a Hitachi 911<sup>®</sup> chemistry analyzer (Boehringer Mannheim, Indianapolis, IN) using a Boehringer Mannheim<sup>®</sup> enzyme immunoassay test system. Thyroid hormone data were summarized using the XYBION system (XYBION Medical Systems Corporation, Cedar Knolls, NJ).

For cell proliferation analysis at 14, 31, and 53 weeks, up to 10 female rats per group (except stop-exposure) received drinking water containing 40 mg BrdU/100 mL Milli-Q water for 5 days. BrdU solutions were administered in amber glass water bottles (Allentown Caging Equipment Company, Inc., Allentown, NJ) equipped with Teflon<sup>®</sup>-lined lids and stainless steel sipper tubes. BrdU solutions were changed after 3 days, and water consumption was measured daily for 5 days. Cell turnover rate in the liver of dosed female rats was compared to the turnover rate in the vehicle control rats by determining the incorporation of BrdU into hepatocytes. A sample of duodenum and liver was fixed in 10% neutral buffered formalin for 18 to 24 hours then transferred to 70% ethanol. Representative sections of the duodenum and liver were trimmed and embedded, and two sections were cut. One of these sections was stained with hematoxylin and eosin and the other with anti-BrdU antibody complexed with avidin and biotin. At the 14-week interim evaluation, potential interlobular variation was determined in the vehicle control and 100 ng/kg groups by counting stained cells in the left lobe and right median lobe. Interlobular variation greater than 25% was considered significant. For the remaining rats, stained cells were counted only in the left lobe. Two thousand labeled or unlabeled hepatocyte nuclei were counted using a  $20\times$  objective and ocular grid. The labeling index is expressed as the percentage of total nuclei that were labeled with BrdU.

For determination of cytochrome P450 activities, liver and lung tissue samples were collected from up to 10 rats per group (except stop-exposure) at 14, 31, and 53 weeks and stored frozen at  $-70^{\circ}$  C. Microsomal suspensions were prepared using the Pearce method (Pearce *et al*, 1996). The concentration of protein in each suspension was determined using the microtiter plate method of the Coomassie Plus Protein Assay (Pierce Chemical Co., Rockford, IL) with bovine serum albumin as the standard. Cytochrome P450 1A1 (CYP1A1)-associated 7-ethoxyresorufin-*O*-deethylase (EROD), CYP1A2associated acetanilide-4-hydroxylase (A4H), and CYP2B-associated pentoxyresorufin-*O*-deethylase (PROD) activities were determined in microsomal proteins, isolated from frozen liver or lung tissue according to established procedures. Data are shown as pmol/minute per mg (EROD and PROD) or nmol/minute per mg (A4H) microsomal protein.

For analysis of tissue concentrations of TCDD, samples of fat, liver, lung, and blood were taken from up to 10 female rats per group at 14, 31, and 53 weeks and at 2 years. Analysis of fat, lung, and blood samples included overnight saponification with potassium hydroxide (blood samples were sonicated following digestion), extraction of the saponificate with hexanes: acetone (1:4 for fat and blood, 1:1 for lung) using vortexing and centrifugation, sample extract cleanup using open-bed silica gel/sodium sulfate columns, and elution with hexanes. Liver samples were homogenized in saturated aqueous sodium sulfate, extracted with hexanes: acetone (1:9), and cleaned and eluted using the same techniques as for the other tissues. Concentrations of TCDD in the tissue extracts were measured by capillary gas chromatography with high resolution mass spectrometry detection.

Complete necropsies and microscopic examinations were performed on all rats. At the interim evaluations, the left kidney, liver, lung, left ovary, spleen, thymus (14 weeks only), and thyroid gland were weighed. At necropsy, all organs and tissues were examined for grossly visible lesions, and all major tissues were fixed and preserved in 10% neutral buffered formalin, processed and trimmed, embedded in paraffin, sectioned to a thickness of 4 to 6  $\mu$ m, and stained with hematoxylin and eosin for microscopic examination. For all paired organs (e.g., adrenal gland, kidney, ovary), samples from each organ were examined. Tissues examined microscopically are listed in Table 7.

Microscopic evaluations were completed by the study laboratory pathologist, and the pathology data were entered into the Toxicology Data Management System. The slides, paraffin blocks, and residual wet tissues were sent to the NTP Archives for inventory, slide/block match, and wet tissue audit. The slides, individual animal data records, and pathology tables were evaluated by an independent quality assessment laboratory. The individual animal records and tables were compared for accuracy, the slide and tissue counts were verified, and the histotechnique was evaluated. A quality assessment pathologist evaluated slides from all tumors and all potential target organs, which included the adrenal cortex, liver, lung, oral mucosa, pancreas, thymus, thyroid gland, and uterus.

The quality assessment report and the reviewed slides were submitted to the NTP Pathology Working Group (PWG) chairperson, who reviewed the selected tissues and addressed any inconsistencies in the diagnoses made by the laboratory and quality assessment pathologists. Representative histopathology slides containing examples of lesions related to chemical administration, examples of disagreements in diagnoses between the laboratory and quality assessment pathologists, or lesions of general interest were presented by the chairperson to the PWG for review. The PWG consisted of the quality assessment pathologist and other pathologists experienced in rodent toxicologic pathology. This group examined the tissues without any knowledge of dose groups or previously rendered diagnoses. When the PWG consensus differed from the opinion of the laboratory pathologist, the diagnosis was changed. Final diagnoses for reviewed lesions represent a consensus between the laboratory pathologist, reviewing pathologist(s), and the PWG. Details of these review procedures have been described, in part, by Maronpot and Boorman (1982) and Boorman et al. (1985). For subsequent analyses of the pathology data, the decision of whether to evaluate the diagnosed lesions for each tissue type separately or combined was generally based on the guidelines of McConnell et al. (1986).

To maintain consistency of diagnoses within and among all the studies on dioxin-like compounds (DLCs) conducted as part of the dioxin TEF evaluation, the same pathologists were involved in all phases of the pathology evaluation including the initial examination and the pathology peer review. Because of the need for a consistent diagnostic approach across all studies and the unusual nature of some of the lesions, this study of TCDD, along with three other studies (PCB126, the TEF mixture, and PeCDF), were subjected to additional PWG reviews. Within many of these studies, there were hepatocellular proliferative lesions for which the criteria used for common diagnoses did not appear to fit. Furthermore, classification was sometimes confounded by significant liver damage (toxic hepatopathy) that was present in many animals from these studies. With the consecutive pathology peer review of each of these
studies, the morphological spectrum of proliferative lesions became more apparent to those involved and the diagnostic criteria for the proliferative lesion further refined. Therefore, a PWG review was held to ensure that these important proliferative lesions were sufficiently and consistently categorized across all seven studies for which data are to be compared. PWG participants for this review were primarily those involved in previous PWGs. A different group of pathologists was also convened to provide additional guidance on the most appropriate classification of the hepatocellular proliferative lesions from these studies of DLCs (Hailey *et al.*, 2005). Participants included Drs. Jerrold Ward, Ernest McConnell, James Swenberg, Michael Elwell, Peter Bannasch, Douglas Wolf, John Cullen, and Rick Hailey. Final diagnoses for the hepatocellular proliferative lesions reflect the consensus of this complete review process.

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### TABLE 7 Experimental Design and Materials and Methods in the 2-Year Gavage Study of TCDD

#### Study Laboratory Battelle Columbus Operations (Columbus, OH)

#### Strain and Species

Harlan Sprague-Dawley rats: Hsd; Sprague-Dawley<sup>TM</sup>

#### Animal Source Harlan Sprague-Dawley, Inc. (Indianapolis, IN)

**Time Held Before Study** 11 days

Average Age When Study Began 8 weeks

**Date of First Dose (female rats only)** June 1, 1998

**Duration of Dosing** 5 days/week for 14, 31, or 53 (interim evaluation), or 30 (stop-exposure), or 105 weeks

#### Date of Last Dose May 30-June 1, 2000 (core study), December 30, 1998 (stop-exposure group)

Necropsy Dates May 31-June 2, 2000

Average Age at Necropsy 112 weeks

#### Size of Study Groups

81 (vehicle control, 10, 22, 46, and 100 ng/kg), 82 (3 ng/kg), or 50 (100 ng/kg stop-exposure)

#### Method of Distribution

Animals were distributed randomly into groups of approximately equal initial mean body weights.

Animals per Cage 3 males; 3 or 5 females

#### Method of Animal Identification

Tail tattoo

#### Diet

Irradiated NTP-2000 pelleted diet (Zeigler Brothers, Inc., Gardners, PA), available ad libitum

#### Water

Tap water (Columbus municipal supply) via automatic watering system, except via amber glass bottles during BrdU administration, available *ad libitum* 

#### Cages

Solid polycarbonate (Lab Products, Inc., Maywood, NJ), changed twice weekly

#### Bedding

Irradiated Sani-Chips® hardwood chips (P.J. Murphy Forest Products Corp., Montville, NJ), changed twice weekly

#### **Cage Filters**

Dupont 2024 spun-bonded polyester sheets (Snow Filtration Co., Cincinnati, OH), changed every 2 weeks

### TABLE 7 Experimental Design and Materials and Methods in the 2-Year Gavage Study of TCDD

#### Racks

Stainless steel (Lab Products, Inc., Maywood, NJ), changed and rotated every 2 weeks

#### **Animal Room Environment**

Temperature:  $72^{\circ} \pm 3^{\circ}$  F Relative humidity:  $50\% \pm 15\%$ Room fluorescent light: 12 hours/day Room air changes: 10/hour

#### Doses

0, 3, 10, 22, 46, or 100 ng/kg

#### **Type and Frequency of Observation**

Observed twice daily; animals were weighed initially, weekly for 13 weeks, monthly thereafter, and at the end of the study. Clinical findings were recorded on day 29, monthly thereafter, and at the end of the study.

#### Method of Sacrifice

Carbon dioxide asphyxiation

#### Necropsy

Necropsy was performed on all female rats. At the 14-, 31-, and 53-week interim evaluations, the left kidney, liver, lung, left ovary, spleen, thymus (14 weeks only), and thyroid gland were weighed.

#### **Thyroid Hormone Analysis**

At 14, 31, and 53 weeks, blood was collected from the retroorbital sinus of up to 10 rats per group for thyroid stimulating hormone, triiodothyronine, and total and free thyroxine determinations.

#### **Cell Proliferation**

At 14, 31, and 53 weeks, up to 10 rats per group received BrdU in drinking water for 5 days. Samples from the liver and duodenum were measured for BrdU labeling.

#### **Cytochrome P450 Activities**

At 14, 31, and 53 weeks, tissue samples from the liver were taken from up to 10 rats per group for 7-ethoxyresorufin-*O*-deethylase, 7-pentoxyresorufin-*O*-deethylase, and acetanilide-4-hydroxylase activities. Lung samples from these rats were analyzed for 7-ethoxyresorufin-*O*-deethylase activity.

#### **Tissue Concentration Analysis**

At 14, 31, 53, and 104 weeks, samples of blood, fat, liver, and lung were taken from up to 10 rats per group for analysis of TCDD concentrations.

#### Histopathology

Complete histopathology was performed on all core study and stop-exposure rats at 2 years. In addition to gross lesions and tissue masses, the following tissues were examined: adrenal gland, bone with marrow, brain, clitoral gland, esophagus, eye, harderian gland, heart, large intestine (cecum, colon, rectum), small intestine (duodenum, jejunum, ileum), kidney, liver, lung with mainstem bronchi, lymph nodes (mandibular and mesenteric), mammary gland, nose, ovary, pancreas, parathyroid gland, pituitary gland, salivary gland, skin, spleen, stomach (forestomach and glandular), thymus, thyroid gland, trachea, urinary bladder, and uterus. The adrenal gland, liver, lung, mammary gland, ovary, pancreas, pituitary gland, spleen, stomach (forestomach and glandular), thymus, thyroid gland, uterus, and vagina were examined at 14, 31, and 53 weeks.

#### **STATISTICAL METHODS**

#### **Survival Analyses**

The probability of survival was estimated by the product-limit procedure of Kaplan and Meier (1958) and is presented in the form of graphs. Animals found dead of other than natural causes or missing were censored from the survival analyses; animals dying from natural causes were not censored. Statistical analyses for possible dose-related effects on survival used Cox's (1972) method for testing two groups for equality and Tarone's (1975) life table test to identify dose-related trends. All reported P values for the survival analyses are two sided.

#### **Calculation of Incidence**

The incidences of neoplasms or nonneoplastic lesions are presented in Tables A1a, A1b, A1c, A5a, A5b, and A5c as the numbers of animals bearing such lesions at a specific anatomic site and the numbers of animals with that site examined microscopically. For calculation of statistical significance, the incidences of most neoplasms (Table A3a and A3b) and all nonneoplastic lesions are given as the numbers of animals affected at each site examined microscopically. However, when macroscopic examination was required to detect neoplasms in certain tissues (e.g., harderian gland, intestine, mammary gland, and skin) before microscopic evaluation, or when neoplasms had multiple potential sites of occurrence (e.g., leukemia or lymphoma), the denominators consist of the numbers of animals on which a necropsy was performed. Tables A3a and A3b also give the survival-adjusted neoplasm rate for each group and each site-specific neoplasm. This survival-adjusted rate (based on the Poly-3 method described below) accounts for differential mortality by assigning a reduced risk of neoplasm, proportional to the third power of the fraction of time on study, to animals that do not reach terminal sacrifice.

#### Analysis of Neoplasm and Nonneoplastic Lesion Incidences

The Poly-k test (Bailer and Portier, 1988; Portier and Bailer, 1989; Piegorsch and Bailer, 1997) was used to assess neoplasm and nonneoplastic lesion prevalence. This test is a survival-adjusted quantal-response procedure that modifies the Cochran-Armitage linear trend test to take survival differences into account. More specifically, this method modifies the denominator in the quantal estimate of lesion incidence to approximate more closely the total number of animal years at risk. For analysis of a given site, each animal is assigned a risk weight. This value is one if the animal had a lesion at that site or if it survived until terminal sacrifice; if the animal died prior to terminal sacrifice and did not have a lesion at that site, its risk weight is the fraction of the entire study time that it survived, raised to the kth power.

This method yields a lesion prevalence rate that depends only upon the choice of a shape parameter for a Weibull hazard function describing cumulative lesion incidence over time (Bailer and Portier, 1988). Unless otherwise specified, a value of k=3 was used in the analysis of sitespecific lesions. This value was recommended by Bailer and Portier (1988) following an evaluation of neoplasm onset time distributions for a variety of site-specific neoplasms in control F344 rats and B6C3F1 mice (Portier et al., 1986). Bailer and Portier (1988) showed that the Poly-3 test gave valid results if the true value of k was anywhere in the range from 1 to 5. A further advantage of the Poly-3 method is that it does not require lesion lethality assumptions. Variation introduced by the use of risk weights, which reflect differential mortality, was accommodated by adjusting the variance of the Poly-3 statistic as recommended by Bieler and Williams (1993).

Tests of significance included pairwise comparisons of each dosed group with controls and a test for an overall dose-related trend. Continuity-corrected Poly-3 tests were used in the analysis of lesion incidence, and reported P values are one sided. The significance of lower incidences or decreasing trends in lesions is represented as 1–P with the letter N added (e.g., P=0.99 is presented as P=0.01N). For neoplasms and nonneoplastic lesions detected at the interim evaluations, the Fisher exact test (Gart *et al.*, 1979), a procedure based on the overall proportion of affected animals, was used.

#### Analysis of Continuous Variables

Two approaches were employed to assess the significance of pairwise comparisons between dosed and control groups in the analysis of continuous variables. Organ and body weight data, which historically have approximately normal distributions, were analyzed with the parametric multiple comparison procedures of Dunnett (1955) and Williams (1971, 1972). Thyroid hormone, cell proliferation, and cytochrome P450 data, which have typically skewed distributions, were analyzed using the nonparametric multiple comparison methods of Shirley (1977) (as modified by Williams, 1986) and Dunn (1964). Jonckheere's test (Jonckheere, 1954) was used to assess the significance of the doserelated trends and to determine whether a trend-sensitive test (Williams' or Shirley's test) was more appropriate for pairwise comparisons than a test that does not assume a monotonic dose-related trend (Dunnett's or Dunn's test). Prior to statistical analysis, extreme values identified by the outlier test of Dixon and Massey (1957) were examined by NTP personnel, and implausible values were eliminated from the analysis. Average severity values were analyzed for significance with the Mann-Whitney U test (Hollander and Wolfe, 1973).

#### **Historical Control Data**

The concurrent control group represents the most valid comparison to the treated groups and is the only control group analyzed statistically in NTP bioassays. However, historical control data are often helpful in interpreting potential treatment-related effects, particularly for uncommon or rare neoplasm types. For meaningful comparisons, the conditions for studies in the historical database must be generally similar. For female Sprague-Dawley rats, the NTP historical database is currently limited to the seven gavage studies conducted as part of the dioxin TEF evaluation (the current TCDD study, PCB 126, the TEF mixture, PeCDF, PCB 153, the PCB 126/PCB 153 mixture, and the PCB 126/PCB 118 mixture; NTP, 2006a,b,c,d,e,f).

#### **QUALITY ASSURANCE METHODS**

The 2-year study was conducted in compliance with Food and Drug Administration Good Laboratory Practice Regulations (21 CFR, Part 58). In addition, as records from the 2-year study were submitted to the NTP Archives, this study was audited retrospectively by an independent quality assurance contractor. Separate audits covered completeness and accuracy of the final pathology tables and a draft of this NTP Technical Report. Audit procedures and findings are presented in the reports and are on file at NIEHS. The audit findings were reviewed and assessed by NTP staff, and all comments were resolved or otherwise addressed during the preparation of this Technical Report.

#### **GENETIC TOXICOLOGY**

The genetic toxicity of TCDD was assessed by testing the ability of the chemical to induce mutations in various strains of *Salmonella typhimurium*, mutations in L5178Y mouse lymphoma cells, sister chromatid exchanges and chromosomal aberrations in cultured Chinese hamster ovary cells, sex-linked recessive lethal mutations in *Drosophila melanogaster*, and micronucleated erythrocytes in mouse bone marrow. The protocols for these studies and the results are given in Appendix B. The genetic toxicity studies have evolved from an earlier effort by the NTP to develop a comprehensive database permitting a critical anticipation of a chemical's carcinogenicity in experimental animals based on numerous considerations, including the molecular structure of the chemical and its observed effects in short-term *in vitro* and *in vivo* genetic toxicity tests (structure-activity relationships). The short-term tests were originally developed to clarify proposed mechanisms of chemicalinduced DNA damage based on the relationship between electrophilicity and mutagenicity (Miller and Miller, 1977) and the somatic mutation theory of cancer (Straus, 1981; Crawford, 1985). However, it should be noted that not all cancers arise through genotoxic mechanisms.

DNA reactivity combined with *Salmonella* mutagenicity is highly correlated with induction of carcinogenicity in multiple species/sexes of rodents and at multiple tissue sites (Ashby and Tennant, 1991). A positive response in the *Salmonella* test was shown to be the most predictive *in vitro* indicator for rodent carcinogenicity (89% of the *Salmonella* mutagens are rodent carcinogens) (Tennant *et al.*, 1987; Zeiger *et al.*, 1990). Additionally, no battery of tests that included the *Salmonella* test improved the predictivity of the *Salmonella* test alone. However, these other tests can provide useful information on the types of DNA and chromosomal damage induced by the chemical under investigation.

The predictivity for carcinogenicity of a positive response in acute in vivo bone marrow chromosome aberration or micronucleus tests appears to be less than that in the Salmonella test (Shelby et al., 1993; Shelby and Witt, 1995). However, clearly positive results in long-term peripheral blood micronucleus tests have high predictivity for rodent carcinogenicity (Witt et al., 2000); negative results in this assay do not correlate well with either negative or positive results in rodent carcino-Because of the theoretical and genicity studies. observed associations between induced genetic damage and adverse effects in somatic and germ cells, the determination of in vivo genetic effects is important to the overall understanding of the risks associated with exposure to a particular chemical. Most organic chemicals that are identified by the International Agency for Research on Cancer as human carcinogens, other than hormones, are genotoxic. The vast majority of these are detected by both the Salmonella assay and rodent bone marrow cytogenetics tests (Shelby, 1988; Shelby and Zeiger, 1990).

### RESULTS

#### **2-YEAR STUDY**

#### Survival

Estimates of 2-year survival probabilities for female rats are shown in Table 8 and in the Kaplan-Meier survival curves (Figure 1). Survival of dosed groups of rats was similar to that of the vehicle control group.

#### **Body Weights and Clinical Findings**

Mean body weights of the 100 ng/kg core study group and the 100 ng/kg stop-exposure group were less than those of the vehicle control group after week 13 of the study (Figure 2 and Table 9). Mean body weights of the 46 ng/kg group were less than those of the vehicle control group during year 2 of the study, and mean body weights of 22 ng/kg rats were less than those of the vehicle controls the last 10 weeks of the study. No clinical findings related to TCDD administration were observed.

 TABLE 8

 Survival of Female Rats in the 2-Year Gavage Study of TCDD

	Vehicle Control	3 ng/kg	10 ng/kg	22 ng/kg	46 ng/kg	100 ng/kg	100 ng/kg (Stop- Exposure)
Animals initially in study	81	82	81	81	81	81	50
14-Week interim evaluation	10	10	10	10	10	10	
31-Week interim evaluation <sup>a</sup>	10	10	10	10	10	10	
53-Week interim evaluation <sup>a</sup>	8	8	8	8	8	8	
Accidental deaths <sup>a</sup>	0	0	5	0	0	0	0
Moribund	19	27	15	19	17	17	16
Natural deaths	9	6	10	15	14	15	13
Animals surviving							
to study termination	25	21	23	19	22	21	21
Percent probability of surviv	al						
at end of study <sup>b</sup>	47	39	49	36	42	40	42
Mean survival (days) <sup>c</sup>	637	598	619	599	634	672	650
Survival analysis <sup>d</sup> P=	=0.736N	P=0.289	P=0.840N	P=0.184	P=0.728	P=0.807	P=0.860

<sup>a</sup>, Censored from survival analyses

b Kaplan-Meier determinations

Mean of all deaths (uncensored, censored, and terminal sacrifice)

The result of the life table trend test (Tarone, 1975) is in the vehicle control column, and the results of the life table pairwise comparisons (Cox, 1972) with the vehicle controls are in the dosed group columns. A negative trend or a lower mortality in a dosed group is indicated by **N**. The trend test does not include the 100 ng/kg stop-exposure group.

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FIGURE 1 Kaplan-Meier Survival Curves for Female Rats Administered TCDD by Gavage for 2 Years



FIGURE 2 Growth Curves for Female Rats Administered TCDD by Gavage for 2 Years

TABLE 9

Mean Body Weights and Survival of Female Rats in the 2-Year Gavage Study of TCDD

Weeks	Vehicle	Control		3 ng/kg			10 ng/kg			22 ng/kg	
on	Av. Wt.	No. of	Av. Wt.	Wt. (% of	No. of	Av. Wt.	Wt. (% of	No. of	Av. Wt.	Wt. (% of	No. of
Study	(g)	Survivors	(g)	controls)	Survivors	(g)	controls)	Survivors	(g)	controls)	Survivors
1	177	98	175	99	98	175	99	98	175	99	98
2	108	98	196	99	98	108	100	98	196	99	08
3	216	98	214	99	98	216	100	98	214	99	98
4	231	98	214	99	98	231	100	98	229	99	98
5	237	98	237	100	98	241	102	98	241	102	98
6	247	98	245	99	98	248	101	98	246	100	98
7	255	98	249	98	98	252	99	98	252	99	98
8	260	98	258	99	98	260	100	97	258	99	98
9	265	98	262	99	98	262	99	96	260	98	97
10	270	98	267	99	98	268	99	96	265	98	97
11	274	98	271	99	98	273	100	96	268	98	97
12	278	98	273	98	98	275	99	96	273	98	97
13	279	98	275	99	98	278	100	96	275	98	97
$17^{a}$	290	82	284	98	82	284	98	80	280	97	81
21	296	82	292	99	82	292	99	80	288	97	81
25	300	81	298	100	82	298	99	80	290	97	81
29	302	80	302	100	82	300	100	79	295	98	80
33 <sup>a</sup>	306	64	308	101	65	304	99	63	298	97	64
37	311	64	314	101	64	309	99	61	304	98	64
41	315	64	318	101	62	314	100	61	309	98	64
45	321	64	322	100	62	319	99	61	314	98	64
49	328	64	330	101	62	328	100	61	317	97	64
52	329	62	334	102	60	330	100	59	320	97	63
57 <sup>a</sup>	337	48	338	100	48	332	99	46	328	97	47
61	345	48	349	101	47	341	99	46	337	98	44
65	353	48	356	101	45	350	99	45	342	97	44
69	359	46	365	102	44	360	100	45	352	98	42
73	360	44	368	102	41	361	100	45	353	98	41
77	364	41	377	103	40	367	101	45	355	98	39
81	368	40	373	102	33	372	101	44	359	98	36
85	371	40	375	101	31	368	99	39	363	98	33
89	375	39	375	100	27	368	98	36	359	96	29
93	382	37	375	98	26	372	97	33	359	94	25
97	381	31	376	99	24	377	99	30	356	94	24
101	383	27	373	98	23	377	99	28	360	94	21
Mean for	weeks										
1-13	245		242	99		244	100		242	99	
14-52	310		310	100		308	99		302	97	
53-101	365		367	101		363	99		352	96	

TABLE 9

Mean Body Weights and Survival of Female Rats in the 2-Year Gavage Study of TCDD

Weeks	ks 46 ng/kg 100 ng/kg 100 ng/kg (St						ıg/kg (Stop-E	op-Exposure)		
on	Av. Wt.	Wt. (% of	No. of	Av. Wt.	Wt. (% of	No. of	Av. Wt.	Wt. (% of	No. of	
Study	(g)	controls)	Survivors	(g)	controls)	Survivors	(g)	controls)	Survivors	
1	175	99	98	176	100	98	174	99	50	
2	197	99	98	198	100	98	195	98	50	
3	216	100	98	215	99	98	214	99	50	
4	229	99	98	228	98	98	226	98	50	
5	237	100	98	234	99	98	232	98	50	
6	244	99	98	240	97	98	232	96	50	
0 7	250	98	98	247	97	98	244	96	50	
8	256	98	98	252	97	98	251	96	50	
9	260	98	98	252	96	98	253	95	50	
10	263	97	98	257	95	98	255	95	50	
11	265	97	98	260	95	98	255	93	50	
12	268	96	98	264	95	98	260	94	50	
13	270	97	98	264	95	98	262	94	50	
$17^{a}$	276	95	82	272	94	82	267	92	50	
21	284	96	82	279	95	82	273	92	50	
25	286	96	81	282	94	82	274	91	50	
29	288	96	80	281	93	82	277	92	50	
$\overline{33}^{a}$	293	96	64	282	92	66	279	91	49	
37	297	95	64	286	92	66	284	91	49	
41	300	95	64	288	92	66	291	93	49	
45	306	95	64	289	90	66	296	92	49	
49	309	94	63	293	89	66	300	92	49	
52	310	94	62	294	89	66	302	92	48	
$57^{a}$	320	95	49	300	89	53	306	91	48	
61	322	93	48	302	88	52	312	91	46	
65	326	92	47	305	86	52	319	91	46	
69	330	92	46	310	86	51	320	89	45	
73	324	90	44	307	85	51	321	89	44	
77	327	90	43	307	84	51	329	90	43	
81	327	89	42	307	83	49	335	91	42	
85	335	90	40	311	84	48	343	92	38	
89	335	89	38	299	80	41	347	93	36	
93	340	89	35	303	79	36	345	90	31	
97	344	90	31	304	80	29	356	93	29	
101	337	88	25	309	81	26	351	92	28	
Mean for week	5									
1-13	241	98		238	97		235	96		
14-52	295	95		285	92		284	92		
53-101	331	91		305	84		332	91		

<sup>a</sup> Interim evaluations occurred during weeks 14, 31, and 53; until week 53, number of survivors includes 17 or 16 (3 ng/kg group) special study animals (except stop-exposure group).

#### **Thyroid Hormone Concentrations**

Assays for total thyroxine  $(T_4)$ , free  $T_4$ , total triiodothyronine  $(T_3)$ , and thyroid stimulating hormone (TSH) were conducted at the 14-, 31-, and 53-week interim evaluations (Table 10). A downward trend in serum total T<sub>4</sub> concentrations with higher TCDD concentrations was evident at 14, 31, and 53 weeks. At 14, 31, and 53 weeks, all dosed groups showed a decrease in mean total T<sub>4</sub> concentrations relative to the vehicle controls. At 14 weeks, the  $T_4$  concentrations in the 22, 46, and 100 ng/kg groups were significantly lower than in vehicle controls by 25.1%, 28.1%, and 31.7%, respectively. At 31 weeks, total  $T_4$  concentrations in the 22, 46, and 100 ng/kg groups were significantly lower than those in the vehicle controls. These decreases were 34.4%, 33.9%, and 21.8% lower relative to vehicle controls in the 22, 46, and 100 ng/kg groups, respectively. At 53 weeks, significant depressions in mean total  $T_{4}$  concentrations relative to vehicle controls of 46.3% and 32.7% were observed in the 46 and 100 ng/kg groups, respectively.

At 14 weeks, a significant decrease in serum free  $T_4$  concentrations relative to the vehicle controls was observed at doses of 22 ng/kg or greater with the high dose group showing the greatest change (25.0%). At 31 weeks, serum free  $T_4$  concentrations were lower at doses greater than 10 ng/kg compared to vehicle controls. These changes were significant for the 22, 46, and 100 ng/kg groups, which were 25.1%, 16.0%, and 26.8% lower than vehicle controls, respectively. At the 53-week interim evaluation, no significant differences in free  $T_4$  concentrations were observed between vehicle control and TCDD-treated groups, although there was a slight trend toward lower levels at the higher doses.

There was a trend for serum concentrations of  $T_3$  to increase with increasing dose in the 10 ng/kg or greater groups at 14 weeks. In the 46 and 100 ng/kg groups, these increases were significantly higher than vehicle controls by 18.3% and 33.0%, respectively. At 31 weeks, serum  $T_3$  was significantly higher in the 46 and 100 ng/kg groups than in the vehicle controls by 30.0% and 22.6%, respectively. At the 53-week interim evaluation, an increasing trend in serum T<sub>3</sub> with higher TCDD concentrations was observed. These increases were dose-dependent and significant for the 10, 22, 46, and 100 ng/kg groups. Serum T<sub>3</sub> concentrations were 13.8%, 23.6%, 27.9%, and 35.4% higher than vehicle controls in the 10, 22, 46, and 100 ng/kg groups, respectively.

There was a trend toward increased serum TSH levels at doses of 10 ng/kg or greater at 14 weeks and 3 ng/kg or greater at 31 weeks. At 14 weeks, serum TSH levels were significantly higher in the 46 and 100 ng/kg groups relative to vehicle controls. No significant differences were observed in serum TSH levels at 31 or 53 weeks. At the 53-week interim evaluation, the mean serum TSH level in the 100 ng/kg group was 43.5% higher, but not significantly, than the vehicle controls.

	Vehicle Control	3 ng/kg	10 ng/kg	22 ng/kg	46 ng/kg	100 ng/kg
Week 14						
n	10	10	10	10	10	10
Total $T_{4}$ (µg/dL)	$6.370 \pm 0.372$	$5.940 \pm 0.321$	$5.480 \pm 0.371$	$4.770 \pm 0.310$ **	$4.580 \pm 0.309 **$	$4.350 \pm 0.289 **$
Free $T_4$ (ng/dL)	$2.478 \pm 0.175$	$2.404 \pm 0.130$	$2.513 \pm 0.242$	$1.983 \pm 0.144*$	$2.040 \pm 0.130 *$	$1.858 \pm 0.131 *$
Total $T_2$ (ng/dL)	$155.685 \pm 9.739$	$149.141 \pm 4.180$	$163.995 \pm 4.995$	$159.066 \pm 4.888$	$184.171 \pm 6.961*$	$207.026 \pm 6.149 **$
TSH (ng/mL)	$14.981 \pm 0.949$	$13.282 \pm 0.718$	$16.018 \pm 1.047$	$17.434 \pm 0.737$	$19.765 \pm 1.587 *$	$18.836 \pm 1.656 *$
Week 31						
n	10	10	10	10	10	10
Total T <sub>4</sub> ( $\mu$ g/dL)	$3.810 \pm 0.414$	$3.650 \pm 0.201$	$3.260 \pm 0.178$	$2.500 \pm 0.153 **$	$2.520 \pm 0.237 **$	$2.980 \pm 0.205 **$
Free $T_4$ (ng/dL)	$1.734 \pm 0.171$	$1.750 \pm 0.094$	$1.573 \pm 0.105$	$1.299 \pm 0.072 **$	$1.457 \pm 0.124*$	$1.269 \pm 0.080 **$
Total $T_2$ (ng/dL)	$138.281 \pm 4.658$	$134.433 \pm 6.867$	$137.681 \pm 4.858$	$142.253 \pm 3.073$	$179.900 \pm 7.440 **$	$169.465 \pm 8.805 **$
TSH (ng/mL)	$12.321 \pm 0.552^{b}$	$14.828 \pm 1.008$	$15.740 \pm 0.864$	$13.654 \pm 0.998$	$16.640 \pm 1.988$	$16.172 \pm 1.685$
Week 53						
n	8	8	8	8	8	8
Total T <sub>4</sub> ( $\mu$ g/dL)	$3.213 \pm 0.236$	$3.038\pm0.204$	$2.550 \pm 0.200$	$2.800 \pm 0.089$	$1.725 \pm 0.161 **$	$2.163 \pm 0.167 **$
Free $T_{4}$ (ng/dL)	$1.553 \pm 0.153$	$1.593 \pm 0.149$	$1.441 \pm 0.109$	$1.781 \pm 0.149$	$1.221 \pm 0.144$	$1.329 \pm 0.076$
Total $\vec{T}_{2}$ (ng/dL)	$129.815 \pm 3.540$	$134.334 \pm 7.868$	$147.760 \pm 3.393 **$	$160.403 \pm 7.226 **$	$166.051 \pm 8.173 **$	175.767 ± 12.596**
TSH (ng/mL)	$13.193 \pm 1.107$	$12.562 \pm 0.902$	$12.390 \pm 0.936$	$12.113 \pm 0.939$	$13.318\pm1.251$	$18.930\pm1.985$

#### TABLE 10 Serum Concentrations of Thyroid Hormones in Female Rats at the 14-, 31-, and 53-Week Interim Evaluations in the 2-Year Gavage Study of TCDD<sup>a</sup>

\* Significantly different (P $\le 0.05$ ) from the vehicle control group by Dunn's or Shirley's test \*\* P $\le 0.01$ Data are presented as mean  $\pm$  standard error. T<sub>4</sub>=thyroxine; T<sub>3</sub>=triiodothyronine; TSH=thyroid stimulating hormone n=9

#### Hepatic Cell Proliferation Data

Hepatocellular proliferation data from the 14-, 31-, and 53-week interim evaluations are shown in Table 11. The consumption of the BrdU drinking water solutions prior to each interim evaluation was similar across groups. Labeling indexes in controls varied from 0.327 to 1.410. Treatment-related effects in dosed groups were observed relative to duration-of-exposure-matched vehicle controls. At 14 weeks, the hepatocellular labeling index was significantly higher in the 22 ng/kg group compared to the vehicle controls. A 2-fold increase in the 22 ng/kg group was observed relative to vehicle controls. The labeling index was significantly higher in all dose groups at the 31-week interim evaluation than in the

vehicle controls. A 2- to 3-fold increase was observed at doses of 3, 10, and 22 ng/kg compared to the vehicle controls. The hepatocyte labeling index was 4-fold higher in the 46 ng/kg group and 11-fold higher in the 100 ng/kg group compared to vehicle controls. At 53 weeks, the labeling index was elevated by nearly 3-fold at 10 ng/kg; however, this increase was not statistically different than vehicle controls due to a large variance in response. The labeling index was significantly higher in the 46 and 100 ng/kg groups than in vehicle controls at the 53-week interim evaluation. The labeling index was nearly 2-fold higher at 46 ng/kg and greater than 5-fold higher at 100 ng/kg relative to vehicle controls.

 TABLE 11

 Hepatic Cell Proliferation Data for Female Rats

 at the 14-, 31-, and 53-Week Interim Evaluations in the 2-Year Gavage Study of TCDD<sup>a</sup>

	Vehicle Control	3 ng/kg	10 ng/kg	22 ng/kg	46 ng/kg	100 ng/kg
		0.0	5.5	5.5	0.0	
n						
Week 14	10	10	10	10	10	10
Week 31	9	10	10	10	10	10
Week 53	8	8	8	8	8	8
Labeling index (%)						
Week 14	$1.410\pm0.271$	$1.675 \pm 0.471$	$1.916 \pm 0.251$	$2.858 \pm 0.395 *$	$2.405\pm0.485$	$1.694 \pm 0.313$
Week 31	$0.327 \pm 0.063$	$0.852 \pm 0.206 *$	$0.956 \pm 0.233*$	$0.792 \pm 0.146 *$	$1.333 \pm 0.355 **$	$3.846 \pm 0.974 **$
Week 53	$0.948\pm0.190$	$0.976\pm0.121$	$2.736 \pm 1.159$	$1.289\pm0.192$	$1.844 \pm 0.293*$	$4.896 \pm 0.883 **$

\* Significantly different (P≤0.05) from the vehicle control group by Shirley's test

\*\* P≤0.01

<sup>a</sup> Data are presented as mean  $\pm$  standard error.

#### Cytochrome P450 Enzyme Activities

At each interim evaluation, liver and lung samples were collected for determinations of P450 enzyme activity. Microsomal suspensions were prepared from liver samples and were assayed for 7-ethoxyresorufin-*O*-deethylase (EROD, CYP1A1) activity, 7-pentoxyresorufin-*O*-deethylase (PROD, CYP2B) activity, and acetanilide-4-hydroxylase (A4H, CYP1A2) activity. Microsomal samples from lung were analyzed for EROD activity only (Table 12).

Hepatic EROD, PROD, and A4H activities were significantly higher in all groups administered TCDD relative to the vehicle controls at the 14-, 31-, and 53-week interim evaluations. Significant induction of hepatic P450 activities occurred at the lowest dose (3 ng/kg) for all three sampling times (14, 31, and 53 weeks). Hepatic EROD activities were maximally induced approximately 70- and 62-fold over vehicle controls in the 100 ng/kg group at 14 and 53 weeks, respectively, and 82-fold over vehicle controls in the 46 ng/kg dose group at 31 weeks. Similarly, hepatic PROD activity generally increased with increasing dose and was significantly

higher in all dosed groups relative to vehicle controls at all three interim evaluations (14, 31, and 53 weeks). Hepatic PROD activities were maximally induced approximately 8- and 6-fold over vehicle controls in the 100 ng/kg group at 14 and 53 weeks, respectively, and 8.5-fold over vehicle controls in the 46 ng/kg group at 31 weeks. Hepatic A4H activity generally increased with increasing dose. Hepatic A4H activities were maximally induced approximately 6- and 4.5-fold over vehicle controls in the 100 ng/kg group at 14 and 53 weeks, respectively, and 5.5-fold over vehicle controls in the 46 ng/kg group at 31 weeks.

Pulmonary EROD activities were approximately 10-fold lower than hepatic EROD activities in the vehicle control groups. EROD activity in the lung was significantly higher in all dosed groups compared to vehicle controls at the 14-, 31-, and 53-week interim evaluations. The maximum degree of induction relative to the vehicle control group was approximately 20-fold at 14 weeks (100 ng/kg), 24-fold at 31 weeks (22, 46, and 100 ng/kg), and 14-fold (10, 46, and 100 ng/kg) at 53 weeks.

	Vehicle					
	Control	3 ng/kg	10 ng/kg	22 ng/kg	46 ng/kg	100 ng/kg
n						
Week 14	10	10	10	10	10	10
Week 31	10	10	10	10	10	10
Week 53	8	8	8	8	8	8
Liver Microso	mes					
Acetanilide-4-hyd	droxylase (A4H) (nmol/n	ninute per mg microsoma	al protein)			
Week 14	$0.448 \pm 0.017$	$0.787 \pm 0.052 **$	$1.004 \pm 0.078 **$	$1.391 \pm 0.092 **$	$2.340 \pm 0.122 **$	$2.660 \pm 0.087 **$
Week 31	$0.394 \pm 0.020$	$0.728 \pm 0.045 **$	$0.791 \pm 0.059 **$	$1.207 \pm 0.071$ **	$2.174 \pm 0.172 **$	$1.882 \pm 0.103 **$
Week 53	$0.531 \pm 0.023$	$0.882 \pm 0.032$ **	$1.484 \pm 0.073 **$	$1.711 \pm 0.133 **$	$2.193 \pm 0.179 **$	$2.385 \pm 0.141$ **
7-Ethoxyresorufi	n-O-deethylase (EROD)	(pmol/minute per mg mi	crosomal protein)			
Week 14	$30.523 \pm 1.475$	925.193 ± 80.785**	1,550.31 ± 116.116**	$1,\!819.68 \pm 108.267 {**}$	2,064.69 ± 134.623**	$2,129.73 \pm 88.569 **$
Week 31	$30.932 \pm 1.263$	$926.292 \pm 64.816 **$	$1,238.36 \pm 74.002 **$	$1,700.49 \pm 101.296 ^{**}$	2,548.92 ± 161.798**	2,074.85 ± 156.032**
Week 53	$30.215 \pm 1.590$	$569.377 \pm 24.622 **$	$1,280.00 \pm 95.303 **$	$1,551.16 \pm 112.364 ^{**}$	$1,726.81 \pm 107.575 **$	$1,\!871.47 \pm 109.140^{**}$
7-Pentoxyresoruf	in-O-deethylase (PROD)	(pmol/minute per mg m	icrosomal protein)			
Week 14	$3.784 \pm 0.246$	$9.143 \pm 0.514$ **	$16.571 \pm 0.843 **$	$23.772 \pm 0.960 **$	$27.346 \pm 1.443 **$	$30.914 \pm 0.847 \text{**}$
Week 31	$3.261 \pm 0.260$	$9.634 \pm 0.413 **$	$14.769 \pm 0.502 **$	$20.421 \pm 0.905 **$	$27.717 \pm 1.248 **$	$25.257 \pm 1.674 **$
Week 53	$3.614\pm0.172$	$7.271 \pm 0.197 **$	$14.760 \pm 0.569 **$	17.281 ± 0.562**	$20.581 \pm 1.078 **$	21.213 ± 1.352**
Lung Microso	mes					
7-Ethoxyresorufi	n-O-deethvlase (EROD)	(pmol/minute per mg mi	crosomal protein)			
Week 14	$3.004 \pm 0.509$	$27.396 \pm 1.602 **$	$38.483 \pm 2.463 **$	44.194 ± 3.493**	44.309 ± 3.794**	$62.214 \pm 4.899 **$
Week 31	$2.071 \pm 0.307$	$25.342 \pm 0.806^{**}$	$30.387 \pm 1.844 **$	$50.190 \pm 2.745 **$	$49.066 \pm 4.399 **$	$48.421 \pm 2.825 **$
Week 53	$3.011 \pm 0.560$	$27.149 \pm 1.863 **$	42.847 ± 3.943**	36.569 ± 4.591**	43.746 ± 6.558**	43.713 ± 2.235**

### TABLE 12 Liver and Lung Cytochrome P450 Data for Female Rats at the 14-, 31-, and 53-Week Interim Evaluations in the 2-Year Gavage Study of TCDD<sup>a</sup>

\*\* Significantly different (P $\le$ 0.01) from the vehicle control group by Shirley's test Data are presented as mean  $\pm$  standard error.

### Determinations of TCDD Concentrations in Tissues

Concentrations of TCDD were determined in liver, lung, fat, and blood at the 14-, 31-, and 53-week interim evaluations and at the end of the 2-year study (105 weeks) (Table 13). The highest concentrations of TCDD were observed in the liver, followed by fat tissue. In vehicle control liver, TCDD concentrations were below the limits of quantitation (113 pg/g) at 14, 31, and 105 weeks. At the 53-week interim evaluation, liver concentrations of TCDD in vehicle control liver were 249 pg/g. Hepatic concentrations increased with increasing doses of TCDD. The highest hepatic concentrations were observed in the 100 ng/kg group with 18,280, 21,180, 15,763, and 9,325 pg/g at 14, 31, 53, and 105 weeks, respectively. In the liver tissue from the stop-exposure group, TCDD concentrations were 737 pg/g, which was slightly higher than levels observed in the 3 ng/kg group (681 pg/g).

In fat tissue of controls, TCDD concentrations were below the experimental limit of quantitation (200 pg/g)

at all of the interim evaluations. In treated groups, fat concentrations increased with increasing doses of TCDD. The highest concentrations were observed in the 100 ng/kg group with 6,395, 7,848, 5,816, and 3,177 pg/g at 14, 31, 53, and 105 weeks, respectively. In the fat tissue of the stop-exposure group, TCDD concentrations were below the limits of quantitation.

No measurable concentrations of TCDD were observed in blood from vehicle control or treated rats at any of the interim evaluations. No measurable concentrations of TCDD were observed in the lung at 14, 53, or 105 weeks. At 31 weeks, TCDD concentrations of 10,500 and 963 pg/g were observed in the lungs of single rats in the 10 and 46 ng/kg groups, respectively. Given the high levels, this may have been due to accidental inhalation of dosing material during gavage. No measurable concentrations of TCDD were observed in the lungs of rats from the other TCDD-treated groups or the vehicle control group.

	Vehicle Control	3 ng/kg	10 ng/kg	22 ng/kg	46 ng/kg	100 ng/kg	100 ng/kg (Stop- Exposure)
n							
Week 14	10	10	10	10	10	10	
Week 31	10	10	10	10	10	10	
Week 53	8	8	8	8	8	8	
Week 105	10	8	10	10	10	10	10
Fat							
Week 14	BLOQ	$295.33 \pm 27.31^{b}$	$708.60 \pm 45.87$	$1,274.80 \pm 120.20$	$2,620.00 \pm 131.39$	$6,395.00 \pm 390.03$	
Week 31	BLOQ	$378.10 \pm 48.00$	$767.50 \pm 23.74$	$1,572.00 \pm 72.00$	$3,089.00 \pm 140.98$	$7,848.00 \pm 554.10$	
Week 53	BLOQ	$304.29 \pm 26.95^{\circ}$	$633.50 \pm 41.03$	$1,\!272.63\pm 66.08$	$2{,}502.50 \pm 126.25$	$5{,}816.25 \pm 417.12$	
Week 105	BLOQ	$505.38\pm91.94$	$753.00 \pm 67.60^{\text{D}}$	$1,\!403.80 \pm 117.01$	$1,996.00 \pm 122.68$	$3,177.00 \pm 505.59$	BLOQ
Liver							
Week 14	BLOQ	$676.20 \pm 23.37$	$2,166.67 \pm 85.52^{b}$	$4,\!931.00\pm212.01$	$9,\!970.00\pm675.44$	$18{,}280.00\pm805.37$	
Week 31	BLOQ,	$598.90 \pm 28.62$	$2,079.00 \pm 130.82$	$4,\!489.00 \pm 193.12$	$9,522.00 \pm 334.03$	$21,180.00 \pm 817.01$	
Week 53	249.00 <sup>a</sup>	$499.38 \pm 27.15$	$1,940.00 \pm 124.57$	$4,\!437.50 \pm 132.07$	$7,818.75 \pm 367.21$	$15,\!762.50\pm437.09$	
Week 105	BLOQ	$680.71 \pm 57.95^{\circ}$	2,213.00 ± 163.28	$4,364.00\pm 267.99$	6,413.00 ± 241.93	9,325.00 ± 1,163.11	$737.00 \pm 270.04^{e}$
Lung							
Week 14	BLOQ	BLOQ	BLOQ,	BLOQ	BLOQ	BLOQ	
Week 31	BLOQ	BLOQ	10,500 <sup>°d</sup>	BLOQ	963 <sup>d</sup>	BLOQ	
Week 53	BLOQ	BLOQ	BLOQ	BLOQ	BLOQ	BLOQ	
Week 105	BLOQ	BLOQ	BLOQ	BLOQ	BLOQ	BLOQ	BLOQ
Blood							
Week 14	NA	NA	NA	NA	BLOQ	BLOQ	
Week 31	NA	NA	NA	NA	NA	BLOQ	
Week 53	NA	NA	NA	NA	NA	NA	
Week 105	NA	NA	NA	NA	NA	NA	NA

TABLE 13
Tissue Concentrations of TCDD in Female Rats in the 2-Year Gavage Study of TCDD <sup>a</sup>

Data are given in pg/g tissue (fat, liver, lung) or pg/mL (blood) as the mean  $\pm$  standard error. Mean values do not include values that were below the experimental limit of quantitation. NA=not analyzed. BLOQ=below the limit of quantitation; LOQ<sub>fat</sub>=200 pg/g, LOQ<sub>liver</sub>=113 pg/g, LOQ<sub>lung</sub>=500 pg/g, а LOQ<sub>blood</sub>=103 pg/mL.

b n=9

c d n=7

n=1e

n=5

#### **Pathology and Statistical Analyses**

This section describes the statistically significant or biologically noteworthy changes in the incidences of neoplasms and/or nonneoplastic lesions of the liver, lung, oral mucosa, uterus, pancreas, thymus, adrenal cortex, heart, clitoral gland, ovary, kidney, forestomach, bone marrow, mesentery, mandibular lymph node, thyroid gland, mammary gland, and pituitary gland. Summaries of the incidences of neoplasms and nonneoplastic lesions, individual animal tumor diagnoses, statistical analyses of primary neoplasms that occurred with an incidence of at least 5% in at least one animal group, and historical incidences for the neoplasms mentioned in this section are presented in Appendix A.

*Liver:* Liver weights were significantly increased in all dosed groups at 14 and 31 weeks (Table C1). At 53 weeks, relative liver weights of rats administered 10 ng/kg or greater were significantly increased as was the absolute liver weight of 100 ng/kg rats. At the 14-week interim evaluation, changes in the liver consisted of increased incidences and/or severities of hepatocellular hypertrophy in groups administered 10 ng/kg or greater (Tables 14 and A5a); the increased liver weight. Multinucleated hepatocytes and diffuse fatty change were seen in 100 ng/kg rats.

At 31 weeks, hepatocellular hypertrophy occurred in all dosed groups and tended to correlate with increased liver weight (Tables 14 and A5a). Dose-related increased incidences of multinucleated hepatocytes were seen in the 46 and 100 ng/kg groups. Increased incidences of pigmentation occurred in rats administered 10 ng/kg or greater, and the severity was increased in the 100 ng/kg group. Mixed cell focus, toxic hepatopathy, and diffuse fatty change were significantly increased in the 100 ng/kg group. The severities of inflammation were increased in all dosed groups.

At the 53-week interim evaluation, hepatocellular hypertrophy occurred in all dosed groups and tended to correlate with increased liver weight (Tables 14 and A5a). The incidences and severities of multinucleated hepatocytes and bile duct hyperplasia were increased in the 100 ng/kg group. The incidences of pigmentation were increased in rats administered 10 ng/kg or greater, and the severity of this lesion increased with increasing dose. Increased incidences of multiple mixed cell focus occurred in the 46 and 100 ng/kg groups. Dose-related increased incidences of toxic hepatopathy and diffuse fatty change occurred in the 46 and 100 ng/kg groups. The severities of inflammation were increased in rats administered 22 ng/kg or greater. Cholangiofibrosis and bile duct fibrosis were seen in the 46 and 100 ng/kg groups.

At 2 years, the incidence of hepatocellular adenoma was significantly increased in the 100 ng/kg core study group and exceeded the historical vehicle control range (Tables 15, A3a, and A4a). Dose-related increased incidences of cholangiocarcinoma were seen in the core study rats administered 22 ng/kg or greater. The highest incidence of cholangiocarcinoma was seen in the 100 ng/kg core study group and included a significant number of animals with multiple cholangiocarcinomas. No cholangiocarcinomas have occurred in the historical vehicle controls (Tables 15 and A4a). Two cholangiocarcinomas and two hepatocellular adenomas were seen in the 100 ng/kg stop-exposure group. Two hepatocholangiomas were seen in the 100 ng/kg core study group, and one cholangioma occurred in a 100 ng/kg stop-exposure rat (Tables 15, A1b, and A1c); no hepatocholangiomas or cholangiomas have occurred in the historical vehicle controls (Tables 15 and A4a). incidences of cholangiofibrosis were significantly increased in the 46 and 100 ng/kg core study groups, and the incidence in the 100 ng/kg stop-exposure group was significantly lower than the 100 ng/kg core study group (Tables 15, A5b, and A5c).

Hepatocellular adenoma was a nodular mass that usually was larger than a focus, had a distinct border, and produced more compression of surrounding normal parenchyma (Plate 1). Adenoma was composed of a rather uniform population of mildly to moderately pleomorphic hepatocytes that generally were normal in size or slightly larger than normal and were arranged in abnormal lobular patterns. The hepatic cords within an adenoma usually intersected the surrounding normal hepatic cords at an oblique angle or sometimes even at a right angle. A few small proliferating bile ducts or oval cells were sometimes seen but were not as numerous as in nodular hyperplasia. The uniform population of relatively normal sized, somewhat pleomorphic hepatocytes that were arranged in abnormal lobular patterns and the lack of proliferating bile ducts were important features differentiating adenoma from nodular hyperplasia.

#### TABLE 14 Incidences of Nonneoplastic Lesions of the Liver in Female Rats at the 14-, 31-, and 53-Week Interim Evaluations in the 2-Year Gavage Study of TCDD

	Vel Cor	hicle ntrol	3 n	g/kg	10 ng/	/kg	22 n	g/kg	46 n	g/kg	100 1	ng/kg
14-Week Interim Evaluation												
Number Examined Microscopically	10	h	10		10		10		10		10	
Hepatocyte, Hypertrophy"	1	$(1.0)^{\circ}$	1	(1.0)	4 (	1.3)	7**	(1.0)	10**	* (1.7)	10*	* (2.3)
Hepatocyte, Multinucleated Fatty Change, Diffuse	0 0		0 0		0 0		0 0		0 0		3 2	(1.0) (1.0)
												()
<b>31-Week Interim Evaluation</b>												
Number Examined Microscopically	10		10		10		10		10		10	
Hepatocyte, Hypertrophy	0		2	(1.0)	3 (	1.0)	6**	(1.2)	9*:	* (1.3)	10*	* (2.7)
Hepatocyte, Multinucleated	0		0		0		0		5*	(1.0)	9*	* (2.0)
Pigmentation Mixed Cell Focus	0		1	(1.0)	8** (	(1.0)	9**	(1.0)	10**	* (1.0)	10*	* (2.1)
(includes multiple)	3		7		4		2		4		10*	*
Toxic Henatonathy	0		0		0		0		0		10*	* (2 0)
Fatty Change Diffuse	Ő		Ő		Ő		Ő		Ő		6*	*(1.0)
Inflammation	9	(1.0)	9	(1 3)	10 (	13)	10	(1.5)	10	(1.5)	10	(1.8)
Oval Cell Hyperplasia		(110)	-	(110)	10 (	)	10	(110)	10	(1.0)	1	(1.0)
Bile Duct, Hyperplasia											1	(2.0)
53-Week Interim Evaluation												
Number Examined Microscopically	8		8		8		8		8		8	
Hepatocyte, Hypertrophy	0		4	(1.0)	7** (	1.1)	8**	(1.3)	8**	* (2.0)	8*	* (3.3)
Hepatocyte, Multinucleated	0		0	. /	0		0	. /	2	(1.0)	8*	* (1.6)
Pigmentation	0		0		5* (	1.0)	8**	(1.0)	8*:	* (1.8)	8*	* (2.6)
Mixed Cell Focus										. ,		
(includes multiple)	2		5		0		6		7*		8*	*
Toxic Hepatopathy	0		0		1 (	1.0)	0		6**	* (1.5)	8*	* (3.4)
Fatty Change, Diffuse	0		0		0		0		2	(1.0)	6*	* (1.7)
Bile Duct, Fibrosis	0		0		0		0		1	(1.0)	2	(2.0)
Bile Duct, Hyperplasia	0		0		1 (	1.0)	2	(1.0)	3	(1.3)	8*	* (1.9)
Inflammation	8	(1.0)	8	(1.0)	8 (	1.0)	8	(1.5)	8	(1.5)	8	(2.0)
Cholangiofibrosis	0		0		0		0		1	(3.0)	1	(2.0)

\* Significantly different (P≤0.05) from the vehicle control group by the Fisher exact test
 \*\* P≤0.01

 a Number of animals with lesion
 b Average severity grade of lesions in affected animals: 1=minimal, 2-mild, 3-moderate

Average severity grade of lesions in affected animals: 1=minimal, 2=mild, 3=moderate, 4=marked

	Vehicle Control	3 ng/kg	10 ng/kg	22 ng/kg	46 ng/kg	100 ng/kg	100 ng/kg (Stop- Exposure)
Number Examined							
Microscopically	53	54 b	53	53	53	53	50
Hepatocyte, Hypertrophy <sup>a</sup>	0	19** (1.2)	19** (1.2)	42** (1.5)	41** (2.0)	52** (3.2)	22**▲▲(1.7)
Hepatocyte, Multinucleated Eosinophilic Focus	0	0	16** (1.2)	26** (1.2)	36** (1.4)	51** (1.8)	32**▲▲(1.3)
(includes multiple)	11	14	21*	27**	27**	44**	27**▲
Inflammation	33 (1.1)	46* (1.2)	47** (1.1)	50** (1.4)	52** (1.2)	49** (1.3)	43** (1.2)
Pigmentation	4 (1.0)	9 (1.2)	34** (1.4)	48** (1.7)	52** (2.0)	53** (2.2)	45** (1.8)
Fatty Change, Diffuse	0	2 (1.0)	12** (1.1)	17** (1.3)	30** (1.2)	48** (1.9)	10**▲▲(1.4)
Necrosis	1 (2.0)	4 (2.3)	4 (2.0)	8* (2.3)	10** (1.9)	17** (2.5)	8* (1.8)
Oval Cell, Hyperplasia	0	4* (1.0)	3 (1.3)	20** (1.3)	38* (1.5)	53** (2.3)	1▲▲ (1.0)
Bile Duct, Hyperplasia	5 (1.6)	4 (1.5)	7 (1.1)	22** (1.3)	40** (1.4)	53** (2.1)	7▲▲ (1.3)
Bile Duct, Cyst	3 (3.3)	1 (3.0)	2 (2.0)	2 (1.5)	0	21** (2.9)	6 <sup>▲▲</sup> (2.3)
Hyperplasia, Nodular	0	0	0	3	7**	36**	0▲▲
Portal, Fibrosis	0	0	0	0	5* (1.8)	27** (2.0)	1 (2.0)
Toxic Hepatopathy	0	2 (1.0)	8** (1.3)	30** (1.2)	45** (1.8)	53** (3.5)	16**▲▲(1.3)
Cholangiofibrosis	1 (1.0)	1 (1.0)	2 (1.5)	1 (1.0)	11** (2.5)	31** (2.4)	1 (2.0)
Henatocellular Adenoma (ii	ncludes multiple	e) <sup>c</sup>					
Overall rate	0/53 (0%)	0/54 (0%)	0/53 (0%)	0/53 (0%)	1/53 (2%)	13/53 (25%)	2/50 (4%)
Adjusted rate <sup>e</sup>	0.0%	0.0%	0.0%	0.0%	2.6%	29.9%	5.2%
Terminal rate f	0/25 (0%)	0/21 (0%)	0/23 (0%)	0/19 (0%)	0/22 (0%)	7/21 (33%)	0/21(0%)
First incidence (days)	i				720	642	702
Poly-3 test <sup>g</sup>	P<0.001	j	_		P=0 497	P<0.001	P=0.233
Poly-3 test	1 -0.001				1 0.177	1 0.001	P=0.004N
Henatocholangioma	0	0	0	0	0	2	0
Cholangioma	0	0	0	0	0	0	1
Cholangiocarcinoma Multi	nle 0	0	0	ů	1	17**	1▲▲
Cholangiocaremonia, Multi		v	Ū	0	1	17	1
Cholangiocarcinoma (inclue	des multiple) <sup>k</sup>						
Overall rate	0/53 (0%)	0/54 (0%)	0/53 (0%)	1/53 (2%)	4/53 (8%)	25/53 (47%)	2/50 (4%)
Adjusted rate	0.0%	0.0%	0.0%	2.9%	10.3%	54.9%	5.2%
Terminal rate	0/25 (0%)	0/21 (0%)	0/23 (0%)	0/19 (0%)	4/22 (18%)	13/21 (62%)	1/21 (5%)
First incidence (days)	_ ` ´	_ ` ´	_ ` ´	529	731 (T)	610	729
Poly-3 test	P<0.001	_	_	P=0.474	P=0.057	P<0.001	P=0.232
Poly-3 test							P<0.001N

### TABLE 15Incidences of Neoplasms and Nonneoplastic Lesions of the Liver in Female Ratsin the 2-Year Gavage Study of TCDD

\* Significantly different (P≤0.05) from the vehicle control group by the Poly-3 test

\*\* P≤0.01

▲ Significantly different (P≤0.05) from the 100 ng/kg core study group by the Poly-3 test

▲▲ P≤0.01

(T) Terminal sacrifice

<sup>a</sup> Number of animals with lesion

<sup>b</sup> Average severity grade of lesions in affected animals: 1=minimal, 2=mild, 3=moderate, 4=marked

Historical incidence for 2 year gavage studies with Sprague-Dawley vehicle control groups: 4/371, range 0%-4%

<sup>d</sup> Number of animals with neoplasm per number of animals with liver examined microscopically

e Poly-3 estimated neoplasm incidence after adjustment for intercurrent mortality

<sup>1</sup> Observed incidence at terminal kill

<sup>g</sup> Beneath the vehicle control incidence is the P value associated with the trend test; the stop-exposure group is excluded from the trend test. Beneath the dosed group incidence are the P values corresponding to pairwise comparisons between the vehicle controls and that dosed group. The Poly-3 test accounts for differential mortality in animals that do not reach terminal sacrifice.

Pairwise comparison between the 100 ng/kg core and stop-exposure groups. A lower incidence in the stop-exposure group is indicated by N.

Not applicable; no neoplasms in animal group

Value of statistic cannot be computed.

k Historical incidence: 0/371

Cholangiocarcinoma consisted of an irregular, relatively large, noncircumscribed lesion that replaced normal liver parenchyma (Plate 2). The lesion consisted of fibrous connective tissue stroma containing numerous atypical bile ducts, which frequently contained mucinous material and cellular debris. The epithelium forming the atypical bile ducts was often discontinuous, consisted of large atypical cells (Plate 3), and displayed degenerative changes. Mitotic figures and localized invasion of adjacent liver parenchyma were also observed.

Cholangiofibrosis appeared relatively small in size, was well demarcated, and did not show evidence of localized invasion (Plates 4 and 5).

Cholangioma was a well-demarcated mass consisting of multiple densely packed, irregular bile duct structures, some of which were moderately dilated, within a small amount of fibrous stroma. The bile ducts were composed of a single layer of densely packed, columnar, somewhat pleomorphic bile duct epithelial cells (Plates 6 and 7). Hepatocholangioma was composed of a mixture of proliferating hepatocellular and bile duct elements. Hepatocholangioma was a rather large, nodular mass with a distinct border that produced compression of surrounding normal parenchyma. The hepatocellular element appeared similar to that seen in hepatocellular adenoma and consisted of a rather uniform population of mildly to moderately pleomorphic hepatocytes that were generally normal sized or slightly larger than normal and were arranged in abnormal hepatic cords. Intermixed with the proliferating hepatocytes were numerous small to large biliary structures, surrounded by small amounts of dense fibrous tissue stroma, that appeared similar to the biliary structures seen within a cholangioma. The smaller biliary structures resembled proliferating small bile ducts while the large structures were generally irregular and sometimes moderately to markedly dilated. Some of the large structures became confluent, producing highly irregular cystic biliary structures that were incompletely separated by short septae projecting into the lumen. Some of the ductular lumens contained homogeneous, lightly eosinophilic material, but most were empty. The biliary structures were composed of a single layer of flattened to cuboidal to low columnar, somewhat pleomorphic, but otherwise relatively normal-appearing, bile duct epithelial cells.

At 2 years, the incidences of hepatocyte hypertrophy, multinucleated hepatocytes, eosinophilic focus, inflammation, pigmentation, diffuse fatty change, and toxic hepatopathy were significantly greater in all dosed groups administered 10 ng/kg or greater than in the vehicle controls, and the severity of these lesions generally increased with increasing dose in the core study groups (Tables 15, A5b, and A5c). Dose-related increased incidences of necrosis, oval cell hyperplasia, and bile duct hyperplasia occurred in core study rats administered 22 ng/kg or greater. The incidences of nodular hyperplasia and portal fibrosis in the 46 and 100 ng/kg core study groups and of bile duct cyst in the 100 ng/kg core study group were significantly increased. The incidences of nonneoplastic liver lesions were generally significantly decreased in the 100 ng/kg stop-exposure group compared to the 100 ng/kg core study group.

Hepatocyte hypertrophy was characterized by hepatocytes that were enlarged with increased amounts of eosinophilic cytoplasm. Minimal hypertrophy affected periportal hepatocytes, and as severity increased, hepatocytes in other areas of the hepatic lobule were also affected. The hypertrophy usually was not confined to periportal hepatocytes, and therefore, the general diagnosis of hepatocyte hypertrophy was used. Multinucleated hepatocytes were characterized by scattered hepatocytes that were enlarged and contained multiple (more than two and often four to six) nuclei. The presence of binucleated hepatocytes was not sufficient to make this diagnosis (Plate 8).

Eosinophilic, mixed, basophilic, and clear cell foci appeared similar and were characterized by a focus of hepatocytes with altered tinctorial properties. Eosinophilic focus was composed of cells with eosinophilic cytoplasm. Mixed cell focus was composed of a mixture of cells with different staining properties, generally a mixture of eosinophilic cells and cells with clear cytoplasm (clear cells). To be classified as an eosinophilic focus, at least 80% of the cells within the focus had to be eosinophilic cells. Otherwise, the focus was classified as a mixed cell focus. Basophilic focus consisted of hepatocytes with basophilic cytoplasm, occasionally with basophilic linear (tigroid) intracytoplasmic aggregates. Clear cell focus was composed of cells having clear cytoplasm. If two or more foci of a given type were present in a liver, it was diagnosed as multiple.

The treatment-related foci were of the eosinophilic and mixed cell type and often differed somewhat from those in control animals. Foci in controls consisted of hepatocytes that were generally somewhat larger than normal but appeared otherwise normal and were arranged in a relatively normal lobular pattern. The hepatic cords at the periphery of these foci generally merged imperceptibly with the surrounding normal liver, resulting in an indistinct border and little or no compression of the adjacent liver parenchyma. In contrast, foci in treated animals generally had a more definite border, the cords within the focus often were not smoothly continuous with those in the surrounding parenchyma, and the foci consisted of cells that were often prominently enlarged with abundant eosinophilic or clear vacuolated cytoplasm. If more than 20% of the cells were vacuolated, the focus was classified as the mixed cell type; otherwise it was classified as an eosinophilic focus. In addition, some larger foci caused variable degrees of compression of the surrounding hepatic parenchyma. The cells were arranged in a relatively normal lobular pattern, and foci sometimes contained large blood vessels and/or portal areas. The presence of proliferating bile ducts or oval cells was not considered characteristic of a focus. If proliferating bile ducts were present, this was considered indicative of nodular hyperplasia, described below.

Inflammation was generally minimal to mild, consisting of accumulation of mononuclear cells (predominantly lymphocytes and plasma cells with occasional macrophages) most often within portal areas but also sometimes scattered randomly throughout the liver. Pigmentation consisted of light brown to golden pigment present within macrophages and occasionally hepatocytes. The pigmented macrophages were often seen in portal areas but were also seen scattered randomly within the liver. The pigment was shown to stain positive for iron with Perl's stain. Diffuse fatty change was generally a minor change consisting of discrete clear vacuoles (consistent with lipid) in the cytoplasm of hepatocytes scattered diffusely throughout the liver. Necrosis consisted of scattered necrotic areas of hepatic parenchyma that were often randomly distributed but occasionally, in more severe cases, were distributed more diffusely. Oval cell hyperplasia consisted of small ovoid cells with basophilic cytoplasm and a round to ovoid nucleus that were arranged in single or double rows and located predominantly in the portal areas. Bile

duct hyperplasia consisted of increased numbers of portal bile ducts. Bile duct cyst was characterized by either single or multiple dilated bile ducts that were lined by attenuated epithelium (Plate 9). Portal fibrosis consisted of fibrous connective tissue accumulation that extended between adjacent portal areas.

Nodular hyperplasia was characterized by few to numerous, small to large nodular foci generally composed of hepatocytes that were considerably larger than normal hepatocytes (hepatocyte hypertrophy) sometimes mixed with areas of increased numbers of small hepatocytes (hepatocyte hyperplasia) (Plate 10). Areas of nodular hyperplasia sometimes blended with the surrounding parenchyma, although they often had a distinct border. Large focal to multifocal areas of nodular hyperplasia were sometimes seen that caused compression of surrounding tissue and/or bulging of the capsular surface. The cells within nodular hyperplasia generally were very large, larger than cells seen within adenomas and usually larger than cells seen within foci, with abundant eosinophilic cytoplasm and often with variable degrees of cytoplasmic vacuolization. In a few areas of nodular hyperplasia, however, the cells were of more normal size or sometimes slightly smaller than normal. The cells appeared to be arranged in normal cords, but the cells often were so large as to obscure the sinusoids between the cords, giving the appearance of solid sheets of hepatocytes. Biliary epithelium and portal areas were usually present within nodular hyperplasia. Blood vessels and/or central veins were also sometimes seen within areas of nodular hyperplasia, usually when hepatocytes were not so hypertrophic as to completely obscure the normal architecture. The presence of hypertrophic vacuolated hepatocytes together with proliferating biliary epithelium was considered to be useful in differentiating nodular hyperplasia (Plate 11) from hepatocellular adenoma (Plate 1).

Toxic hepatopathy included all nonneoplastic liver changes under one overall term. The severity of the toxic hepatopathy was graded in order to give an overall severity grade for the degree of toxicity in a liver. This was to allow for easier comparison of the degree of toxic change among different dosed groups than would be possible if the severities of all the individual nonneoplastic changes were compared among the different groups. This diagnosis was used in addition to, not instead of, any of the nonneoplastic diagnoses already made. The changes included under the diagnosis included focal cellular alteration, multinucleated hepatocytes, cystic degeneration, fatty change, inflammation, necrosis, pigmentation, nodular hyperplasia, bile duct cysts, bile duct hyperplasia, hepatocyte degeneration, hepatocyte hypertrophy, oval cell hyperplasia, and portal fibrosis. Some dosed animals occasionally had just a few of these changes present, but this was not considered to be sufficient liver involvement to warrant a diagnosis of toxic hepatopathy.

*Lung:* The absolute lung weights of 10 ng/kg rats at 14 and 31 weeks and of 46 ng/kg rats at 14 weeks were significantly increased; relative lung weights of all dosed groups at 31 weeks, of 100 ng/kg rats at 53 weeks, and of 10 ng/kg rats at 14 weeks were significantly increased (Table C1).

At 2 years, the incidence of cystic keratinizing epithelioma was significantly increased in the 100 ng/kg core study group, and multiple epitheliomas occurred in two animals in the 100 ng/kg core study group (Tables 16, A1b, and A3a). There were no epitheliomas in the 100 ng/kg stop-exposure group, and cystic keratinizing epitheliomas have not occurred in the historical vehicle controls (Tables 16 and A4b). The epitheliomas ranged from relatively small to very large lesions that replaced much of the normal lung parenchyma. They were cystic structures consisting of a highly irregular wall of highly keratinized stratified squamous epithelium and a center filled with keratin. The outer portion of the lesion grew by expansion into the adjacent lung, but evidence of invasion was not observed (Plates 12 and 13).

The incidences of bronchiolar metaplasia of the alveolar epithelium were significantly greater in all core study dosed groups and in the 100 ng/kg stop-exposure group than those in the vehicle control group; the incidence in the stop-exposure group was significantly less than that in the 100 ng/kg core study group (Tables 16, A5b, and A5c). Squamous metaplasia was present in the 46 and 100 ng/kg core study groups and in the 100 ng/kg stop-exposure group. Bronchiolar metaplasia of the alveolar epithelium consisted of replacement of the normal alveolar epithelium by cuboidal to columnar, sometimes ciliated cells and was often accompanied by abundant mucus production in the affected area (Plate 14). The lesion generally diffusely affected the epithelium located at the bronchiolar-alveolar junction and adjacent alveoli. The incidences of histiocytic infiltration were significantly increased in the 22, 46, and 100 ng/kg groups. Aggregates of large alveolar macrophages were sometimes present in areas of bronchiolar metaplasia. Bronchiolar metaplasia was differentiated from alveolar epithelial hyperplasia, which was seen only in the vehicle controls. In alveolar hyperplasia, the alveoli were lined by cuboidal epithelium and, unlike bronchiolar metaplasia in dosed animals, prominent mucus production was not observed, and very prominent inflammatory cell infiltrate, consisting of large aggregates of alveolar macrophages commonly mixed with focal aggregates of neutrophils, was usually associated with the affected areas.

Squamous metaplasia of the alveolar epithelium was generally a minor change consisting of one or more small, irregular foci of keratinizing stratified squamous epithelium that had replaced the normal alveolar epithelium (Plate 15).

Oral Mucosa: There was a positive trend in the incidence of gingival squamous cell carcinoma of the oral cavity, and the incidence in the 100 ng/kg group was significantly increased. In addition, the incidences in the 46 ng/kg and 100 ng/kg core study and stop-exposure groups exceeded the historical control range (Tables 17, A3a, A3b, and A4c). The incidences of gingival squamous hyperplasia were significantly increased in all dosed groups including the stop-exposure group (Tables 17, A5b, and A5c). Gingival squamous hyperplasia and squamous cell carcinoma, although reduced in incidences when compared to the 100 ng/kg core study group, were still present in the 100 ng/kg stop-exposure group. Squamous cell carcinoma occurred within the oral mucosa of the palate and was located adjacent to the incisor tooth in nasal section III. It was characterized by irregular cords and clusters of stratified squamous epithelial cells that invaded deep into the underlying connective tissue and often invaded the bone of the maxilla (Plate 16). Squamous hyperplasia was a focal lesion that occurred in the stratified squamous epithelium of the gingival oral mucosa adjacent to the molars in nasal section III. It consisted of varying degrees of thickening of the epithelium, often with the formation of epithelial rete pegs that extended a short distance into the underlying connective tissue. Ends of hair shafts and/or some degree of inflammation were often present in the areas of squamous hyperplasia, suggesting, at least in these cases, that the hyperplasia was secondary to the presence of the hair shafts and associated inflammation. It was unclear whether there was an association between squamous hyperplasia and squamous cell carcinoma.

	Veh Con	icle trol	3 ng/kg	10 ng/kg	22 ng/kg	46 ng/kg	100 ng/kg	100 ng/kg (Stop- Exposure)
Number Examined								
Microscopically	53		54	53	52	53	52	50
Alveolar Epithelium.								
Metaplasia, Bronchiolar <sup>a</sup>	2	$(1.0)^{b}$	19** (1.6)	33** (1.5)	35** (1.4)	45** (1.7)	46** (2.0)	31**▲(1.3)
Infiltration Cellular.								
Histiocyte	41	(1.9)	46 (1.8)	43 (1.8)	43* (1.6)	49* (1.8)	50* (2.0)	41 (2.0)
Metaplasia, Squamous	1	(1.0)	0	0	0	5 (2.2)	6 (2.2)	3 (1.3)
Alveolar Epithelium,						× /	× /	× /
Hyperplasia	12	(1.1)	0**	0**	0**	0**	0**	0**
Cystic Keratinizing Epitheli	oma.							
Multiple	0		0	0	0	0	2	0
Cystic Keratinizing Epitheli Overall rate Adjusted rate Terminal rate First incidence (days) Poly-3 test <sup>g</sup> Poly-3 test	oma ( 0/53 0.0% 0/25  P<0.	includes n (0%) (0%) 001	nultiple) <sup>c</sup> 0/54 (0%) 0.0% 0/21 (0%) j	0/53 (0%) 0.0% 0/23 (0%) 	0/52 (0%) 0.0% 0/19 (0%) 	0/53 (0%) 0.0% 0/22 (0%) 	9/52 (17%) 21.1% 4/21 (19%) 610 P=0.002	0/50 (0%) 0.0% 0/21 (0%)  P=0.003N

# TABLE 16Incidences of Neoplasms and Nonneoplastic Lesions of the Lung in Female Ratsin the 2-Year Gavage Study of TCDD

\* Significantly different (P≤0.05) from the vehicle control group by the Poly-3 test

\*\*  $P \le 0.01$ 

▲ Significantly different (P≤0.05) from the 100 ng/kg core study group by the Poly-3 test

<sup>a</sup> Number of animals with lesion

Average severity grade of lesions in affected animals: 1=minimal, 2=mild, 3=moderate, 4=marked

Historical incidence for 2-year gavage studies with Sprague-Dawley vehicle control groups: 0/370

<sup>a</sup>Number of animals with neoplasm per number of animals with lung examined microscopically

e Poly-3 estimated neoplasm incidence after adjustment for intercurrent mortality

<sup>1</sup> Observed incidence at terminal kill

<sup>g</sup> Beneath the vehicle control incidence is the P value associated with the trend test; the stop-exposure group is excluded from the trend test. Beneath the dosed group incidence are the P values corresponding to pairwise comparisons between the vehicle controls and that dosed group. The Poly-3 test accounts for differential mortality in animals that do not reach terminal sacrifice.

Pairwise comparison between the 100 ng/kg core and stop-exposure groups. A lower incidence in the stop-exposure group is indicated by N.

<sup>1</sup> Not applicable; no neoplasms in animal group

<sup>j</sup> Value of statistic cannot be computed.

TABLE 17
Incidences of Neoplasms and Nonneoplastic Lesions of the Oral Mucosa in Female Rats
in the 2-Year Gavage Study of TCDD

	Vehicle Control	3 ng/kg	10 ng/kg	22 ng/kg	46 ng/kg	100 ng/kg	100 ng/kg (Stop- Exposure)
Number Necropsied Gingival, Hyperplasia,	53	54	53	53	53	53	50
Squamous Cell Carcinoma Overall rate Adjusted rate Terminal rate First incidence (days) Poly-3 test Poly-3 test	1/53 (2%) 2.5% 0/25 (0%) 366 P<0.001	2/54 (4%) 5.7% 1/21 (5%) 647 P=0.449	1/53 (2%) 2.6% 0/23 (0%) 578 P=0.755	0/53 (0%) 0.0% 0/19 (0%) 	4/53 (8%) 10.2% 1/22 (5%) 659 P=0.169	10/53 (19%) 22.0% 2/21 (10%) 546 P=0.007	5/50 (10%) 12.4% 0/21 (0%) 520 P=0.099 P=0.193N

\* Significantly different (P≤0.05) from the vehicle control group by the Poly-3 test

\*\* P≤0.01

<sup>a</sup> Number of animals with lesion

Average severity grade of lesions in affected animals: 1=minimal, 2=mild, 3=moderate, 4=marked

Historical incidence for 2-year gavage studies with Sprague-Dawley vehicle control groups: 4/371, range 0%-2%

Number of animals with neoplasm per number of animals necropsied

Poly-3 estimated neoplasm incidence after adjustment for intercurrent mortality

Observed incidence at terminal kill

<sup>g</sup> Beneath the vehicle control incidence is the P value associated with the trend test; the stop-exposure group is excluded from the trend test. Beneath the dosed group incidence are the P values corresponding to pairwise comparisons between the vehicle controls and that dosed group. The Poly-3 test accounts for differential mortality in animals that do not reach terminal sacrifice. A lower incidence in a dosed group is indicated by N.

Pairwise comparison between the 100 ng/kg core and stop-exposure groups. A lower incidence in the stop-exposure group is indicated by N.

<sup>1</sup> Not applicable; no neoplasms in animal group

*Uterus:* The incidence of squamous cell carcinoma in the 46 ng/kg group at 2 years was significantly greater than that in the vehicle controls, and there were two squamous cell carcinomas in the 100 ng/kg stop-exposure group (Tables 18, A3a, and A1c). No squamous cell carcinomas have occurred in the historical vehicle controls (Tables 18 and A4d). Squamous cell carcinoma occurred on the endometrial surface, often caused dilatation of the uterus, and was characterized by irregular cords and clusters of atypical stratified squamous epithelial cells that invaded the underlying myometrium (Plates 17 and 18). One squamous cell papilloma was seen in each of the vehicle control and 10 ng/kg groups.

At 2 years, the incidence of squamous metaplasia was significantly less in the 100 ng/kg core study group compared to vehicle controls and the 100 ng/kg stop-exposure group (Tables 18, A5b, and A5c). The significance of this finding is unclear. Squamous metaplasia was generally a minimal to mild, multifocal change consisting of tubular structures within the endometrium that were lined by stratified squamous epithelium.

*Pancreas:* At 31 and 53 weeks, the incidences of acinar cytoplasmic vacuolization were increased in the 100 ng/kg group. Chronic active inflammation and

50
33▲▲(2.1)
0
2/50 (4%)
5.2%
2/21 (10%)
731 (T)
P=0.232
P=0.225

### TABLE 18Incidences of Neoplasms and Nonneoplastic Lesions of the Uterus in Female Ratsin the 2-Year Gavage Study of TCDD

\*\* Significantly different (P≤0.01) from the vehicle control group by the Poly-3 test

▲ Significantly different (P≤0.01) from the 100 ng/kg core study group by the Poly-3 test

(T) Terminal sacrifice

a Number of animals with lesion

Average severity grade of lesions in affected animals: 1=minimal, 2=mild, 3=moderate, 4=marked

Historical incidence for 2-year gavage studies with Sprague-Dawley vehicle control groups: 1/371, range 0%-2%

Number of animals with neoplasm per number of animals necropsied

Poly-3 estimated neoplasm incidence after adjustment for intercurrent mortality

<sup>1</sup> Observed incidence at terminal kill

<sup>g</sup> Beneath the vehicle control incidence is the P value associated with the trend test; the stop-exposure group is excluded from the trend test. Beneath the dosed group incidence are the P values corresponding to pairwise comparisons between the vehicle controls and that dosed group. The Poly-3 test accounts for differential mortality in animals that do not reach terminal sacrifice.

Pairwise comparison between the 100 ng/kg core and stop-exposure groups

Not applicable; no neoplasms in animal group

j Value of statistic cannot be computed.

acinar atrophy were present in 100 ng/kg rats at 14 and 53 weeks (Tables 19 and A5a).

At 2 years, one acinar adenoma and two acinar cell carcinomas were seen in the 100 ng/kg core study group (Tables 19 and A3a). The incidences of acinar cell carcinoma and adenoma or carcinoma (combined) exceeded the historical control range (Tables 19 and A4e). The incidences of acinar cytoplasmic vacuolization were significantly increased in the 46 and 100 ng/kg core study groups, and the incidences of chronic active inflammation, acinar atrophy, and arterial chronic active inflammation were significantly increased in the 100 ng/kg core study group. One acinar carcinoma was seen in a 100 ng/kg stop-exposure rat; the incidences of nonneoplastic lesions were not significantly different from the vehicle controls. The incidences of the nonneoplastic lesions in the stop-exposure group were less than those in the 100 ng/kg core study group, and the incidence of acinar cytoplasmic vacuolization in the stop-exposure group was significantly decreased compared to the 100 ng/kg core study group.

Adenoma of the acinar cells was characterized by a discrete mass of tubular and acinar structures composed of small acinar cells with brightly eosinophilic cytoplasm and lacking zymogen granules. Carcinoma was a large, multinodular lesion with moderate amounts of dense fibrous stroma. Carcinomas were composed of densely packed clusters of poorly formed acinar structures consisting of small acinar cells with prominent vesicular nuclei and small amounts of eosinophilic cytoplasm with indistinct borders. Scattered solid areas composed of densely packed, highly pleomorphic, round to ovoid acinar cells with large vesicular nuclei and scant cytoplasm were also seen.

# TABLE 19Incidences of Neoplasms and Nonneoplastic Lesions of the Pancreas in Female Ratsin the 2-Year Gavage Study of TCDD

	Vehicle Control	3 ng/kg	10 ng/kg	22 ng/kg	46 ng/kg	100 ng/kg
14-Week Interim Evaluation						
Number Examined Microscopically	10	10	10	10	10	10
Inflammation Chronic Active	0	0	0	0	0	$2(15)^{b}$
Acinus, Atrophy	0	0	0	0	0	2 (1.5)
31-Week Interim Evaluation						
Number Examined Microscopically Acinus, Vacuolization	10	10	10	10	10	10
Cytoplasmic	0	0	0	0	0	5* (1.0)
53-Week Interim Evaluation						
Number Examined Microscopically Acinus, Vacuolization	8	8	8	8	8	8
Cytoplasmic	0	0	0	0	0	7** (1.0)
Inflammation, Chronic Active	0	0	0	0	0	2 (2.0)
Acinus Atrophy	0	0	0	0	0	2 (2.0)

	Veh Con	icle trol	3 ng	g/kg	10 n	g/kg	22 n	g/kg	46 n	g/kg	100 n	ıg/kg	100 (St Expo	ng/kg op- osure)
2-Year Study														
Number Examined														
Microscopically	51		54		52		53		52		51		49	
Acinus, Vacuolization														
Cytoplasmic	1	(2.0)	0		0		1	(1.0)	15*	* (1.1)	42**	* (1.8)	0▲	<b>A</b>
Inflammation														
Chronic Active	0		0		2	(1.5)	2	(1.0)	3	(1.3)	6*	(2.0)	4	(1.5)
Acinus, Atrophy	1	(1.0)	2	(1.5)	4	(1.5)	4	(1.5)	4	(1.5)	9*	(2.2)	4	(1.8)
Artery, Inflammation,														
Chronic Active	0		1	(3.0)	1	(2.0)	2	(2.5)	2	(3.0)	7*	(2.3)	2	(2.5)
Acinus, Adenoma <sup>c</sup>	0		0		0		0		0		1		0	
Acinus, Carcinoma <sup>d</sup>	0		0		0		0		0		2		1	
Adenoma or Carcinoma <sup>c</sup> Overall rate <sup>e</sup> Adjusted rate <sup>g</sup> Terminal rate <sup>g</sup> First incidence (days) Poly-3 test <sup>i</sup> Poly-3 test	0/51 0.0% 0/25 j P<0.	(0%) (0%) 001	0/54 0.0% 0/21 k	(0%) 6 (0%)	0/52 0.0% 0/23 	(0%) % (0%)	0/53 0.0% 0/19 	(0%) 6 (0%)	0/52 0.09 0/22 	2 (0%) % 2 (0%)	3/51 7.3% 2/21 641 P=0.	(6%) 6 (10%) .129	1/49 2.79 1/21 731 P=0 P=0	9 (2%) % l (5%) (T) 9.494 9.345N

## TABLE 19Incidences of Neoplastic and Nonneoplastic Lesions of the Pancreas in Female Ratsin the 2-Year Gavage Study of TCDD

\* Significantly different ( $P \le 0.05$ ) from the vehicle control group by the Fisher exact test (interim evaluations) or the Poly-3 test (2-year study) \*\*  $P \le 0.01$ 

▲ Significantly different (P≤0.01) from the 100 ng/kg core study group by the Poly-3 test

(T) Terminal sacrifice

<sup>a</sup> Number of animals with lesion

Average severity grade of lesions in affected animals: 1=minimal, 2=mild, 3=moderate, 4=marked

<sup>c</sup> Historical incidence for 2-year gavage studies with Sprague-Dawley vehicle control groups: 1/366, range 0%-2%

<sup>d</sup> Historical incidence: 0/366

e Number of animals with neoplasm per number of animals with pancreas examined microscopically

Poly-3 estimated neoplasm incidence after adjustment for intercurrent mortality

<sup>g</sup> Observed incidence at terminal kill

Beneath the vehicle control incidence is the P value associated with the trend test; the stop-exposure group is excluded from the trend test.
 Beneath the dosed group incidence are the P values corresponding to pairwise comparisons between the vehicle controls and that dosed group. The Poly-3 test accounts for differential mortality in animals that do not reach terminal sacrifice.

<sup>1</sup> Pairwise comparison between the 100 ng/kg core and stop-exposure groups. A lower incidence in the stop-exposure group is indicated <sup>2</sup> by N.

 $\frac{J}{k}$  Not applicable; no neoplasms in animal group

<sup>K</sup> Value of statistic cannot be computed.

Atrophy was a focal to multifocal to diffuse change consisting of a reduction in the amount of acinar tissue with an associated increase in stromal fibrous connective tissue. Chronic active inflammation was generally associated with atrophy and consisted of an infiltrate of mononuclear cells with occasional neutrophils within the stroma. Cytoplasmic vacuolation consisted of small, clear, discrete intracytoplasmic vacuoles within pancreatic acinar cells. Sometimes these vacuoles coalesced to form larger single vacuoles. The severity of the change was determined by the degree of vacuolization per cell and the amount of tissue involved. Arterial chronic active inflammation was a focal to multifocal change characterized by a thick mantle of macrophages, lymphocytes, and plasma cells around the arteries, with infiltration into the muscular layers of the artery. There was often fibrinoid necrosis of the vessel, and the tunica intima was frequently thickened. Endothelial cells were swollen or decreased in number (Plate 19). This inflammatory reaction often extended into the surrounding parenchyma.

*Thymus:* The thymus weights of 46 and 100 ng/kg rats were significantly decreased at 14 weeks (Table C1). At 14, 31, and 53 weeks, the incidences of atrophy were significantly increased in the 46 and 100 ng/kg groups, and the severity was increased in the 100 ng/kg group (Tables 20 and A5a). At 2 years, atrophy occurred in all groups including the vehicle controls (Tables 20, A5b, and A5c) and was significantly increased in the 22 ng/kg and greater groups. Severity was greatest in the 100 ng/kg core study group. The incidence and severity of atrophy in the 100 ng/kg stop-exposure group were greater than those in the vehicle controls, which suggested a treatment-related effect. Atrophy consisted to varying degrees of loss of lymphoid cells from the cortex resulting in reduction of cortical thickness.

*Adrenal Gland:* Treatment-related changes included increased incidences of cortical atrophy and hyperplasia in core study groups administered 10 ng/kg or greater. The cortical atrophy was significant in the 100 ng/kg group, while hyperplasia was significant in the 22, 46, and 100 ng/kg groups. The incidences of cytoplasmic vacuolization were increased in the 22 ng/kg or greater groups (Tables 21, A5b, and A5c). The incidences of these lesions in the 100 ng/kg stop-exposure group were less than those in the 100 ng/kg core study group, and the difference in the incidence of cortical atrophy was significant. Cortical cystic degeneration was seen in all groups, including controls, the incidence being higher in treated groups, and was significantly increased in the 10 and 22 ng/kg groups.

Cortical atrophy was a locally extensive to diffuse change characterized by loss of cortical epithelial cells within the zona fasciculata and zona reticularis with a subsequent reduction in cortical thickness. The zona glomerulosa was spared. The remaining cells were sometimes vacuolated, especially in the more severe lesions. In severe cases, the entire cortex was considerably reduced in thickness, resulting in a smaller gland that often was surrounded by a thickened capsule. Cortical hyperplasia was a focal to multifocal change, generally located in the zona fasciculata, consisting of a discrete area containing increased numbers of cortical cells. The hyperplastic cells were the same size as or somewhat smaller than surrounding normal cortical cells and had slightly basophilic cytoplasm. In some cases, especially with large lesions, there was compression of the surrounding tissue. However, these were distinguishable as hyperplasia by the fact that the cells still formed normal cords, particularly in the upper zona fasciculata. Cortical cytoplasmic vacuolation was a focal to multifocal to diffuse change consisting of small, discrete, clear intracytoplasmic vacuoles. Sometimes the cytoplasm contained a large, single vacuole that displaced the nucleus. The changes were morphologically consistent with the accumulation of lipid.

Heart: At 2 years, significantly increased incidences of minimal to mild cardiomyopathy were seen in rats administered 10 ng/kg or greater (Tables 21, A5b, and A5c). The incidence of cardiomyopathy was significantly lower in the 100 ng/kg stop-exposure group compared to the 100 ng/kg core study group but was significantly greater than that in the vehicle controls. Cardiomyopathy had the typical microscopic appearance of this lesion seen in aging rats and appeared similar to the cardiomyopathy seen in aging F344/N rats (MacKenzie and Alison, 1990). It was a multifocal, generally minimal to mild lesion consisting of hypereosinophilic myofibers that lacked cross striations, infiltrates of mononuclear cells, separation of myofibers by myxomatous material (bluish material on H&E stain), and eventually replacement of myofibers by fibrous connective tissue. The severity was graded based upon the number and extent of foci of myocardial degeneration. Minimal cardiomyopathy consisted of a few scattered foci while mild cardiomyopathy consisted of a greater number of lesions more diffusely scattered within the myocardium.

*Clitoral Gland and Ovary:* The incidences of cystic dilatation of the clitoral gland ducts were significantly increased in 22 and 100 ng/kg core study rats at 2 years (Tables 21, A5b, and A5c); the incidence of this lesion

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Incidences of Nonneoplastic Lesions of the Thymus in Female Rats in the 2-Year Gavage Study of TCDD

	Vel Coi	hicle ntrol	3 n	g/kg	10 r	ıg/kg	22 r	ıg/kg	46 ng/kg	100 ng/kg
14-Week Interim Evaluation Number Examined Microscopically	10	, , , b	10		10		10		10	10
Atrophy	91	(1.0)	0		0		4*	(1.3)	10** (1.6)	
<b>31-Week Interim Evaluation</b> Number Examined Microscopically Atrophy	10 2	(2.0)	10 1	(1.0)	10 5	(1.6)	10 6	(1.8)	10 7* (2.0)	10 10** (3.1)
53-Week Interim Evaluation										
Number Examined Microscopically Atrophy	8 4	(2.0)	8 2	(2.0)	7 3	(2.3)	8 7	(2.3)	8 8* (3.4)	8 8* (3.4)
Ve Co	nicle ntrol	3	ng/kg	10 n	g/kg	22 ng/k	۲g	46 ng/kg	100 ng/kg	100 ng/kg (Stop- Exposure)
2-Year Study Number Examined Microscopically 51 Atrophy 36	(2.	5 6) 4	2 1 (2.7)	52 44	(3.0)	49 41* (	3.1)	46 44** (3.6)	42 42** (3.9)	49 45** (3.3)

\* Significantly different (P $\le$ 0.05) from the vehicle control group by the Fisher exact test (interim evaluations) or the Poly-3 test (2-year study) \*\* P $\le$ 0.01

b Number of animals with lesion

Average severity grade of lesions in affected animals: 1=minimal, 2=mild, 3=moderate, 4=marked

was significantly lower in the 100 ng/kg stop-exposure group compared to the 100 ng/kg core study group. This lesion consisted of dilated ducts that were filled with keratin and lined by attenuated epithelium. The severity varied from minimal to marked and was graded depending upon the size of the dilated ducts. Minimal lesions consisted of ducts dilated to approximately 2 to 3 mm, and marked lesions consisted of ducts dilated to approximately 1 cm or more in diameter. The incidence of ovarian atrophy was significantly lower in the 100 ng/kg core study group than in vehicle controls, and the incidence in the 100 ng/kg stop-exposure group was significantly higher compared to the 100 ng/kg core study group. Atrophy was characterized by absence of ovarian structures, primarily corpora lutea, but also by lack of follicles in some cases.

*Kidney:* At 2 years, the incidences of nephropathy were significantly increased in the 100 ng/kg core study and stop-exposure groups (Tables 21, A5b, and A5c); the incidence of nephropathy was significantly lower in the stop-exposure group compared to the 100 ng/kg core

study group. Increased incidences of transitional epithelial hyperplasia occurred in the 46 and 100 ng/kg core study groups.

Nephropathy was generally a minimal to mild change, although sometimes moderate to marked nephropathy was seen. The appearance of this lesion was typical of that seen in aging rats and was similar to that observed in F344/N rats (Barthold, 1998). Nephropathy was characterized by scattered foci of regenerative tubules lined by basophilic epithelium and sometimes surrounded by increased basement membrane, dilated tubules filled with proteinaceous casts and surrounded by fibrous connective tissue, and scattered foci of mixed inflammatory cells. Severity was graded based upon the number and extent of changes described previously. Minimal nephropathy was characterized by small numbers of scattered affected tubules, usually involving less than 10% of the renal tubules. However, marked nephropathy involved approximately 50% to 60% or more of the tubules.

#### TABLE 21

Incidences of Selected Nonneoplastic	Lesions in Female Rats in the 2-Year	Gavage Study of TCDD
--------------------------------------	--------------------------------------	----------------------

	Veh Con	icle trol	3 nş	g/kg	10 ng	g/kg	22 ng	g/kg	46 n	g/kg	100 n	g/kg	100 n (Sto Expo	ig/kg op- sure)
Adrenal Cortex <sup>a</sup>	53		54		53		53		53		53		50	
Atrophy <sup>b</sup>	2	$(2.5)^{c}$	0		4	(1.5)	5	(2.6)	5	(1.4)	27**	(2.4)	4▲4	(1.8)
Degeneration, Cystic	11	(2.5)	15	(2.1)	21*	(2.1)	18*	(2.3)	17	(2.2)	17	(2.3)	17	(2.2)
Hyperplasia	16	(2.0)	16	(2.2)	18	(2.4)	25*	(2.3)	29**	* (2.3)	30*	(2.5)	20	(2.6)
Vacuolization Cytoplasmic	11	(1.4)	7	(1.3)	12	(1.3)	21**	(1.6)	18	(1.7)	15	(2.1)	13	(1.3)
Heart	53		54		53		52		53		52		50	
Cardiomyopathy	10	(1.2)	12	(1.0)	22**	* (1.0)	25**	(1.1)	32**	* (1.1)	36**	(1.3)	22**	▲(1.4)
Mesentery Chronic Active Artery														
Inflammation	0		1	(3.0)	0		0		4	(2.3)	7	(2.7)	1	(3.0)
Clitoral Gland	50		52		53		52		51		53		49	
Duct, Cyst	34	(2.2)	37	(2.1)	41	(2.2)	42*	(2.0)	41	(2.2)	48**	(2.3)	35	(2.0)
Ovary	51		53		53		53		53		53		49	
Atrophy	49	(4.0)	46	(4.0)	50	(4.0)	44	(4.0)	50	(4.0)	31**	(3.9)	45	<b>(4.0)</b>
Kidney	53		54		53		53		53		53		50	
Nephropathy Transitional Epithelium	34	(1.2)	26	(1.1)	32	(1.3)	36	(1.4)	39	(1.4)	52**	(2.2)	41*	(1.4)
Hyperplasia	3	(2.3)	6	(1.7)	8	(1.8)	8	(1.3)	11*	(2.0)	11*	(2.0)	5	(1.4)
Stomach, Forestomach	53		54		53		53		53		53		50	
Hyperplasia, Squamous	3	(1.3)	4	(1.5)	4	(1.8)	2	(2.0)	7	(2.1)	11*	(1.7)	5	(1.8)
Bone Marrow	53		54		53		53		53		53		50	
Hyperplasia	36	(3.0)	41	(3.2)	32	(3.2)	35	(3.1)	37	(2.9)	43*	(2.8)	36	(2.9)
Lymph Node, Mandibular	51		54		52		50		51		52		49	
Ectasia	0		0		0		0		1	(2.0)	6*	(2.7)	2	(2.0)

 \* Significantly different (P≤0.05) from the vehicle control group by the Poly-3 test
 \*\* P≤0.01
 A Significantly different (P≤0.05) from the 100 ng/kg core study group by the Poly-3 test
 ▲ P≤0.01
 a Number of animals with tissue examined microscopically
 b Number of animals with lesion
 c Average severity grade of lesions in affected animals: 1=minimal, 2=mild, 3=moderate, Average severity grade of lesions in affected animals: 1=minimal, 2=mild, 3=moderate, 4=marked Transitional epithelium hyperplasia was sometimes focal to multifocal, but generally a diffuse, usually minimal to mild change consisting of varying degrees of thickening of the renal pelvic or papillary epithelium up to approximately 1.5 to 2 times the normal thickness. The significance of this change was unclear; it did not appear to correlate with the increased severity of nephropathy because the animals with hyperplasia often had minimal nephropathy.

*Forestomach:* The incidence of squamous hyperplasia was significantly increased in the 100 ng/kg core study group at 2 years (Tables 21, A5b, and A5c). Squamous hyperplasia of the forestomach epithelium was generally a minimal to mild, focal or occasionally multifocal change characterized by varying degrees of thickening of the stratified squamous epithelium up to approximately five times normal thickness in more severe cases. Sometimes the hyperplasia occurred around a focal ulcer, although most cases occurred without the presence of an apparent ulcer.

Bone Marrow: At 2 years, dose-related increased incidences of hyperplasia occurred in the 3 and 100 ng/kg groups, and the incidence was significantly increased in the 100 ng/kg group (Tables 21 and A5b). Bone marrow hyperplasia was graded as follows: grade 4 (marked) was used when the entire marrow cavity was filled with dense marrow. Grade 3 (moderate) hyperplasia was recorded when marrow elements composed about 90% of the cavity (the remaining 10% was fat). Grade 2 (mild) hyperplasia was recorded when marrow elements composed approximately 60% to 90% of the marrow cavity, and grade 1 (minimal) hyperplasia was rarely recorded because of the normal variation in the amount of bone marrow. Normal bone marrow was used when the distal end of the femur section contained 20% to 60% marrow.

*Mesentery:* Compared to the vehicle controls, the incidences of chronic active inflammation were increased in the 100 ng/kg core study group and the 100 ng/kg stop-exposure group.

*Mandibular Lymph Node:* The incidence of ectasia was significantly increased in the 100 ng/kg core study group at 2 years (Tables 21, A5b, and A5c). Ectasia consisted of mild to moderate, focal to multifocal dilatation of medullary sinuses (lymphangiectasis). The significance of this lesion was unclear.

*Thyroid Gland:* Dose-dependent increases in the incidences of follicular cell hypertrophy of the thyroid gland were observed at 14, 31, and 53 weeks and at 2 years

(Tables 22, A5a, A5b, and A5c). At 2 years, doserelated increased incidences of minimal follicular cell hypertrophy were noted in core study rats administered 22 ng/kg or greater, and the incidences were significantly increased in the 46 and 100 ng/kg groups. The incidence of this lesion was significantly lower in the 100 ng/kg stop-exposure group compared to the 100 ng/kg core study group. Hypertrophy was a localized to diffuse change characterized by follicles that were decreased in size and contained decreased amounts of colloid in which aggregates of amphophilic, flocculant appearing material were often present. The affected follicles were lined by large, prominent, cuboidal follicular epithelial cells that were approximately two to three times normal size, usually with abundant pale cytoplasm sometimes containing small, clear, resorption vacuoles. Since some degree of this change can occur spontaneously, the severity grade of minimal was recorded when 50% to 60% of the follicles were involved, mild severity when 60% to 75% of the follicles were involved, moderate when 75% to 90% of the follicles were involved, and marked when over 90% of the follicles were involved. The incidence of thyroid gland C-cell adenoma was significantly less in the 100 ng/kg core study group than in the vehicle controls (Tables 22 and A1b).

Mammary and Pituitary Glands: At 2 years, the incidences of mammary gland fibroadenoma [vehicle control, 33/53; 3 ng/kg, 40/54; 10 ng/kg, 34/53; 22 ng/kg, 29/53; 46 ng/kg, 36/53; 100 ng/kg (core study), 24/53; 100 ng/kg (stop-exposure), 32/50] and of adenoma or carcinoma (combined) (6/53, 8/54, 5/53, 2/53, 2/53, 1/53, 1/50) were significantly decreased in the 100 ng/kg core study group (Tables A3a and A3b). The incidence of pituitary gland (pars distalis or unspecified site) adenoma was significantly decreased in the 100 ng/kg core study group at 2 years (25/53, 20/54, 26/52, 15/53, 20/53, 11/52, 19/50). The incidences of mammary gland fibroadenoma and pituitary gland adenoma in the 100 ng/kg stop-exposure group were significantly greater than those in the 100 ng/kg core study group but not significantly different than the vehicle controls.

#### **GENETIC TOXICOLOGY**

TCDD (0.1 to 1,000  $\mu$ g/plate) was not mutagenic in *Salmonella typhimurium* strains TA98, TA100, TA1535, or TA1537 with or without hamster or rat liver metabolic activation enzymes (S9 fraction) (Table B1; Mortelmans *et al.*, 1984). No induction of trifluorothymidine resistance was noted in L5178Y tk<sup>+/-</sup> mouse lymphoma cells treated with 0.0625 to 1  $\mu$ g/mL TCDD

# TABLE 22Incidences of Neoplasms and Nonneoplastic Lesions of the Thyroid Gland in Female Ratsin the 2-Year Gavage Study of TCDD

		Vehicle Control		3 n	g/kg	10 n	ıg/kg	22	ng/kg	46 ng/kg	100 ng/kg
14-Week Interim Evalua Number Examined Microscop	<b>tion</b> vically	10		10		10		10		10	10
Follicular Cell, Hypertrophy <sup>a</sup>		0		1	(1.0) <sup>b</sup>	3	(3.0)	4	* (2.5)	4* (2.8)	9**(2.1)
<b>31-Week Interim Evalua</b> Number Examined Microscop	<b>tion</b> vically	10		10		10		10		10	10
Follicular Cell, Hypertrophy		0		3	(1.7)	3	(1.7)	3	(1.3)	4* (1.3)	6**(1.8)
53-Week Interim Evalua Number Examined Microscop Follicular Cell,	<b>tion</b> bically	10		10	(1.0)	10	(1.0)	10	(1.5)	10	10
	Veh Con	icle trol	3 ng	g/kg	10 n;	g/kg	22 ng	/kg	46 ng/kg	100 ng/kg	100 ng/kg (Stop- Exposure)
2-Year Study Number Examined Microscopically Follicular Cell, Hypertrophy	52 3	(2.3)	54 4	(1.5)	53 4	(2.0)	51 7	(1.1)	53 10* (1.2)	52 17** (1.2)	49 6▲ (1.5)
C-cell Adenoma Overall rate <sup>c</sup> Adjusted rate <sup>d</sup> Terminal rate <sup>e</sup> First incidence (days) Poly-3 test <sup>g</sup> Poly-3 test <sup>g</sup>	20/52 46.79 10/2: 474 P=0.	2 (38%) % 5 (40%) 010N	15/5/ 40.4 <sup>4</sup> 10/2 462 P=0.	4 (28%) % 1 (48%) 366N	17/5 43.1 13/2 626 P=0.	3 (32%) % 3 (57%) 459N	16/51 43.2% 8/19 ( 338 P=0.4	(31%) 5 42%) 64N	13/53 (25% 31.6% 8/22 (36%) 486 P=0.111N	<ul> <li>b) 11/52 (21%)</li> <li>25.6%</li> <li>6/21 (29%)</li> <li>638</li> <li>P=0.030N</li> </ul>	) 13/49 (27%) 33.7% 7/21 (33%) 584 P=0.166N P=0.288

## TABLE 22Incidences of Neoplastic and Nonneoplastic Lesions of the Thyroid Gland in Female Ratsin the 2-Year Gavage Study of TCDD

	Vehicle Control	3 ng/kg	10 ng/kg	22 ng/kg	46 ng/kg	100 ng/kg	100 ng/kg (Stop- Exposure)
C-cell Adenoma or Carcin	oma						
Overall rate	21/52 (40%)	15/54 (28%)	17/53 (32%)	16/51 (31%)	14/53 (26%)	11/52 (21%)	14/49 (29%)
Adjusted rate	49.0%	40.4%	43.1%	43.2%	34.0%	25.6%	36.3%
Terminal rate	11/25 (44%)	10/21 (48%)	13/23 (57%)	8/19 (42%)	9/22 (41%)	6/21 (29%)	8/21 (38%)
First incidence (days)	474	462	626	338	486	638	584
Poly-3 test	P=0.008N	P=0.288N	P=0.374N	P=0.380N	P=0.115N	P=0.017N	P=0.173N
Poly-3 test							P=0.208

\* Significantly different ( $P \le 0.05$ ) from the vehicle control group by the Fisher exact test (interim evaluations) or the Poly-3 test (2-year study)

\*\* P≤0.01

▲ Significantly different (P≤0.05) from the 100 ng/kg core study group by the Poly-3 test  $a^{a}$ 

<sup>a</sup> Number of animals with lesion

<sup>b</sup> Average severity grade of lesions in affected animals: 1=minimal, 2=mild, 3=moderate, 4=marked

Number of animals with neoplasm per number of animals with thyroid gland examined microscopically

<sup>d</sup> Poly-3 estimated neoplasm incidence after adjustment for intercurrent mortality

e Observed incidence at terminal kill

<sup>1</sup> Beneath the vehicle control incidence is the P value associated with the trend test; the stop-exposure group is excluded from the trend test. Beneath the dosed group incidence are the P values corresponding to pairwise comparisons between the vehicle controls and that dosed group. The Poly-3 test accounts for differential mortality in animals that do not reach terminal sacrifice. A negative trend or a lower incidence in a dosed group is indicated by N.

<sup>g</sup> Pairwise comparison between the 100 ng/kg core and stop-exposure groups

with or without S9 (Table B2; McGregor *et al.*, 1991). In tests for chromosomal damage in cultured Chinese hamster ovary cells, TCDD did not induce sister chromatid exchanges (SCEs) or chromosomal aberrations (Abs) when tested up to toxic levels ( $0.8 \mu g/mL$ ) with or without S9 activation enzymes (Tables B3 and B4; Galloway *et al.*, 1987). Without S9, TCDD induced significant increases in SCEs at concentrations of 0.6 to 0.8  $\mu g/mL$  in two of five independent trials when a prolonged culture time was employed to compensate for TCDD-induced cell cycle delay. However, because the response was not consistently reproducible, the SCE test was concluded to be negative overall. In addition, there was a complication from the ethanol solvent in these

SCE experiments. A comparison of the true negative control culture and an ethanol solvent control culture without TCDD indicated that ethanol alone, in the amount needed to dissolve TCDD, increased SCE frequencies. No induction of sex-linked recessive lethal mutations was observed in germ cells of male *Drosophila melanogaster* following treatment with 50, 250, or 500 ppm TCDD (Table B5; Zimmering *et al.*, 1985). Finally, no induction of Abs was observed in bone marrow cells of male B6C3F<sub>1</sub> mice 17 or 36 hours after intraperitoneal injection of 250, 500, or 1,000  $\mu$ g/kg TCDD (Table B6). In summary, no mutagenic activity was detected with TCDD in a variety of *in vitro* and *in vivo* short-term tests.





#### PLATE 1

Hepatocellular adenoma (asterisk) in the liver of a female rat administered 100 ng/kg TCDD by gavage for 2 years. The borders are distinct, and there is compression of the surrounding hepatic parenchyma (arrows). H&E;  $2.5\times$ 





#### PLATE 3

Cholangiocarcinoma in the liver of a female rat administered 100 ng/kg TCDD by gavage for 2 years. Note the atypical, multilayered aspect of the neoplastic epithelium (arrows). H&E;  $66 \times$ 



#### PLATE 4

Cholangiofibrosis (grade 3) in the liver of a female rat administered 100 ng/kg TCDD by gavage for 2 years. The lesion is small and well circumscribed (arrow). H&E;  $5\times$


Higher magnification of Plate 4. The proliferating biliary epithelium forming mucus-producing, crescent-shaped, glandular structures (asterisks) are surrounded by concentric layers of connective tissue. The glands are lined by a single layer of cells. H&E;  $33 \times$ 





Cholangioma in the liver of a female rat in the 100 ng/kg stop-exposure group. The lesion is well circumscribed, and the margins are indicated by arrows. H&E;  $3.3\times$ 



#### PLATE 7

Cholangioma in the liver of a female rat in the 100 ng/kg stop-exposure group. The mass consists of multiple often dilated bile ducts with little associated fibrous connective tissue stroma. The bile ducts are composed of a single layer of epithelium (arrow) that sometimes forms acinar or papillary structures. H&E;  $66 \times$ 



#### PLATE 8

Multinucleated hepatocytes (arrows) in the liver of a female rat administered 46 ng/kg TCDD by gavage for 2 years. H&E;  $66 \times$ 



Bile duct cyst in the liver of a female rat administered 100 ng/kg TCDD by gavage for 2 years. Multiple dilated bile ducts are lined by attenuated epithelium (arrow). H&E;  $5\times$ 



#### PLATE 10

Nodular hyperplasia in the liver of a female rat administered 100 ng/kg TCDD by gavage for 2 years. There are multiple nodular foci (asterisks) composed of hepatocytes that vary in size. H&E;  $13.2 \times$ 



#### PLATE 11

Nodular hyperplasia in a female rat administered 100 ng/kg TCDD by gavage for 2 years. The nodule is circumscribed (dark arrows), slightly compressing the surrounding normal hepatic tissue, and there are occasional bile ducts (open arrows) within the nodule. H&E;  $100\times$ 



#### PLATE 12

Cystic keratinizing epithelioma in the lung of a female rat administered 100 ng/kg TCDD by gavage for 2 years. The single cystic structure consists of a highly irregular wall of keratinized stratified squamous epithelium and a center (asterisk) filled with keratin. H&E;  $5\times$ 



Cystic keratinizing epithelioma in the lung of a female rat administered 100 ng/kg TCDD by gavage for 2 years. The outer portion of the lesion is composed of keratinized stratified squamous epithelium that grows by expansion into the adjacent lung without evidence of invasion (arrows). H&E;  $66 \times$ 



#### PLATE 14

Alveolar epithelial-bronchiolar metaplasia (grade 3) in the lung of a female rat administered 100 ng/kg TCDD by gavage for 2 years. The normal alveolar epithelium is replaced by cuboidal to columnar (arrow), ciliated epithelium intersparsed with dome-shaped multiciliated cells. H&E;  $33 \times$ 



#### PLATE 15

Squamous metaplasia of the alveolar epithelium (grade 2) in the lung of a female rat administered 100 ng/kg TCDD by gavage for 2 years. Alveoli are lined with mature squamous cells (arrows). H&E;  $66\times$ 



#### PLATE 16

Squamous cell carcinoma in the gingival epithelium of a female rat administered 100 ng/kg TCDD by gavage for 2 years. Nests of neoplastic squamous cells (arrows) are invading the adjacent stroma. H&E;  $33 \times$ 



Squamous cell carcinoma in the uterus of a female rat administered 46 ng/kg TCDD by gavage for 2 years. Note the invasion of cords and clusters of squamous cells (arrows) into the underlying myometrium. Arrowhead indicates the surface lining epithelium. H&E;  $10\times$ 



#### PLATE 18

Higher magnification of Plate 17. Note the invasion of cords and clusters of squamous cells (arrows) into the underlying myometrium. H&E;  $66 \times$ 



#### PLATE 19

Arterial inflammation (arrows) in the pancreas of a female rat administered 100 ng/kg TCDD by gavage for 2 years. H&E;  $5\times$ 

## **DISCUSSION AND CONCLUSIONS**

This 2-year study of the chronic toxicity and carcinogenicity of TCDD in female Harlan Sprague-Dawley rats is one in a series of studies carried out as part of a multistudy NTP initiative examining the relative chronic toxicity and carcinogenicity of dioxin-like compounds (DLCs) and structurally related polychlorinated biphenyls (PCBs) (see Overview section). While one of the primary aims of this dioxin toxic equivalency factor (TEF) evaluation was to analyze the comparative carcinogenicity of TCDD, PeCDF, and PCB126, in this Technical Report, only the results of the TCDD toxicology and carcinogenicity study are described. A quantitative analysis of the effects observed in this study to responses observed with other compounds studied as part of the dioxin TEF evaluation are presented by Toyoshiba et al. (2004) and Walker et al. (2005).

Dose selection for this study of TCDD was based on prior observations made in a 2-year carcinogenicity study of TCDD conducted by Dow Chemical Company (Kociba *et al.*, 1978). This prior study in Spartan Sprague-Dawley rats used dosed feed corresponding to dosage levels of 1, 10, or 100 ng TCDD/kg body weight per day. Increases in liver tumors were observed at doses of 10 and 100 ng TCDD/kg body weight per day (Goodman and Sauer, 1992). For the current TCDD study, animals were treated by gavage with TCDD in corn oil:acetone (99:1) 5 days per week for up to 2 years at doses up to 100 ng/kg (3, 10, 22, 46, or 100 ng/kg). Two additional doses between 10 and 100 ng/kg were used to provide more information on the predicted dose response for induction of neoplasms.

In this study, there was no significant effect on survival as a result of TCDD exposure. Daily administration of TCDD at doses above 46 ng/kg led to a reduction in body weight gain over the course of the 2-year study, with a greater magnitude of reduction observed with increasing dose and duration of dosing. At 31 and 53 weeks, body weights of the 100 ng/kg group were 90% and 85% those of vehicle control group weights, respectively. Reduction in body weight gain is an often seen toxic response to DLCs. The reduction in body weight gain was treatment related and required daily administration. In the stop-exposure group, daily administration of compound ceased after 30 weeks of treatment and was followed by daily (5 days per week) gavage administration of corn oil:acetone (99:1) alone for the remainder of the study. Thereafter, the rate of body weight gain in these animals returned to normal and was similar to vehicle control animals; however, the terminal body weights were 92% of that observed in vehicle controls.

The principal findings of this study were increased incidences of benign and malignant neoplasms in several organs, with specific increased incidences of neoplasms in the lung (cystic keratinizing epithelioma), liver (cholangiocarcinoma and hepatocellular adenoma), and oral mucosa (gingival squamous cell carcinoma), following chronic administration of 100 ng/kg TCDD in corn oil:acetone (99:1) by gavage, five times per week for 2 years. In addition, there was an increase in the incidence of squamous cell carcinoma of the uterus in the 46 ng/kg group that was considered to be treatment related. A significant trend for increased incidence of pancreatic adenoma or carcinoma (combined) may have been related to treatment. Occurrences of hepatocholangioma and cholangioma of the liver may also have been related to TCDD treatment.

The principal nonneoplastic finding in this study was a significant increase in incidence and severity of hepatotoxicity. In addition, numerous organs exhibited increased incidences of nonneoplastic lesions, notably the lung, oral mucosa, pancreas, thymus, adrenal cortex, heart, clitoral gland, kidney, forestomach, mesentery, and thyroid gland.

Administration for 2 years led to significant accumulation of TCDD in liver and adipose tissue. The significant accumulation in adipose tissue is consistent with the lipophilic nature of this compound. Previous studies of TCDD indicate that the liver and adipose are the main depots for TCDD in rodents and, combined, contribute approximately 70% to 80% of the total body burden within the animal (DeVito et al., 1995). As expected, the mean levels in the liver were higher than those in adipose on a wet weight basis. This is likely caused by the sequestration of TCDD in the liver as a result of binding to CYP1A2, which is also inducible by TCDD in the liver (Voorman and Aust, 1989; Diliberto et al., 1997). Because of high detection limits, levels of TCDD in lung and blood were undetectable in almost all animals examined. Prior studies with lower detection limits indicate that TCDD also accumulates in these tissues in Sprague-Dawley rats after chronic exposure to similar doses used in this study, albeit at much lower levels than those seen in liver and adipose (Kohn et al., 1993; Tritscher et al., 2000). The low detection limits in the present study as compared to detectable levels of TCDD in the liver and fat of controls in prior studies and also in the NTP study of the mixture of TCDD, PCB 126, and PeCDF (NTP, 2006b) were caused by the fact that a lowresolution gas chromatography/mass spectrometry system was used instead of a high resolution gas chromatography/mass spectrometry system.

Analysis of liver tissue levels of TCDD in this study indicated that 2-year administration at dose levels of 10 and 100 ng/kg resulted in mean liver levels of 2.2 and 9.3 ng/g tissue, respectively. The relationship between intake and liver levels was linear. By comparison, in the TCDD dosed feed study by Kociba *et al.* (1978), terminal liver TCDD levels were 5.1 and 24 ng/g at the 10 and 100 ng/kg doses, respectively, and over twofold higher than those seen in the present study.

The higher tissue levels of TCDD in the Kociba *et al.* (1978) study compared to the present study may be due to the different dosing regimens used. In the Kociba *et al.* (1978) study, animals were exposed to TCDD 7 days per week via dosed feed. In the present study, animals were treated 5 days per week by oral gavage. Modeling of the tissue concentrations produced by these two dosing scenarios using the pharmacokinetic model of TCDD indicated that the liver levels of TCDD in the dosed feed study would be higher than those by oral gavage (data not shown) over the course of the study, as was observed.

In the current study, cessation of daily treatment with TCDD in the stop-exposure group led to a decline in levels of TCDD in all tissues examined. At the end of the study, the mean level of TCDD in the liver of the stop-exposure animals was 0.7 ng/g (five of 10 animals had undetectable levels below 50 pg/g) compared to

9.3 ng/g in animals exposed for the full 2 years at the same administered dose. This level was similar to that observed at the end of the study in the 3 ng/kg group (mean liver level of 0.7 ng/g). Therefore, interpretation of the pathology results in the stop-exposure animals has to be made in light of the fact that exposure per se does not end on cessation of daily administration of compounds. TCDD levels do decline significantly over the remainder of the study, but because of the lipophilic nature of the compound, animals are still continually exposed throughout the course of the study. Using the pharmacokinetic model for TCDD, the predicted areas under the curve (AUC) for liver TCDD for the 22 and 46 ng/kg groups were  $0.30 \times 10^7$  and  $0.70 \times 10^7$  pg days/g, respectively. By comparison, the AUC at 2 years for the stop-exposure group was predicted to be  $0.52 \times 10^7$  pg days/g. This 2-year AUC seen in the stopexposure group is comparable to that predicted using a daily administered dose of 36 ng/kg per day. In addition, exposure to DLCs occurs as a result of exposure to low levels of these compounds that are present in rodent feed. Previous studies have shown that TCDD can be detected in control animals at low levels (Vanden Heuvel et al., 1994; Walker et al., 1998). These concentrations are attributed to the ingestion of very low levels of polyhalogenated aromatic hydrocarbons present in rodent chow. TCDD in the liver of control animals in this study was below the limits of detection (50 pg/g) except in a single animal at the 53-week time point. All experimental treatments in the TEF studies were made in addition to a background of exposure to DLCs that are normally present in feed, and the control group exposure is not zero. Control levels of TCDD at 2 years in the TEF mixture study (NTP, 2006b) in liver and fat were 7.7 and 12.6 pg/g wet weight, respectively. While the level of TCDD in the feed used in the present study was not analyzed, subsequent analysis of polyhalogenated aromatic hydrocarbons in the NTP 2000 feed indicated that TCDD was below the limits of detection (mean level of quantitation of 0.0596 pg/g feed) (Table E5). However, predicted background intake is at least 100- to 1,000-fold lower than the lowest experimental doses used; therefore, the additional contribution of this background exposure rate to the observed neoplastic and nonneoplastic responses is likely to be negligible.

Increases in CYP1A1 and CYP1A2 are characteristic responses to DLCs and are directly linked to binding and activation of the aryl hydrocarbon receptor (AhR) (Whitlock, 1993). In many cases, the relative potency for induction of CYP1A1 *in vivo* is an appropriate surrogate

for the dioxin-like activity of a given compound and provides the basis for many TEFs (Van den Berg et al., 1998). In this study, dose-dependent increases in CYP1A1- and CYP1A2-associated enzyme activity as a result of TCDD administration were observed at all time points. The finding that the liver was a target following TCDD administration was expected. It was also expected that administration of this compound would lead to increases in these specific responses. While not discussed in this report, data on increased expression of CYP1A1 and CYP1A2 from the present study together with data from the other studies of DLCs conducted as part of the TEF evaluation have been used for a quantitative evaluation of the additivity of relative potency for DLCs for these endpoints (Toyoshiba et al., 2004).

Numerous studies have examined the toxicity of DLCs and have demonstrated that the liver is a principal target organ site for the action of these compounds (Poland and Knutson, 1982; Kociba *et al.*, 1978; NTP, 1982a). In the present study, the principal hepatic neoplasms observed were cholangiocarcinoma and hepatocellular adenoma. Two cases of hepatocholangioma were seen only in the 100 ng/kg core study group, and a single case of cholangioma occurred in the stop-exposure group.

One of the principal findings of the study was the extensive hepatotoxicity induced by TCDD treatment. The incidence and pattern of hepatic toxicity exhibited a clear dose and duration dependence and preceded neoplastic effects on the liver. In this study, there was a significant increase in hepatic toxicity with increases in severity occurring at higher doses and longer durations of exposure. Hepatic toxicity was characterized by foci of cellular alterations, multinucleated hepatocytes, diffuse fatty change, inflammation, necrosis, pigmentation, nodular hyperplasia, bile duct cysts, bile duct hyperplasia, hepatocyte hypertrophy, oval cell hyperplasia, and portal fibrosis. A comprehensive term of toxic hepatopathy was also used, reflecting the overall severity grade of the nonneoplastic lesions. This term allowed for easier comparison of the degree of toxic change among different dose groups than would be possible if the severities of all the individual nonneoplastic changes had to be compared among the different groups. This diagnosis was used in addition to, not instead of, any of the nonneoplastic diagnoses already made. Some treated animals occasionally had just a few of these changes present, but this was not considered to be sufficient liver involvement to warrant a diagnosis of toxic hepatopathy. Treatment-related effects observed in the liver at the 14-week interim evaluation were restricted to hepatocyte hypertrophy, multinucleated hepatocytes, and diffuse fatty change. At 31 weeks, there was an increased incidence and severity of hepatocyte hypertrophy and increased incidence of fatty change; a broader spectrum of effects were observed including toxic hepatopathy, multinucleated hepatocytes, increased severity of inflammation, foci of cellular alterations, and pigmentation deposits. These changes continued to be observed at 53 weeks together with bile duct hyperplasia. At the end of the 2-year study, there were additional toxic changes including increased bile duct cysts, necrosis, cholangiofibrosis, inflammation, portal fibrosis, oval cell hyperplasia, and nodular hyperplasia. At the 10 ng/kg dose, where there were no observed liver neoplasms, significant increases in the incidence of nonneoplastic effects and toxic hepatopathy were seen, including foci of cellular alterations, diffuse fatty change, hepatocyte hypertrophy, multinucleated hepatocytes, inflammation, and pigmentation.

The increases in the incidence and severity of hepatotoxicity and increased incidence of hepatocellular adenoma are consistent with previously observed effects of TCDD and hexachlorodioxins on the liver (Kociba et al., 1978; NCI, 1980; NTP, 1982a). However, in this study, the most significant effect was on the incidence of cholangiocarcinoma. Previous studies of DLCs and PCBs have only rarely seen these cholangial lesions, despite the data showing that bile ducts are targets for DLCs. In an initiation-promotion study, cholangiocarcinoma was seen in 1/14 diethylnitrosamine-initiated female rats exposed to 100 ng TCDD/kg per day for 60 weeks (Walker et al., 2000). In the 2-year bioassay of Aroclor 1254, there were no observed cholangiocarcinomas (Mayes et al., 1998). In addition, there was no increased incidence of cholangioarcinoma in the TCDD feed study by Kociba et al. (1978). It is not clear why there was such a marked difference between the present study and these previous studies of TCDD and related DLCs with respect to the increased incidence of cholangiocarcinoma.

The incidence of hepatocellular adenoma in the 100 ng/kg core study group in the present study (13/53, 25%) was less than that seen in the Kociba *et al.* (1978) dosed feed study where the incidences of hepatocellular adenomas were 2/86, 9/50, and 14/45 (31%) in the 0, 10, and 100 ng/kg groups, respectively (Goodman and Sauer, 1992). In addition, no hepatocellular carcinomas were observed in the present study compared to 4/45 in the 100 ng/kg group in the TCDD dosed feed study (Goodman and Sauer, 1992). In the NTP (1982a) gavage

study of Osborne-Mendel rats, the incidence of liver "neoplastic nodules" in female rats was 12/49 (24%) at a weekly dose of 500 ng/kg per day; the incidence of neoplastic nodules or hepatocellular carcinoma was 14/49 (29%). There was no significant effect in male rats. In the NCI (1980) study of a mixture of 1,2,3,6,7,8hexachlorodibenzo-p-dioxin and 1,2,3,7,8,9-hexachlorodibenzo-p-dioxin in Osborne-Mendel rats, the incidences of neoplastic nodules or hepatocellular carcinoma (combined) were 5/75 (7%), 10/50 (20%), 12/50 (24%), and 30/50 (60%) at doses of 0, 1,250, 2,500, or 5,000 ng/kg per week, respectively. Given the TEF of 0.1 for hexachlorodioxins (Van den Berg et al., 1998), these weekly doses are over a similar range of total toxic equivalents (TEQs) as used in the present gavage study.

There are several explanations for the lower incidence of hepatocellular adenoma relative to the earlier TCDD studies. The terminal liver levels of TCDD in the present study are over twofold lower than those observed in the Kociba et al. (1978) dosed feed study, most likely because of the different dosing regimens used. Consequently, while the administered dose was the same between the two studies, the internal dose to the liver over the course of the study was likely lower than in the Kociba et al. (1978) study; therefore, the lower response may simply be a dose-response issue. In addition, the Kociba study used the Spartan substrain of Sprague-Dawley rats compared to the Harlan Sprague-Dawley rat strain used in the present study. Therefore, the lower incidence may in part be due to substrain differences in susceptibility.

Another aspect to consider with the liver neoplasms is that pathology nomenclature has changed. The Kociba et al. (1978) study indicated a 47% incidence of "hepatocellular hyperplastic nodules" in the 100 ng TCDD/kg group compared to a 9% incidence in control animals. Subsequent to that study, there was an evolution of nomenclature for hepatocellular proliferative lesions, and the slides from that study were reevaluated. In that reevaluation, neoplastic lesions were classified as adenoma or carcinoma. Using the newer nomenclature, the incidence of hepatocellular adenoma was 31% at the highest dose of 100 ng TCDD/kg (Goodman and Sauer, 1992). It is clear from the pathologic reevaluations that some of the hyperplastic nodules originally seen in the Kociba et al. (1978) study were indeed nonneoplastic. Significant hepatotoxicity was noted in the Kociba et al. (1978) TCDD study (Goodman and Sauer, 1992), in the NCI (1980) TCDD study, and in the NTP (1982a) TCDD studies.

The spectrum of hepatocellular proliferative lesions observed in the present study is not common in NTP studies, and there is a lack of biological information relative to the progression and behavior of these lesions. These lesions generally occurred on a background of toxic hepatopathy, the components of which are listed above and described in the Results section. It is generally accepted that, in the rat, hepatocellular adenoma and hepatocellular carcinoma represent a morphological and biological continuum (Narama et al., 2003; Hailey et al., 2005). Foci of cellular alteration are often part of that continuum but not always. In high dose animals, proper categorization of the lesions was further complicated by the presence of the toxic hepatopathy. While the biological behavior of hepatocellular lesions within this study and other studies conducted as part of the dioxin TEF evaluation is uncertain, the morphology suggests that in this study, eosinophilic foci and mixed cell foci, nodular hyperplasia, and potentially adenoma were a continuum. Carcinomas were not observed in the present study.

The foci of cellular alterations seen in dosed animals generally differed from the typical foci seen in vehicle controls. Foci seen in vehicle controls were usually smaller, lacked discrete borders and blended with the surrounding parenchyma, produced little or no compression, and consisted of cells that were normal-sized or slightly smaller or larger than normal. In contrast, foci in the livers of dosed animals generally had discrete borders, produced some compression of the adjacent parenchyma, and consisted of large, hypertrophic, often vacuolated cells. The significantly increased incidences of hypertrophy resulted in a greater degree of compression of adjacent hepatic parenchyma than is often seen with foci of hepatocellular alteration. At 2 years, focal lesions were observed that resembled foci of hepatocellular alteration but were larger and often nodular with greater compression of surrounding hepatic parenchyma and more disorganization of hepatic cords. As with foci, these lesions generally contained a somewhat normal hepatic structure including portal triads with biliary tracts. Additionally, these focal lesions contained variable numbers of randomly scattered biliary epithelium that often formed profiles of small glands/ductules. The large size of the lesions and presence of scattered biliary epithelium suggested a proliferative response of both hepatocellular and biliary cells; therefore, these lesions were considered to have progressed beyond a simple focus of cellular alteration. However, because of the somewhat normal hepatic structure and the dual cellular composition, the lesions were considered to be hyperplastic rather than neoplastic and were diagnosed as nodular hyperplasia.

In the higher dose animals with severe toxic hepatopathy, there was evidence of hepatocyte degeneration and loss and a regenerative response by the damaged liver. The term of "hyperplasia, nodular" was selected as the inclusive term and was characterized by areas of focal hypertrophy and hyperplasia of hepatocytes that also contained proliferating biliary epithelium. This lesion was considered to be the result of the presence of a proliferative stimulus. In the dioxin TEF evaluation studies, nodular hyperplasia varied in size but generally appeared morphologically similar whether in a high dose animal with severe toxic hepatopathy or in a lower dose animal where the toxic hepatopathy was minimal to nonexistent. Nodular hyperplasia was seen most commonly in the higher dose groups in which prominent toxic changes were present. However, a lesser degree of nodular hyperplasia was sometimes seen in lower dose animals in which the only evidence of liver pathology may have been hepatocellular hypertrophy.

Morphologically, a hyperplastic nodule associated with regeneration cannot be distinguished from a hyperplastic nodule of another pathogenesis. The morphological alterations suggest that regeneration is a significant contributor to the proliferative response in animals with toxic hepatopathy. However, this does not explain these responses in animals that lack toxic hepatopathy. This indicates that some type of stimulus, other than regeneration secondary to degeneration and necrosis of the hepatic parenchyma, may have contributed to the proliferative lesions observed in this study.

Dealing with the potential pathogenesis for the foci and nodular hyperplasia, the earliest treatment-related hepatocellular change seen in these studies, noted at the interim 14-, 31-, and 53-week evaluations, was a diffuse hepatocyte hypertrophy. With continued dosing, poorly demarcated foci of prominent hypertrophic, often vacuolated hepatocytes resembling those seen in foci and nodular hyperplasia were seen superimposed on the background of diffuse hypertrophy. It appeared that the poorly demarcated foci of hypertrophic cells grew with continued dosing, giving rise to lesions diagnosed as foci, and that continued dosing, in some instances aided by toxic changes, may have progressed to nodular hyperplasia.

In contrast to nodular hyperplasia, hepatocellular adenoma was a nodular mass that usually was larger than a focus, had a distinct border, and produced more compression of surrounding normal hepatic parenchyma. Adenomas were composed of mildly to moderately pleomorphic hepatocytes with a subjectively increased nucleus to cytoplasmic ratio. Cells lacked the normal architectural arrangements of hepatic lobules, and while a few bile ducts may have been present within an adenoma, they were usually found at the periphery of the lesion and were considered entrapped. Proliferating biliary epithelium or oval cells were generally absent. The presence of proliferating bile duct epithelium was an important feature, useful in differentiating adenoma from nodular hyperplasia.

The increased incidence of cholangiocarcinoma following exposure was an unexpected finding but consistent with observations made in other studies conducted as part of the dioxin TEF evaluation (NTP, 2006a,b,c,d,e,f). Spontaneous cholangioma and cholangiocarcinoma are apparently rare in the Harlan Sprague-Dawley rat and were not observed in 371 vehicle control animals from these seven studies. These neoplasms are characterized by glandular structures lined by a single layer of welldifferentiated epithelium (benign lesions) or single or multiple layers of epithelial cells that have malignant characteristics (e.g., high nuclear to cytoplasmic ratio, pleomorphism and anisokaryosis, and an increased mitotic rate).

In the present study, cholangiocarcinoma was diagnosed, and while it differed morphologically from spontaneous cholangiocarcinoma, it was similar to chemicallyinduced cholangiocarcinoma in another study (Maronpot et al., 1991). In this study, cholangiocarcinomas were variably sized, often multiple lesions composed of irregular and atypical bile ducts in a matrix of fibrous connective tissue. The bile ducts themselves were often incomplete or crescent-shaped and lined by very basophilic, cuboidal to columnar cells with large, euchromatic nuclei. Stratification of these epithelial cells was present in some areas. Atypical biliary epithelium was often identified within the adjacent hepatic parenchyma, suggesting invasion. The fibrous connective tissue component was frequently profound, much more than that seen in the scirrhous reaction that may be observed with spontaneous cholangiocarcinoma. The lesions seen in this study were sometimes large, effacing an entire liver lobe.

Cholangiofibrosis was the term used to describe small lesions that were less aggressive in appearance. Cholangiofibrosis often originated in the portal area and tended to have a more mature fibrous connective tissue component and less atypia associated with the epithelial cells. Most often, cholangiofibrosis and cholangiocarinomas seen in this study did not compress the surrounding hepatic parenchyma or expand beyond the existing hepatic profile. However, cholangiocarcinomas often did expand within the liver lobe.

While cholangiofibrosis and cholangiocarcinoma appear to be a morphological continuum, there is limited biological information relative to the pathogenesis or progression of these lesions. As a result, the most appropriate classification scheme for these lesions is somewhat uncertain and controversial. While the characteristic of malignancy, distant metastasis, was not observed in any animals in the present study, other characteristics of malignancy were present, such as atypical appearance of the epithelial cells and apparent localized invasion. It was clear that some of these cholangiolar lesions were small, very benign appearing, and warranted a nonneoplastic diagnosis, and there were lesions at the other end of the spectrum that appeared aggressive. While there were specific diagnostic criteria for cholangiofibrosis versus cholangiocarcinoma, some of the lesions did not readily fit the criteria and posed a diagnostic challenge.

Other chemicals, including furan, have increased the incidences of lesions similar to those observed in the present study (Maronpot *et al.*, 1991). In the Maronpot *et al.* (1991) furan study, the lesions appeared more aggressive, yet even in that study, where there was nearly a 100% incidence in treated animals, there were few metastases. In the present study, it appears that the cholangiocarcinomas were slowly growing neoplasms of relatively low-grade malignancy. Transplantation studies done in the furan study were positive for growth and metastases. Transplantation studies were not done with lesions from the present study.

There was a single occurrence of cholangioma in the 100 ng/kg stop-exposure group, and two hepatocholangiomas occurred in the 100 ng/kg group. Both neoplasm types appear to be rare and did not occur in 371 vehicle control animals from this study or the other six dioxin TEF evaluation studies (NTP, 2006a,b,c,d,e,f). Hepatocholangiomas were mixed neoplasms with areas of hepatocytes that appeared identical to hepatocellular adenoma and areas of ductular structures lined by biliary epithelium that appeared identical to cholangioma. The pluripotent nature of these neoplasms was demonstrated by occasional ductular structures lined by cells resembling both hepatocytes and biliary epithelium. In contrast to the cholangiofibrosis and cholangiocarcinomas, a scirrhous response was not present within these neoplasms. While the histogenesis of hepatocholangioma is not clear, there was evidence of proliferation of hepatocytes, biliary epithelium, and oval cells within these studies. Therefore, the occurrence of these rare hepatocholangiomas in the 100 ng/kg group and the single incidence of cholangioma in the 100 ng/kg stop-exposure group may have been related to administration of TCDD.

The mechanism underlying the increase in incidence of cholangiocarcinoma is likely to be multifactorial. There was clearly an effect on bile duct proliferation in this study. This may be an indirect response to the toxicity observed as a result of the action of the DLC on the hepatocytes or a direct effect on the biliary cells themselves. Tritscher et al. (1995) showed a high degree of staining for TGF alpha in bile duct cells after TCDD administration in female rats. The observed bile duct proliferation may represent a process of excessive and long-term repair following specific damage to hepatocytes, leading to the death of hepatocytes and perhaps also of the bile duct epithelium. The proliferative response may be a reparative response of proliferating hepatocytes, bile duct cells, and scarring tissue (cholangiofibrosis). The inflammation observed can produce oxidative stress that may also result in promotion of DNA damage. Consequently, the oxidative stress may be only a secondary phenomenon of the ongoing response to the toxic hepatopathy. In addition, there may be a direct stimulatory effect on the oval cells themselves. This is supported by the observed increases in oval cell hyperplasia. The oval cells may differentiate into hepatocytes or biliary cells. This may explain why both hepatocellular proliferative and biliary lesions are associated with exposure.

There has been a considerable amount of research conducted examining the potential mode of action of DLCs in the liver. There is a general scientific consensus that almost all responses of TCDD and related compounds require initial binding to the AhR. Recent data indicate that the acute toxic responses (including hepatotoxicity) to TCDD require AhR binding and nuclear localization (Bunger *et al.*, 2003). In addition, transgenic mouse studies indicate that constitutive activation of the AhR alone can lead to induction of stomach tumors (Andersson *et al.*, 2002). Numerous studies have shown that TCDD is a potent promoter of altered hepatic foci in initiation-promotion models of hepatocarcinogenesis (Pitot et al., 1980; Maronpot et al., 1993). Given that TCDD and related compounds are not direct-acting genotoxic agents and are potent growth dysregulators, it is believed that their predominant mode of action is as tumor promoters. Within a conceptual multistage model of carcinogenesis, promotion mediated via the AhR may be caused by an increase in net growth rate of initiated cells because of selective growth advantage or decreased rate of cell death via suppression of apoptosis. In the current study with TCDD and in previous tumor promotion studies of TCDD, there were significant increases in hepatocyte replication as judged by BrdU labeling studies (Maronpot et al., 1993; Walker et al., 1998; Wyde et al., 2001a). Studies by Stinchcombe et al. (1995), Worner and Schrenk (1996), and Bohnenberger et al. (2001) have also shown a suppression of apoptosis by TCDD and PCBs. In addition, altered growth regulation may be caused by alterations in intercellular communication, which have been observed in the livers of rats exposed to DLCs (Baker et al., 1995; Warngard et al., 1996; Bager et al., 1997). While TCDD is not a direct acting genotoxic agent, there are data indicating that persistent AhR active compounds may be indirectly genotoxic. This may contribute to an increase in the number of cells within the liver capable of undergoing promotion (Moolgavkar et al., 1996; Portier et al., 1996). It is hypothesized that the indirect genotoxicity may be via AhR-dependent induction of CYP1 family cytochromes P450 that leads to induction of oxidative stress caused by either inefficient electron transfer during P450 metabolism (Park et al., 1996) or the production of redox active estradiol metabolites as a result of CYP1 mediated estrogen metabolism (Lucier et al., 1991; Kohn et al., 1993). Studies have shown induction of oxidative stress and DNA damage by high dose acute exposure to TCDD (Stohs et al., 1990). The induction of lipid peroxidation and single stranded DNA breaks was also observed in tissues from the present TCDD study (Hassoun et al., 2000, 2002). Other studies on the female specific tumor promotion response in rats have shown induction of oxidative DNA damage and hepatocyte replication by TCDD that is female specific and estrogen dependent (Lucier et al., 1991; Tritscher et al., 1996; Wyde et al., 2001a,b).

In the present study of TCDD, an increase in the incidence of cystic keratinizing epithelioma (CKE) of the lung was observed at the highest dose of 100 ng/kg. 75

Histopathologically, these lesions varied in size and number and appeared as cystic structures consisting of a highly irregular wall of highly keratinized stratified squamous epithelium, with a center filled with keratin. These lesions were absent in vehicle control and low dose animals.

In the 2-year feed study by Kociba et al. (1978), an increased incidence of keratinizing squamous cell carcinoma of the lung was observed following exposure to 100 ng TCDD/kg body weight per day. In the evaluation of tissues in the dioxin TEF evaluation studies, squamous cell carcinoma was distinguished from CKE by the presence of areas of solid growth and evidence of inva-While no direct comparison has been made sion. between CKE and the keratinizing squamous cell carcinoma observed in the Kociba et al. (1978) study, given the keratinizing nature of the CKE, it is possible that these represent the same lesions. At the time of the Kociba et al. (1978) evaluation, CKE was not a consistently used diagnostic term. Diagnostic criteria for classification of CKE as a lesion distinct from squamous cell carcinoma were later developed at a workshop held in the mid 1990s (Boorman et al., 1996).

In contrast to the present study, a recent study of the carcinogenicity of the high TEQ PCB mixture Aroclor 1254 in Charles River Sprague-Dawley rats demonstrated no increases in any type of lung tumor, despite a high incidence of hepatocellular tumors (Mayes *et al.*, 1998). While Aroclor 1254 contains a significant TEQ contribution by coplanar PCBs that have dioxin-like activity, this mixture also contains mono-ortho and di-ortho PCBs.

In addition to an increase in the incidence of CKE, there was a significant increase in the incidences of bronchiolar metaplasia of the alveolar epithelium at all doses at 2 years including the stop-exposure group. These findings are consistent with prior observations of an increase in the incidences of alveolar-bronchiolar metaplasia following exposure to TCDD within the framework of a two stage initiation-promotion model in Sprague-Dawley rat lung (Tritscher et al., 2000). Of note was the significant reduction in the incidences of alveolar epithelial hyperplasia in all TCDD treated groups, with no incidences reported in the TCDD treated groups. Dose-related increased incidences and severities of squamous metaplasia and alveolar histiocytic infiltration were also noted in the 46 and 100 ng/kg groups.

Alveolar ducts and alveoli are normally lined by type I alveolar epithelial cells and type II alveolar epithelial cells, which are cuboidal. Type I cells are very susceptible to damage, and the typical response in the lung, subsequent to damage to the type I cells, is a proliferation of the type II cells. This is often diagnosed as alveolar epithelial hyperplasia. In the present study, alveolar epithelial hyperplasia was observed only in vehicle control animals, and TCDD induced a multifocal lesion that was found throughout the lung at the junction of the terminal bronchioles and alveolar ducts. The epithelium was cuboidal to columnar and ciliated, in contrast to type II alveolar epithelial cells. Scattered throughout the ciliated cells were dome-shaped nonciliated cells, consistent with Clara cells. Clara cells are normally found in the lining of the bronchioles but not alveoli or alveolar ducts. Histochemical analyses of mucin and glutathione S-transferase pi in lung tissue from the dioxin TEF evaluation studies indicate that this appears to be similar to bronchiolar epithelium and is distinct from alveolar epithelial hyperplasia (Brix et al., 2004). It is not clear, however, if this lesion represents a destruction of type I alveolar epithelial cells with replacement by bronchiolar type epithelium (bronchiolar metaplasia) or an extension of bronchiolar epithelium from the terminal bronchiole (bronchiolar hyperplasia).

There are likely at least two potential mechanisms involved in the lung. CYP1A1 is inducible in the lung by TCDD in several species (Beebe et al., 1990; Walker et al., 1995). This was confirmed in the present study by the observed increase in lung CYP1A1-associated 7-ethoxyresorufin-O-deethylase activity. The inducibility of CYP1A1 by TCDD is observable in Clara cells, bronchiolar cells, and, to a lesser degree, in type II alveolar epithelial cells (Tritscher et al., 2000). This indicates that the bronchiolar epithelium is clearly responsive to AhR ligands and suggests a potential for a direct effect of TCDD on the lung. In vitro studies of normal human lung epithelial cells (mixed type II, Clara cell type) demonstrate the altered expression by TCDD of genes involved in numerous cell signaling pathways, indicating altered retinoid signaling and altered cytokine signaling pathways (Martinez et al., 2002).

Another possible mechanism for the action of TCDD on the lung may be an indirect effect caused by the disruption of retinoid homeostasis in the liver. In rodents, mobilization of retinoid stores by TCDD and DLCs leads to a disruption in retinoid homeostasis and vitamin A deficiency (Van Birgelen *et al.*, 1994, 1995b; Fiorella *et al.*, 1995; Fattore *et al.*, 2000; Schmidt *et al.*, 2003). A characteristic of retinoid deficiency is abnormal epithelial differentiation to a keratinized squamous phenotype (Lancillotti *et al.*, 1992; Lotan, 1994). The action of DLC may therefore be a disruption of retinoid action leading to altered growth and differentiation of the lung epithelium, resulting in squamous metaplasia and CKE.

Gingival squamous cell carcinoma of the oral mucosa occurred in all dose groups except the 22 ng/kg group. The incidence of this lesion was significantly elevated in the 100 ng/kg core study group. In the TCDD feed study by Kociba et al. (1978), there were increases in the incidences of stratified squamous cell carcinoma of the hard palate/nasal turbinates in both male and female rats. The location of the squamous cell carcinomas in the present study was adjacent to the molars and invaded the hard palate/nasal turbinate areas, suggesting that the lesions seen in this and the Kociba et al. (1978) studies are similar. In addition to the squamous effects in the oral mucosa and lung, there were also increased incidences of forestomach squamous hyperplasia in the 46 and 100 ng/kg core study groups. All these squamous lesions may be related to the alteration in retinoid homeostasis that is known to be induced by TCDD.

In recent years, there has been an increasing awareness of the sensitivity of the oral cavity to the effects of DLCs. In two PCB/PCDF human poisoning episodes, one of the toxic responses observed in humans was early tooth eruption (Grassman et al., 1998). More recent studies have shown that TCDD can accelerate incisor tooth eruption and delay molar eruption. Proliferation of the periodontal squamous epithelium has been seen in juvenile mink exposed to TCDD (Render et al., 2001) but not in juvenile Long Evans rats exposed to 100 ppb PCB 126 or 10 ppb TCDD for 101 days (Aulerich et al., 2001). Studies suggest that the effect of TCDD on tooth development is caused by a disruption in epidermal growth factor receptor-mediated signaling (Partanen et al., 1998) as has been shown for other developmental effects of TCDD such as a cleft palate (Abbott et al., 2003).

In the present study, there were increased incidences of adrenal cortical atrophy in the 100 ng/kg core study group and of cortical hyperplasia in the 22, 46, and 100 ng/kg groups. The incidence of cytoplasmic vacuolization was significantly increased in the 22 ng/kg group. Cortical atrophy was a prominent effect and may reflect the continued stress in these animals, leading to depletion of corticosteroid hormones (Sapolsky *et al.*, 1987). It may also be caused by other unknown mechanisms.

In the present study, there was a trend for increased incidence of pancreatic neoplasms associated with TCDD exposure. Majeed (1997) showed that acinar pancreatic carcinoma is a rare tumor seen in the pancreas of female Sprague-Dawley CD rats and has a spontaneous background rate of 0.02%. These data suggest that increased incidences of pancreatic neoplasms may have been related to TCDD treatment. By comparison, in the 2-year feed study by Kociba *et al.* (1978), there was a significant reduction in the incidence of acinar adenoma of the pancreas in male rats relative to the high spontaneous incidence seen in male rats in that study.

Dose-related increases in the incidences of acinar atrophy were seen in all treated groups, and the incidence was significantly elevated in the 100 ng/kg core study group. The incidences of acinar chronic active inflammation, acinar cytoplasmic vacuolization, and chronic arterial inflammation in the 100 ng/kg core study group and the incidence of acinar cytoplasmic vacuolization in the 46 ng/kg group were all significantly greater than those in the vehicle controls at 2 years.

Acinar atrophy of the pancreas may be related to the down-regulation of cholecystokinin (CCK). As shown by Lee et al. (2000) in samples from the present TCDD study, intestinal CCK is reduced by TCDD exposure. Down-regulation of CCK is likely caused by a general endocrine effect, resulting from the reduction in body weight gain following exposure to TCDD. CCK is an important regulator of pancreatic growth and function (Baldwin, 1995; Varga et al., 1998). Previous studies have shown that increased incidences of apoptosis and pancreatic acinar atrophy occurred in Otsuka Long-Evans Tokushima Fatty rats that lack the CCK-A receptor (Jimi et al., 1997). In addition, antagonism of CCK action can lead to reduced pancreatic growth (Ohlsson et al., 1995). The relationship among acinar atrophy, cytoplasmic vacuolization, and development of pancreatic neoplasms is unclear because these lesions were also observed in the PCB 126 study (NTP, 2006a), which was conducted as part of the dioxin TEF evaluation, yet there were no increases in the incidences of pancreatic neoplasms in the that study.

In the present study, the incidences of nephropathy were significantly greater in the 100 ng/kg core study and stop-exposure groups than in the vehicle controls. Doserelated increases in the incidences of hyperplasia of the transitional epithelium were noted in all treated groups and were significantly elevated in the 46 and 100 ng/kg core study groups. The significance of this difference was unclear; it did not appear to correlate with the increased severity of nephropathy because the animals with hyperplasia often had minimal nephropathy. While TCDD is known to induce biochemical responses in the kidney (Walker *et al.*, 1995), the kidney historically has not been a target for TCDD-induced neoplasia.

Administration of TCDD to female Harlan Sprague-Dawley rats in this study significantly increased the incidences of cardiomyopathy in all dose groups administered 10 ng/kg or greater. However, the average severity of cardiomyopathy was unaffected. Cardiomyopathy is a common, spontaneously occurring degenerative change of myocardial fibers that is seen in rats as they age. Its cause in the rat is unknown, but age of onset and severity are affected by diet, environment, and stress (Jokinen et al., 2003). The microscopic appearance of cardiomyopathy was the same in both the vehicle control and treated animals and was typical of that described for spontaneous lesions. This finding may suggest that exposure to the chemical increased the occurrence of the spontaneous change. The heart is a target for TCDD and related DLCs in both rodents and humans (Peterson et al., 1993; Flesch-Janys et al., 1995; Walker and Catron, 2000; Heid et al., 2001). In the 2-year feed study in which Sprague-Dawley rats were administered up to 100 ng/kg per day of TCDD, Kociba et al. (1978) also reported an increase of myocardial degenerative change above background levels in females only.

In this study, the incidence of thymic atrophy was significantly elevated at 46 and 100 ng/kg at 14, 31, and 53 weeks and at 22 ng/kg or greater at the end of the study. Thymic atrophy is one of the characteristic immunotoxic responses to DLCs (Poland and Knutson, 1982) and is due to an AhR-mediated alteration in lymphocyte growth and differentiation (Staples *et al.*, 1998; Gasiewicz *et al.*, 2000). Thymic atrophy may also be related to the reduction in body weight gain observed in these animals as seen in short-term feed restriction studies (Levin *et al.*, 1993).

In this 2-year study, there was a significantly lower adjusted incidence of thyroid gland C-cell adenomas in the 100 ng/kg core study group. Similarly, there was a lower incidence of C-cell adenomas in male rats exposed to TCDD in feed (Kociba *et al.*, 1978). The reduced incidence of C-cell adenomas may be related to the reduced body weight gain seen in the 100 ng/kg group.

In the present study, there were no significant increases in the incidences of thyroid gland follicular cell neoplasms, although there were increases in the incidences of follicular cell hypertrophy in the 46 and 100 ng/kg core study groups. By comparison, in the NTP 2-year gavage study of TCDD in Osborne-Mendel rats, there were significant increases in the incidences of thyroid gland follicular cell adenomas (NTP, 1982a) in male rats and a nonsignificant increase in female rats.

Alteration in thyroid hormone homeostasis by TCDD is well established (Van Birgelen et al., 1994, 1995a; Sewall et al., 1995). Analyses of thyroid hormones in the present study confirmed the alterations in thyroid hormone homeostasis by TCDD. The disruption of thyroid hormone homeostasis by DLCs is believed to be caused by the increase in thyroxine  $(T_4)$  glucuronidation as a result of increased expression of UDP-glucuronosyltransferases. This leads to a reduction in circulating  $T_4$ , decreased negative feedback inhibition of the thyroid gland, and overexpression of thyroid stimulating hormone (TSH) (Curran and DeGroot, 1991). Kohn et al. (1996) developed a mathematical model of the effects of TCDD on UDP-glucuronosyltransferase expression and thyroid hormone homeostasis that is consistent with this mechanism. It has been hypothesized that overstimulation of the thyroid gland by TSH may be involved in the mechanism of follicular cell carcinogenesis (Hill et al., 1989). In the present study, significant alterations in T4 and TSH were observed at the early time points, but a significant increase in TSH levels was seen only at 14 weeks, despite a significant effect on  $T_4$ . It is possible, therefore, that the lack of follicular cell neoplasia in this study reflects a lack of a sustained long-term increase on TSH sufficient to promote neoplasia.

In this study, there was a significantly lower adjusted incidence of mammary gland neoplasms following TCDD administration at 100 ng/kg per day. Fibroadenoma is a spontaneous lesion in female Sprague-Dawley rats and occurred at a high incidence (71%) in vehicle control animals. Mammary gland carcinoma was also seen in vehicle control rats at a lower incidence (10%). In the 100 ng/kg core study group, the incidences of fibroadenoma and carcinoma were 51% and 0%, respectively. In addition, there was a significantly lower incidence of spontaneous pituitary gland adenoma in the 100 ng/kg core study group (25%) than in the vehicle control group (58%). The lower incidences of mammary gland and pituitary gland neoplasms in treated rodents are believed to be related to a general endocrine effect as a result of reductions in body weight gain associated with treatment. An association between reduced body weight gain and lower incidences of mammary gland and pituitary gland tumors has been observed in many NTP studies of F344/N rats (Seilkop, 1995). Significantly lower incidences of mammary gland and pituitary gland tumors were also observed in animals exposed to 100 ng/kg in the 2-year feed study by Kociba *et al.*, (1978).

Reductions in insulin-like growth factor 1 (IGF-1) may underlie the inhibitory effect of reduced body weight gain on tumor development. Caloric restriction leads to lower levels of IGF-1 and reduction in background tumor rates (Hursting et al., 2003). One of the major intestinal hormones expressed in the proximal gastrointestinal tract is CCK, which regulates gallbladder contraction, pancreatic secretion, stomach emptying, and intestinal motility; CCK can also inhibit food intake. In an analysis of intestinal tissue obtained from the current study, Lee et al. (2000) showed lower levels of intestinal CCK and induction of insulin-like growth factor binding protein 3 (IGFBP3) by TCDD in this NTP study. Alterations in CCK processing enzymes by TCDD were also observed in cultured intestinal cells, suggesting a direct effect of TCDD on intestinal cells. The authors hypothesized that changes in CCK may be caused by alterations in processing enzymes and lower IGF-1 levels as a result of alterations in IGFBP3.

In the present TCDD study, there was a significantly increased incidence of squamous cell carcinoma of the uterus in animals administered 46 ng/kg. There were also two animals with squamous cell carcinomas in the stop-exposure group. The observed neoplasms were considered to be related to TCDD treatment because of the observed increased incidences of squamous neoplasms seen in other tissues in this study. There were no neoplasms in the 100 ng/kg core study group, which may be because of a suppressive effect of reduced body weight gain seen at the highest dose. Within this context, there was a significant reduction in the incidence of uterine squamous metaplasia in the 100 ng/kg core study group. By comparison, there was no effect on uterine squamous cell carcinoma in the TCDD dosed-feed study by Kociba et al. (1978). However, there was a significantly lower incidence of benign tumors of the uterus in the 100 ng/kg group.

#### **CONCLUSIONS**

Under the conditions of this 2-year gavage study, there was *clear evidence of carcinogenic activity*\* of TCDD in female Harlan Sprague-Dawley rats based on increased incidences of cholangiocarcinoma and hepatocellular adenoma of the liver, cystic keratinizing epithelioma of the lung, and gingival squamous cell carcinoma of the oral mucosa. The increased incidence of squamous cell carcinoma of the uterus was also considered to be related to TCDD administration. The marginally increased incidence is the statement of the set of the set

dences of pancreatic acinar neoplasms and occurrences of hepatocholangioma and cholangioma of the liver may have been related to TCDD administration.

TCDD administration caused increased incidences of nonneoplastic lesions of the liver, lung, oral mucosa, pancreas, thymus, adrenal cortex, heart, clitoral gland, kidney, forestomach, mesentery, and thyroid gland in female rats.

<sup>\*</sup> Explanation of Levels of Evidence of Carcinogenic Activity is on page 11. A summary of the Technical Report Review Subcommittee comments and public discussion on this Technical Report appears on page 13.

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## APPENDIX A SUMMARY OF LESIONS IN FEMALE RATS IN THE 2-YEAR GAVAGE STUDY OF TCDD

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#### TABLE A1a Summary of the Incidence of Neoplasms in Female Rats at the 14-, 31-, and 53-Week Interim Evaluations in the 2-Year Gavage Study of TCDD<sup>a</sup>

	Vehicle Control	3 ng/kg	10 ng/kg	22 ng/kg	46 ng/kg	100 ng/kg
Disposition Summary						
Animals initially in study	28	28	28	28	28	28
14-Week interim evaluation	10	10	10	10	10	10
31-Week interim evaluation	10	10	10	10	10	10
53-Week interim evaluation	8	8	8	8	8	8
Animals examined microscopically	28	28	28	28	28	28
14-Week Interim Evaluati	ion					
Genital System						
Uterus	(10)	(1)				(10)
Deciduoma benign	()	1 (100%)				()
Integumentary System						
Mammary gland	(10)					(10)
Carcinoma	1 (10%)					
Systems Examined at 14 Weeks	s with No Neoplasms	Observed				
Alimentary System	· · · · · · · · · · · · · · · · · · ·					
Cardiovascular System						
Endocrine System						
Conoral Body System						
General Bouy System						
Museulogkolotal System						
Nauseuro System						
Nervous System						
Respiratory System						
Special Senses System						
Urinary System						
31-Week Interim Evaluati	ion					
Integumentary System						
Mammary gland	(10)				(1)	(10)
Fibroadenoma	(**)				1 (100%)	(10)
Skin	(1)				()	
Sarcoma	1 (100%)					
	. /					

# TABLE A1aSummary of the Incidence of Neoplasms in Female Rats at the 14-, 31-, and 53-Week Interim Evaluationsin the 2-Year Gavage Study of TCDD

	Vehicle Control	3 ng/kg	10 ng/kg	22 ng/kg	46 ng/kg	100 ng/kg
Systems Examined at 31 Wee	ks with No Neoplasms	Observed				
Alimentary System						
Cardiovascular System						
Endocrine System						
General Body System						
Genital System						
Musculoskeletal System						
Nervous System						
Respiratory System						
Special Senses System						
Urinary System						
53-Week Interim Evalua	tion					
Endocrine System						
Adrenal medulla	(8)		(1)			(8)
Pheochromocytoma benign	(2)		1 (100%)			-
Pituitary gland	(8)		(1) (100%)			(7)
Adenoma			1 (100%)			
Genital System						
Uterus	(8)			(1)		(8)
Polyp stromal	1 (13%)					1 (13%)
Integumentary System						
Mammary gland	(8)	(2)	(2)			(8)
Fibroadenoma		2 (100%)	1 (50%)			
Fibroadenoma, multiple			1 (50%)			
		<u>.</u>				
Systems Examined at 53 Wee	ks with No Neoplasms	Observed				
Alimentary System						
Carulovascular System						

Cardiovascular System General Body System Hematopoietic System Musculoskeletal System Nervous System Respiratory System Special Senses System Urinary System

#### TABLE A1a

#### Summary of the Incidence of Neoplasms in Female Rats at the 14-, 31-, and 53-Week Interim Evaluations in the 2-Year Gavage Study of TCDD

	Vehicle Control	3 ng/kg	10 ng/kg	22 ng/kg	46 ng/kg	100 ng/kg
Neoplasm Summary						
Total animals with primary neoplasms <sup>c</sup>						
14-Week interim evaluation	1	1				
31-Week interim evaluation	1				1	
53-Week interim evaluation	1	2	3			1
Total primary neoplasms						
14-Week interim evaluation	1	1				
31-Week interim evaluation	1				1	
53-Week interim evaluation	1	2	4			1
Total animals with benign neoplasms						
14-Week interim evaluation		1				
31-Week interim evaluation					1	
53-Week interim evaluation	1	2	3			1
Total benign neoplasms						
14-Week interim evaluation		1				
31-Week interim evaluation					1	
53-Week interim evaluation	1	2	4			1
Total animals with malignant neoplasms						
14-Week interim evaluation	1					
31-Week interim evaluation	1					
Total malignant neoplasms						
14-Week interim evaluation	1					
31-Week interim evaluation	1					

a Number of animals examined microscopically at the site and the number of animals with neoplasm
b Number of animals with any tissue examined microscopically
c Primary neoplasms: all neoplasms except metastatic neoplasms
Summary of the Incidence of Neoplasms in Female Rats in the 2-Year Gavage Study of TCDD<sup>a</sup>

			8	ť		
	Vehicle Control	3 ng/kg	10 ng/kg	22 ng/kg	46 ng/kg	100 ng/kg
Disposition Summary						
Animals initially in study	53	54	53	53	53	53
Early deaths						
Accidental deaths			5			
Moribund	19	27	15	19	17	17
Natural deaths	9	6	10	15	14	15
Survivors						
Terminal sacrifice	25	21	23	19	22	21
Animals examined microscopically	53	54	53	53	53	53
Alimentary System						
Intestine large, rectum	(52)	(54)	(53)	(52)	(53)	(53)
Carcinoma	(51)	1 (2%)	(52)	(52)	(52)	(52)
Intestine small, jejunum	(51)	(54)	(53)	(53)	(53)	(52)
Leiomyoma	(52)	(54)	(52)	(52)	(52)	(52)
Carcinoma metastatic nancreas	(55)	(34)	(33)	(55)	(55)	(33)
Carcinoma, metastatic, uterus					2(4%)	2 (470)
Cholangiocarcinoma				1 (2%)	3 (6%)	8 (15%)
Cholangiocarcinoma multiple				1 (270)	1 (2%)	17 (32%)
Henatocellular adenoma					1 (2%)	10 (19%)
Henatocellular adenoma multiple					1 (270)	3 (6%)
Hepatocholangioma						2(4%)
Mesenterv	(2)	(2)		(1)	(6)	(8)
Carcinoma, metastatic, uterus	(-)	(_)		(1)	1 (17%)	(0)
Schwannoma malignant	1 (50%)					
Oral mucosa	(2)	(9)	(16)	(16)	(22)	(29)
Gingival, squamous cell carcinoma	1 (50%)	2 (22%)	1 (6%)		4 (18%)	10 (34%)
Pancreas	(51)	(54)	(52)	(53)	(52)	(51)
Carcinoma, metastatic, uterus				~ /	1 (2%)	
Acinus, adenoma					× /	1 (2%)
Acinus, carcinoma						2 (4%)
Stomach, forestomach	(53)	(54)	(53)	(53)	(53)	(53)
Squamous cell carcinoma	1 (2%)				1 (2%)	
Squamous cell papilloma					1 (2%)	1 (2%)
Stomach, glandular	(53)	(54)	(53)	(53)	(53)	(53)
Cardiovascular System						
Heart	(53)	(54)	(53)	(52)	(53)	(52)
Schwannoma malignant		1 (2%)				
Endocrine System						
Adrenal cortex	(53)	(54)	(53)	(53)	(53)	(53)
Adenoma	1 (2%)	(- )	2 (4%)	2 (4%)	1 (2%)	1 (2%)
Carcinoma	- (-/)		= ()	1 (2%)	- (-/)	- (=/)
Pheochromocytoma benign				(=)		1 (2%)
Capsule, sarcoma, metastatic, skeletal musc	le		1 (2%)			()
Adrenal medulla	(53)	(54)	(53)	(53)	(53)	(53)
Pheochromocytoma malignant	<u> </u>	(- )	X 7	()	<>	1 (2%)
Pheochromocytoma benign	3 (6%)	2 (4%)	2 (4%)	3 (6%)	3 (6%)	3 (6%)
	- (*,*)	()	()	- (*,*)	- (***)	- ()

Summary of the Incidence of Neoplasms in Female Rats in the 2-Year Gavage Study of TCDD

	Vehicle Control	3 ng/kg	10 ng/kg	22 ng/kg	46 ng/kg	100 ng/kg
Endocrine System (continued)						
Islets, pancreatic	(51)	(54)	(53)	(53)	(52)	(52)
Adenoma	1 (2%)		2 (4%)			
Carcinoma	1 (2%)					
Parathyroid gland	(46)	(47)	(47)	(44)	(45)	(45)
Adenoma	(52)	(5.4)	(50)	1 (2%)	(52)	(50)
Pituitary gland	(53)	(54)	(52)	(53)	(53)	(52)
Adenoma Meningioma malignant metastatic brain	4 (8%)			1 (2%)		1 (2%)
Pars distalis adenoma	18 (34%)	20 (37%)	26 (50%)	15(28%)	20 (38%)	10 (19%)
Pars distalis, adenoma, multiple	3 (6%)	(	(	()	(	
Pars intermedia, adenoma	2 (4%)					
Thyroid gland	(52)	(54)	(53)	(51)	(53)	(52)
Bilateral, C-cell, adenoma	3 (6%)	3 (6%)	2 (4%)	2 (4%)	1 (2%)	3 (6%)
C-cell, adenoma	17 (33%)	12 (22%)	15 (28%)	14 (27%)	12 (23%)	8 (15%)
C-cell, carcinoma	1 (2%)	1 (2%)			2 (4%)	
Follicular cell, adenoma	1 (2%)				1 (20/)	
Follicular cell, carcinoma					1 (2%)	
General Body System None						
Genital System						
Clitoral gland	(50)	(52)	(53)	(52)	(51)	(53)
Ovary	(51)	(53)	(53)	(52)	(53)	(53)
Granulosa cell tumor malignant	(01)	(00)	(00)	(55)	1 (2%)	(00)
Thecoma benign					1 (2%)	
Periovarian tissue, schwannoma malignant,	,					
metastatic, uterus	1 (2%)					
Uterus	(52)	(53)	(53)	(53)	(53)	(53)
Adenoma		1 (201)	1 (2%)		<b>a</b> (10/)	1 (201)
Carcinoma		1 (2%)		1 (20/)	2 (4%)	1 (2%)
Carcinoma, multiple				1 (2%)	1 (294)	
Leiomyoma	1 (2%)				1 (270)	
Polyn stromal	9 $(17\%)$	7 (13%)	4 (8%)	8 (15%)	9 (17%)	7 (13%)
Polyp stromal, multiple	2 (4%)	(10,0)	. (0/0)	3 (6%)	1 (2%)	1 (2%)
Squamous cell carcinoma					4 (8%)	
Squamous cell carcinoma, multiple					1 (2%)	
Squamous cell papilloma	1 (2%)		1 (2%)			
Cervix, carcinoma		1 (2%)				
Cervix, schwannoma malignant	1 (2%)	(4)				
Vagina		(1)	(1)	(3)	(3)	(1) (1000()
Schwannoma malignant metastatic skin				1 (33%)	1 (55%)	1 (100%)
Squamous cell carcinoma				1 (5570)	1 (33%)	
Hamadan diatis Conduct						
nematopoietic System	(52)	(5.4)	(52)	(52)	(52)	(52)
Done marrow	(33)	(54)	(33)	(53)	(33)	(55)
Deen cervical carcinoma metastatic	(2)	(0)	(3)	(3)	(0)	(9)
thyroid gland	1 (50%)					
Lumbar, carcinoma, metastatic, uterus	. (3070)				1 (17%)	
Lymph node, mandibular	(51)	(54)	(52)	(50)	(51)	(52)

# Summary of the Incidence of Neoplasms in Female Rats in the 2-Year Gavage Study of TCDD

	Vehicle Control	3 ng/kg	10 ng/kg	22 ng/kg	46 ng/kg	100 ng/kg
Hematopoietic System (continued)						
Lymph node, mesenteric	(52)	(53)	(53)	(53)	(53)	(51)
Spleen	(51)	(54)	(53)	(53)	(52)	(52)
Hemangiosarcoma	1 (20)				1 (2%)	
Schwannoma malignant, metastatic, uterus	1 (2%)	(52)	(52)	(40)	(40)	(12)
Carcinoma metastatic lung	(51)	(52) 1 (2%)	(52)	(49)	(40)	(42)
Histiocytic sarcoma, metastatic, lung		1 (270)	1 (2%)			
Integumentary System						
Mammary gland	(53)	(54)	(53)	(53)	(53)	(53)
Adenoma	2 (4%)	()	1 (2%)	()	()	1 (2%)
Carcinoma	3 (6%)	7 (13%)	4 (8%)	2 (4%)	2 (4%)	
Carcinoma, multiple	1 (2%)	1 (2%)				
Fibroadenoma	21 (40%)	26 (48%)	17 (32%)	17 (32%)	24 (45%)	20 (38%)
Fibroadenoma, multiple	12 (23%)	14 (26%)	17 (32%)	12 (23%)	12 (23%)	4 (8%)
Skin	(53)	(54)	(53)	(53)	(53)	(53)
Basal cell carcinoma	1 (2%)		1 (2%)			
Fibroma	1 (2%)		2 (4%)	1 (20/)		1 (2%)
Fibrosarcoma				1 (2%)		
Linoma		1 (2%)		1 (2%)		
Myxosarcoma		1 (270)	1 (2%)			
Schwannoma malignant	1 (2%)		1 (270)	2 (4%)		1 (2%)
Musculoskeletal System						
Bone	(53)	(54)	(53)	(53)	(53)	(53)
Cranium, schwannoma malignant, metastat skin	tic,	(34)	(55)	1 (2%)	(55)	(55)
Periosteum, fibrosarcoma				1 (2%)		
Skeletal muscle			(1)	(2)		
Fibrous histiocytoma				1 (50%)		
Sarcoma			1 (100%)			
Nervous System						
Brain	(53)	(54)	(53)	(53)	(53)	(53)
Glioma malignant			1 (2%)			
Granular cell tumor malignant			1 (2%)			
Respiratory System						
Lung	(53)	(54)	(53)	(52)	(53)	(52)
Alveolar/bronchiolar adenoma, multiple					1 (2%)	
Alveolar/bronchiolar carcinoma		1 (2%)				
Carcinoma, metastatic, mammary gland		1 (2%)				4 (00/)
Carcinoma, metastatic, pancreas	1 (2011)					1 (2%)
Carcinoma, metastatic, thyroid gland	1 (2%)					7 (120/)
Cystic keratinizing epithelioma						7 (13%)
Nenhroblastoma metastatic kidney		1 (2%)				2 (470)
Sarcoma, metastatic, skeletal muscle		1 (270)	1 (2%)			
Mediastinum, sarcoma			1 (2%)			1 (2%)
Nose	(53)	(54)	(53)	(53)	(53)	(53)
Squamous cell carcinoma, metastatic,	× /	× /	× /			~ /
oral mucosa						1 (2%)

Summary of the Incidence of Neoplasms in Female Rats in the 2-Year Gavage Study of TCDD

	Vehicle Control	3 ng/kg	10 ng/kg	22 ng/kg	46 ng/kg	100 ng/kg
Special Senses System						
Ear	(2)		(2)			(2)
Pinna, neural crest tumor	(52) (50%)	(54)	(52)	(52)	(52)	(52)
Squamous cell carcinoma, metastatic, oral mucosa	(55)	(34)	(33)	(32)	(33)	3 (6%)
Zymbal's gland	(1)					- (c,c)
Carcinoma	1 (100%)					
Urinary System						
Kidney	(53)	(54)	(53)	(53)	(53)	(53)
Nephroblastoma		2 (4%)	1 (2%)			
Schwannoma malignant, metastatic, uterus	1 (2%)			1 (20())		
Stromal nephroma	(52)	(52)	(52)	1 (2%)	(52)	(52)
Polyn	(32)	(33)	(55)	(32)	(55)	(33)
Schwannoma malignant metastatic uterus	1 (2%)	1 (270)				
Squamous cell carcinoma, metastatic, uteru	15				1 (2%)	
Transitional epithelium, carcinoma					1 (2%)	
Systemic Lesions						
Multiple organs <sup>b</sup>	(53)	(54)	(53)	(53)	(53)	(53)
Leukemia granulocytic		× /	1 (2%)		· /	
Lymphoma malignant		1 (2%)	1 (2%)		1 (2%)	2 (4%)
Neoplasm Summary						
Total animals with primary neoplasms <sup>c</sup>	47	54	48	44	50	50
Total primary neoplasms	116	105	106	89	116	131
Total animals with benign neoplasms	45	49	45	40	46	42
Total benign neoplasms	102	86	92	78	88	87
Total animals with malignant neoplasms	13	19	13	11	24	34
Total malignant neoplasms	13	19	14	11	28	44
Total animals with metastatic neoplasms	2	3	2	2	3	6
Iotal metastatic neoplasms	6	3	3	3	6	1
Iotal animals with uncertain neoplasms -	1					
Total uncertain neoplasms	1					
rotar uncertain neopiasins	1					

a Number of animals examined microscopically at the site and the number of animals with neoplasm
 b Number of animals with any tissue examined microscopically
 b Primary neoplasms: all neoplasms except metastatic neoplasms

Summary of the Incidence of Neoplasms in Female Rats in the 2-Year Gavage Study of TCDD (Stop-Exposure)<sup>a</sup>

	Vehicle	e Control	100	ng/kg	100 (Stop-E	ng/kg Exposure)
Disposition Summary						
Animals initially in study	53		53		50	
Early deaths						
Moribund	19		17		16	
Natural deaths	9		15		13	
Survivors						
Terminal sacrifice	25		21		21	
Animals examined microscopically	53		53		50	
Alimentary System						
Intestine large, rectum	(52)		(53)		(50)	
Fibrous histiocytoma metastatic skin	(52)		(55)		(30)	(2%)
Intestine small jejunum	(51)		(52)		(50)	(270)
Schwannoma malignant metastatic heart	(51)		(32)		(30)	(2%)
Liver	(53)		(53)		(50)	(270)
Carcinoma metastatic pancreas	(00)		2	(4%)	(20)	
Cholangiocarcinoma				(15%)	1	(2%)
Cholangiocarcinoma multiple			17	(32%)	1	(2%)
Cholangioma			17	(5270)	1	(2%)
Fibrous histiocytoma, metastatic, skin					1	(2%)
Hepatocellular adenoma			10	(19%)	2	(4%)
Hepatocellular adenoma, multiple			3	(6%)	_	
Hepatocholangioma			2	(4%)		
Schwannoma malignant, metastatic, heart			_	(1))	1	(2%)
Mesentery	(2)		(8)		(1)	(270)
Schwannoma malignant	(_)	(50%)	(0)		(1)	
Oral mucosa	(2)	(0070)	(29)		(11)	
Gingiyal squamous cell carcinoma	(_)	(50%)	10	(34%)	(11)	(45%)
Pancreas	(51)	(3070)	(51)	(31/0)	(49)	(1070)
Acinus adenoma	(31)		(51)	(2%)	(4))	
Acinus, carcinoma			2	(2%)	1	(2%)
Stomach forestomach	(53)		(53)	(470)	(50)	(270)
Sauamous cell carcinoma	(33)	(2%)	(55)		(50)	
Squamous cell papilloma	1	(270)	1	(2%)		
Stomach, glandular	(53)		(53)	(270)	(50)	
Tongue	(55)		(55)		(30)	
Squamous cell carcinoma					(1)	(100%)
Cardiovascular System						
Blood vessel	(53)		(53)			(49)
Aarta fibrous histiogytoma metastatic skin	(55)		(55)		1	(49)
Heart	(53)		(52)		(50)	(270)
Carainama matastatia mammary gland	(55)		(32)		(30)	(29/)
Schwannoma malignant					3	(276) (6%)
Endocrine System						
Adrenal cortex	(53)		(53)		(50)	
Adenoma	(55)	(2%)	(55)	(2%)	(30)	
Aucholita Dheochromocytoma henica	1	(270)	1	(270)		
Cansula, schwannoma malignant, matagtatic, haart			1	(270)	1	(2%)
Capsure, senwannonna mangnant, metastatic, neart					1	(270)

# Summary of the Incidence of Neoplasms in Female Rats in the 2-Year Gavage Study of TCDD (Stop-Exposure)

	Vehicle	Control	100	ng/kg	100 (Stop-E	ng/kg xposure)
Endocrine System (continued)						
Adrenal medulla	(53)		(53)		(50)	
Pheochromocytoma complex					1	(2%)
Pheochromocytoma malignant			1	(2%)		
Pheochromocytoma benign	3	(6%)	3	(6%)	1	(2%)
Islets, pancreatic	(51)		(52)		(49)	
Adenoma	1	(2%)				
Carcinoma	1	(2%)				
Parathyroid gland	(46)		(45)		(44)	
Pituitary gland	(53)		(52)		(50)	
Adenoma	4	(8%)	1	(2%)		
Pars distalis, adenoma	18	(34%)	10	(19%)	19	(38%)
Pars distalis, adenoma, multiple	3	(6%)				
Pars intermedia, adenoma	2	(4%)				
Thyroid gland	(52)		(52)		(49)	
Bilateral, C-cell, adenoma	3	(6%)	3	(6%)	2	(4%)
C-cell, adenoma	17	(33%)	8	(15%)	11	(22%)
C-cell, carcinoma Follicular cell, adenoma	1	(2%) (2%)			1	(2%)
None						
Genital System						
Clitoral gland	(50)		(53)		(49)	
Ovary	(51)		(53)		(49)	
Periovarian tissue, schwannoma malignant,						
metastatic, uterus	1	(2%)				
Carcinoma, metastatic, uterus					1	(2%)
Uterus	(52)		(53)	(	(50)	
Carcinoma			1	(2%)	2	(4%)
Leiomyoma	1	(2%)	-	(100)		
Polyp stromal	9	(17%)	1	(13%)	3	(6%)
Polyp stromal, multiple	2	(4%)	1	(2%)	1	(2%)
Squamous cell carcinoma	1	(20/)			2	(4%)
Squamous cell papilloma	1	(2%)				
Vagina	1	(270)	(1)			
Schwannoma malignant			(1)	(100%)		
Hematopoietic System						
Bone marrow	(53)		(53)		(50)	
Fibrous histiocytoma, metastatic, skin					1	(2%)
Lymph node	(2)		(9)		(5)	
Deep cervical, carcinoma, metastatic,						
thyroid gland	1	(50%)	/			
Lymph node, mandibular	(51)		(52)		(49)	
Lymph node, mesenteric	(52)		(51)		(49)	
Fibrous histiocytoma, metastatic, skin					1	(2%)

# Summary of the Incidence of Neoplasms in Female Rats in the 2-Year Gavage Study of TCDD (Stop-Exposure)

	Vehicle	e Control	100	ng/kg	100 (Stop-E	ng/kg Exposure)
Hematopoietic System (continued) Spleen Fibrous histiocytoma, metastatic, skin Hemangiosarcoma Schwannoma malignant, metastatic, heart Schwannoma malignant, metastatic, uterus Thymus Fibrous histiocytoma, metastatic, skin Thymoma malignant	(51) 1 (51)	(2%)	(52) (42)		(49) 1 1 1 (49) 1 1	(2%) (2%) (2%) (2%)
Integumentary System Mammary gland Adenolipoma Adenoma Carcinoma Carcinoma, multiple Fibroadenoma, multiple Skin Basal cell carcinoma Fibroma Fibroma Fibrous histiocytoma Liposarcoma Schwannoma malignant	(53) 2 3 1 21 12 (53) 1 1	(4%) (6%) (2%) (40%) (23%) (2%) (2%) (2%)	(53) 1 20 4 (53) 1 1	(2%) (38%) (8%) (2%) (2%)	(50) 1 19 13 (50) 1 1 1	(2%) (2%) (38%) (26%) (2%) (2%) (2%)
Musculoskeletal System Bone Squamous cell carcinoma, metastatic, tongue Skeletal muscle Fibrous histiocytoma, metastatic, skin	(53)		(53)		(50) 1 (1) 1	(2%) (100%)
Nervous System Brain	(53)		(53)		(50)	
Respiratory System Lung Carcinoma, metastatic, mammary gland Carcinoma, metastatic, pancreas Carcinoma, metastatic, thyroid gland Cystic keratinizing epithelioma Cystic keratinizing epithelioma, multiple Fibrous histiocytoma, metastatic, skin Mediastinum, sarcoma Schwannoma malignant, metastatic, heart Nose Squamous cell carcinoma, metastatic, oral mucosa	(53) 1 (53)	(2%)	(52) 1 7 2 1 (53) 1	(2%) (13%) (4%) (2%) (2%)	(50) 1 1 (50)	(2%) (2%) (2%)

Summary of the Incidence of Neoplasms in Female Rats in the 2-Year Gavage Study of TCDD (Stop-Exposure)

	Vehicle	e Control	100 ng/kş	g 100 ng/kg (Stop-Exposure)	
Special Senses System					
Ear	(2)		(2)		
Pinna, neural crest tumor	1	(50%)			
Harderian gland	(53)		(53)	(50)	
Squamous cell carcinoma, metastatic,			2 ((0/)		
oral mucosa	(1)		3 (6%)		
Zymbal s gland	(1)	(100%)			
	1	(100%)			
Urinary System					
Kidney	(53)		(53)	(50)	
Fibrous histiocytoma, metastatic, skin				1 (2%)	
Schwannoma malignant, metastatic, uterus	1	(2%)			
Urinary bladder	(52)		(53)	(50)	
Schwannoma malignant, metastatic, uterus	1	(2%)			
Systemic Lesions					
Multiple organs <sup>b</sup>	(53)		(53)	(50)	
Adenolipoma	()		()	1 (2%)	
Lymphoma malignant			2 (4%)	1 (2%)	
Neoplasm Summary					
Total animals with primary neoplasms <sup>c</sup>	47		50	46	
Total primary neoplasms	116		131	98	
Total animals with benign neoplasms	45		42	41	
Total benign neoplasms	102		87	74	
Total animals with malignant neoplasms	13		34	19	
Total malignant neoplasms	13		44	24	
Total animals with metastatic neoplasms	2		6	5	
Total metastatic neoplasms	6		7	19	
Total animals with uncertain neoplasms -					
benign or malignant	1				
Total uncertain neoplasms	1				

a Number of animals examined microscopically at the site and the number of animals with neoplasm
 b Number of animals with any tissue examined microscopically
 b Primary neoplasms: all neoplasms except metastatic neoplasms

TABLE A2 Individual A

Individual Animal Tumor Patholog	gy of Fe	ma	le I	Rat	s i	n tl	he	2-\	lea	ar (	Ga	vaş	ge S	Stu	dy	of	тс	CD	D:	V	ehi	icle	e C	on	tro	l		
	1	1	3	3	3	4	4	4	4	5	5	5	5	6	6	6	6	6	6	6	6	6	6	6	6	7	7	7
Number of Days on Study	6	8	5	6	6	6	7	8	9	1	2	3	4	1	3	4	4	4	4	4	6	6	7	9	9	0	1	2
	9	5	1	1	6	9	4	7	2	2	6	3	8	0	1	5	6	6	7	8	6	8	5	4	7	1	7	2
	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Carcass ID Number	8	2	2	2	8	4	8	0	3	6	1	0	8	7	6	2	7	8	4	3	0	8	0	2	4	7	6	3
	6	5	1	4	2	2	4	7	5	3	3	6	3	3	1	3	5	7	1	9	4	1	9	2	3	1	4	4
Alimentary System																												
Esophagus	+	+	+	+	+	+	+	+	+	+	+	+	+	+	М	+	+	+	+	+	+	+	+	+	+	+	+	+
intestine large, colon	+	+	М	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
intestine large, rectum	+	+	+	+	+	+	+	+	+	+	+	+	+	M	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Intestine large, cecum	+	+	M	+	+	+	+	+	+	+	+	+	+	M	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Intestine small duodenum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
intestine small jejunum	+	+	м	+	+	+	+	+	+	+	+	+	+	Ń	+	+	+	+	+	+	+	+	+	+	+	+	+	+
ntestine small, jejunum	+	+	M	+	+	+	+	+	+	+	+	+	+	M	+	+	+	+	+	+	+	+	+	+	+	+	+	+
iver	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Mesentery	+		'			'	'	'	'				'		'	'				+			'	'			'	
Schwannoma malignant	v																			'								
Oral mucosa	Л				+																							
Gingival squamous cell carcinoma					v																							
Ongival, squamous cen caremonia	1	+	м	+	л _	-	-	+	-	+	+	+	+	м	+	+	+	-	-	-	+	+	+	+	+	+	+	+
alivery glands	+	+	1VI +	+	+	+	+	+	+	M	+	+	+	1VI +	M	+	+	+	+	+	+	+	+	+	+	+	+	+
Sanvary granus		+ +	+ +	+	+	+ +	+ +	+ +	+ +	1VI	+	+	+	+ +	111	+ +	+ +	+ +	+ +	+ +	+ +	+	+	+ +	+	+	+	+ +
Someona call concinents	т	Ŧ	т	Ŧ	Ŧ	Ŧ	Ŧ	т	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	т	Ŧ	Ŧ	$\mathbf{v}$	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	т	Ŧ	Ŧ	т
Squamous cell carcinoma	1																л 											
	т	-	т	Ŧ	Ŧ	Ŧ	Ŧ	т	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	т	-	- -	Ŧ	Ŧ	Ŧ	-	- -	Ŧ	- -	Ŧ	т	-	Ŧ	т
lootn		+													+	+				+	+		+			+		
Cardiovascular System																												
Blood vessel	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Ieart	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Endocrine System																												
Adrenal cortex	+	+	+	+	+	$^+$	$^+$	+	$^+$	+	+	+	+	+	+	$^+$	+	+	+	+	$^+$	+	$^+$	+	+	+	$^+$	+
Adenoma																												
Adrenal medulla	+	+	+	+	+	$^+$	$^+$	+	$^+$	+	+	+	+	$^+$	+	$^+$	+	+	+	+	$^+$	+	$^+$	+	+	+	$^+$	+
Pheochromocytoma																												
slets, pancreatic Adenoma	+	+	М	+	+	+	+	+	+	+	+	+	+	М	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Carcinoma																х												
Parathyroid gland	+	м	+	+	+	+	+	М	+	+	+	+	+	+	М	+	+	+	М	+	+	м	+	+	+	+	+	+
Pituitary gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Adenoma	1														ÿ	ÿ			x									
Pars distalis adenoma							x			x					1	1	x	x	1						x	x		x
Pars distalis, adenoma multiple							1			Λ							<b>2 1</b>	1							Λ	Δ		Λ
Pars intermedia adenoma																											x	
Thyroid gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	м	+	+	+	+	+	+	+	+	+	+	+	л +	+
Bilateral C-cell adenoma	т	1-						1	'						141	1					$\mathbf{v}$	v	'					
C cell adenoma							v		v		v	v				v					л	л	v	v	v			
C-cell carcinoma							л		л		л	л				л							л	л	л			
C-teri, careinoma																												

+: Tissue examined microscopically A: Autolysis precludes examination

M: Missing tissue I: Insufficient tissue

X: Lesion present Blank: Not examined

TABLE A2 Individual Animal Tu

Individual Animal Tumor Patholog	gy of Fei	na	le l	<b>k</b> at	S 11	n t	ne .	2-1	Yea	ar (	Ga	vaş	ge s	Stu	dy	01	10	ĴD	D:	V	eh	icl	e C	on	trol	
	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	
Number of Days on Study	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	
<i>v v</i>	1	1	1	1	1	1	1	1	1	1	2	2	2	2	2	2	2	3	3	3	3	3	3	3	3	
	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	Tot
Carcass ID Number	0	0	1	3	3	3	6	9	9	9	1	1	3	4	4	8	9	0	0	1	4	4	4	9	9	Tissue
	2	3	4	6	7	8	2	2	3	8	0	1	1	0	4	8	1	1	5	2	7	8	9	6	7	Tumor
Alimentary System																										
Esophagus	+	+	+	+	+	+	+	+	+	+	+	+	$^+$	+	$^+$	$^+$	$^+$	+	$^+$	$^+$	+	+	+	+	+	5
Intestine large, colon	+	+	+	+	+	$^+$	$^+$	+	+	+	+	+	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	+	+	+	+	+	5
Intestine large, rectum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	5
Intestine large, cecum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	5
Intestine small duodenum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	5
Intestine small jejunum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	5
Intestine small ileum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	5
Liver	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	5
Mesentery	1							'					'		'	'				'						5
Schwannoma malignant																										
Oral mucosa									+																	
Gingival, squamous cell carcinoma																										
Pancreas	+	+	+	+	+	$^+$	+	+	+	+	+	+	+	+	+	+	+	+	+	$^+$	+	+	+	+	+	5
Salivary glands	+	+	+	+	+	$^+$	+	+	+	+	+	+	+	+	+	+	+	+	+	$^+$	+	+	+	+	+	5
Stomach, forestomach	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	5
Squamous cell carcinoma																										-
Stomach glandular	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	5
Tooth			+	+	+			+	+		+	+		+			+									1
Cardiovascular System																										
Blood vessel	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	5
Heart	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	5
Endocrine System																										
Adrenal cortex	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	5
Adenoma											Х															
Adrenal medulla	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	5
Pheochromocytoma benign												Х		Х								Х				
Islets, pancreatic	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	5
Adenoma	Х																									
Carcinoma																										
Parathyroid gland	+	+	$^+$	$^+$	$^+$	$^+$	Μ	$^+$	+	+	+	+	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	+	+	+	$^+$	Μ	4
Pituitary gland	+	+	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	+	+	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	+	$^+$	$^+$	+	5
Adenoma																					Х					
Pars distalis, adenoma		Х	Х		Х				Х	Х	Х	Х					Х			Х				Х	Х	1
Pars distalis, adenoma, multiple													Х		Х			Х								
Pars intermedia, adenoma					Х																					
Thyroid gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	5
Bilateral, C-cell, adenoma										x																5
C-cell adenoma			x		x					- 1	x			x		x		x	x		x				х	1
C-cell carcinoma			11								21			- 1	x	2 <b>x</b>			- 1		11					1
c con, caromonia															11											

individual Annial Tuniol Tathology	or rei											6	,															
Number of Days on Study	1 6 9	1 8 5	3 5 1	3 6 1	3 6 6	4 6 9	4 7 4	4 8 7	4 9 2	5 1 2	5 2 6	5 3 3	5 4 8	6 1 0	6 3 1	6 4 5	6 4 6	6 4 6	6 4 7	6 4 8	6 6 6	6 6 8	6 7 5	6 9 4	6 9 7	7 0 1	7 1 7	7 2 2
Carcass ID Number	0 8 6	0 2 5	0 2 1	0 2 4	0 8 2	0 4 2	0 8 4	0 0 7	0 3 5	0 6 3	0 1 3	0 0 6	0 8 3	0 7 3	0 6 1	0 2 3	0 7 5	0 8 7	0 4 1	0 3 9	0 0 4	0 8 1	0 0 9	0 2 2	0 4 3	0 7 1	0 6 4	0 3 4
General Body System None																												
Genital System																												
Clitoral gland	+	+	+	+	+	+	+	+	+	+	+	+	М	М	+	+	+	+	+	+	+	+	+	+	+	М	+	+
Dvary	+	+	М	+	+	+	+	+	+	+	+	+	+	Μ	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Periovarian tissue, schwannoma malignant, metastatic, uterus												Х																
Dviduct																											+	
Jterus	+	+	+	+	+	+	+	+	+	+	+	+	+	М	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Leiomyoma Polyp stromal Polyp stromal, multiple Squamous cell papilloma																X X									X	х		
Cervix, schwannoma malignant												Х																
Iematopoietic System																												
Bone marrow	+	+	+	+	+	+	+	+	+	+	$^+$	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
ymph node Deep cervical, carcinoma, metastatic, thyroid gland																											+	
ymph node, mandibular	+	+	+	+	+	+	+	+	+	М	+	+	+	+	М	+	+	+	+	+	+	+	+	+	+	+	+	+
ymph node, mesenteric	+	+	Μ	+	+	+	$^+$	+	+	$^+$	$^+$	+	$^+$	+	$^+$	$^+$	$^+$	$^+$	+	$^+$	$^+$	+	$^+$	$^+$	+	+	$^+$	+
pleen	+	+	М	+	+	+	+	М	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Schwannoma malignant, metastatic, uterus Thymus	+	+	+	+	+	+	+	+	+	+	+	X +	+	+	+	+	+	+	+	+	+	М	+	+	+	М	+	+
ntegumentary System																												
Mammary gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Adenoma Carcinoma						x									Х													
Carcinoma, multiple																									Х			
Fibroadenoma		Х		Х		Х	Х									Х		Х	Х				Х		Х		Х	
Fibroadenoma, multiple									Х		Х						Х				Х	Х		Х				Х
Skin	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Basal cell carcinoma Fibroma													х						Х									
Schwannoma malignant													-													Х		
Musculoskeletal System																												
Bone	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Nervous System																												
Brain	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Respiratory System																												
Respiratory System	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Respiratory System Jung Carcinoma, metastatic, thyroid gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
<b>Respiratory System</b> Lung Carcinoma, metastatic, thyroid gland Nose	+ +	+	++	+	+	+	++	+	+	+	+	++	+	++	+	+	++	++	+	+	+	+	+	++	+	+	+	+

Individual Animal Tumor Pathology of	of Fei	ma	le ]	Rat	ts i	n tl	he	2-1	Yea	ar (	Ga	vaş	ge S	Stu	ldy	of	T (	CD	D:	V	eh	icle	e C	on	trol	<u>l</u>	
Number of Days on Study	7 3 1	7 3 2	7 3 2	7 3 2	7 3 2	7 3 2	7 3 2	7 3 2	7 3 3																		
Carcass ID Number	0 0 2	0 0 3	0 1 4	0 3 6	0 3 7	0 3 8	0 6 2	0 9 2	0 9 3	0 9 8	0 1 0	0 1 1	0 3 1	0 4 0	0 4 4	0 8 8	0 9 1	0 0 1	0 0 5	0 1 2	0 4 7	0 4 8	0 4 9	0 9 6	0 9 7	Tota Tissues Tumor	al s/ rs
General Body System None																											
<b>Genital System</b> Clitoral gland Ovary Periovarian tissue, schwannoma malignant,	+ +	+ +	+ +	+++	+++	+ +	+ +	+ +	+ +	+++	+ +	+++	+ +	+ +	+ +	+ +	+ +	+ +	+ +	+ +	+ +	+ +	+ +	+ +	+ +	5 5	0
Oviduct Uterus Leiomyoma Polyp stromal Polyp stromal, multiple Squamous cell papilloma	+	+	++	+ X	+	+	+	+	+	+ X	+	+ X X	+	+ X	+	+	+	+	+ X	+ X	+	+	+ X	+ X	+	5	1 2 2 1 9 2 1
Cervix, schwannoma malignant Hematopoietic System Bone marrow	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	5	1
Lymph node Deep cervical, carcinoma, metastatic, thyroid gland Lymph node, mandibular Lymph node, mesenteric	++	+++	+++	+++	++++	++++	++++	++++	++++	+++	+++	+++	+++	+++	+ X + +	++++	++++	++++	++++	++++	++++	++++	++++	++++	+++	5	1
Spleen Schwannoma malignant, metastatic, uterus Thymus	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	5	1
Integumentary System Mammary gland Adenoma Carcinoma Carcinoma multiple	+	+	+ X	+	+	+	+	+	+	+	+ X	+	+	+	+	+	+	+	+	+	+	+	+ X	+	+	5	3 2 3 1
Fibroadenoma Fibroadenoma, multiple Skin Basal cell carcinoma Fibroma Schwannoma malignant	+	X +	+	X +	X +	+	+	X +	X +	+	X +	X +	X +	X +	X +	+	+	X +	X +	X +	+	X +	X +	+	X +	2 1 5	1 2 3 1 1
Musculoskeletal System Bone	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	5	3
Nervous System Brain	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	5	3
Respiratory System Lung Carcinoma, metastatic, thyroid gland Nose Trachea	+ + +	+ + +	+ + +	+++++	+++++	+ + +	+ + +	+++++	+++++	+++++	+ + +	+++++	+ + +	+++++	+ X + +	+ + +	+++++	+++++	+ + +	+++++	+ + +	+++++	+++++	+++++	+ + +	5 5 5	1 1 3 2

TABLE A2

Individual Animal Tumor Pathology	of Fer	nal	le F	Rat	s ir	n th	ie 2	2-Y	ea	r (	Gav	vag	je S	Stu	dy	of	тс	CD	D:	V	ehi	icle	e C	on	tro	1		
	1	1	3	3	3	4	4	4	4	5	5	5	5	6	6	6	6	6	6	6	6	6	6	6	6	7	7	7
Number of Days on Study	6	8	5	6	6	6	7	8	9	1	2	3	4	1	3	4	4	4	4	4	6	6	7	9	9	0	1	2
	9	5	1	1	6	9	4	7	2	2	6	3	8	0	1	5	6	6	7	8	6	8	5	4	7	1	7	2
	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Carcass ID Number	8	2	2	2	8	4	8	0	3	6	1	0	8	7	6	2	7	8	4	3	0	8	0	2	4	7	6	3
	6	5	1	4	2	2	4	7	5	3	3	6	3	3	1	3	5	7	1	9	4	1	9	2	3	1	4	4
Special Senses System																												
Ear																										+	+	
Pinna, neural crest tumor																										Х		
Eye	+	$^+$	$^+$	+	$^+$	+	+	+	$^+$	$^+$	$^+$	$^+$	$^+$	+	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	+	$^+$	+
Harderian gland	+	+	+	$^+$	+	+	+	+	+	+	+	$^+$	+	+	+	$^+$	+	+	+	+	+	$^+$	$^+$	$^+$	+	+	+	+
Zymbal's gland																												
Carcinoma																												
Urinary System																												
Kidney	+	$^+$	$^+$	$^+$	+	+	+	+	+	$^+$	$^+$	$^+$	$^+$	+	+	$^+$	+	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	+	$^+$	+
Schwannoma malignant, metastatic, uterus												Х																
Ureter																												
Urinary bladder	+	$^+$	+	+	+	+	+	+	+	$^+$	$^+$	+	+	М	+	+	+	+	+	+	$^+$	$^+$	$^+$	+	+	+	$^+$	+
Schwannoma malignant, metastatic, uterus												Х																
Systemic Lesions																												
Multiple organs	+	$^+$	$^+$	+	+	+	+	+	$^+$	$^+$	$^+$	$^+$	+	+	+	+	+	+	+	$^+$	$^+$	$^+$	$^+$	+	+	+	$^+$	+

Individual Animal Tumor Pathology	of Fer	na	le l	Rat	ts i	n t	he	2-1	Yea	ar (	Ga	vaş	ge (	Stu	ıdy	of	T	CD	D:	V	eh	icle	e C	on	trol	
Number of Days on Study	7 3																									
	1	1	1	1	1	1	1	1	1	1	2	2	2	2	2	2	2	3	3	3	3	3	3	3	3	
	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	Total
Carcass ID Number	0 2	0 3	1 4	3 6	3 7	3 8	6 2	9 2	9 3	9 8	1 0	1 1	3 1	4 0	4 4	8 8	9 1	0 1	0 5	1 2	4 7	4 8	4 9	9 6	9 7	Tissues/ Tumors
Special Senses System																										2
Ear Pinna, neural crest tumor																										2
Eye	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Harderian gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Zymbal's gland		+																								1
Carcinoma		Х																								1
Urinary System																										
Kidney Schwannoma malianant, metastatic, uterus	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Ureter											+															1
Urinary bladder	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	52
Schwannoma malignant, metastatic, uterus																										1
Systemic Lesions																										
Multiple organs	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53

Individual Animal Tumor Patholog	gy of Fei	ma	le l	Rat	ts i	n tl	he	2-}	Yea	ar (	Ga	vag	e S	Stu	dy	of	ТС	CD	D:	3	ng	/kg	5						
Number of Days on Study	2 0	2 4	2 6	2 6	3 4	36	3 9	4 4	4 4	4	4 8	4 9	4 9	5 2	5 4	5 5	5 5	5 5	5 5	5 5	5 5	5 7	5 7	6 0	6 0	6 0	6 1	6 2	6 4
	0	1	4	4	1	1	4	8	9	2	5	0	2	6	8	6	6	6	6	7	7	8	8	4	4	7	3	0	7
	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
Carcass ID Number	6 5	1 4	2 8	4 1	2 0	4 2	1 7	1 5	0 6	8 1	5 5	1 2	2 9	0 8	0 3	0 2	0 5	1 3	8 4	5 4	6 3	1 6	7 9	8 2	8 3	7 7	6 4	6 8	1 0
Alimentary System																													
Esophagus	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Intestine large, colon	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	$^+$	Μ	$^+$	+	+	+	+	+	$^+$	$^+$	+	+
Intestine large, rectum	+	+	+	+	+	+	$^+$	+	+	+	$^+$	+	+	+	+	+	$^+$	$^+$	+	+	+	+	+	+	+	+	$^+$	$^+$	+
Carcinoma																													
Intestine large, cecum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	$^+$	М	$^+$	+	+	+	+	+	+	$^+$	$^+$	+
Intestine small, duodenum	+	+	+	+	$^+$	+	+	+	+	+	+	+	+	+	+	+	+	$^+$	$^+$	$^+$	$^+$	+	+	$^+$	$^+$	$^+$	$^+$	$^+$	+
Intestine small, jejunum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Intestine small, ileum	+	+	+	+	$^+$	+	+	+	+	+	+	+	+	+	+	+	+	$^+$	Μ	$^+$	+	$^+$	$^+$	+	$^+$	$^+$	$^+$	$^+$	+
Liver	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Mesentery	+																											+	
Oral mucosa								+	+												+								+
Gingival, squamous cell carcinoma																													Х
Pancreas	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Salivary glands	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Stomach, forestomach	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Stomach, glandular Footh	+	+	+	+	+	+	+	+	+ +	+	+	+	+	+	+	+ +	+	+	+	+ +	+	+	+ +	+	+	+ +	+ +	+ +	+
Cardiovascular System																													
Blood vessel	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Heart Schwannoma malignant	+	+	+	+	+	+	+	+	$^+$ X	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Endocrine System																													
Adrenal cortex	+	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	+	+	$^+$	+	$^+$	+	+	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	+
Adrenal medulla	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	$^+$	+	+	+	+	+	+	+	+	$^+$	+	+
Pheochromocytoma benign																													
Islets, pancreatic	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Parathyroid gland	+	+	+	+	+	+	+	М	+	+	+	+	+	+	+	+	+	М	+	+	+	+	+	+	М	+	+	+	+
Pituitary gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Pars distalis, adenoma																								Х	Х		Х		Х
Fhyroid gland Bilateral, C-cell, adenoma	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
C-cell, adenoma C-cell, carcinoma										Х			Х			Х								X X					
General Body System None																													
Genital System																													
Clitoral gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	М	+	+	+	+	+	+	+	$^+$	+	+
Ovary	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	М	$^+$	+	+	+	+	+	+	$^+$	+	+
Oviduct																													
Uterus	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	М	$^+$	+	+	+	+	+	+	$^+$	+	+
Carcinoma																													
Polyp stromal														Х								Х							
~ 1																													
Cervix, carcinoma																													

Individual Animal Tumor Patholog	gy of Fer	na	le l	Rat	ts i	n t	he	2-1	Yea	ar (	Ga	vaş	ge S	Stu	dy	of	T (	CD	D:	3	ng	g/kg	g			
Number of Days on Study	6 4 8	6 8 1	7 1 8	7 2 9	7 3 1	7 3 2	7 3 2	7 3 2	7 3 3																	
	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	Total
Carcass ID Number	9	1 8	6 2	2 6	0 4	2	2 4	3 1	3 4	5 1	6 1	0	6	9 8	1	2 5	5 3	0 7	2	2 3	3 0	3	4 4	4 5	9 7	Tumors
Alimentary System																										
Esophagus	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	54
Intestine large, colon	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Intestine large, rectum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	54
Latesting large accure				X																						1
Intestine large, cecum	- -	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+ +	55
Intestine small jejunum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	54
Intestine small ileum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Liver	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	54
Mesenterv																										2
Oral mucosa					+								+					+	+				+			9
Gingival, squamous cell carcinoma													Х													2
Pancreas	+	$^+$	$^+$	+	+	+	+	+	+	+	+	+	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	+	+	+	+	+	54
Salivary glands	+	$^+$	+	+	+	+	+	+	+	+	+	+	$^+$	+	+	$^+$	$^+$	+	$^+$	$^+$	+	+	+	+	+	54
Stomach, forestomach	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	54
Stomach, glandular	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	54
Tooth						+				+		+	+			+		+	+	+			+			16
Cardiovascular System																										
Blood vessel	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	54
Heart Schwannoma malignant	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	54 1
Endoaring System																										
A dramal contray																										54
Adrenal medulla	- -	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	54
Pheochromocytoma benign	1											'			x		'		x	'						24
Islets pancreatic	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	54
Parathyroid gland	+	+	+	+	+	+	+	М	+	+	+	+	+	+	M	+	+	+	+	+	+	+	M	+	M	47
Pituitary gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	54
Pars distalis, adenoma					Х	Х	Х	Х	Х	Х		Х	Х		Х	Х			Х	Х	Х	Х	Х		Х	20
Thyroid gland	+	$^+$	$^+$	+	+	+	+	+	+	$^+$	+	+	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	+	+	+	+	+	54
Bilateral, C-cell, adenoma									Х		Х										Х					3
C-cell, adenoma			Х			Х	Х								Х		Х	Х		Х			Х			12
C-cell, carcinoma																										1
General Body System None																										
Genital System																										
Clitoral gland	+	+	М	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	52
Ovary	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Oviduct				+																						1
Uterus	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Carcinoma														Х												1
Polyp stromal					Х				Х			Х						Х							Х	7
Cervix,carcinoma											Х															1
Vagina																										1

Individual Animal Tumor Pathology	of Fei	na	le l	<b>l</b> at	s ii	n tl	1e I	2-1	l ea	ar (	Ga	vag	ge S	Stu	dy	of	T	ĴD	D:	3	ng	/kg	5						
Number of Days on Study	2 0 0	2 4 1	2 6 4	2 6 4	3 4 1	3 6 1	3 9 4	4 4 8	4 4 9	4 6 2	4 8 5	4 9 0	4 9 2	5 2 6	5 4 8	5 5 6	5 5 6	5 5 6	5 5 6	5 5 7	5 5 7	5 7 8	5 7 8	6 0 4	6 0 4	6 0 7	6 1 3	6 2 0	6 4 7
Carcass ID Number	1 6 5	1 1 4	1 2 8	1 4 1	1 2 0	1 4 2	1 1 7	1 1 5	1 0 6	1 8 1	1 5 5	1 1 2	1 2 9	1 0 8	1 0 3	1 0 2	1 0 5	1 1 3	1 8 4	1 5 4	1 6 3	1 1 6	1 7 9	1 8 2	1 8 3	1 7 7	1 6 4	1 6 8	1 1 0
Hematopoietic System																													
Bone marrow	+	+	+	+	+	+	+	+	+	+	+	+	$^+$	+	+	$^+$	+	$^+$	+	+	$^+$	+	+	$^+$	+	+	+	$^+$	+
Lymph node			$^+$		+			$^+$				+	$^+$										+						
Lymph node, mandibular	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	$^+$	+	+	+	+	+	$^+$	$^+$	+	+	$^+$	+
Lymph node, mesenteric	+	+	$^+$	$^+$	+	+	+	+	$^+$	+	+	+	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	Μ	$^+$	$^+$	+	+	$^+$	$^+$	$^+$	$^+$	$^+$	+
Spleen	+	+	$^+$	$^+$	+	+	+	$^+$	$^+$	+	+	+	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	+	+	$^+$	$^+$	$^+$	$^+$	$^+$	+
Thymus	+	+	$^+$	$^+$	+	+	+	$^+$	$^+$	+	+	+	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	+	+	М	$^+$	$^+$	$^+$	$^+$	+
Carcinoma, metastatic, lung										Х																			
Integumentary System																													
Mammary gland	+	$^+$	$^+$	$^+$	$^+$	+	+	+	$^+$	$^+$	+	+	$^+$	+	+	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	+	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$
Carcinoma		Х			Х																		Х						
Carcinoma, multiple																													
Fibroadenoma		Х				Х	Х	Х	Х				Х		Х	Х	Х		Х	Х		Х	Х		Х	Х		Х	
Fibroadenoma, multiple				Х							Х			Х				Х			Х			Х					
Skin	+	+	$^+$	$^+$	+	+	$^+$	+	$^+$	+	+	+	$^+$	+	+	$^+$	$^+$	$^+$	+	$^+$	+	+	+	+	$^+$	+	$^+$	$^+$	+
Lipoma																													
Musculoskeletal System																													
Bone	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Nervous System																													
Brain	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Respiratory System																													
Lung	+	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$
Alveolar/bronchiolar carcinoma										Х																			
Carcinoma, metastatic, mammary gland					Х																								
Nephroblastoma, metastatic, kidney			Х																										
Nose	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Trachea	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Special Senses System																													
Eye	+	+	+	+	+	+	+	+	+	+	+	+	$^+$	+	+	$^+$	+	$^+$	+	+	+	+	+	+	+	+	+	$^+$	+
Harderian gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Urinary System																													
Kidney	+	+	$^+$	$^+$	+	+	+	+	$^+$	+	+	+	$^+$	+	+	$^+$	+	$^+$	+	$^+$	+	+	+	$^+$	+	+	+	$^+$	+
Nephroblastoma	Х		Х																										
Urinary bladder Polyp	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	М	+	+	+	+	+	+	+	+	+	+
Systemic Lesions																													
Multiple organs	.1	<u>ـــ</u>	_L_	_L_	_L	+	+	+	_L	<u>ـــ</u>	<i>_</i> ⊥	-L	_L	_L	<b>_</b> L	_L	4	_L	_L	_L	<i>_</i> ⊥_	-L	_L		<u>ــــ</u>	_L	_L	_L	+
Lymphome malignant	+	т	-	-	-	-	7	-	Ŧ	т	т	$\mathbf{v}$	-	-	-	-	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	-	Ŧ	T	T	-	7	T
Lymphoma mangnant												л																	

Individual Animal Tumor Pathology	of Fe	ma	le	Rat	s ii	n tl	he	2-1	Y ea	ar (	Ga	vag	ge S	Stu	dy	of	T	CD	D:	3	ng	/kg	5			
Number of Days on Study	6 4 8	6 8 1	7 1 8	7 2 9	7 3 1	7 3 2	7 3 2	7 3 2	7 3 3																	
Carcass ID Number	1 0 9	1 1 8	1 6 2	1 2 6	1 0 4	1 2 1	1 2 4	1 3 1	1 3 4	1 5 1	1 6 1	1 7 0	1 7 6	1 9 8	1 1 1	1 2 5	1 5 3	1 0 7	1 2 2	1 2 3	1 3 0	1 3 3	1 4 4	1 4 5	1 9 7	Total Tissues/ Tumors
Hematopoietic System																										
Bone marrow	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	54
Lymph node																										6
Lymph node, mandibular	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	54
Lymph node, mesenteric	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Spleen	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	54
Thymus Carcinoma, metastatic, lung	+	+	+	+	+	+	+	+	М	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	52 1
Integumentary System																										
Mammary gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	54
Carcinoma					Х	Х										Х					Х					7
Carcinoma, multiple			Х																							1
Fibroadenoma	Х	Х							Х	Х	Х				Х	Х		Х		Х			Х			26
Fibroadenoma, multiple						Х	Х						Х	Х			Х				Х	Х		Х		14
Skin	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	54
Lipoma							Х																			1
Musculoskeletal System Bone	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	54
Nervous System Brain	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	54
Respiratory System																										
Lung Alveolar/bronchiolar carcinoma Carcinoma, metastatic, mammary gland Nephroblastoma metastatic, kidney	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	54 1 1
Nose	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	54
Trachea	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	54
Special Senses System																										
Eye	+	+	+	+	+	+	$^+$	+	$^+$	+	+	+	$^+$	+	+	+	$^+$	+	+	$^+$	+	$^+$	$^+$	$^+$	+	54
Harderian gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	54
Urniary System																										
Kidney	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	54
Nephroblastoma																										2
Urinary bladder	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Polyp	Х																									1
Systemic Lesions																										
Multiple organs	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	54
Lymphoma malignant																										1

TABLE A2 Individual

															-													
	0	0	1	2	2	3	3	4	5	5	5	5	5	5	6	6	6	6	6	6	6	6	6	6	6	7	7	7
Number of Days on Study	4	5	7	3	4	4	4	2	5	7	7	7	7	7	0	1	1	2	3	4	5	5	5	7	9	1	1	2
	7	4	7	6	0	4	8	9	0	8	8	8	8	8	4	3	4	6	5	5	0	9	9	6	7	7	8	0
	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2
arcass ID Number	0	5	2	9	8	3	7	2	2	0	4	4	6	8	6	4	2	6	9	0	1	7	9	1	3	8	5	4
	5	8	0	0	6	5	4	4	5	1	4	6	4	2	3	2	3	0	6	4	1	0	7	7	1	9	9	1
Providence Oracle and																												
annentary System	+	+	-	+	-	-	1	+	-	-	+	+	+	+	+	-	+	+	+	-	+	+	+	+	+	-	-	+
sopilagus	+	+	+	+	+	+	+	+	+	+	+	+	+	+ +	+	+	+	+	+ +	+	+	+	+ +	+	+	+	+	+
ntestine large, colon	+ +	+ +	+ +	+ +	+	+ +	+ +	+	+	+	+ +	+	+ +	+ +	+ +	+	+	+ +										
nestine large, rectum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
nestine large, cecum	+ +	+ +	+ +	+ +	+	+ +	+ +	+	+	+	+ +	+	+ +	+ +	+ +	+	+	+ +										
testine small, duodenum	- T	т 1	т 1	т 1	т 1	- -	т 1	т 1	- T	т 1	т 1	T	- -	т 1	т 1	т 1	т 1	т 1	т 1	- -	т 1	т 1	т 1	т 1	т 1	- -	т 1	т 1
testine small, jejunum	+ +	+	+	+ +	- -	+	+ +	+ +	+ +	+	+	+ +																
iven	- T	т 1	т 1	т 1	т 1	- -	т 1	т 1	- T	т 1	т 1	T	т 1	- -	т 1	т 1	т 1	т 1	т 1	- -	т 1	т 1						
	+	+	+	+	+	+	+	+	+	+	+	+	Ŧ	Ŧ	Ŧ	+	+ +	+	+	+	+	+	+	+	+	+	+	+
Gingival, squamous cell carcinoma					+			+		+	+	+ X					т					+	+		+			
ancreas	+	+	+	+	+	A	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
alivary glands	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	Μ	+	+	+	+
tomach, forestomach	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
omach, glandular both	+	+	+	+	+	+	+	+ +	+ +	+	++	+ +	+	+	+	+	+ +	+	+	+	+	+	++	+	+	+	+ +	+
ardiovascular System																												
lood vessel	+	$^+$	+	+	+	$^+$	$^+$	$^+$	$^+$	+	$^+$	+	$^+$	+	$^+$	+	$^+$	+	$^+$	$^+$	$^+$	$^+$	+	$^+$	+	+	$^+$	$^+$
eart	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
ndocrine System																												
Adenoma	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Capsule, sarcoma, metastatic, skeletal muscle								Х																				
drenal medulla	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Pheochromocytoma benign																												
lets, pancreatic Adenoma	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
arathyroid gland	+	+	+	+	+	+	+	М	М	+	+	+	+	+	+	+	М	+	+	+	+	+	+	+	+	+	+	+
ituitary gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	М	+	+	+	+	+
Pars distalis, adenoma									Х			Х							Х		Х	Х		Х	Х	Х	Х	
hyroid gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Bilateral, C-cell, adenoma C-cell, adenoma																		х						Х			Х	Х
General Body System																												
one																												
Senital System																												
ntorar giana	+	+	+	+	+	+	+	+	+	+	+	+	+	т _	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Valy	+	+	+	+	+	+	+	+	+	+	+	+	Ŧ	Ŧ	Ŧ	+	Ŧ	+	+	+	+	+	+	+	+	+	+	+
															,	,		,										
A demonstra	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Adenoma																						37						
POIND SITOMAL																						Å						

TABLE A2 Individual Animal Tu

Individual Animal Tumor Pathology o	f Fei	ma	le 1	Rat	ts i	n t	he	2-Y	Yea	ar (	Ga	vag	ge S	Stu	ldy	of	TC	CD	D:	1	0 n	g/ŀ	٢g			
Number of Days on Study	7 2 4	7 2 8	7 3 1	7 3 2	7 3 3	7 3 3	7 3 3	7 3 3	7 3 3																	
Carcass ID Number	2 8 7	2 1 8	2 0 3	2 1 3	2 1 6	2 2 1	2 3 4	2 4 5	2 4 9	2 6 6	2 6 8	2 7 2	2 2 2	2 3 3	2 4 7	2 5 6	2 5 7	2 6 1	2 6 2	2 7 3	2 0 2	2 1 2	2 1 5	2 8 4	2 8 8	Total Tissues/ Tumors
Alimentary System																										
Esophagus Intestine large, colon	++	++	+++	++	+++	+++	+++	++	+++	+++	+++	+++	+++	++++	+++	+++	+++	++	+++	+++	+++	+++	+++	+++	+++	53 53
Intestine large, could	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Intestine large, cecum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Intestine small, duodenum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Intestine small, ieiunum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Intestine small, ileum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Liver	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Oral mucosa Gingival, squamous cell carcinoma		+						+				+		+			+				+	+				16 1
Pancreas	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	52
Salivary glands	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	52
Stomach, forestomach	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Stomach, glandular Tooth	+	+	+	+	+	+	+ +	+ +	+	+	+	+	+	+	+ +	+	+	+	+	+	+ +	+ +	+	+	+	53 12
Cardiovascular System																										
Blood vessel	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Heart	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Endocrine System																										
Adrenal cortex	+	$^+$	+	+	+	$^+$	$^+$	+	$^+$	+	+	+	$^+$	+	+	+	+	$^+$	$^+$	$^+$	+	+	+	$^+$	+	53
Adenoma						Х										Х										2
Capsule, sarcoma, metastatic, skeletal muscle																										1
Adrenal medulla	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Pheochromocytoma benign				Х								Х														2
Islets, pancreatic	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Adenoma																					Х	Х				2
Parathyroid gland	+	+	+	+	+	+	Μ	+	+	+	+	Μ	+	+	+	+	М	+	+	+	+	+	+	+	+	47
Pituitary gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	52
Pars distalis, adenoma	Х	Х	Х	Х	Х	Х	Х						Х	Х	Х	Х	Х	Х				Х	Х	Х	Х	26
Thyroid gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
C-cell, adenoma					Х	л	х	х		Х	Х	л	х		х		Х			х				х	Х	15
General Body System																										
None																										
Genital System																										
Clitoral gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Ovary	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Oviduct																		+					+			2
Uterus	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Adenoma	_							Х																		1
Polyp stromal	Х							Х						Х												4
Squamous cell papilloma										Х																1
vagina	+																									1

Individual Animal Tumor Pathology	y of Fe	ma	le l	Rat	ts ii	n tl	ne 2	2-Y	ea	r (	Gav	vag	e S	tuc	ły c	of T	CD	D:	1(	) n	g/k	g					
Number of Days on Study	0 4 7	0 5 4	1 7 7	2 3 6	2 4 0	3 4 4	3 4 8	4 2 9	5 5 0	5 7 8	5 7 8	5 7 8	5 7 8	5 7 8	6 0 4	56 11 34	6 2 6	6 3 5	6 4 5	6 5 0	6 5 9	6 5 9	6 7 6	6 9 7	7 1 7	7 1 8	7 2 0
Carcass ID Number	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2 2	2 2	2	2	2	2	2	2	2	2	2	2	2
	5	8	0	0	6	5	4	4	5	1	4	6	4	2	3	2 3	0	6	4	1	0	7	7	1	9	9	1
Hematopoietic System																											
Bone marrow	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+ -	+ +	+	+	+	+	+	+	+	+	+	+	+
Lymph node										+							+										
Lymph node, mandibular	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+ -	+ +	+	+	+	+	+	+	М	+	+	+	+
Lymph node, mesenteric	+	+	+	+	+	+	+	+	+	$^+$	+	+	+	+	+ -	+ +	+	+	+	$^+$	+	+	+	+	+	+	+
Spleen	+	+	+	+	+	+	+	+	+	$^+$	+	+	+	+	+ -	+ +	+	+	$^+$	$^+$	+	+	$^+$	+	+	$^+$	+
Thymus	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+ -	+ +	+	+	$^+$	$^+$	+	$^+$	+	+	М	+	+
Histiocytic sarcoma, metastatic, lung																											
ntegumentary System																											
Mammary gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+ -	+ +	+	+	+	+	+	+	+	+	+	+	+
Adenoma																											
Carcinoma																						Х				Х	
Fibroadenoma				Х		Х			Х		Х			Х	XZ	Χ				Х	Х				Х		Х
Fibroadenoma, multiple										Х			Х					Х					Х	Х			
Skin	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+ -	+ +	+	+	+	+	+	+	+	+	+	+	+
Basal cell carcinoma												_							_			Х					
Fibroma												Х				_			Х								
Myxosarcoma															2	K											
Musculoskeletal System																											
Bone	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+ -	+ +	+	+	+	+	+	+	+	+	+	+	+
Skeletal muscle								+																			
Sarcoma								Х																			
Nervous System																											
Brain	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+ -	+ +	+	+	+	+	+	+	+	+	+	+	+
Glioma malignant																Х	-										
Granular cell tumor malignant																											
Peripheral nerve																+											
spinai cord																+											
Respiratory System																											
Lung	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+ -	+ +	+	+	+	+	+	+	+	+	+	+	+
Sarcoma, metastatic, skeletal muscle								Х																			
Mediastinum, sarcoma																											
	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+ -	+ +	+	+	+	+	+	+	+	+	+	+	+
Nose			+	+	+	+	+	+	+	+	+	+	+	+	+ -	+ +	+	+	+	+	+	+	+	+	+	+	+
Nose Frachea	+	+																									
Nose Frachea Special Senses System	+	+																									
Nose Frachea Special Senses System Bar	+	+																						+			
Nose Trachea <b>Special Senses System</b> Ear Eye	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+ -	+ +	+	+	+	+	+	+	+	+++	+	+	+

Individual Animal Tumor Patholog	gy of Fe	ma	le l	Rat	ts ii	n t	he	2-3	Yea	ar (	Ga	vaş	ge S	Stu	dy	of	T	CD	D:	1	0 n	g/l	ĸg			
Number of Days on Study	7 2 4	7 2 8	7 3 1	7 3 1	7 3 1	7 3 1	7 3 1	7 3 1	7 3 1	7 3 1	7 3 1	7 3 1	7 3 2	7 3 3	7 3 3	7 3 3	7 3 3	7 3 3								
Carcass ID Number	2 8 7	2 1 8	2 0 3	2 1 3	2 1 6	2 2 1	2 3 4	2 4 5	2 4 9	2 6 6	2 6 8	2 7 2	2 2 2	2 3 3	2 4 7	2 5 6	2 5 7	2 6 1	2 6 2	2 7 3	2 0 2	2 1 2	2 1 5	2 8 4	2 8 8	Total Tissues/ Tumors
Hematopoietic System																										
Bone marrow	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Lymph node		+																								3
Lymph node, mandibular	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	52
Lymph node, mesenteric	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Thumus		+	+	+	+ +	+	+	+ +	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Histiocytic sarcoma, metastatic, lung	Ŧ	т	т	т	т	т	т Х	T	Ŧ	т	т	т	т	т	Ŧ	Ŧ	т	т	т	Ŧ	т	т	т	т	Ŧ	1
Integumentary System																										
Mammary gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Adenoma																						Х				1
Carcinoma		X		v											Х	v	v							v		4
Fibroadenoma multiple		л	v	л	v	v	v	$\mathbf{v}$	v					v	v	л	л			v	v		v	л	v	17
Skin	+	+	л +	+	л +	л +	л +	л +	л +	+	+	+	+	л +	л +	+	+	+	+	л +	л +	+	л +	+	л +	53
Basal cell carcinoma					'			'					'													1
Fibroma																										2
Myxosarcoma																										1
Musculoskeletal System																										
Bone	+	+	+	+	+	+	+	+	$^+$	$^+$	+	+	+	+	+	$^+$	$^+$	$^+$	+	$^+$	+	$^+$	+	$^+$	+	53
Skeletal muscle Sarcoma																										1 1
Nervous System																										
Brain	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Glioma malignant																										1
Granular cell tumor malignant												Х														1
Spinal cord																										1
Respiratory System																										
Lung	+	$^+$	+	$^+$	$^+$	+	$^+$	+	$^+$	$^+$	$^+$	+	$^+$	+	$^+$	$^+$	$^+$	$^+$	+	$^+$	+	+	+	$^+$	$^+$	53
Sarcoma, metastatic, skeletal muscle																										1
Mediastinum, sarcoma							Х																			1
Nose	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Trachea	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Special Senses System											+															2
Eve	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Harderian gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
											-							-			-					

TABLE A2 Individual

Individual Animal Tumor Path	ology of F	em	ale	R	ats	s in	ı tł	1e î	2-}	Yea	ar (	Ga	vag	ge S	Stu	ıdy	of	Т	CD	D:	1(	) n	g/k	g						
Number of Days on Study		0 0 4 : 7 ·	0 5 4	1 7 7	2 3 6	2 4 0	3 4 4	3 4 8	4 2 9	5 5 0	5 7 8	5 7 8	5 7 8	5 7 8	5 7 8	6 0 4	6 1 3	6 1 4	6 2 6	6 3 5	6 4 5	6 5 0	6 5 9	6 5 9	6 7 6	6 9 7	7 1 7	7 1 8	7 2 0	
Carcass ID Number		2 2 0 2 5 2	2 5 8	2 2 0	2 9 0	2 8 6	2 3 5	2 7 4	2 2 4	2 2 5	2 0 1	2 4 4	2 4 6	2 6 4	2 8 2	2 6 3	2 4 2	2 2 3	2 6 0	2 9 6	2 0 4	2 1 1	2 7 0	2 9 7	2 1 7	2 3 1	2 8 9	2 5 9	2 4 1	
Urinary System Kidney Nephroblastoma Urinary bladder		+ -	+	+	+	+ X +	+	+	++	++	++	+	+	++	++	++	+	++	++	++	++	++	++	++	++	+	++	+	++	
Systemic Lesions Multiple organs Leukemia granulocytic Lymphoma malignant		+ -	+	+	+	+	+	+	+	+	+	+	+	+	+	+ X	+	+	+ X	+	+	+	+	+	+	+	+	+	+	

# TABLE A2 Individual Animal Tr

Individual Animal Tumor Path	ology of Fe	ma	le ]	Rat	ts i	n ti	he	2-1	Yea	ar (	Ga	vag	ge S	Stu	ıdy	of	T	CD	D:	1(	) n	g/ŀ	٢g			
Number of Days on Study	7 2 4	7 2 8	7 3 1	7 3 2	7 3 3	7 3 3	7 3 3	7 3 3	7 3 3																	
Carcass ID Number	2 8 7	2 1 8	2 0 3	2 1 3	2 1 6	2 2 1	2 3 4	2 4 5	2 4 9	2 6 6	2 6 8	2 7 2	2 2 2	2 3 3	2 4 7	2 5 6	2 5 7	2 6 1	2 6 2	2 7 3	2 0 2	2 1 2	2 1 5	2 8 4	2 8 8	Total Tissues/ Tumors
Urinary System Kidney Nephroblastoma Urinary bladder	+	+	+	++	++	++	++	++	++	++	++	+	++	++	++	++	++	+	+	+	++	++	++	++	++	53 1 53
Systemic Lesions Multiple organs Leukemia granulocytic Lymphoma malignant	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53 1 1

TABLE A2 Individual Animal Tu

Number of Days on Study       0       1       3       3       3       4       4       4       4       5 <th>Individual Animal Tumor Pathology</th> <th>of Fer</th> <th>nal</th> <th>le I</th> <th>Rat</th> <th>s i</th> <th>n t</th> <th>he</th> <th>2-\</th> <th>l ea</th> <th>ar (</th> <th>Ga</th> <th>vaş</th> <th>ge S</th> <th>Stu</th> <th>ıdy</th> <th>of</th> <th>T (</th> <th>CD</th> <th>D:</th> <th>22</th> <th>2 n</th> <th>g/ŀ</th> <th>g</th> <th></th> <th></th> <th></th> <th></th> <th></th>	Individual Animal Tumor Pathology	of Fer	nal	le I	Rat	s i	n t	he	2-\	l ea	ar (	Ga	vaş	ge S	Stu	ıdy	of	T (	CD	D:	22	2 n	g/ŀ	g					
$ \begin{array}{c} \text{Carcass ID Number} & \begin{array}{ccccccccccccccccccccccccccccccccccc$	Number of Days on Study	0 5 3	1 9 7	3 3 8	3 7 3	3 8 0	3 8 9	4 0 6	4 1 6	4 1 7	4 5 3	4 6 2	4 8 5	5 2 2	5 3 3	5 4 5	5 5 7	5 6 1	5 7 8	5 7 8	5 7 8	5 9 2	6 0 1	6 0 7	6 1 5	6 2 6	6 2 6	6 2 6	6 4 1
Careass ID Number $3 \ 5 \ 3 \ 5 \ 5 \ 5 \ 5 \ 5 \ 5 \ 5 \ $		2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2		2	2	2
Allmentary System           Esophagus         + + + + + + + + + + + + + + + + + + +	Carcass ID Number	3 9	5 4 9	3 0 7	3 1 7	5 1 1	3 0	5 1 8	3 7	5 1 4	3 0 4	5 6 1	5 0 6	5 8 4	3 9 7	5 0 9	3 5	5 8 3	5 0	5 6 3	5 7 6	5 4 0	5 8 0	3 2 9	5 0 3	5 1 9	3 2 0	3 1	5 9 8
Suppland       +<	Alimentary System																												
nutstine large, colon       +	Esophagus	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	Μ	+	+	+	+	+	+	+	+	+	+	+
Intestine large, rortum       + <td>ntestine large, colon</td> <td>+</td>	ntestine large, colon	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
intesting lange, occum       + <td>ntestine large, rectum</td> <td>+</td> <td>М</td> <td>+</td>	ntestine large, rectum	+	+	+	+	+	+	+	+	+	+	+	+	М	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
the standing duodenum       +	ntestine large, cecum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
the standill regurum       +	ntestine small, duodenum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	М	+	+	+	+	+	+	+	+	+	+	+
Interm       + <td>ntestine small, jejunum</td> <td>+</td>	ntestine small, jejunum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Liver       + <td>ntestine small, ileum</td> <td>+</td> <td>M</td> <td>+</td>	ntestine small, ileum	+	+	+	+	+	+	+	+	+	+	+	+	M	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Carding location in Messandi       + <td< td=""><td>Cholongiocarcinoma</td><td>+</td><td>+</td><td>+</td><td>+</td><td>+</td><td>+</td><td>+</td><td>+</td><td>+</td><td>+</td><td>+</td><td>+</td><td>+</td><td>+</td><td>+</td><td>+</td><td>+</td><td>+</td><td>+</td><td>+</td><td>+</td><td>+</td><td>+</td><td>+</td><td>+</td><td>+</td><td>+</td><td>+</td></td<>	Cholongiocarcinoma	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Attraction       +	Aecentery																					л							
Tarrener behavious and the set of the set o	)ral mucosa							+							+	+					+	+	+		+		+	+	+
alrary glands       + + + + + + + + + + + + + + + + + + +	ancreas	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
$ \begin{array}{c}  + + + + + + + + + + + + + + + + + + +$	alivary glands	+	+	+	M	+	M	+	+	+	+	+	+	+	+	+	+	M	+	+	+	+	+	+	+	+	+	+	+
tomach, glandular       +	tomach, forestomach	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
interview       +	tomach, glandular	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Cardiovascular System         lood vessel       +	ooth	+		+			+									+					+	+							+
Carcinovascular bystem         Bood vessel         + + + + + + + + + + + + + + + + + + +	Pardiovasqular System																												
10000 (vssel)       + + + + + + + + + + + + + + + + + + +	Lai ulovascular System																												
Adenoma       + </td <td>Blood Vessel</td> <td>+</td>	Blood Vessel	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Cholocrine System       +		т	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	т	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	т	Ŧ	Ŧ	IVI	т	Ŧ	Ŧ	Ŧ	т	т	Ŧ	Ŧ	Ŧ	Ŧ	т
Adenoma       + + + + + + + + + + + + + + + + + + +	Endocrine System																												
Adenoma       X       X         Carcinoma       +	Adrenal cortex	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Caracinoma Adrenal medulla $+ + + + + + + + + + + + + + + + + + +$	Adenoma																									Х			
Adrenal medulla       +	Carcinoma																												
Phecohromocytoma benign         slets, pancreatic       + + + + + + + + + + + + + + + + + + +	Adrenal medulla	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
slets, parcreatic       + + + + + + + + + + + + + + + + + + +	Pheochromocytoma benign																												
Arathyroid gland AdenomaM++M++<	slets, pancreatic	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Action matrix gland $+ + + + + + + + + + + + + + + + + + +$	arathyroid gland	Μ	+	+	Μ	+	Μ	+	+	+	Μ	+	+	+	+	Μ	+	Μ	+	+	+	+	+	+	+	Μ	Μ	+	+
ManagementXXX	Adenoma Situitary gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Pars distalis, adenoma       X <td>Meningioma malignant metastatic brain</td> <td>Г</td> <td>'</td> <td></td> <td></td> <td>x</td> <td>1</td> <td></td> <td>'</td> <td>'</td> <td>1.</td> <td>1.</td> <td>1.</td> <td>1</td> <td></td> <td></td> <td>1</td> <td>1</td> <td></td> <td>1</td> <td>'</td> <td></td> <td>'</td> <td></td> <td>'</td> <td>1</td> <td>'</td> <td>'</td> <td>1</td>	Meningioma malignant metastatic brain	Г	'			x	1		'	'	1.	1.	1.	1			1	1		1	'		'		'	1	'	'	1
The formation of the f	Pars distalis, adenoma					Λ										х						х				х		х	х
Bilateral, C-cell, adenomaXXXXXGeneral Body System NoneClitoral gland $+ + + + + + + + + + + + + + + + + + + $	Thyroid gland	+	+	+	м	+	+	+	+	+	+	+	+	+	+	+	+	м	+	+	+	+	+	+	+	+	+	+	+
C-cell, adenoma X X X X X X General Body System None $C-cell, adenoma X X X X X X X X X X X X X X X X X X X$	Bilateral, C-cell, adenoma				.,,													.,,								<i>.</i>			,
General Body System         None         Genital System         Clitoral gland       + + + + + + + + + + + + + + + + + + +	C-cell, adenoma			Х					Х												Х		Х			Х			
Genital System         Clitoral gland       + + + + + + + + + + + + + + + + + + +	General Body System																												
Clitral gland       + + + + + + + + + + + + + + + + + + +	Conital System																												
x + y + y + y + y + y + y + y + y + y +	Vitoral gland					.1		5						М						J			.1	J	5				
$\begin{array}{cccccccccccccccccccccccccccccccccccc$		+	+	+	+	+	+	+	+	+	+	+	+	IVI	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Carcinoma, multiple Polyp stromal, multiple Vagina Schwannoma malignant metastatic skin X	Iterus	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Polyp stromal, multiple X X X X X Polyp stromal, multiple X Agina + Schwannoma malignant metastatic skin X	Carcinoma multinle	Ŧ	-	-	-1"	Ŧ	-1-	-T*	7*	7	-	-	-	-1-	7*	-1-	-1-	-1-	7*	Τ'	7	-	-	Τ'	7*	Τ.	7	-	Τ'
Polyp stromal, multiple A A A A A A A A Polyp stromal, multiple X A A A A A A A A A A A A A A A A A A	Polyn stromal																x						v	x					x
Agina + Schwannoma malignant metastatic skin Y	Polyn stromal multiple																Λ						Λ	Δ		x			1
Schwannoma malionant metastatic skin Y	/agina										+															1			
A A A A A A A A A A A A A A A A A A A	Schwannoma malignant metastatic skin										x																		

Number of Days on Study       6       6       6       7 </th <th>Individual Animal Tumor Pathology</th> <th>of Fe</th> <th>na</th> <th>le l</th> <th>Rat</th> <th>ts i</th> <th>n t</th> <th>he</th> <th>2-}</th> <th>Yea</th> <th>ar</th> <th>Ga</th> <th>va</th> <th>ge S</th> <th>Stu</th> <th>ıdy</th> <th>of</th> <th>T</th> <th>CD</th> <th>D:</th> <th>2</th> <th>2 n</th> <th>ıg/l</th> <th>٢g</th> <th></th> <th></th> <th></th>	Individual Animal Tumor Pathology	of Fe	na	le l	Rat	ts i	n t	he	2-}	Yea	ar	Ga	va	ge S	Stu	ıdy	of	T	CD	D:	2	2 n	ıg/l	٢g			
$ \begin{array}{c} \text{Carcass ID Number} & \begin{array}{ccccccccccccccccccccccccccccccccccc$	Number of Days on Study	6 4 5	6 7 5	6 8 0	6 9 0	7 0 8	7 0 8	7 3 1	7 3 1	7 3 1	7 3 1	7 3 1	7 3 1	7 3 1	7 3 1	7 3 1	7 3 1	7 3 2	7 3 2	7 3 3							
Alianetary System         Esophagus       + + + + + + + + + + + + + + + + + + +	Carcass ID Number	3 6 5	3 1 6	3 4 3	3 4 5	3 4 8	3 6 2	3 0 1	3 0 2	3 0 5	3 2 6	3 3 4	3 4 1	3 4 2	3 8 1	3 9 4	3 9 6	3 1 2	3 3 2	3 3 3	3 3 6	3 4 6	3 8 2	3 9 2	3 9 5	3 6 4	Total Tissues/ Tumors
Esphagus       + + + + + + + + + + + + + + + + + + +	Alimentary System																										
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Esophagus	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	52
Intestine large, cetum       + <td>Intestine large, colon</td> <td>+</td> <td>53</td>	Intestine large, colon	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
$\begin{array}{c} \text{Incsine and, contains and, loudenum } & + & + & + & + & + & + & + & + & + &$	Intestine large, rectum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	52
$\begin{array}{c} 1 & 1 & 1 & 1 & 1 & 1 & 1 & 1 & 1 & 1 $	Intestine mall duodenum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	52
Intestine small, leum       + + + + + + + + + + + + + + + + + + +	Intestine small, ieiunum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Liver the second secon	Intestine small, jejunum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	52
Chalagiocarcinoma       1         Mesentery       +       +       +       1         Oral mucosa       +       +       +       +       +       1         Pancreas       +	Liver	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Mesentry       +       +       1         Oral mucosa       +	Cholangiocarcinoma																										1
Oral mucosa       + <td< td=""><td>Mesentery</td><td></td><td></td><td></td><td>+</td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td>1</td></td<>	Mesentery				+																						1
Pancreas $+ + + + + + + + + + + + + + + + + + +$	Oral mucosa		+	+		+									+					$^+$							16
Salivary glands $+ + + + + + + + + + + + + + + + + + +$	Pancreas	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Stomach, torestomach       + + + + + + + + + + + + + + + + + + +	Salivary glands	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	50
Stomach, glandular       + + + + + + + + + + + + + + + + + + +	Stomach, forestomach	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Cardiovascular System         Blood vessel       + + + + + + + + + + + + + + + + + + +	Stomach, glandular Tooth	+	+	+	+	++	+	+	+	+	+	+	+	++	+	+	+	+	+	+	+	+	+	+	+	++	53
Blood vessel       + + + + + + + + + + + + + + + + + + +	Cardiovascular System																										
Heart       + + + + + + + + + + + + + + + + + + +	Blood vessel	+	+	$^+$	+	+	$^+$	$^+$	+	+	+	+	+	+	+	$^+$	$^+$	+	$^+$	$^+$	$^+$	+	+	+	$^+$	$^+$	53
Endocrine System         Adrenal cortex       +	Heart	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	52
Adrenal cortex       + + + + + + + + + + + + + + + + + + +	Endocrine System																										
Carcinoma       X       X       X       X         Adrenal medulla       + + + + + + + + + + + + + + + + + + +	Adrenal cortex Adenoma	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	$^+$ X	+	+	53 2
Addream medula       + + + + + + + + + + + + + + + + + + +	Carcinoma																		X								1
Information of type in a being in       X	Adrenial medulia Phaeshromoautoma honign	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	$\mathbf{v}$	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	$\mathbf{v}$	$\mathbf{v}$	Ŧ	23
Data by parathyroid gland       + + + + + + + + + + + + + + + + + + +	Islets nancreatic	+	+	+	+	+	+	+	+	+	+	+	л +	+	+	+	+	+	+	+	+	+	+	+	л +	+	53
Adenoma       X       Image: State of the state	Parathyroid gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	M	44
Pituitary gland       + + + + + + + + + + + + + + + + + + +	Adenoma		Х																								1
Meningioma malignant, metastatic, brain Pars distalis, adenomaXXXXXXXXXX15Thyroid gland Bilateral, C-cell, adenoma $+ + + + + + + + + + + + + + + + + + + $	Pituitary gland	+	+	+	+	+	$^+$	+	$^+$	+	+	+	+	+	+	$^+$	$^+$	+	$^+$	$^+$	$^+$	+	+	+	$^+$	+	53
Pars distalis, adenoma       X <td>Meningioma malignant, metastatic, brain</td> <td></td> <td>1</td>	Meningioma malignant, metastatic, brain																										1
Thyroid gland $+ + + + + + + + + + + + + + + + + + + $	Pars distalis, adenoma						Х				Х		Х	Х	Х			Х			Х	Х		Х	Х		15
Bilateral, C-cell, adenoma       X	Thyroid gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	51
General Body System         None         Genital System         Clitoral gland       + + + + + + + + + + + + + + + + + + +	C-cell, adenoma			Х	Х		Х			Х	Х		Х		Х	Х	Х				Х				Х		2 14
Genital System         Clitoral gland       + + + + + + + + + + + + + + + + + + +	General Body System None																										
Clitoral gland       + + + + + + + + + + + + + + + + + + +	Genital System																										
Ovary       + + + + + + + + + + + + + + + + + + +	Clitoral gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	52
Uterus       + + + + + + + + + + + + + + + + + + +	Ovary	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Carcinoma, multipleX1Polyp stromalXXX8	Uterus	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Polyp stromal X X X X 8	Carcinoma, multiple									Х																	1
	Polyp stromal										Х		Х						Х			Х					8
Polyp stromal, multiple X X 3	Polyp stromal, multiple														Х		Х										3
vagina + 3 Schwannoma malignant, metastatic, skin 1	vagina Schwannoma malignant, metastatic, skin				+																					+	3

Individual Animal Tumor Patholo	gy of F	em	al	e F	Rat	s i	n tl	he	2-1	Yea	ar (	Ga	vaş	ge S	Stu	dy	of	T	CD	D:	22	2 n	g/k	g						
Number of Days on Study		0 5 3	1 9 7	3 3 8	3 7 3	3 8 0	3 8 9	4 0 6	4 1 6	4 1 7	4 5 3	4 6 2	4 8 5	5 2 2	5 3 3	5 4 5	5 5 7	5 6 1	5 7 8	5 7 8	5 7 8	5 9 2	6 0 1	6 0 7	6 1 5	6 2 6	6 2 6	6 2 6	6 4 1	
Carcass ID Number		3 3 9	3 4 9	3 0 7	3 1 7	3 1 1	3 3 0	3 1 8	3 3 7	3 1 4	3 0 4	3 6 1	3 0 6	3 8 4	3 9 7	3 0 9	3 3 5	3 8 3	3 5 0	3 6 3	3 7 6	3 4 0	3 8 0	3 2 9	3 0 3	3 1 9	3 2 0	3 3 1	3 9 8	
Hematopoietic System Bone marrow Lymph node Lymph node, mandibular Lymph node, mesenteric Spleen Thymus		+++++++++++++++++++++++++++++++++++++++	+++++++++++++++++++++++++++++++++++++++	+++++++++++++++++++++++++++++++++++++++	+ M + +	+ + + M	+ M + +	++++++	+++++++	+++++++	+++++++	+++++++	+++++++	+ + + + +	+++++++	++++++	++++++	+ + + M	+++++++	+ + + + + +	+++++++	++++++	+ + + M	++++++++	+++++++	+++++++	+ + + + + +	+++++++	++++++++	
Integumentary System Mammary gland Carcinoma Fibroadenoma Fibroadenoma, multiple Skin Fibrosarcoma Keratoacanthoma Schwannoma malignant		+	+	+ X +	+	+ + X	++	++	++	+	+ X + X	++	+ X +	++	+ X +	++	+ X +	++	++	+ X +	+ X +	+ X +	++	+ X +	+ X +	+ X +	+ X +	+ X +	+ X +	
Musculoskeletal System Bone Cranium, schwannoma malignant, metastatic, skin Periosteum, fibrosarcoma Skeletal muscle Fibrous histiocytoma		+	+	+	+	+ X	+	+	+	+	+	+ + X	+	+ X	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	
Nervous System Brain		+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	
Respiratory System Lung Nose Trachea		++++++	+ + +	++++++	+++++	+++++	+++++	+++++	+++++	+++++	++++	+++++	+++++	+++++	+++++	+ + +	+++++	M + M	++++++	+++++	+++++	+++++	+++++	++++++	++++++	+++++	++++++	++++++	++++++	
<b>Special Senses System</b> Eye Harderian gland		+ +	+ +	+ +	+++	+ +	+ +	+ +	+++	++	+ +	++	+++	+++	+++	+ +	++	M M	+++	+++	++	+ +	+ +	+++	+++	+ +	+ +	+ +	+++	
Urinary System Kidney Stromal nephroma Urinary bladder		+	+	+	++	++	++	++	++	++	++	++	++	+ M	++	++	++	+	++	++	++	++	++	+	+	+	+	+	+	
Systemic Lesions Multiple organs		+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	

Individual Animal Tumor Patholog	gy of Fe	ma	le	Rat	ts i	n tl	he	2-Y	Yea	ar (	Ga	vag	ge S	Stu	ldy	of	TC	CD	D:	22	2 n	g/ŀ	g			
Number of Days on Study	6 4 5	6 7 5	6 8 0	6 9 0	7 0 8	7 0 8	7 3 1	7 3 2	7 3 2	7 3 2	7 3 2	7 3 2	7 3 2	7 3 2	7 3 2	7 3 3										
Carcass ID Number	3 6 5	3 1 6	3 4 3	3 4 5	3 4 8	3 6 2	3 0 1	3 0 2	3 0 5	3 2 6	3 3 4	3 4 1	3 4 2	3 8 1	3 9 4	3 9 6	3 1 2	3 3 2	3 3 3	3 3 6	3 4 6	3 8 2	3 9 2	3 9 5	3 6 4	Total Tissues/ Tumors
Hematopoietic System Bone marrow Lymph node Lymph node, mandibular Lymph node, mesenteric Spleen Thymus	+ + + + + +	· + · + · +	++++++	+ + + + + +	+ + + + + +	+++++++	+++++++	++++++	+ + + + +	+++++++	++++++	++++++	+ + + + +	++++++	++++++	+++++++	+++++++	++++++	+++++++	+ + + M	+++++++	+++++++	+++++++	+++++++	++++++++	53 5 50 53 53 49
Integumentary System Mammary gland Carcinoma Fibroadenoma Fibroadenoma, multiple Skin Fibrosarcoma Keratoacanthoma Schwannoma malignant	+ X +	+ X	+ X + X	+	+ X +	+ X +	+ X +	+ X +	+	+ X +	+	+ X +	+ X +	+	+ X +	+	+ X +	+ X +	+ + X	+ X +	+	+ X +	+ X +	++	+ X +	53 2 17 12 53 1 1 2
Musculoskeletal System Bone Cranium, schwannoma malignant, Metastatic, skin Periosteum, fibrosarcoma Skeletal muscle Fibrous histiocytoma	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53 1 1 2 1
Nervous System Brain	+	· +	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
<b>Respiratory System</b> Lung Nose Trachea	+ + +	- + - + - +	+++++	+++++	++++++	+++++	+++++	+++++	+++++	+++++	+++++	++++++	+++++	+++++	+++++	+++++	+++++	+++++	+++++	+++++	+++++	+++++	+++++	+++++	+ + +	52 53 52
<b>Special Senses System</b> Eye Harderian gland	+ +	· + · +	++	+ +	+++	+++	+++	+++	+++	+++	+++	++	+++	+++	+++	+++	+++	+++	+++	+++	+++	+++	+++	+++	+++	52 52
Urinary System Kidney Stromal nephroma Urinary bladder	+	· + · +	+	++	+ X +	++	+	+	++	++	+	+	+	+	+	++	+	+	++	++	++	++	++	++	++	53 1 52
Systemic Lesions Multiple organs	+	• +	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53

TABLE A2 Individual Animal TI

Individual Animal Tumor Patholog	gy of Fer	nal	le I	Rat	ts i	n tl	he	2-1	Yea	ar (	Ga	vag	ge S	Stu	dy	of	TC	CD	D:	4	6 n	g/k	ĸg					
Number of Days on Study	1 4 1	1 8 5	3 1 7	3 4 1	3 9 6	4 4 4	4 7 4	4 8 6	4 8 6	5 1 8	5 5 7	5 7 8	5 8 9	5 9 0	5 9 9	6 2 1	6 4 2	6 4 5	6 5 7	6 5 9	6 7 2	6 7 3	6 7 3	6 7 5	6 7 7	6 8 3	6 9 4	7 0 1
																										—		
Carcass ID Number	4 2 6	4 4 5	4 4 7	4 1 3	4 8 3	4 6 5	4 1 6	4 6 6	4 8 1	4 1 0	4 7 9	4 5 2	4 1 7	4 1 8	4 5 1	4 6 7	4 9 7	4 7 1	4 4 6	4 0 9	4 1 2	4 0 7	4 6 9	4 7 7	4 8 2	4 7 4	4 2 8	4 1 9
Alimentary System																												
Sophagus	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
ntestine large colon	+ +	+ +	+ +	+	+ +	+ +	+	+	+ +	+ +	+	+	+ +	+ +	+	+ +	+ +	+ +	+	+ +	+	+ +	+	+ +	+ +	+	+ +	+
ntestine large, colon	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
ntestine large, rectum	+ +	+ +	+ +	+	+	+ +	+	+	+ +	+ +	+	+	+ +	+ +	+	+ +	+ +	+ +	+	+ +	+	+	+	+ +	+ +	+	+ +	+
ntestine small duodenum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
ntestine small jejunum	- <del>-</del> -	г ,	т "⊥	т ,⊥	т 	т "L		г 	т 	+ +	+ +	т _	т "⊥	т "⊥	т "⊥	г "	۔ +	г -	г 	т "⊥	т ,⊥	т 	т "⊥	т "L	г "⊥	г -	т "L	- -
L ejomyoma	-	Г	т	т	т	Г	Г	Г	Τ,	т	T	т	т	т	Г	Г	ſ	F	Г	т	т	т	т	$\mathbf{v}$	F	Г	Г	Г
Intesting small iloum	+	+	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	+	+	+	+	+	л 	+	+	+	+
iver	+ +	г +	T +	+	+	+	+	+	+	+	+	+	+ +	T +	+	+	+	+	+	+ +	+	+	+	+	г +	- -	+	+
Carcinoma metastatic uterus	Г	1			'			'		1.			x	1					'			'					x	'
Cholangiocarcinoma Cholangiocarcinoma, multiple													Λ														Λ	
Hepatocellular adenoma Jesentery													+				+											
Carcinoma, metastatic, uterus													Х															
Dral mucosa			$^+$				$^+$		$^+$		+				$^+$		$^+$		$^+$	$^+$			$^+$		$^+$	$^+$		
Gingival, squamous cell carcinoma																				Х								
ancreas	+	$^+$	+	+	+	$^+$	$^+$	$^+$	+	+	+	+	$^+$	+	М	+	$^+$	+	$^+$	$^+$	+	+	$^+$	+	+	$^+$	+	$^+$
Carcinoma, metastatic, uterus													Х															
alivary glands	+	$^+$	+	+	+	$^+$	$^+$	$^+$	+	+	+	+	Μ	+	$^+$	М	$^+$	+	$^+$	$^+$	+	+	$^+$	+	+	$^+$	+	$^+$
tomach, forestomach	+	$^+$	+	+	+	$^+$	$^+$	$^+$	+	+	+	+	$^+$	+	$^+$	+	$^+$	+	$^+$	$^+$	+	+	$^+$	+	+	$^+$	+	$^+$
Squamous cell carcinoma																												
Squamous cell papilloma																												
tomach, glandular	+	$^+$	$^+$	+	$^+$	$^+$	$^+$	$^+$	$^+$	+	+	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	+	$^+$	$^+$	$^+$	$^+$	$^+$	+	$^+$
Tooth			+				+						+			+				+					+			
Cardiovascular System																												
Blood vessel	+	$^+$	+	+	$^+$	+	+	$^+$	+	+	+	+	$^+$	+	+	$^+$	+	$^+$	$^+$	$^+$	+	$^+$	$^+$	+	+	+	+	+
leart	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Endocrine System																												
drenal cortex	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
denoma																												
drenal medulla	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Pheochromocytoma benign																												
elets, pancreatic	+	+	+	+	+	+	+	+	+	+	+	+	+	+	М	+	+	+	+	+	+	+	+	+	+	+	+	+
arathyroid gland	М	+	+	+	+	+	+	+	М	+	+	+	Μ	+	Μ	+	+	$^+$	+	+	Μ	+	М	+	+	+	+	+
ituitary gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	$^+$	+	+	+	+	$^+$	+	+	+	+	+
Pars distalis, adenoma						Х						Х	Х	Х					Х		Х	Х					Х	
hyroid gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Bilateral, C-cell, adenoma																												
C-cell, adenoma									Х		Х			Х											Х	Х		
C-cell, carcinoma																												
Follicular cell, carcinoma													Х															
General Body System																												

None

	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	
Number of Dava on Study	/		2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	
Number of Days on Study	0	0	2	3	1	3	3	3	1	3	1	1	3	2 2	3 2	2 2	с С	2	2	2	2	2	2	2	2	
	1	0	0	1	1	1	1	1	1	1	1	1	1	Ζ	2	2	2	3	3	3	3	3	3	3	3	
	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	Total
Carcass ID Number	8	0	7	0	0	1	1	3	5	5	6	7	8	4	5	6	7	0	2	4	6	7	8	9	9	Tissues
	9	2	8	5	6	4	5	0	4	5	2	2	8	9	0	3	5	1	7	8	4	0	7	0	6	Tumors
Alimontory System																										
Esophagus	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Intestine large colon	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Intestine large, colon	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Intestine large, rectain	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Intestine small duodenum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Intestine small, jejunum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Leiomvoma																										1
Intestine small, ileum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Liver	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Carcinoma, metastatic, uterus																										2
Cholangiocarcinoma											Х				Х					Х						3
Cholangiocarcinoma, multiple				Х																						1
Hepatocellular adenoma			Х																							1
Mesentery						+			+														+	+		6
Carcinoma, metastatic, uterus																										1
Oral mucosa	+	+	+			+		+				+	+					$^+$			+		+		+	22
Gingival, squamous cell carcinoma		Х	Х																						Х	4
Pancreas	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	$^+$	+	+	+	+	+	+	+	52
Carcinoma, metastatic, uterus																										1
Salivary glands	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	$^+$	$^+$	$^+$	+	+	+	+	+	51
Stomach, forestomach	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	$^+$	+	+	+	+	+	+	+	53
Squamous cell carcinoma	Х																									1
Squamous cell papilloma		Х																								1
Stomach, glandular	+	+	+	+	+	$^+$	+	+	+	+	+	+	$^+$	+	+	$^+$	$^+$	+	$^+$	$^+$	+	+	+	+	+	53
Tooth	+	+				+		+					+			+	+				+					14
Cardiovascular System																										
Blood vessel	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Heart	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Endocrine System																										
Adrenal cortex	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Adenoma							X																			l
Adrenal medulla	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Pheochromocytoma benign		X				X											X									3
Islets, pancreatic	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	52
Parathyroid gland	+	+	+	+	+	M	+	+	+	+	+	+	+	+	+	+	M	+	+	+	+	+	+	+	+	45
Pituitary gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Pars distalis, adenoma	X		X		X			X		X		X		X	X	X	X							X	X	20
nyroid gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Bilateral, C-cell, adenoma																	Х								v	1
C-cell, adenoma				Х		Х	X			37	Х		Х	Х											Х	12
C-cell, carcinoma							Х			Х																2
Fonicular cell, carcinoma																										1
Follicular cell, carcinoma General Body System																										

None

Fable A2 Individual Animal Tumor Pathology	of Fei	na	le ]	Rat	is i	n tl	he	2-}	l ea	ar (	Ga	vag	ge S	Stu	dy	of	ТС	CD	D:	4	6 n	g/k	g					
Number of Days on Study	1 4 1	1 8 5	3 1 7	3 4 1	3 9 6	4 4 4	4 7 4	4 8 6	4 8 6	5 1 8	5 5 7	5 7 8	5 8 9	5 9 0	5 9 9	6 2 1	6 4 2	6 4 5	6 5 7	6 5 9	6 7 2	6 7 3	6 7 3	6 7 5	6 7 7	6 8 3	6 9 4	7 0 1
	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
Carcass ID Number	2 6	4 5	4 7	1 3	8 3	6 5	1 6	6 6	8 1	1 0	7 9	5 2	1 7	1 8	5 1	6 7	9 7	7 1	4 6	0 9	1 2	0 7	6 9	7 7	8 2	7 4	2 8	1 9
Genital System																												
Clitoral gland	+	+	+	+	+	Μ	+	+	+	+	+	+	+	+	+	+	+	+	М	+	+	+	+	+	+	+	+	+
Ovary	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Granulosa cell tumor malignant																									v			
necoma benign																+									Х			
Jterus	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Carcinoma																												
Hemangiosarcoma Polyp stromal														х		х									х			
Polyp stromal, multiple														-		-	Х								-			
Squamous cell carcinoma													Х		Х				Х								Х	
Squamous cell carcinoma, multiple																								Х				
yagina Schwannoma malignant											+ X									+								
Squamous cell carcinoma											Λ																	
Jematonoietic System																												
Sone marrow	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
ymph node										+		+	+	+		+												+
Lumbar, carcinoma, metastatic, uterus													Х															
ymph node, mandibular	+	+	+	+	+	+	+	+	+	+	+	+	Μ	+	+	М	+	+	+	+	+	+	+	+	+	+	+	+
ymph node, mesenteric	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Hemangiosarcoma	+	+	+	+	+	+	+	+	+	+	+	÷	+	+	IVI	+	+	+	+	+	+	+	+	+	+	+	+	+
Thymus	+	+	+	+	+	М	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	М	+	+	+	+	М
ntogumontour System																												
Mammary gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Carcinoma																							Х					
Fibroadenoma	Х			Х	Х	Х	Х	Х	Х		Х	Х		Х					Х			Х				Х	Х	Х
Fibroadenoma, multiple				.1	.1	J	J	L		.1		.1	J		J	J	X	X	L	5		.1	.1	X	X	J	.1	5
KIII	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Musculoskeletal System				,			,	,			,						,	,	,					,	,			
Solie	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Nervous System																												
Brain	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Respiratory System																												
ung	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Alveolar/bronchiolar adenoma, multiple							,	,									,	,	,					,	,			
nose Trachea	++	++	++	++	++	++	++	++	+ +	++	++	++	++	++	++	++	++	++	++	++	++	++	++	++	++	++	++	++
Special Sonses System																												
ve	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Harderian gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
0																					-				-			-

Individual Animal Tumor Pathology	of Fe	ma	le l	Rat	ts i	n t	he	2-Y	Yea	ar (	Ga	vaş	ge S	Stu	ldy	of	T	CD	D:	4	6 n	g/ŀ	٢g			
Number of Days on Study	7 0 1	7 0 8	7 2 0	7 3 1	7 3 2	7 3 2	7 3 2	7 3 2	7 3 3																	
Carcass ID Number	4 8 9	4 0 2	4 7 8	4 0 5	4 0 6	4 1 4	4 1 5	4 3 0	4 5 4	4 5 5	4 6 2	4 7 2	4 8 8	4 4 9	4 5 0	4 6 3	4 7 5	4 0 1	4 2 7	4 4 8	4 6 4	4 7 0	4 8 7	4 9 0	4 9 6	Total Tissues/ Tumors
Genital System																										
Clitoral gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	51
Ovary	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Granulosa cell tumor malignant Thecoma benign						X																				1
Oviduct																										1
Uterus	+	+	+	+	+	+	+	+	$^+$	+	+	+	$^+$	+	+	+	$^+$	+	+	+	+	+	+	+	+	53
Carcinoma						Х							Х													2
Hemangiosarcoma										Х																1
Polyp stromal Polyp stromal, multiple			Х				Х								Х		Х			Х	Х					9
Squamous cell carcinoma Squamous cell carcinoma, multiple Vagina																					+					4 1 3
Schwannoma malignant Squamous cell carcinoma																					x					1
Hematopoietic System																										
Bone marrow	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Lymph node																										6
Lumbar carcinoma metastatic uterus																										1
Lymph node mandibular	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	51
Lymph node, mesenteric	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Spleen	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	52
Hemangiosarcoma																			x							1
Thymus	+	М	Μ	+	+	+	+	+	+	М	+	+	+	+	+	+	+	+	+	+	+	+	+	+	М	46
Integumentary System																										52
Mammary gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Carcinoma			v	v	Х					v	37			v	v	v		v						v		2
Fibroadenoma		v	Х	Х	v		v			Х	Х		v	Х	Х	Х		Х	v		v	v	v	Х		24
Skin	+	X +	+	+	X +	+	х +	+	+	+	+	+	х +	+	+	+	+	+	Х +	+	Х +	X +	X +	+	+	12 53
Musculoskeletal System																										
Bone	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Nervous System Brain	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Respiratory System																										
Lung	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Alveolar/bronchiolar adenoma, multiple																				Х						1
Nose	+	+	+	+	+	+	+	+	$^+$	+	+	+	+	$^+$	+	+	+	$^+$	$^+$	+	$^+$	$^+$	$^+$	+	+	53
Trachea	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Special Senses System																										
Eye	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Harderian gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53

TABLE A2 Individual

Individual Animal Tumor Pathology of	Fei	na	le l	Rat	s i	n tl	he	2-1	Yea	ar (	Ga	vag	ge S	Stu	ıdy	of	Т	CD	D:	40	6 n	g/k	g						
	1	1	3	3	3	4	4	4	4	5	5	5	5	5	5	6	6	6	6	6	6	6	6	6	6	6	6	7	
Number of Days on Study	4	8	1	4	9	4	7	8	8	1	5	7	8	9	9	2	4	4	5	5	7	7	7	7	7	8	9	0	
	1	5	7	1	6	4	4	6	6	8	7	8	9	0	9	1	2	5	7	9	2	3	3	5	7	3	4	1	
	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	
Carcass ID Number	2	4	4	1	8	6	1	6	8	1	7	5	1	1	5	6	9	7	4	0	1	0	6	7	8	7	2	1	
	6	5	7	3	3	5	6	6	1	0	9	2	7	8	1	7	7	1	6	9	2	7	9	7	2	4	8	9	
Urinary System																													
Kidney	+	+	+	+	+	+	$^+$	+	+	+	+	+	+	+	+	$^+$	$^+$	$^+$	$^+$	$^+$	+	+	+	$^+$	$^+$	+	+	+	
Urinary bladder	+	+	+	+	+	+	$^+$	+	+	+	+	+	+	+	+	$^+$	$^+$	$^+$	+	$^+$	+	+	+	$^+$	$^+$	+	+	+	
Squamous cell carcinoma, metastatic, uterus															Х														
Transitional epithelium, carcinoma													Х																
Systemic Lesions																													
Multiple organs	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	
Lymphoma malignant										Х																			

Individual Animal Tumor Pathology of	of Fei	na	le l	Rat	ts i	n t	he	2-1	Yea	ar (	Ga	vag	ge S	Stu	ıdy	of	Т	CD	D:	40	6 n	g/k	g			
Number of Days on Study	7 0 1	7 0 8	7 2 0	7 3 1	7 3 2	7 3 2	7 3 2	7 3 2	7 3 3																	
Carcass ID Number	4 8 9	4 0 2	4 7 8	4 0 5	4 0 6	4 1 4	4 1 5	4 3 0	4 5 4	4 5 5	4 6 2	4 7 2	4 8 8	4 4 9	4 5 0	4 6 3	4 7 5	4 0 1	4 2 7	4 4 8	4 6 4	4 7 0	4 8 7	4 9 0	4 9 6	Total Tissues/ Tumors
Urinary System Kidney Urinary bladder Squamous cell carcinoma, metastatic, uterus Transitional epithelium, carcinoma	+ +	++	++	++	++	+++	++	++	++	+++	++	++	++	++	+++	++	++	++	++	++	++	++	++	++	+++	53 53 1 1
Systemic Lesions Multiple organs Lymphoma malignant	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53 1

TABLE A2 Individual Anim

Individual Annial Tumor Tatholog	sy of ref		-							-					J	-						8	8					
	3	4	5	5	5	5	5	6	6	6	6	6	6	6	6	6	6	6	6	6	6	6	6	6	6	6	7	7
Number of Days on Study	9	7	4	6	8	9	9	0	1	1	1	1	3	3	4	4	4	4	4	5	5	6	6	7	8	9	0	1
	6	4	6	1	1	8	9	4	0	0	2	4	8	8	1	1	2	7	8	2	5	7	8	3	0	3	1	6
Conservation ID Normalian	5	5	6	5	5	6	6	5	5	5	5	5	5	6	5	6	5	6	6	5	6	5	5	6	6	5	5	5
Carcass ID Number	2	2	3	9	1	0	1	8	1	1	2	8	1	0	0	3	9	0	4	0	I	8	1	0	4	8	2	1
	5	I	6	9	0	6	4	4	2	4	4	6	1	9	7	8	8	2	4	9	6	9	I	4	2	5	2	9
Alimentary System																												
Esophagus	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
ntestine large, colon	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	М	М	+	+
ntestine large, rectum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
ntestine large, cecum	+	+	+	+	$^+$	+	+	+	+	+	$^+$	+	+	+	+	+	+	+	$^+$	+	+	+	$^+$	$^+$	М	М	+	+
ntestine small, duodenum	+	+	+	+	$^+$	+	+	+	+	+	$^+$	+	+	+	+	+	+	+	$^+$	+	+	+	$^+$	$^+$	+	+	+	+
ntestine small, jejunum	+	+	+	+	$^+$	+	+	+	+	+	$^+$	+	+	+	+	+	+	+	$^+$	+	+	+	$^+$	$^+$	М	+	+	+
ntestine small, ileum	+	+	$^+$	$^+$	+	+	$^+$	+	$^+$	+	$^+$	+	+	+	+	+	+	+	+	+	$^+$	+	+	$^+$	М	М	+	$^+$
Liver	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	$^+$	+	+	+	+
Carcinoma, metastatic, pancreas																Х												
Cholangiocarcinoma										Х			Х										Х					
Cholangiocarcinoma, multiple												Х		Х		Х			Х		Х					Х	Х	
Hepatocellular adenoma																	Х		Х							Х		Х
Hepatocellular adenoma, multiple Hepatocholangioma														x														
lesentery					+				+					+							+			+				
Dral mucosa		+	+	+	+	+	+	+	+	+			+	+	+	+				+	+	+		+	+			+
Gingival squamous cell carcinoma			x	x				x		x			x								x				x			x
ancreas	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	А	+	+	+	+
Acinus adenoma																								11				
Acinus, carcinoma																x												
alivary glands	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	м
Stomach forestomach	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Squamous cell papilloma																												
Stomach glandular	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Sooth							+			+	+			+	+	+		+		+		+						
Cardiovascular System																												
Blood vessel	+	$^+$	$^+$	$^+$	+	+	$^+$	+	$^+$	$^+$	$^+$	+	+	+	+	+	+	+	+	$^+$	$^+$	+	+	$^+$	+	+	$^+$	$^+$
leart	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Endocrine System																												
Adrenal cortex	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Adenoma																			Х									
Pheochromocytoma benign																	Х											
Adrenal medulla	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Pheochromocytoma malignant																	Х											
Pheochromocytoma benign																	Х							Х				
slets, pancreatic	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
arathyroid gland	+	+	М	+	+	+	+	+	+	+	+	+	+	М	Μ	+	+	+	+	+	+	+	+	+	+	+	+	М
Pituitary gland	М	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Adenoma																												
Pars distalis, adenoma								Х								Х		Х		Х							Х	_
Thyroid gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	М
Bilateral, C-cell, adenoma													Х		Х													
																			37						37			

#### **General Body System**

None

TABLE A2           Individual Animal Tumor Patholo	ogy of Fe	ma	le ]	Rat	ts i	n ti	he	2-Y	Yea	ar (	Ga	vag	ge S	Stu	ıdy	of	T	CD	D:	1	00	ng	/kg			
Number of Days on Study	7 1 7	7 2 1	7 2 2	7 2 3	7 3 1	7 3 2	7 3 2	7 3 2	7 3 2	7 3 2	7 3 3	7 3 3	7 3 3	7 3 3	7 3 3	7 3 3										
Carcass ID Number	5 9 7	5 1 8	6 3 3	6 1 2	5 1 3	5 1 6	5 8 1	5 8 3	6 0 8	6 1 3	6 1 5	6 1 7	6 3 1	6 3 2	5 0 6	5 8 7	5 9 6	6 0 1	6 1 8	5 1 5	5 2 0	6 0 3	6 3 5	6 3 7	6 4 1	Total Tissues/ Tumors
Alimentary System																										
Esophagus	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Intestine large, colon	+	• +	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	51
Intestine large, rectum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Intestine large, cecum	+	• +	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	51
Intestine small, duodenum	+	• +	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Intestine small, jejunum	+	• +	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	52
Intestine small, ileum	+	• +	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	51
Liver	+	• +	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Carcinoma, metastatic, pancreas																		Х								2
Cholangiocarcinoma		Х		•••				Х			Х			•••	•••		Х	•••			Х		•••		•••	8
Cholangiocarcinoma, multiple	-			Х									X	Х	Х			Х		Х		Х	X	X	X	17
Hepatocellular adenoma	Х			•••				•••			Х		Х										Х	Х	Х	10
Hepatocellular adenoma, multiple				Х				Х								Х						77				3
Hepatocnolangioma																						л				2
Mesentery										+					+					+						8
Cincinal annual call consistents	+	-				+	+			+	+	+		+ V	+				+ V			+				29
Gingival, squamous cell carcinoma		м												<u>л</u>					<u>л</u>							10
Acinus adenoma	+	· IVI	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+ X	+	+	+	+	+	+	+	+	51
Acinus carcinoma																		х								2
Salivary glands	+	· +	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	52
Stomach forestomach	+	· +	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Squamous cell papilloma											x															1
Stomach, glandular	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Tooth		+				+			+			+			+	+		+		+		+				18
Cardiovascular System																										
Blood vessel	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Heart	+	M	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	52
Endocrine System																										
Adrenal cortex	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Adenoma																										1
Pheochromocytoma benign																										1
Adrenal medulla	+	+	+	+	+	$^+$	$^+$	$^+$	+	+	+	+	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	+	$^+$	+	$^+$	$^+$	$^+$	53
Pheochromocytoma malignant																										1
Pheochromocytoma benign			Х																							3
Islets, pancreatic	+	M	+	+	+	+	+	+	+	+	+	+	$^+$	+	+	+	$^+$	$^+$	$^+$	+	+	+	$^+$	$^+$	+	52
Parathyroid gland	+	+	+	Μ	+	М	+	+	Μ	$^+$	+	+	$^+$	+	+	+	$^+$	$^+$	$^+$	+	+	Μ	$^+$	$^+$	+	45
Pituitary gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	52
Adenoma												Х														1
Pars distalis, adenoma	Х	2	Х						Х						Х						Х					10
Thyroid gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	52
Bilateral, C-cell, adenoma C-cell, adenoma										Х	Х	Х								Х	Х	Х				3 8
Concerned De des Secretoria																										

**General Body System** 

None
Individual Animal Tumor Pathology	of Fei	nal	le I	Rat	s iı	n tł	ie 2	2-¥	lea	r (	Gav	vag	e S	tu	dy	of	тс	CD]	D:	10	)0 :	ng/	'kg					
	3	4	5	5	5	5	5	6	6	6	6	6	6	6	6	6	6	6	6	6	6	6	6	6	6	6	7	7
Number of Days on Study	9	7	4	6	8	9	9	0	1	1	1	1	3	3	4	4	4	4	4	5	5	6	6	7	8	9	Ó	1
	6	4	6	1	1	8	9	4	0	0	2	4	8	8	1	1	2	7	8	2	5	7	8	3	0	3	1	6
Conserve ID Normalian	5	5	6	5	5	6	6	5	5	5	5	5	5	6	5	6	5	6	6	5	6	5	5	6	6	5	5	5
Carcass ID Number	2	2	5	9	1	0	1	8 4	1	1	2	8	1	0	0	3	9	0	4 4	0	1	8	1	0	4	8	2	1
	5	1	0	,	0	0	4	4	2	4	4	0	/	,	/	0	0	2	4	,	0	,	1	4	2	5	2	,
Genital System																												
Clitoral gland	+	$^+$	$^+$	$^+$	+	$^+$	+	+	$^+$	$^+$	$^+$	+	+	+	+	+	+	+	+	+	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	$^+$	+
Ovary	+	$^+$	+	$^+$	+	$^+$	+	+	$^+$	$^+$	+	+	+	+	+	+	+	+	+	+	$^+$	$^+$	$^+$	$^+$	+	$^+$	$^+$	+
Dviduct																												
Uterus	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Carcinoma																												Х
Polyp stromal						х								х				х										
Polyp stromal, multiple														••			х											
Vagina								+																				
Schwannoma malignant								x																				
Hematopoietic System																												
Bone marrow	+	$^+$	+	$^+$	+	$^+$	+	$^+$	+	+	+	+	+	+	+	+	+	+	+	+	$^+$	$^+$	+	$^+$	+	$^+$	$^+$	+
Lymph node		$^+$	$^+$			$^+$													+					$^+$		$^+$		
ymph node, mandibular	+	+	$^+$	$^+$	+	+	+	+	$^+$	$^+$	+	$^+$	+	+	+	+	+	+	+	+	$^+$	$^+$	$^+$	$^+$	+	$^+$	$^+$	Ν
ymph node, mesenteric	+	+	$^+$	$^+$	+	+	+	+	$^+$	$^+$	+	$^+$	+	+	+	+	+	+	+	+	$^+$	$^+$	$^+$	$^+$	М	М	$^+$	+
Spleen	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
ĥymus	+	М	+	М	+	М	+	+	+	+	М	+	+	+	+	+	+	+	Μ	Μ	+	М	+	М	+	+	+	+
Integumentary System																												
Mammary gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Adenoma	v			37		37		v	37		37				v					37			37					
Fibroadenoma	А			л		л		л	л		л				л	v				л			л				v	
Skin	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	Х +	+	+	+	+	+	+	+	+	+	+	X +	+
Fibroma																												
Schwannoma malignant							Х																					
Musculoskeletal System																												
Bone	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Nervous System																												
Brain	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Respiratory System									,		,														,			
Lung Carcinoma, metastatic, pancreas	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Cystic keratinizing epithelioma multiple									Х													x		Х		Х		Х
Mediastinum, sarcoma						Х																						
Nose	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Squamous cell carcinoma metastatic																												
Squamous cell carcinoma, metastatic, oral mucosa																									,			X

Individual Animal Tumor Pathology	of Fe	ma	le l	Rat	s i	n tl	he	2-3	Yea	ar (	Ga	vag	ge S	Stu	ldy	of	T C	CD	D:	1	00	ng	/kg	5			
Number of Days on Study	7 1 7	7 2 1	7 2 2	7 2 3	7 3 1	7 3 2	7 3 2	7 3 2	7 3 2	7 3 2	7 3 3	7 3 3	7 3 3	7 3 3	7 3 3	7 3 3											
Carcass ID Number	5 9 7	5 1 8	6 3 3	6 1 2	5 1 3	5 1 6	5 8 1	5 8 3	6 0 8	6 1 3	6 1 5	6 1 7	6 3 1	6 3 2	5 0 6	5 8 7	5 9 6	6 0 1	6 1 8	5 1 5	5 2 0	6 0 3	6 3 5	6 3 7	6 4 1	To Tissu Tum	otal 1es/ 1ors
Genital System	1																									5	· 7
Clitoral gland	+	+	+	+	+	+	+	++	+	+	+	+	+	+	++	+	+	++	++	++	+	++	++	+	++	5	3
Oviduct													+						'	'		'	'			5	1
Uterus	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	5	3
Carcinoma																											1
Polyp stromal			Х						Х										Х			Х					7
Polyp stromal, multiple																											1
Vagina Schwannoma malignant																											1
Hematopoietic System																											
Bone marrow	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	5	3
Lymph node								+										+				+				5	9
Lymph node, mandibular	+	+	+	++	+	+	+	++	+	+	+	++	+	+	++	+	+	++	++	++	+	++	++	+	++	5	2 1
Spleen	+	Ń	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	5	2
Thymus	+	+	+	+	+	+	+	+	+	+	+	+	М	+	+	М	+	+	+	+	М	+	+	+	+	4	2
Integumentary System																											
Mammary gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	5	3
Adenoma																								X	17	2	1
Fibroadenoma multinle			Х			Х	Х	v	Х	Х	v	Х			Х		Х			Х				Х	Х	2	20 - 4
Skin	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	5	3
Fibroma	Х																										1
Schwannoma malignant																											1
Musculoskeletal System	Т	+	-	-	+	-	-	+	-	-	-	-	-	-	+	+	+	+	-	-	-	-	-	-	-	5	2
Bolle	T	Ŧ	т	т	т	т	т	т	т	т	т	т	т	т	т	т	т	т	т	т	т	т	т	т	т	5	5
Nervous System																											
Brain	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	5	3
Respiratory System																										_	
Lung	+	М	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	5	1
Custic keratinizing enithelioma																		л					x	v	v		1
Cystic keratinizing epithelioma. multiple																	х						Λ	Λ	1		2
Mediastinum, sarcoma																	-										1
Nose	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	5	3
Squamous cell carcinoma, metastatic,																											
oral mucosa																									,	-	1
Haunda	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	3	5

TABLE A2

Individual Animal Tumor Pathology	of Fei	na	le l	Rat	s ii	n tl	he	2-}	Yea	ar (	Gar	vag	ge S	Stu	dy	of	т	CD	D:	1(	<b>)0</b> 1	ng/	kg					
	3	4	5	5	5	5	5	6	6	6	6	6	6	6	6	6	6	6	6	6	6	6	6	6	6	6	7	7
Number of Days on Study	9	7	4	6	8	9	9	0	1	1	1	1	3	3	4	4	4	4	4	5	5	6	6	7	8	9	0	1
	6	4	6	1	1	8	9	4	0	0	2	4	8	8	1	1	2	7	8	2	5	7	8	3	0	3	1	6
	5	5	6	5	5	6	6	5	5	5	5	5	5	6	5	6	5	6	6	5	6	5	5	6	6	5	5	5
Carcass ID Number	2	2	3	9	1	0	1	8	1	1	2	8	1	0	0	3	9	0	4	0	1	8	1	0	4	8	2	1
	5	1	6	9	0	6	4	4	2	4	4	6	7	9	7	8	8	2	4	9	6	9	1	4	2	5	2	9
Special Senses System																												
Ear	+																+											
Eye	+	+	+	+	$^+$	+	$^+$	+	$^+$	+	+	+	+	$^+$	+	$^+$	+	+	+	+	$^+$	$^+$	$^+$	$^+$	$^+$	+	+	+
Iarderian gland	+	+	+	+	$^+$	+	$^+$	+	$^+$	+	+	+	+	$^+$	+	$^+$	+	+	+	+	$^+$	$^+$	$^+$	$^+$	$^+$	+	+	+
Squamous cell carcinoma, metastatic,																												
oral mucosa			Х							Х																		
Urinary System																												
Kidney	+	$^+$	+	+	+	+	$^+$	+	+	+	$^+$	+	$^+$	+	+	+	+	+	$^+$	$^+$	$^+$	+	+	$^+$	$^+$	+	$^+$	+
Urinary bladder	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Systemic Lesions																												
Multiple organs Lymphoma malignant	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+

Individual Animal Tumor Patholog	y of Fei	na	le l	Rat	s i	n t	he	2-	Yea	ar (	Ga	vaş	ge	Stu	ıdy	of	T	CD	D:	1	00	ng	/kg	ç		
	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	
Number of Days on Study	1	2	2	2	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	
	7	1	2	3	1	1	1	1	1	1	1	1	1	1	2	2	2	2	2	3	3	3	3	3	3	
	5	5	6	6	5	5	5	5	6	6	6	6	6	6	5	5	5	6	6	5	5	6	6	6	6	Total
Carcass ID Number	9	1	3	1	1	1	8	8	0	1	1	1	3	3	0	8	9	0	1	1	2	0	3	3	4	Tissues/
	7	8	3	2	3	6	1	3	8	3	5	7	1	2	6	7	6	1	8	5	0	3	5	7	1	Tumors
Special Senses System																										
Ear	1																									2 52
Eye Uardarian aland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Squamous cell carcinoma metastatic	т	Ŧ	Ŧ	Ŧ	т	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	т	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	Ŧ	55
oral mucosa														Х												3
Urinary System																										
Kidney	+	$^+$	+	+	$^+$	+	$^+$	+	+	+	+	+	+	+	+	$^+$	$^+$	$^+$	$^+$	+	+	+	+	+	+	53
Urinary bladder	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Systemic Lesions																										
Multiple organs	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	53
Lymphoma malignant																		Х				Х				2

TABLE A2           Individual Animal Tumor Pathology of	of Fei	na	le 1	Rat	ts i	n tl	he	2-}	lea	ar (	Ga	vaş	ge S	Stu	ıdy	of	T	CD	D:	1	00	ng/	/kg	(S	top-Exposure)
Number of Days on Study	2 1 5	3 4 5	3 9 4	4 1 0	4 7 4	4 8 4	5 2 0	5 3 7	5 7 8	5 7 8	5 7 8	5 8 4	6 0 4	6 1 4	6 2 9	6 4 1	6 4 3	6 4 5	6 4 5	6 5 3	6 5 6	6 7 3	7 0 2	7 0 2	7 1 8
Carcass ID Number	5 4 6	5 6 6	5 4 7	5 3 7	5 4 2	5 3 2	5 6 1	5 5 5	5 2 9	5 7 0	5 7 4	5 6 4	5 6 8	5 5 0	5 4 0	5 7 2	5 3 6	5 4 3	5 6 3	5 3 0	5 7 3	5 3 1	5 2 7	5 5 7	5 5 2
Alimentary System																									
Esophagus	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	$^+$	+	+	+	+	+	+	$^+$	+
Intestine large, colon	+	+	+	+	+	+	+	М	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Intestine large, rectum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Fibrous histiocytoma, metastatic, skin			X					м																	
Intestine large, cecum	+	+	+	+	+	+	+	M	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Intestine small, iciunum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Schwannoma malignant, metastatic, heart																								x	
Intestine small, ileum	+	+	+	+	+	+	+	М	+	+	М	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Liver	+	+	+	+	+	+	$^+$	+	$^+$	+	+	+	+	+	+	$^+$	+	+	+	+	+	+	+	$^+$	+
Cholangiocarcinoma Cholangiocarcinoma, multiple																									
Fibrous histiocytoma, metastatic, skin Hepatocellular adenoma			Х																				x		
Schwannoma malignant, metastatic, heart																								х	
Mesentery	+																								
Oral mucosa		+					+	+				+					+				+		+	+	+
Gingival, squamous cell carcinoma							Х	Х				Х									Х		Х		
Pancreas	+	+	+	+	+	+	+	Μ	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Acinus, carcinoma																									
Stomach forestomach	+	+	+	+	+	+	+	+	+	+	+	+	+	+		+	+	+	+	+	+	+	+	+	+
Stomach, glandular	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Tongue																							+		
Squamous cell carcinoma																							Х		
Tooth		+				+	+				+	+					+						+		
Cardiovascular System																									
Blood vessel	+	+	+	+	+	+	+	+	+	+	Μ	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Aorta, fibrous histiocytoma, metastatic, skin			X																						
Caroinoma matastatic mammany gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Schwannoma malignant						Х		л																Х	
Endocrine System																									
Adrenal cortex	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Capsule, schwannoma malignant, metastatic, heart																								X	
Aurenai medulla Dhaochromocytoma complay	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	т
Pheochromocytoma benign																								x	
Islets, pancreatic	+	+	+	+	+	+	+	М	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Parathyroid gland	+	+	+	+	М	+	+	+	+	+	М	+	+	+	М	+	+	+	+	+	+	+	+	+	+
Pituitary gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Pars distalis, adenoma												Х		Х		Х					Х		Х	Х	Х
Thyroid gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	М	+	+	+	+	+	+	+	+	+	+
Bilateral, C-cell, adenoma												v					v						v		v
C-cell, carcinoma												Х					Х						Х		Λ

															·							-	-	-	-	
	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	7	
Number of Days on Study	2	2	2	2	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	
	4	6	6	9	1	1	1	1	1	1	1	1	1	1	2	2	2	2	2	2	3	3	3	3	3	
	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5	Total
Carcass ID Number	6	3	4	5	2	3	3	3	5	5	5	6	6	6	3	4	4	4	5	7	2	4	5	6	7	Tissues/
	0	3	1	9	8	4	5	8	3	4	6	2	5	9	9	4	5	9	8	5	6	8	1	7	1	Tumors
Alimentary System																										
Esophagus	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	50
Intestine large, colon	+	+	$^+$	$^+$	+	+	+	+	+	+	$^+$	+	+	+	+	$^+$	$^+$	+	+	+	$^+$	+	+	+	+	49
Intestine large, rectum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	$^+$	+	+	+	+	50
Fibrous histiocytoma, metastatic, skin																										1
Intestine large, cecum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	49
Intestine small, duodenum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	49
Intestine small, jejunum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	50
Schwannoma malignant, metastatic, heart																										1
Intestine small, ileum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	48
Liver	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	50
Cholangiocarcinoma Cholangiocarcinoma multinla				Х															v							1
Cholangiocarcinoma, multiple																			л		v					1
Fibrous histiocytoma metastatic skin																					Λ					1
Henatocellular adenoma		x																								2
Schwannoma malignant metastatic heart		11																								- 1
Mesentery																										1
Oral mucosa													+								+					11
Gingival, squamous cell carcinoma																										5
Pancreas	+	+	$^+$	$^+$	+	+	+	+	+	+	$^+$	+	+	+	+	$^+$	$^+$	+	+	+	$^+$	+	+	+	+	49
Acinus, carcinoma																									Х	1
Salivary glands	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	$^+$	+	+	+	+	49
Stomach, forestomach	+	$^+$	+	+	$^+$	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	$^+$	+	+	+	+	50
Stomach, glandular	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	50
Tongue																										1
Squamous cell carcinoma																										1
Tooth						+			+				+	+							+					12
Cardiovascular System	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	49
Aorta fibrous histiocytoma metastatic skin		·			·							·			·	·	·									1
Heart	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	50
Carcinoma, metastatic, mammary gland																										1
Schwannoma malignant												Х														3
Endocrine System																										
Adrenal cortex	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	50
heart																										1
Adrenal medulla	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	50
Pheochromocytoma complex					Х																					1
Pheochromocytoma benign																										1
Islets, pancreatic	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	49
Parathyroid gland	Μ	+	+	+	+	+	М	+	+	+	+	+	+	+	+	+	+	+	+	+	М	+	+	+	+	44
Pituitary gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	50
Pars distalis, adenoma	Х		Х	Х			Х			Х		Х	Х			Х			Х	Х				Х	Х	19
Thyroid gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	49
Bilateral, C-cell, adenoma	_				Х						Х															2
C-cell, adenoma	Х		Х			<b>1</b> 7		Х				Х				Х						Х			Х	11
I CALL CORCIDODO						X																				1

Individual Animal Tumor Pathology o	f Fei	nal	le l	Rat	ts i	n tl	he	2-1	Yea	ar (	Ga	vaş	ge S	Stu	ıdy	of	T	CD	D:	10	00 1	ng/	kg	(S	top-Exposure)
Number of Days on Study	2 1 5	3 4 5	3 9 4	4 1 0	4 7 4	4 8 4	5 2 0	5 3 7	5 7 8	5 7 8	5 7 8	5 8 4	6 0 4	6 1 4	6 2 9	6 4 1	6 4 3	6 4 5	6 4 5	6 5 3	6 5 6	6 7 3	7 0 2	7 0 2	7 1 8
Carcass ID Number	5 4 6	5 6 6	5 4 7	5 3 7	5 4 2	5 3 2	5 6 1	5 5 5	5 2 9	5 7 0	5 7 4	5 6 4	5 6 8	5 5 0	5 4 0	5 7 2	5 3 6	5 4 3	5 6 3	5 3 0	5 7 3	5 3 1	5 2 7	5 5 7	5 5 2
General Body System None																									
Genital System Clitoral gland Ovary Carcinoma, metastatic, uterus Oviduct Uterus Carcinoma Polyp stromal Polyp stromal, multiple Squamous cell carcinoma	+++++	++++	+++++	M + +	+++++	+ + +	++++++	+ M +	+++++	+++++	+++++	+++++	+++++	+++++	+++++	+++++	++++++	+++++	+++++	+++++	+++++	++++++	+++++	++++++	+ + +
Hematopoietic System Bone marrow Fibrous histiocytoma, metastatic, skin Lymph node Lymph node, mandibular Lymph node, mesenteric Fibrous histiocytoma, metastatic, skin Spleen Fibrous histiocytoma, metastatic, skin Hemangiosarcoma Schwannoma malignant, metastatic, heart	++++++	+ + + +	+ X + + X + X	++++++	+ + + +	+++++	++++++	+ + M M	+ + + +	+++++	+ + + + +	+ + + + +	+ + + +	++++++	+ + +	+ + + +	+ + + +	+++++	+++++	+++++	++++++	++++++	++++++	+ + + + +	+ + + +
Thymus Fibrous histiocytoma, metastatic, skin Thymoma malignant	+	+	+ X	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	М	+	+
Integumentary System Mammary gland Adenolipoma Carcinoma Fibroadenoma Fibroadenoma, multiple	+	+ X	+	+ X	+ X	+	+	+ X	+ X X	+ X	+ X	+	+ X	+	+	+ X	+ X	+ X	+ X	+ X	+	+ X	+ X	+	+ X
Skin Fibroma Fibrous histiocytoma Liposarcoma	+	+	+ X	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Musculoskeletal System Bone Squamous cell carcinoma, metastatic, tongue Skeletal muscle Fibrous histiocytoma, metastatic, skin	+	+	+ + X	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+ X	+	+
Nervous System Brain	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+

TABLE A2           Individual Animal Tumor Pathology of	f Fei	na	le I	Rat	ts ii	n tl	he	2-3	Yea	ar (	Ga	vag	ge S	Stu	ıdy	of	т	CD	D:	1	00	ng/	/kg	(S	top	)-Exposure)
Number of Days on Study	7 2 4	7 2 6	7 2 6	7 2 9	7 3 1	7 3 1	7 3 1	7 3 1	7 3 1	7 3 1	7 3 1	7 3 1	7 3 1	7 3 1	7 3 2	7 3 2	7 3 2	7 3 2	7 3 2	7 3 2	7 3 3	7 3 3	7 3 3	7 3 3	7 3 3	
Carcass ID Number	5 6 0	5 3 3	5 4 1	5 5 9	5 2 8	5 3 4	5 3 5	5 3 8	5 5 3	5 5 4	5 5 6	5 6 2	5 6 5	5 6 9	5 3 9	5 4 4	5 4 5	5 4 9	5 5 8	5 7 5	5 2 6	5 4 8	5 5 1	5 6 7	5 7 1	Total Tissues/ Tumors
General Body System None																										
Genital System Clitoral gland Ovary Carcinoma, metastatic, uterus Oviduct Uterus Carcinoma Polyp stromal Polyp stromal, multiple Squamous cell carcinoma	++++	+++++	+++++	+ + X	+++++	+ + X + X X	+++++	+++++	+++++	+++++	+++++	+++++	+++++	+++++	+++++	+++++	+ + X X	++++++	+++++	+ + +	+ + X	+++++	+++++	+ + X	+++++	49 49 1 2 50 2 3 1 2
Hematopoietic System Bone marrow Fibrous histiocytoma, metastatic, skin Lymph node Lymph node, mandibular Lymph node, mesenteric Fibrous histiocytoma, metastatic, skin Spleen Fibrous histiocytoma, metastatic, skin Hemangiosarcoma Schwannoma malignant, metastatic, heart Thymus Fibrous histiocytoma, metastatic, skin Thymoma malignant	+ + + +	+ + + +	+ + + +	+ + + X +	+ + + +	+ + + +	+ + + +	+ + + +	+ + + + X	+ + + +	+ + + +	+ + + +	+ + + +	+ + + +	+ + + +	+ + + +	+ + + +	+ + + +	+ + + +	+ + + +	+ + + +	+ + + +	+ + + +	+ + + +	+ + + + +	50 1 5 49 49 1 49 1 1 1 1 1 1 1 1
Integumentary System Mammary gland Adenolipoma Carcinoma Fibroadenoma Fibroadenoma, multiple Skin Fibroma Fibroma Fibrous histiocytoma Liposarcoma	+ X +	+ X +	+ X +	+ X +	+	+	+ X +	+ X +	+ X +	+ +	+ X +	+ X +	+	+ X +	+	+	+ X +	+ X + X	+ X +	+ X +	+ X +	+ X +	+	+ X +	+ + X	50 1 19 13 50 1 1 1
Musculoskeletal System Bone Squamous cell carcinoma, metastatic, tongue Skeletal muscle Fibrous histiocytoma, metastatic, skin	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	50 1 1 1
Nervous System Brain	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	50

TABLE A2

Individual Animal Tumor Pathology of I	Fer	nal	e I	Rat	s ii	n tl	he	2-}	Yea	ar (	Ga	vag	ge S	Stu	ıdy	of	т	CD	D:	1(	<b>)0</b> :	ng/	kg	(S	top-Exposure)
Number of Days on Study	2 1 5	3 4 5	3 9 4	4 1 0	4 7 4	4 8 4	5 2 0	5 3 7	5 7 8	5 7 8	5 7 8	5 8 4	6 0 4	6 1 4	6 2 9	6 4 1	6 4 3	6 4 5	6 4 5	6 5 3	6 5 6	6 7 3	7 0 2	7 0 2	7 1 8
Carcass ID Number	5 4 6	5 6 6	5 4 7	5 3 7	5 4 2	5 3 2	5 6 1	5 5 5	5 2 9	5 7 0	5 7 4	5 6 4	5 6 8	5 5 0	5 4 0	5 7 2	5 3 6	5 4 3	5 6 3	5 3 0	5 7 3	5 3 1	5 2 7	5 5 7	5 5 2
Respiratory System																									
Lung Carcinoma, metastatic, mammary gland Fibrous histiocytoma, metastatic, skin	+	+	+ X	+	+	+	+	+ X	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Schwannoma malignant, metastatic, heart																								Х	
Nose Trachea	+ +																								
Special Senses System																									
Eye Harderian gland	+ +																								
Urinary System Kidney	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Fibrous histiocytoma, metastatic, skin Urinary bladder	+	+	X +	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Systemic Lesions Multiple organs Adenolipoma Lymphoma malignant	+	+ X	+	+	+	+	+	+	+ X	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+

Individual Animal Tumor Pathology	of Fer	na	le l	Rat	ts ii	n t	he	2-	Yea	ar (	Ga	vag	ge S	Stu	ıdy	of	T T	CD	D:	1	00	ng/	′kg	; (S	top	-Exposure)
Number of Days on Study	7 2 4	7 2 6	7 2 6	7 2 9	7 3 1	7 3 2	7 3 2	7 3 2	7 3 2	7 3 2	7 3 2	7 3 3	7 3 3	7 3 3	7 3 3	7 3 3										
Carcass ID Number	5 6 0	5 3 3	5 4 1	5 5 9	5 2 8	5 3 4	5 3 5	5 3 8	5 5 3	5 5 4	5 5 6	5 6 2	5 6 5	5 6 9	5 3 9	5 4 4	5 4 5	5 4 9	5 5 8	5 7 5	5 2 6	5 4 8	5 5 1	5 6 7	5 7 1	Total Tissues/ Tumors
Respiratory System																										
Lung Carcinoma, metastatic, mammary gland Fibrous histiocytoma, metastatic, skin Schwannoma malignant, metastatic, heart	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	50 1 1 1
Nose Trachea	++	+ +	++	++	+ +	+ +	+ +	++	+ +	++	++	+++	+ +	++	++	+ +	++	+++	++	+ +	+ +	+ +	+++	++	+ +	50 50
Special Senses System																										
Eye Harderian gland	+++	+ +	++	+ +	++	+ +	+ +	+ +	++	+ +	50 50															
Urinary System Kidney Fibrous histiocytoma metastatic skin	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	50
Urinary bladder	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	50
Systemic Lesions Multiple organs Adenolipoma Lymphoma malignant	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	50 1 1

# TABLE A3a Statistical Analysis of Primary Neoplasms in Female Rats in the 2-Year Gavage Study of TCDD

	Vehicle Control	3 ng/kg	10 ng/kg	22 ng/kg	46 ng/kg	100 ng/kg
Adrenal Medulla: Benign Pheochro	omocytoma					
Overall rate <sup>a</sup> .	3/53 (6%)	2/54 (4%)	2/53 (4%)	3/53 (6%)	3/53 (6%)	3/53 (6%)e
Adjusted rate <sup>b</sup>	7.6%	5.8%	5.2%	8.8%	7.7%	7.0%
Terminal rate <sup>c</sup>	3/25 (12%)	2/21 (10%)	2/23 (9%)	3/19 (16%)	2/22 (9%)	0/21 (0%)
First incidence (days)	731 (T)	731 (T)	731 (T)	731 (T)	708	642
Poly-3 test <sup>u</sup>	P=0.542	P=0.561N	P=0.509N	P=0.594	P=0.658	P=0.622N
Liver: Cholangiocarcinoma						
Overall rate	0/53 (0%)	0/54 (0%)	0/53 (0%)	1/53 (2%)	4/53 (8%)	25/53 (47%)
Adjusted rate	0.0%	0.0%	0.0%	2.9%	10.3%	54.9%
Terminal rate	0/2,5 (0%)	0/21 (0%)	0/23 (0%)	0/19 (0%)	4/22 (18%)	13/21 (62%)
First incidence (days)	1	_	—	592	731 (T)	610
Poly-3 test	P<0.001	g	—	P=0.474	P=0.057	P<0.001
Liver: Hepatocellular Adenoma						
Overall rate	0/53 (0%)	0/54 (0%)	0/53 (0%)	0/53 (0%)	1/53 (2%)	13/53 (25%)
Adjusted rate	0.0%	0.0%	0.0%	0.0%	2.6%	29.9%
Terminal rate	0/25 (0%)	0/21 (0%)	0/23 (0%)	0/19 (0%)	0/22 (0%)	7/21 (33%)
First incidence (days)		—	—	—	720	642
Poly-3 test	P<0.001	—	—	—	P=0.497	P<0.001
Lung: Cystic Keratinizing Epitheli	oma					
Overall rate	0/53 (0%)	0/54 (0%)	0/53 (0%)	0/52 (0%)	0/53 (0%)	9/52 (17%)
Adjusted rate	0.0%	0.0%	0.0%	0.0%	0.0%	21.1%
Terminal rate	0/25 (0%)	0/21 (0%)	0/23 (0%)	0/19 (0%)	0/22 (0%)	4/21 (19%)
First incidence (days)	-	—	—	—	—	610
Poly-3 test	P<0.001	_	_	_	_	P=0.002
Mammary Gland: Fibroadenoma						
Overall rate	33/53 (62%)	40/54 (74%)	34/53 (64%)	29/53 (55%)	36/53 (68%)	24/53 (45%)
Adjusted rate	71.0%	81.7%	74.2%	70.3%	74.5%	51.0%
Terminal rate	16/25 (64%)	16/21 (76%)	16/23 (70%)	12/19 (63%)	15/22 (68%)	12/21 (57%)
First incidence (days)	185	241	236	453	141	396
Poly-3 test	P<0.001N	P=0.147	P=0.452	P=0.570N	P=0.435	P=0.031N
Mammary Gland: Fibroadenoma	or Adenoma					
Overall rate	35/53 (66%)	40/54 (74%)	35/53 (66%)	29/53 (55%)	36/53 (68%)	24/53 (45%)
Adjusted rate	74.7%	81.7%	76.4%	70.3%	74.5%	51.0%
Terminal rate	17/25 (68%)	16/21 (76%)	17/23 (74%)	12/19 (63%)	15/22 (68%)	12/21 (5/%)
Poly-3 test	185 P<0.001N	241 P=0.269	236 P=0.522	453 P=0.407N	141 P=0.589N	396 P=0.010N
Mammary Gland: Carcinoma						
Overall rate	4/53 (8%)	8/54 (15%)	4/53 (8%)	2/53 (4%)	2/53 (4%)	0/53 (0%)
Adjusted rate	9.9%	21.6%	10.2%	5.7%	5.1%	0.0%
Terminal rate	2/25 (8%)	4/21 (19%)	1/23 (4%)	0/19 (0%)	1/22 (5%)	0/21 (0%)
Poly-3 test	469 P=0.004N	241 P=0.132	659 P=0.626	338 P=0.401N	673 P=0.350N	 P=0.053N
Mammary Gland: Adenoma or Ca	6/53 (11%)	8/5/ (150/)	5/53 (00/)	2/53 (40/)	2/53 (40/)	1/53 (20/)
Adjusted rate	0/33 (1170) 14 70/	0/5+(15%) 21.6%	12.8%	5 7%	2/33 (470) 5 10%	2 40%
Terminal rate	14./70 3/25 (120/2)	21.070 <u>4/21 (1004</u> )	12.070 2/23 (00/2)	0/10 (0%)	J.170 1/22 (5%)	2.470 1/21 (5%)
First incidence (days)	469	241	659	338	673	731 (T)
Polv-3 test	P=0.005N	P=0.310	P=0.530N	P=0.182N	P=0.143N	P=0.048N
· · · · · ·						

### TABLE A3a

Statistical Analysis of Primary Neoplasms in Female Rats in the 2-Year Gavage Study of TCDD

	Vehicle Control	3 ng/kg	10 ng/kg	22 ng/kg	46 ng/kg	100 ng/kg
Mammary Gland: Fibroadenoma, A	denoma, or Car	cinoma				
Overall rate	35/53 (66%)	43/54 (80%)	37/53 (70%)	31/53 (58%)	37/53 (70%)	24/53 (45%)
Adjusted rate	74.7%	86.2%	80.2%	73.2%	76.2%	51.0%
Terminal rate	17/25 (68%)	17/21 (81%)	17/23 (74%)	12/19 (63%)	15/22 (68%)	12/21 (57%)
First incidence (days)	185	241	236	338	141	396
Poly-3 test	P<0.001N	P=0.107	P=0.343	P=0.533N	P=0.526	P=0.010N
Oral Mucosa: Squamous Cell Carci	noma					
Overall rate	1/53 (2%)	2/54 (4%)	1/53 (2%)	0/53 (0%)	4/53 (8%)	10/53 (19%)
Adjusted rate	2.5%	5.7%	2.6%	0.0%	10.2%	22.0%
Terminal rate	0/25(0%)	1/21 (5%)	0/23(0%)	0/19 (0%)	1/22 (5%)	2/21 (10%)
First incidence (days)	366	647	578		659	546
Poly-3 test	P<0.001	P=0.449	P=0.755	P=0.533N	P=0.169	P=0.007
Pancreas: Adenoma or Carcinoma						
Overall rate	0/51 (0%)	0/54 (0%)	0/52 (0%)	0/53 (0%)	0/52 (0%)	3/51 (6%)
Adjusted rate	0.0%	0.0%	0.0%	0.0%	0.0%	7 3%
Terminal rate	0/25(0%)	0/21 (0%)	0/23(0%)	0/19 (0%)	0/22 (0%)	2/21 (10%)
First incidence (days)						641
Poly-3 test	P<0.001	_	—	—	—	P=0.129
Pituitary Gland (Pars Distalis): Ade	noma					
Overall rate	25/53 (47%)	20/54 (37%)	26/52 (50%)	15/53 (28%)	20/53 (38%)	11/52 (21%)
Adjusted rate	58.4%	55.3%	64.2%	41.2%	47.3%	25.0%
Terminal rate	15/25 (60%)	16/21 (76%)	15/23 (65%)	9/19 (47%)	10/22 (46%)	4/21 (19%)
First incidence (days)	474	604	550	545	444	604
Poly-3 test	P<0.001N	P=0.478N	P=0.369	P=0.087N	P=0.202N	P<0.001N
Skin: Fibroma, Myxosarcoma, or Fi	brosarcoma					
Overall rate	1/53 (2%)	0/54 (0%)	3/53 (6%)	1/53 (2%)	0/53 (0%)	1/53 (2%)
Adjusted rate	2.5%	0.0%	7.5%	2.9%	0.0%	2.3%
Terminal rate	0/25 (0%)	0/21 (0%)	0/23 (0%)	1/19 (5%)	0/22 (0%)	0/21 (0%)
First incidence (days)	548	_ ``	578	731 (T)	_ ` `	717
Poly-3 test	P=0.439N	P=0.529N	P=0.303	P=0.724	P=0.506N	P=0.747N
Thyroid Gland (C-cell): Adenoma						
Overall rate	20/52 (38%)	15/54 (28%)	17/53 (32%)	16/51 (31%)	13/53 (25%)	11/52 (21%)
Adjusted rate	46.7%	40.4%	43.1%	43.2%	31.6%	25.6%
Terminal rate	10/25 (40%)	10/21 (48%)	13/23 (57%)	8/19 (42%)	8/22 (36%)	6/21 (29%)
First incidence (days)	474	462	626	338	486	638
Poly-3 test	P=0.010N	P=0.366N	P=0.459N	P=0.464N	P=0.111N	P=0.030N
Thyroid Gland (C-cell): Adenoma of	r Carcinoma					
Overall rate	21/52 (40%)	15/54 (28%)	17/53 (32%)	16/51 (31%)	14/53 (26%)	11/52 (21%)
Adjusted rate	49.0%	40.4%	43.1%	43.2%	34.0%	25.6%
Terminal rate	11/25 (44%)	10/21 (48%)	13/23 (57%)	8/19 (42%)	9/22 (41%)	6/21 (29%)
First incidence (days)	474	462	626	338	486	638
Poly-3 test	P=0.008N	P=0.288N	P=0.374N	P=0.380N	P=0.115N	P=0.017N
Uterus: Stromal Polyp						
Overall rate	11/53 (21%)	7/53 (13%)	4/53 (8%)	11/53 (21%)	10/53 (19%)	8/53 (15%)
Adjusted rate	27.4%	19.6%	10.3%	30.3%	24.7%	18.2%
Terminal rate	8/25 (32%)	5/21 (24%)	2/23 (9%)	6/19 (32%)	5/22 (23%)	3/21 (14%)
First incidence (days)	645	526	659	557	590	598
Poly-3 test	P=0.404N	P=0.296N	P=0.045N	P=0.490	P=0.492N	P=0.223N

#### TABLE A3a

Statistical Analysis of Primary Neoplasms in Female Rats in the 2-Year Gavage Study of TCDD

	Vehicle Control	3 ng/kg	10 ng/kg	22 ng/kg	46 ng/kg	100 ng/kg
Uterus: Squamous Cell Carcinoma						
Overall rate	0/53 (0%)	0/53 (0%)	0/53 (0%)	0/53 (0%)	5/53 (9%)	0/53 (0%)
Adjusted rate	0.0%	0.0%	0.0%	0.0%	12.3%	0.0%
Terminal rate	0/25 (0%)	0/21 (0%)	0/23 (0%)	0/19 (0%)	0/22 (0%)	0/21 (0%)
First incidence (days)	_	_	_	_	589	_
Poly-3 test	P=0.337	—	—		P=0.032	—
All Organs: Benign Neoplasms						
Overall rate	45/53 (85%)	49/54 (91%)	45/53 (85%)	40/53 (75%)	46/53 (87%)	42/53 (79%)
Adjusted rate	92.2%	97.1%	95.6%	90.2%	92.9%	84.7%
Terminal rate	23/25 (92%)	21/21 (100%)	22/23 (96%)	17/19 (90%)	21/22 (96%)	18/21 (86%)
First incidence (days)	185	241	236	338	141	396
Poly-3 test	P=0.010N	P=0.232	P=0.383	P=0.511N	P=0.610	P=0.173N
All Organs: Malignant Neoplasms						
Overall rate	13/53 (25%)	19/54 (35%)	13/53 (25%)	11/53 (21%)	24/53 (45%)	34/53 (64%)
Adjusted rate	29.6%	45.3%	30.3%	28.3%	56.5%	69.9%
Terminal rate	4/25 (16%)	7/21 (33%)	3/23 (13%)	3/19 (16%)	12/22 (55%)	14/21 (67%)
First incidence (days)	169	200	240	338	518	546
Poly-3 test	P<0.001	P=0.093	P=0.566	P=0.547N	P=0.008	P<0.001
All Organs: Benign or Malignant Neoplasms						
Overall rate	47/53 (89%)	54/54 (100%)	48/53 (91%)	44/53 (83%)	50/53 (94%)	50/53 (94%)
Adjusted rate	92.7%	100.0%	97.7%	94.4%	97.9%	96.6%
Terminal rate	23/25 (92%)	21/21 (100%)	22/23 (96%)	18/19 (95%)	21/22 (96%)	20/21 (95%)
First incidence (days)	169	200	236	338	141	396
Poly-3 test	P=0.560	P=0.046	P=0.223	P=0.535	P=0.205	P=0.317

(T) Terminal sacrifice

<sup>a</sup> Number of neoplasm-bearing animals/number of animals examined. Denominator is number of animals examined microscopically for adrenal gland, liver, lung, pancreas, pituitary gland, and thyroid gland; for other tissues, denominator is number of animals necropsied.

Poly-3 estimated neoplasm incidence after adjustment for intercurrent mortality

<sup>c</sup> Observed incidence at terminal kill

<sup>d</sup> Beneath the vehicle control incidence is the P value associated with the trend test. Beneath the dosed group incidence are the P values corresponding to pairwise comparisons between the vehicle controls and that dosed group. The Poly-3 test accounts for the differential mortality in animals that do not reach terminal sacrifice. A negative trend or a lower incidence in a dose group is indicated by N.

<sup>e</sup> One malignant pheochromocytoma occurred in an animal that also had a benign pheochromocytoma.

<sup>1</sup> Not applicable; no neoplasms in animal group

<sup>g</sup> Value of statistic cannot be computed.

TABLE A	A3b
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### Statistical Analysis of Primary Neoplasms in Female Rats in the Stop-Exposure Evaluation of TCDD

	Vehicle	100 ng/kg	100 ng/kg	
	Control		(Stop-Exposure)	
Adrenal Medulla: Benign Pheochromocytoma				
Overall rate <sup>a</sup> .	3/53 (6%)	3/53 (6%)	1/50 (2%)	
Adjusted rate	7.6%	7.0%	2.6%	
Terminal rate <sup>C</sup>	3/25 (12%)	0/21 (0%)	0/21 (0%)	
First incidence (days)	731 (T)	642	702	
Poly-3 test	(01(1)	P=0.622N	P=0.319N	
Poly-3 test			P=0.361N	
Adrenal Medulla: Benign or Malignant Pheochromo	cytoma			
Overall rate	3/53 (6%)	3/53 (6%)	2/50 (4%)	
Adjusted rate	7.6%	7.0%	5.2%	
Terminal rate	3/25 (12%)	0/21 (0%)	1/21 (5%)	
First incidence (days)	731 (T)	642	702	
Poly-3 test	(1)	P=0.622N	P=0.518N	
Poly-3 test			P=0.567N	
Heart: Schwannoma Malignant				
Overall rate	0/53 (0%)	0/52 (0%)	3/50 (6%)	
Adjusted rate	0.0%	0.0%	7 7%	
Terminal rate	0/25 (0%)	0/21(0%)	1/21 (5%)	
First incidence (days)	f	_	484	
Polv-3 test		g	P=0.116	
Poly-3 test			P=0.110	
Liver: Cholangiocarcinoma				
Overall rate	0/53 (0%)	25/53 (47%)	2/50 (4%)	
Adjusted rate	0.0%	54.9%	5.2%	
Terminal rate	0/25 (0%)	13/21 (62%)	1/21 (5%)	
First incidence (days)	_ `	610	729	
Poly-3 test		P<0.001	P=0.232	
Poly-3 test			P<0.001N	
Liver: Hepatocellular Adenoma				
Overall rate	0/53 (0%)	13/53 (25%)	2/50 (4%)	
Adjusted rate	0.0%	29.9%	5.2%	
Terminal rate	0/25 (0%)	7/21 (33%)	0/21 (0%)	
First incidence (days)	_ ` `	642	702	
Poly-3 test		P<0.001	P=0.233	
Poly-3 test			P=0.004N	
Lung: Cystic Keratinizing Epithelioma				
Overall rate	0/53 (0%)	9/52 (17%)	0/50 (0%)	
Adjusted rate	0.0%	21.1%	0.0%	
Terminal rate	0/25 (0%)	4/21 (19%)	0/21 (0%)	
First incidence (days)	_ `	610	_ ``	
Poly-3 test		P=0.002	_	
Poly-3 test			P=0.003N	
Mammary Gland: Fibroadenoma				
Overall rate	33/53 (62%)	24/53 (45%)	32/50 (64%)	
Adjusted rate	71.0%	51.0%	71.8%	
Terminal rate	16/25 (64%)	12/21 (57%)	13/21 (62%)	
First incidence (days)	185	396	345	
Poly-3 test		P=0.031N	P=0.562	
Poly-3 test			P=0.027	
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TABLE	A3b
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### Statistical Analysis of Primary Neoplasms in Female Rats in the Stop-Exposure Evaluation of TCDD

	Vehicle Control	100 ng/kg	100 ng/kg (Stop-Exposure)	
Mammary Gland: Fibroadenoma or Adenoma				
Overall rate	35/53 (66%)	24/53 (45%)	32/50 (64%)	
Adjusted rate	74.7%	51.0%	71.8%	
Terminal rate	17/25 (68%)	12/21 (57%)	13/21 (62%)	
First incidence (days)	185	396	345	
Poly-3 test		P=0.010N	P=0.470N	
Poly-3 test			P=0.027	
Mammary Gland: Carcinoma				
Overall rate	4/53 (8%)	0/53 (0%)	1/50 (2%)	
Adjusted rate	9.9%	0.0%	2.6%	
Terminal rate	2/25 (8%)	0/21 (0%)	0/21 (0%)	
First incidence (days)	469	—	537	
Poly-3 test		P=0.053N	P=0.192N	
Poly-3 test			P=0.481	
Mammary Gland: Adenoma or Carcinoma				
Overall rate	6/53 (11%)	1/53 (2%)	1/50 (2%)	
Adjusted rate	14.7%	2.4%	2.6%	
Terminal rate	3/25 (12%)	1/21 (5%)	0/21 (0%)	
First incidence (days)	469	731 (T)	537	
Poly-3 test		P=0.048N	P=0.064N	
Poly-3 test			P=0.740	
Mammary Gland: Fibroadenoma, Adenoma, or Ca	rcinoma			
Overall rate	35/53 (66%)	24/53 (45%)	33/50 (66%)	
Adjusted rate	74.7%	51.0%	73.0%	
Terminal rate	17/25 (68%)	12/21 (57%)	13/21 (62%)	
First incidence (days)	185	396 D. 0.010M	345 D. 0 52501	
Poly-3 test		P=0.010N	P=0.525N	
Poly-3 test			P=0.019	
Oral Mucosa: Squamous Cell Carcinoma				
Overall rate	1/53 (2%)	10/53 (19%)	5/50 (10%)	
Adjusted rate	2.5%	22.0%	12.4%	
Terminal rate	0/25 (0%)	2/21 (10%)	0/21 (0%)	
First incidence (days)	366	546 D-0.007	520 P=0.000	
Poly-3 test		P=0.007	P=0.099 P=0.193N	
Pancreas: Adenoma or Carcinoma	0/51 (00/)	2/51 ((0))	1/10/20/2	
Overall rate	0/51 (0%)	3/51 (6%)	1/49 (2%)	
Adjusted rate	0.0%	/.3%	$\frac{2.1\%}{0}$	
First incidence (days)	0/23 (0%)	2/21 (10%)	1/21(5%)	
Poly_3 test		P=0.129	P=0.494	
Poly-3 test		1-0.129	P=0.345N	
Pituitam Cland (Dans Distalia on Unspecified Site).	Adanama			
Guarall rate		11/52 (210/)	10/50 (289/)	
Adjusted rote	25/33 (41%) 50 10/	11/32 (21%) 25.00/	19/30 (38%) A7 50/	
Aujusicu raie Terminal rate	20.4% 15/25 (60%)	23.0% 2/21 (10%)	47.5% 9/21 (43%)	
First incidence (days)	15/25 (00%) 474		584	
Poly-3 test	т,т	P<0.001N	P=0.213N	
Poly-3 test		1 0.00110	P=0.023	

### TABLE A3b

## Statistical Analysis of Primary Neoplasms in Female Rats in the Stop-Exposure Evaluation of TCDD

	Vehicle Control	100 ng/kg	100 ng/kg (Stop-Exposure)	
Thyroid Gland (C-cell): Adenoma				
Overall rate	20/52 (38%)	11/52 (21%)	13/49 (27%)	
Adjusted rate	46.7%	25.6%	33.7%	
Terminal rate	10/25 (40%)	6/21 (29%)	7/21 (33%)	
First incidence (days)	474	638	584	
Poly-3 test		P=0.030N	P=0.166N	
Poly-3 test			P=0.288	
Thyroid Gland (C-cell): Adenoma or Carcinoma				
Overall rate	21/52 (40%)	11/52 (21%)	14/49 (29%)	
Adjusted rate	49.0%	25.6%	36.3%	
Terminal rate	11/25 (44%)	6/21 (29%)	8/21 (38%)	
First incidence (days)	474	638	584	
Poly-3 test		P=0.017N	P=0.173N	
Poly-3 test			P=0.208	
Uterus: Squamous Cell Carcinoma				
Overall rate	0/53 (0%)	0/53 (0%)	2/50 (4%)	
Adjusted rate	0.0%	0.0%	5.2%	
Terminal rate	0/25 (0%)	0/21 (0%)	2/21 (10%)	
First incidence (days)	_ ` `	_ ` `	731 (T)	
Poly-3 test		_	P=0.232	
Poly-3 test			P=0.225	
Uterus: Stromal Polyp				
Overall rate	11/53 (21%)	8/53 (15%)	4/50 (8%)	
Adjusted rate	27.4%	18.2%	10.5%	
Terminal rate	8/25 (32%)	3/21 (14%)	3/21 (14%)	
First incidence (days)	645	598	729	
Poly-3 test		P=0.223N	P=0.050N	
Poly-3 test			P=0.259N	
All Organs: Benign Neoplasms				
Overall rate	45/53 (85%)	42/53 (79%)	41/50 (82%)	
Adjusted rate	92.2%	84.7%	89.4%	
Terminal rate	23/25 (92%)	18/21 (86%)	18/21 (86%)	
First incidence (days)	185	396	345	
Poly-3 test		P=0.173N	P=0.455N	
Poly-3 test			P=0.343	
All Organs: Malignant Neoplasms				
Overall rate	13/53 (25%)	34/53 (64%)	19/50 (38%)	
Adjusted rate	29.6%	69.9%	44.3%	
Terminal rate	4/25 (16%)	14/21 (67%)	9/21 (43%)	
First incidence (days)	169	546	345	
Poly-3 test		P<0.001	P=0.110	
Polv-3 test			P-0.008N	

#### TABLE A3b

Statistical Analysis of Primary Neoplasms in Female Rats in the Stop-Exposure Evaluation of TCDD

	Vehicle Control	100 ng/kg	100 ng/kg (Stop-Exposure)	
All Organs: Benign or Malignant Neoplasms				
Overall rate	47/53 (89%)	50/53 (94%)	46/50 (92%)	
Adjusted rate	92.7%	96.6%	94.5%	
Terminal rate	23/25 (92%)	20/21 (95%)	19/21 (91%)	
First incidence (days)	169	396	345	
Poly-3 test		P=0.317	P=0.522	
Poly-3 test			P=0.501N	

(T) Terminal sacrifice

<sup>a</sup> Number of neoplasm-bearing animals/number of animals examined. Denominator is number of animals examined microscopically for adrenal gland, heart, liver, lung, pancreas, pituitary gland, and thyroid gland; for other tissues, denominator is number of animals necropsied.

<sup>b</sup> Poly-3 estimated neoplasm incidence after adjustment for intercurrent mortality

<sup>c</sup> Observed incidence at terminal kill

<sup>a</sup> Beneath the dosed group incidence are the P values corresponding to pairwise comparisons between the vehicle controls and that dosed group. The Poly-3 test accounts for the differential mortality in animals that do not reach terminal sacrifice. A lower incidence in a dose group is indicated by N.

Pairwise comparison between the 100 ng/kg core and stop-exposure groups. A lower incidence in the stop-exposure group is indicated by N.

<sup>1</sup> Not applicable; no neoplasms in animal group

<sup>g</sup> Value of statistic cannot be computed.

#### TABLE A4a

Historical Incidence of Liver Neoplasms in Vehicle Control Female Sprague-Dawley Rats<sup>a</sup>

Study	Incidence in Controls					
	Cholangioma	Hepatocholangioma	Hepatocellular Adenoma	Cholangiocarcinoma		
Historical Incide	nce					
PCB 126	0/53	0/53	1/53	0/53		
PCB 126/PCB 118 Mixture	0/53	0/53	2/53	0/53		
PCB 126/PCB 153						
Binary Mixture	0/53	0/53	0/53	0/53		
PCB 153	0/53	0/53	0/53	0/53		
PeCDF	0/53	0/53	1/53	0/53		
TCDD	0/53	0/53	0/53	0/53		
TEF Mixture	0/53	0/53	0/53	0/53		
Overall Historica	l Incidence					
Total (%) Mean ± standard Range	0/371 deviation	0/371	$\begin{array}{c} 4/371 \ (1.1\%) \\ 1.1\% \pm 1.5\% \\ (0\%-4\%) \end{array}$	0/371		

<sup>a</sup> Data as of February 24, 2005

# TABLE A4bHistorical Incidence of Cystic Keratinizing Epithelioma in the Lungof Vehicle Control Female Sprague-Dawley Rats<sup>a</sup>

Study	Incidence in Controls
Historical Incidence	
PCB 126 PCB 126/PCB 118 Mixture PCB 126/PCB 153 Binary Mixture PCB 153 PeCDF TCDD	0/53 0/53 0/53 0/52 0/53 0/53
TEF Mixture Overall Historical Incidence Total	0/53 0/370

<sup>a</sup> Data as of February 24, 2005

Study	Incidence in Controls		
Historical Incidence			
PCB 126 PCB 126/PCB 118 Mixture PCB 126/PCB 153 Binary Mixture PCB 153 PeCDF TCDD TEF Mixture	0/53 1/53 0/53 0/53 1/53 1/53 1/53		
Overall Historical Incidence			
Total (%) Mean ± standard deviation Range	$\begin{array}{c} 4/371 \ (1.1\%) \\ 1.1\% \pm 1.0\% \\ 0\%-2\% \end{array}$		

# TABLE A4cHistorical Incidence of Squamous Cell Carcinoma in the Oral Mucosaof Vehicle Control Female Sprague-Dawley Rats<sup>a</sup>

<sup>a</sup> Data as of February 24, 2005

# TABLE A4dHistorical Incidence of Squamous Cell Carcinoma in the Uterusof Vehicle Control Female Sprague-Dawley Rats<sup>a</sup>

Study	Incidence in Controls
Historical Incidence	
PCB 126 PCB 126/PCB 118 Mixture PCB 126/PCB 153 Binary Mixture PCB 153 PeCDF TCDD TEF Mixture	0/53 0/53 1/53 0/53 0/53 0/53 0/53
Overall Historical Incidence	
Total (%) Mean ± standard deviation Range	$\begin{array}{c} 1/371 \ (0.3\%) \\ 0.3\% \pm 0.7\% \\ 0\% - 2\% \end{array}$

<sup>a</sup> Data as of February 24, 2005

## TABLE A4e

Historical Incidence of Pancreas Neoplasms in Vehicle Control Female Sprague-Dawley Rats<sup>a</sup>

Study	Adenoma	Carinoma	Adenoma or Carcinoma
Historical Incidence			
PCB 126	1/51	0/51	1/51
PCB 126/PCB 118 Mixture	0/53	0/53	0/53
PCB 126/PCB 153 Binary Mixture	0/53	0/53	0/53
PCB 153	0/53	0/53	0/53
PeCDF	0/53	0/53	0/53
TCDD	0/51	0/51	0/51
TEF Mixture	0/52	0/52	0/52
Overall Historical Incidence			
Total (%)	1/366 (0.3%)	0/366	1/366 (0.3%)
Mean $\pm$ standard deviation	$0.3\%\pm0.7\%$		$0.3\% \pm 0.7\%$
Range	0%-2%		0%-2%

<sup>a</sup> Data as of February 24, 2005

# Summary of the Incidence of Nonneoplastic Lesions in Female Rats at the 14-, 31-, and 53-Week Interim Evaluations in the 2-Year Gavage Study of TCDD<sup>a</sup>

	Vehicle Control	3 ng/kg	10 ng/kg	22 ng/kg	46 ng/kg	100 ng/kg
Disposition Summary						
Animals initially in study	28	28	28	28	28	28
14-Week interim evaluation	10	10	10	10	10	10
31-Week interim evaluation	10	10	10	10	10	10
53-Week interim evaluation	8	8	8	8	8	8
Animals examined microscopically	28	28	28	28	28	28
14-Week Interim Evaluatio	n					
Alimentary System						
Liver	(10)	(10)	(10)	(10)	(10)	(10)
Fatty change, diffuse						2 (20%)
Inflammation	10 (100%)	10 (100%)	8 (80%)	10 (100%)	10 (100%)	10 (100%)
Mixed cell focus		2 (20%)		1 (10%)	1 (10%)	2 (20%)
Pigmentation	1 (10%)					1 (10%)
Hepatocyte, hypertrophy	1 (10%)	1 (10%)	4 (40%)	7 (70%)	10 (100%)	10 (100%)
Hepatocyte, multinucleated						3 (30%)
Pancreas	(10)	(10)	(10)	(10)	(10)	(10)
Acinus, atrophy						2 (20%) 2 (20%)
Endocrine System						
Adrenal cortex	(10)	(10)	(10)	(10)	(10)	(10)
Hypertrophy	1 (10%)	2 (20%)	2 (20%)	1 (10%)	1 (10%)	
Thyroid gland	(10)	(10)	(10)	(10)	(10)	(10)
Follicular cell, hypertrophy		1 (10%)	3 (30%)	4 (40%)	4 (40%)	9 (90%)
Genital System						
Ovary	(10)					(10)
Atrophy	2 (20%)					1 (10%)
Uterus	(10)	(1)				(10)
Metaplasia, squamous	1 (10%)					4 (400/)
Endometrium, hyperplasia, cystic	5 (50%)					4 (40%)
Hematopoietic System						
Spleen	(10)					(10)
Pigmentation	10 (100%)					10 (100%)
Thymus	(10)	(10)	(10)	(10)	(10)	(10)
Atrophy		1 (10%)			4 (40%)	10 (100%)
Respiratory System						
Lung	(10)	(10)	(10)	(10)	(10)	(10)
Inflammation, chronic					1 (10%)	

<sup>a</sup> Number of animals examined microscopically at the site and the number of animals with lesion

Summary of the Incidence of Nonneoplastic Lesions in Female Rats at the 14-, 31-, and 53-Week Interim Evaluations in the 2-Year Gavage Study of TCDD

	Vehicle	Control	3 n	g/kg	10 r	ng/kg	22 I	ng/kg	46 1	ng/kg	100	ng/kg
Systems Examined at 14 Weeks Cardiovascular System General Body System Integumentary System Musculoskeletal System Nervous System Special Senses System Urinary System	with No Le	esions Ob	servea	l								
31-Week Interim Evaluation	on											
Alimentary System												
Liver	(10)		(10)		(10)		(10)		(10)		(10)	
Angiectasis			1	(10%)					1	(10%)		
Fatty change, diffuse											6	(60%)
Hepatodiaphragmatic nodule			1	(10%)								
Inflammation	9	(90%)	9	(90%)	10	(100%)	10	(100%)	10	(100%)	10	(100%)
Mitotic alteration	1	(10%)	2	(200/)	2	(200/)	2	(200/)	2	(200/)	2	(200/)
Mixed cell focus multiple	1	(10%)	3	(30%)	3	(30%)	Z	(20%)	5	(30%)	37	(30%)
Necrosis	2	(20%)	4	(40%)	1	(10%)			1	(10%)	/	(10%)
Pigmentation			1	(10%)	8	(80%)	9	(90%)	10	(100%)	10	(100%)
Toxic hepatopathy			-	()		(00,0)	-	(,,,,,)		()	10	(100%)
Bile duct, hyperplasia											1	(10%)
Hepatocyte, hypertrophy			2	(20%)	3	(30%)	6	(60%)	9	(90%)	10	(100%)
Hepatocyte, multinucleated									5	(50%)	9	(90%)
Oval cell, hyperplasia											1	(10%)
Pancreas	(10)	(2001)	(10)		(10)	(100())	(10)		(10)	(100/)	(10)	
Acinus, atrophy Acinus, vacuolization cytoplasmic	2	(20%)			1	(10%)			1	(10%)	5	(50%)
Endocrine System												
Adrenal cortex	(10)		(10)		(10)		(10)		(10)		(10)	
Degeneration, cystic							1	(10%)	1	(10%)		
Hyperplasia							1	(10%)				
Hypertrophy			2	(20%)	3	(30%)	3	(30%)	3	(30%)	3	(30%)
Necrosis	(10)		(10)		(10)		1	(10%)	(10)		(10)	
Thyroid gland	(10)		(10)	(200/)	(10)	(200/)	(10)	(200/)	(10)	(400/)	(10)	(600/)
Foncular cen, nypertrophy			3	(30%)	3	(30%)	3	(30%)	4	(40%)	0	(60%)
Genital System												
Ovary	(10)	(000/)									(10)	(700/)
Atrophy	(10)	(90%)	(1)								(10)	(70%)
Inflammation suppurative	(10)		(1)	(100%)							(10)	
Metaplasia, squamous	8	(80%)	1	(10070)							4	(40%)
Endometrium, hyperplasia, cystic	1	(10%)	1	(100%)							2	(20%)
Hematopoietic System												
Spleen	(10)										(10)	
Pigmentation	10	(100%)									10	(100%)

## Summary of the Incidence of Nonneoplastic Lesions at Female Rats in the 14-, 31-, and 53-Week Interim Evaluations in the 2-Year Gavage Study of TCDD

	Vehicle Control	3 ng/kg	10 ng/kg	22 ng/kg	46 ng/kg	100 ng/kg
Hematopoietic System (continued) Thymus Atrophy	(10) 2 (20%)	(10) 1 (10%)	(10) 5 (50%)	(10) 6 (60%)	(10) 7 (70%)	(10) 10 (100%)
Respiratory System Lung Infiltration cellular, histiocyte	(10) 2 (20%)	(10) 3 (30%)	(10) 1 (10%)	(10) 1 (10%)	(10)	(10) 1 (10%)

#### Systems Examined at 31 Weeks with No Lesions Observed

Cardiovascular System General Body System Integumentary System Musculoskeletal System Nervous System Special Senses System Urinary System

#### 53-Week Interim Evaluation

Alimentary System						
Liver	(8)	(8)	(8)	(8)	(8)	(8)
Basophilic focus			1 (13%)			2 (25%)
Basophilic focus, multiple		1 (13%)				1 (13%)
Cholangiofibrosis					1 (13%)	1 (13%)
Clear cell focus	2 (25%)					· · · ·
Fatty change, diffuse					2 (25%)	6 (75%)
Hepatodiaphragmatic nodule				1 (13%)		
Inflammation	8 (100%)	8 (100%)	8 (100%)	8 (100%)	8 (100%)	8 (100%)
Mixed cell focus	2 (25%)	1 (13%)		2 (25%)	1 (13%)	
Mixed cell focus, multiple		4 (50%)		4 (50%)	6 (75%)	8 (100%)
Pigmentation			5 (63%)	8 (100%)	8 (100%)	8 (100%)
Toxic hepatopathy			1 (13%)		6 (75%)	8 (100%)
Bile duct, cyst			1 (13%)			
Bile duct, fibrosis					1 (13%)	2 (25%)
Bile duct, hyperplasia			1 (13%)	2 (25%)	3 (38%)	8 (100%)
Hepatocyte, hypertrophy		4 (50%)	7 (88%)	8 (100%)	8 (100%)	8 (100%)
Hepatocyte, multinucleated					2 (25%)	8 (100%)
Pancreas	(8)	(8)	(8)	(8)	(8)	(8)
Inflammation, chronic active						2 (25%)
Acinus, atrophy						2 (25%)
Acinus, vacuolization cytoplasmic						7 (88%)
Endocrine System						
Adrenal cortex	(8)	(8)	(8)	(8)	(8)	(8)
Degeneration, cystic	1 (13%)	1 (13%)	1 (13%)	2 (25%)		1 (13%)
Hyperplasia	1 (13%)	~ /	~ /	~ /	3 (38%)	2 (25%)
Hypertrophy	5 (63%)	3 (38%)	4 (50%)	4 (50%)	5 (63%)	7 (88%)
Vacuolization cytoplasmic		1 (13%)				
Pituitary gland	(8)	( /	(1)			(7)
Cyst	1 (13%)		~ /			X-7

# Summary of the Incidence of Nonneoplastic Lesions in Female Rats at the 14-, 31-, and 53-Week Interim Evaluations in the 2-Year Gavage Study of TCDD

	Vehicle Control	3 ng/kg	10 ng/kg	22 ng/kg	46 ng/kg	100 ng/kg
Endocrine System (continued) Thyroid gland Follicular cell, hypertrophy	(8)	(8) 1 (13%)	(8) 2 (25%)	(8) 2 (25%)	(8) 3 (38%)	(8) 5 (63%)
Genital System Ovary Atrophy Uterus Metaplasia, squamous Cervix, cyst, squamous Endometrium, hyperplasia, cystic	(8) 8 (100%) (8) 7 (88%) 1 (13%) 5 (63%)			(1) 1 (100%)		(7) 5 (71%) (8) 4 (50%) 2 (25%)
Hematopoietic System Spleen Hyperplasia, focal, lymphoid Pigmentation Thymus Atrophy	(8) 1 (13%) 8 (100%) (8) 4 (50%)	(8) 2 (25%)	(7) 3 (43%)	(8) 7 (88%)	(8) 8 (100%)	(8) 8 (100%) (8) 8 (100%)
Integumentary System Mammary gland Cyst	(8) 3 (38%)	(2)	(2)			(8)
Respiratory System Lung Infiltration cellular, histiocyte	(8) 3 (38%)	(8) 1 (13%)	(8) 2 (25%)	(8) 1 (13%)	(8) 4 (50%)	(8) 3 (38%)
Urinary System Kidney Cyst Inflammation, chronic active Nephropathy Pelvis, dilatation Ureter Mineralization Transitional epithelium, hyperplasia Urinary bladder Inflammation Transitional epithelium, hyperplasia		(2) 1 (50%) 1 (50%) 1 (50%) (1) 1 (100%) 1 (100%) 1 (100%)				(1) 1 (100%) 1 (100%) (1) 1 (100%) 1 (100%)

Systems Examined at 53 Weeks with No Lesions Observed Cardiovascular System General Body System Musculoskeletal System Nervous System Special Senses System

Summary of the Incidence of Nonneoplastic Lesions in Female Rats in the 2-Year Gavage Study of TCDD<sup>a</sup>

	Vehicle Control	3 ng/kg	10 ng/kg	22 ng/kg	46 ng/kg	100 ng/kg
Disposition Summary						
Animals initially in study	53	54	53	53	53	53
Early deaths						
Accidental deaths			5			
Moribund	19	27	15	19	17	17
Natural deaths	9	6	10	15	14	15
Survivors						
Terminal sacrifice	25	21	23	19	22	21
Animals examined microscopically	53	54	53	53	53	53
Alimentary System						
Esophagus	(52)	(54)	(53)	(52)	(53)	(53)
Hemorrhage			1 (2%)			
Perforation			2 (4%)			
Muscularis, inflammation	1 (2%)	2 (4%)	1 (2%)	1 (2%)	2 (4%)	
Periesophageal tissue, inflammation			3 (6%)			
Intestine large, colon	(52)	(53)	(53)	(53)	(53)	(51)
Parasite metazoan	1 (2%)	2 (4%)	1 (2%)			2 (4%)
Intestine large, rectum	(52)	(54)	(53)	(52)	(53)	(53)
Parasite metazoan	6 (12%)	4 (7%)			2 (4%)	1 (2%)
Intestine small, duodenum	(53)	(54)	(53)	(52)	(53)	(53)
Cyst		1 (20/)				1 (2%)
Ulcer Enithalium humamlasia		1 (2%)	1 (20/)			
Liver	(53)	(54)	(53)	(53)	(53)	(53)
Angiectasis	(33) 2 (4%)	1 (2%)	(55)	(55)	3 (6%)	5 (0%)
Basophilic focus	8 (15%)	6 (11%)	8 (15%)	7 (13%)	3 (6%)	3 (6%)
Basophilic focus, multiple	4 (8%)	3 (6%)	5 (9%)	1 (2%)	2(4%)	5 (9%)
Cholangiofibrosis	1 (2%)	1 (2%)	2 (4%)	1 (2%)	11 (21%)	31 (58%)
Clear cell focus	4 (8%)	1 (2%)	1 (2%)	2 (4%)	1 (2%)	1 (2%)
Clear cell focus, multiple	1 (2%)			2 (4%)	1 (2%)	1 (2%)
Congestion	2 (4%)					2 (4%)
Cytoplasmic alteration						2 (4%)
Degeneration, cystic				2 (4%)		4 (8%)
Eosinophilic focus	8 (15%)	6 (11%)	7 (13%)	10 (19%)	5 (9%)	2 (4%)
Eosinophilic focus, multiple	3 (6%)	8 (15%)	14 (26%)	17 (32%)	22 (42%)	42 (79%)
Fatty change, diffuse	<b>a</b> (10.0	2 (4%)	12 (23%)	17 (32%)	30 (57%)	48 (91%)
Fatty change, focal	2 (4%)	6 (11%)	4 (8%)	4 (8%)	1 (2%)	2 (4%)
Hematopoletic cell proliferation	1 (2%)	2 (4%)	2 (4%)	4 (8%)	2 (4%)	2 (4%)
Hemorrhage		2(40/)		1 (20/)	1 (2%)	
Hyperplasia, nodular		2 (4%)		1 (2%) 3 (6%)	2(4%) 7(13%)	36 (68%)
Inflammation	33 (62%)	46 (85%)	47 (89%)	5(070)	52 (98%)	49 (02%)
Mitotic alteration	55 (0270)	40 (0570)	47 (0)70)	1 (2%)	1 (2%)	4) ()2/0)
Mixed cell focus	4 (8%)	8 (15%)	8 (15%)	1 (270)	2 (4%)	4 (8%)
Mixed cell focus, multiple	26 (49%)	11 (20%)	21 (40%)	23 (43%)	28 (53%)	17 (32%)
Necrosis	1 (2%)	4 (7%)	4 (8%)	8 (15%)	10 (19%)	17 (32%)
Pigmentation	4 (8%)	9 (17%)	34 (64%)	48 (91%)	52 (98%)	53 (100%)
Thrombosis	(···)	× · · · ·	(* · · ·)		<u> </u>	1 (2%)
Toxic hepatopathy		2 (4%)	8 (15%)	30 (57%)	45 (85%)	53 (100%)
Artery, inflammation, chronic active						2 (4%)

<sup>a</sup> Number of animals examined microscopically at the site and the number of animals with lesion

	Vehicle Control		hicle Control 3 ng/kg 10 ng/k		ng/kg	22 1	ng/kg	46 1	ng/kg	100	ng/kg	
Alimentary System (continued)												
Liver (continued)	(53)		(54)		(53)		(53)		(53)		(53)	
Bile duct, cyst	3 (	(6%)	1	(2%)	2	(4%)	2	(4%)	()		21	(40%)
Bile duct, fibrosis			2	(4%)			1	(2%)	3	(6%)	4	(8%)
Bile duct, hyperplasia	5 (	(9%)	4	(7%)	7	(13%)	22	(42%)	40	(75%)	53	(100%)
Centrilobular, degeneration	2 (	(4%)	2	(4%)			4	(8%)	3	(6%)	5	(9%)
Hepatocyte, hypertrophy			19	(35%)	19	(36%)	42	(79%)	41	(77%)	52	(98%)
Hepatocyte, multinucleated					16	(30%)	26	(49%)	36	(68%)	51	(96%)
Hepatocyte, centrilobular, atrophy									1	(2%)		
Oval cell, hyperplasia			4	(7%)	3	(6%)	20	(38%)	38	(72%)	53	(100%)
Portal, fibrosis									5	(9%)	27	(51%)
Mesentery	(2)		(2)				(1)		(6)		(8)	
Metaplasia, osseous	1 (	(50%)										
Artery, inflammation, chronic active			1	(50%)				(1000())	4	(67%)	7	(88%)
Fat, necrosis		(=00/)	1	(50%)			1	(100%)	1	(17%)	1	(13%)
Vein, thrombosis		(50%)	( <b>0</b> )		(10)		(10)		(22)		(20)	
Oral mucosa	(2)	(500/)	(9)	(700/)	(16)	(000/)	(16)	(010/)	(22)	((00/))	(29)	(550())
Gingival, hyperplasia, squamous	1 (	(50%)	(5.4)	(78%)	14	(88%)	(52)	(81%)	(52)	(68%)	10	(55%)
Pancreas	(51)		(54)		(52)		(53)		(52)		(51)	(20/)
Inflammation courts							1	(20/)			1	(270)
Inflammation, active					2	(10/2)	2	(270) (404)	3	(6%)	6	(12%)
Necrosis					2	(470)	1	(470)	5	(070)	1	(1270)
Acinus atrophy	1 (	(2%)	2	(4%)	4	(8%)	1	(270) (8%)	4	(8%)	9	(270)
Acinus, hyperplasia	2 (	(2%)	2	(470)	7	(070)	1	(2%)	- - 2	(3/0)	,	(10/0)
Acinus, vacuolization cytoplasmic	1 (	(2%)					1	(2%)	15	(29%)	42	(82%)
Artery, inflammation, chronic active		(=/0)	1	(2%)	1	(2%)	2	(4%)	2	(4%)	.=	(14%)
Salivary glands	(51)		(54)		(52)	()	(50)		(51)		(52)	(
Atrophy	× ,		· · ·		· · ·		2	(4%)	1	(2%)	· · /	
Inflammation					2	(4%)	3	(6%)			1	(2%)
Mineralization	1 (	(2%)										
Stomach, forestomach	(53)		(54)		(53)		(53)		(53)		(53)	
Cyst, squamous					1	(2%)						
Edema							1	(2%)			1	(2%)
Erosion					1	(2%)			1	(2%)		
Hyperkeratosis	1 (	(2%)	1	(2%)	1	(2%)	1	(2%)	2	(4%)	3	(6%)
Hyperplasia, squamous	3 (	(6%)	4	(7%)	4	(8%)	2	(4%)	7	(13%)	11	(21%)
Inflammation	1 (	(2%)	1	(2%)	2	(4%)			2	(4%)	5	(9%)
Mineralization					1	(2%)						
Necrosis		(10)		(20)				(20)		(20)	1	(2%)
Ulcer	2 (	(4%)	1	(2%)	(52)		(52)	(2%)	(52)	(2%)	4	(8%)
Stomach, glandular	(53)		(54)		(53)	(20/)	(53)		(53)		(53)	
Diverticulum	1 (	(20/)			1	(2%)						
Erosion Motoplacia	1 (	(2%)							1	(20%)		
Mineralization	2 (	(60/)					1	(20/)	1	(270)		
	5 (	(0%)					1	(270)	3	(0%)	2	(10/2)
Tooth	(16)		(16)		(12)		(13)		(14)		(18)	(470)
Peridontal tissue fibrosis	(10)	(6%)	(10)		(12)		(15)		(14)		(10)	
Peridontal tissue, inflammation	15 (	(94%)	16	(100%)	12	(100%)	13	(100%)	14	(100%)	18	(100%)
Cardiovascular System												
Blood vessel	(53)		(54)		(53)		(53)		(53)		(53)	
Thrombosis	(55)		(0))		(33)	(2%)	(33)		(55)		(33)	
Aorta, mineralization	1 (	(2%)	1	(2%)	-	~ /			1	(2%)		

TABLE	A5b
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Summary of the Incidence of Nonneoplastic Lesions in Female Rats in the 2-Year Gavage Study of TCDD

	Vehicle	Control	3 n	g/kg	10 r	ng/kg	22 1	ng/kg	46 1	ng/kg	100	ng/kg
Cardiovascular System (continued) Heart Cardiomyopathy Mineralization	(53) 10 1	(19%) (2%)	(54) 12	(22%)	(53) 22	(42%)	(52) 25	(48%)	(53) 32 1	(60%) (2%)	(52) 36	(69%)
Artery, inflammation Artery, mineralization Artery, thrombosis			1	(2%)							1	(2%) (2%)
Endocrine System												
Adrenal cortex	(53)		(54)		(53)		(53)		(53)		(53)	
Angiectasis	11	(21%)	21	(39%)	18	(34%)	17	(32%)	17	(32%)	11	(21%)
Atrophy	2	(4%)			4	(8%)	5	(9%)	5	(9%)	27	(51%)
Degeneration, cystic	11	(21%)	15	(28%)	21	(40%)	18	(34%)	17	(32%)	17	(32%)
Hyperplasia	16	(30%)	16	(30%)	18	(34%)	25	(47%)	29	(55%)	30	(57%)
Hypertrophy	41	(77%)	43	(80%)	46	(87%)	40	(75%)	45	(85%)	47	(89%)
Inflammation, chronic active Inflammation, suppurative Minoralization	1 1	(2%) (2%)									1	(29/)
Necrosis			1	(2%)	2	(40/2)	1	(2%)	3	(6%)	1	(270)
Thrombosis			1	(270)	2	(470)	1	(2%)	5	(070)	1	(270)
Vacuolization extonlasmic	11	(21%)	7	(13%)	12	(23%)	21	(270)	18	(34%)	15	(28%)
Adrenal medulla	(53)	(2170)	(54)	(1570)	(53)	(2370)	(53)	(1070)	(53)	(31/0)	(53)	(2070)
Angiectasis	(00)		1	(2%)	2	(4%)	(00)	(2%)	(00)		(00)	
Hyperplasia	19	(36%)	10	(19%)	18	(34%)	7	(13%)	10	(19%)	9	(17%)
Inflammation, suppurative		()		( )		()	1	(2%)		( )		( )
Islets, pancreatic	(51)		(54)		(53)		(53)		(52)		(52)	
Hyperplasia	· · ·		· · /		1	(2%)	1	(2%)	· · ·			
Parathyroid gland	(46)		(47)		(47)		(44)		(45)		(45)	
Fibrosis					1	(2%)						
Hyperplasia	1	(2%)										
Pituitary gland	(53)		(54)		(52)		(53)		(53)		(52)	
Angiectasis	25	(47%)	18	(33%)	25	(48%)	20	(38%)	21	(40%)	9	(17%)
Atypia cellular	1	(2%)										
Cyst	1	(2%)	1	(2%)							1	(2%)
Cytoplasmic alteration	1	(2%)			1	(2%)	4	(8%)	3	(6%)		
Developmental malformation	1	(2%)										
Inflammation, chronic											1	(2%)
Vacuolization cytoplasmic	10	(2(0))	3	(6%)	l	(2%)	2	(4%)	2	(4%)	10	(270/)
Pars distalis, hyperplasia	19	(36%)	19	(35%)	16	(31%)	21	(40%)	26	(49%)	19	(37%)
Pars intermedia, hyperplasia	(52)	(4%)	(5.4)		(52)		(51)		(52)	(2%)	(52)	(2%)
Angiostosis	(52)		(54)	(20/)	(53)	(20/)	(51)	(20/)	(55)	(80/)	(52)	(20/)
Augreetasis	1	(20/)	1	(270)	1	(270)	1	(270)	4	(070)	1	(270)
Inflammation chronic active	1	(270)			1	(2%)	1	(270)			1	(270)
Inflammation, suppurative					1	(2%)						
C-Cell hyperplasia	19	(37%)	17	(31%)	22	(42%)	19	(37%)	16	(30%)	23	(44%)
Follicular cell, hyperplasia	2	(4%)	2	(4%)		(12/0)	1)	(3770)	10	(3070)	23	(
Follicular cell, hyperfronhy	3	(6%)	4	(7%)	4	(8%)	7	(14%)	10	(19%)	17	(33%)
	5	(***)				()	,	()	10	()	- /	()

**General Body System** 

None

	Vehicle Control		ol 3 ng/kg		10 1	ng/kg	22 ng/kg		/kg 46 ng/kg		100 ng/kg	
Genital System												
Clitoral gland	(50)		(52)		(53)		(52)		(51)		(53)	
Hyperplasia, basal cell	1	(2%)	(- )		1	(2%)	(- )		(- )		()	
Inflammation	41	(82%)	40	(77%)	35	(66%)	34	(65%)	28	(55%)	26	(49%)
Duct, cyst	34	(68%)	37	(71%)	41	(77%)	42	(81%)	41	(80%)	48	(91%)
Ovary	(51)		(53)		(53)		(53)		(53)		(53)	
Atrophy	49	(96%)	46	(87%)	50	(94%)	44	(83%)	50	(94%)	31	(58%)
Congestion							1	(2%)				
Cyst	14	(27%)	16	(30%)	13	(25%)	15	(28%)	14	(26%)	13	(25%)
Fibrosis					1	(2%)			1	(2%)		
Inflammation, chronic active							1	(2%)			1	(2%)
Inflammation, suppurative							1	(2%)	1	(2%)		
Artery, inflammation, chronic active											1	(2%)
Corpus luteum, cyst			1	(2%)								
Periovarian tissue, inflammation, suppura	tive						1	(2%)				
Oviduct	(2)		(1)		(2)				(1)		(1)	
Inflammation	2	(100%)	1	(100%)	1	(50%)					1	(100%)
Necrosis						(500/)			1	(100%)		
Epithelium, hyperplasia	(52)		(52)		(52)	(50%)	(52)		(52)		(52)	
Uterus	(52)		(53)		(53)		(53)		(53)	(20/)	(53)	
Adenomyosis									1	(2%)	1	(20/)
Cyst	1	(20/)									1	(2%)
Inflammation abronia activa	1	(270)	2	(40/)	2	(60/)			2	(60/.)	1	(2%)
Inflammation, curputative	7	(130/2)	2	(470) (1104)	12	(0%) (23%)	0	(170/)	11	(070) (2104)	1	(2%)
Metaplasia squamous	20	(1570)	31	(1170)	28	(2370)	23	(1770) (43%)	32	(2170)	17	(37%)
Necrosis	29	(3070)	51	(3870)	28	(3370)	23	(73)	52	(0070)	17	(3270)
Iller					1	(2%)	1	(270)				
Cervix hyperplasia stromal			1	(2%)	1	(2%)	1	(2%)	1	(2%)		
Cervix inflammation suppurative			1	(270)	1	(270)	1	(270)	1	(2%)		
Endometrium, adenomyosis					1	(2%)			-	(_, ))		
Endometrium, hyperplasia, cystic	30	(58%)	33	(62%)	35	(66%)	27	(51%)	31	(58%)	19	(36%)
Epithelium, hyperplasia		()		()		()	1	(2%)		()		()
Serosa, inflammation, chronic active					1	(2%)						
Vagina			(1)		(1)		(3)		(3)		(1)	
Cyst					1	(100%)			1	(33%)		
Hyperplasia, stromal			1	(100%)								
Inflammation							2	(67%)				
Necrosis							1	(33%)				
Hematopoietic System												
Bone marrow	(53)		(54)		(53)		(53)		(53)		(53)	
Atrophy											1	(2%)
Hyperplasia	36	(68%)	41	(76%)	32	(60%)	35	(66%)	37	(70%)	43	(81%)
Lymph node	(2)		(6)		(3)		(5)		(6)		(9)	
Deep cervical, hemorrhage							1	(20%)				
Inguinal, ectasia									1	(17%)		
Lumbar, ectasia	1	(50%)	2	(33%)	1	(33%)	2	(40%)				
Lumbar, hemorrhage							1	(20%)				
Lumbar, hyperplasia, plasma cell			2	(33%)			1	(20%)				
Lumbar, pigmentation							1	(20%)				
Mediastinal, congestion									1	(17%)		
Mediastinal, ectasia				(1 = 0 ))				(2001)	1	(17%)	3	(33%)
Mediastinal, hemorrhage			1	(17%)			1	(20%)		(1 = 0 ) )	5	(56%)
Mediastinal, hyperplasia						(220.()			1	(17%)		(110.)
Mediastinal, hyperplasia, histiocytic					1	(33%)			1	(17%)	1	(11%)

		Vehicle	Control	3 n	g/kg	10 1	ng/kg	22 ו	ng/kg	46 1	ng/kg	100	ng/kg
$ \begin{array}{llllllllllllllllllllllllllllllllllll$	Hematopoietic System (continued)												
	Bone marrow (continued)	(53)		(54)		(53)		(53)		(53)		(53)	
	Mediastinal, hyperplasia, lymphoid			1	(17%)								
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Mediastinal, hyperplasia, plasma cell					1	(220/)	1	(20%)			1	(110/)
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	Mediastinal, pigmentation			1	(170/)	1	(33%)					1	(11%)
	Renal hyperplasia histiocytic			1	(17%)								
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Lymph node, mandibular	(51)		(54)	(1770)	(52)		(50)		(51)		(52)	
	Congestion	1	(2%)	(- )		(- )		()		(- )		(- )	
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Ectasia									1	(2%)	6	(12%)
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	Hyperplasia, lymphoid			3	(6%)	3	(6%)					2	(4%)
	Hyperplasia, plasma cell	25	(49%)	31	(57%)	23	(44%)	22	(44%)	15	(29%)	16	(31%)
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	Inflammation, suppurative	(50)		(50)		(50)		(50)		(50)		1	(2%)
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Lymph node, mesenteric	(52)		(53)	(20/)	(53)		(53)	(20/)	(53)		(51)	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	Atrophy			1	(2%)			1	(2%)			1	(20%)
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	Hemorrhage									1	(2%)	1	(270)
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Hyperplasia histiocytic	1	(2%)							1	(2%)	2	(4%)
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Hyperplasia, plasma cell	-	(2/0)							1	(2%)	-	(1/0)
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Spleen	(51)		(54)		(53)		(53)		(52)		(52)	
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	Hematopoietic cell proliferation	46	(90%)	50	(93%)	38	(72%)	42	(79%)	44	(85%)	43	(83%)
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	Hyperplasia, lymphoid	1	(2%)									4	(8%)
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	Necrosis					1	(2%)						
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	Pigmentation	45	(88%)	49	(91%)	49	(92%)	51	(96%)	49	(94%)	47	(90%)
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	Lymphoid follicle, atrophy	4	(8%)	(50)		1	(2%)	(10)		1	(2%)	1	(2%)
Autophy       35 (11/a)       41 (97/a)       41 (97/a)	1 hymus A trophy	(51)	(710/)	(52)	(700/)	(52)	(850/)	(49)	(840/)	(46)	(069/)	(42)	(100%)
Cyst       1 (2%)       1 (2%)       1 (2%)       1 (2%)         Inflammation, chronic active       1 (2%)       1 (2%)       1 (2%)         Integumentary System       1 (2%)       1 (2%)       1 (2%)         Mammary gland       (53)       (54)       (53)       (53)       (53)         Cyst       4 (8%)       3 (6%)       1 (2%)       1 (2%)       1 (2%)         Hyperplasia       25 (47%)       21 (39%)       24 (45%)       22 (42%)       18 (34%)       16 (30%)         Inflammation, chronic active       1 (2%)       1 (2%)       1 (2%)       1 (2%)       1 (2%)         Inflammation, granulomatous       1 (2%)       1 (2%)       1 (2%)       1 (2%)       1 (2%)         Skin       (53)       (53)       (53)       (53)       (53)       (53)       (53)         Cyst epithelial inclusion       1 (2%)       1 (2%)       1 (2%)       1 (2%)       1 (2%)         Hemorrhage       1 (2%)       1 (2%)       1 (2%)       1 (2%)       1 (2%)         Ulcer       1 (2%)       1 (2%)       1 (2%)       1 (2%)       1 (2%)         Bone       (53)       (54)       (53)       (53)       (53)       (53)       1 (2%)	Cyst	50	(71%)	41	(79%)	44	(83%)	41	(84%)	44	(90%)	42	(100%)
Inflammation, chronic active       1 (2%)         Inflammation, chronic active       1 (2%)         Integumentary System       1 (2%)         Mammary gland       (53)       (54)       (53)       (53)       (53)         Cyst       4 (8%)       3 (6%)       1 (2%)       1 (2%)       1 (2%)         Hyperplasia       25 (47%)       21 (39%)       24 (45%)       22 (42%)       18 (34%)       16 (30%)         Inflammation, chronic active       1 (2%)       1 (2%)       1 (2%)       1 (2%)       1 (2%)         Inflammation, granulomatous       1 (2%)       1 (2%)       1 (2%)       1 (2%)         Skin       (53)       (53)       (53)       (53)       (53)       (53)       (53)         Hemorrhage       1 (2%)       1 (2%)       1 (2%)       1 (2%)       1 (2%)       1 (2%)         Hemorrhage       1 (2%)       1 (2%)       1 (2%)       1 (2%)       1 (2%)         Ulcer       2 (4%)       1 (2%)       1 (2%)       1 (2%)       1 (2%)         Musculoskeletal System       1       1 (2%)       1 (2%)       1 (2%)       1 (2%)         Skeletal muscle       (1)       (2)       1 (2%)       1 (2%)       1 (2%)	Hemorrhage	1	(2%)			1	(2%)			1	(270)	1	(270)
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	Inflammation, chronic active	1	(270)			1	(270)			1	(2%)		
Marmary gland       (53)       (54)       (53)       (53)       (53)       (53)       (53)         Cyst       4 (8%)       3 (6%)       1 (2%)       1 (2%)       1 (2%)       1 (2%)         Hyperplasia       25 (47%)       21 (39%)       24 (45%)       22 (42%)       18 (34%)       16 (30%)         Inflammation, dronic active       1 (2%)       1 (2%)       1 (2%)       1 (2%)         Skin       (53)       (53)       (53)       (53)       (53)       (53)         Cyst epithelial inclusion       1 (2%)       1 (2%)       2 (4%)       2 (4%)       2 (4%)         Hemorrhage       1 (2%)       1 (2%)       1 (2%)       1 (2%)       1 (2%)       1 (2%)         Inflammation, chronic active       2 (4%)       1 (2%)       1 (2%)       1 (2%)       1 (2%)         Inflammation       1 (2%)       1 (2%)       1 (2%)       1 (2%)       1 (2%)       1 (2%)         Musculoskeletal System       1 (2%)       1 (2%)       1 (2%)       1 (2%)       1 (2%)         Skeletal muscle       (1)       (2)       1 (2%)       1 (2%)       1 (2%)         Ketter       1 (2%)       1 (2%)       1 (2%)       1 (2%)       1 (2%)	Integumentary System												
Multicity guild       (b)       (c)	Mammary gland	(53)		(54)		(53)		(53)		(53)		(53)	
Hyperplasia       25 (47%)       21 (39%)       24 (45%)       22 (42%)       18 (34%)       16 (30%)         Inflammation, chronic active       1 (2%)       1 (2%)       1 (2%)       1 (2%)         Inflammation, chronic active       1 (2%)       1 (2%)       1 (2%)       1 (2%)         Skin       (53)       (54)       (53)       (53)       (53)       (53)       (53)         Cyst epithelial inclusion       1 (2%)       1 (2%)       1 (2%)       1 (2%)       1 (2%)         Hyperplasia, squamous       1 (2%)       1 (2%)       1 (2%)       1 (2%)         Inflammation, chronic active       2 (4%)       1 (2%)       1 (2%)         Hyperplasia, squamous       1 (2%)       1 (2%)       1 (2%)         Inflammation, chronic active       2 (4%)       1 (2%)       1 (2%)         Ulcer       2 (4%)       1 (2%)       1 (2%)         Epidermis, inflammation       1 (2%)       1 (2%)       1 (2%)         Masculoskeletal System       1 (2%)       1 (2%)       1 (2%)         Skeletal muscle       (1) (2)       1 (2%)       1 (2%)         Hemorrhage       1 (50%)       1 (50%)       1 (50%)	Cyst	4	(8%)	3	(6%)	(00)		1	(2%)	1	(2%)	1	(2%)
Inflammation, chronic active       1 (2%)       1 (2%)       1 (2%)         Inflammation, granulomatous       1 (2%)       1 (2%)       1 (2%)         Skin       (53)       (54)       (53)       (53)       (53)         Cyst epithelial inclusion       1 (2%)       2 (4%)       2 (4%)       2 (4%)         Hemorrhage       1 (2%)       1 (2%)       1 (2%)       1 (2%)         Hyperplasia, squamous       1 (2%)       1 (2%)       1 (2%)       1 (2%)         Inflammation, chronic active       2 (4%)       1 (2%)       1 (2%)         Necrosis       1 (2%)       1 (2%)       1 (2%)         Ulcer       2 (4%)       1 (2%)       1 (2%)         Epidermis, inflammation       1 (2%)       1 (2%)       1 (2%)         Musculoskeletal System       1 (2%)       1 (2%)       1 (2%)         Bone       (53)       (54)       (53)       (53)       (53)       (53)         Fracture       1 (2%)       1 (2%)       1 (2%)       1 (2%)       1 (2%)         Skeletal muscle       1 (2%)       1 (2%)       1 (2%)       1 (2%)       1 (2%)         Hemorrhage       1 (50%)       1 (50%)       1 (50%)       1 (50%)       1 (50%)	Hyperplasia	25	(47%)	21	(39%)	24	(45%)	22	(42%)	18	(34%)	16	(30%)
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	Inflammation, chronic active									1	(2%)		
Skin $(53)$ $(54)$ $(53)$ $(53)$ $(53)$ $(53)$ $(53)$ Cyst epithelial inclusion       1 $(2\%)$ 1 $(2\%)$ 2 $(4\%)$ Hemorrhage       1 $(2\%)$ 1 $(2\%)$ 1 $(2\%)$ Inflammation, chronic active       2 $(4\%)$ 1 $(2\%)$ 1 $(2\%)$ Ulcer       2 $(4\%)$ 1 $(2\%)$ 1 $(2\%)$ 1 $(2\%)$ Musculoskeletal System       8       8       8       8       1 $(2\%)$ 1 $(2\%)$ Fracture       (53)       (54)       (53)       (53) $(53)$ <	Inflammation, granulomatous	1	(2%)	1	(2%)	1	(2%)					1	(2%)
Cyst epithelial inclusion       1 (2%)       2 (4%)         Hemorrhage       1 (2%)       1 (2%)         Hyperplasia, squamous       1 (2%)       1 (2%)         Inflammation, chronic active       2 (4%)       1 (2%)         Necrosis       1 (2%)       1 (2%)         Ulcer       1 (2%)       1 (2%)         Epidermis, inflammation       1 (2%)       1 (2%)         Musculoskeletal System       1 (2%)       1 (2%)         Bone       (53)       (54)       (53)       (53)       (53)         Fracture       1 (2%)       1 (2%)       1 (2%)         Skeletal muscle       (1)       (2)       1 (2%)         Hemorrhage       1 (50%)       1 (50%)       1 (50%)	Skin	(53)		(54)		(53)		(53)		(53)		(53)	
Hemorrhage       1 (2%)         Hyperplasia, squamous       1 (2%)         Inflammation, chronic active       2 (4%)         Necrosis       1 (2%)         Ulcer       1 (2%)         Epidermis, inflammation       1 (2%)         Musculoskeletal System         Bone       (53)         Fracture       1 (2%)         Maxilla, inflammation, focal, suppurative       1 (2%)         Skeletal muscle       1 (2%)         Hemorrhage       1 (50%)         Inflammation, chronic active       1 (50%)	Cyst epithelial inclusion	1	(2%)				(20)					2	(4%)
Hyperplasia, squamous       1 (2%)       1 (2%)         Inflammation, chronic active       2 (4%)       1 (2%)         Necrosis       1 (2%)       1 (2%)         Ulcer       1 (2%)       1 (2%)         Epidermis, inflammation       1 (2%)       1 (2%)         Musculoskeletal System       1 (2%)       1 (2%)         Bone       (53)       (54)       (53)       (53)       (53)         Fracture       1 (2%)       1 (2%)       1 (2%)         Skeletal muscle       (1)       (2)       1 (2%)         Hemorrhage       1 (50%)       1 (50%)       1 (50%)	Hemorrhage					1	(2%)			1	(20/)		
Initialimation, chronic active     1 (2%)     1 (2%)       Necrosis     1 (2%)     1 (2%)       Ulcer     1 (2%)     1 (2%)       Epidermis, inflammation     1 (2%)       Musculoskeletal System     1 (2%)       Bone     (53)     (54)       Fracture     1 (2%)       Maxilla, inflammation, focal, suppurative     1 (2%)       Skeletal muscle     (1)     (2)       Hemorrhage     1 (50%)       Inflammation, chronic active     1 (50%)	Hyperplasia, squamous					1	(2%)			1	(2%)	1	(20/)
Interiors       1 (2%)         Ulcer       1 (2%)         Epidermis, inflammation       1 (2%)         Musculoskeletal System       1 (2%)         Bone       (53)       (54)       (53)       (53)       (53)         Fracture       1 (2%)         Maxilla, inflammation, focal, suppurative       1 (2%)         Skeletal muscle       (1)       (2)         Hemorrhage       1 (50%)         Inflammation, chronic active       1 (50%)	Necrosis					2	(4%)					1	(270)
Epidermis, inflammation       1 (2%)         Musculoskeletal System         Bone       (53)       (54)       (53)       (53)       (53)         Fracture       1 (2%)         Maxilla, inflammation, focal, suppurative       1 (2%)         Skeletal muscle       (1)       (2)         Hemorrhage       1 (50%)         Inflammation, chronic active       1 (50%)	Illeer					1	(2%)			1	(2%)		
Musculoskeletal System           Bone         (53)         (54)         (53)         (53)         (53)           Fracture         1         1         (2%)           Maxilla, inflammation, focal, suppurative         1         (2%)           Skeletal muscle         (1)         (2)           Hemorrhage         1         (50%)           Inflammation, chronic active         1         (50%)	Epidermis, inflammation					1	(270)			1	(270)	1	(2%)
Bone       (53)       (54)       (53)       (53)       (53)       (53)         Fracture       1       (2%)       1       (2%)         Maxilla, inflammation, focal, suppurative       1       (2)       1       (2%)         Skeletal muscle       (1)       (2)       1       (50%)         Inflammation, chronic active       1       (50%)       1       (50%)	Musculoskeletal System												
Fracture(60) <t< td=""><td>Bone</td><td>(53)</td><td></td><td>(54)</td><td></td><td>(53)</td><td></td><td>(53)</td><td></td><td>(53)</td><td></td><td>(53)</td><td></td></t<>	Bone	(53)		(54)		(53)		(53)		(53)		(53)	
Maxilla, inflammation, focal, suppurative1 (2%)Skeletal muscle(1)Hemorrhage1 (50%)Inflammation, chronic active1 (50%)	Fracture	(55)		(54)		(55)		(55)		(55)		1	(2%)
Skeletal muscle(1)(2)Hemorrhage1(50%)Inflammation, chronic active1(50%)	Maxilla, inflammation, focal, suppurative			1	(2%)								
Hemorrhage1 (50%)Inflammation, chronic active1 (50%)	Skeletal muscle					(1)		(2)					
Inflammation, chronic active 1 (50%)	Hemorrhage							1	(50%)				
	Inflammation, chronic active							1	(50%)				

	Vehicle Control		l 3 ng/kg		10 1	ng/kg	22 ng/kg		kg 46 ng/kg		100 ng/kg	
Nervous System												
Brain	(53)		(54)		(53)		(53)		(53)		(53)	
Hemorrhage					1	(2%)						
Hydrocephalus			1	(2%)	1	(2%)						
Inflammation, suppurative									1	(2%)		
Mineralization			1	(2%)	1	(2%)			2	(4%)	2	(4%)
Vacuolization cytoplasmic, focal					1	(2%)						
Artery, thrombosis									1	(2%)		
Cerebellum, developmental malformation	1	(2%)										
Cerebellum, necrosis					1	(2%)						
Glial cell, hyperplasia			1	(2%)	(4)						1	(2%)
Spinal cord					(1)	(1000())						
Gliosis					1	(100%)						
Respiratory System												
Lung	(53)		(54)		(53)		(52)		(53)		(52)	
Congestion	()		(- )		2	(4%)	(- )		()		(- )	
Edema					2	(4%)			1	(2%)		
Hemorrhage			1	(2%)							2	(4%)
Infiltration cellular, histiocyte	41	(77%)	46	(85%)	43	(81%)	43	(83%)	49	(92%)	50	(96%)
Inflammation, chronic active											1	(2%)
Inflammation, granulomatous	2	(4%)	1	(2%)	2	(4%)	2	(4%)				
Inflammation, suppurative	1	(2%)					2	(4%)				
Metaplasia, squamous	1	(2%)							5	(9%)	6	(12%)
Mineralization	1	(2%)							1	(2%)	1	(2%)
Necrosis							1	(2%)			1	(2%)
Pigmentation			1	(2%)								
Alveolar epithelium, hyperplasia	12	(23%)										
Alveolar epithelium, metaplasia, bronchiolar	2	(4%)	19	(35%)	33	(62%)	35	(67%)	45	(85%)	46	(88%)
Bronchiole, dilatation					1	(2%)						
Serosa, inflammation, focal, suppurative							1	(2%)				
Nose	(53)		(54)		(53)		(53)		(53)		(53)	
Cyst, squamous		(10)						((0))	1	(2%)		(00)
Inflammation	2	(4%)					3	(6%)	5	(9%)	4	(8%)
Goblet cell, septum, hyperplasia		(10)							1	(2%)		
Nasolacrimal duct, inflammation	2	(4%)										
Nasolacrimal duct, respiratory epithelium,									1	(20/)		
Negenhammagal dust inflammation							1	(20/)	1	(2%)	1	(20/)
Nasopharyngeal duct, mhammation							1	(270)			1	(270)
Nasopharyngeal duct, necrosis							1	(2%)	1	(20/)		
Olfactory epithelium, degeneration									1	(270)	1	(20%)
Begniratory epithelium, anat									1	(270)	1	(270)
Respiratory epithelium, hyperplasia							1	(2%)	2	(4%)		
Septum hyperplasia squamous							1	(270)	2	(470)	1	(2%)
Septum, hyperplasia, squamous											2	(270)
Squamous enithelium inflammation											2	(1/0)
suppurative	1	(2%)										
Turbinate, hyperplasia, squamous	1	(2/0)									1	(2%)
Turbinate, septum, inflammation					1	(2%)	1	(2%)			2	(4%)
Turbinate, respiratory epithelium.					1	(=)	1	(=, -)			-	(
hyperplasia					1	(2%)	1	(2%)				
~ 1 I					-	` '	-	. /				

	Vehicle	Control	3 n	g/kg	10 r	ng/kg	22 ו	ng/kg	46 1	ng/kg	100	ng/kg
Special Senses System												
Eve	(53)		(54)		(53)		(52)		(53)		(53)	
Degeneration	1	(2%)	(- )		()		(- )		()		()	
Hemorrhage	1	(2%)										
Anterior chamber, inflammation, suppura	ative	(_, , ,									1	(2%)
Cornea, inflammation, suppurative					1	(2%)					1	(2%)
Lens, degeneration						(= / •)			1	(2%)		(_,,,)
Retina, atrophy	5	(9%)	5	(9%)	5	(9%)	3	(6%)	4	(8%)	6	(11%)
Retina, degeneration		(,,,,)		(,,,,)		(,,,,)		(0,0)	1	(2%)		()
Harderian gland	(53)		(54)		(53)		(52)		(53)	(= / •)	(53)	
Atrophy, focal	1	(2%)	(4.1)		()		(==)		()		()	
Inflammation	12	(23%)	3	(6%)	2	(4%)	2	(4%)	8	(15%)	6	(11%)
Urinary System												
Kidney	(53)		(54)		(53)		(53)		(53)		(53)	
Calculus gross observation	(55)		(31)		(55)		(55)		(55)		(33)	(2%)
Calculus microscopic observation only	1	(2%)	1	(2%)	1	(2%)	3	(6%)	2	(4%)	1	(270)
Casts protein	3	(6%)	5	(9%)	2	(4%)	5	(0,0)	- 1	(2%)	1	(2%)
Cvst	2	(4%)	-	(,,,,)	_	(1,0)			1	(2%)	1	(2%)
Fibrosis		(1))	1	(2%)						(= / •)		(_,,,)
Infarct			-	(=, •)	1	(2%)						
Inflammation, chronic active	5	(9%)	1	(2%)		(=,0)	1	(2%)	1	(2%)		
Inflammation suppurative		(,,,,)		(=, •)	2	(4%)	1	(2%)	1	(2%)	2	(4%)
Mineralization	40	(75%)	39	(72%)	30	(57%)	32	(60%)	42	(79%)	42	(79%)
Necrosis		((2,3))		(,=,,)		(2 / / 0)	1	(2%)	1	(2%)		(,,,,,)
Nephropathy	34	(64%)	26	(48%)	32	(60%)	36	(68%)	39	(74%)	52	(98%)
Papilla, necrosis	5.	(01/0)	20	(10/0)	1	(2%)	20	(00/0)	0,7	(, . , 0)		(,,,,,)
Pelvis dilatation	2	(4%)	1	(2%)	1	(2%)			2	(4%)		
Pelvis, inflammation	2	(4%)	1	(2%)	1	(2%)	1	(2%)	4	(8%)	2	(4%)
Renal tubule, degeneration		(1,0)		(=, •)		(=, •)		(_/*)		(0,0)	1	(2%)
Transitional epithelium hyperplasia	3	(6%)	6	(11%)	8	(15%)	8	(15%)	11	(21%)	11	(21%)
Ureter	(1)	(0,0)	Ũ	(11/0)	0	(10,0)	Ũ	(10,0)		(21/0)		(21/0)
Inflammation	1	(100%)										
Transitional epithelium, hyperplasia	1	(100%)										
Urinary bladder	(52)	()	(53)		(53)		(52)		(53)		(53)	
Calculus microscopic observation only	(==)		1	(2%)	()		(==)		()		()	
Hemorrhage			1	(2%)								
Inflammation	6	(12%)	10	(19%)	4	(8%)			8	(15%)	3	(6%)
Metaplasia, squamous	Ũ	( = )	- 0	()	•	()			0	()	1	(2%)
Transitional epithelium, hyperplasia	1	(2%)	2	(4%)	1	(2%)			6	(11%)	2	(4%)
······································		< · · · /	-						0		-	

#### TABLE A5c Summary of the Incidence of Nonneoplastic Lesions in Female Rats in the 2-Year Gavage Study of TCDD (Stop-Exposure)<sup>a</sup>

Deposition Summary         Signal Active Signal Science		Vehicle Control		100	100 ng/kg		100 ng/kg (Stop-Exposure)	
Animals initially in study     53     53     50       Morband     19     17     16       Natural deaths     9     15     13       Survivors	Disposition Summary							
Early deaths in the second se	Animals initially in study		53		53		50	
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Early deaths							
Natural deaths91513Terminal secrifice252121Animals examined microscopically535350Terminal secrifice252121Animals examined microscopically535350Temoty System1(2%)1Henstine microscopically53(3)(3)Mascularis, inflammation1(2%)2(4%)Intestine large, colon(5)(3)(5)Intestine large, colon(5)(3)(5)Intestine microscopically(5)(5)(10%)Intestine small, jejunam(5)(5)(5)Infammation, chronic1(2%)1Cyst1(2%)(3)(6)Angiectasis2<(4%)	Moribund		19		17		16	
Survivors         25         21         21           Animals examined microscopically         53         53         50           Animals examined microscopically         12%         63         60           Insertine large, colon         12%         24%         12%           Parsite metazoan         6 (12%)         1 (2%)         500           Insertine range, colon         63         (2%)         60           Insertine range, colon         63         (2%)         60           Insertine range, colon         63         (2%)         60           Insertine range, colon         63         (2%)         1 (2%)           Insertine range, colon         63         (2%)         1 (2%)           Insertine range, colon         63         (2%)         1 (2%)           Color cell focus, multiple         4 (8%)         3 (6%)         7 (14%)           Color cell focus, multiple         2 (4%)         6 (12%)         2 (4%)           Color cell focus	Natural deaths		9		15		13	
Terminal scarifice       25       21       21         Animals examined microscopically       53       53       50         Animals examined microscopically       53       53       50         Amimals examined microscopically       53       53       50         Amimals examined microscopically       63       (50)       (49)         Instaine large colon       (52)       (51)       (49)         Therstine large colon       (52)       (53)       (50)         Parasite metazoan       6       (12%)       1       (2%)         Instaine sange rotum       (51)       (29)       (50)         Instaine sange rotum       (51)       (29)       (50)         Instaine sange rotum       (51)       (29)       (50)         Instaine sange rotum       (53)       (53)       (49)         Cyst       1       (2%)       1       (2%)         Instaine sange rotum       (53)       (50)       9       (18%)         Ange costs       8       (15%)       3       (5%)       7       (14%)         Cyst       1       (2%)       6       (12%)       6       (12%)       6       (12%)       6       (12%) <td>Survivors</td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td>	Survivors							
Animals examined microscopically         53         53         50           Atimetery System         Esphagas         (52)         (53)         (50)           Muscularis, inflammation         1         (24)         (49)           Parasite metazoan         1         (24)         1         (27)           Insettine large, scloan         (52)         (53)         (50)           Parasite metazoan         (6)         (26)         (12)         (50)           Insettine large, scloan         (6)         (26)         (12)         (50)           Insettine stang, liquum         (51)         (22)         (50)           Insettine stang, liquum         (53)         (30)         (49)           Cyst         1         (2%)         (2%)         (51)         (28)           Insertine stang, liquud         (53)         (49)         (48)         (58)         (49)           Cyst         1         (2%)         (2%)         (28)         (48)         (58)         (12%)           Chorage/ofboxis         1         (2%)         3         (58%)         1<(2%)	Terminal sacrifice		25		21	21		
Alimentary SystemEsophagus(52)(53)(50)Muscularis, inflammation1(2%)1Intestine large, colon(2%)(50)(50)Parasite netazoan1(2%)(2%)(50)Intestine large, colun(52)(50)(50)Parasite metazoan6(1%)(2%)(50)Intestine salle, figurum(51)(23)(50)Inflammation, chronic1(2%)(50)Cyst1(2%)(50)Cyst1(2%)(50)Angicetasis(3)(58)(74%)Apria cellular(3)(5%)1Basophilic focus, multiple4(8%)5Chear cell focus, multiple1(2%)2Cyst2(4%)2(4%)Congestion2(4%)2(4%)Cytoplasmic alteration2(4%)2(4%)Degeneration, cystic4(8%)4(8%)Eastry change, focal2(4%)2(4%)Hypatodiaphrgunatic nodule1(2%)2(4%)Hypatodiaphrgunatic nodule1(2%)1(2%)Hypatodiaphrgunatic nodule1(2%)1(2%)Hypatodiaphrgunatic nodule1(2%)1(2%)Hypatodiaphrgunatic nodule1(2%)1(2%)Hypatodiaphrgunatic nodule1(2%)1(2%)Hypatodiaphrgunatic nodule <td>Animals examined microscopically</td> <td colspan="2">53</td> <td></td> <td>53</td> <td colspan="2">50</td> <td></td>	Animals examined microscopically	53			53	50		
Esophage(52)(53)(50)Mascularis, inflammation1(2%)(51)(49)Parasite metazoan1(2%)2(4%)1Itestine large, cetum(52)(51)(2%)(50)Parasite metazoan6(12%)1(2%)Intestine alley, cetum(51)(52)(50)Intestine salley, liquam(51)(53)(49)Cyst1(2%)(4%)Cyst1(2%)(53)Intestine salley, duodenum(53)(53)(50)Appia cellular1(2%)(4%)Basophilic focus8(15%)3(6%)Aupia cellular1(2%)1(2%)Charley focus, multiple1(2%)2(4%)Charley focus, multiple1(2%)2(4%)Cogestion2(4%)1(2%)Cogestion2(4%)1(2%)Cogestion2(4%)1(2%)Degeneration, cystic4(8%)1(2%)Eastyphiliz focus, multiple3(6%)4(4%)Hepatodiaphragmatic nodule1(2%)2(4%)Hepatodiaphragmatic nodule1(2%)1(2%)Fatty change, focal2(4%)4(8%)1Hepatodiaphragmatic nodule1(2%)1(2%)Hintomation33(6%)4(8%)1Hepato	Alimentary System							
Massularis, inflammation       (2%)       (2%)       (2%)         Intestine large, colon       (22)       (51)       (49)         Parasite metazoan       (6       (2%)       1       (2%)         Intestine large, rectum       (52)       (53)       (50)         Parasite metazoan       (6       (12%)       1       (2%)         Intestine small, jejonum       (51)       (52)       (50)         Inflammation, chronic       1       (2%)       1       (2%)         Liver       (53)       (53)       (50)       (49)         Cyst       1       (2%)       1       (2%)         Liver       (53)       (53)       (50)       1       (2%)         Basophilic focus       8       (15%)       3       (6%)       7       (14%)         Basophilic focus       4       (8%)       1       (2%)       1       (2%)         Char cell focus       1       (2%)       1       (2%)       1       (2%)         Congestion       2       (4%)       1       (2%)       1       (2%)       1       (2%)       1       (2%)       1       (2%)       1       (2%)       1       <	Esophagus	(52)		(53)		(50)		
	Muscularis, inflammation	1	(2%)	()		()		
Parasite metazoan       (1 $(2\%)$ (2 $(4\%)$ (1 $(2\%)$ Intestine large, rectum       (52)       (53)       (50)         Parasite metazoan       (6       (12\%)       (1       (2%)         Intestine small, jejunum       (51)       (52)       (50)         Inflammation, chronic       1       (2%)       (49)         Cyst       1       (2%)       (53)       (50)         Angicetasis       2       (4%)       5       (9%)       4       (8%)         Atypia cellular       1       (2%)       1       (2%)       1       (2%)         Basophilic focus, multiple       4       (8%)       1       (2%)       1       (2%)         Clear cell focus       8       (15%)       3       (5%)       7       (14%)         Clear cell focus       4       (8%)       1       (2%)       1       (2%)         Cogestion       2       (4%)       1       (2%)       1       (2%)         Cogestion cystic       4       (8%)       1       (2%)       1       (2%)         Eosinophilic focus, multiple       3       (6%)       4       (8/9)	Intestine large, colon	(52)	(_,*)	(51)		(49)		
Intestine large, rectum       (52)       (53)       (50)       (50)         Parasite metazoan       6       (12%)       1       (2%)       5       (10%)         Intestine small, jejnum       (51)       (52)       (50)       (10%)       Intestine small, jejnum       (2%)         Intestine small, judoenum       (53)       (53)       (49)       (2%)       (49)         Cyst       1       (2%)       1       (2%)       1       (2%)         Angiectasis       2       (4%)       5       (9%)       4       (8%)         Atypia cellular       1       (2%)       3       (6%)       7       (14%)         Basophilic focus, multiple       4       (8%)       5       (9%)       9       (15%)         Chear cell focus       4       (8%)       1       (2%)       6       (12%)         Congestion       2       (4%)       2       (4%)       1       (2%)         Degeneration, cystic       4       (8%)       1       (2%)       1       (2%)         Eastry change, focal       2       (4%)       6       (10%)       1       (2%)         Degeneration, cystic       4       (8%) </td <td>Parasite metazoan</td> <td>(02)</td> <td>(2%)</td> <td>2</td> <td>(4%)</td> <td>()</td> <td>(2%)</td> <td></td>	Parasite metazoan	(02)	(2%)	2	(4%)	()	(2%)	
Image retazoan(b)(b)(b)Parasite metazoan6 (12%)1 (2%)5 (10%)Intestine small, jejumun(51)(52)(50)Inflammation, chronic1 (2%)1 (2%)Intestine small, duodenum(53)(53)(50)Angiectasis2 (4%)5 (9%)4 (8%)Atypia cellular1 (2%)1 (2%)Basophilic focus, multiple8 (15%)3 (6%)7 (14%)Basophilic focus, multiple4 (8%)1 (2%)2 (4%)Clear cell focus, multiple1 (2%)1 (2%)2 (4%)Cytoplasmic alteration2 (4%)2 (4%)2 (4%)Cytoplasmic alteration2 (4%)2 (4%)6 (12%)Degeneration, cystic4 (8%)1 (2%)1 (2%)Fatty change, focal2 (4%)2 (4%)2 (4%)Heyatoophilic focus, multiple3 (6%)42 (79%)21 (42%)Fatty change, focal2 (4%)2 (4%)2 (4%)Heyatoophilic focus, multiple3 (6%)40 (92%)43 (86%)Heyatodaphargamatic nodular1 (2%)2 (4%)2 (4%)Heyatodaphargamatic nodular1 (2%)1 (2%)1 (2%)Horeplasia, nodular5 (10%)4 (8%)1 (2%)Horeplasia, nodular6 (8%)1 (2%)1 (2%)Hindammation33 (6%)4 (8%)1 (2%)Mixed cell focus4 (8%)5 (10%)6 (12%)Mixed cell focus4 (8%)5 (10%)6 (12%)Mixed cell focus4 (8%)5 (10%)<	Intestine large rectum	(52)	(2,0)	(53)	(170)	(50)	(270)	
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Parasite metazoan	(32)	(12%)	(33)	(2%)	(30)	(10%)	
Inflammation, chronic       1 (2%)         Inflammation, chronic       1 (2%)         Intestine small, duodenum       (53)       (53)       (50)         Angiectasis       2 (4%)       5 (9%)       4 (8%)         Angiectasis       2 (4%)       5 (9%)       4 (8%)         Basophilic focus, multiple       8 (15%)       3 (6%)       7 (14%)         Basophilic focus, multiple       4 (8%)       1 (2%)       2 (4%)         Changiofibrosis       1 (2%)       1 (2%)       2 (4%)         Clar cell focus, multiple       1 (2%)       1 (2%)       2 (4%)         Congestion       2 (4%)       2 (4%)       2 (4%)         Degeneration, cystic       4 (8%)       1 (2%)       2 (4%)         Eosinophilic focus, multiple       3 (6%)       4 (10%)       8 (16%)         Eosinophilic focus, multiple       3 (6%)       4 (10%)       8 (16%)         Degeneration, cystic       4 (8%)       1 (2%)       2 (4%)         Easinophilic focus, multiple       3 (6%)       2 (4%)       2 (4%)         Haty change, diffuse       1 (2%)       2 (4%)       2 (4%)         Heatodiaphynagmatic nodule       1 (2%)       3 (6%)       4 (8%)         Hyperplasia, nodular <td>Intestine small jejunum</td> <td>(51)</td> <td>(1270)</td> <td>(52)</td> <td>(270)</td> <td>(50)</td> <td>(10/0)</td> <td></td>	Intestine small jejunum	(51)	(1270)	(52)	(270)	(50)	(10/0)	
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Inflammation chronic	(51)		(52)		(50)	(20/2)	
	Intestine small duodenum	(53)		(53)		(40)	(270)	
Cyst       (53)       (53)       (50)         Angicetasis       2 (4%)       5 (9%)       4 (8%)         Atypia cellular       1       (2%)         Basophilic focus       8 (15%)       3 (6%)       7 (14%)         Basophilic focus, multiple       4 (8%)       5 (9%)       9 (18%)         Cholangiofibrosis       1 (2%)       1 (2%)       6 (12%)         Clear cell focus, multiple       1 (2%)       1 (2%)       6 (12%)         Congestion       2 (4%)       2 (4%)       1 (2%)         Cytoplasmic alteration       2 (4%)       1 (2%)       1 (2%)         Degeneration, cystic       4 (8%)       1 (2%)       1 (2%)         Eosinophilic focus, multiple       3 (6%)       42 (79%)       1 (2%)         Fatty change, diffuse       4 (8%)       1 (2%)       2 (4%)         Fatty change, focal       2 (4%)       2 (4%)       8 (16%)         Hyperplasia, nodular       1 (2%)       2 (4%)       1 (2%)         Hyperplasia, nodular       3 (62%)       49 (92%)       43 (8%)         Hyperplasia, nodular       1 (2%)       17 (32%)       8 (16%)         Nixed cell focus, multiple       26 (49%)       17 (32%)       8 (5%)	Cyst	(55)		(55)	(294)	(49)		
$\begin{array}{c c c c c c c } \begin{array}{c c c c c c } (1,5) & (2,5) & (3,0) & (3,0) \\ \hline Angiectasis & 2 & (4\%) & 5 & (9\%) & 4 & (8\%) \\ \hline Arypia cellular & & & & & & & & & & & & & & & & & & &$	Liver	(52)		(52)	(270)	(50)		
Augice calls2294 $(8\%)$ Basophilic focus8 $(15\%)$ 3 $(6\%)$ 7 $(14\%)$ Basophilic focus, multiple4 $(8\%)$ 5 $(9\%)$ 9 $(18\%)$ Cholangi offbrosis1 $(2\%)$ 31 $(58\%)$ 1 $(2\%)$ Clear cell focus, multiple1 $(2\%)$ 1 $(2\%)$ 2 $(4\%)$ Congestion2 $(4\%)$ 1 $(2\%)$ 2 $(4\%)$ Degeneration, cystic486 $(12\%)$ Eosinophilic focus, multiple3 $(6\%)$ 42 $(79\%)$ 21 $(42\%)$ Fatty change, diffuse4 $(8\%)$ 1 $(2\%)$ 2 $(4\%)$ Fatty change, focal2 $(4\%)$ 2 $(4\%)$ 8 $(16\%)$ Hepatodia/pragmatic nodular1 $(2\%)$ 2 $(4\%)$ 8 $(16\%)$ Hyperplasia, nodular33 $(62\%)$ 4 $(8\%)$ 1 $(2\%)$ Mixed cell focus, multiple26 $(49\%)$ 1 $(2\%)$ 2 $(4\%)$ Hyperplasia, nodular33 $(62\%)$ 4 $(8\%)$ 1 $(2\%)$ Inflammation33 $(62\%)$ 17 $(32\%)$ 28 $(5\%)$ Nice cosis1 $(2\%)$ 1 $(2\%)$ 1 $(2\%)$ Nice cosis1 $(2\%)$ 1 $(2\%)$ 1 $(2\%)$ Thrombosis1 $(2\%)$ 1 $(2\%)$ 1 $(2\%)$ Thrombosis1 $(2\%)$ 1<	Ancientaria	(33)	(40/)	(33)	(00/)	(30)	(80/)	
Arypia centual1 (2%)Basophilic focus8 (15%)3 (6%)7 (14%)Basophilic focus, multiple4 (8%)5 (9%)9 (18%)Chalagiofibrosis1 (2%)31 (58%)1 (2%)Clear cell focus, multiple1 (2%)6 (12%)Clear cell focus, multiple1 (2%)2 (4%)Cytoplasmic alteration2 (4%)2 (4%)Degeneration, cystic4 (8%)1 (2%)Eosinophilic focus, multiple3 (6%)42 (79%)Eosinophilic focus, multiple3 (6%)42 (79%)Eosinophilic focus, multiple3 (6%)42 (79%)Fatty change, diffuse48 (91%)10 (20%)Fatty change, focal2 (4%)2 (4%)Hematopoictic cell proliferation1 (2%)2 (4%)Hematopoictic cell proliferation1 (2%)2 (4%)Hepatodiaphragmatic nodule1 (2%)2 (4%)Hyperplasia, nodular6 (62%)1 (2%)Inflammation33 (62%)49 (92%)Mixed cell focus4 (8%)4 (8%)Necrosis1 (2%)12 (2%)Pigmentation4 (8%)53 (100%)Necrosis1 (2%)1 (2%)Thrombosis1 (2%)2 (4%)Thrombosis1 (2%)1 (2%)Attery, inflammation, chronic active2 (4%)Attery, inflammation, chronic active2 (4%)Bile duct, fibrosis4 (8%)5 (10%)Bile duct, fibrosis4 (8%)5 (10%)Bile duct, fibrosis5 (9%)52 (9%)Bile	Augiculasis	2	(470)	5	(970)	4	(070)	
Dasophilic focus Basophilic focus, multiple $\delta$ (12%) $\delta$ (12%) $\delta$ 	Atypia cenular Degenhilie feeug	0	(150/)	2	(60/)	1	(270)	
Basephilic focus, multiple4 ( $8\%$ )5 ( $9\%$ )9 ( $18\%$ )Cholangiofibrosis1 ( $2\%$ )31 ( $28\%$ )1 ( $2\%$ )Clear cell focus4 ( $8\%$ )1 ( $2\%$ )2 ( $4\%$ )Congestion2 ( $4\%$ )2 ( $4\%$ )Cytoplasmic alteration2 ( $4\%$ )2 ( $4\%$ )Degeneration, cystic4 ( $8\%$ )1 ( $2\%$ )Eosinophilic focus, multiple3 ( $6\%$ )42 ( $79\%$ )21 ( $42\%$ )Fatty change, focal2 ( $4\%$ )10 ( $20\%$ )Fatty change, focal2 ( $4\%$ )2 ( $4\%$ )8 ( $16\%$ )Hematopoietic cell proliferation1 ( $2\%$ )2 ( $4\%$ )2 ( $4\%$ )Hyperplasia, nodular1 ( $2\%$ )2 ( $4\%$ )2 ( $4\%$ )Inflammation33 ( $62\%$ )49 ( $92\%$ )43 ( $86\%$ )Mixed cell focus, multiple26 ( $49\%$ )1 ( $2\%$ )Mixed cell focus, multiple26 ( $49\%$ )1 ( $2\%$ )Mixed cell focus, multiple26 ( $49\%$ )4 ( $8\%$ )1 ( $2\%$ )Mixed cell focus, multiple26 ( $49\%$ )4 ( $8\%$ )1 ( $2\%$ )Mixed cell focus, multiple26 ( $49\%$ )4 ( $8\%$ )1 ( $2\%$ )Mixed cell focus, multiple26 ( $49\%$ )53 ( $100\%$ )45 ( $90\%$ )Thrombosis1 ( $2\%$ )17 ( $32\%$ )8 ( $16\%$ )Pigmentation4 ( $8\%$ )53 ( $100\%$ )45 ( $90\%$ )Thrombosis1 ( $2\%$ )1 ( $2\%$ )1 ( $2\%$ )Hyperplasia5 ( $9\%$ )21 ( $4\%$ )5 ( $10\%$ )Bile duct, fibrosis4 ( $8\%$ )53 ( $100\%$ )7 ( $14\%$ )Bile duct, fibrosis5 ( $9\%$ ) </td <td>Dasophilic focus</td> <td>0</td> <td>(1370)</td> <td>5</td> <td>(0%)</td> <td>/</td> <td>(14%)</td> <td></td>	Dasophilic focus	0	(1370)	5	(0%)	/	(14%)	
Cloar coll focus1 $(2\%)$ 31 $(2\%)$ 1 $(2\%)$ Clear cell focus, multiple1 $(2\%)$ 1 $(2\%)$ 2 $(4\%)$ Congestion2 $(4\%)$ 1 $(2\%)$ 1 $(2\%)$ Cytoplasmic alteration2 $(4\%)$ 1 $(2\%)$ Degeneration, cystic4 $(8\%)$ 6 $(12\%)$ Eosinophilic focus, multiple3 $(6\%)$ 2 $(4\%)$ 6 $(12\%)$ Fatty change, diffuse48 $(91\%)$ 10 $(22\%)$ Fatty change, diffuse48 $(91\%)$ 10 $(22\%)$ Hepatodiaphragmatic nodule1 $(2\%)$ 2 $(4\%)$ 2Hyperplasia, nodular36 $(62\%)$ 2 $(4\%)$ 2Hyperplasia, nodular36 $(62\%)$ 43 $(86\%)$ Mixed cell focus4 $(8\%)$ 4 $(2\%)$ 28Necrosis1 $(2\%)$ 17 $(32\%)$ 8 $(16\%)$ Pigmentation4 $(8\%)$ 53 $(100\%)$ 45 $(90\%)$ Necrosis1 $(2\%)$ 17 $(32\%)$ 8 $(16\%)$ Pigmentation, chronic active2 $(4\%)$ 1 $(2\%)$ Thrombosis1 $(2\%)$ 16 $(32\%)$ Artery, inflammation, chronic active2 $(4\%)$ 5 $(10\%)$ Bile duct, fibrosis4 $(8\%)$ 5 $(10\%)$ Bile duct, fibrosis5 $(9\%)$ 53 $(100\%)$ 7Bile duct, fibrosis5 <td>Basophilic locus, multiple</td> <td>4</td> <td>(8%)</td> <td>5</td> <td>(9%)</td> <td>9</td> <td>(18%)</td> <td></td>	Basophilic locus, multiple	4	(8%)	5	(9%)	9	(18%)	
Clear cell focus4 (8%)1 (2%)6 (12%)Cara cell focus, multiple1 (2%)2 (4%)2 (4%)Congestion2 (4%)2 (4%)1 (2%)Degeneration, cystic2 (4%)6 (12%)Eosinophilic focus, multiple3 (6%)42 (79%)21 (42%)Fatty change, diffuse4 (8%)10 (20%)Fatty change, focal2 (4%)2 (4%)8 (16%)Hematopoietic cell proliferation1 (2%)2 (4%)2 (4%)Hepatodiaphragmatic nodule33 (62%)49 (92%)43 (86%)Inflammation33 (62%)49 (92%)43 (86%)Mixed cell focus, multiple26 (49%)17 (32%)28 (56%)Necrosis1 (2%)17 (32%)8 (16%)Pigenetation4 (8%)4 (8%)1 (2%)Necrosis1 (2%)17 (32%)8 (16%)Pigenetation4 (8%)1 (2%)1 (2%)Necrosis1 (2%)1 (2%)1 (2%)Thrombosis1 (2%)1 (2%)1 (2%)Thrombosis1 (2%)1 (2%)1 (2%)Artery, inflammation, chronic active2 (4%)5 (10%)Bile duct, fibrosis4 (8%)5 (10%)5 (10%)Bile duct, fibrosis5 (9%)5 (9%)1Hepatocyte, hypertrophy5 (9%)5 (9%)22 (4%)	Cholangiofibrosis	1	(2%)	31	(58%)	I	(2%)	
Clear cell focus, multiple1 $(2\%)$ 2 $(4\%)$ Congestion2 $(4\%)$ 2 $(4\%)$ Cytoplasmic alteration2 $(4\%)$ 1 $(2\%)$ Degeneration, cystic4 $(8\%)$ $(12\%)$ Eosinophilic focus, multiple3 $(6\%)$ 42 $(79\%)$ 21Fatty change, diffuse48 $(91\%)$ 10 $(20\%)$ Fatty change, focal2 $(4\%)$ 8 $(16\%)$ Hematopoietic cell proliferation1 $(2\%)$ 2 $(4\%)$ Hepatodiaphragmatic nodula3 $(62\%)$ 49 $(92\%)$ 43Mixed cell focus, multiple26 $(49\%)$ 17 $(32\%)$ 28Mixed cell focus, multiple26 $(49\%)$ 17 $(32\%)$ 8 $(16\%)$ Necrosis1 $(2\%)$ 1 $(2\%)$ 1 $(2\%)$ Thrombosis1 $(2\%)$ 1 $(2\%)$ 1 $(2\%)$ Vacuolization cytoplasmic2 $(4\%)$ 53 $(100\%)$ 45 $(90\%)$ Vacuolization cytoplasmic2 $(4\%)$ 53 $(100\%)$ 16 $(32\%)$ Vacuolization cytoplasmic2 $(4\%)$ 53 $(100\%)$ 5 $(10\%)$ Bile duct, fibrosis4 $(8\%)$ 53 $(100\%)$ 7 $(14\%)$ Bile duct, fibrosis5 $(9\%)$ 53 $(100\%)$ 7 $(14\%)$ Bile duct, fibrosis5 $(9\%)$ 53 $(100\%)$ 7 $(14\%)$ Bile duct, fibrosis5 <td< td=""><td>Clear cell focus</td><td>4</td><td>(8%)</td><td>1</td><td>(2%)</td><td>6</td><td>(12%)</td><td></td></td<>	Clear cell focus	4	(8%)	1	(2%)	6	(12%)	
Conjection       2 (4%)       2 (4%)         Cytoplasmic alteration       2 (4%)       1 (2%)         Degeneration, cystic       4 (8%)         Eosinophilic focus       8 (15%)       2 (4%)       6 (12%)         Eosinophilic focus, multiple       3 (6%)       42 (79%)       21 (42%)         Fatty change, focal       2 (4%)       2 (4%)       8 (16%)         Hematopoietic cell proliferation       1 (2%)       2 (4%)       2 (4%)         Hepatodiaphragmatic nodule       1 (2%)       2 (4%)       2 (4%)         Hyperplasia, nodular       33 (62%)       49 (92%)       43 (86%)         Mixed cell focus       4 (8%)       1 (2%)       28 (56%)         Necrosis       1 (2%)       17 (32%)       28 (56%)         Necrosis       1 (2%)       17 (32%)       8 (16%)         Pigmentation       4 (8%)       53 (100%)       45 (90%)         Thrombosis       1 (2%)       1 (2%)       1 (2%)         Thrombosis       1 (2%)       1 (2%)       1 (2%)         Bile duct, cyst       3 (6%)       21 (40%)       6 (12%)         Bile duct, fibrosis       4 (8%)       5 (10%)       6 (12%)         Bile duct, fibrosisis       5 (9%)       5	Clear cell focus, multiple	1	(2%)	1	(2%)	2	(4%)	
Cytoplasmic alteration2 $(4\%)$ 1 $(2\%)$ Degeneration, cystic4 $(8\%)$ Eosinophilic focus8 $(15\%)$ 2 $(4\%)$ 6 $(12\%)$ Eosinophilic focus, multiple3 $(6\%)$ 42 $(79\%)$ 21 $(42\%)$ Fatty change, diffuse48 $(91\%)$ 10 $(20\%)$ Fatty change, focal2 $(4\%)$ 2 $(4\%)$ 8 $(16\%)$ Hematopoietic cell proliferation1 $(2\%)$ 2 $(4\%)$ 2 $(4\%)$ Hepatodiaphragmatic nodule1 $(2\%)$ 1 $(2\%)$ Hyperplasia, nodular33 $(62\%)$ 49 $(92\%)$ 43 $(86\%)$ Mixed cell focus4 $(8\%)$ 4 $(8\%)$ 1 $(2\%)$ Necrosis1 $(2\%)$ 17 $(32\%)$ 28 $(56\%)$ Pigmentation4 $(8\%)$ 53 $(100\%)$ 45 $(90\%)$ Thrombosis1 $(2\%)$ 1 $(2\%)$ $(2\%)$ Vacuolization cytoplasmic2 $(4\%)$ 53 $(100\%)$ 45 $(90\%)$ Artery, inflammation, chronic active2 $(4\%)$ 53 $(10\%)$ $(2\%)$ Bile duct, fibrosis4 $(8\%)$ 53 $(10\%)$ $(14\%)$ Bile duct, fibrosis4 $(8\%)$ 53 $(10\%)$ $(14\%)$ Bile duct, fibrosis5 $(9\%)$ 53 $(10\%)$ $(14\%)$ Bile duct, fibrosis5 $(2\%)$ $(2\%)$ $(2\%)$ Hepatocyte, hypertrophy <td>Congestion</td> <td>2</td> <td>(4%)</td> <td>2</td> <td>(4%)</td> <td></td> <td></td> <td></td>	Congestion	2	(4%)	2	(4%)			
Degeneration, cystic4 (8%)Eosinophilic focus, multiple3 (6%)42 (7%)6 (12%)Eosinophilic focus, multiple3 (6%)42 (7%)21 (42%)Fatty change, diffuse48 (91%)10 (20%)Fatty change, focal2 (4%)2 (4%)8 (16%)Hematopoietic cell proliferation1 (2%)2 (4%)2 (4%)Hepatodiaphragmatic nodule1 (2%)2 (4%)4 (8%)Hyperplasia, nodular33 (62%)49 (92%)43 (86%)Inflammation33 (62%)49 (92%)43 (86%)Mixed cell focus, multiple26 (49%)17 (32%)28 (56%)Necrosis1 (2%)17 (32%)8 (16%)Pigmentation4 (8%)53 (100%)45 (90%)Thrombosis1 (2%)1 (2%)Toxic hepatopathy53 (100%)16 (32%)Vacuolization cytoplasmic2 (4%)2 (4%)Bile duct, cyst3 (6%)21 (40%)6 (12%)Bile duct, fibrosis4 (8%)53 (100%)7 (14%)Centrilobular, degeneration2 (4%)5 (9%)22 (44%)	Cytoplasmic alteration			2	(4%)	1	(2%)	
Eosinophilic focus8 (15%)2 (4%)6 (12%)Eosinophilic focus, multiple3 (6%)42 (79%)21 (42%)Fatty change, diffuse48 (91%)10 (20%)Fatty change, focal2 (4%)2 (4%)8 (16%)Hematopoietic cell proliferation1 (2%)2 (4%)2 (4%)Hepatodiaphragmatic nodule1 (2%)2 (4%)1 (2%)Hyperplasia, nodular36 (68%)1 (2%)Inflammation33 (62%)49 (92%)43 (86%)Mixed cell focus4 (8%)1 (2%)Mixed cell focus, multiple26 (49%)17 (32%)28 (56%)Necrosis1 (2%)17 (32%)8 (16%)Pigmentation4 (8%)53 (100%)45 (90%)Thrombosis1 (2%)1 (2%)1Toxic hepatopathy53 (100%)16 (32%)Vacuolization cytoplasmic2 (4%)2 (4%)Bile duct, cyst3 (6%)51 (10%)6 (12%)Bile duct, thyperplasia5 (9%)53 (100%)7 (14%)Centrilobular, degeneration2 (4%)5 (9%)Hepatocyte, hypertrophy52 (98%)22 (44%)	Degeneration, cystic			4	(8%)			
Eosinophilic focus, multiple3 $(6\%)$ 42 $(79\%)$ 21 $(42\%)$ Fatty change, focal2 $(4\%)$ 10 $(20\%)$ Fatty change, focal2 $(4\%)$ 2 $(4\%)$ Hematopoietic cell proliferation1 $(2\%)$ 2 $(4\%)$ Hepatodiaphragmatic nodule1 $(2\%)$ 2 $(4\%)$ Hyperplasia, nodular36 $(68\%)$ 1 $(2\%)$ Inflammation33 $(62\%)$ 49 $(92\%)$ 43 $(86\%)$ Mixed cell focus4 $(8\%)$ 1 $(2\%)$ Mixed cell focus, multiple26 $(49\%)$ 17 $(32\%)$ 28 $(56\%)$ Necrosis1 $(2\%)$ 17 $(32\%)$ 8 $(16\%)$ Pigmentation4 $(8\%)$ 53 $(100\%)$ 45 $(90\%)$ Toxic hepatopathy53 $(100\%)$ 16 $(32\%)$ Vacuolization cytoplasmic1 $(2\%)$ 1 $(2\%)$ Artery, inflammation, chronic active2 $(4\%)$ 5 $(10\%)$ Bile duct, cyst3 $(6\%)$ 53 $(100\%)$ 5 $(10\%)$ Bile duct, thyperplasia5 $(9\%)$ 53 $(100\%)$ 7 $(14\%)$ Centrilobular, degeneration2 $(4\%)$ 5 $(9\%)$ $(14\%)$ Hepatocyte, hypertrophy52 $(9\%)$ 22 $(44\%)$	Eosinophilic focus	8	(15%)	2	(4%)	6	(12%)	
Fatty change, diffuse48 (91%)10 (20%)Fatty change, focal2 (4%)2 (4%)8 (16%)Hematopoietic cell proliferation1 (2%)2 (4%)2 (4%)Hepatodiaphragmatic nodule1 (2%)1 (2%)Hyperplasia, nodular36 (68%)Inflammation33 (62%)49 (92%)43 (86%)Mixed cell focus4 (8%)4 (8%)1 (2%)Mixed cell focus, multiple26 (49%)17 (32%)28 (56%)Necrosis1 (2%)17 (32%)8 (16%)Pigmentation4 (8%)53 (100%)45 (90%)Thrombosis1 (2%)1 (2%)Toxic hepatopathy53 (100%)16 (32%)Vacuolization cytoplasmic2 (4%)2 (4%)Bile duct, cyst3 (6%)21 (40%)6 (12%)Bile duct, throrsis4 (8%)53 (100%)7 (14%)Centrilobular, degeneration2 (4%)5 (9%)22 (44%)	Eosinophilic focus, multiple	3	(6%)	42	(79%)	21	(42%)	
Fatty change, focal2 (4%)2 (4%)8 (16%)Hematopoietic cell proliferation1 (2%)2 (4%)2 (4%)Hepatodiaphragmatic nodule1 (2%)2 (4%)1 (2%)Hyperplasia, nodular36 (68%)36(68%)Inflammation33 (62%)49 (92%)43 (86%)Mixed cell focus4 (8%)4 (8%)1 (2%)Mixed cell focus, multiple26 (49%)17 (32%)28 (56%)Necrosis1 (2%)17 (32%)8 (16%)Pigmentation4 (8%)53 (100%)45 (90%)Thrombosis1 (2%)1 (2%)1 (2%)Toxic hepatopathy53 (100%)16 (32%)Vacuolization cytoplasmic2 (4%)1 (2%)Bile duct, cyst3 (6%)21 (40%)6 (12%)Bile duct, thyperplasia5 (9%)53 (100%)7 (14%)Centrilobular, degeneration2 (4%)5 (9%)22 (44%)	Fatty change, diffuse			48	(91%)	10	(20%)	
Hematopoietic cell proliferation1 $(2\%)$ 2 $(4\%)$ 2 $(4\%)$ Hepatodiaphragmatic nodule1 $(2\%)$ 1 $(2\%)$ Hyperplasia, nodular33 $(62\%)$ 49 $(92\%)$ 43Inflammation33 $(62\%)$ 4 $(8\%)$ 1Mixed cell focus4 $(8\%)$ 4 $(8\%)$ 1Mixed cell focus, multiple26 $(49\%)$ 17 $(32\%)$ 28Necrosis1 $(2\%)$ 17 $(32\%)$ 8 $(16\%)$ Pigmentation4 $(8\%)$ 53 $(100\%)$ 45 $(90\%)$ Thrombosis1 $(2\%)$ 16 $(32\%)$ Toxic hepatopathy53 $(100\%)$ 16 $(32\%)$ Vacuolization cytoplasmic2 $(4\%)$ 51 $(10\%)$ Bile duct, fibrosis4 $(8\%)$ 53 $(100\%)$ 6Bile duct, fibrosis4 $(8\%)$ 53 $(100\%)$ 7Bile duct, hyperplasia5 $(9\%)$ 53 $(100\%)$ 7Centrilobular, degeneration2 $(4\%)$ 5 $(9\%)$ Hepatocyte, hypertrophy52 $(98\%)$ 22 $(44\%)$	Fatty change, focal	2	(4%)	2	(4%)	8	(16%)	
Hepatodiaphragmatic nodule1 (2%)Hyperplasia, nodular36 (68%)Inflammation33 (62%)49 (92%)43 (86%)Mixed cell focus4 (8%)4 (8%)1 (2%)Mixed cell focus, multiple26 (49%)17 (32%)28 (56%)Necrosis1 (2%)17 (32%)8 (16%)Pigmentation4 (8%)53 (100%)45 (90%)Thrombosis1 (2%)16 (32%)Toxic hepatopathy53 (100%)16 (32%)Vacuolization cytoplasmic1 (2%)Artery, inflammation, chronic active2 (4%)Bile duct, cyst3 (6%)21 (40%)6 (12%)Bile duct, fibrosis4 (8%)53 (100%)7 (14%)Centrilobular, degeneration2 (4%)5 (9%)22 (44%)	Hematopoietic cell proliferation	1	(2%)	2	(4%)	2	(4%)	
Hyperplasia, nodular36 $68\%$ )Inflammation33 $(62\%)$ 49 $(92\%)$ 43 $(86\%)$ Mixed cell focus4 $(8\%)$ 4 $(8\%)$ 1 $(2\%)$ Mixed cell focus, multiple26 $(49\%)$ 17 $(32\%)$ 28 $(56\%)$ Necrosis1 $(2\%)$ 17 $(32\%)$ 8 $(16\%)$ Pigmentation4 $(8\%)$ 53 $(100\%)$ 45 $(90\%)$ Thrombosis1 $(2\%)$ 1 $(2\%)$ Toxic hepatopathy53 $(100\%)$ 16 $(32\%)$ Vacuolization cytoplasmic1 $(2\%)$ 1 $(2\%)$ Artery, inflammation, chronic active2 $(4\%)$ 5 $(10\%)$ Bile duct, fibrosis4 $(8\%)$ 53 $(100\%)$ 7Bile duct, hyperplasia5 $(9\%)$ 53 $(100\%)$ 7Bile duct, hyperplasia5 $(9\%)$ 53 $(100\%)$ 7Hepatocyte, hypertrophy52 $(98\%)$ 22 $(44\%)$	Hepatodiaphragmatic nodule					1	(2%)	
Inflammation33 $(62\%)$ 49 $(92\%)$ 43 $(86\%)$ Mixed cell focus4 $(8\%)$ 4 $(8\%)$ 1 $(2\%)$ Mixed cell focus, multiple26 $(49\%)$ 17 $(32\%)$ 28 $(56\%)$ Necrosis1 $(2\%)$ 17 $(32\%)$ 8 $(16\%)$ Pigmentation4 $(8\%)$ 53 $(100\%)$ 45 $(90\%)$ Thrombosis1 $(2\%)$ 1 $(2\%)$ Toxic hepatopathy53 $(100\%)$ 16 $(32\%)$ Vacuolization cytoplasmic1 $(2\%)$ 1 $(2\%)$ Artery, inflammation, chronic active2 $(4\%)$ 6 $(12\%)$ Bile duct, cyst3 $(6\%)$ 21 $(40\%)$ 6 $(12\%)$ Bile duct, fibrosis4 $(8\%)$ 5 $(10\%)$ 7 $(14\%)$ Centrilobular, degeneration2 $(4\%)$ 5 $(9\%)$ 7 $(14\%)$ Hepatocyte, hypertrophy52 $(98\%)$ 22 $(44\%)$	Hyperplasia, nodular			36	(68%)			
Mixed cell focus4 (8%)4 (8%)1 (2%)Mixed cell focus, multiple26 (49%)17 (32%)28 (56%)Necrosis1 (2%)17 (32%)8 (16%)Pigmentation4 (8%)53 (100%)45 (90%)Thrombosis1 (2%)1 (2%)Toxic hepatopathy53 (100%)16 (32%)Vacuolization cytoplasmic1 (2%)Artery, inflammation, chronic active2 (4%)Bile duct, cyst3 (6%)21 (40%)Bile duct, fibrosis4 (8%)5 (10%)Bile duct, hyperplasia5 (9%)53 (100%)Centrilobular, degeneration2 (4%)5 (9%)Hepatocyte, hypertrophy52 (98%)22 (44%)	Inflammation	33	(62%)	49	(92%)	43	(86%)	
Mixed cell focus, multiple $26$ (49%) $17$ (32%) $28$ (56%)Necrosis1 (2%)17 (32%)8 (16%)Pigmentation4 (8%)53 (100%)45 (90%)Thrombosis1 (2%)1< (2%)	Mixed cell focus	4	(8%)	4	(8%)	1	(2%)	
Necrosis1 $(2\%)$ 17 $(32\%)$ 8 $(16\%)$ Pigmentation4 $(8\%)$ 53 $(100\%)$ 45 $(90\%)$ Thrombosis1 $(2\%)$ 1 $(2\%)$ Toxic hepatopathy53 $(100\%)$ 16 $(32\%)$ Vacuolization cytoplasmic1 $(2\%)$ 1 $(2\%)$ Artery, inflammation, chronic active2 $(4\%)$ 6 $(12\%)$ Bile duct, cyst3 $(6\%)$ 21 $(40\%)$ 6 $(12\%)$ Bile duct, fibrosis4 $(8\%)$ 5 $(10\%)$ Bile duct, hyperplasia5 $(9\%)$ 53 $(100\%)$ 7Centrilobular, degeneration2 $(4\%)$ 5 $(9\%)$ Hepatocyte, hypertrophy52 $(98\%)$ 22 $(44\%)$	Mixed cell focus, multiple	26	(49%)	17	(32%)	28	(56%)	
Pigmentation4 (8%)53 (100%)45 (90%)Thrombosis1 (2%)Toxic hepatopathy53 (100%)16 (32%)Vacuolization cytoplasmic1 (2%)Artery, inflammation, chronic active2 (4%)Bile duct, cyst3 (6%)21 (40%)Bile duct, fibrosis4 (8%)5 (10%)Bile duct, hyperplasia5 (9%)53 (100%)Centrilobular, degeneration2 (4%)5 (9%)Hepatocyte, hypertrophy52 (98%)22 (44%)	Necrosis	1	(2%)	17	(32%)	8	(16%)	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	Pigmentation	4	(8%)	53	(100%)	45	(90%)	
Toxic hepatopathy53 (100%)16 (32%)Vacuolization cytoplasmic1 (2%)Artery, inflammation, chronic active2 (4%)Bile duct, cyst3 (6%)21 (40%)Bile duct, fibrosis4 (8%)5 (10%)Bile duct, hyperplasia5 (9%)53 (100%)Centrilobular, degeneration2 (4%)5 (9%)Hepatocyte, hypertrophy52 (98%)22 (44%)	Thrombosis			1	(2%)			
Vacualization cytoplasmic1 (2%)Artery, inflammation, chronic active2 (4%)Bile duct, cyst3 (6%)21 (40%)Bile duct, fibrosis4 (8%)5 (10%)Bile duct, hyperplasia5 (9%)53 (100%)Centrilobular, degeneration2 (4%)5 (9%)Hepatocyte, hypertrophy52 (98%)22 (44%)	Toxic hepatopathy			53	(100%)	16	(32%)	
Artery, inflammation, chronic active2 (4%)Bile duct, cyst3 (6%)21 (40%)6 (12%)Bile duct, fibrosis4 (8%)5 (10%)Bile duct, hyperplasia5 (9%)53 (100%)7 (14%)Centrilobular, degeneration2 (4%)5 (9%)Hepatocyte, hypertrophy52 (98%)22 (44%)	Vacuolization cytoplasmic				·	1	(2%)	
Bile duct, cyst       3 (6%)       21 (40%)       6 (12%)         Bile duct, fibrosis       4 (8%)       5 (10%)         Bile duct, hyperplasia       5 (9%)       53 (100%)       7 (14%)         Centrilobular, degeneration       2 (4%)       5 (9%)       7 (14%)         Hepatocyte, hypertrophy       52 (98%)       22 (44%)	Artery, inflammation, chronic active			2	(4%)			
Bile duct, fibrosis       4 (8%)       5 (10%)         Bile duct, hyperplasia       5 (9%)       53 (100%)       7 (14%)         Centrilobular, degeneration       2 (4%)       5 (9%)       7         Hepatocyte, hypertrophy       52 (98%)       22 (44%)	Bile duct, cyst	3	(6%)	21	(40%)	6	(12%)	
Bile duct, hyperplasia       5 (9%)       53 (100%)       7 (14%)         Centrilobular, degeneration       2 (4%)       5 (9%)         Hepatocyte, hypertrophy       52 (98%)       22 (44%)	Bile duct, fibrosis	-	. /	4	(8%)	5	(10%)	
Centrilobular, degeneration       2 (4%)       5 (9%)         Hepatocyte, hypertrophy       52 (98%)       22 (44%)	Bile duct, hyperplasia	5	(9%)	53	(100%)	7	(14%)	
Hepatocyte, hypertrophy 52 (98%) 22 (44%)	Centrilobular, degeneration	2	(4%)	5	(9%)		× · · ·	
	Hepatocyte, hypertrophy	2	()	52	(98%)	22	(44%)	

<sup>a</sup> Number of animals examined microscopically at the site and the number of animals with lesion

#### TABLE A5c Summary of the Incidence of Nonneoplastic Lesions in Female Rats in the 2-Year Gavage Study of TCDD (Stop-Exposure)

	Vehicle	e Control	100 ng/kg		100 (Stop-E	ng/kg xposure)
Alimentary System (continued)						
Liver (continued)	(53)		(53)		(50)	
Hepatocyte, multinucleated	× /		51	(96%)	32	(64%)
Oval cell, hyperplasia			53	(100%)	1	(2%)
Portal, fibrosis			27	(51%)	1	(2%)
Serosa, inflammation, chronic active				· /	1	(2%)
Mesentery	(2)		(8)		(1)	
Metaplasia, osseous	1	(50%)				
Artery, inflammation, chronic active			7	(88%)	1	(100%)
Fat, necrosis			1	(13%)		
Vein, thrombosis	1	(50%)		· /		
Oral mucosa	(2)		(29)		(11)	
Gingival, hyperplasia, squamous	1	(50%)	16	(55%)	8	(73%)
Pancreas	(51)	()	(51)	(22,0)	(49)	
Angiectasis	(- )		(- )		1	(2%)
Basophilic focus					1	(2%)
Hemorrhage			1	(2%)	1	(270)
Inflammation chronic active			6	(12%)	4	(8%)
Necrosis			1	(2%)		(0,0)
Acinus atrophy	1	(2%)	9	(18%)	4	(8%)
Acinus, hyperplasia	2	(2%)		(10/0)		(0,0)
Acinus, hyperplasta Acinus, vacualization extenlasmic	1	(2%)	42	(82%)		
Artery inflammation chronic active	1	(270)	7	(14%)	2	(4%)
Saliyary glands	(51)		(52)	(1470)	(49)	(470)
Atrophy	(51)		(52)		((-)) 2	(4%)
Fibrosis					1	(2%)
Inflammation			1	(20%)	1	(270)
Mineralization	1	$(2^{0/2})$	1	(270)	1	(270)
Stomach forestomach	(53)	(270)	(53)		(50)	
Edomo	(55)		(55)	(20/)	(50)	
Euclia Usum entremeteorie	1	(20/)	1	(270)	1	(29/)
Hyperkeratosis	1	(2%)	11	(070)	1	(270)
Information	5	(0%)	11	(21%)	5	(10%)
Necessia	1	(2%)	5	(9%)	4	(8%)
I Hear	2	(40/)	1	(270)	4	(90/)
	(52)	(4%)	4	(8%)	4	(8%)
Stomacn, glandular	(53)	(20())	(53)		(50)	(20/)
Erosion	1	(2%)			1	(2%)
Mineralization	3	(6%)	2	(40/)		
Ulcer			2	(4%)	(1)	
longue					(1)	(1000/)
Inflammation, chronic active	(10)		(10)			(100%)
looth	(16)	((0))	(18)		(12)	
Peridontal tissue, fibrosis	1	(6%)	10	(1000/)	10	(1000/)
Peridontal tissue, inflammation	15	(94%)	18	(100%)	12	(100%)
Cardiovascular System						
Blood vessel	(53)		(53)		(49)	
Aorta mineralization	(00)	(2%)	(00)		()	
Heart	(53)	(270)	(52)		(50)	
Cardiomyopathy	10	(19%)	36	(69%)	22	(44%)
Mineralization	10	(2%)	50	(0270)		(
Inflammation	1	(=/0)			1	(2%)
Atrium thrombosis					1	(4%)
Enicardium inflammation chronic active					1	(2%)
Artery inflammation			1	(2%)	1	
Artery thrombosis			1	(2%)		
. i.e., uioinoosis			1	(270)		

# TABLE A5cSummary of the Incidence of Nonneoplastic Lesions in Female Rats in the 2-Year Gavage Studyof TCDD (Stop-Exposure)

	Vehicle	e Control	100 ng/kg		100 (Stop-E		
Endocrine System							
Adrenal cortex	(53)		(53)		(50)		
Angiectasis	11	(21%)	11	(21%)	15	(30%)	
Atrophy	2	(4%)	27	(51%)	4	(8%)	
Degeneration, cystic	11	(21%)	1/	(32%)	1/	(34%)	
Hyperplasia	10	(30%)	30 47	(57%)	20	(40%)	
Inflammation, chronic active	41	(7770)	4/	(89%)	40	(9270)	
Inflammation, enfonce active	1	(2%)					
Mineralization	1	(270)	1	(2%)			
Necrosis			1	(2%)	3	(6%)	
Vacuolization cytoplasmic	11	(21%)	15	(270)	13	(26%)	
Adrenal medulla	(53)	()	(53)	(_0,0)	(50)	()	
Hyperplasia	19	(36%)	9	(17%)	15	(30%)	
Islets, pancreatic	(51)	()	(52)		(49)	()	
Hyperplasia					1	(2%)	
Parathyroid gland	(46)		(45)		(44)	× /	
Hyperplasia	1	(2%)					
Pituitary gland	(53)		(52)		(50)		
Angiectasis	25	(47%)	9	(17%)	15	(30%)	
Atypia cellular	1	(2%)					
Cyst	1	(2%)	1	(2%)			
Cytoplasmic alteration	1	(2%)			1	(2%)	
Developmental malformation	1	(2%)					
Inflammation, chronic			1	(2%)			
Vacuolization cytoplasmic					3	(6%)	
Pars distalis, hyperplasia	19	(36%)	19	(37%)	18	(36%)	
Pars intermedia, hyperplasia	2	(4%)	1	(2%)			
Thyroid gland	(52)		(52)		(49)		
Angiectasis			1	(2%)	1	(2%)	
Cyst	1	(2%)	1	(2%)			
C-Cell, hyperplasia	19	(37%)	23	(44%)	15	(31%)	
Follicular cell, hyperplasia	2	(4%)	17	(220)		(120/)	
Follicular cell, hypertrophy	3	(6%)	17	(33%)	6	(12%)	
General Body System None							
Conital System							
Cliteral gland	(50)		(52)		(40)		
Atrophy	(30)		(55)		(49)	$(2^{0/2})$	
Hyperplasia basal cell	1	(2%)			1	(270)	
Hyperplasia, basar celi Hyperplasia, squamous	1	(270)			1	(2%)	
Inflammation	41	(82%)	26	(49%)	35	(71%)	
Duct cyst	34	(68%)	20 48	(91%)	35	(71%)	
Ovary	(51)	(0070)	(53)	()1/0)	(49)	(/1/0)	
Atrophy	(31)	(96%)	31	(58%)	45	(92%)	
Cvst	14	(27%)	13	(25%)	16	(33%)	
Inflammation, chronic active	11	()	1	(2%)	10	()	
Inflammation, granulomatous			-	< · · ·	1	(2%)	
Artery, inflammation, chronic active			1	(2%)	-	· /	
Oviduct	(2)		(1)	. ,	(2)		
Inflammation	2	(100%)	ĺ	(100%)	2	(100%)	

# TABLE A5c Summary of the Incidence of Nonneoplastic Lesions in Female Rats in the 2-Year Gavage Study of TCDD (Stop-Exposure)

	Vehicle	e Control	100	ng/kg	100 (Stop-E	ng/kg xposure)
Genital System (continued)						
Uterus	(52)		(53)		(50)	
Cyst			1	(2%)		
Hemorrhage	1	(2%)	1	(2%)		
Inflammation, chronic active	_		1	(2%)	1	(2%)
Inflammation, suppurative	7	(13%)	5	(9%)	9	(18%)
Metaplasia, squamous	29	(56%)	17	(32%)	33	(66%)
Endometrium hyperplacia evictic	20	(590/)	10	(260/)	1	(2%)
	30	(38%)	19	(3076)	52	(0470)
Hematopoietic System						
Bone marrow	(53)		(53)		(50)	
Atrophy			1	(2%)	í	(2%)
Hyperplasia	36	(68%)	43	(81%)	36	(72%)
Myelofibrosis					1	(2%)
Lymph node	(2)		(9)		(5)	
Lumbar, ectasia	1	(50%)			2	(40%)
Lumbar, hyperplasia, plasma cell					2	(40%)
Mediastinal, ectasia			3	(33%)		
Mediastinal, hemorrhage			5	(56%)	1	(20%)
Mediastinal, hyperplasia, histocytic			1	(11%)		(200/)
Mediastinal, hyperplasia, plasma cell			1	(110/)	1	(20%)
Mediastinal, pigmentation	(51)		(52)	(11%)	(40)	
Congestion	(51)	(2%)	(32)		(49)	
Ectasia	1	(270)	6	(12%)	2	(4%)
Hyperplasia lymphoid			2	(4%)	2	(4%)
Hyperplasia, plasma cell	25	(49%)	16	(31%)	30	(61%)
Inflammation, suppurative		(,.)	1	(2%)		((()))
Lymph node, mesenteric	(52)		(51)		(49)	
Ectasia			1	(2%)		
Hyperplasia, histiocytic	1	(2%)	2	(4%)	1	(2%)
Spleen	(51)		(52)		(49)	
Hematopoietic cell proliferation	46	(90%)	43	(83%)	42	(86%)
Hyperplasia, lymphoid	1	(2%)	4	(8%)	1	(2%)
Pigmentation	45	(88%)	47	(90%)	42	(86%)
Lymphoid follicle, atrophy	4	(8%)	1	(2%)	1	(2%)
Red pulp, atrophy	(51)		(10)		1	(2%)
I nymus	(51)	(710/)	(42)	(1009/)	(49)	(02%)
Atrophy	30	(/1%)	42	(100%)	45	(92%)
Hemorrhage	1	(2%)	1	(270)		
Integumentary System						
Mammary gland	(53)		(53)		(50)	
Cyst	4	(8%)	1	(2%)	4	(8%)
Hyperplasia	25	(47%)	16	(30%)	19	(38%)
Inflammation, granulomatous	1	(2%)	1	(2%)	(50)	
SKIN Court an ithalial in aluai	(53)	(20/)	(53)	(40/)	(50)	
Cyst epitnelial inclusion	1	(2%)	2	(4%) (2%)		
Illianmation, enronic active			1	(2%)	1	(29/)
Enidermis inflammation			1	(2%)	1	(270)
			1	(270)		

#### TABLE A5c Summary of the Incidence of Nonneoplastic Lesions in Female Rats in the 2-Year Gavage Study of TCDD (Stop-Exposure)

	Vehicle	e Control	100	ng/kg	100 (Stop-F	ng/kg Exposure)	
Musculoskeletal System							
Bone Fracture	(53)		(53) 1	(2%)	(50)		
Nervous System							
Brain	(53)		(53)		(50)		
Hemorrhage					1	(2%)	
Inflammation, suppurative					1	(2%)	
Mineralization			2	(4%)			
Cerebellum, developmental malformation	1	(2%)					
Glial cell, hyperplasia			1	(2%)			
Respiratory System							
Lung	(53)		(52)		(50)		
Hemorrhage	· · · ·		2	(4%)	× ,		
Infiltration cellular, histiocyte	41	(77%)	50	(96%)	41	(82%)	
Inflammation, chronic active			1	(2%)			
Inflammation, granulomatous	2	(4%)					
Inflammation, suppurative	1	(2%)			1	(2%)	
Metaplasia, squamous	1	(2%)	6	(12%)	3	(6%)	
Mineralization	1	(2%)	1	(2%)			
Necrosis			1	(2%)			
Alveolar epithelium, hyperplasia	12	(23%)					
Alveolar epithelium, metaplasia, bronchiolar	2	(4%)	46	(88%)	31	(62%)	
Serosa, inflammation, suppurative					1	(2%)	
Nose	(53)		(53)		(50)		
Inflammation	2	(4%)	4	(8%)	1	(2%)	
Goblet cell, respiratory epithelium, hyperplasia					1	(2%)	
Nasolacrimal duct, inflammation	2	(4%)			1	(2%)	
Nasopharyngeal duct, inflammation			1	(2%)			
Olfactory epithelium, degeneration				(20)	1	(2%)	
Olfactory epithelium, inflammation			I	(2%)	1	(2%)	
Offactory epithelium, glands, hyperplasia					1	(2%)	
Respiratory epithelium, cyst					1	(2%)	
Sentum hymeralisis agreements			1	(20/)	1	(2%)	
Septum, hyperplasia, squamous			1	(2%)			
Septum, initialination Squamous enithelium inflammation suppurative	1	(2%)	2	(4%)			
Turbinate hyperplasia squamous	1	(270)	1	(2%)			
Turbinate, septum, inflammation			2	(4%)			
special Senses System	(50)		(50)				
Eye	(53)	(20/)	(53)		(50)		
Degeneration	1	(2%) (2%)					
Antarior chamber inflammation supportive	1	(270)	1	(20/2)			
Cornea inflammation supportive			1	(2%)			
Retinal detachment			1	(270)	1	(2%)	
Rilateral cataract					1	(2%)	
Retina atronhy	5	(9%)	6	(11%)	1	(12%)	
Retina, degeneration	5	(270)	0	(11/0)	1	(2%)	
, accontation					1	(=, *)	
# TABLE A5c Summary of the Incidence of Nonneoplastic Lesions in Female Rats in the 2-Year Gavage Study of TCDD (Stop-Exposure)

	Vehicle	e Control	100	ng/kg	100 (Stop-E	ng/kg xposure)	
Special Senses System (continued)							
Harderian gland	(53)		(53)		(50)		
Atrophy, focal	1	(2%)					
Hemorrhage		~ /			1	(2%)	
Inflammation	12	(23%)	6	(11%)	9	(18%)	
Urinary System							
Kidney	(53)		(53)		(50)		
Accumulation, hyaline droplet	( )		× ,		2	(4%)	
Calculus gross observation			1	(2%)			
Calculus microscopic observation only	1	(2%)					
Casts protein	3	(6%)	1	(2%)	1	(2%)	
Cyst	2	(4%)	1	(2%)			
Inflammation, chronic active	5	(9%)					
Inflammation, suppurative			2	(4%)			
Mineralization	40	(75%)	42	(79%)	42	(84%)	
Nephropathy	34	(64%)	52	(98%)	41	(82%)	
Papilla, transitional epithelium, hyperplasia					1	(2%)	
Pelvis, dilatation	2	(4%)			1	(2%)	
Pelvis, inflammation	2	(4%)	2	(4%)			
Renal tubule, degeneration			1	(2%)			
Transitional epithelium, hyperplasia	3	(6%)	11	(21%)	5	(10%)	
Ureter	(1)						
Inflammation	1	(100%)					
Transitional epithelium, hyperplasia	1	(100%)					
Urinary bladder	(52)		(53)		(50)		
Inflammation	6	(12%)	3	(6%)	2	(4%)	
Metaplasia, squamous			1	(2%)			
Transitional epithelium, hyperplasia	1	(2%)	2	(4%)			

# APPENDIX B GENETIC TOXICOLOGY

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# **GENETIC TOXICOLOGY**

## SALMONELLA TYPHIMURIUM MUTAGENICITY TEST PROTOCOL

Testing was performed as reported by Mortelmans *et al.* (1984). 2,3,7,8-Tetrachlorodibenzo-*p*-dioxin (TCDD) was sent to the laboratory as a coded aliquot from Radian Corporation (Austin, TX). It was incubated with the *Salmonella typhimurium* tester strains TA98, TA100, TA1535, and TA1537 either in buffer or S9 mix (metabolic activation enzymes and cofactors from Aroclor 1254-induced male Sprague-Dawley rat or Syrian hamster liver) for 20 minutes at 37° C. Top agar supplemented with L-histidine and d-biotin was added, and the contents of the tubes were mixed and poured onto the surfaces of minimal glucose agar plates. Histidine-independent mutant colonies arising on these plates were counted following incubation for 2 days at 37° C.

Each trial consisted of triplicate plates of concurrent positive and negative controls and five doses of TCDD. The high dose was limited by toxicity. All trials were repeated.

In this assay, a positive response is defined as a reproducible, dose-related increase in histidine-independent (revertant) colonies in any one strain/activation combination. An equivocal response is defined as an increase in revertants that is not dose related, is not reproducible, or is not of sufficient magnitude to support a determination of mutagenicity. A negative response is obtained when no increase in revertant colonies is observed following chemical treatment. There is no minimum percentage or fold increase required for a chemical to be judged positive or weakly positive.

## MOUSE LYMPHOMA MUTAGENICITY TEST PROTOCOL

The experimental protocol is presented in detail by McGregor *et al.* (1991). TCDD was supplied as a coded aliquot by Radian Corporation. The high dose of 1  $\mu$ g/mL was determined by toxicity. L5178Y mouse lymphoma cells were maintained at 37° C as suspension cultures in supplemented Fischer's medium; normal cycling time was approximately 10 hours. To reduce the number of spontaneously occurring cells resistant to trifluorothymidine (TFT), subcultures were exposed to medium containing thymidine, hypoxanthine, methotrexate, and glycine for 1 day; to medium containing thymidine, and glycine for 1 day; and to normal medium for 3 to 5 days. For cloning, the horse serum content was increased and Noble agar was added.

All treatment levels within an experiment, including concurrent positive and solvent controls, were replicated. Treated cultures contained  $6 \times 10^6$  cells in 10 mL medium. This volume included the S9 fraction in those experiments performed with metabolic activation. Incubation with TCDD continued for 4 hours, at which time the medium plus TCDD was removed, and the cells were resuspended in fresh medium and incubated for an additional 2 days to express the mutant phenotype. Cell density was monitored so that log phase growth was maintained. After the 48-hour expression period, cells were plated in medium and soft agar supplemented with TFT for selection of TFT-resistant cells, and cells were plated in nonselective medium and soft agar to determine cloning efficiency. Plates were incubated at 37° C in 5% CO<sub>2</sub> for 10 to 12 days. The test was initially performed without S9. Because a clearly positive response was not obtained, the test was repeated using freshly prepared S9 from the livers of Aroclor 1254-induced male F344 rats.

Minimum criteria for accepting an experiment as valid and a detailed description of the statistical analysis and data evaluation are presented by Caspary *et al.* (1988). All data were evaluated statistically for trend and peak responses. Both responses had to be significant ( $P \le 0.05$ ) for TCDD to be considered positive, i.e., capable of inducing TFT resistance. A single significant response led to a call of "questionable," and the absence of both a trend and peak response resulted in a "negative" call.

## **CHINESE HAMSTER OVARY CELL CYTOGENETICS PROTOCOLS**

Testing was performed as reported by Galloway *et al.* (1987). TCDD was sent to the laboratory as a coded aliquot by Radian Corporation. It was tested in cultured Chinese hamster ovary (CHO) cells for induction of sister chromatid exchanges (SCEs) and chromosomal aberrations (Abs), both in the presence and absence of Aroclor 1254-induced male Sprague-Dawley rat liver S9 and cofactor mix. Cultures were handled under gold lights to prevent photolysis of bromodeoxyuridine-substituted DNA. Each test consisted of concurrent solvent and positive controls and of three doses of TCDD; the high dose was limited by toxicity. A single flask per dose was used, and tests yielding equivocal or positive results were repeated.

*Sister Chromatid Exchange Test:* In the SCE test without S9, CHO cells were incubated for 25.5 hours with TCDD in supplemented McCoy's 5A medium. Bromodeoxyuridine (BrdU) was added 2 hours after culture initiation. After 25.5 hours, the medium containing TCDD was removed and replaced with fresh medium plus BrdU and Colcemid, and incubation was continued for 2 hours. Cells were then harvested by mitotic shake-off, fixed, and stained with Hoechst 33258 and Giemsa. In the SCE test with S9, cells were incubated with TCDD, serum-free medium, and S9 for 2 hours. The medium was then removed and replaced with medium containing serum and BrdU and no TCDD. Incubation proceeded for an additional 25.5 hours, with Colcemid present for the final 2 hours. Harvesting and staining were the same as for cells treated without S9. All slides were scored blind, and those from a single test were read by the same person. Fifty second-division metaphase cells were scored for frequency of SCEs/cell from each dose level. Because significant chemical-induced cell cycle delay was seen, incubation time was lengthened to ensure a sufficient number of scorable (second-division metaphase) cells.

Statistical analyses were conducted on the slopes of the dose-response curves and the individual dose points (Galloway *et al.*, 1987). An SCE frequency 20% above the concurrent solvent control value was chosen as a statistically conservative positive response. The probability of this level of difference occurring by chance at one dose point is less than 0.01; the probability for such a chance occurrence at two dose points is less than 0.001. An increase of 20% or greater at any single dose was considered weak evidence of activity; increases at two or more doses resulted in a determination that the trial was positive. A statistically significant trend (P<0.005) in the absence of any responses reaching 20% above background led to a call of equivocal.

*Chromosomal Aberrations Test:* In the Abs test without S9, cells were incubated in McCoy's 5A medium with TCDD for 19.5 hours; Colcemid was added and incubation continued for 2 hours. The cells were then harvested by mitotic shake-off, fixed, and stained with Giemsa. For the Abs test with S9, cells were treated with TCDD and S9 for 2 hours, after which the treatment medium was removed and the cells were incubated for 8.5 hours in fresh medium, with Colcemid present for the final 2 hours. Cells were harvested in the same manner as for the treatment without S9.

Cells were selected for scoring on the basis of good morphology and completeness of karyotype  $(21 \pm 2 \text{ chromosomes})$ . All slides were scored blind, and those from a single test were read by the same person. One hundred first-division metaphase cells were scored at each dose level. Classes of aberrations included simple (breaks and terminal deletions), complex (rearrangements and translocations), and other (pulverized cells, despiralized chromosomes, and cells containing 10 or more aberrations).

Chromosomal aberration data are presented as percentage of cells with aberrations. To arrive at a statistical call for a trial, analyses were conducted on both the dose response curve and individual dose points. For a single trial, a statistically significant ( $P \le 0.05$ ) difference for one dose point and a significant trend ( $P \le 0.015$ ) were considered weak evidence for a positive response; significant differences for two or more doses indicated the trial was positive. A positive trend test in the absence of a statistically significant increase at any one dose resulted in an equivocal call (Galloway *et al.*, 1987). Ultimately, the trial calls were based on a consideration of the statistical analyses as well as the biological information available to the reviewers.

## **DROSOPHILA MELANOGASTER TEST PROTOCOL**

The assay for induction of sex-linked recessive lethal (SLRL) mutations was performed with adult flies as described by Zimmering *et al.* (1985). TCDD was supplied as a coded aliquot by Radian Corporation. TCDD was assayed in the SLRL test by injection into adult Canton-S wild-type males 24 to 72 hours old at the beginning of treatment.

To administer TCDD by injection, a glass Pasteur pipette was drawn out in a flame to a microfine filament, and the tip was broken off to allow delivery of the test solution. Injection was performed either manually, by attaching a rubber bulb to the other end of the pipette and forcing through sufficient solution (0.2 to  $0.3 \,\mu$ L) to slightly distend the abdomen of the fly, or by attaching the pipette to a microinjector that automatically delivered a calibrated volume. Flies were anesthetized with ether and immobilized on a strip of tape. Injection into the thorax, under the wing, was performed with the aid of a dissecting microscope.

Toxicity tests were performed to set concentrations of TCDD at a level that would induce 30% mortality 24 hours after injection, while keeping induced sterility at an acceptable level. Canton-S males were treated with a solution of TCDD dissolved in peanut oil and allowed to recover for 24 hours. A concurrent peanut oil control group was also included. Treated males were mated to three *Basc* females for 3 days and were given fresh females at 2-day intervals to produce three matings of 3, 2, and 2 days (in each case, sample sperm from successive matings was treated at successively earlier postmeiotic stages).  $F_1$  heterozygous females were mated with their siblings and then placed in individual vials.  $F_1$  daughters from the same parental male were kept together to identify clusters. (A cluster occurs when a number of mutants from a given male result from a single spontaneous premeiotic mutation event and is identified when the number of mutants from that male exceeds the number predicted by a Poisson distribution.) If a cluster was identified, all data from the male in question were discarded. Presumptive lethal mutations were identified as vials containing fewer than 5% of the expected number of wild-type males after 17 days; these were retested to confirm the response.

SLRL data were analyzed by simultaneous comparison with the concurrent and historical controls (Mason *et al.*, 1992) using a normal approximation to the binomial test (Margolin *et al.*, 1983). A test result was considered positive if the P value was less than or equal to 0.01 and the mutation frequency in the tested group was greater than 0.10% or if the P value was less than or equal to 0.05 and the frequency in the treatment group was greater than 0.15%. A test was considered to be inconclusive if the P value was between 0.05 and 0.01 but the frequency in the treatment group was between 0.10% and 0.15% or if the P value was between 0.10 and 0.05 but the frequency in the treatment group was greater than 0.10%. A test was considered negative if the P value was greater than 0.10% or if the frequency in the treatment group was greater than 0.10%. A test was considered negative if the P value was greater than 0.10% or if the frequency in the treatment group was greater than 0.10%. A test was considered negative if the P value was greater than 0.10% or if the P value was between 0.10 and 0.05 but the frequency in the treatment group was greater than 0.10%. A test was considered negative if the P value was greater than 0.100%.

## **MOUSE BONE MARROW CYTOGENETICS TEST PROTOCOL**

A dose range-finding study was performed along with an analysis of published  $LD_{50}$  information to set the doses used in the study. The highest dose was 1,000 µg/kg. TCDD was tested for induction of Abs in mouse bone marrow by two different protocols. The first protocol used a standard harvest time of 17 hours, and the second protocol used a delayed harvest time of 36 hours.

Male  $B6C3F_1$  mice (10 animals per dose group) were injected intraperitoneally with TCDD dissolved in corn oil. Solvent control animals received injections of corn oil only. The positive control was dimethylbenzanthracene. The mice were subcutaneously implanted with a BrdU tablet (McFee *et al.*, 1983) 18 hours before the scheduled harvest. (For the standard protocol, this required BrdU implantation to precede injection with TCDD by 1 hour.) The use of BrdU allowed selection of the appropriate cell population for scoring. (Abs induced by chemical administration are present in maximum number at the first metaphase following treatment; they decline in number during subsequent nuclear divisions due to cell death.) Two hours before sacrifice, the animals received an intraperitoneal injection of colchicine in saline. The animals were killed 17 or 36 hours after TCDD injection (18 hours after BrdU dosing). One or both femurs were removed, and the marrow was flushed out with phosphatebuffered saline (pH 7.0). Cells were treated with a hypotonic salt solution, fixed, and dropped onto chilled slides. After a 24-hour drying period, the slides were stained (with fluorescence-plus-Giemsa) and scored.

Fifty first-division metaphase cells were scored from up to eight animals per treatment group. Responses were evaluated as the percentage of aberrant metaphase cells, excluding gaps. The data were analyzed by a trend test (Margolin *et al.*, 1986).

## **EVALUATION PROTOCOL**

These are the basic guidelines for arriving at an overall assay result for assays performed by the National Toxicology Program. Statistical as well as biological factors are considered. For an individual assay, the statistical procedures for data analysis have been described in the preceding protocols. There have been instances, however, in which multiple aliquots of a chemical were tested in the same assay, and different results were obtained among aliquots and/or among laboratories. Results from more than one aliquot or from more than one laboratory are not simply combined into an overall result. Rather, all the data are critically evaluated, particularly with regard to pertinent protocol variations, in determining the weight of evidence for an overall conclusion of chemical activity in an assay. In addition to multiple aliquots, the *in vitro* assays have another variable that must be considered in arriving at an overall test result. *In vitro* assays are conducted with and without exogenous metabolic activation. Results obtained in the absence of activation are not combined with results obtained in the presence of activation; each testing condition is evaluated separately. The summary table in the Abstract of this Technical Report presents a result that represents a scientific judgement of the overall evidence for activity of the chemical in an assay.

### RESULTS

TCDD (0.1 to 1,000 µg/plate) was not mutagenic in S. typhimurium strains TA98, TA100, TA1535, or TA1537, with or without hamster or rat liver metabolic activation enzymes (S9 fraction) (Table B1; Mortelmans et al., 1984). No induction of TFT resistance was noted in L5178Y tk<sup>+/-</sup> mouse lymphoma cells treated with 0.0625 to 1 µg/mL TCDD with or without S9 (Table B2; McGregor et al., 1991). In tests for chromosomal damage in cultured CHO cells, TCDD did not induce SCEs or Abs when tested up to toxic levels (0.8 µg/mL) with or without S9 activation enzymes (Tables B3 and B4; Galloway et al., 1987). Without S9, TCDD induced significant increases in SCEs at concentrations of 0.6 to 0.8 µg/mL, in two of five independent trials when a prolonged culture time was employed to compensate for TCDD-induced cell cycle delay. However, because the response was not consistently reproducible, the SCE test was concluded to be negative overall. In addition, there was a complication from the ethanol solvent in these SCE experiments. A comparison of a true negative control culture and an ethanol solvent control culture without TCDD indicated that ethanol alone, in the amount needed to dissolve TCDD, increased SCE frequencies. No induction of SLRL mutations was observed in germ cells of male D. melanogaster following treatment with 50, 250, or 500 ppm TCDD (Table B5; Zimmering et al., 1985). Finally, no induction of Abs was observed in bone marrow cells of male B6C3F, mice 17 or 36 hours after intraperitoneal injection of 250, 500, or 1,000 µg/kg TCDD (Table B6). In summary, no mutagenic activity was detected with TCDD in a variety of in vitro and in vivo short-term tests.

				Reverta	nts/Plate <sup>b</sup>		
Strain	Dose	-8	9	+10% ł	amster S9	+10%	rat S9
	(µg/plate)	Trial 1	Trial 2	Trial 1	Trial 2	Trial 1	Trial 2
TA100	0	119 ± 4.5	141 ± 10.1	$98 \pm 6.9$	155 ± 12.8	$86 \pm 2.8$	154 ± 9.0
	0.1	$93 \pm 10.0$		$96 \pm 18.2$		$101 \pm 8.7$	
	1	$83 \pm 8.1$		$94 \pm 12.5$		$96 \pm 4.8$	
	10	$93 \pm 5.5$	$170\pm14.9$	$94 \pm 4.7$	$117\pm12.3$	$96 \pm 8.6$	$188 \pm 7.3$
	33	$85\pm7.0$	$178\pm8.6$	$78\pm5.5$	$117 \pm 7.5$	$84 \pm 2.0$	$173 \pm 2.5$
	100	$98 \pm 5.5$	$162 \pm 5.7$	$103 \pm 7.3$	$111 \pm 0.6$	$94 \pm 3.4$	$195 \pm 5.0$
	333		$176\pm0.9$		$124 \pm 12.4$		$155 \pm 5.5$
	1,000		$149\pm7.8$		$116\pm9.4$		$177 \pm 6.8$
Trial sun	nmary	Negative	Negative	Negative	Negative	Negative	Negative
Positive	control	$527 \pm 7.4$	$374 \pm 21.2$	$1,449 \pm 87.8$	$1,527 \pm 66.5$	867 ± 18.7	$853 \pm 45.0$
TA1535	5 0	$16 \pm 2.1$	$13 \pm 1.3$	$12\pm1.8$	$5\pm1.0$	$15\pm1.7$	$8 \pm 1.0$
	0.1	$18 \pm 2.2$		$6 \pm 0.3$		$8 \pm 0.6$	
	1	$20 \pm 3.5$		$7 \pm 0.6$		$13 \pm 3.2$	
	10	$20 \pm 2.9$	$15 \pm 3.5$	$6 \pm 1.2$	$8\pm0.7$	$8 \pm 0.7$	$11 \pm 2.5$
	33	$17 \pm 1.2$	$17 \pm 0.6$	$8 \pm 1.2$	$11 \pm 1.2$	$7 \pm 0.7$	$11 \pm 3.7$
	100	$18 \pm 2.3$	$17 \pm 0.7$	$9 \pm 2.0$	$8 \pm 3.4$	$6 \pm 0.9$	$8 \pm 3.4$
	333		$18 \pm 0.6$		$9 \pm 3.2$		$7 \pm 1.2$
	1,000		$18 \pm 3.9$		8 ± 2.3		$8 \pm 2.0$
Trial sun Positive	nmary control	Negative $494 \pm 14.6$	Negative $308 \pm 17.0$	Negative $265 \pm 12.7$	Negative $242 \pm 12.2$	Negative $417 \pm 26.1$	Negative $121 \pm 14.4$
	_						
TA1537	7 0	$15 \pm 2.5$	$5 \pm 1.3$	$19 \pm 1.5$	$5 \pm 0.0$	$15 \pm 1.7$	$9 \pm 1.7$
	0.1	$11 \pm 2.3$		$23 \pm 1.7$		$14 \pm 1.2$	
	1	$11 \pm 1.2$		$14 \pm 3.0$		$16 \pm 6.4$	
	10	$10 \pm 1.8$	$5 \pm 0.3$	$17 \pm 1.8$	$5 \pm 1.5$	$10 \pm 2.3$	$7 \pm 0.9$
	33	$13 \pm 3.1$	$6\pm0.0$	$18 \pm 1.2$	$4 \pm 0.6$	$11 \pm 1.5$	$11 \pm 1.0$
	100	$10 \pm 2.5$	$7 \pm 0.6$	$13 \pm 2.4$	$6 \pm 3.0$	$15 \pm 1.5$	$4 \pm 1.2$
	333		$5 \pm 0.7$		$8 \pm 0.5$		$8 \pm 1.0$ 7 + 1.2
	1,000		$7 \pm 2.0$		/ ± 1.5		/ ± 1.2
Trial sun	nmary	Negative	Negative	Negative	Negative	Negative	Negative
Positive	control	$147 \pm 7.5$	$122 \pm 11.1$	$383 \pm 63.4$	$413 \pm 22.8$	$441 \pm 26.9$	$164 \pm 4.2$
TA98	0	$21\pm1.7$	$19\pm3.5$	$33\pm4.7$	$21\pm5.2$	$46\pm7.1$	$24\pm2.1$
	0.1	$22 \pm 2.5$		$27 \pm 6.7$		$35\pm4.8$	
	1	$18 \pm 1.5$		$26\pm0.9$		$26 \pm 0.3$	
	10	$16\pm0.0$	$22 \pm 3.3$	$27 \pm 1.8$	$23\pm3.6$	$21 \pm 2.2$	$27 \pm 3.5$
	33	$20 \pm 3.5$	$20 \pm 4.6$	$29 \pm 2.7$	$28 \pm 2.0$	$22 \pm 3.2$	$29 \pm 2.7$
	100	$16 \pm 1.9$	$18 \pm 3.7$	$27 \pm 2.7$	$24 \pm 3.5$	$29 \pm 4.0$	$26 \pm 5.8$
	333		$18 \pm 3.1$		$23 \pm 2.2$		$21 \pm 1.5$
	1,000		$18 \pm 3.1$		$18 \pm 2.8$		$26 \pm 0.6$
Trial sun	nmary	Negative	Negative	Negative	Negative	Negative	Negative
Positive	control	$623 \pm 85.1$	$725 \pm 32.2$	$1,184 \pm 18.8$	$1,204 \pm 10.8$	$636 \pm 80.3$	$384 \pm 22.8$

#### TABLE B1 Mutagenicity of TCDD in Salmonella typhimurium<sup>a</sup>

а Study was performed at SRI International. The detailed protocol and these data are presented by Mortelmans et al. (1984). b

c

Revertants are presented as mean  $\pm$  standard error from three plates. The positive controls in the absence of metabolic activation were sodium azide (TA100 and TA1535), 9-aminoacridine (TA1537), and 4-nitro-o-phenylenediamine (TA98). The positive control for metabolic activation with all strains was 2-aminoanthracene.

Induction of Trifluorothymidine Resistance in L5178Y Mouse Lymphoma Cells by TCDD<sup>a</sup>

Compound	Concentration (µg/mL)	Cloning Efficiency (%)	Relative Total Growth (%)	Mutant Count	Mutant Fraction <sup>b</sup>	Average Mutant Fraction
-89						
Trial 1						
Dimethylsulfoxi	de <sup>c</sup>	77	94	82	35	
Dimenijiounom		67	98	68	34	
		62	90	112	60	
		76	118	98	43	43
TCDD	0.0625	76	110	108	47	
		85	118	80	31	39
	0.125	64	101	72	38	
		64	112	56	29	33
	0.25	65	108	60	31	
		80	104	71	30	30
	0.5	71	105	61	29	
		76	98	58	26	27
	1	70	90	58	28	
		86	101	70	27	27
Methyl methanes	sulfonate <sup>d</sup>					
, , , , , , , , , , , , , , , , , , ,	15	46	45	176	129	
		47	34	208	148	
Trial 2						
Dimethylsulfoxi	de	86	114	48	19	
		76	98	44	19	
		72	99	79	36	
		89	88	60	23	24
TCDD	0.0625	80	110	49	20	
		75	119	53	23	22
	0.125	42	54	47	37	
		78	120	72	31	34
	0.25	63	83	81	43	
		80	176	56	23	33
	0.5	75	98	61	27	
		90	127	49	18	23
	1	98	88	98	33	
		97	115	70	24	29
Methyl methanes	sulfonate					
	15	60	43	217	122	
		51	41	190	124	23

Induction of Trifluorothymidine Resistance in L5178Y Mouse Lymphoma Cells by TCDD

Compound	Concentration (μg/mL)	Cloning Efficiency (%)	Relative Total Growth (%)	Mutant Count	Mutant Fraction <sup>b</sup>	Average Mutant Fraction
+\$9						
Trial 1						
Dimethylsulfoxide		83	90	80	32	
		75	96	46	20	
		87	103	98	37	20
		85	111	80	31	30
TCDD						
1000	0.0625	88	92	132	50	
	010020	92	87	106	39	44
	0.125	86	86	123	47	
		81	79	119	49	48
	0.25	92	85	110	40	
		99	86	104	35	37
	0.5	83	90	105	42	
		102	101	90	30	36
	1	87	93	97	37	
		103	96	112	36	37
<b>X</b> A A A A A	daa	-	72	227	1.45	
Methylcholanthrene	2.5	/8	73	337	145	10
		93	/3	3/3	134	19
Trial 2						
Dimethylsulfoxide		88	103	82	31	
·	0	70	96	51	24	
		86	109	67	26	
		84	91	52	21	26
ТСОО	0.0625	101	110	89	29	
TODD	0.0025	76	110	71	31	30
	0.125	88	117	79	30	50
	0.125	82	123	64	26	28
	0.25	71	105	58	27	
		96	150	107	37	32
	0.5	92	114	100	36	
		75	107	81	36	36
	1	91	126	79	29	
		94	109	79	28	29
Methylcholanthrene	2.5	81	97	339	139	
		92	98	330	120	10

а Study was performed at Inveresk Research International. The detailed protocol and these data are presented by McGregor *et al.* (1991). Mutant fraction=mutant cells/ $10^6$  clonable cells

b

с Solvent control d

Positive control

Induction of Sister Chromatid Exchanges in Chinese Hamster Ovary Cells by TCDD<sup>a</sup>

Compound	Dose (µg/mL)	Total Cells Scored	No. of Chromosomes	No. of SCEs	SCEs/ Chromosomes	SCEs/ Cell	Hrs in BrdU	Relative Change of SCEs/ Chromosome <sup>b</sup> (%)
-S9 Trial 1 Summary: Nega	utive							
Ethanol <sup>c</sup>		50	1,031	422	0.41	8.44	25.5	
TCDD	0.04 0.133 0.4	50 50 50	1,059 1,067 1,047	432 454 402	0.41 0.43 0.38	8.64 9.08 8.04	25.5 25.5 25.5	-0.34 3.95 -6.20
Mitomycin-C <sup>e</sup>	0.005	25	520	722	P=0.738 1.39	28.88	25.5	239.22
Trial 2 Summary: Posit	ive							
Ethanol		50	1,053	385	0.37	7.70	25.5	
TCDD	0.4 0.6 0.6 0.8	50 50 50 50	1,045 1,033 1,045 1,045	430 429 508 563	0.41 0.42 0.49 0.54 P=0.000	8.60 8.58 10.16 11.26	25.5 25.5 36.3 <sup>f</sup> 36.3 <sup>f</sup>	12.84 13.59 32.96* 47.35*
Mitomycin-C	0.005	25	524	762	1.45	30.48	25.5	297.73
Trial 3 Summary: Nega	ıtive							
None Ethanol		50 50	1,059 1,052	400 527	0.38 0.50	8.00 10.54	$\begin{array}{c} 25.8\\ 32.5^{\mathrm{f}} \end{array}$	
TCDD	0.6 0.7 0.8	50 50 50	1,048 1,059 1,045	513 537 597	0.49 0.51 0.57 P=0.012	10.26 10.74 11.94	$32.5^{f}$ $32.5^{f}$ $32.5^{f}$	-2.29 1.22 14.04
Mitomycin-C	0.005	25	517	705	1.36	28.20	25.8	172.21

Induction of Sister Chromatid Exchanges in Chinese Hamster Ovary Cells by TCDD

Compound (	Dose µg/mL)	Total Cells Scored	No. of Chromosomes	No. of SCEs	SCEs/ Chromosomes	SCEs/ Cell	Hrs in BrdU	Relative Change of SCEs/ Chromosome (%)
+89								
Trial 1 Summary: Negativ	e							
Summary. Regativ	C							
Ethanol		50	1,038	625	0.60	12.50	25.5	
TCDD								
	0.04	50	1,034	580	0.56	11.60	25.5	-6.84
	0.133	50	1,045	552	0.53	11.04	25.5	-12.27
	0.4	50	1,042	582	0.56	11.64	25.5	-7.24
					P=0.944			
Cyclophosphamide	<sup>e</sup> 1.5	25	523	897	1.72	35.88	25.5	184.85
<b>Trial 2</b> Summary: Negativ	e							
Ethanol		50	1,043	534	0.51	10.68	25.5	
TCDD								
	0.4	50	1,037	525	0.51	10.50	25.5	-1.12
	0.6	50	1,035	569	0.55	11.38	25.5	7.38
	0.8	50	1,045	530	0.51	10.60	25.5	-0.94
					P=0.350			
Cyclophosphamide	1.5	25	526	834	1.59	33.36	25.5	209.69

\* Positive response (≥20% increase over solvent control)
 \* Study was performed at Litton Bionetics, Inc. The detailed protocol and these data are presented by Galloway *et al.* (1987).

b

SCE=sister chromatid exchange; BrdU=bromodeoxyuridine. SCEs/chromosome in treated cells versus SCEs/chromosome in solvent control cells

c Solvent control d

Significance of SCEs/chromosome tested by the linear regression trend test versus log of the dose

e Positive control  $\mathbf{f}$ 

Due to cell cycle delay, harvest time was extended to maximize the number of second-division metaphase cells available for analysis.

Induction of Chromosomal Aberrations in Chinese Hamster Ovary Cells by TCDD<sup>a</sup>

			<b>Total Aberrations</b>			
	Dose	<b>Total Cells</b>	Number	Aberrations/	Cells with	
Compound	(µg/mL)	Scored	of Aberrations	Cell	Aberrations (%)	
- <b>S9</b> Harvest time: 21.5 hours Summary: Negative						
Ethanol <sup>b</sup>		100 100	0 4	0.00 0.04	0.0 4.0	
TCDD	0.6 0.7 0.8	100 100 100	2 1 2	0.02 0.01 0.02	2.0 1.0 2.0	
					P=0.859 <sup>c</sup>	
Mitomycin-C <sup>d</sup>	0.062	50	14	0.28	22.0	
			Con	nplex Aberrati	ons	
Ethanol		100 100	0 2	0.00 0.02	0.0 2.0	
TCDD	0.6 0.7 0.8	100 100 100	0 1 2	0.00 0.01 0.02	0.0 1.0 2.0	
					P=0.424	
Mitomycin-C	0.062	50	7	0.14	14.0	
			Sin	nple Aberratio	ns	
Ethanol		100 100	0 2	0.00 0.02	0.0 2.0	
TCDD	0.6 0.7 0.8	100 100 100	1 0 0	0.01 0.00 0.00	1.0 0.0 0.0	
					P=0.966	
Mitomycin-C	0.062	50	7	0.14	10.0	
			Ot	her Aberration	18	
Ethanol		100 100	0 0		0.0 0.0	
TCDD	0.6 0.7 0.8	100 100 100	1 0 0		1.0 0.0 0.0	
Mitomycin-C	0.062	50	0		0.0	

Induction of Chromosomal Aberrations in Chinese Hamster Ovary Cells by TCDD

			Total Aberrations			
Compound	Dose (µg/mL)	Total Cells Scored	Number of Aberrations	Aberrations/ Cell	Cells with Aberrations (%)	
+ <b>S9</b> Harvest time: 10.5 hours Summary: Negative						
Ethanol		100 100	5 3	0.05 0.03	5.0 3.0	
TCDD	0.4 0.6 0.8	100 100 100	5 0 6	0.05 0.00 0.06	5.0 0.0 5.0	
Cyclophosphamida <sup>d</sup>	50	50	13	0.26	P=0.505	
Cyclophosphannae	50	50	15	0.20	22.0	
				mplex Aberratio	0118	
Ethanol		100 100	3 1	0.03 0.01	3.0 1.0	
TCDD	0.4 0.6 0.8	100 100 100	1 0 1	$0.01 \\ 0.00 \\ 0.01$	1.0 0.0 1.0	
					P=0.625	
Cyclophosphamide	50	50	6	0.12	10.0	
			S	imple Aberration	18	
Ethanol		100 100	0 1	0.00 0.01	0.0 1.0	
TCDD	0.4 0.6 0.8	100 100 100	4 0 3	0.04 0.00 0.03	4.0 0.0 2.0	
					P=0.605	
Cyclophosphamide	50	50	7	0.14	12.0	
			(	Other Aberration	15	
Ethanol		100 100	2 1		2.0 1.0	
TCDD	0.4 0.6 0.8	100 100 100	0 0 2		0.0 0.0 2.0	
Cyclophosphamide	50	50	0		0.0	

Study was performed at Litton Bionetics, Inc. The detailed protocol and these data are presented by Galloway *et al.* (1987). Solvent control Significance of percent cells with aberrations tested by the linear regression trend test versus log of the dose Positive control а

b

c d

Route of	Dose	Incidence of	No. of Lethals/N	o. of X Chroi	nosomes Tested	
Exposure	(ppm)	Sterility (%)	Mating 1	Mating 2	Mating 3	Total <sup>b</sup>
Injection	50	1	1/5,345	2/2,845	2/1,328	5/9,518 (0.05%)
-	$0^{c}$		3/3,592	0/2,258	1/1,371	4/7,221 (0.06%)
	250	27	0/283	0/43	0/0	0/326 (0.00%)
	0		0/227	0/124	0/82	0/433 (0.00%)
	500	35	0/606	0/54	0/107	0/767 (0.00%)
	0		2/1,762	0/457	0/200	2/2,419 (0.08%)
						P=0.502 <sup>d</sup>

TABLE B5	
nduction of Sex-Linked Recessive Lethal Mutations in Drosophila melanogaster by TC	<b>DD</b> <sup>a</sup>

а Study was performed at Bowling Green State University. The detailed protocol and these data are presented by Zimmering *et al.* (1985). The mean mutant frequency from 518 negative control experiments is 0.074% (Mason *et al.*, 1992). b

Total number of lethal mutations/number of X chromosomes tested for three mating trials

с Data were corrected for the occurrence of spontaneous clusters. d

Significance of total number of lethal mutations/total number of X chromosomes tested by a normal approximation to the binomial test (Margolin et al., 1983).

Compound	Dose	Number of Mice with Erythrocytes Scored	Aberrations/Cell <sup>b</sup>	P Value <sup>c</sup>
Trial 1 (17 hours)				
Corn oil <sup>d</sup>		8	$4.75\pm0.84$	
TCDD (µg/kg)	250 500 1,000	8 7 8	$8.50 \pm 1.12$ 10.00 ± 1.57 7.75 ± 1.44	0.0165 0.0028 0.0398
Dimethylbenzanthrac	tene (mg/kg) <sup>f</sup> 100	8	P=0.094 <sup>e</sup> 13.00 ± 0.85	0.0000
Trial 2 (17 hours)	)			
Corn oil		8	$1.25 \pm 0.53$	
TCDD (µg/kg)	250 500 1,000	8 8 8	$2.75 \pm 0.65$ $2.50 \pm 0.73$ $2.25 \pm 0.70$ P=0.253	0.0649 0.0962 0.1404
Dimethylbenzanthrac	cene (mg/kg) 100	8	8.25 ± 1.67	0.0000
Trial 1 (36 hours)	1			
Corn oil		8	$2.25\pm0.88$	
TCDD (µg/kg)	250 500 1,000	8 8 8	$\begin{array}{c} 1.75 \pm 0.70 \\ 0.50 \pm 0.33 \\ 1.25 \pm 0.37 \end{array}$ P=0.906	0.6962 0.9832 0.8596
Dimethylbenzanthrac	cene (mg/kg) 50	8	$28.25 \pm 2.19$	0.0000

# TABLE B6Induction of Chromosomal Aberrations in Bone Marrow Cells of Male Mice Treatedwith TCDD by Intraperitoneal Injection<sup>a</sup>

<sup>a</sup> Study was performed at Oak Ridge Associated Universities. The detailed protocol is presented by McFee *et al.* (1983).

<sup>b</sup> Mean  $\pm$  standard error

<sup>c</sup> Pairwise comparison with the vehicle control. Dosed group values are significant at P $\leq$ 0.008; positive control values are significant at P $\leq$ 0.005 (Margolin *et al.*, 1986)

d Vehicle control

<sup>e</sup> Significance tested by the one-tailed trend test; significant at P $\leq$ 0.025 (Margolin *et al.*, 1986)

f Positive control

# APPENDIX C ORGAN WEIGHTS AND ORGAN-WEIGHT-TO-BODY-WEIGHT RATIOS

TABLE C1	Organ Weights and Organ-Weight-to-Body-Weight Ratios for Female Rats	
	at the 14-, 31-, and 53-Week Interim Evaluations in the 2-Year Gavage Study	
	of TCDD	188

	Vehicle Control	3 ng/kg	10 ng/kg	22 ng/kg	46 ng/kg	100 ng/kg
n						
Week 14	10	10	10	10	10	10
Week 31	10	10	10	10	10	10
Week 53	8	8	8	8	8	8
Necropsy body wt						
Week 14	$270 \pm 5$	$266 \pm 5$	$283 \pm 7$	$280 \pm 9$	$269 \pm 6$	$256 \pm 6$
Week 31	$307 \pm 7$	$300 \pm 7$	$298 \pm 5$	$290 \pm 7$	$294 \pm 7$	$274 \pm 6**$
Week 53	$338 \pm 6$	$333 \pm 6$	$343\pm16$	$322 \pm 12$	$301 \pm 8*$	$288 \pm 10**$
L. Kidney						
Week 14						
Absolute	$0.761 \pm 0.016$	$0.800 \pm 0.015$	$0.790 \pm 0.028$	$0.764\pm0.020$	$0.758 \pm 0.027$	$0.753 \pm 0.019$
Relative	$2.823\pm0.046$	$3.014 \pm 0.052$	$2.793 \pm 0.065$	$2.734\pm0.052$	$2.811 \pm 0.060$	$2.944 \pm 0.051$
Week 31						
Absolute	$0.862 \pm 0.031$	$0.906 \pm 0.019$	$0.874\pm0.020$	$0.854\pm0.033$	$0.842 \pm 0.021$	$0.819 \pm 0.021$
Relative	$2.802 \pm 0.068$	$3.030 \pm 0.066 *$	$2.939 \pm 0.048$	$2.939\pm0.070$	$2.866 \pm 0.039$	$2.988 \pm 0.044$
Week 53						
Absolute	$1.004\pm0.021$	$0.997\pm0.030$	$1.025 \pm 0.043$	$0.983 \pm 0.038$	$0.952 \pm 0.021$	$0.931 \pm 0.027$
Relative	$2.971\pm0.070$	$2.991\pm0.076$	$3.003\pm0.100$	$3.052\pm0.016$	$3.173\pm0.066$	$3.248 \pm 0.083*$
Liver						
Week 14						
Absolute	$7.936 \pm 0.304$	$9.261 \pm 0.302*$	$10.277 \pm 0.399 **$	$9.806 \pm 0.514$ **	$10.179 \pm 0.296 **$	$10.315 \pm 0.361 **$
Relative Week 31	$29.344 \pm 0.64$	$34.836 \pm 0.858 **$	$36.240 \pm 0.738 **$	$34.856 \pm 0.837 **$	$37.766 \pm 0.465 **$	$40.273 \pm 0.979 **$
Absolute	$0.188 \pm 0.277b$	$10.543 \pm 0.345**$	$10.840 \pm 0.206**$	$10.630 \pm 0.357 **$	$11.561 \pm 0.386**$	$10.070 \pm 0.238 **$
Absolute D. 1. C	$9.100 \pm 0.277^{\circ}$	$10.343 \pm 0.343$	$10.040 \pm 0.290$	$10.039 \pm 0.337$	$11.301 \pm 0.380$	$10.379 \pm 0.238$
Kelative	$30.1/0 \pm 0.415^{\circ}$	$35.131 \pm 0.541$ **	$36.411 \pm 0.713$ **	$36.619 \pm 0.55 / **$	$39.265 \pm 0.633$ **	$40.11 / \pm 0.641 **$
Week 55	$10.760 \pm 0.206$	$10.075 \pm 0.212$	$12.117 \pm 1.060$	12 269 1 0 592	12 246 + 0 429	12 010 + 0 450*
Pelative	$10.700 \pm 0.290$ 31.815 ± 0.742	$10.975 \pm 0.315$ 32.032 ± 0.760	$12.117 \pm 1.000$ $34.006 \pm 1.570*$	$12.208 \pm 0.383$ 38.030 ± 0.801**	$12.340 \pm 0.428$ $41.051 \pm 0.724 **$	$12.910 \pm 0.430^{\circ}$ $44.020 \pm 0.861**$
Kelative	$51.815 \pm 0.742$	$52.952 \pm 0.709$	54.900 ± 1.570	58.050 ± 0.801	41.031 ± 0.724	44.929 ± 0.801
Lung						
week 14	1 (42 + 0.040	$1.9(1 \pm 0.072)$	1 005 + 0 004**	1 790 + 0.046	1 077 + 0 072*	1 747 + 0 040
Absolute	$1.043 \pm 0.040$	$1.801 \pm 0.072$	$1.995 \pm 0.084^{**}$	$1.789 \pm 0.046$	$1.8//\pm 0.0/2^{*}$	$1./4/\pm 0.040$
Week 21	$0.092 \pm 0.111$	$7.014 \pm 0.270$	/.10/ ± 0.384	$0.409 \pm 0.147$	$7.013 \pm 0.331$	$0.849 \pm 0.187$
Absolute	$1.803 \pm 0.065$	$2325 \pm 0.125$	$2.378 \pm 0.152*$	$1.940 \pm 0.100$	$2.236 \pm 0.140$	$2300 \pm 0.117$
Relative	$1.893 \pm 0.003$ $6.194 \pm 0.278$	$2.325 \pm 0.125$ 7 789 ± 0.440*	$2.378 \pm 0.132$ 7 998 + 0.497*	$1.940 \pm 0.100$ 6 674 ± 0.270*	$2.230 \pm 0.149$ 7 584 + 0.438*	$2.309 \pm 0.117$ 8 415 + 0 358**
Week 53	$0.174 \pm 0.278$	7.769 ± 0.440	1.))0 ± 0.4)1	$0.074 \pm 0.270$	7.564 ± 0.456	0.415 ± 0.556
Absolute	$2\ 104 \pm 0\ 146$	$1.943 \pm 0.080$	$1.991 \pm 0.088$	$2.057 \pm 0.106$	$2.151 \pm 0.188$	$2451 \pm 0110$
Relative	$6.243 \pm 0.469$	$5.851 \pm 0.287$	$5.865 \pm 0.326$	$6.397 \pm 0.254$	$7.104 \pm 0.483$	$8.526 \pm 0.280^{**}$
Ovaries						
Week 14						
Absolute	$0.054 \pm 0.004$	$0.053 \pm 0.002^{b}$	$0.061 \pm 0.003$	$0.059 \pm 0.005$	$0.053 \pm 0.003$	$0.054 \pm 0.004$
Relative	$0.200 \pm 0.015$	0.204 + 0.002	$0.217 \pm 0.011$	$0.208 \pm 0.015$	$0.196 \pm 0.007$	$0.211 \pm 0.016$
Week 31	0.200 ± 0.013	$0.207 \pm 0.000^{\circ}$	$0.217 \pm 0.011$	0.200 ± 0.015	0.170 ± 0.007	0.211 ± 0.010
Absolute	$0.047\pm0.003$	$0.055\pm0.003$	$0.048\pm0.003$	$0.050\pm0.005$	$0.050\pm0.003$	$0.044\pm0.004$
Relative	$0.152\pm0.010$	$0.184\pm0.010$	$0.161\pm0.007$	$0.172\pm0.013$	$0.171\pm0.010$	$0.161\pm0.014$
Week 53						
Absolute	$0.049\pm0.003$	$0.053\pm0.003$	$0.053 \pm 0.003$	$0.056\pm0.003$	$0.049\pm0.003$	$0.054\pm0.006$
Relative	$0.144\pm0.010$	$0.160\pm0.008$	$0.155 \pm 0.011$	$0.174\pm0.009$	$0.164\pm0.008$	$0.186 \pm 0.015 *$

# TABLE C1Organ Weights and Organ-Weight-to-Body-Weight Ratios for Female Ratsat the 14-, 31-, and 53-Week Interim Evaluations in the 2-Year Gavage Study of TCDD<sup>a</sup>

	Vehicle Control	3 ng/kg	10 ng/kg	22 ng/kg	46 ng/kg	100 ng/kg
n						
Week 14	10	10	10	10	10	10
Week 31	10	10	10	10	10	10
Week 53	8	8	8	8	8	8
Necropsy body wt						
Week 14	$270 \pm 5$	$266 \pm 5$	$283 \pm 7$	$280 \pm 9$	$269 \pm 6$	$256 \pm 6$
Week 31	$307 \pm 7$	$300 \pm 7$	$298 \pm 5$	$290 \pm 7$	$294 \pm 7$	$274 \pm 6**$
Week 53	$338\pm 6$	$333\pm 6$	$343\pm16$	$322\pm12$	$301 \pm 8*$	$288 \pm 10^{\boldsymbol{*}\boldsymbol{*}}$
Spleen Week 14						
Absolute	$0.538 \pm 0.015^{b}$	$0.558 \pm 0.033$	$0.570 \pm 0.029$	$0.565 \pm 0.024$	$0.528 \pm 0.022$	$0.467 \pm 0.025$
Relative Week 31	$2.007 \pm 0.046^{b}$	$2.091\pm0.098$	$2.013\pm0.089$	$2.019\pm0.063$	$1.960\pm0.065$	$1.828\pm0.089$
Absolute	$0.562 \pm 0.026$	$0.563 \pm 0.023$	$0.520 \pm 0.017$	$0.511 \pm 0.025$	$0.513 \pm 0.021$	$0.434 \pm 0.011 **$
Relative	$1.825 \pm 0.066$	$1.874 \pm 0.047$	$1.748 \pm 0.047$	$1.759 \pm 0.067$	$1.747 \pm 0.059$	$1.586 \pm 0.033 **$
Week 53						
Absolute	$0.544\pm0.028$	$0.549 \pm 0.024$	$0.508\pm0.025$	$0.512\pm0.021$	$0.480\pm0.026$	$0.452 \pm 0.023 *$
Relative	$1.613\pm0.091$	$1.645\pm0.058$	$1.488\pm0.060$	$1.599\pm0.063$	$1.592\pm0.060$	$1.578\pm0.092$
Thymus						
Week 14	0.200 + 0.010	0.007 + 0.010	0.215 + 0.014	0.100 + 0.012	0.1(0.10.00(*	0 125 + 0 010**
Adsolute	$0.209 \pm 0.019$ 0.772 ± 0.062	$0.227 \pm 0.012$ 0.854 ± 0.044	$0.215 \pm 0.014$ 0.756 ± 0.027	$0.190 \pm 0.013$ 0.601 ± 0.061	$0.108 \pm 0.000^{\circ}$ 0.626 ± 0.024*	$0.125 \pm 0.010^{**}$ $0.480 \pm 0.026^{**}$
Relative	$0.772 \pm 0.002$	$0.834 \pm 0.044$	$0.730 \pm 0.037$	$0.091 \pm 0.001$	$0.020 \pm 0.024^{\circ}$	$0.489 \pm 0.030^{+1}$
Thyroid gland Week 14						
Absolute	$0.029\pm0.002$	$0.025\pm0.002$	$0.028\pm0.002$	$0.026\pm0.001$	$0.026\pm0.002$	$0.028\pm0.001$
Relative	$0.108\pm0.005$	$0.094\pm0.009$	$0.100\pm0.004$	$0.093 \pm 0.003$	$0.098\pm0.006$	$0.109\pm0.004$
Week 31						
Absolute	$0.025\pm0.002$	$0.021\pm0.002$	$0.020\pm0.001$	$0.021\pm0.002$	$0.024\pm0.002$	$0.021 \pm 0.002$
Relative	$0.083\pm0.007$	$0.071\pm0.006$	$0.067\pm0.004$	$0.072\pm0.008$	$0.081\pm0.007$	$0.076\pm0.006$
Week 53						
Absolute	$0.039\pm0.002$	$0.039\pm0.004$	$0.033\pm0.002$	$0.032\pm0.002$	$0.031 \pm 0.002 \texttt{*}$	$0.031 \pm 0.002*$
Relative	$0.117\pm0.006$	$0.116\pm0.012$	$0.097\pm0.006$	$0.101\pm0.005$	$0.105\pm0.008$	$0.108\pm0.006$

#### TABLE C1 Organ Weights and Organ-Weight-to-Body-Weight Ratios for Female Rats at the 14-, 31-, and 53-Week Interim Evaluations in the 2-Year Gavage Study of TCDD

\* Significantly different ( $P \le 0.05$ ) from the vehicle control group by Williams' or Dunnett's test. \*\*  $P \le 0.01$ <sup>a</sup> Organ weights (absolute weights) and body weights are given in grams; organ-weight-to-body-weight ratios (relative weights) are given as

b mg organ weight/g body weight (mean  $\pm$  standard error). n=9

# APPENDIX D CHEMICAL CHARACTERIZATION AND DOSE FORMULATION STUDIES

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# CHEMICAL CHARACTERIZATION AND DOSE FORMULATION STUDIES

## **PROCUREMENT AND CHARACTERIZATION** *TCDD*

TCDD was obtained from IIT Research Institute (Chicago, IL) in one lot (CR82-2-2) that was used for the 2-year study. Identity and purity analyses were conducted by the analytical chemistry laboratory, Research Triangle Institute (Research Triangle Park, NC), and the study laboratory, Battelle Columbus Operations (Columbus, OH). Reports on analyses performed in support of the TCDD study are on file at the National Institute of Environmental Health Sciences.

Lot CR82-2-2 of the chemical, a white crystalline powder, was identified as TCDD by the analytical chemistry laboratory using infrared spectroscopy, proton nuclear magnetic resonance (NMR) spectroscopy, direct probe mass spectroscopy (MS), low resolution gas chromotography (GC) coupled with MS by system A (Table D1), and melting point determination. In addition, identity analysis was conducted by the study laboratory using proton NMR. All spectra were consistent with the structure of TCDD. Infrared and mass spectra matched reference spectra of TCDD, and although a reference proton NMR spectrum was not available, the observed chemical shift agreed with that reported in the literature (Gurka *et al.*, 1985; Ashley *et al.*, 1989). A precise melting point range was not determined as the chemical appeared to sublime at approximately 260° C. The infrared, proton NMR, and mass spectra are present in Figures D1, D2, and D3, respectively.

The purity of lot CR82-2-2 was determined by the analytical chemistry laboratory using GC systems A, B, and C and by the study laboratory using system D. The purity profile obtained by system B detected two impurities with a combined relative area of 2.0% and that obtained by system C detected two impurities with a combined relative area of 1.6%. The major impurity detected by each system (1.5% of the major peak) was identified using GC/MS by system A as 1,2,4-trichlorodibenzo-*p*-dioxin. A small peak eluting immediately after the main component was believed to be a dimethyl isomer of trichloro-*p*-dioxin (positional substitution unknown). Also, a trace amount of a higher molecular weight tetrachlorinated dioxin (parent ion = 426) was observed, but due to the relatively weak intensity of the signal, precise identification could not be made. The purity profile obtained by system D indicated that the test article had a purity of 101.6% relative to a reference sample of the same lot. The overall purity of lot CR82-2-2 was determined to be 98% or greater.

#### Formulation Materials

USP-grade acetone was obtained from Spectrum Quality Products (Gardena, CA) in five lots and was used with corn oil as the vehicle in the 2-year gavage study. The identity of each lot was confirmed by the study laboratory using infrared spectroscopy prior to its use. The purity of each lot was determined by GC (system E) prior to initial use and at intervals of no more than 6 months thereafter. All acetone lots showed a purity of at least 99.9% except one that had a single impurity of 0.125%. Periodic analyses of the corn oil vehicle performed by the study laboratory using potentiometric titration demonstrated peroxide concentrations less than 3 mEq/kg.

### **PREPARATION OF STOCK SAMPLES**

Lot CR82-2-2 was dissolved in acetone and prealiquotted for use as analytical stock or formulation stock in the study because of the very small amount of chemical that was required to prepare the dose formulations at the intended concentrations. An analytical stock solution was prepared at a target concentration of 10  $\mu$ g/mL by dissolving approximately 10 mg of accurately weighed TCDD in 1,000 mL of acetone. A formulation stock solution was prepared at a target concentration of 15  $\mu$ g/mL by dissolving approximately 15 mg of accurately weighed TCDD in 1,000 mL of acetone. Following analysis to confirm proper concentration, these solutions were

used to prepare analytical standard stocks of 50 and 100  $\mu$ g, frozen reference stocks and chemical reference stocks of 100  $\mu$ g for periodic purity determinations, and dose formulation working stocks. They were prepared by transferring the required volumes of respective solutions into appropriately sized glass containers and evaporating the solvent. The dried aliquots were stored at room temperature and protected from light in amber glass bottles. Purity was monitored with periodic reanalysis by the study laboratory using system E. No degradation was observed during the course of the study.

## **PREPARATION AND ANALYSIS OF DOSE FORMULATIONS**

The dose formulations were prepared by dissolving TCDD working stocks in acetone first and then diluting in corn oil (Spectrum Quality Products) such that the final concentration contained 1.0% acetone (Table D2). The dose formulations were stored at room temperature in amber glass bottles with minimal headspace, sealed with Teflon<sup>®</sup>-lined lids, for up to 35 days.

Homogeneity studies of 1.2 and 40 ng/mL dose formulations and simulated animal room stability studies of a 1.2 ng/mL dose formulation were performed by the study laboratory using GC/MS by system F. Homogeneity was confirmed, and stability was confirmed for 3 hours under simulated animal room conditions. An additional stability study of a 0.2 ng/mL dose formulation was performed by the analytical laboratory using GC by system G. In this study, stability was confirmed for at least 35 days for the dose formulation stored in sealed glass containers at 5° C and room temperature (approximately 25° C). Gavagability was confirmed by the study laboratory for the 40 ng/mL formulation.

Periodic analyses of the dose formulations of TCDD were conducted by the study laboratory using GC/MS by system F. During the 2-year study, the dose formulations were analyzed at least every 3 months (Table D3). Of the dose formulations analyzed, 97% (56/58) were within 10% of the target concentrations; all were within 14% of target. Ninety percent (19/21) of the animal room samples were within 10% of the target concentrations; all were within 16% of target.

TABLE D1

# Gas Chromatography Systems Used in the 2-Year Gavage Study of TCDD<sup>a</sup>

Detection System	Column	Carrier Gas	Oven Temperature Program
System A Mass spectrometry, selected ion recording	J&W DB-5 MS, 30 m × 0.32 mm, 0.5-µm film thickness (J&W Scientific, Folsom, CA)	Helium at 2 mL/minute	150° C to 300° C at 10° C/minute, then held for 45 minutes
System B Electron capture	J&W DB-5 MS, 30 m × 0.32 mm, 0.5-µm film thickness (J&W Scientific)	Helium at 2.1 mL/minute	150° C to 300° C at 10° C/minute, then held for 45 minutes
<b>System C</b> Flame ionization	J&W DB-5 MS, 30 m × 0.32 mm, 0.5-µm film thickness (J&W Scientific)	Helium at 1.1 mL/minute	150° C to 300° C at 10° C/minute, then held for 15 minutes
System D Electron capture	Supelco PTE-5, 30 m × 0.32 mm, 1-µm film thickness (Supelco Inc., Bellefonte, PA)	Helium at 1.5 mL/minute	200° C to 300° C at 10° C/minute, then held for 8 minutes
<b>System E</b> Flame ionization	20% SP-2401/0.1% Carbowax 1500 on 100/120 Supelcoport, 2.4 m × 2 mm (Supelco, Inc.)	Helium at 30 mL/minute	40° C for 4 minutes, then 10° C/minute to 170° C
System F Mass spectrometry, selected ion recording	J&W DB-5 MS, 15 m × 0.25 mm, 0.25-µm film thickness (J&W Scientific)	Helium, ultra high purity at 2 mL/minute	140° C for 1 minute, then 25° C/minute to 310° C
System G Mass spectrometry, selected ion recording	J&W DB-5 MS, 30 m × 0.32 mm, 0.25-µm film thickness (J&W Scientific)	Helium at 1.2 mL/minute	100° C to 300° C at 8° C/minute, then held for 10 minutes

<sup>a</sup> Gas chromatographs were manufactured by Hewlett-Packard (Palo Alto, CA) (systems A, B, C, D, E, and G), and Carlo Erba/Fisons (Valencia, CA) (system F). Mass spectrometers were manufactured by Hewlett-Packard (systems A and G) and VG (Cheshire, UK) (system F).



FIGURE D1 Infrared Spectrum of TCDD



FIGURE D2 Proton Nuclear Magnetic Resonance Spectrum of TCDD



FIGURE D3 Direct Probe Mass Spectrum of TCDD

# TABLE D2Preparation and Storage of Dose Formulations in the 2-Year Gavage Study of TCDD

#### Preparation

Dose formulation working stocks were prepared by transferring the appropriate volumes from a 15  $\mu$ g/mL formulation stock solution into 15 mL amber glass bottles, evaporating the acetone, and sealing the bottles with Teflon<sup>®</sup>-lined lids.

Dose formulations were prepared by filling a 2 L volumetric flask approximately half full with corn oil. 20 mL of acetone was added in three aliquots to the appropriate dose formulation working stock bottle; capped, vortexed, sonicated, and transferred to the volumetric flask after each rinse. The contents of the volumetric flask were diluted to volume with corn oil, capped, shaken vigorously, and stirred on a stirplate for 3 or 24 hours, with periodic inverting and shaking.

**Chemical Lot Number** CR82-2-2

**Maximum Storage Time** 

 $35 \ days$ 

#### **Storage Conditions**

Working stocks of TCDD were stored in 15 mL amber glass vials, and sealed with Teflon<sup>®</sup>-lined lids at room temperature (approximately 25° C). Dose formulations were stored in 60, 120, or 250 mL amber glass screw-cap bottles with Teflon<sup>®</sup>-lined lids at room temperature (approximately 25° C).

#### **Study Laboratory**

Battelle Columbus Operations (Columbus, OH)

# TABLE D3Results of Analyses of Dose Formulations Administered to Female Rats in the 2-Year Gavage Studyof TCDD

Date Prepared	Date Analyzed	Target Concentration (ng/mL)	Determined Concentration <sup>a</sup> (ng/mL)	Difference from Target (%)
May 22, 1998	May 27, 1998	1.2	1.189	-1
		4	4.032	+1
		8.8	9.070	+3
		18.4	18.31	0
		40	40.90	+2
	July 2, 1998 <sup>b</sup>	1.2	1.243	+4
		4	4.083	+2
		8.8	8.633	-2
		18.4	18.35	0
		40	40.97	+2
July 17, 1998	July 22, 1998	1.2	1.193	-1
		4	3.788	-5
		8.8	8.697	-1
		18.4	17.67	-4
		40	38.47	-4
		40	37.93	-5
October 9, 1998	October 13, 1998	1.2	1.113	-7
		4	3.836	-4
		8.8	8 646	-2
		18.4	16.66	_9
		40	37.89	-5
		40	40.16	-0
December 4, 1998	December 9-10, 1998	1.2	1.124	6
		4	3.529 <sup>c</sup>	-12
		8.8	8.202	-7
		18.4	15.88 <sup>c</sup>	-14
		40	35.93	-10
		40	37.85	-5
	January 12-13, 1999 <sup>b</sup>	1.2	1.127	6
		4	3.722	-7
		8.8	8.110	-8
		18.4	16.85	-8
		40	36.50	-9
		40	38.31	-4
February 26, 1999	March 4, 1999	1.2	1.318	+10
		4	4.021	+1
		8.8	9.262	+5
		18.4	18.14	-1
		40	37.86	-5
April 23, 1999	April 28-29, 1999	1.2	1.120	-7
		4	3.816	-5
		8.8	8.218	-7
		18.4	17.19	-7
		40	36.47	0

Date Prepared	Date Analyzed	Target Concentration (ng/mL)	Determined Concentration (ng/mL)	Difference from Target (%)
July 16, 1999	July 20, 1999	1.2	1.196	0
		4	3.827	-4
		8.8	8.832	0
		18.4	17.19	-7
		40	38.55	-4
	September 1, 1999 <sup>b</sup>	1.2	1.179	-2
	•	4	3.740	-7
		8.8	8.531	-3
		18.4	17.14	-7
		40	38.57	-4
September 10, 1999	September 17, 1999	1.2	1.203	0
1	1	4	3.746	6
		8.8	8.807	0
		18.4	17.73	-4
		40	38.20	-5
December 6 and 10, 1999	December 17, 1999	1.2	1,174	-2
		4	3.609	-10
		8.8	8.555	-3
		18.4	16.89	8
		40	37.12	-7
January 28, 2000	February 2, 2000	1.2	$1.179 \pm 0.015$	-2
······································	,	4	$3.780 \pm 0.028$	-6
		8.8	$8.435 \pm 0.101$	-4
		18.4	$17.08 \pm 0.26$	-7
		40	$38.22 \pm 0.41$	-4
	March 7 2000 <sup>b</sup>	12	$1.074 \pm 0.023$	-11
		4	$3.701 \pm 0.088$	-7
		8.8	$8.608 \pm 0.180$	-2
		18.4	$15.40 \pm 0.22$	-16
		40	$36.29 \pm 1.06$	-9
April 21, 2000	April 26, 2000	1.2	$1.154 \pm 0.015$	-4
	- p.m =0, 2000	4	$3.779 \pm 0.078$	-6
		8.8	$8.605 \pm 0.102$	-2
		18.4	$16.78 \pm 0.49$	_9
		40	$38.47 \pm 0.46$	-4

#### TABLE D3 Results of Analyses of Dose Formulations Administered to Female Rats in the 2-Year Gavage Study of TCDD

<sup>a</sup> Reported value is the average of duplicate analyses or the average ± standard deviation of quadruplicate analyses. Dosing volume=2.5 mL/kg; 1.2 ng/mL=3 ng/kg, 4 ng/mL=10 ng/kg, 8.8 ng/mL=22 ng/kg, 18.4 ng/mL=46 ng/kg, 40 ng/mL=100 ng/kg.
 <sup>b</sup> Animal room samples
 <sup>c</sup> Formulation was outside the acceptable range of ± 10% of target concentration, but was used at NTP's direction.

# APPENDIX E INGREDIENTS, NUTRIENT COMPOSITION, AND CONTAMINANT LEVELS IN NTP-2000 RAT AND MOUSE RATION

TABLE E1	Ingredients of NTP-2000 Rat and Mouse Ration	202
TABLE E2	Vitamins and Minerals in NTP-2000 Rat and Mouse Ration	202
TABLE E3	Nutrient Composition of NTP-2000 Rat and Mouse Ration	203
TABLE E4	Contaminant Levels in NTP-2000 Rat and Mouse Ration	204
TABLE E5	Concentrations of PCBs and Dioxins in NTP-2000 Rat and Mouse Ration	205

Ingredients	Percent by Weight	
Ground hard winter wheat	22.26	
Ground #2 yellow shelled corn	22.18	
Wheat middlings	15.0	
Oat hulls	8.5	
Alfalfa meal (dehydrated, 17% protein)	7.5	
Purified cellulose	5.5	
Soybean meal (49% protein)	5.0	
Fish meal (60% protein)	4.0	
Corn oil (without preservatives)	3.0	
Soy oil (without preservatives)	3.0	
Dried brewer's yeast	1.0	
Calcium carbonate (USP)	0.9	
Vitamin premix, <sup>a</sup>	0.5	
Mineral premix <sup>b</sup>	0.5	
Calcium phosphate, dibasic (USP)	0.4	
Sodium chloride	0.3	
Choline chloride (70% choline)	0.26	
Methionine	0.2	

### TABLE E1 Ingredients of NTP-2000 Rat and Mouse Ration

a b

Wheat middlings as carrier Calcium carbonate as carrier

	Amount	Source
Vitamins		
Α	4,000 IU	Stabilized vitamin A palmitate or acetate
D	1,000 IU	D-activated animal sterol
Κ	1.0 mg	Menadione sodium bisulfite complex
∝-Tocopheryl acetate	100 IU	
Niacin	23 mg	
Folic acid	1.1 mg	
d-Pantothenic acid	10 mg	d-Calcium pantothenate
Riboflavin	3.3 mg	*
Thiamine	4 mg	Thiamine mononitrate
B <sub>12</sub>	52 µg	
Pyridoxine	6.3 mg	Pyridoxine hydrochloride
Biotin	0.2 mg	<i>d</i> -Biotin
Minerals		
Magnesium	514 mg	Magnesium oxide
Iron	35 mg	Iron sulfate
Zinc	12 mg	Zinc oxide
Manganese	10 mg	Manganese oxide
Copper	2.0 mg	Copper sulfate
Iodine	0.2 mg	Calcium iodate
Chromium	0.2 mg	Chromium acetate

#### TABLE E2 Vitamins and Minerals in NTP-2000 Rat and Mouse Ration<sup>a</sup>

<sup>a</sup> Per kg of finished product

	Mean ± Standard		
Nutrient	Deviation	Range	Number of Samples
Protein (% by weight)	$13.4 \pm 0.42$	12.7 – 14.5	24
Crude fat (% by weight)	$8.1 \pm 0.23$	7.6 - 8.6	24
Crude fiber (% by weight)	$9.1 \pm 0.61$	7.9 - 10.0	24
Ash (% by weight)	$4.9\pm0.17$	4.7 - 5.4	24
Amino Acids (% of total di	iet)		
Arginine	$0.731 \pm 0.050$	0.670 - 0.800	8
Cystine	$0.224 \pm 0.012$	0.210 - 0.240	8
Glycine	$0.684 \pm 0.041$	0.620 - 0.740	8
Histidine	$0.333 \pm 0.018$	0.310 - 0.350	8
Isoleucine	$0.524 \pm 0.046$	0.430 - 0.590	8
Leucine	$1.061 \pm 0.061$	0.960 - 1.130	8
Lysine	$0.708 \pm 0.056$	0.620 - 0.790	8
Methionine	$0.401 \pm 0.035$	0.350 - 0.460	8
Phenylalanine	$0.598 \pm 0.036$	0.540 - 0.640	8
Threonine	$0.501 \pm 0.051$	0.430 - 0.590	8
Tryptophan	$0.126 \pm 0.014$	0.110 - 0.150	8
Tyrosine	$0.390 \pm 0.056$	0.280 - 0.460	8
Valine	$0.640 \pm 0.049$	0.550 - 0.690	8
Essential Fatty Acids (% o	f total diet)		
Linoleic	$3.97 \pm 0.284$	3.59 - 4.54	8
Linolenic	$0.30 \pm 0.042$	0.21 - 0.35	8
Vitamins			
Vitamin $\Lambda$ (III/kg)	$5.773 \pm 0.40$	4 220 7 790	24
Vitamin D (IU/kg)	$1,000^{a}$	4,220 - 7,790	24
Vitanini D (IO/Kg)	$82.2 \pm 14.08$	62.2 107.0	0
Thisming (nnm) <sup>b</sup>	$7.8 \pm 0.85$	61 - 02	24
Pihoflavin (nnm)	$7.8 \pm 0.83$	0.1 - 9.3	24
Niacin (npm)	$5.0 \pm 1.12$ $74.3 \pm 5.04$	4.20 - 7.70	8 8
Pantothania agid (nnm)	$74.5 \pm 3.94$ 22.5 $\pm 3.06$	17.4 - 20.1	8
Puridovine (npm) <sup>b</sup>	$22.5 \pm 3.90$ 9.04 + 2.37	64 124	8
Falia agid (npm)	$9.04 \pm 2.37$	0.4 - 12.4	8
Piotin (nnm)	$1.04 \pm 0.38$	1.20 - 2.32	8
Vitamin P (nnh)	$0.535 \pm 0.15$	0.223 - 0.704	8 9
Chaling $(ppb)$	$08.7 \pm 03.0$	18.3 - 174.0	8
Chonne (ppm)*	$5,155 \pm 525$	2,700 - 3,790	8
Minerals	0.004 + 0.020	0.002 1.000	24
Calcium (%)	$0.984 \pm 0.039$	0.903 - 1.060	24
Phosphorus (%)	$0.554 \pm 0.025$	0.505 - 0.592	24
Potassium (%)	$0.659 \pm 0.022$	0.627 - 0.691	8
Chloride (%)	$0.35 / \pm 0.02 /$	0.300 - 0.392	8
Sodium (%)	$0.189 \pm 0.019$	0.160 - 0.212	8
Magnesium (%)	$0.199 \pm 0.009$	0.185 - 0.213	8
Sultur (%)	$0.1/8 \pm 0.021$	0.153 - 0.209	8
Iron (ppm)	$160 \pm 14.7$	135 - 177	8
Manganese (ppm)	$50.3 \pm 4.82$	42.1 - 56.0	8
Zinc (ppm)	$50.7 \pm 6.59$	43.3 - 61.1	8
Copper (ppm)	$6.29 \pm 0.828$	5.08 - 7.59	8
lodine (ppm)	$0.461 \pm 0.187$	0.233 - 0.843	8
Chromium (ppm)	$0.542 \pm 0.128$	0.330 - 0.707	7
Cobalt (ppm)	$0.23 \pm 0.049$	0.20 - 0.30	7

#### TABLE E3 Nutrient Composition of NTP-2000 Rat and Mouse Ration

<sup>a</sup> From formulation
 <sup>b</sup> As hydrochloride (thiamine and pyridoxine) or chloride (choline)

	Mean ± Standard		
	Deviation	Range	Number of Samples
Contaminants			
Arsenic (ppm)	$0.16 \pm 0.080$	0.10 - 0.37	24
Cadmium (ppm)	$0.04 \pm 0.007$	0.04 - 0.07	24
Lead (ppm)	$0.09 \pm 0.053$	0.05 - 0.25	24
Mercury (ppm)	<0.02		24
Selenium (ppm)	$0.19 \pm 0.033$	0.15 - 0.28	24
Aflatoxins (ppb)	<5.00		24
Nitrate nitrogen (ppm) <sup>c</sup>	$10.4 \pm 2.07$	9.04 - 16.8	24
Nitrite nitrogen (ppm) <sup>c</sup>	<0.61		24
BHA (ppm).	<1.0		24
BHT (ppm)	<1.0		24
Aerobic plate count (CFU/g)	<10		24
Coliform (MPN/g)	0		24
Escherichia coli (MPN/g)	<10		24
Salmonella (MPN/g)	Negative		24
Total nitrosoamines (ppb) <sup>e</sup>	$4.6 \pm 1.54$	2.1 - 8.8	24
<i>N</i> -Nitrosodimethylamine $(ppb)^{e}$	$1.9 \pm 0.93$	10 - 51	24
<i>N</i> -Nitrosopyrrolidine (ppb)	$2.7 \pm 0.97$	1.0 - 5.6	24
Pesticides (ppm)			
Q-BHC	< 0.01		24
B-BHC	<0.02		24
V-BHC	<0.01		24
δ-BHC	<0.01		24
Heptachlor	<0.01		24
Aldrin	<0.01		24
Hentachlor enoxide	<0.01		24
DDE	<0.01		24
ממס	<0.01		24
DDD	<0.01		24
HCB	<0.01		24
Mirex	<0.01		24
Methoxychlor	<0.01		24
Dieldrin	<0.03		24
Endrin	<0.01		24
Telodrin	<0.01		24
Chlordane	<0.01		24
Toxanhene	<0.10		24
Estimated PCBs	<0.10		24
Ronnel	<0.20		24
Ethion	<0.01		24
Trithion	<0.02		24
Diazinon	<0.05		24
Methyl chlorpyrifos	$0.126 \pm 0.115$	0.020 - 0.499	24
Methyl parathion	<0.02	0.020 0.499	24
Ethyl parathion	<0.02		24
Malathion	$0.231 \pm 0.218$	0.020 - 0.826	24
Endosulfan I	<0.01	0.020 0.020	24
Endosulfan II	<0.01		24
Endosulfan sulfate	<0.03		24
	0.00		- '

TABLE E4 Contaminant Levels in NTP-2000 Rat and Mouse Ration<sup>a</sup>

а All samples were irradiated. CFU=colony-forming units; MPN=most probable number; BHC=hexachlorocyclohexane or benzene hexachloride b

For values less than the limit of detection, the detection limit is given as the mean. Sources of contamination: alfalfa, grains, and fish meal Sources of contamination: soy oil and fish meal All values were corrected for percent recovery. с

d

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TABLE E5

Concentrations of PCBs and Dioxins in NTP-2000 Rat and Mouse Ration<sup>a</sup>

Analyte	Mean Concentration <sup>b</sup>	Standard Deviation	Mean LOQ	Standard Deviation
2,3,7,8-TCDD			0.0592	0.0106
1,2,3,7,8-PeCDD			0.119	0.0498
1,2,3,4,7,8-HxCDD			0.124	0.0366
1,2,3,6,7,8-HxCDD			0.120	0.0345
1,2,3,7,8,9-HxCDD			0.124	0.0387
1,2,3,4,6,7,8-HpCDD	0.573	0.417	0.573	0.417
OCDD	3.47	2.00	3.47	2.00
2,3,4,7,8-PeCDF	0.0413	0.0821	0.0934	0.0545
2,3,7,8-TCDF	0.0102		0.0692	0.0187
1,2,3,4,7,8-HxCDF	0.00753		0.0492	0.0213
1,2,3,6,7,8-HxCDF			0.0445	0.0155
1,2,3,7,8,9-HxCDF			0.0712	0.0259
2,3,4,6,7,8-HxCDF			0.0485	0.0176
1,2,3,7,8-PeCDF	0.00707		0.0871	0.0275
1,2,3,4,6,7,8-HpCDF	0.115	0.425	0.162	0.254
1,2,3,4,7,8,9-HpCDF			0.0870	0.0212
OCDF	0.207	0.272	0.330	0.211
2-Chlorobiphenyl	19.2	11.0	19.2	11.0
3-Chlorobiphenyl	1.73	0.465	4.99	0.893
4-Chlorobiphenyl	15.6	8.68	15.6	8.68
2,2'-Dichlorobiphenyl	62.0	54.3	62.0	54.3
2,3-Dichlorobiphenyl	267	244	267	244
2,3' -Dichlorobiphenyl	46.5	41.7	46.5	41.7
2,4-Dichlorobiphenyl/2,5-Dichlorobiphenyl	26.9	24.6	28.5	24.1
3,3'-Dichlorobiphenyl	101	108	101	108
3,4-Dichlorobiphenyl/3,4 -Dichlorobiphenyl	11.7	9.48	16.5	10.6
3,5-Dichlorobiphenyl	(2.5	(1.0	8.96	0.314
$4,4^{\circ}$ -Dichlorobiphenyl	63.5	64.8	78.5	67.8
2,2, ,5-1richlorobipnenyl/2,4, ,6-1richlorobipnenyl	112	102	112	103
	82.4	/5.3	82.4	/5.3
2,2, ,5-1 fichlorobiphenyl	202	185	202	185
2,2, ,0-1 fichlorobiphenyl $2,2,2'$ Trichlorobiphenyl $2,2,2'$ Trichlorobiphenyl $2,2,4'$ Trichlorobiphenyl $2,2,4'$ Trichlorobiphenyl $2,2,4'$ Trichlorobiphenyl $2,2,4'$ Trichlorobiphenyl $2,4'$	13./	14.8	14.9	14.1
2,5,5 - Inchlorobiphenyl/2,5,4-Inchlorobiphenyl/2,5,4-Inchlorobiphenyl	90 5	76.2	80.5	76.2
2.3.5 Trichlorobinhenvl	80.5	70.5	4.48	0.158
2.3.6 Trichlorobinhenyl/2.3' 6 Trichlorobinhenyl	13.3	12.0	14.1	12.5
2.3' 4-Trichlorobinhenvl	21.4	20.2	21.8	20.0
2.3' 5-Trichlorobinhenyl	44.9	39.1	44.9	39.1
2.4.4'-Trichlorobinhenyl	222	215	222	215
2.4.5-Trichlorobinhenvl	1 11	213	4 78	0.945
2.4.6-Trichlorobinhenvl	1.11	2.11	4 48	0.158
2.4' 5-Trichlorobinhenvl	223	195	223	195
2' 3 5-Trichlorobinhenvl	220	170	4 48	0 158
3.3'.4-Trichlorobiphenyl	4.29	2.71	6.32	2.62
3.3'.5-Trichlorobiphenyl	,		4.48	0.158
3 4 4'-Trichlorobinhenyl	30.1	25.9	30.1	25.9
3.4.5-Trichlorobiphenyl	2011	2017	4.48	0.158
3.4'.5-Trichlorobiphenyl			4.48	0.158
2.2′.3.3′-TeCB	14.4	15.4	19.2	15.4
2,2'3,4-TeCB/2,3,4',6-TeCB/2,3',4'.6-TeCB/2,3'.5.5'-TeCB	108	106	108	106
2.2′.3.4′-TeCB/2.3.3′.6-TeCB	35.7	35.5	37.3	34.8
2,2',3,5-TeCB/2,2'4,5'-TeCB	141	142	141	142
2,2',3,5'-TeCB	173	192	173	192
		-		-

TABLE E5

## Concentrations of PCBs and Dioxins in NTP-2000 Rat and Mouse Ration

Analyte	Mean Concentration	Standard Deviation	Mean LOQ	Standard Deviation	
2,2',3,6-TeCB	17.7	18.1	21.7	17.8	
2,2',3,6'-TeCB	5.75	3.36	11.4	3.97	
2,2',4,4'-TeCB	45.1	39.3	45.1	39.3	
2,2',4,5-TeCB/2,4,4',6-TeCB	26.1	27.2	29.4	26.6	
2,2′,4,6-TeCB			8.96	0.314	
2,2′,4,6′-TeCB	6.15	3.60	11.8	4.51	
2,2',5,5'-TeCB/2,3',4,6-TeCB	371	441	371	441	
2,2′,5,6′-TeCB	20.0	19.3	24.1	19.9	
2,2′,6,6′-TeCB			8.96	0.314	
2,3,3',4-TeCB			8.96	0.314	
2,3,3',4,'-TeCB/2,3,4,4'-TeCB	70.4	80.9	70.4	80.9	
2,3,3',5-TeCB			8.96	0.314	
2,3,3',5'-TeCB			8.96	0.314	
2,3,4,5-TeCB			8.96	0.314	
2,3,4,6-TeCB			8.96	0.314	
2,3,4',5-TeCB	1.25		9.40	1.49	
2,3,5,6-TeCB			8.96	0.314	
2,3',4,4'-TeCB	104	116	104	116	
2,3',4,5-TeCB			8.96	0.314	
2,3',4,5'-TeCB			8.96	0.314	
2,3',4',5-TeCB	197	238	197	238	
2,3',5',6-TeCB			8.96	0.314	
2,4,4',5-TeCB	67.2	80.3	68.0	78.7	
2',3,4,5-TeCB			8.96	0.314	
3,3',4,4'-TeCB	6.95	3.92	12.6	5.59	
3,3′,4,5-TeCB			8.96	0.314	
3,3',4,5'-TeCB			8.96	0.314	
3,3',5,5'-TeCB			8.96	0.314	
3,4,4',5-TeCB			8.96	0.314	
2,2',3,3',4-PeCB	16.7	24.2	20.8	20.5	
2,2',3,3',5-PeCB			8.96	0.314	
2,2',3,3',6-PeCB/2,2',3,5,5'-PeCB	106	124	106	124	
2,2',3,4,4'-PeCB	27.6	38.1	30.9	34.3	
2,2',3,4,5-PeCB			8.96	0.314	
2,2',3,4,5'-PeCB/2,3,4',5,6-PeCB/2',3,4,5,6'-PeCB	66.5	79.2	66.5	79.2	
2,2',3,4,6-PeCB/2,2',3,4',6-PeCB	38.1	47.7	41.4	45.0	
2,2',3,4,6'-PeCB	0.882		9.03	0.385	
2,2',3,4',5-PeCB/2,2',4,5,5'-PeCB	233	252	233	252	
2,2',3,5,6-PeCB			8.96	0.314	
2,2',3,5,6'-PeCB			8.96	0.314	
2,2',3,5',6-PeCB/2,2',3',4,6-PeCB/2,2',4,5,6'-PeCB	237	287	237	287	
2,2',3,6,6'-PeCB			8.96	0.314	
2,2',3',4,5-PeCB	61.3	77.5	62.9	74.3	
2,2',4,4',5-PeCB	109	116	109	116	
2,2',4,4',6-PeCB			8.96	0.314	
2,2',4,5',6-PeCB			8.96	0.314	
2,2',4,6,6'-PeCB			8.96	0.314	
2,3,3',4,4'-PeCB	32.4	31.4	32.4	31.4	
2,3,3',4,5-PeCB	142	187	142	187	
2,3,3',4',5-PeCB/2,3,3',4,6-PeCB	7.59	6.23	13.2	6.96	
2,3,3',4,5'PeCB/2,3,3',5,6-PeCB	6.10	7.90	12.5	7.23	
2.3.3'.4'.6-PeCB	127	142	127	142	
2,3,3',5,5'-PeCB/2,3,4,4',6-PeCB	3.88	6.58	10.3	3.86	
2,3,3',5',6-PeCB		**	8.96	0.314	

#### TABLE E5

# Concentrations of PCBs and Dioxins in NTP-2000 Rat and Mouse Ration

$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	Analyte	Mean Concentration	Standard Deviation	Mean LOQ	Standard Deviation	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,3,4,4′,5-PeCB	0.927		9.08	0.487	
2.3'4.5'-bCB       1.26       9.40       1.49         2.3'4.5'-bCB       8.96       0.314         2'3.4', 5-PCB       8.96       0.314         2'3.3', 4.5'PCB       8.96       0.314         2.3', 4.5'PCB       8.96       0.314         2.7'3.4', 5-PCB       8.96       0.314         2.7'3.4', 5-PCB       8.96       0.314         2.7'3.4', 4.5'HCB       7.48       7.04       1.31       7.06         2.7'3.4', 4.5'HCB       8.9       8.6       2.13       1.75         2.7'3.4', 4.5'HCB22', 4.3.5, 6'HCB       1.89       1.86       2.13       1.75         2.7'3.4', 5.6'HCB22', 4.3.5, 6'HCB       1.40       1.29       1.88       2.13       1.26         2.7'3.4', 5.6'HCB22', 4.3.5, 6'HCB       1.40       1.29       1.88       2.2'3.4', 5'HCB       2.0       1.88       6.5         2.7'3.4', 5'HCB22', 4.3.5, 6'HCB       1.40       1.29       1.88       2.2'3.4', 5'HCB       2.14       2.14       2.14       1.12	2,3',4,4',5-PeCB	130	198	131	192	
2.3/4.5/-PCB       8.06       0.314         2.3/4.5/-PCB       8.06       0.314         2/3.4/5/-PCB       8.06       0.314         2/3.4/5/-PCB       8.06       0.314         2/3.4/5/-PCB       8.06       0.314         2/3.4/5/-PCB       8.06       0.314         3/4.4/5/-PCB       8.06       0.314         2/3.5/4/-HCB2.3/4/5/-HCB       8.06       0.314         2/3.3/4/-HCB2.3/4/5/-HCB       8.06       0.314         2/3.3/4/-HCB2.3/4/5/-HCB       8.06       0.314         2/3.3/4/-HCB2.3/4/5/-HCB       8.07       0.01         2/3.3/4/-HCB2.3/4/5/-HCB       8.07       0.01         2/3.3/4/-HCB2.3/4/5/-HCB       3.45       1.45       9.00         2/3.3/5/-HCB2.3/4/5/-HCB       3.45       1.45       9.00       1.83         2/3.3/5/-HCB2.3/4/5/-HCB       3.46       1.45       9.00       1.83         2/3.3/5/-HCB2.3/4/5/-HCB       3.46       1.45       9.00       1.83         2/3.3/5/-HCB2.3/4/5/-HCB       8.83       6.5       8.83       6.5         2/3.3/4/-S/-HCB       8.83       6.5       8.83       6.5         2/3.4/4/-HCB2.3/4/-S/-HCB       8.80       9.0       9.0	2,3',4,4',6-PeCB	1.26		9.40	1.49	
2.3/4.5/PCB       8.96       0.314         2/3.4/5-PCB       8.96       0.314         2/3.4/5-PCB       8.96       0.314         2/3.4/5-PCB       8.96       0.314         3/4.4/5-PCB       8.96       0.314         2/3.4/5-PCB       8.96       0.314         2/3.4/5-PCB       8.96       0.314         2/3.4/4-HCR2.3/4/5.5'-HCB       7.48       7.04       13.1       7.06         2/3.3/4,5'-HCB       2.52       0.495       8.96       0.314         2/3.3/4,5'-HCB       2.52       0.495       8.96       0.314         2/3.3/4,5'-HCB       2.52       0.495       9.01       18         2/3.3/4,5'-HCB       3.45       1.45       9.0       1.88         2/3.3/5,5'-HCB2.3/4,5',6'HCB       8.9       1.6.1       1.89       1.8       1.1         2/3.3/5,5'-HCB2.3/4,4',5',6-HXCB       88.3       6.5.5       8.3       6.5.5         2/3.3/5,5'-HCB2.3/4,4',5',6-HXCB       88.3       6.5.5       8.3       6.5.5         2/3.4/5,4'-HXCB       82.0       8.4       8.2       6.4         2/3.4/4,4'-HXCB       8.9       0.14       2.2       2.4,4/5,4'-HXCB       8.96       0.314	2,3',4,5,5'-PeCB			8.96	0.314	
2'3.3', 4.5-PCB860.3142'3.4.4', 5-PCB1.499.642.262'3.4.4', 5-PCB8.660.3143', 4.4', 5'-PCB8.660.3143', 4.4', 5'-PCB8.660.3142', 3', 4.4', HCB2, 3', 4', 5, 5'-HCB8.660.3142', 3', 4.4', HCB2, 3', 4', 5, 5'-HCB8.600.3142', 3', 4.5', HCB2.520.4950.3142', 3', 4.5', HCB2.520.4950.3142', 3', 4.5', HCB2, 3', 4', 5, 6+HCB8.891.317.52', 3', 4', 5+HCB2, 2', 4, 4, 5, 6+HCB3.451.459.001.882', 3', 4', 5+HCB2, 2', 4, 4, 5', 6+HCB1.611.892.091.832', 3', 4', 5+HCB2, 2', 4, 4, 5', 6+HCB88.365.58.6552', 3', 4', 5+HCB2, 2', 4, 4, 5', 6+HCB89.268.489.268.42', 3, 4', 5+HCB2, 3', 4', 5', 6+HCB89.268.489.60.3142', 3, 4', 5+HCB2, 3', 4', 5', 6+HCB89.365.52.142.14 <t< td=""><td>2,3',4,5',6-PeCB</td><td></td><td></td><td>8.96</td><td>0.314</td><td></td></t<>	2,3',4,5',6-PeCB			8.96	0.314	
2'3.4',5'-PCCB       8.96       0.314         33',44',5-PCCB       8.96       0.314         22',33',44',5-PCCB       8.96       0.314         22',33',44',5-PCCB       8.96       0.314         22',33',45'-HACB       8.97       0.48         22',33',45'-HACB       8.97       0.10       2.75         22',33',56,4-HCB2,2',34,56'-HACB       8.92       0.84       126         22',33',56,4-HCB2,2',34,56'-HACB       8.92       0.84       126         22',34',56'-HACB       16.1       18.9       20.9       18.3         22',34,46'-HACB       8.92       0.84       892       0.84         22',34,46'-HACB       8.92       0.84       126         22',34,46'-HACB       8.92       0.84       126         22',34,46'-HACB       8.92       0.84       127         22',34,46'-HACB       8.92       0.84       127         22',34,46'-HACB       8.96 </td <td>2',3,3',4,5-PeCB</td> <td></td> <td></td> <td>8.96</td> <td>0.314</td> <td></td>	2',3,3',4,5-PeCB			8.96	0.314	
2',34,5'-PCB 8,96 0.314 33',44,5'-PCB 8,96 0.314 22',33',45'-PCB 8,96 0.314 22',33',45'-PCB 8,96 0.314 22',33',45'-PCB 8,96 0.314 22',33',46'-PCB2,33',4',5,6'HCB 18,9 18,6 2,13 17.5 22',33',46'-PCB2,33',4',5,6'HCB 2,79 2,62 10,1 2,75 22',33',56'-PCB 2,79 2,62 10,1 2,75 22',33',56'-PCB 14,0 12,9 18,0 12,6 22',33',56'-PCB 14,0 12,9 18,0 12,6 22',33',56'-PCB 14,0 12,9 18,0 12,6 22',33',56'-PCB 14,0 12,9 18,0 12,6 22',34,4',5'-PCB 8,3 65,5 8,83 65,5 8,83 65,5 8,83 65,5 8,84 65,5 22',34,4',5'-PCB 8,10 8,83 65,5 8,83 65,5 22',34,4',5'-PCB 8,10 8,83 65,5 8,83 65,5 22',34,4',5'-PCB 8,10 8,83 65,5 8,83 65,5 22',34,4',5'-PCB 8,10 8,81,7 22',34,4',5'-PCB 8,10 8,81,7 22',34,4',5'-PCB 8,10 8,81,7 22',34,4',5'-PCB 8,10 8,10 8,10 7,68 22',34,4',5'-PCB 8,10 8,10 8,10 7,68 22',34,4',5'-PCB 8,10 9,11 8,10 7,68 22',34,4',5'-PCB 8,10 9,11 8,10 7,68 22',34,4',5'-PCB 8,10 9,11 8,10 7,68 22',34,4',5'-PCB 8,10 9,11 8,11 8,11 8,11 8,11 8,11 8,11 8,11	2',3,4,4',5-PeCB			8.96	0.314	
33/44/5-PCCB       896       0.314         22/33/44/5-HCCB.2.33/4/5.5'-HKCB       7.48       7.04       13.1       7.06         22/33/44/5-HKCB       2.52       0.495       9.86       2.00         22/33/4,45'-HKCB       2.52       0.495       9.86       0.314         22/33/4,45'-HKCB       18.9       18.6       2.13       17.5         22/33/4,5'-HKCB,22/3,4,5'-HKCB       3.45       1.45       9.90       1.88         22/33/5,5'-HKCB,22/3,4,5'-HKCB       3.45       1.45       9.90       1.83         22/33/5,6'-HKCB,22/3,4,4,5'-HKCB       16.1       18.9       2.6       8.96       0.314         22/3,4/4'-SHKCB,22/3,4/4,5-HKCB,23,3'4',5',6-HKCB       88.3       65.5       88.3       65.5         22/3,4/4/-SHKCB,23/3,4/5,6-HKCB,23,3',4',5',6-HKCB       89.2       68.4       89.2       68.4         22/3,4/4/-SHKCB,23/4,4/5-HKCB       89.2       68.4       89.2       68.4         22/3,4/4/-SHKCB,23/4,4/5-HKCB       89.2       68.4       89.6       0.314         22/3,4/4/-SHKCB,23/4,4/5-HKCB       89.6       0.314       22/3,4/5/5-HKCB       89.6       0.314         22/3,4/4/-SHKCB,23/4,4/5-HKCB       1.31       9.16       0.768       22/3,4/5/5-HKCB       <	2',3,4,5,5'-PeCB	1.49		9.64	2.26	
3.3/4.5/-PCB	3,3',4,4',5-PeCB			8.96	0.314	
22/3,4/4/-HxCB2,33/4/5/-HxCB       7.48       7.04       13.1       7.06         22/3,4/4/-HxCB       2.52       0.495       9.86       0.314         22/3,4/4/-HxCB       2.52       0.495       9.86       0.314         22/3,4/4/-HxCB2,3,3/4,5'-6HxCB       18.9       18.6       21.3       17.5         22/3,3/4,6/-HxCB2,3,3/4,5'-6HxCB       3.45       1.45       9.90       1.83         22/3,3/5,5'-HxCB2,2/3,4,5,6'HxCB       16.1       12.9       18.0       12.6         22/3,3/5,6'HxCB2,3/3/4,5,6'HxCB       8.3       6.5       8.3       6.5         22/3,4/5,6'HxCB2,3/3/4,5,6'HxCB2,3,3'4,5,6'HxCB       8.92       6.84       89.2       6.84         22/3,4/4,6'HxCB2,3/3,4',5,6'HxCB       8.92       6.84       89.2       6.84         22/3,4/4,6'HxCB2,3/3,4',5,6'HxCB       8.3       6.5       8.3       6.5         22/3,4/4,6'HxCB2,3/4,5,6'HxCB       8.92       6.84       89.2       6.84       89.2       6.84       89.2       6.84       89.2       6.84       89.2       6.84       89.2       6.84       89.2       6.84       89.2       6.84       89.2       6.84       89.2       6.84       89.2       6.84       89.6       0.314       22/3,4/5,6'	3,3',4,4,5'-PeCB			8.96	0.314	
22/33/4/5-HxCB     8.96     0.314       22/33/4/5-HxCB     8.96     0.314       22/33/4/5-HxCB     8.96     0.314       22/33/4/5-HxCB22/3/4,5/6-HxCB     18.9     18.6     21.3       22/33/5/5-HxCB22/3,4,5/6-HxCB     2.79     2.62     10.1     2.75       22/33/5/5/FxCB     16.1     18.9     18.6     12.6       22/33/5/5/FxCB     16.1     18.9     18.6     12.6       22/34/5/5/FxCB     16.1     18.9     2.62     10.1     2.75       22/34/5/FxCB     16.1     18.9     2.09     18.3       22/34/5/FxCB     16.1     18.9     2.09     18.3       22/34/5/FxCB     13.0     86     0.314       22/34/5/FxCB     88.3     65.5     88.3     65.5       22/34/5/FxCB     13.1     9.46     1.67       22/34/5/FxCB     1.31     9.46     1.67       22/34/5/FxCB     1.03     9.18     0.768       22/34/5/FxCB     1.03     9.18     0.768       22/34/5/FxCB     1.03     9.18     0.768       22/34/5/FxCB     1.03     9.18     0.768       22/34/5/FxCB     1.03     9.75     2.59       22/34/5/FxCB     1.03     9.75     2.59	2,2',3,3',4,4'-HxCB/2,3,3',4',5,5'-HxCB	7.48	7.04	13.1	7.06	
2/2/3/4/5/-HxCB       2.52       0.495       9.86       2.00         2/2/3/4/6/HxCB       18.9       18.6       2.1.3       17.5         2/3/3/4/6/HxCB/2/3/3/4/5/6-HxCB       3.45       1.45       9.90       1.88         2/3/3/5/-HxCB/2/3/3/5/-HxCB       3.45       1.45       9.90       1.88         2/3/3/5/-HxCB/2/3/3/5/-HxCB       2.70       10.1       2.75         2/3/3/5/-HxCB/2/3/4/5/-HxCB       16.0       12.9       18.0       12.6         2/3/3/5/-HxCB/2/3/4/5/-HxCB       8.65       8.83       65.5         2/3/3/6/-HxCB/2/3/4/5/-HxCB       8.60       0.314       22.7         2/3/4/6/-HxCB/2/3/4/5/-HxCB       8.90       6.84       89.2       68.4         2/3/4/6/-HxCB       8.96       0.314       22.3/4/5/-HxCB       2.27/3/4/5/-HxCB       8.96       0.314         2/3/4/5/-HxCB       1.31       9.46       1.67       2.2/3/4/5/-HxCB       2.2/3/4/5/-HxCB       2.1       2.1       2.2/3/4/5/-HxCB       2.1       2.1       2.2/3/4/5/-HxCB       2.1       2.1	2.2′.3.3′.4.5-HxCB			8.96	0.314	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2.2′.3.3′.4.5′-HxCB	2.52	0.495	9.86	2.00	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2.2′.3.3′.4.6-HxCB			8.96	0.314	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2 2' 3 3' 4 6'-HxCB/2 3 3' 4 5' 6-HxCB	18.9	18.6	21.3	17.5	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2 2' 3 3' 5 5'-HxCB/2 2' 3 4 5 6-HxCB	3 45	1 45	9.90	1.88	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2 2' 3 3' 5 6-HxCB/2 2' 3 4 5 6'-HxCB	2 79	2.62	10.1	2 75	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2 2' 3 3' 5 6'-HyCB	14.0	12.02	18.0	12.6	
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	2,2',3,3',5,6'-HxCB	16.1	18.9	20.9	18.3	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,2',3,5',0,0' -IIXCD 2,2',3,4,4',5' HyCR	10.1	10.9	8.96	0.314	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,2,3,4,4,5' Hy(D/) $2,2',4',5,6$ Hy(D/) $2,2',4',5',6$ Hy(D	<u> </u>	65 5	8.90	0.314	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,2',3,4,4',5' -fixed/ $2,3,5',4',5,0'$ -fixed/ $2,3,5',4',5',0'$ -fixed	80.3	69.5	80.3	68.4	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,2',3,4,4',6' Hy/CP	69.2	08.4	8.06	0314	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,2',3,4,4',0' -fixed	6.01	1 99	0.90	0.314	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,2,3,4,5,3 - <b>I</b> XCD	0.01	4.00	11./	4.70	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,2,3,4,5,0-HXCB	1.51		9.46	1.0/	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,2,3,4,0,0 -fixed	25.0	21.5	8.90 25.9	0.514	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,2, 3,4, 5,5 -HXCB/2,5,5, 4, 5 0-HXCB	25.0	21.5	23.8	21.2	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,2, 3,4, 5,0-HXCB	1.05		9.18	0.768	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,2,3,4,5,6 -HXCB			8.96	0.314	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,2,3,4,6,6 -HXCB	21.0	10.0	8.96	0.314	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,2 ,3,5,5 ,6-HXCB	21.9	18.2	24.3	18.1	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,2,3,5,6,6 -HxCB	507	1 510	8.96	0.314	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,2 ,4,4 ,5,5 -HxCB	58/	1,513	587	1,514	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,2 <sup>°</sup> ,4,4 <sup>°</sup> ,5,6 <sup>°</sup> -HxCB	1.59		9.75	2.59	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,2 <sup>°</sup> ,4,4 <sup>°</sup> ,6,6 <sup>°</sup> -HxCB	1 =0		8.96	0.314	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,3,3',4,4',5-HxCB	1.79	0.382	9.05	0.423	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,3,3',4,4',5'-HxCB			8.96	0.314	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,3,3 <sup>°</sup> ,4,4 <sup>°</sup> ,6-HxCB/2,3,3 <sup>°</sup> ,4,5,6-HxCB	3.79	2.82	10.2	2.67	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,3,3',4,5,5'-HxCB			8.96	0.314	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,3,4,4',5,6-HxCB			8.96	0.314	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,3',4,4',5,5'-HxCB	0.865		9.02	0.352	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,3',4,4',5',6-HxCB			8.96	0.314	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	3,3',4,4',5,5'-HxCB			8.96	0.314	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,2',3,3',4,4',5-HpCB	10.9	9.25	14.1	8.29	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,2',3,3',4,4',6-HpCB	0.945		9.10	0.532	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,2',3,3',4,5,5'-HpCB			8.96	0.314	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,2',3,3',4,5,6-HpCB			8.96	0.314	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2,2',3,3',4,5,6'-HpCB	9.18	8.79	13.2	7.48	
2,2',3,3',4,6,6'-HpCB       8.96       0.314         2,2',3,3',4',5,6-HpCB       8.07       9.24       12.9       7.46         2,2',3,3',5,5',6-HpCB       4.98       7.90       11.4       5.64         2,2',3,3',5,5',6-HpCB       4.77       8.51       11.3       5.51         2,2',3,4,4',5,5'-HpCB       33.4       21.9       33.4       21.9         2,2',3,4,4',5,6-HpCB       8.96       0.314	2,2′,3,3′,4,5′,6-HpCB			8.96	0.314	
2,2',3,3',4',5,6-HpCB       8.07       9.24       12.9       7.46         2,2',3,3',5,5',6-HpCB       4.98       7.90       11.4       5.64         2,2',3,3',5,5',6-HpCB       4.77       8.51       11.3       5.51         2,2',3,4,4',5,5'-HpCB       33.4       21.9       33.4       21.9         2,2',3,4,4',5,6-HpCB       8.96       0.314	2,2',3,3',4,6,6'-HpCB			8.96	0.314	
2,2',3,3',5,5',6-HpCB4.987.9011.45.642,2',3,3',5,6,6'-HpCB4.778.5111.35.512,2',3,4,4',5,5'-HpCB33.421.933.421.92,2',3,4,4',5,6-HpCB8.960.314	2,2',3,3',4',5,6-HpCB	8.07	9.24	12.9	7.46	
2,2',3,3',5,6,6'-HpCB       4.77       8.51       11.3       5.51         2,2',3,4,4',5,5'-HpCB       33.4       21.9       33.4       21.9         2,2',3,4,4',5,6-HpCB       8.96       0.314	2,2′,3,3′,5,5′,6-НрСВ	4.98	7.90	11.4	5.64	
2,2',3,4,4',5,5'-HpCB 33.4 21.9 33.4 21.9 22',3,4,4',5,6-HpCB 896 0.314	2,2',3,3',5,6,6'-HpCB	4.77	8.51	11.3	5.51	
2.2', 3.4.4', 5.6-HpCB 8.96 0.314	2,2',3,4,4',5,5'-HpCB	33.4	21.9	33.4	21.9	
	2,2',3,4,4',5,6-HpCB			8.96	0.314	
TABLE	E5					
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IADLL	1.5					

#### Concentrations of PCBs and Dioxins in NTP-2000 Rat and Mouse Ration

Analyte	Mean Concentration	Standard Deviation	Mean LOQ	Standard Deviation	
2.2'.3.4.4'.5.6'-HpCB/2.2'.3.4'.5.5'.6-HpCB	38.1	34.0	38.1	34.0	
2.2'.3.4.4'.5'.6-HpCB	7.49	9.53	12.3	7.22	
2,2',3,4,4',6,6'-HpCB			8.96	0.314	
2,2',3,4,5,5',6-HpCB			8.96	0.314	
2,2',3,4,5,6,6'-HpCB			8.96	0.314	
2,2',3,4',5,6,6'-HpCB			8.96	0.314	
2,3,3',4,4',5,5'-HpCB			8.96	0.314	
2,3,3',4,4',5,6-HpCB			8.96	0.314	
2,3,3',4,4',5',6-HpCB			8.96	0.314	
2,3,3',4,5,5',6-HpCB			8.96	0.314	
2,3,3',4',5,5',6-HpCB			8.96	0.314	
2,2',3,3',4,4',5,5'-OCB	2.41		14.2	4.22	
2,2',3,3',4,4',5,6-OCB			13.0	1.07	
2,2',3,3',4,4',5,6'-OCB/2,2',3,4,4',5,5',6-OCB	6.94	15.4	16.6	8.94	
2,2',3,3',4,4',6,6'-OCB			13.0	1.07	
2,2',3,3',4,5,5',6-OCB			13.0	1.07	
2,2',3,3',4,5,6,6'-OCB	7.65	17.5	17.3	10.4	
2,2',3,3',4,5',6,6'-OCB			13.0	1.07	
2,2',3,3',4,5,5',6'-OCB	1.64		13.4	1.85	
2,2',3,3',5,5',6,6'-OCB	3.18		15.0	6.73	
2,2',3,4,4',5,6,6'-OCB			13.0	1.07	
2,3,3',4,4',5,5',6-OCB			13.0	1.07	
2,2',3,3',4,4',5,5',6-NCB	6.15		18.0	16.5	
2,2',3,3',4,4',5,6,6'-NCB	1.65		13.4	1.90	
2,2',3,3',4,5,5',6,6'-NCB	4.36		16.1	10.6	
DeCB	6.17		18.0	16.6	

Data presented as pg analyte/g feed; LOQ=Limit of quantitation. Dioxin and dibenzofuran congeners were analyzed by EPA Method 1613, using GC with high resolution mass spectrometry and isotope dilution. PCB congeners were analyzed by EPA Method 1668, using GC with high resolution mass а spectrometry. b

Mean concentration of samples with measurable concentrations; blanks indicate concentrations below the limit of detection in all samples.

## APPENDIX F SENTINEL ANIMAL PROGRAM

Methods	 210
Results	 210

### SENTINEL ANIMAL PROGRAM

### **METHODS**

Rodents used in the Carcinogenesis Program of the National Toxicology Program are produced in optimally clean facilities to eliminate potential pathogens that may affect study results. The Sentinel Animal Program is part of the periodic monitoring of animal health that occurs during the toxicologic evaluation of chemical compounds. Under this program, the disease state of the rodents is monitored via serology on sera from extra (sentinel) animals in the study rooms. These animals and the study animals are subject to identical environmental conditions. The sentinel animals come from the same production source and weanling groups as the animals used for the studies of chemical compounds.

Serum samples were collected from randomly selected male and female rats during the 2-year study. Blood from each animal was collected and allowed to clot, and the serum was separated. The samples were processed appropriately and sent to Microbiological Associates, Inc./BioReliance Corp. (Rockville, MD), for determination of antibody titers. The laboratory serology methods and viral agents for which testing was performed are tabulated below; the times at which blood was collected during the studies are also listed.

#### **Method and Test** RATS

#### **ELISA**

*Mycoplasma arthritidis* Mycoplasma pulmonis PVM (pneumonia virus of mice) RCV/SDA (rat coronavirus/sialodacryoadenitis virus) Sendai

### **Time of Analysis**

Study termination Study termination 1, 6, 12, and 18 months, study termination 1, 6, 12, and 18 months, study termination 1, 6, 12, and 18 months, study termination

Immunofluorescence Assay Parvovirus M. arthritidis

#### RESULTS

All serology tests were negative.

1, 6, 12, and 18 months, study termination Study termination

## APPENDIX G SINGLE-DOSE TOXICOKINETIC STUDY IN FEMALE SPRAGUE-DAWLEY RATS

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### SINGLE-DOSE TOXICOKINETIC STUDY IN FEMALE SPRAGUE-DAWLEY RATS

### INTRODUCTION

A single dose of TCDD was administered by gavage to Harlan Sprague-Dawley rats at 50 or 100 ng/kg. TCDD levels were determined in postdose blood, lung, liver and fat tissue samples at time points up to 100 days. The results were analyzed to establish basic toxicokinetic parameters. Because the dose volume used in the bioassay was 2.5 mL/kg rather than the 5.0 mL/kg used in most studies of TCDD, both volumes were used at the high dose to determine the effect of reducing the dose volume. Bioavailability was not determined in this study because no intravenous administration was conducted.

#### **MATERIALS AND METHODS**

TCDD was procured in one lot (970401R-AC) from AccuStandard, Inc. (New Haven, CT), and was characterized by nuclear magnetic resonance and mass spectrometry. It was found to be 94% pure with one impurity with a mass spectrum consistent with triphenylphosphine. Because it was a single administration study and the dose would be so low (about 6 ng triphenylphosphine/kg), it was decided that the material would be suitable for the study. Formulations for the study were prepared in corn oil at concentrations of either 2.5 or 5.0 mL/kg with 1% acetone as described in Appendix D.

Female rats (11 weeks of age at the start of the study) were used for the study. They were administered 50 or 100 ng/kg of formulation in a volume of 2.5 mL/kg or 100 ng/kg of formulation in a volume of 5.0 mL/kg. At 0.25, 0.5, 1.0, 1.5, 2, 3, 4, 5, 6, 8, 10, 13, 16, and 24 hours and 2, 4, 6, 10, 15, 30, 45, 60, 75, 90, 105, 120, and 150 days, postdosing groups of five rats per time point were anesthetized with a  $CO_2/O_2$  mixture and as much blood as possible was collected in EDTA tubes by cardiac puncture and stored at 5° C for analysis. At 8 and 24 hours and 2, 30, 60, 90, 120, and 150 days, the rats were euthanized with  $CO_2$  after the blood was collected and liver, lung, and mesenteric fat were collected, weighed, and frozen at  $-20^{\circ}$  C until analysis.

For analysis, 20  $\mu$ L of a 500 ng/mL solution of  ${}^{13}C_{12}$ -TCDD (internal standard) was added to a 1 mL aliquot of blood or 0.5 g aliquot of lung or liver and 2 mL of 50% potassium hydroxide solution. The samples were placed in a 40° C water bath for 15 minutes and digested overnight on a horizonal shaker. The digested samples were extracted with 10 mL of acetone:hexane (80:20) by vortexing followed by centrifugation. Fat tissue (0.5 g) was extracted with 10 mL of hexane without saponification after addition of the internal standard. For all tissues, the extraction solvent was removed with a nitrogen stream. Each residue was dissolved in about 1 mL of hexane and eluted through a silica gel column. The eluates were reduced to about 200  $\mu$ L and transferred to an autosampler. Samples were analyzed by gas chromatography/mass spectrometry using an HP 6890 gas chromatograph and an HP 5973 MSD mass spectrometer. A DB-5MS column, 30 m × 0.25 mm ID, 0.25- $\mu$ m film thickness was used; the oven temperature started at 100° C, increased at 8° C/minute to 300° C, and was then held for 10 minutes. Mass spectra were collected in single ion monitoring mode at 70 eV, scanning m/z 334, 332 (TCDD), 322, and 320 (internal standard). Responses for TCDD were quantitated using least squares linear regression of a calibration curve generated from matched tissues from untreated Sprague-Dawley rats spiked with TCDD. Table G1 provides figures of merit from the validation of the methods used.

Noncompartmental modeling with WinNonlin<sup>®</sup> (Pharsight Corp., Apex, NC) was used to derive toxicokinetic parameters from concentration versus time data.

#### **RESULTS AND DISCUSSION**

There were insufficient data for toxicokinetic modeling of blood and lung (Table G2). Measured blood concentrations of TCDD were generally below the limit of quantitation, suggesting rapid absorption and distribution to tissues. The limited number of timepoints limited the toxicokinetic analysis of the remaining tissues. Elimination half-lives and terminal elimination rate constants could not be estimated. While half-lives could not be calculated, it appears that TCDD is eliminated more rapidly from liver than from fat. There was no effect of dose volume on toxicokinetic parameters. Tissue concentration data are presented in Figures G1 to G4.

Toxicokinetic modeling of the data in this study was limited to simple noncompartmental analysis, primarily area under the concentration versus time curve (AUC), due to the limited data set. Doses to liver and fat as estimated by AUC/dose were approximately linear with administered dose. The AUCs for liver and fat were similar. In previously reported studies, tissue concentrations of <sup>14</sup>C-TCDD-derived radioactivity were also nearly equal in liver and fat 3 weeks after a single dose (Rose *et al.*, 1976). This same study reported three to five times as much TCDD in liver than in fat after daily dosing for 1, 3, or 7 weeks. In the present study, TCDD concentration in liver was maximal at day 1, but  $C_{max}$  in fat occurred 20 to 40 days later, implying redistribution to fat. Because the redistribution to fat was slow with respect to the dosing interval, TCDD concentrations would be expected to be higher in liver after multiple daily doses.

Tissue	Linearity <sup>a</sup>	Limit of Quantitation	Precision <sup>b</sup>	Accuracy <sup>c</sup>
Blood	> 0.99	200 pg/g	Within 15%	34% or less
Lung	> 0.99	200 pg/g	Within 15%	15% or less
Liver	> 0.99	200 pg/g	Within 15%	15% or less
Fat	> 0.99	120 pg/g	Within 15%	22% or less

#### TABLE G1 Figures of Merit for Assays of TCDD in the Single Gavage Dose Toxicokinetic Study in Female Sprague-Dawley Rats

а b

c

Correlation coefficient Standard deviation of quality control samples Relative error in determined versus prepared concentration of calibration standards

TABLE G2
Foxicokinetic Parameter Estimates in Female Sprague-Dawley Rats after a Single Gavage Dose of TCDD

Parameter	Dose <sup>a</sup>	Tissue Param	eter Estimate	
	(ng/kg)	Liver	Fat	
C <sub>max</sub> (pg/g)	50	494	105	
max 4 C C	100	943	483	
	100 <sup>b</sup>	1,350	397	
T <sub>max</sub> (days)	50	1	40	
max	100	1	20	
	100 <sup>b</sup>	1	1	
$AUC_{lost}$ (days • pg/g)	50	8,600	5,300	
last	100	14,600	19,200	
	100 <sup>b</sup>	12,900	27,500	
$AUC_{lost}/Dose (days \cdot pg/g)/(ng/kg)$	50	172	106	
	100	146	192	
	100 <sup>b</sup>	129	275	

Dose volume=2.5 mL/kg except where otherwise indicated Dose volume=5.0 mL/kg a b



FIGURE G1 Lung Concentrations of TCDD in Female Sprague-Dawley Rats after a Single Gavage Dose of TCDD



FIGURE G2 Liver Concentrations of TCDD in Female Sprague-Dawley Rats after a Single Gavage Dose of TCDD







FIGURE G4 Fat Concentrations of TCDD in Female Sprague-Dawley Rats after a Single Gavage Dose of TCDD

## APPENDIX H PHYSIOLOGICALLY BASED PHARMACOKINETIC MODEL

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### PHYSIOLOGICALLY BASED PHARMACOKINETIC MODEL

### INTRODUCTION

A goal for the physiologically based pharmacokinetic (PBPK) modeling of the disposition data from the dioxin toxic equivalency factor (TEF) evaluation studies is a general model for the tissue distribution of dioxin-like compounds (DLCs) and mixtures of compounds that interact with the aryl hydrocarbon receptor (AhR) in the Sprague-Dawley rat.

One key aspect to understanding the toxicity of an agent is how dose is related to the toxicity of concern. The utility of a PBPK model is in its ability to predict alternate measures of "dose" other than those that are readily measured (such as administered dose or tissue concentrations). In addition, the kinetics of tissue distribution of a compound can be compared between different routes and patterns of exposure. Also, an understanding of the factors that govern the tissue distribution of a compound, its metabolites, and subsequent molecular/biochemical responses may provide insights into the factors governing the dose response of toxicity, site specificity, and mode of action of the compound under study.

In general, PBPK models have been validated in the observable response range for numerous compounds in both animals and humans, making them useful for risk assessment, especially for cross-species extrapolation. They also aid in extrapolation from one chemical to other structurally related chemicals because many of the components of the model are the same or can be deduced for related compounds.

The disposition of a chemical within the body is governed by the absorption of an administered chemical and its distribution among tissues, metabolism, and elimination from the body (ADME). These processes for TCDD and related DLCs in part depend upon their physicochemical properties (e.g., tissue permeation and partition coefficients, kinetic constants, and biochemical parameters) and physiological parameters (e.g., organ volumes and blood flow rates).

A PBPK model for TCDD is a mathematical structure that describes the relationship between these factors and ADME. This model describes the pharmacokinetics of TCDD by a series of mass-balance differential equations in which the state variables represent the concentration of TCDD in anatomically distinct regions, "compartments" of the body. These tissue compartments are linked by a physiologically realistic pattern of blood perfusion and flow through the different tissue compartments.

The time course of behavior in each compartment of a PBPK model is defined by equations and model parameters for input and loss of chemical. The specific structure of a PBPK model and the assumptions used to develop the model are encoded in the equations. The model's physiological parameters are, in many cases, compound independent, well established, and available in the literature (e.g., rates of blood flow, blood volume, tissue volumes, etc.). Physicochemical parameters are used that are often specific to a given compound but can be measured experimentally and may be available in the literature. Some of these parameters may not be available *a priori* and so have to be determined within the framework of the model by an iterative process of changing the parameter, fitting the model to a given dataset, and evaluating the goodness of the fit of the model to the data. Careful evaluation of any PBPK model must involve the adequacy of its fit to the data, the relationship of its structure to the underlying biology, and the mathematical details linking these two. In addition, the biological plausibility of optimized parameters needs to be considered. Validation of the model using datasets that were not used in its construction lends more credence to the predictive power of a model.

### **MODEL DEVELOPMENT**

For the current dioxin TEF evaluation model, the same basic model structure was used for all compounds studied, with some of the model parameters, such as metabolism or binding to the AhR, unique to each model. The model was based upon the model of Kohn *et al.* (2001). The Kohn model is an extension of earlier PBPK models for TCDD in rats (Kohn *et al.*, 1993, 1996) that with each iteration has gone through further rounds of refinement and inclusion of increased biological complexity. A thorough summary of PBPK modeling for TCDD and the evolution of these and other PBPK models of TCDD can be found elsewhere (USEPA, 2000c).

Kohn's model included compartments for fat, liver, kidney, gastrointestinal tract, muscle, and viscera. Blood is distributed among arterial, venous, and tissue capillary spaces. The model also includes equations for the liver amounts of AhR, CYP1A1, CYP1A2, and CYP1B1, as well as equations describing basal expression, induction by TCDD, and degradation of the mRNA for each of these. The amount of each enzyme depends on the time-lagged concentration of the corresponding mRNA. TCDD in the liver may bind to CYP1A2 and the AhR. A key to the model is that the induction rates for all four represented mRNAs depend on the time-lagged concentration of AhR bound to TCDD. Induction increases from zero to a maximum rate as the concentration of AhR-TCDD increases. Since transthyretin (also known as prealbumin) can bind hydroxylated polychlorinated dibenzodioxins and single doses of TCDD can cause a prolonged decrease in this protein, a dose-dependent decrease was included in the model. This bound TCDD cannot enter the tissues in the model and may become free in the blood by dissociation or proteolysis. To allow the model to fit data at both low and high doses, the model includes loss of TCDD from the liver by lysis of dead cells (as the result of hepatotoxicity) where the rate of cell death was assumed to increase as a hyperbolic function of the cumulative amount of unbound hepatic TCDD.

There were three main steps to building a PBPK model for the dioxin TEF evaluation studies; conversion of Kohn's model, addition of a lung compartment, and addition of the study-specific body weights to the model. A copy of Kohn's model coded in the ScoP simulation was converted to both a Matlab model and a Simulink model. Simulations from the three models were compared to each other to confirm that the conversion to Matlab/Simulink was accurate. Next, a lung compartment was added to the Simulink model because the NTP data for the TEF studies include lung tissue concentrations. The lung compartment is diffusion-limited and includes the same equations used in the liver for the AhR, CYP1A1, and CYP1B1. The lung and liver compartments use the same gene expression parameters on a per liter basis. The final step was to include the rat body weights from each study rather than the body weight function from Kohn's model. Body weights were available weekly for the first 12 weeks of the studies and then monthly for the remainder of the studies. Interpolation of the mean body weights was used to estimate the body weight as a function of time.

All the parameters from Kohn's model (Kohn *et al.*, 2001) were used in the new model. The only new parameters were the lung partition coefficient and the lung permeability factor. Values of the lung partition coefficient and the lung permeability factor parameters were estimated by optimization, fitting the model predictions to the tissue data (liver, lung, fat, blood).

### **RESULTS AND DISCUSSION**

The model predictions match the data over the dose range and across tissues. Kohn's model (Kohn *et al.*, 2001) makes accurate predictions of the new NTP data for liver and adipose (results not shown). The model predictions for lung and blood are correctly below the limit of detection. The parameter estimates for the lung partition coefficient and lung permeability are 4.57 and 0.85 respectively. The lung partition coefficient is very close to the liver value and the permeability value is between the liver and kidney permeability values. All of these tissues are well perfused and should have similar values. The accuracy of the predictions suggests that it was reasonable to use the parameters from the liver to describe the cytochrome P450 and AhR dynamics in the lung. Lung data with a lower limit of detection would be useful to better evaluate the model. Overall, the model succeeds in describing the data and can be used as a base model for modeling similar chemicals that bind to the AhR.

## TABLE H1TCDD Model Parameters

Parameter	Model Value	Unit	
<i>lt</i> (lag time for induced expression)	0.2	day	
Cardiac Output	14.7	L/hr per kg <sup>0.7</sup>	
<i>V</i> <sub>Protein</sub> (blood binding protein)	300	nmole/L per day	
Ki <sub>Protein</sub> (inhibition of blood protein production)	0.0006	nM	
Kd <sub>Protein</sub> (blood binding protein)	10	nM	
K <sub>AhR</sub>	0.27	nM	
K <sub>CYP1A2</sub> TCDD	30	nM	
V <sub>metabolism</sub>	9.12	nmole/L per day	
K <sub>metabolism</sub>	0.968	nM	
n <sub>metabolism</sub>	1.12	_	
k <sub>subchronic absorption</sub>	0.65	kg <sup>0.75</sup> /day	
k <sub>absorption</sub>	4.8	kg <sup>0.75</sup> /day	
k <sub>binding</sub>	1,000	/nmole per day	
k <sub>Ah</sub> degradation	2.16	/day	
k <sub>Ah</sub> TCDD degradation	5.15	/day	
k <sub>proteolysis</sub>	0.2727	/day	
k <sub>deadenvlation</sub>	576	nt/day	
k <sub>mRNA</sub> degradation	13.4	/day	
k <sub>urine</sub>	5.36	/day	
k <sub>bile</sub>	3.81	/day	
k <sub>feces</sub>	1.152	/day	
k <sub>lysis</sub>	200	/day	
critical <sub>accumulation</sub>	0.6	nmole	
k <sub>recoverv</sub>	0.01	/day	
critical concentration	2	nM	

# TABLE H2Partition Coefficients and Permeability Coefficients

	Partition	Permeability	
Fat	187.0	0.024	
Muscle	4.48	0.171	
Viscera	3.35	0.334	
Liver	4.60	1.49	
Kidney	3.35	0.559	
Gastrointestinal tract	3.35	0.248	
Lung	4.57 <sup>a</sup>	0.85 <sup>a</sup>	

<sup>a</sup> These coefficients are estimated

## TABLE H3 Gene Expression Parameters Given in the Kohn *et al.* (2001) Model

	Aryl Hydrocarbon				
	Receptor	CYP1A1	CYP1A2	CYP1B1	
Expression nmole/L per day	0.0177	0.023	0.42	0.00001	
Induction (V) nmole/L per day	$0.08^{a}$	13.5	55.0 <sup>a</sup>	3.92	
Induction (K) nM	8.3 <sup>a</sup>	2.04	4.41	13	
Induction (N) nt	128	156	196	74	
Induction (Ka) nM	NA	1.16	NA	10.0	
Synthesis (V) nmole/L per day	3,000	3,000	2,400	520	
K <sub>ribosome</sub> nM	2.68	NA	NA	NA	

<sup>a</sup> From optimization

## TABLE H4Physiological Parameters

	Fraction of Body Weight	Fraction Capillary	Fraction of Cardiac Output
<b>.</b>	0.0272	0.129	0.020
Liver	0.03/3	0.138	0.039
Fat	0.07	0.02	0.065
Muscle	0.542	0.02	0.334
Viscera	0.163	0.075	0.248
Kidney	0.0148	0.16	0.133
Gastrointestinal tract	0.075	0.0265	0.181
Lung	0.005	0.36	_
Arterial	0.0044	_	_
Venous	0.0132	_	_
Liver (hepatic artery only)			

# TABLE H5Initial Conditions for Protein in Blood<sup>a</sup>

Arterial	0.539
Venous	1.616
Gastrointestinal tract	0.0755
Fat	0.283
Muscle	0.741
Viscera	0.413
Liver	0.328
Kidney	0.0778

<sup>a</sup> Conditions are measured in nmoles.

TABLE H6
nitial Conditions for Cytochrome P450s and the Aryl Hydrocarbon Receptor (AhR) <sup>a</sup>

<sup>a</sup> Conditions are measured in nmoles.



Model Predictions (-) and Individual Tissue Data (O) for the Vehicle Control Group in the 2-Year Study The limit of detection is shown with a dotted line.



FIGURE H2 Model Predictions (-) and Individual Tissue Data (O) for the 3 ng/kg Group in the 2-Year Study The limit of detection is shown with a dotted line.



FIGURE H3 Model Predictions (-) and Individual Tissue Data (O) for the 10 ng/kg Group in the 2-Year Study The limit of detection is shown with a dotted line.









Model Predictions (-) and Individual Tissue Data (O) for the 46 ng/kg Group in the 2-Year Study The limit of detection is shown with a dotted line.



FIGURE H6

Model Predictions (-) and Individual Tissue Data (O) for the 100 ng/kg Group in the 2-Year Study The limit of detection is shown with a dotted line.





Model Predictions (-) and Individual Tissue Data (O) for the 100 ng/kg Stop-Exposure Group in the 2-Year Study The limit of detection is shown with a dotted line.

### APPENDIX I ASSOCIATED PUBLICATIONS

The following peer reviewed journal publications have been published using data or special study samples obtained from this study and other studies carried out as part of the dioxin TEF evaluation.

Brix, A.E., Jokinen, M.P., Walker, N.J., Sells, D.M., and Nyska, A. (2004). Characterization of bronchiolar metaplasia of the alveolar epithelium in female Sprague-Dawley rats exposed to 3,3',4,4',5-pentachlorobiphenyl (PCB126). *Toxicol. Pathol.* **32**, 333-337.

Brix, A.E., Nyska, A., Haseman, J.K., Sells, D.M., Jokinen, M.P., and Walker, N.J. (2005). Incidences of selected lesions in control female Harlan Sprague-Dawley rats from two-year studies performed by the National Toxicology Program. *Toxicol. Pathol.* **33**, 477-483.

Hailey, J.R., Walker, N.J., Sells, D.M., Brix, A.E., Jokinen, M.P., and Nyska, A. (2005). Classification of proliferative hepatocellular lesions in Harlan Sprague-Dawley rats chronically exposed to dioxin-like compounds. *Toxicol. Pathol.* **33**, 165-174.

Hassoun, E.A., Li, F., Abushaban, A., and Stohs, S.J. (2000). The relative abilities of TCDD and its congeners to induce oxidative stress in the hepatic and brain tissues of rats after subchronic exposure. *Toxicology* **145**, 103-113.

Hassoun, E.A., Li, F., Abushaban, A., and Stohs, S.J. (2001). Production of superoxide anion, lipid peroxidation and DNA damage in the hepatic and brain tissues of rats after subchronic exposure to mixtures of TCDD and its congeners. *J. Appl. Toxicol.* **21**, 211-219.

Hassoun, E.A., Wang, H., Abushaban, A., and Stohs, S.J. (2002). Induction of oxidative stress in the tissues of rats after chronic exposure to TCDD, 2,3,4,7,8-pentachlorodibenzofuran, and 3,3',4,4',5-pentachlorobiphenyl. *J. Toxicol. Environ. Health* A **65**, 825-842.

Jokinen, M.P., Walker, N.J., Brix, A.E., Sells, D.M., Haseman, J.K., and Nyska, A. (2003). Increase in cardiovascular pathology in female Sprague-Dawley rats following chronic treatment with 2,3,7,8-tetrachlorodibenzo-p-dioxin and 3,3',4,4',5-pentachlorobiphenyl. *Cardiovasc. Toxicol.* **3**, 299-310.

Lee, H.M., He, Q., Englander, E.W., and Greeley, G.H., Jr. (2000). Endocrine disruptive effects of polychlorinated aromatic hydrocarbons on intestinal cholecystokinin in rats. *Endocrinology* **141**, 2938-2944.

Nyska, A., Jokinen, M.P., Brix, A.E., Sells, D.M., Wyde, M.E., Orzech, D., Haseman, J.K., Flake, G., and Walker, N.J. (2004). Exocrine pancreatic pathology in female Harlan Sprague-Dawley rats after chronic treatment with 2,3,7,8-tetrachlorodibenzo-p-dioxin and dioxin-like compounds. *Environ. Health Perspect.* **112**, 903-909.

Nyska, A., Yoshizawa, K., Jokinen, M.P., Brix, A.E., Sells, D.M., Wyde, M.E., Orzech, D.P., Kissling, G.E., and Walker, N.J. (2005). Olfactory epithelial metaplasia and hyperplasia in female Harlan Sprague-Dawley rats following chronic treatment with polychlorinated biphenyls. *Toxicol. Pathol.* **33**, 371-377.

Tani, Y., Maronpot, R.R., Foley, J.F., Haseman, J.K., Walker, N.J., and Nyska, A. (2004). Follicular epithelial cell hypertrophy induced by chronic oral administration of 2,3,7,8-tetrachlorodibenzo-p-dioxin in female Harlan Sprague-Dawley rats. *Toxicol. Pathol.* **32**, 41-49.

Toyoshiba, H., Walker, N.J., Bailer, A.J., and Portier, C.J. (2004). Evaluation of toxic equivalency factors for induction of cytochromes P450 CYP1A1 and CYP1A2 enzyme activity by dioxin-like compounds. *Toxicol. Appl. Pharmacol.* **194**, 156-168.

Vezina, C.M., Walker, N.J., and Olson, J.R. (2004). Subchronic exposure to TCDD, PeCDF, PCB126, and PCB153: Effect on hepatic gene expression. *Environ. Health Perspect.* **112**, 1636-1644.

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Yoshizawa, K., Walker, N.J., Jokinen, M.P., Brix, A.E., Sells, D.M., Marsh, T., Wyde, M.E., Orzech, D., Haseman, J.K., and Nyska, A. (2005). Gingival carcinogenicity in female Harlan Sprague-Dawley rats following two-year oral treatment with 2,3,7,8-tetrachlorodibenzo-p-dioxin and dioxin-like compounds. *Toxicol. Sci.* **83**, 64-77.



### National Toxicology Program National Institute of Environmental Health Sciences

National Institute of Environmental Health Sciences National Institutes of Health P.O. Box 12233, MD K2-05 Durham, NC 27709 Tel: 984-287-3211 ntpwebrequest@niehs.nih.gov

https://ntp.niehs.nih.gov

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